



HAL
open science

Characterization of a combined batch-continuous procedure for the culture of anammox biomass

R. Connan, P. Dabert, S. Le Roux, O. Chapleur, G. Bridoux, M. Vanotti, Fabrice Béline, A. Magri

► To cite this version:

R. Connan, P. Dabert, S. Le Roux, O. Chapleur, G. Bridoux, et al.. Characterization of a combined batch-continuous procedure for the culture of anammox biomass. *Ecological Engineering*, 2017, 106, pp.231-241. 10.1016/j.ecoleng.2017.06.003 . hal-02606365

HAL Id: hal-02606365

<https://hal.inrae.fr/hal-02606365v1>

Submitted on 17 May 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

Characterization of a combined batch-continuous procedure for the culture of anammox biomass

Romain Connan^a, Patrick Dabert^a, Sophie Le Roux^a, Olivier Chapleur^b, Gilbert Bridoux^c, Matias B. Vanotti^d, Fabrice Béline^a, Albert Magrí^{a,*}

^a Irstea, UR OPAALE, 17 Avenue de Cucillé-CS 64427, F-35044 Rennes, France

^b Irstea, UR HBAN, 1 rue Pierre-Gilles de Gennes-CS 10030, F-92761 Antony Cedex, France

^c SAUR, Recherche et développement, 2 rue de la Bresle, F-78201 Maurepas, France

^d USDA-ARS Coastal Plains Soil, Water and Plant Research Center, 2611 W. Lucas St., Florence, SC 29501, USA

Interest in autotrophic nitrogen (N) removal through anaerobic ammonium oxidation (anammox) is high in the field of wastewater treatment as a more economic and sustainable alternative than conventional nitrification-denitrification. However, anammox biomass is difficult to enrich, and this can hinder the start-up of new applications. We carried out experimental work to characterize a combined batch-continuous procedure for the enrichment and culture of anammox biomass. In the first stage (time span: 120 d), the enrichment was started in batch mode (sealed vial) using suspended activated sludge as inoculum. Anammox activity was clearly developed since the specific ammonium (NH_4^+) conversion rate increased from 0 to $118 \pm 1 \text{ mg NH}_4^+ \text{-N}/(\text{g VS d})$ (VS, volatile solids); i.e., $560 \pm 11 \text{ mg N}/(\text{L d})$ in terms of N-conversion rate (NCR). Subsequently, the sludge was transferred into a continuous upflow reactor packed with a polyester non-woven material to promote the attached growth of the biomass. Such bioreactor was operated without interruption during 400 d. Under an appropriate feeding regime, the anammox activity increased fast, and a sustained NCR of $1183 \pm 100 \text{ mg N}/(\text{L d})$ was reached according to the N-loading rate applied. Evolution of the microbial community structure was characterized using high-throughput DNA sequencing. The overall procedure prompted the selection of a community enriched in the anammox bacterial species *Candidatus Brocadia sinica* (up to ~70% of the total DNA sequences). Other coexisting microbial groups belonged to *Rhodocyclaceae* (class β -Proteobacteria), *Anaerolineae* (phylum *Chloroflexi*) and *Ignavibacteriaceae* (phylum *Chlorobi*).

1. Introduction

The anaerobic ammonium oxidation (anammox) is an interesting bioprocess for the removal of nitrogen (N) from wastewaters

Abbreviations: anammox, anaerobic ammonium oxidation; DO, dissolved oxygen; HRT, hydraulic residence time; N, nitrogen; NCE, nitrogen conversion efficiency; NCR, nitrogen conversion rate; NLR, nitrogen loading rate; NMDS, non-metric multidimensional scaling; OTU, operational taxonomic unit; PCR, polymerase chain reaction; SBR, sequencing batch reactor; VS, volatile solids; WWTP, wastewater treatment plant.

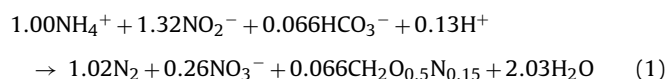
* Corresponding author at: Institut national de Recherche en Sciences et Technologies pour l'Environnement et l'Agriculture (Irstea), 17 Avenue de Cucillé - CS 64427, F-35044 Rennes, France.

E-mail addresses: connan.romain@gmail.com (R. Connan), patrick.dabert@irstea.fr (P. Dabert), sophie.leroux@irstea.fr (S. Le Roux), olivier.chapleur@irstea.fr (O. Chapleur), gbridoux@saur.fr (G. Bridoux), matias.vanotti@ars.usda.gov (M.B. Vanotti), fabrice.beline@irstea.fr (F. Béline), albert.magri@gmail.com (A. Magrí).

since it allows more economic and sustainable treatment than conventional approaches based on heterotrophic denitrification (Ma et al., 2016; Siegrist et al., 2008). In engineered systems, its coupling with partial nitritation results in a complete autotrophic deammonification, which reduces by 60% the oxygen requirement, 100% the organic carbon requirement, and 90% the biosolids production with respect to classical nitrification-denitrification. In addition, it offers the chance of working with more compact reactors at higher loading rates (Magrí et al., 2013; Van Hulle et al., 2010).

The anammox process is mediated by chemolithoautotrophic bacteria that oxidize ammonium (NH_4^+) into dinitrogen gas (N_2) using nitrite (NO_2^-) as the electron acceptor (Strous et al., 1998). Nitric oxide (NO) and hydrazine (N_2H_4) are two known intermediates of such reaction (Kartal et al., 2011). This bioconversion takes place under absence of oxygen and presence of inorganic carbon. A small amount of nitrate (NO_3^-) is also produced due to the oxidation of nitrite linked to the fixation of inorganic carbon in anabolism (de Almeida et al., 2011). According to the full stoichiometry pro-

posed by Strous et al. (1998) –Eq. (1)–, ammonium and nitrite are converted into dinitrogen gas and nitrate under the molar ratios 1.00/1.32/1.02/0.26 for NH_4^+ consumption, NO_2^- consumption, N_2 production, and NO_3^- production, respectively –values obtained through mass balance in a sequencing batch reactor (SBR) running under stable conditions–.



