

Inhibition by the ionic strength of hydrogen production from the Organic Fraction of Municipal Solid Waste

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Abstract

Composition of the Organic Fraction of Municipal Solid Waste (OFMSW) in organic compounds and inorganic ions is highly variable and might impact the microbial activity in dark fermentation processes. In this study, the effect of the total amount of inorganic ions on fermentative hydrogen production was investigated. Batch experiments were carried out at pH 6 and under a temperature of 37°C. A freshly reconstituted organic fraction of municipal solid waste (OFMSW) was used as model substrate. At low concentrations in ammonium or chloride ions (2.9 - 5.1 g.L⁻¹, respectively), the hydrogen yield reached a maximum of 40.8±0.5. mLH₂.gVS⁻¹ and 25.1±5.6 mLH₂.gVS⁻¹. In contrast, at high total ionic concentrations of ammonium and chloride (11.1 - 35.5 g.L⁻¹ respectively), a strong inhibition of the fermentative microbial activity and more particularly hydrogen production, was observed. When considering the ionic strength of each ion, the effects of ammonia, chloride or a mixture of different ions (Na⁺, K⁺, H⁺, Li⁺, NH₄⁺, Mn²⁺, NH₄⁺, Mg²⁺, Cl⁻, PO₄³⁻, Br⁻, I⁻, SO₄²⁻) showed very similar inhibitory trends regardless the type of ion or the composition of the ionic mixture. A threshold inhibitory value of the ionic strength was estimated at 0.75±0.13 M with a substantial impact on the fermentative activity from 0.81±0.12 M, with hydrogen yields of 18.1±3.3 and 6.2±4.1 mLH₂.gVS⁻¹, respectively.

27 Microbial community composition was also significantly impacted with a specific decrease
28 in relative abundance of hydrogen-producing bacteria from the genus *Clostridium* sp. at
29 high ionic strength.

30 *Keywords: Biohydrogen; Dark fermentation; Total ammoniacal nitrogen; Ionic strength*

31

32 **1. Introduction**

33

34 Nowadays, fossil fuels are the main resources in primary energy, and their usages have
35 important effects on global warming. Due to a rapid decrease of their stocks and a constant
36 increase of the energy demand, it is now necessary to find new alternatives that are not only
37 sustainable and environment-friendly but also widely available. In this context, organic
38 biomasses, including the organic fraction of municipal solid waste (OFMSW) are well
39 adapted resources. In 2012, Hoornweg and Bhada-Tata [1] reported that urban cities
40 generated about 1.3 billion tons of solid waste per year worldwide. This amount is expected
41 to increase up to 2.2 billion tons in 2025. In addition, the organic fraction of municipal
42 solid waste (OFMSW) has a high calorific value making the OFMSW highly suitable for
43 bioenergy production [2,3]. Historically, methane has been the main gas biologically
44 produced from OFMSW [3], due to the maturity of the technology in terms of
45 performances and conversion efficiency with advanced automation and modelling [4].
46 Indeed, Anaerobic Digestion (AD) is a well-established biological process used to convert
47 organic matter into a biogas that can further used as renewable energy. AD-based waste
48 treatment plants are promoted by the European Parliament [5] to use “the waste as a source
49 of energy”. In more recent years, hydrogen production by dark fermentation has gained an

50 increasing interest since H₂ is considered as an important energy carrier for next-generation
51 technologies towards decarbonation of the industrial sector [6]. Indeed, H₂ has a higher heat
52 of combustion (119.93 kJ.g⁻¹) than methane (50.02 kJ.g⁻¹) [7] and water is the only end-
53 product generated by its combustion or its use in fuel cells [8]. Hydrogen can also be used
54 as a fuel for vehicles and as a reactant for many chemical reactions (crack hydrocarbons,
55 sulfur or nitrogen compounds removing) [9]. The biological production of hydrogen by
56 dark fermentation is an intermediate step of AD and could be implemented in AD-based
57 plants treating OFMSW. During the dark fermentation process, hydrolytic bacteria convert
58 complex organic substrate into monomers which can be easily assimilated by fermentative
59 hydrogen producing bacteria (HPB), such as *Clostridium* sp. [10,11]. Many studies have
60 already shown the possibility to produce hydrogen from OFMSW, by using mixed culture
61 in continuous stirred tank or batch reactor, for a H₂ yield ranging from 20 to 70.1
62 mLH₂.gVS⁻¹ [10,12]. In AD treatment plants, hydrogen and methane can be both produced
63 by decoupling dark fermentation and methanogenesis in a two-stage AD process [14]. By
64 separating hydrolysis/acidogenesis and methanogenesis, each process can be optimized
65 with adequate operating conditions for improving the conversion of OFMSW into energy
66 [15]. In the first stage, 7.5 to 15% of the energy contained in the organic substrate can be
67 converted into hydrogen and co-metabolites, such as volatile fatty acids (VFAs) [16]. The
68 remaining organic matter and the VFAs can then be converted into methane in the second
69 stage in order to transform a maximum of anaerobically biodegradable organic matter into
70 energy.

71 However, hydrogen accumulation in a dark fermenter is often limited by the presence of
72 hydrogen-consuming anaerobes such as methanogens [17] or the accumulation of inhibitors

73 such as VFAs, chloride and total ammonia nitrogen (TAN) [18,19]. When using OFMSW
74 as organic substrate, ammonia nitrogen and chloride ions are often found in high
75 concentrations [20] due to, respectively, the decomposition of proteins and amino acids
76 [21] and the addition of sodium chloride (salt) during food preparation. Ammonia nitrogen,
77 defined as the sum of ammoniac gas (NH_3) and ammonium ions (NH_4^+), was already
78 reported as an inhibitor of the microbial activity, especially under its non ionic form (NH_3)
79 [22]. Ammonia inhibition has also been suggested on fermentative hydrogen production,
80 but the effects are not yet well described. As an illustration, Salerno et al. [23] showed a
81 negative effect on hydrogen production at ammonia nitrogen concentrations higher than 1.6
82 gN.L^{-1} , i.e. $0.4 \text{ mgNH}_3.\text{L}^{-1}$, in a continuous reactor. In contrast, a neutral effect on hydrogen
83 yields in batch test was observed in the same study, even at higher concentration of
84 ammonia nitrogen (1.3 and $13 \text{ mgNH}_3.\text{L}^{-1}$). Cavinato et al. [24] reported an inhibitory
85 effect on hydrogen production with a change on metabolic pathways when ammonia
86 nitrogen concentration increased from 0.97 to 1.98 gN.L^{-1} in a two stage process treating
87 food waste.

88 Halophilic conditions can also be detrimental to hydrogen production in particular in
89 OFMSW rich in chloride ions. Pierra et al. [25] showed inhibition of the hydrogen
90 production at NaCl concentrations higher than 9 gNaCl.L^{-1} in the medium, using glucose
91 as substrate.

92 To better understand the effect of these ions on the microbial activity in dark fermentation,
93 the impact of ion addition on hydrogen production from OFMSW was investigated by
94 decoupling the effect of a specific ion and the global ionic strength of the medium. The aim
95 of this study was therefore (a) to evaluate the individual impact of ammonia and chloride

96 ion concentrations on hydrogen production, and (b) to investigate the impact of the total
97 ionic strength on the fermentative metabolism with different types of ions.

98

99 2. Materials and Methods

100 2.1. Feedstock and inoculum preparation

101 A reconstituted organic fraction of MSW was freshly prepared and its composition was
102 representative of the average composition of OFMSW collected in France on a yearly basis,
103 according to the national characterization of MSW (MODECOMTM, 1993). The proportions
104 of each component are described in Table 1.

105

106 Table 1 : Average composition of the reconstituted OFMSW. Data are expressed in % of wet
107 weight (% w/w).

Category	Components	% w/w
Food waste	Meat	7.0
	Coffee grounds	3.9
	Rice	4.3
	Potatoes	20.9
	Bread	5.1
	Yogurt	2.0
Garden waste	Grass	5.0
Papers	Papers	35.1
Cardboards	Cardboards	16.7

108

109 Meat, rice, potatoes and coffee grounds were first cooked and mixed with yogurt and bread.

110 Garden waste, paper and cardboards were shredded and sieved at 1 cm. Total solid (TS)

111 and volatile solid (VS) contents of the reconstituted OFMSW were 0.74 ± 0.01 gTS.g⁻¹ and

112 0.63 ± 0.01 gVS.g⁻¹, respectively.

113 Microbial anaerobic inoculum corresponded to a sample of an anaerobic lagoon treating
114 leachates from methanogenic storage cells in a real MSW landfill (TRIFYL). The initial pH
115 was 7.64, with a TS and VS concentration of 20.5 ± 0.3 gTS.L⁻¹ and 7.4 ± 0.2 gVS.L⁻¹,
116 respectively. Since the leachate initially contained a high quantity of ammonium ions
117 (4.02 ± 0.79 gN.L⁻¹), a stripping pretreatment was carried out prior to experimentation.
118 Stripping corresponded to a flush of the liquid phase of the leachate with air during 3 hours
119 at pH 9, at a temperature of 90°C. Nitrogen removal efficiency was 95 %. Prior stripping,
120 the microbial community was removed by centrifugation (8000 rpm during 30 min) to not
121 be affected by the stripping treatment of the leachate. At the end of the stripping treatment,
122 the temperature was stabilized at 25°C, the pH adjusted at 6 and the microbial community
123 was reintroduced.

124

125 2.2. Experimental procedure

126

127 The batch reaction was carried out in a 500 mL flask with a working volume of 400 mL.
128 Each experiment was carried out at 37°C. Culture medium contained 9.25 gVS of OFMSW
129 inoculated with 136 g of stripping-pretreated leachate diluted with 249.4 g of water to reach
130 a S/X (substrate/biomass) ratio of 10 (on VS basis) [26]. pH was adjusted at 6 with NaOH
131 (1 M). Batch reactors were then flushed with N₂ before experiment to remove oxygen
132 traces. For each experiment, different solutions of ions were added to obtain a large range
133 of ionic concentrations in the reactors, as described in the next section. Samples of 2 mL
134 were collected and stored at -20°C at the beginning and the end of each experiment for

135 metabolite and microbial analyses. A total of 152 batch assays was performed, with
136 triplicates for each condition tested.

137

138 2.2.1. Impact of ammonium and chloride ions on hydrogen production

139

140 The first part of the study was carried out with three different types of ionic solutions. The
141 first experiment consisted in adding NH_4Cl only (Ammonium-Chloride solution), under a
142 concentration ranging from 0 to 18.5 gN.L^{-1} for ammonia ions and from 0 to $42 \text{ gCl}^{-1}.\text{L}^{-1}$ for
143 chloride ions. In the second experiment, a mixture of different anions coupled to NH_4^+
144 (NH_4Cl , $\text{NH}_4\text{H}_2\text{PO}_4$, $(\text{NH}_4)_2\text{SO}_4$) was added to avoid the effect of chloride ion. The final
145 concentration in ammonium ions ranged from 0 to 22.8 gN.L^{-1} (Ammonium-X solution).
146 Finally, in the last experiment, a mixture of different cations coupled to Cl^- (CaCl_2 , KCl ,
147 NaCl) (Chloride-X solution) was evaluated at a concentration range of 0 - $40 \text{ gCl}^{-1}.\text{L}^{-1}$.

148

149 2.2.2. Impact of ionic strength on hydrogen production

150

151 The second part of the study was carried out to evaluate the impact of the total ionic
152 strength regardless the type of ions and minimize the individual concentration. Two
153 different solutions composed of seven and six components were used, corresponding to the
154 main ions found in OFMSW leachate [20] : solution M1 (NaCl , KH_2PO_4 , LiBr , KI ,
155 $(\text{NH}_4)_2\text{SO}_4$, MnCl_2 , MgSO_4) and solution M2 (NaCl , KH_2PO_4 , LiBr , KI , $(\text{NH}_4)_2\text{SO}_4$, LiCl).
156 A range of concentration from 0 to 1.9 M in term of ionic strength was tested. For M1 and
157 M2, each components were added at the same concentrations depending of the fixed ionic
158 strength (i.e. for M1 at ionic strength of 0.161 M, a concentration of 0.016 mol.L^{-1} of each

159 component was added meaning a concentration of 0.9, 2.2, 1.4, 2.7, 2.1, 3.2 and 4.0 g.L⁻¹
160 for NaCl, KH₂PO₄, LiBr, KI, (NH₄)₂SO₄, MnCl₂, MgSO₄ respectively). In this mixture,
161 ammonium and chloride concentrations did not exceed 1.4 gN.L⁻¹ and 8.4 gCl⁻.L⁻¹ (i.e. 0.08
162 and 0.24 M respectively).

163

164 2.3. Analytical methods

165 Gas pressure and composition were periodically measured (every four hours) with an
166 automated multiplexed micro-gas chromatograph (μGC R3000, SRA instrument), as
167 described by Cazier et al. [27]. Cumulated hydrogen was estimated by automatic and
168 periodic gas pressure measurement.

169 Volatile fatty acids (VFAs) were measured at the beginning and at the end of each
170 experiment. Individual VFA concentrations were determined with a gas chromatograph
171 Perkin Clarus 580 with an Elite-FFAP crossbond[®]carbowax[®] 15 m column connected to a
172 flame ionization detector at 280°C and N₂ at 6mL.min⁻¹ as carrier gas, as previously
173 described by Motte et al. [28].

174 For microbial analyses, samples were collected at the end of each experiment. DNA was
175 extracted and purified with the Fast DNA SPIN kit for soil in accordance with
176 manufacturer's instructions (MP Biomedicals). DNA quantity and purity were assessed by
177 spectrophotometry (Infinite NanoQuant M200, Tecan). The V4 and V5 regions of the 16S
178 rRNA genes were then amplified and sequenced by Illumina MiSeq (get.genotoul.fr) and
179 sequences were analysed as described elsewhere [29].

180

181

182 2.4. Data analysis and calculation

183

184 A modified Gompertz equation based model was fitted to the cumulative hydrogen
185 production curve to assess the three following kinetic parameters: the hydrogen yield, the
186 maximal hydrogen production rate and the lag phase [29].

187

188
$$H(t)=H \exp\{-\exp[R_e/H (\lambda-t)+1]\}$$
 (1)

189

190 where $H(t)$ is cumulative hydrogen amount (mL) at time t , H is the hydrogen potential
191 (mL), R is maximal hydrogen production rate (mL.h⁻¹), λ is the lag-phase and t the time
192 (hours).

193 The ionic strength was calculated according to the dimensional equation as described by
194 Solomon [30].

195

196
$$I= (1/2) \times \sum (C_i Z_i^2)$$
 (2)

197

198 where I is the ionic strength of a solution (mol.l⁻¹), C_i the concentration of each individual
199 ion (mol.l⁻¹) and Z_i the charge state of each ion.

200

201 **3. Results and discussion**

202

203 3.1. Effect of ammonium and chloride ions on hydrogen production

204 In all experiments, no methane production was observed confirming that initial heat shock
205 pretreatment of the inoculum was efficient to suppress methanogens. Hydrogen production
206 from OFMSW was first evaluated at different concentrations of NH_4Cl (Ammonium-
207 Chloride) from 0 to 18.5 gN.L^{-1} (Fig. 1(a)). Overall, apart from one sample (containing 2.0
208 gN.L^{-1}), a good fitting of the data to the Gompertz model was observed on the hydrogen
209 response (details are presented in supplementary material).

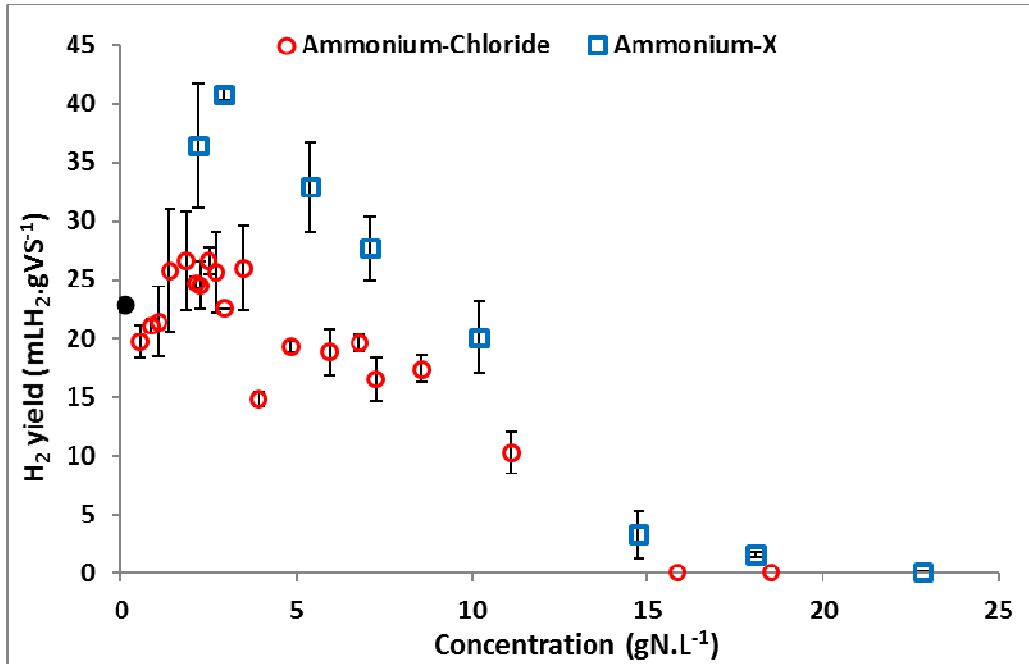
210 In the control where no ion was added, the hydrogen yield reached $22.9 \pm 0.1 \text{ mLH}_2.\text{gVS}^{-1}$
211 with a lag phase of 5.9 ± 0.1 hours and a productivity of $3.29 \pm 0.12 \text{ mLH}_2.\text{gVS}^{-1}.\text{h}^{-1}$.
212 Consistently, Favaro et al. [13] reported a similar hydrogen yield of $23.4 \pm 2.9 \text{ mLH}_2.\text{gVS}^{-1}$
213 using real OFMSW in a reactor operated at mesophilic temperature with a pH buffered at 7.
214 However, this value is lower than the yields reported by Pan et al. [31] or Elbeshbishy et al.
215 [32] who used batch reactors with no heat shock treatment or pH regulation (39 and 40
216 $\text{mLH}_2.\text{gVS}^{-1}$ respectively). In these study, more fermentescible food waste was used as
217 substrate which probably explains these differences in hydrogen production. As shown in
218 Fig. 1, when NH_4Cl was added at low concentration (from 1.4 to 3.5 gN.L^{-1}) and in
219 comparison with the control, a slight but significant increase of 8% ($p\text{-value} < 0.001$) was
220 observed with an average yield of $24.9 \pm 2.5 \text{ mLH}_2.\text{gVS}^{-1}$. The lag phase was also shortened
221 to 5.2 ± 0.5 hours. A further increase of the ammonia nitrogen concentration between 3.5
222 and 8.6 gN.L^{-1} led to a slight decrease of the hydrogen production ($p\text{-value} < 0.001$) to
223 reach a yield of $17.8 \pm 1.5 \text{ mLH}_2.\text{gVS}^{-1}$ at 8.6 gN.L^{-1} . For concentrations higher than 11.1
224 gN.L^{-1} , a substantial decrease of the hydrogen yield down to $1.9 \pm 2.8 \text{ mLH}_2.\text{gVS}^{-1}$, and a
225 significant increase of the hydrogen lag phase up to 44.9 ± 9.3 hours were observed. Finally,