To date, six “Candidatus” anammox bacterial genera have been enriched from samples collected in wastewater treatment facilities and natural environments such as freshwater and marine zones; i.e., *Ca. Brocadia* (Strous et al., 1999), *Ca. Kuenenia* (Schmid et al., 2000), *Ca. Scalindua* (Kuypers et al., 2003), *Ca. Anammoxoglobus* (Kartal et al., 2007), *Ca. Jettenia* (Quan et al., 2008), and *Ca. Anammoximicrobium* (Khramenkov et al., 2013). All these genera belong to the same phylum *Planctomycetes*. In spite of this fact, while the first five aforementioned genera form a deeply branched monophyletic group (family *Brocadiaaceae*), the sixth is closely related to the genus *Pirellula* (family *Planctomycetaceae*). In physiological terms, the anammox bacteria feature a specific cytoplasmic membrane-bound organelle known as anammoxosome, which is the locus of the anammox catabolism (van Niftrik and Jetten, 2012). They are also characterized by a low growth rate, with doubling times (at $\sim 30^\circ\text{C}$) of 2.1–11 days (Lotti et al., 2015; Strous et al., 1998). Owing to this slow biomass development and the specialized metabolism, the anammox bacteria may be difficult to culture.

The anammox bacteria have not been isolated in pure culture yet. Otherwise, such microorganisms have been enriched from various environments up to a culture purity degree of about 80–95% (van Niftrik and Jetten, 2012) –maximum value found in the literature is $98 \pm 1\%$ in a suspended cell anammox culture (Lotti et al., 2014)–. Frequently, those strategies used to enrich anammox biomass consist on the utilization of different types of continuously operated reactors such as the SBR, rotating biological contactor, membrane bioreactor, upflow anaerobic sludge blanket reactor, upflow fixed-bed biofilm reactor, or stirred-tank moving-packed-bed reactor, among others (Egli et al., 2003; Strous et al., 1998; Tsushima et al., 2007; Wang et al., 2009; Xiong et al., 2013; Bae et al., 2015). Alternatively, anammox enrichments have also been performed in batch mode using sealed vials (Bae et al., 2010; Connan et al., 2016; Yasuda et al., 2011). In all cases, appropriate selection of the environmental conditions such as temperature, pH, and levels of ammonium, nitrite, organic carbon, dissolved oxygen (DO), and other nutrients and inhibitors, is critical for a successful enrichment and mass culture (Carvajal-Arroyo et al., 2013). The enrichment of anaerobic ammonium-oxidizing biomass from conventional sludge is time-consuming and may take from several months to years depending on the seeding source, reactor setup, and operational conditions applied (Ibrahim et al., 2016). Thus, biomass enrichment usually is the critical step for the start-up of new anammox applications (especially when pre-enriched sludge is not available).

The objective of this study is to characterize a culture of anammox biomass obtained from activated sludge using a combination of batch and continuous procedures. The enrichment was started under suspended biomass batch mode (sealed vial) and subsequently continued using a continuous upflow reactor packed with a polyester non-woven material to promote the attached growth of the biomass. The evolution of the microbial community structure was characterized throughout the process by means of 16S rRNA gene high-throughput sequencing.

2. Material and methods

2.1. Biomass sources: collection and pretreatment

Activated sludge collected in a municipal wastewater treatment plant (WWTP) that combine the use of a Modified Ludzack-Ettinger bioreactor unit and a membrane filtration loop to perform N-removal was used as inoculum for the enrichment of anammox biomass. Such treatment facility is located in Betton (France). Before starting with the anammox enrichment procedure, denitrification was favored during the first days after sampling in order to promote biodegradation of residual organic matter. Such pretreatment was carried out at room temperature by adding a nitrate source (KNO_3) in pulses equivalent to 100 mg N/L, similarly to Casagrande et al. (2011), and controlling the pH within the range 7.0–8.0 (HCl). The anammox batch enrichment was started after 4 weeks, once denitrification activity decreased. For microbial characterisation purposes, an alternative anammox biomass sample was obtained from a lab-scale 10 L jacketed upflow fixed-bed biofilm reactor running at the USDA-ARS laboratory in Florence, South Carolina, USA (Vanotti et al., 2011). At the time of sludge collection, the reactor was fed with mineral medium containing 153 mg NH_4^+ -N/L and 153 mg NO_2^- -N/L, and was operated with a hydraulic residence time (HRT) of 4 h, N-loading rate (NLR) of about 1800 mg N/(Ld) and water temperature of 30°C , analogously as detailed by Magrí et al. (2012b). Once received at the Irstea laboratory in Rennes, the external sample (E) was stored for 2 years under freezing conditions before proceeding with DNA extraction.

2.2. Batch stage

The enrichment in batch mode was performed using a glass vial which contained the inoculum and mineral medium (working volume of 0.5 L). Such vial was sealed with a rubber stopper plus an aluminium cap and placed into an incubator shaker (model KS4000i control, IKA, Germany) at 150 rpm, 35°C , and in dark conditions. Initial solids content inside the vial was adjusted to 1.50 g VS (VS, volatile solids). Biomass settling was allowed weekly to withdraw the supernatant and, subsequently, to refill the vial with new mineral medium (avoiding the eventual accumulation of inhibitory compounds). The mineral medium was initially prepared with low amount of nitrite and ammonium (25 mg NO_2^- -N/L + 25 mg NH_4^+ -N/L). Once the anammox activity was detected, a second weekly addition of nitrogenous substrates was performed targeting a progressive increase in the concentration of nitrite from 25 to 150 mg NO_2^- -N/L (ammonium was added at a ratio of 1.2 g NO_2^- -N/g NH_4^+ -N). The pH within the vial was controlled in the range from 7.0 to 8.0 (HCl 2 M). N_2 flushing was used to displace air in the vial headspace every time it was opened. Liquid samples were taken before and after each feeding event and filtered using $0.45 \mu\text{m}$ polypropylene membrane filters. A biological sample was taken once per month (B0-B4), centrifuged at 10,000g for 4 min, and the pellet was stored at -20°C (after discarding the supernatant). This enrichment step lasted 120 days (4 months), and final anammox activity was evaluated in a batch test (in duplicates), as described elsewhere (Connan et al., 2016).