226 hydrogen was not detected for concentrations above 15.9 gN.L⁻¹ suggesting a strong impact
227 of the NH₄Cl concentration on hydrogen-producing microbial activity.

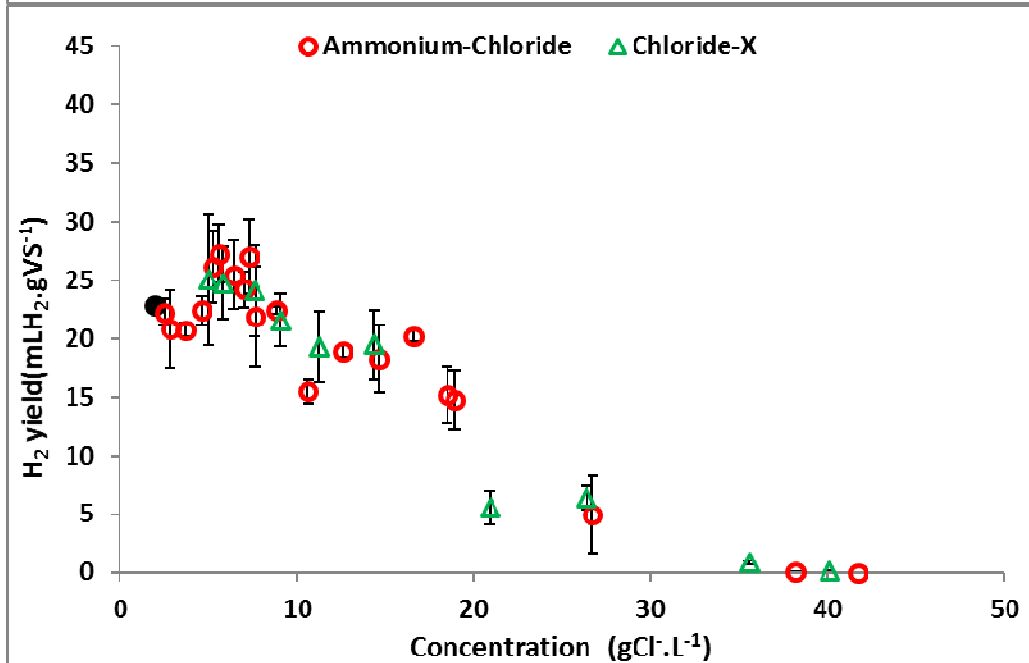
228 Thereafter, different forms of ammonia nitrogen (Ammonium-X) were added (Fig. (1a)) to
229 avoid the effect of the anion. Similarly, a positive effect on hydrogen production was
230 observed for concentrations up to 7.1 gN.L⁻¹, with a maximum at 2.9 gN.L⁻¹ (40.8±0.5.
231 mLH₂.gVS⁻¹). However, the lag phase was longer (12.8±4.7 hours) and the productivity
232 lower (1.65±0.39 mLH₂.gVS⁻¹.h⁻¹) than in the control. At higher values (10.2 gN.L⁻¹), the
233 hydrogen yield decreased to a value lower than in the control (20.1±3 mLH₂.gVS⁻¹). The
234 decrease of hydrogen yield from 10.2 gN.L⁻¹ was concomitant with an increase of the lag
235 phase (28.9±9.6 hours) and a decrease of the hydrogen productivity (0.39±0.08 mLH₂.gVS⁻¹.
236 h⁻¹), suggesting an impact on the microbial activity. At higher concentrations, a drastic
237 drop of the hydrogen production was observed with a trend similar to the experiments
238 supplemented with Ammonium-Chloride. These observations support the fact that the ions
239 added to the medium had a strong impact on the microbial activity for concentrations above
240 a threshold value of 8.6 gN.L⁻¹ for the Ammonium-Chloride solutions, and 10.2 gN.L⁻¹ for
241 Ammonium-X solutions. The relative standard deviation was calculated on the hydrogen
242 yield to evaluate the reproducibility of the triplicates (supplementary material). For NH₄Cl
243 experiments, 57 % of the triplicates had a relative standard deviation below 10 %, while 90
244 % of the triplicates were below 20 %, showing a high reproducibility. For Ammonium-X,
245 25 % of the samples had a relative standard deviation below 10 %, and 63 % below 20 %.
246 During this experiment, higher variability between the triplicates was observed, in
247 particular when ammonia nitrogen was added at low concentration. Interestingly variability

248 decreased with the increase of ammonia nitrogen concentration with a good reproducibility
249 of the impact of ammonia-chloride addition on hydrogen production.

250



251



252 Figure 1: Hydrogen yields estimated by Gompertz model in batch reactors operated at (a)
253 increasing total ammonia nitrogen concentrations in the forms of [NH₄Cl] only (●) or [NH₄Cl,
254 NH₄H₂PO₄, (NH₄)₂SO₄] (□) and (b) increasing chloride concentrations in the forms of [NH₄Cl] only
255 (○) or [CaCl₂, KCl, and NaCl] (Δ), control (●).

256

257 Similar experiments were then performed by adding a mixture of CaCl₂, KCl, NaCl
258 (Chloride-X) to assess the impact of chloride ions (Fig. (1b)). The trend of the inhibition
259 curve was statistically similar (p-value < 0.05) to the experiment where chloride ion was
260 added in the form of NH₄Cl only (Ammonium-Chloride experiments). Indeed, hydrogen
261 production was not affected between 0 to 10.2 gCl⁻.L⁻¹ (24.8±4.5 mLH₂.gVS⁻¹) with an
262 average lag phase and a productivity of 9.9±2.1 hours and 1.56 mLH₂.gVS⁻¹.h⁻¹,
263 respectively. From 20 gCl⁻.L⁻¹ to 40 gCl⁻.L⁻¹ hydrogen production decreased and a total
264 inhibition was observed at 35.5 gCl⁻.L⁻¹. By assessing the relative standard deviation of the
265 Chloride-X experiments, 100% of the triplicates had a relative standard deviation below 20
266 %, showing a high reproducibility between the triplicates.

267 When combined to the results of the experiments carried out with Ammonium-Chloride
268 solutions, it was concluded a strong inhibition of both ammonia nitrogen and chloride ions
269 on hydrogen pathway. Several studies already reported that the non-ionic form of ammonia
270 nitrogen (NH₃) can act as an inhibitor on the hydrogen production due to its high
271 permeability to bacterial cell membrane [33]. In our study, a strong inhibitory effect was
272 observed at an ammonia concentration of 8.6 gN.L⁻¹, equivalent to 11 mgNH₃.L⁻¹.
273 Comparatively, Salerno et al. [23] reported that the maximum hydrogen production rate
274 decreased from 56 mLH₂.h⁻¹ at 2 gN.L⁻¹ to 16 mLH₂.h⁻¹ at 10 gN.L⁻¹ (i.e. 13 mgNH₃.L⁻¹),

275 respectively, in a batch reactor operated at pH 6.2, supplement with NH_4Cl and fed with
276 glucose. However, no impact on hydrogen yield was observed ($1.0 \pm 0.04 \text{ molH}_2 \cdot \text{mol}_{\text{glucose}}^{-1}$,
277 i.e. $120 \text{ mLH}_2 \cdot \text{gVS}^{-1}$). Wang et al. [34] showed an inhibitory effect on hydrogen production
278 at a concentration of ammonia nitrogen of $5 \text{ gN} \cdot \text{L}^{-1}$ ($64 \text{ mgNH}_3 \cdot \text{L}^{-1}$) using glucose as
279 substrate, with an initial pH of 7 and a reactor operated at mesophilic temperature.