2.3. Continuous stage

The biomass enriched following the aforementioned method was seeded in a continuous upflow column reactor (0.94 g VS). This was a jacketed cylindrical reactor made of glass (Trallero & Schlee, Spain) with inner diameter of 9 cm and column height (to the effluent port) of 66 cm. Similarly to other works (Furukawa et al., 2003; Vanotti et al., 2011), a support made of polyester non-woven material coated with pyridinium-type polymer (Japan Vilene Co., Japan)

was placed inside the column reactor to enhance the retention of the biomass (8 strips each one 4.5 cm wide and 57 cm long). Total liquid volume was 4.5 L whereas volume of the reaction zone (excluding the upper 9 cm zone without support) was 3.9 L. Process temperature was controlled at 35 °C using a water heating circulator (model EH-13, Julabo, Germany). Mineral medium was continuously pumped inside the reactor by the bottom-end through a low-flow peristaltic pump (model PD5001, Heidolph, Germany) equipped with a multichannel head (model C4, Heidolph), and treated liquid was discharged near the top of the reactor after passing through the matrix of immobilized biomass. Targeted NLRs ranged from 111 to 1551 mg N/(Ld) according to the nominal volumetric flow rate (3–12 mL/min) and influent concentration (50–175 mg NO₂⁻-N/L at a ratio of 1 g NO₂⁻-N/g NH₄⁺-N). Corresponding HRTs (taking into account the reaction volume) were 21.7–5.4 h. The mineral medium was stored at room temperature in a polyethylene tank sealed at the top to prevent air from entering (a Tedlar gas sampling bag filled with N₂ was connected with tubing to the top of the tank). Such influent tank was refilled weekly with fresh mineral medium. Tygon tubing was used for the setup. The reactor was placed in a dark chamber to avoid light. Liquid samples were regularly taken from the influent and the effluent lines for chemical analysis. Volumetric flow rate was measured by collecting the liquid in a graduated cylinder during a known period of time. When required, gas samples were taken from the headspace of the reactor for analysis. Off-gas flow rate in the effluent line was measured using the water displacement method. Finally, biological samples from the reactor –considering bottom (“B”) and top (“T”) ends of the fabric material– were taken during the experimental period at time 0, 3.5, 6.5, and 13.5 months (C0–C3X, where X corresponds to the letter “B” or “T”), and processed as aforementioned in Section 2.2. The upflow reactor was uninterruptedly operated during 400 days (13.5 months).

2.4. Mineral medium

The synthetic nutritive solution used throughout the experiments was prepared with tap water according to a modification of the mineral medium described by Magrí et al. (2012a); i.e., NH₄Cl (variable: 95–669 mg/L), NaNO₂ (variable: 123–862 mg/L), KNO₃ (variable: 0–361 mg/L), KHCO₃ (variable: 0–1000 mg/L), NaHCO₃ (variable: 0–1000 mg/L), KH₂PO₄ (variable: 0–27 mg/L), FeSO₄·7H₂O (9 mg/L), EDTA (5 mg/L), MgSO₄·7H₂O (240 mg/L), CaCl₂·2H₂O (143 mg/L), and trace element solution 0.3 mL/L. The trace element solution contained ZnSO₄·7H₂O (1247 mg/L), MnSO₄·H₂O (1119 mg/L), CuSO₄·5H₂O (44 mg/L), Al₂(SO₄)₃·14H₂O (201.5 mg/L), Na₂MoO₄·2H₂O (129 mg/L), CoCl₂·6H₂O (30 mg/L), KCl (100 mg/L), and EDTA (975 mg/L). KNO₃ was added to the mineral medium throughout the batch enrichment period in order to help maintaining anoxic conditions in case of total nitrite consumption (preventing sulfate reduction). Unfortunately, because of a mislabeling by the chemical supplier (K₂O was supplied instead of KH₂PO₄) any phosphorus source was added to the mineral medium used during the batch enrichment and also during the first 90 days of operation of the upflow reactor. In this regard, during batch culture, phosphorus released by the decaying biomass seemed to be sufficient to avoid limitation in the availability of this nutrient. Conversely, this was not the case when operating the upflow reactor as it will be discussed later in Section 3.2.1. Once identified after analysis, the aforementioned chemical was replaced by a real source of KH₂PO₄. Finally, the liquid flow regime in the upflow reactor was assessed by punctually replacing the bicarbonate source in the mineral medium (KHCO₃ and NaHCO₃ were interchanged) and subsequently monitoring the evolution of the concentration of the potassium (K⁺) and sodium (Na⁺) ions in the effluent line. Once dis-

solved all the mineral salts, the DO was purged by bubbling with N₂ (<0.2 mg O₂/L) and the pH was adjusted to 6.8–7.0 (HCl).

2.5. Chemical analysis

NH₄⁺, NO₂⁻, NO₃⁻, K⁺, Na⁺, and phosphate (PO₄) were measured by ion chromatography (model 850 Professional IC, Metrohm, Switzerland). VS were measured after sample drying to constant weight at 105 °C and further ignition in a muffle furnace at 550 °C. The pH and DO were measured using portable meters pH 197i and Oxi 197 (WTW, Germany), respectively. Nitrous oxide (N₂O) in gaseous samples was measured by gas chromatography (model 6890N, Agilent Technologies, USA).

2.6. N-transformation calculations

Both, the N-conversion rate (NCR) and the N-conversion efficiency (NCE) were defined according to the removal of ammonium and nitrite from the liquid. During batch enrichment, the NCR was calculated from the corresponding time-dependent slopes for the evolution of the concentrations (obtained through linear regression analysis). Specific activity (specific-NCR) was determined taking into account the VS content in the vial. Under continuous operation of the upflow reactor, the NCR was calculated from the difference in concentrations between the influent and the effluent, divided by the measured HRT. Similarly, the NCE was calculated from the difference in concentrations between the influent and the effluent, divided by the total concentration in the influent. On the other hand, reaction molar ratios were calculated according to the difference in concentrations of ammonium, nitrite, and nitrate between influent and effluent and expressed per unit of ammonium removal. The dinitrogen gas reaction ratio was calculated through mass balance (1 mol N₂ = 2 atoms N) considering the other three measured N-species.