280 In addition, Pan et al. [22] showed an improvement of the hydrogen yield at concentrations
281 of ammonia nitrogen as low as $1.5 \text{ gN} \cdot \text{L}^{-1}$ and $3.5 \text{ gN} \cdot \text{L}^{-1}$, in a batch reactor using food
282 waste as substrate. In the present study, similar improvement was observed at $3.5 \text{ gN} \cdot \text{L}^{-1}$
283 showing the benefic effect of ammonia nitrogen on microbial activity. When different
284 forms of ammonium (Ammonium-X) were added, a higher positive effect on hydrogen
285 production was observed suggesting that the molecules added ($\text{NH}_4\text{H}_2\text{PO}_4$, $(\text{NH}_4)_2\text{SO}_4$)
286 were probably lacking in the OFMSW. Such observation confirms the positive effect of
287 phosphorus and nitrogen on hydrogen producing bacteria activity at low concentration as
288 already shown by Wang and Wan [35].

289 In comparison, no positive effect of chloride ion addition was observed in this study. Such
290 negative impact on hydrogen production of the Chloride-X solutions is consistent with the
291 literature where chloride ions are described as strong inhibitors of the microbial activity in
292 anaerobic bioreactors. Lefebvre et al. [36] showed an inhibitory effect in anaerobic
293 digestion at a concentration of NaCl of $20.0 \text{ g} \cdot \text{L}^{-1}$ (i.e. a chloride concentration of 12.1 gCl^-
294 $\cdot \text{L}^{-1}$), using ethanol as carbon source. In fermentation, Pierra et al. [25] observed a decrease
295 of the hydrogen production at NaCl concentrations as low as $9 \text{ gNaCl} \cdot \text{L}^{-1}$ (i.e. $5.4 \text{ gCl}^- \cdot \text{L}^{-1}$).

296 In comparison, our study showed an inhibitory effect on hydrogen production from 10.2
297 $\text{gCl}^- \cdot \text{L}^{-1}$.

298

299 In conclusion, the comparison of the three experiments clearly shows that ammonia and
300 chloride ions had the same inhibition trend in terms of hydrogen yield, hydrogen lag phase
301 and hydrogen productivity. Indeed the hydrogen yield decreased of about 92% and 89% for
302 the Ammonium-Chloride and Ammonium-X experiments respectively, when increasing the
303 ammonium concentration from $[0-5] \text{ gN} \cdot \text{L}^{-1}$ to $>10 \text{ gN} \cdot \text{L}^{-1}$. A very similar trend was
304 observed with the chloride ions, with a decrease of the hydrogen yield by 89% from the low
305 $[0-10] \text{ gCl}^- \cdot \text{L}^{-1}$ to the high concentrations (up to $20 \text{ gCl}^- \cdot \text{L}^{-1}$) of chloride, in the Chloride-X
306 experiments.

307 Consistently, an increase of the ammonium and chloride ion concentration led to an
308 increase of the lag phase for all the tested solutions. A very similar impact was shown on
309 the hydrogen productivity, regardless the type of ion added, with a maximum of 3.04 ± 0.71 ,
310 1.48 ± 0.52 and $1.56 \pm 0.81 \text{ mLH}_2 \cdot \text{gVS}^{-1} \cdot \text{h}^{-1}$ observed at low concentrations for Ammonium-
311 Chloride, Ammonium-X and Chloride-X solutions, respectively, i.e. within the range of $[0-$
312 $5] \text{ gN} \cdot \text{L}^{-1}$ and $[0-10] \text{ gCl}^- \cdot \text{L}^{-1}$.

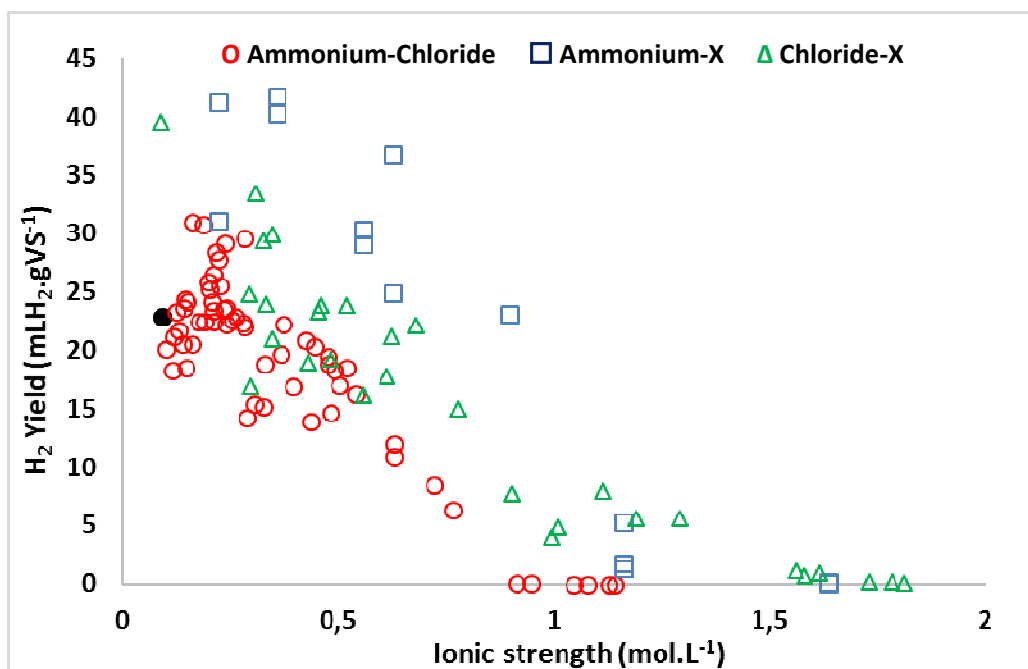
313 These results suggest that a global metabolic inhibition could have occurred and might be
314 responsible of the negative impact on microbial fermentation, independently of the nature
315 of the ion.

316

317 3.2. Impact of the ionic strength on dark fermentation

318 3.2.1. Impact of the ionic strength on hydrogen production

319 In light of these results, an ionic strength index was calculated to evaluate the impact of the
320 total ionic content rather than the effect of individual ion on the fermentative activity. The
321 total ionic strength was estimated based on each individual ionic concentration and their
322 number of charges. By revisiting the data, the impact of the ionic strength on hydrogen
323 production is shown in Fig. 2, for Ammonium-Chloride, Ammonium-X and Chloride-X
324 experiments.



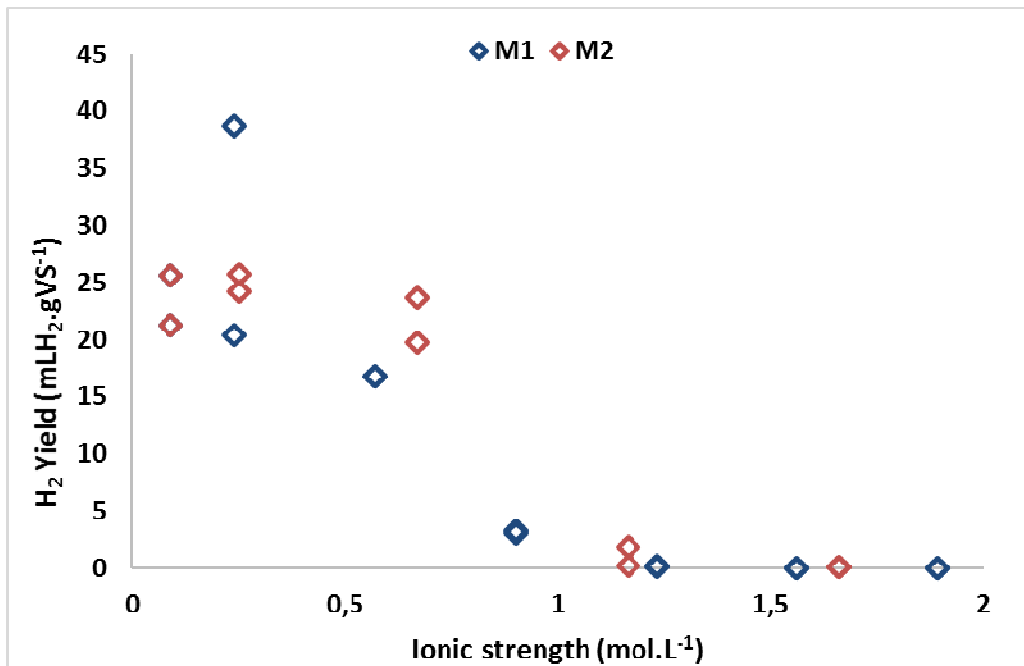
325
326 Figure 2: Effect of ionic strength on biohydrogen production in dark fermentation reactor for
327 Ammonium-Chloride (○), Ammonium-X (□), Chloride-X (△) experiments and the control (●)

328
329 As noticed previously, an improvement of the hydrogen production was observed at low
330 ionic strength, for the Ammonium-Chloride, Ammonium-X and Chloride-X experiments.
331 Such slight positive effect confirms the benefic effect of the molecules added on hydrogen
332 production at low concentration of ammonium. A similar inhibitory trend was observed in
333 all the experiments according to the value of the ionic strength. A threshold value of this

334 inhibition was estimated at 0.75 ± 0.13 M, for an average H_2 yield of 18.1 ± 3.3 mL H_2 .gVS $^{-1}$,
335 whatever the ions considered. The inhibition was significant at ionic strengths higher than
336 0.81 ± 0.12 M with an average H_2 yield of 6.2 ± 4.1 mL H_2 .gVS $^{-1}$. Total inhibition of the
337 hydrogen production was then observed at an ionic strength of 1.04, 1.63 and 1.73 M for
338 Ammonium-Chloride, Ammonium-X and Chloride-X experiments, respectively, suggesting
339 here an effect of the type of ions.

340 To better understand the impact of the ions regardless their nature, the total ionic strength
341 was modified by changing the composition of the ionic solutions and considering several
342 anions and cations, as follows: solution M1 was composed of Na^+ , K^+ , H^+ , Li^+ , NH_4^+ , Cl^- ,
343 PO_4^{3-} , Br^- , I^- , SO_4^{2-} , while solution M2 was composed of Na^+ , K^+ , H^+ , Li^+ , NH_4^+ , Mn^{2+} ,
344 Mg^{2+} , Cl^- , PO_4^{3-} , Br^- , I^- , SO_4^{2-} (Fig. 3). By increasing the number of ion types, the
345 concentration of each individual element remained low (< 0.18 M) while the ionic strength
346 increased from 0.18 to 1.9 M.

347



348

349 Figure 3: Effect of ionic strength on biohydrogen production for M1: NaCl, KH₂PO₄, LiBr, KI,
 350 (NH₄)₂SO₄, MnCl₂.4H₂O, MgSO₄.7H₂O (◊) and M2: NaCl, KH₂PO₄, LiBr, KI, (NH₄)₂SO₄, LiCl (◊)
 351 experiments.

352

353 As shown in Fig. 3, the trend was similar than previously observed with an improvement of
 354 the hydrogen production at the lowest ionic strengths of 0.09 and 0.25 M with an average
 355 yield of 26.5±6.1 and 25.0±0.7 mLH₂.gVS⁻¹ for M1 and M2, respectively. Inhibition of the
 356 hydrogen production was significant after values of ionic strength of 0.57 and 0.67 M for
 357 the M1 and M2 solutions, which is similar to the previous observations made with
 358 Ammonium-Chloride, Ammonium-X and Chloride-X solutions (0.75±0.13 M).

359 No hydrogen production was observed for ionic strength higher than 1.56 and 1.66 M for
 360 M1 and M2, respectively. The concentration of ammonia nitrogen and chloride ions at 1.56
 361 M for M1 and at 1.66 M for M2 were 1.7 and 1.1 gN.L⁻¹ and 13.1 and 6.5 gCl⁻.L⁻¹,
 362 respectively. Interestingly, these concentrations in individual components corresponded to

363 the concentrations found at the highest performances of hydrogen production in the
364 Ammonium-Chloride experiments. These observations clearly suggest that hydrogen
365 inhibition was not due to the individual concentration in ammonium and chloride ions, but
366 to the overall ionic strength.

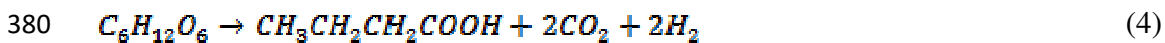
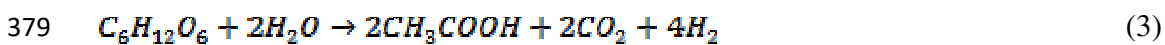
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368 3.2.2. Impact of ionic strength on by-products production

369 Furthermore, soluble fermentation co-products were analyzed at the end of the batch tests
370 to estimate the impact of the ionic solutions on the global fermentation activity. Total
371 amount of metabolic co-products was expressed in mgCOD.gVS⁻¹ (Table 2).

372 In the control experiment, i.e. without addition of ions, the main accumulated metabolites
373 were acetate and butyrate at a concentration of 104.2±8.3 mgCOD.gVS⁻¹ (2.5±0.2 gCOD.L⁻¹)
374 and 133.3±8.3 mgCOD.gVS⁻¹ (3.2±0.2 gCOD.L⁻¹) representing 36 % and 46 % of the
375 total COD, respectively.

376 Theoretically acetate and butyrate are correlated to a high hydrogen yield. Indeed, two
377 moles of acetate and one mole of butyrate leads to four and two mole of hydrogen
378 respectively (Eq. 3 and 4) [37].



381 Many studies show these metabolites as major end product during dark fermentation
382 reaction [39,25]. Cavinato et al. [24] reported the same range of acetate and butyrate
383 concentration in a thermophilic dark fermentation using food waste (3.29±1.64 and
384 4.32±1.48 gCOD.L⁻¹ respectively) which is slightly higher than our results due to

385 thermophilic application which is known to improve the microbial activity [39]. These
386 results suggest that the conditions in the control were favorable for hydrogen producing
387 pathways. Indeed, the maximum H₂ yield is related to acetate as by-product. However,
388 from an experimental point of view, high H₂ yields are usually associated to butyrate in
389 mixed culture [40].

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397 Table 2 : Final concentration of acetate, propionate, butyrate, valerate and caproate during all the experiments (Ammonium-Chloride,
 398 Ammonium-X and Chloride-X) at different ionic strength and estimation of the homoacetogenic activity in all experiments. For technical
 399 reasons, M1 solutions precipitates in the liquid fraction made the analyses no possible without damaging the analyzer.
 400

Experiments	Metabolite (mgCOD.gVS ⁻¹)							Homoacetogenesis (mol/mol)
	Ionic strength (mol.L ⁻¹)	Acetate	Propionate	Butyrate	Valerate	Caproate	Ethanol	
Control	0.09	104.2±8.3	9.6±0.8	133.3±8.3	4.6±0.8	3.3±2.5	17.5±3.7	39.9 %
Ammonium-Chloride	0.25	100.0±33.3	14.6±2.9	145.8±58.3	3.7±0.8	4.6±2.9	7.5±1.4	39.7 %
	0.35	79.2±16.7	17.5±14.2	120.8±20.8	20.8±25.0	6.7±8.3	5.0±10.0	42.1 %
	0.59	58.3±4.2	39.6±9.6	62.5±16.7	0.8±1.7	17.9±0.8	40.0±35.0	31.6 %
	1.02	14.2±2.5	1.7±0.4	-	-	-	-	32.2 %
Ammonium-X	0.28	45.8±8.3	10.0±2.9	116.7±25.0	-	-	16.7±33.3	26.3 %
	0.65	33.3±8.3	11.7±8.7	75.0±20.8	-	-	3.3±6.7	22.7 %
	1.02	23.7±2.5	10.4±8.7	25.4±22.5	-	-	27.5±16.7	21.6 %
	1.39	8.3±2.1	-	-	-	-	-	24.6 %
	1.76	1.7±2.1	-	-	-	-	-	30.1 %
Chloride-X	0.35	145.8±12.5	22.9±7.5	212.5±29.2	1.2±1.7	6.2±10.0	18.3±19.0	42.7 %
	0.55	133.3±12.5	14.2±3.7	233.3±20.8	1.2±1.7	20.0±21.7	19.2±26.2	47.2 %
	0.70	79.2±16.7	8.7±1.2	133.3±25.0	-	-	-	42.8 %
	1.10	83.3±62.5	16.7±12.1	45.8±16.7	-	-	7.9±15.8	35.6 %
	1.71	12.1±5.0	-	19.6±13.7	-	-	13.3±16.7	52.3 %
M2	0.24	66.7±4.2	10.0±0.4	116.7±8.3	5.0±4.2	-	-	39.0 %
	0.57	41.7±4.2	12.9±5.0	120.8±8.3	6.2±4.0	-	-	50.4 %
	0.90	41.2±1.2	9.6±0.4	17.5±1.2	5.0±4.1	-	-	33.3 %
	1.23	31.2±0.4	10.4±0.4	15.0±0.4	5.0±4.2	-	-	36.3 %