2.7. Microbial community analysis

2.7.1. DNA extraction

Total DNA was extracted from approximately 0.25 g of pellet with the PowerSoil™ DNA Isolation Kit (MoBio Laboratories Inc., USA), according to the manufacturer's instructions. The concentration and purity of the extracted DNA were checked spectrophotometrically (model ND-1000, NanoDrop Technologies, USA) and in TAE 1X – 0.7% agarose gel. The extracted DNA was stored at –20 °C until further analysis.

2.7.2. High-throughput DNA sequencing

16S rRNA genes high-throughput DNA sequencing was performed by the BIOMIC team of Irstea (Antony, France) using an Ion PGM™ (Life Technologies, USA) platform, as described by Poirier et al. (2016a, 2016b). Briefly, the bacterial and archaeal hypervariable region V4–V5 of the 16S rRNA gene was amplified using the primer 515F (5'-GTGYCAGCMGCCGCGTA-3') and a modified version of the primer 928R (Wang and Qian, 2009) that was named 928Ramx (5'-CCCCGYCAATTCTTTTRAGT-3'). The primer 928Ramx includes the nucleotide H (T, A or C) instead of the nucleotide M (A or C) in position 13, which does not change its universality but increases its similarity with anammox sequences. Amplification was performed in a 50 µL reaction mixture using from 10 to 200 pg of extracted DNA and the Pfx SuperMix protocol from Life Technologies, as described in Poirier et al. (2016b). Further processing of the polymerase chain reaction (PCR) products –purification, quantification, emulsion PCR, and sequencing on an Ion 316™ chip v2 using the Ion PGM™ System (Life Technologies)– was carried out according to the manufacturer's instructions, as described in Connan et al. (2016). After sequencing, the PGM™ software filtered out low

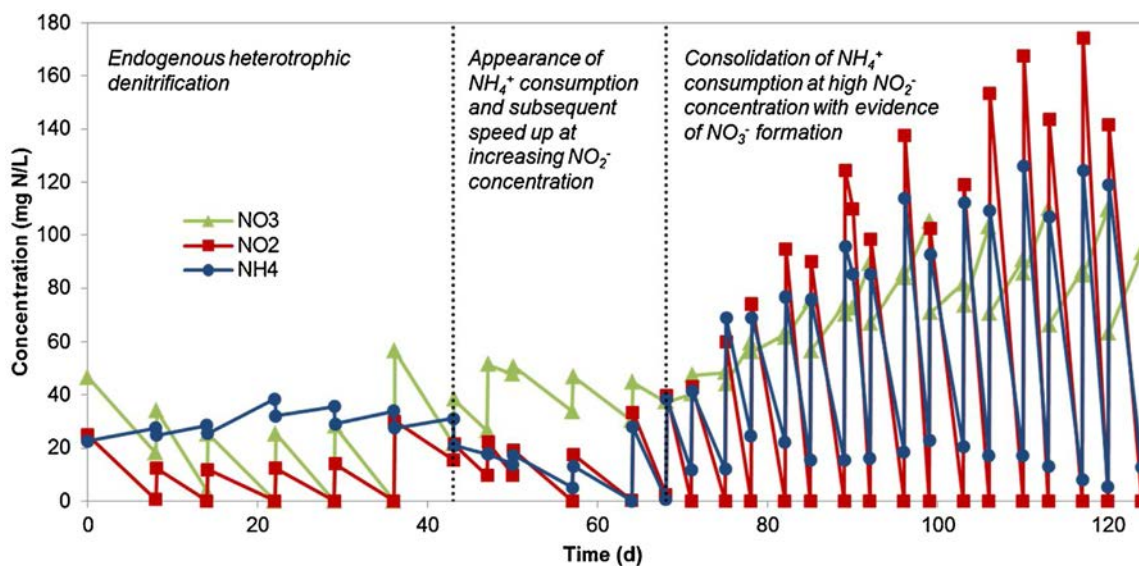


Fig. 1. Evolution of the N-species during the batch enrichment of anammox biomass (4 months).

quality and polyclonal sequence reads. Filtered sequences were analyzed using the QIIME pipeline (v1.8.0) (Caporaso et al., 2010). Sequences shorter than 200 bp or longer than 250 bp, chimeras, and singletons were removed from the dataset. Operational taxonomic units (OTUs) were subsequently defined using UPARSE implemented in USEARCH (v8.0.1623) (Edgar, 2013) at a 97% similarity level. MOTHUR (v1.25.0) (Schloss et al., 2009) and SILVA (v119) (Quast et al., 2013) were used as the classifier tool and database for taxonomic association (with a minimum similarity threshold of 80%), respectively.

2.7.3. Statistical analysis

The statistical analysis to assess the evolution of the microbial community structure was carried out through the non-metric multidimensional scaling (NMDS) method using the open-source software R (v3.2.3) (Venables et al., 2016) including the vegan package (v2.3-2) (Oksanen et al., 2016). The diversity indices Shannon-Weaver and Simpson, as well as the number of OTUs and equitability, were calculated for all samples.

3. Results and discussion

3.1. Batch stage

The batch enrichment developed significant anammox activity after 4 months (Fig. 1), which was measured, in terms of NCR, as 560 ± 11 mg N/(Ld) (ammonium conversion rate of 222 ± 2 mg $\text{NH}_4^+\text{-N}/(\text{Ld})$ at a $\text{NO}_2^-/\text{NH}_4^+$ reaction molar ratio of 1.53 ± 0.03) (Fig. 2). If referred to the VS content, this is equivalent to a specific-NCR of 297 ± 6 mg N/(g VSd). At the end of the enrichment, the biomass still maintained the aspect of activated sludge and brownish colour, but tiny red granules could already be identified in the liquid bulk. Three main phases were observed throughout the enrichment period (Fig. 1): (Phase I) endogenous heterotrophic denitrification was the dominant process and ammonium may even slightly increase during incubation due to the hydrolysis of the remaining organic matter, (Phase II) appearance of ammonium consumption (detected after 43 days) and subsequent speed up at increasing concentrations of nitrite, and (Phase III) consolidation of ammonium consumption at high concentration of nitrite (attaining levels of $150 \text{ mg NO}_2^- \text{-N/L}$) under $\text{NO}_2^-/\text{NH}_4^+$ reaction molar ratios approaching the value of 1.32 given in Eq. (1), and with evi-

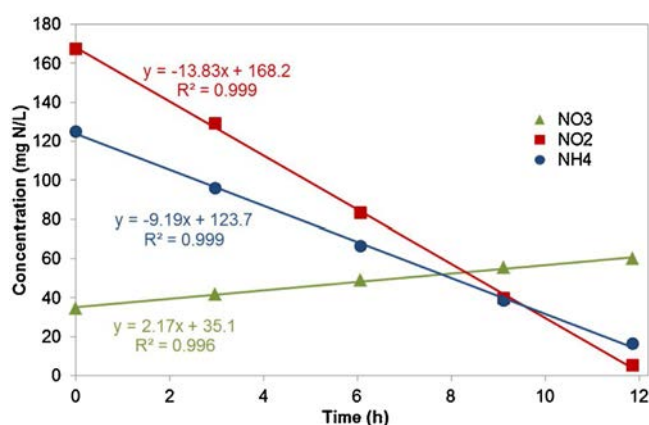


Fig. 2. Batch test carried out to assess the anammox activity after 4 months of enrichment.

dence of nitrate production. As previously reported (Connan et al., 2016; Uyanik et al., 2011), the nitrite supply strategy applied when targeting anammox biomass enrichment is of utmost importance. In this study, nitrite levels of $150 \text{ mg NO}_2^- \text{-N/L}$ were feasible without inhibition (this was corroborated by the linear evolution of the N-forms observed in the final activity test, which was launched at nitrite concentrations slightly higher than $150 \text{ mg NO}_2^- \text{-N/L}$; Fig. 2) although the enrichment was started using low concentrations of nitrite.

3.2. Continuous stage

3.2.1. N-removal performance

The suspended biomass with anaerobic ammonium-oxidizing activity was transferred into the upflow reactor. According to the corresponding reaction volumes, such handling implied an initial biomass dilution at a rate of about 1/8. The upflow reactor was subsequently operated without interruption for a period of 400 days (Figs. 3 and 4, and Table 1). Two main phases can be identified throughout this long experimental period according to the availability of orthophosphate phosphorus in the mineral medium used for feeding the reactor (Section 2.4).

Phase I (90 days). The NLR was progressively increased (Fig. 3A) targeting from 111 to $332 \text{ mg N}/(\text{Ld})$ according to the nominal volu-

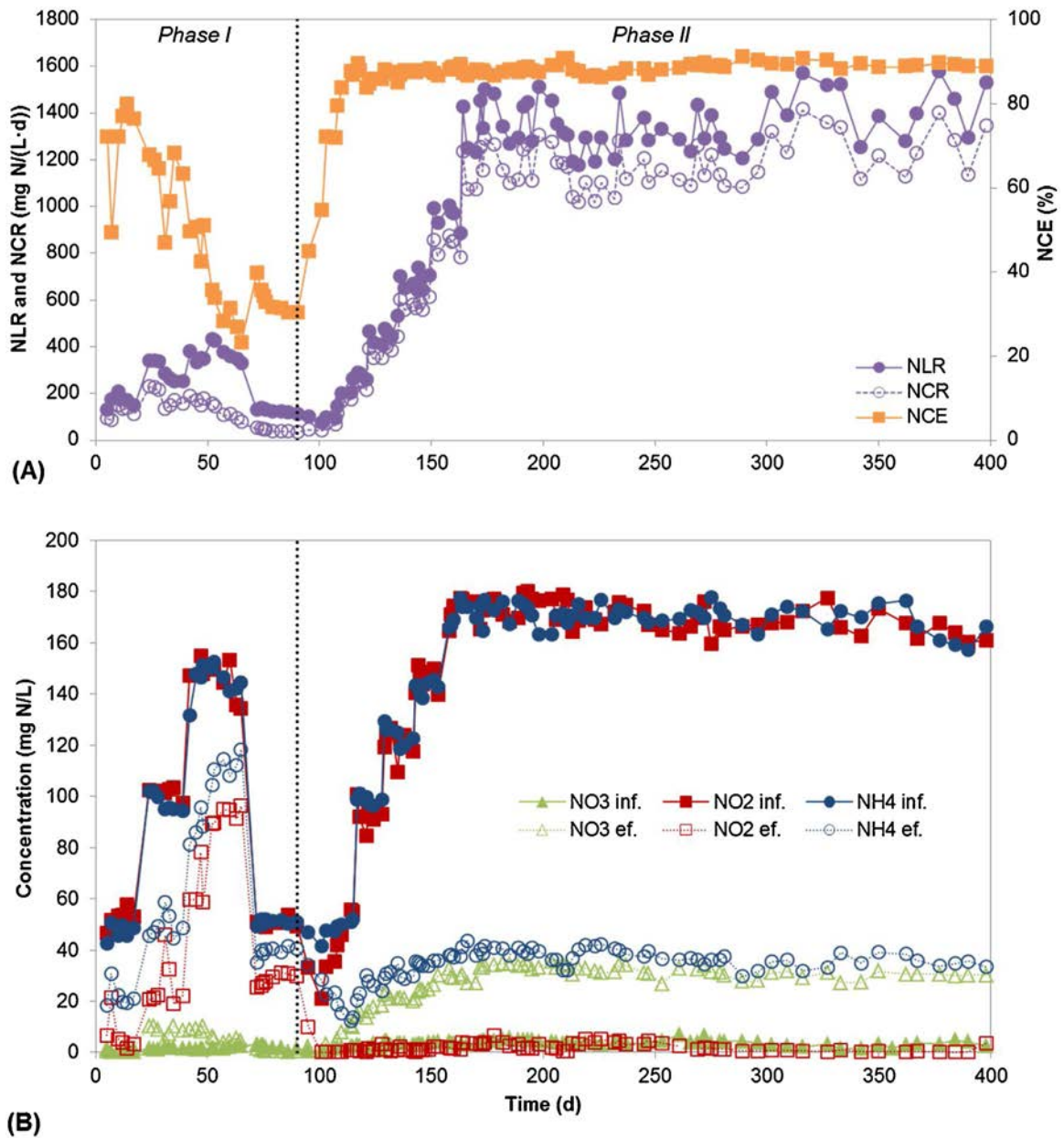


Fig. 3. Time course of the N-removal performance in the continuous upflow reactor. (A) N-loading rate (NLR), N-conversion rate (NCR), and N-conversion efficiency (NCE). (B) Influent (inf.) and effluent (ef.) ammonium, nitrite, and nitrate concentrations.