401 Overall, the increase of ionic strength led to a decrease of the total accumulation in
 402 metabolites regardless the solutions added. Concerning the Ammonium-Chloride
 403 experiments, from 0.25 to 1.02 M, acetate and butyrate concentrations drastically dropped
 404 to nearly no production with concomitant decrease of hydrogen production. Propionate
 405 concentration reached a maximum value of 14.6 ± 2.9 mgCOD.gVS⁻¹ at 0.25 M, and
 406 decreased to 1.7 ± 0.4 mgCOD.gVS⁻¹ at 1.02 M. Valerate and caproate were only present in
 407 trace amounts up to 0.59 M and were not detected at higher ionic strengths. A similar trend
 408 was observed for Ammonium-X and M2 solutions, with a decrease of the metabolite
 409 production according to the increase of the ionic strength. At low ionic strength of 0.28 and
 410 0.24 M, a maximal production of metabolites of 189 ± 69 and 198 ± 13 mgCOD.gVS⁻¹,
 411 respectively, were observed. The total metabolite concentration reached a low value
 412 (100 ± 30 and 60 ± 10 mgCOD.gVS⁻¹) at higher ionic strengths of 1.02 and 1.23 M,
 413 respectively. Similar trends were observed for the Chloride-X experiments with a
 414 maximum of metabolite production (407 ± 80 mgCOD.gVS⁻¹) at low ionic strength (0.35 M)
 415 with acetate and butyrate as main metabolites (145 ± 12 and 212 ± 29 mgCOD.gVS⁻¹
 416 respectively). A decrease of the total concentration was observed with a low metabolite
 417 production of 61 ± 47 mgCOD.gVS⁻¹ at 1.71 M.

418 The consumption of hydrogen by homoacetogenesis to produce acetate was estimated and
 419 is presented in Table 2. The acetate produced by homoacetogenesis can be estimated by the
 420 following two equations (Eq. 5 and 6), as already described by Arooj et al. [41] :

$$421 \quad \textit{Total HAc} = \textit{HAc}_a + \textit{HAc}_b \quad (5)$$

$$422 \quad \textit{H}_2 \textit{ Yield} = 2 \times (\textit{HBu}) - (\textit{HPr}) + 2 \times (\textit{HAc}_a) - 4 \times (\textit{HAc}_b) \quad (6)$$

423 Where Total HAc is the acetate concentration; HAca and Hacb are the acetate
424 concentration from the H₂-producing (HAC_a) and H₂-consuming (HAC_b) pathways; HBu
425 the butyrate concentration and HPr the propionate concentration.

426 Indeed, hydrogen yield is theoretically associated to the stoichiometry of 2 mol of hydrogen
427 generated per mole of acetate and butyrate accumulated, while propionate consumes 1 mol
428 of hydrogen. Eq. (5) considers the production of acetate from two distinct pathways
429 corresponding to H₂-producing (HAC_a) and H₂-consuming (HAC_b) homoacetogenic
430 pathway. Considering Eq.(5) and Eq. (6), the amount of acetate produced by
431 homoacetogenesis was estimated according to molar balances and the results are presented
432 in Table 2. The estimated amount of acetate produced by homoacetogenesis pathway in the
433 control represented about 39.9% of the total COD. Consistently, Castelló et al. [42]
434 reported that 33 to 57% of the acetate was produced by homoacetogenesis using a
435 continuous stirred tank reactor fed with cheese whey. As already reported by Saady [43],
436 homoacetogenesis is associated to fast growing strict anaerobes forming spores such as
437 *Clostridium* sp. that can consume between 11% and 43% of the hydrogen produced in
438 batch. In our experiments, the homoacetogenesis ratio is stable regardless the ionic strength
439 concentration. The percentage of acetate issued from the homoacetogenesis pathway was
440 estimated at 36.4±5.3, 25.1±3.3, 44.1±6 and 39.8±7.5% for Ammonium-Chloride,
441 Ammonium-X, Chloride-X and M2 solutions whatever the ionic strength. Therefore, the
442 decrease of hydrogen performances according to the increasing ionic strengths was not
443 attributed to an increase of hydrogen consumption by homoacetogenic bacteria. Only the
444 Ammonium-X solution showed a lower amount (25.1±3.3%) resulting from a high
445 hydrogen yield observed at low ionic strength (41.1±0.8. mLH₂.gVS⁻¹at 0.31±0.08 mol.L⁻¹).

446 Overall, the analyses of metabolites showed the same impact of the ionic strength than
447 previously observed with hydrogen production, i.e. a global decrease of the metabolite
448 production when the concentration of ions in the medium increased, regardless the
449 solutions added. Further increase of the ionic strength drastically affected the acetate and
450 butyrate pathways which are the two metabolic pathways generating hydrogen [37]. Similar
451 observations were reported by Cavinato et al. [24] who showed a decrease by 72 % of the
452 acetate and butyrate concentrations when a nitrogen accumulated from 970 to 1976 mgN.L⁻¹
453 in dark fermentation, using municipal solid waste as substrate. Zheng et al. [44] observed
454 a similar decrease of the butyrate and H₂ yields by adding NaCl from 0 to 500 mM and
455 using glucose as substrate. In addition, our results showed that the increase of ionic strength
456 did not change the metabolic pathways since acetate and butyrate remain the main
457 metabolites in each experiments, with no accumulation of lactate or other by-products that
458 can be produced during microbial stress [45]. Our observations rather suggest that the
459 global microbial activity was strongly impacted by the increase of the ionic strength.

460

461 3.2.3. Impact of the ionic strength on the microbial community

462 Microbial characterization using phylogenetic affiliation of bacterial 16S rDNA was
463 performed on the Ammonium-Chloride experiment, as model experiment, at different ionic
464 strengths. The results are presented in Table 3. Only sequence having affiliations above
465 98% of similarity with sequences of known species are presented.

466

467 Table 3 : Phylogenetic affiliation of bacterial 16S rDNA from experiments Ammonium-Chloride at
 468 different ionic strength (mol.L⁻¹) using BLAST algorithm.

Closely related to	Sequence similarity (%)	Relative abundance of species in each ecosystem (%)							
		0.25	0.35	0.53	0.63	0.72	0.95	1.02	1.13
<i>Clostridium butyricum</i>	100	32	44	26	7	-	-	-	-
<i>Clostridium tertium</i>	99	3	3	4	6	9	-	-	-
<i>Enterococcus saccharolyticus</i>	99	32	23	48	39	16	-	-	-
<i>Enterococcus faecalis</i>	100	-	7	5	5	15	11	-	-
<i>Pseudomonas caeni</i>	99	5	5	4	5	2	2	1	1
<i>Pseudomonas pertucinogena</i>	99	-	-	-	-	-	20	18	22
<i>Marinocpirillum minutulum</i>	100	-	-	-	-	-	11	3	3
<i>Atopostipes suicloacalis</i>	99	-	-	-	-	-	-	28	1
Others (<i>Clostridiales</i> NC)		10	3	5	27	44	10	7	12
Others (<i>Oceanospirillales</i> NC)		-	-	-	-	-	10	12	15
Others (NC)		12	15	8	11	14	36	38	56

469

470 At low ionic strength (0.25, 0.35 and 0.53 M) the most abundant bacteria were assigned to
 471 known hydrogen producing bacteria, i.e. *Clostridium butyricum* and *Enterococcus*
 472 *saccharolyticus*. Liu et al. [46] reported high hydrogen production using *Clostridium*
 473 *butyricum* in a fermentative batch reactor at pH 6 with glucose as substrate (1.83
 474 mmol/mmol_{glucose}). The ability of *Enterococcus saccharolyticus* to hydrolyze cellobiose and
 475 produce hydrogen was previously described [47]. These observations are consistent with
 476 our experiment where freshly reconstituted OFMSW was composed of 51.8 % of paper and
 477 cardboard. *Clostridium tertium* was found in lower abundance in the experiment, and was
 478 also described as hydrogen producing bacteria [48]. The overall microbial community
 479 characterization suggests that the environment at low ionic strength was ideal for the
 480 development of hydrogen-producing bacteria.

481 A neutral impact of the ionic strength was shown at low concentration (from 0.25 to 0.53
482 M) on the microbial community composition, which is consistent with macroscopic
483 observations of high hydrogen production.

484 At intermediate ionic strength (0.63 and 0.72 M), a modification of the microbial
485 community was observed with a decrease in abundance of *Clostridium butyricum* (7 and
486 0% respectively) and an increase in unclassified *Clostridiales* (27 and 44 % respectively).