Table 1

Summary of the operating conditions applied and performance for the upflow reactor.

Parameter ^a	Phase I (90 d)	Phase II (310 d)	
	–	73 d	237 d
HRT (h)	18.1 (2.6) ^b	22.4–7.2	6.1 (0.5)
NLR (mg N/(L·d))	110–431	73–1002	1353 (109)
NCR (mg N/(L·d))	33–227	39–870	1183 (100)
NCE (%)	23–80	45–89	88 (1)
NH ₄ ⁺ inf. (mg N/L)	43–152	42–178	170 (5)
NO ₂ ⁻ inf. (mg N/L)	47–155	21–178	170 (6)
NO ₃ ⁻ inf. (mg N/L)	1 (1)	3 (1)	4 (1)
NH ₄ ⁺ ef. (mg N/L)	18–118	12–38	37 (3)
NO ₂ ⁻ ef. (mg N/L)	2–96	0–10	2 (2)
NO ₃ ⁻ ef. (mg N/L)	0–10	1–33	32 (2)
NO ₂ ⁻ /NH ₄ ⁺ reaction molar ratio	1.70 (0.28)	1.34 (0.12)	1.27 (0.06)
N ₂ /NH ₄ ⁺ reaction molar ratio	1.33 (0.17)	1.08 (0.08)	1.03 (0.02)
NO ₃ ⁻ /NH ₄ ⁺ reaction molar ratio	0.05 (0.10)	0.18 (0.05)	0.21 (0.02)

^a Abbreviations: HRT, hydraulic residence time; NLR, N-loading rate; NCR, N-conversion rate; NCE, N-conversion efficiency; inf., influent; ef., effluent.

^b Parameter values are provided according to the minimum-maximum range or as average (standard deviation in parentheses).

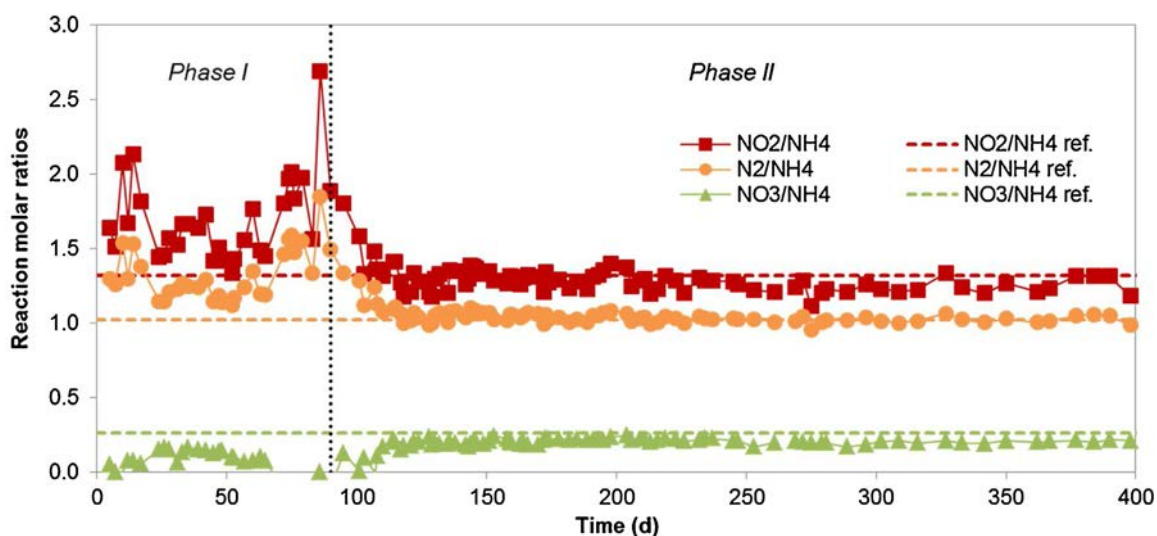


Fig. 4. Time course of the nitrite-to-ammonium, dinitrogen gas-to-ammonium, and nitrate-to-ammonium reaction molar ratios in the continuous upflow reactor. Horizontal dashed lines indicate those reference (ref.) values provided in Eq. (1) (1.32, 1.02, and 0.26, respectively).

metric flow rate (3 mL/min) and influent concentration (50–150 mg NO_2^- -N/L at a ratio of 1 g NO_2^- -N/g NH_4^+ -N). Taking into account the activity of the biomass measured at the end of the batch enrichment and the dilution rate applied, NCRs not lower than 70 mg N/(Ld) were expected. Satisfactory N-removal performance was observed during the first 10 days of operation with low nitrite (limiting substrate) concentration in the effluent (Fig. 3B). However, nitrite and ammonium started to accumulate in the reactor in the following days due to the lack of phosphorus in the feeding stream with the consequent decrease in the NCE. After 70 days of operation the nominal influent concentration of nitrite was returned to 50 mg NO_2^- -N/L but it did not imply the recovery of the system (NCE < 40%). As depicted in Fig. 4, the reaction ratios during this phase did not match well those values given in Eq. (1), with higher values than expected for the $\text{NO}_2^-/\text{NH}_4^+$ and N_2/NH_4^+ reaction molar ratios but with lower values than expected for the $\text{NO}_3^-/\text{NH}_4^+$ reaction molar ratio (which may be explained due to the coexistence of anammox and heterotrophic denitrification processes). Once arrived at this point, the problem with the phosphorus source had already been identified and the corresponding chemical used to prepare the mineral medium was replaced.

Phase II (310 days). In this second phase the NLR was progressively increased again (Fig. 3A). Particularly, the targeted loads ranged initially from 111 to 1034 mg N/(Ld) according to the nominal volumetric flow rate (3–8 mL/min) and influent concentration (50–175 mg NO_2^- -N/L at a ratio of 1 g NO_2^- -N/g NH_4^+ -N). Fast recovery of the N-removal (ammonium and nitrite) performance was observed, resulting in increased NCRs (maximum value of 870 mg N/(Ld)) and NCEs (maximum value of 89%). Nitrite was completely consumed (1 ± 2 mg N/L in the effluent), nitrate was again detected in the outlet (1–33 mg N/L) (Fig. 3B), and the reaction molar ratios evolved towards those given in Eq. (1) (Fig. 4). After 73 days of operation (total time: 163 days) the targeted NLR was further increased up to 1551 mg N/(Ld) according to a nominal volumetric flow rate of 12 mL/min (maximum value planned for this study). Once arrived at this point, it was assumed that the anammox enrichment had been achieved successfully. The biomass growing at the bottom of the reactor and attached to the support material developed red colour. During the following 237 days the reactor run at an average NCR of 1183 ± 100 mg N/(Ld) and NCE of $88 \pm 1\%$ (Table 1) in order to continue the culture of the anammox biomass. During this time, the gas produced within the reactor was repeatedly analyzed aiming to detect N_2O . In this regard, N_2O off-gas

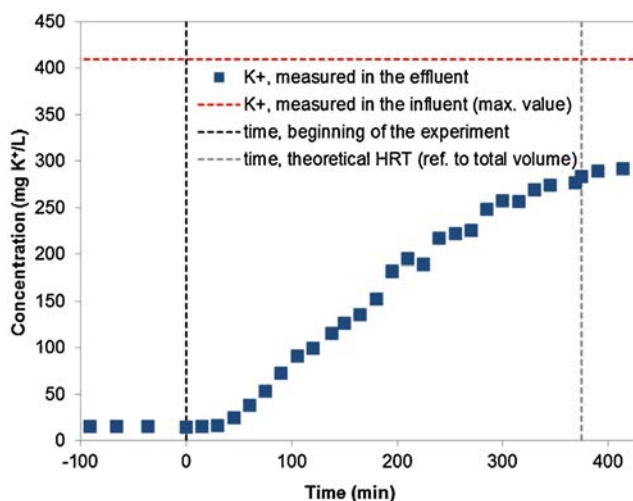


Fig. 5. Assessment of the liquid flow regime in the upflow reactor by introducing a step perturbation into the feed stream entering the reactor (replacement of the bicarbonate source in the mineral medium, from NaHCO_3 to KHCO_3) and subsequent monitoring of the concentration of K^+ in the outlet (day 266; volumetric flow rate of 12 mL/min). HRT, hydraulic residence time (here referred to the total liquid volume).

emissions never accounted for more than 0.2% of the applied N-load, which is quite similar to the values reported by other authors such as Okabe et al. (2011) (0.1% N-load in average) using similar bioreactor technology.

3.2.2. Liquid flow regime assessment

Despite the satisfactory N-removal performance achieved, the existence of the support material, biomass growing, and rising gas bubbles within the reactor did result in a liquid flow regime far from the mixing patterns assumed in an ideal plug-flow reactor without axial mixing (where all the atoms of material leaving the reactor have been inside it for exactly the same amount of time (Fogler, 2006); i.e., the HRT). This was evidenced in a short-term experiment based on the introduction of a step perturbation into the system (Fig. 5); i.e., increase in the influent concentration of K^+ (which does not take part in the anammox reaction). The mineral medium did not flow through the reactor uniformly, as it is shown by the temporal evolution of the concentration of K^+ in the outlet. Thus, at a time smaller than the HRT, the concentration of K^+ in

the effluent started to increase (i.e., early exit of liquid), but at a time equal to the HRT, the concentration of K^+ in the effluent had not reached the inlet levels yet. This fact implies that, there were regions in the fixed-bed that offered little resistance to the flow, and as a result a major portion of the fluid channeled through this pathway. The molecules following this pathway did not spend as much time inside the reactor as those flowing through other regions offering high resistance to the flow.

The aforementioned non-ideal flow pattern did not significantly hinder the conversion of nitrite (limiting substrate in the mineral medium), and very low concentrations were measured in the outlet (~ 2 mg NO_2^- -N/L). Indeed, the NLR applied in this investigation was not pushed to maximum values –e.g., Tsushima et al. (2007) reported N-removal rates as high as 26 g N/(L d) in a similar upflow reactor– since main target was the effective culture of anammox biomass. In addition, uneven biomass accumulation was observed along the fixed-bed. Thus, the biomass especially grew attached at the bottom-end of the fabric material, partially evolving into granular biomass similarly as described elsewhere (Zhang et al., 2015). Such biomass distribution could favor the existence of a gradient for the vertical flow resistance as well as dead zones (in such case implying underestimation of the measured NCR). Alternative shapes and sizes of the non-woven carrier could also be considered for enhancing the culture process (Wang et al., 2016a). Anammox is an autotrophic process and, consequently, produces few amount of biomass. However, and according to this test, under long-term culture periods in upflow fixed-bed biofilm reactors it may still be necessary to periodically extract sludge from inside of the reactor in order to minimize the risk of clogging and to enhance the liquid flow.

3.3. Microbial community characterisation

Changes in the microbial community structure during the enrichment in batch (B) mode, and subsequently in the continuous (C) upflow reactor, were monitored using 16S rRNA high-throughput DNA sequencing. An external (E) sample coming from the upflow fixed-bed biofilm reactor running at the USDA-ARS laboratory (Vanotti et al., 2011) was also analyzed using the same methodology. After quality filtering, the number of sequences per sample ranged from 2648 (sample B3) to 20555 (sample B2) with an average for all the libraries of 11670 ± 5593 sequences per sample (13 samples in total). Coverage of all libraries, calculated after a systematic random depletion conducted to equalize the number of sequences per sample to that of the smallest library, was $\geq 95 \pm 3\%$ (which is high enough to validate the subsequent analysis).