487 This correlates with the variability of hydrogen yields found in this range of ionic strength
488 suggesting that the population structure was disturbed. At higher ionic strength (above 0.72

489 M), the dominant *Clostridium butyricum*, *Clostridium tertium* and *Enterococcus*
490 *saccharolyticus* were not detected and the relative abundance in unclassified *Clostridiales*

491 drastically decreased to 10%, 7% and 12% at 0.95, 1.02, 1.13 mol.L⁻¹, respectively. This
492 observation is consistent with an absence of hydrogen production in these experiments. At

493 high ionic strength, the presence of other microbial populations was detected such as
494 *Pseudomonas pertucinogena*, *Marinocpirillum minutulum* and *Atopostipes suicloacalis*

495 which are not associated with hydrogen production. Therefore, the increase of ionic
496 strength seemed to have highly impacted hydrogen producing bacteria that are not

497 particularly adapted to high ionic environments. As reported by Van Niel et al. [49], an
498 accumulation of ions in the medium (above 175 mM) can cause a cell lysis in *Clostridium*

499 *saccharolyticus*. More generally, high concentrations of ions cause high osmolarity across
500 the bacterial that can lead to an inhibition of the microbial activity, as shown with

501 *Clostridium butyricum* [50].

502

503

504 **4. Conclusion**

505 This study provides new aspects on fermentative hydrogen production using freshly
506 reconstituted OFMSW. During dark fermentation, the presence of ions (chloride or
507 ammonia) at low concentrations did not impact hydrogen performances and was even
508 beneficial with the best performances found when 2.9 gN.L⁻¹ was added (40.8±0.5.
509 mLH₂.gVS⁻¹). However, at higher concentration of ions and whatever the ions added, a
510 very similar trend of inhibition on hydrogen production was observed. For all experiments,
511 the inhibition started after a similar threshold value of ionic strength (0.75±0.13 M)
512 whatever the ions considered from 18.1±3.3 mLH₂.gVS⁻¹ to 6.2±4.1 mLH₂.gVS⁻¹ at
513 0.81±0.12 M. Such inhibition was also observed on the overall microbial activity with a
514 decrease of metabolite production. Moreover, the accumulation of ions caused a stressful
515 environment as shown by microbial community changes with a particular impact on
516 dominant H₂-producing bacteria (*Clostridium butyricum*, *Clostridium tertium* and
517 *Enterococcus saccharolyticus*). In the context of process upscaling and implementation of
518 this technology for waste energy recovery, the ionic strength should be carefully monitored
519 to avoid reaching the critical threshold value and thus, reduce the overall efficiency of the
520 process.

521

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524 undertook the chemical analyses.

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527 **References**

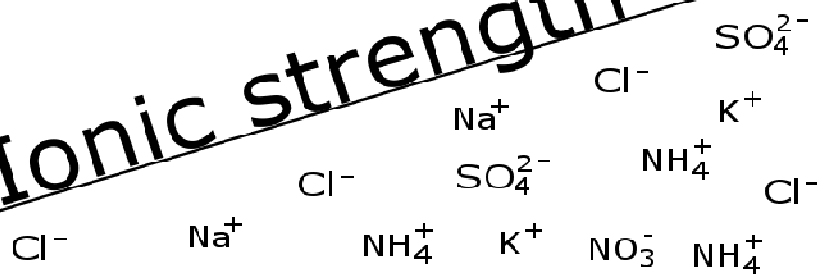
- 528 [1] Hoornweg D, Bhada-Tata P. What a Waste: A Global Review of Solid Waste
529 Management. *Urban Development Ser Knowl Pap* 2012;15:1–98.
530 doi:10.1111/febs.13058.
- 531 [2] Alibardi L, Cossu R. Composition variability of the organic fraction of municipal
532 solid waste and effects on hydrogen and methane production potentials. *Waste*
533 *Manag* 2014;36:147–55. doi:10.1016/j.wasman.2014.11.019.
- 534 [3] Fantozzi F, Buratti C. Anaerobic digestion of mechanically treated OFMSW:
535 Experimental data on biogas/methane production and residues characterization.
536 *Bioresour Technol* 2011;102:8885–92. doi:10.1016/j.biortech.2011.06.077.
- 537 [4] Mata-Alvarez J, Macé S, Llabrés P. Anaerobic digestion of organic solid wastes. An
538 overview of research achievements and perspectives. *Bioresour Technol* 2000;74:3–
539 16. doi:10.1016/S0960-8524(00)00023-7.
- 540 [5] European Parliament. Directive 2006/12/EC of the European Parliament and of the
541 Council of 5 April 2006 on waste. *Off J Eur Union* 2006;April:L 114/9-21.
- 542 [6] Chinellato G, Cavinato C, Bolzonella D, Heaven S, Banks CJ. Biohydrogen
543 production from food waste in batch and semi-continuous conditions: Evaluation of
544 a two-phase approach with digestate recirculation for pH control. *Int J Hydrogen*
545 *Energy* 2013;38:4351–60. doi:10.1016/j.ijhydene.2013.01.078.
- 546 [7] Pasupuleti SB, Venkata Mohan S. Single-stage fermentation process for high-value
547 biohythane production with the treatment of distillery spent-wash. *Bioresour*
548 *Technol* 2015;189:177–85. doi:10.1016/j.biortech.2015.03.128.
- 549 [8] Meher Kotay S, Das D. Biohydrogen as a renewable energy resource-Prospects and
550 potentials. *Int J Hydrogen Energy* 2008;33:258–63.
551 doi:10.1016/j.ijhydene.2007.07.031.
- 552 [9] Das D. Hydrogen production by biological processes: a survey of literature. *Int J*
553 *Hydrogen Energy* 2001;26:13–28. doi:10.1016/S0360-3199(00)00058-6.
- 554 [10] Yukesh Kannah R, Kavitha S, Sivashanmugham P, Kumar G, Nguyen DD, Chang
555 SW, et al. Biohydrogen production from rice straw: Effect of combinative
556 pretreatment, modelling assessment and energy balance consideration. *Int J*
557 *Hydrogen Energy* 2019;44:2203–15. doi:10.1016/j.ijhydene.2018.07.201.
- 558 [11] Chatellard L, Trably E, Carrère H. The type of carbohydrates specifically selects
559 microbial community structures and fermentation patterns. *Bioresour Technol*
560 2016;221:541–9. doi:10.1016/j.biortech.2016.09.084.
- 561 [12] Romero Aguilar M a., Fdez-Güelfo L a., Álvarez-Gallego CJ, Romero García LI.
562 Effect of HRT on hydrogen production and organic matter solubilization in
563 acidogenic anaerobic digestion of OFMSW. *Chem Eng J* 2013;219:443–9.
564 doi:10.1016/j.cej.2012.12.090.
- 565 [13] Favaro L, Alibardi L, Lavagnolo MC, Casella S, Basaglia M. Effects of inoculum

- 566 and indigenous microflora on hydrogen production from the organic fraction of
 567 municipal solid waste. *Int J Hydrogen Energy* 2013;38:11774–9.
 568 doi:10.1016/j.ijhydene.2013.06.137.
- 569 [14] Cavinato C, Bolzonella D, Fatone F, Cecchi F, Pavan P. Optimization of two-phase
 570 thermophilic anaerobic digestion of biowaste for hydrogen and methane production
 571 through reject water recirculation. *Bioresour Technol* 2011;102:8605–11.
 572 doi:10.1016/j.biortech.2011.03.084.
- 573 [15] Blonskaja V, Menert A, Vilu R. Use of two-stage anaerobic treatment for distillery
 574 waste. *Adv Environ Res* 2003;7:671–8. doi:10.1016/S1093-0191(02)00038-2.
- 575 [16] Hallenbeck PC, Ghosh D. Advances in fermentative biohydrogen production: the
 576 way forward? *Trends Biotechnol* 2009;27:287–97.
 577 doi:10.1016/j.tibtech.2009.02.004.
- 578 [17] Lay JJ. Biohydrogen generation by mesophilic anaerobic fermentation of
 579 microcrystalline cellulose. *Biotechnol Bioeng* 2001;74:280–7. doi:10.1002/bit.1118.
- 580 [18] Wang B, Wan W, Wang J. Inhibitory effect of ethanol, acetic acid, propionic acid
 581 and butyric acid on fermentative hydrogen production. *Int J Hydrogen Energy*
 582 2008;33:7013–9. doi:10.1016/j.ijhydene.2008.09.027.
- 583 [19] Zhang S, Kim TH, Lee Y, Hwang SJ. Effects of VFAs concentration on bio-
 584 hydrogen production with *Clostridium bifermentans* 3AT-ma. *Energy Procedia*
 585 2012;14:518–23. doi:10.1016/j.egypro.2011.12.968.
- 586 [20] Bakonyi P, Dharmaraja J, Shobana S, Koók L, Rózsensberszki T, Nemestóthy N, et
 587 al. Leachate valorization in anaerobic biosystems: Towards the realization of waste-
 588 to-energy concept via biohydrogen, biogas and bioelectrochemical processes. *Int J*
 589 *Hydrogen Energy* 2019. doi:10.1016/j.ijhydene.2019.02.028.
- 590 [21] Ramsay IR, Pullammanappallil PC. Protein degradation during anaerobic wastewater
 591 treatment: Derivation of stoichiometry. *Biodegradation* 2001;12:247–57.
 592 doi:10.1023/A:1013116728817.
- 593 [22] Pan J, Chen X, Sheng K, Yu Y, Zhang C, Ying Y. Effect of ammonia on
 594 biohydrogen production from food waste via anaerobic fermentation. *Int. J.*
 595 *Hydrogen Energy*, vol. 38, Elsevier Ltd; 2013, p. 12747–54.
 596 doi:10.1016/j.ijhydene.2013.06.093.
- 597 [23] Salerno MB, Park W, Zuo Y, Logan BE. Inhibition of biohydrogen production by
 598 ammonia. *Water Res* 2006;40:1167–72. doi:10.1016/j.watres.2006.01.024.
- 599 [24] Cavinato C, Giuliano A, Bolzonella D, Pavan P, Cecchi F. Bio-hythane production
 600 from food waste by dark fermentation coupled with anaerobic digestion process: A
 601 long-term pilot scale experience. *Int J Hydrogen Energy* 2012;37:11549–55.
 602 doi:10.1016/j.ijhydene.2012.03.065.
- 603 [25] Pierra M, Trably E, Godon JJ, Bernet N. Fermentative hydrogen production under
 604 moderate halophilic conditions. *Int J Hydrogen Energy* 2013;9:1–10.
 605 doi:10.1016/j.ijhydene.2013.08.035.

- 606 [26] Cappai G, De Gioannis G, Muntoni A, Poletini A, Pomi R, Spiga D. Effect of
607 inoculum to substrate ratio (ISR) on hydrogen production through dark fermentation
608 of food waste. Fifteenth International Waste Management and Landfill Symposium,
609 2015.
- 610 [27] Cazier E, Trably E, Steyer JP, Escudie R. Biomass hydrolysis inhibition at high
611 hydrogen partial pressure in solid-state anaerobic digestion. *Bioresour Technol*
612 2015;190:106–13. doi:10.1016/j.biortech.2015.04.055.
- 613 [28] Motte JC, Escudie R, Beauvils N, Steyer JP, Bernet N, Delgenes JP, et al.
614 Morphological structures of wheat straw strongly impacts its anaerobic digestion.
615 *Ind Crops Prod* 2014;52:695–701. doi:10.1016/j.indcrop.2013.11.038.
- 616 [29] Moscoviz R, Trably E, Bernet N. Consistent 1,3-propanediol production from
617 glycerol in mixed culture fermentation over a wide range of pH. *Biotechnol Biofuels*
618 2016;9:1–11. doi:10.1186/s13068-016-0447-8.
- 619 [30] Solomon T. The Definition and Unit of Ionic Strength. *J Chem Educ* 2001;78:1691–
620 2. doi:10.1021/ed078p1691.
- 621 [31] Pan J, Zhang R, El-Mashad HM, Sun H, Ying Y. Effect of food to microorganism
622 ratio on biohydrogen production from food waste via anaerobic fermentation. *Int J*
623 *Hydrogen Energy* 2008;33:6968–75. doi:10.1016/j.ijhydene.2008.07.130.
- 624 [32] Elbeshbishy E, Hafez H, Dhar BR, Nakhla G. Single and combined effect of various
625 pretreatment methods for biohydrogen production from food waste. *Int J Hydrogen*
626 *Energy* 2011;36:11379–87. doi:10.1016/j.ijhydene.2011.02.067.
- 627 [33] Müller T, Walter B, Wirtz A, Burkovski A. Ammonium toxicity in bacteria. *Curr*
628 *Microbiol* 2006;52:400–6. doi:10.1007/s00284-005-0370-x.
- 629 [34] Wang B, Wan W, Wang J. Effect of ammonia concentration on fermentative
630 hydrogen production by mixed cultures. *Bioresour Technol* 2009;100:1211–3.
631 doi:10.1016/j.biortech.2008.08.018.
- 632 [35] Wang J, Wan W. Factors influencing fermentative hydrogen production: A review.
633 *Int J Hydrogen Energy* 2009;34:799–811. doi:10.1016/j.ijhydene.2008.11.015.
- 634 [36] Lefebvre O, Quentin S, Torrijos M, Godon JJ, Delgenès JP, Moletta R. Impact of
635 increasing NaCl concentrations on the performance and community composition of
636 two anaerobic reactors. *Appl Microbiol Biotechnol* 2007;75:61–9.
637 doi:10.1007/s00253-006-0799-2.
- 638 [37] Hawkes F, Hussy I, Kyazze G, Dinsdale R, Hawkes D. Continuous dark
639 fermentative hydrogen production by mesophilic microflora: Principles and progress.
640 *Int J Hydrogen Energy* 2007;32:172–84. doi:10.1016/j.ijhydene.2006.08.014.
- 641 [38] Mamimin C, Singkhala A, Kongjan P, Suraraksa B, Prasertsan P, Imai T, et al. Two-
642 stage thermophilic fermentation and mesophilic methanogen process for biohythane
643 production from palm oil mill effluent. *Int J Hydrogen Energy* 2015;40:6319–28.
644 doi:10.1016/j.ijhydene.2015.03.068.
- 645 [39] Wang J, Wan W. Effect of temperature on fermentative hydrogen production by

- 646 mixed cultures. *Int J Hydrogen Energy* 2008;33:5392–7.
647 doi:10.1016/j.ijhydene.2008.07.010.
- 648 [40] Hawkes FR, Dinsdale R, Hawkes DL, Hussy I. Sustainable fermentative hydrogen
649 production: Challenges for process optimisation. *Int J Hydrogen Energy*
650 2002;27:1339–47. doi:10.1016/S0360-3199(02)00090-3.
- 651 [41] Arooj MF, Han SK, Kim SH, Kim DH, Shin HS. Continuous biohydrogen
652 production in a CSTR using starch as a substrate. *Int J Hydrogen Energy*
653 2008;33:3289–94. doi:10.1016/j.ijhydene.2008.04.022.
- 654 [42] Castelló E, Braga L, Fuentes L, Etchebehere C. Possible causes for the instability in
655 the H₂ production from cheese whey in a CSTR. *Int J Hydrogen Energy*
656 2018;43:2654–65. doi:10.1016/j.ijhydene.2017.12.104.
- 657 [43] Saady NMC. Homoacetogenesis during hydrogen production by mixed cultures dark
658 fermentation: Unresolved challenge. *Int J Hydrogen Energy* 2013;38:13172–91.
659 doi:10.1016/j.ijhydene.2013.07.122.
- 660 [44] Zheng XJ, Zheng YM, Yu HQ. Influence of NaCl on Hydrogen Production from
661 Glucose by Anaerobic Cultures. *Environ Technol* 2005;26:1073–80.
- 662 [45] Noike T, Takabatake H, Mizuno O, Ohba M. Inhibition of hydrogen fermentation of
663 organic wastes by lactic acid bacteria. *Int J Hydrogen Energy* 2002;27:1367–71.
664 doi:10.1016/S0360-3199(02)00120-9.
- 665 [46] Liu IC, Whang LM, Ren WJ, Lin PY. The effect of pH on the production of
666 biohydrogen by clostridia: Thermodynamic and metabolic considerations. *Int J*
667 *Hydrogen Energy* 2011;36:439–49. doi:10.1016/j.ijhydene.2010.10.045.
- 668 [47] Adav SS, Lee DJ, Wang A, Ren N. Functional consortium for hydrogen production
669 from cellobiose: Concentration-to-extinction approach. *Bioresour Technol*
670 2009;100:2546–50. doi:10.1016/j.biortech.2008.12.014.
- 671 [48] Sivagurunathan P, Sen B, Lin CY. Batch fermentative hydrogen production by
672 enriched mixed culture: Combination strategy and their microbial composition. *J*
673 *Biosci Bioeng* 2014;117:222–8. doi:10.1016/j.jbiosc.2013.07.015.
- 674 [49] Van Niel EWJ, Claassen P, Stams A. Substrate and product inhibition of hydrogen
675 production by the extreme thermophile, *Caldicellulosiruptor saccharolyticus*.
676 *Biotechnol Bioeng* 2003;81:255–62. doi:10.1002/bit.10463.
- 677 [50] Szymanowska-Powałowska D. The effect of high concentrations of glycerol on the
678 growth, metabolism and adaptation capacity of *Clostridium butyricum* DSP1.
679 *Electron J Biotechnol* 2015;18:128–33. doi:10.1016/j.ejbt.2015.01.006.
- 680
- 681

Ionic strength



0.09 M

0.75 M

0.81 M



<p>37°C pH : 6 S/X : 10</p>