3.3.1. Global evolution of the microbial community

Microbial diversity throughout the entire experimental period (i.e., ~ 18 months) is assessed in Table 2 using the indices Shannon-Weaver, Simpson and number of OTUs. Such diversity indices showed a systematic decrease during both the batch stage (i.e., reductions of 23.3%, 5.1%, and 32.2%, respectively, were confirmed between B0 and B4) and the continuous stage (i.e., index reductions of 29.8–40.4%, 11.1–22.2%, and 34.5–68.3%, respectively, were confirmed between C0 and C3X). Particularly, for the batch stage, a sharp decrease occurred after 2 months of enrichment (e.g., the number of OTUs dropped about 50% between B2 and B3). Indeed, in such time span it was expected an ecological transition linked to the changes in the dominant metabolic pathway; i.e., from denitrification to anammox (Fig. 1). Concerning the continuous stage, the index reductions were more variable, eventually depending on the sampling position within the reactor (e.g., the number of OTUs decreased between 34.5% and 68.3%). According to this consideration, higher index reductions were found at the bottom side (where ammonium and nitrite concentrations were higher). Equitability

also underwent systematic decrease during both stages; i.e., 22.2% and 28.6%, respectively. Such data may imply concomitant disappearance of some species and larger segregation between the low and highly represented taxa (Magurran, 2004; Oksanen, 2016).

Relative evolution of the microbial community structure throughout the experimental period was also assessed using the NMDS method (Fig. 6). A progressive evolution of the microbial communities was observed for both batch and continuous stages. Regarding the batch stage, the dynamic conditions applied during the first 4 months induced a strong evolution of the microbial community, especially between B1 and B3 (transition from a denitrifying to an anammox ecosystem). After transferring the biomass to the continuous stage, the microbial community continued evolving. Initial lack of phosphorus in the feed stream resulted in a smaller relative change of the microbial community structure during the first 3.5 months (C0 vs. C1X) than in the following 3 months (C1X vs. C2X) after restoring the phosphorus supply. Concerning the next 7 months of culture (C2X vs. C3X), the microbial community structure still evolved, but at a lower rate. Fast stabilization of the microbial diversity in anammox reactors was reported by previous studies, even after perturbation episodes in the feed stream (Pereira et al., 2014). The external sample coming from the USDA-ARS anammox reactor (E) was also included in the NMDS analysis. Its microbial community remains apart from the final culture samples. The position of the different samples in the NMDS plot suggests that their vertical distribution is at least partly correlated with microbial diversity.

3.3.2. Description of the microbial community

Because of the large set of data, only OTUs containing more than 1% of the total sequences are considered. The proportion of OTUs excluded from the analysis by choosing this threshold progressively decreased during the experimental period; i.e., from 29% at initial time (B0) to 11% at the end of the batch stage (B4), and to 3% at the end of the continuous stage (C3X). Any of the currently known anammox genus was found within the excluded OTUs. Dominant microbial groups (OTUs with relative abundance $>1\%$ of the total number of sequences) progressively evolved during both batch and continuous stages (Fig. 7). Such operational procedures could affect differently the evolution of the microbial community.

The phylum *Proteobacteria*, which is commonly retrieved in wastewater treatment ecosystems (Bertrand et al., 2011; Hu et al., 2012), was dominant in the activated sludge used as inoculum, but underwent a progressive decline, with a relative abundance decreasing from 36% (B0) to about 10% (C3X). Initially present as α -, β -, and γ -*Proteobacteria* according to 7%, 15%, and 13% of the total sequences, respectively, the experimental conditions applied favored the selection of genera belonging to the family *Rhodocyclaceae* (class β -*proteobacteria*). Thus, at the end of the experimental period, relative abundances for this family ranged from 2% (C3T) to 11% (C3B), while α - and γ -*Proteobacteria* represented less than 1% of the total community. The most frequent genus belonging to *Rhodocyclaceae* was related to the uncultured bacterium clone Dok59 (FJ710778), which has previously been retrieved from a long-term operated anammox biofilm reactor, and it is closely related to the denitrifying bacteria *Denitratisoma oestradiolicum* clone 20b.15 (KF810114, 99% similarity). Such genus was also identified as the dominant proteobacterial genus in the external sample coming from the USDA-ARS anammox reactor (with relative abundance of 9%).

The phyla *Bacteroidetes* (16%), *Chloroflexi* (15%), *Actinobacteria* (9%), *Acidobacteria* (5%), and *Planctomycetes* (5%) were also abundant in the seeding sludge. Regarding *Bacteroidetes*, a progressive reduction in its relative abundance was observed (up to about 2% of the total sequences at the end of the experimental period). It appears that the provided conditions were not favorable for *Bac-*

Table 2
Diversity indices for the microbial community during the batch enrichment (B0-B4) and in the continuous upflow reactor (C0-C3X). Sampling position regarding the carrier within the upflow reactor is identified with the letter "B" for bottom or "T" for top. The external sample coming from USDA-ARS (E) is also included.

Time (months) Sample	Batch stage					Continuous stage							
	0	1	2	3	4	0	3.5		6.5		13.5		USDA-ARS
	B0	B1	B2	B3	B4	C0	C1T	C1B	C2T	C2B	C3T	C3B	E
Shannon-Weaver	7.8	7.6	7.7	5.7	6.0	5.7	4.9	4.6	3.6	4.8	4.0	3.4	5.8
Simpson	0.99	0.99	0.99	0.95	0.94	0.90	0.90	0.90	0.70	0.90	0.80	0.70	0.96
Number of OTUs	552	515	576	271	374	319	192	154	147	207	209	101	231
Equitability	0.9	0.8	0.8	0.7	0.7	0.7	0.6	0.6	0.5	0.6	0.5	0.5	0.7

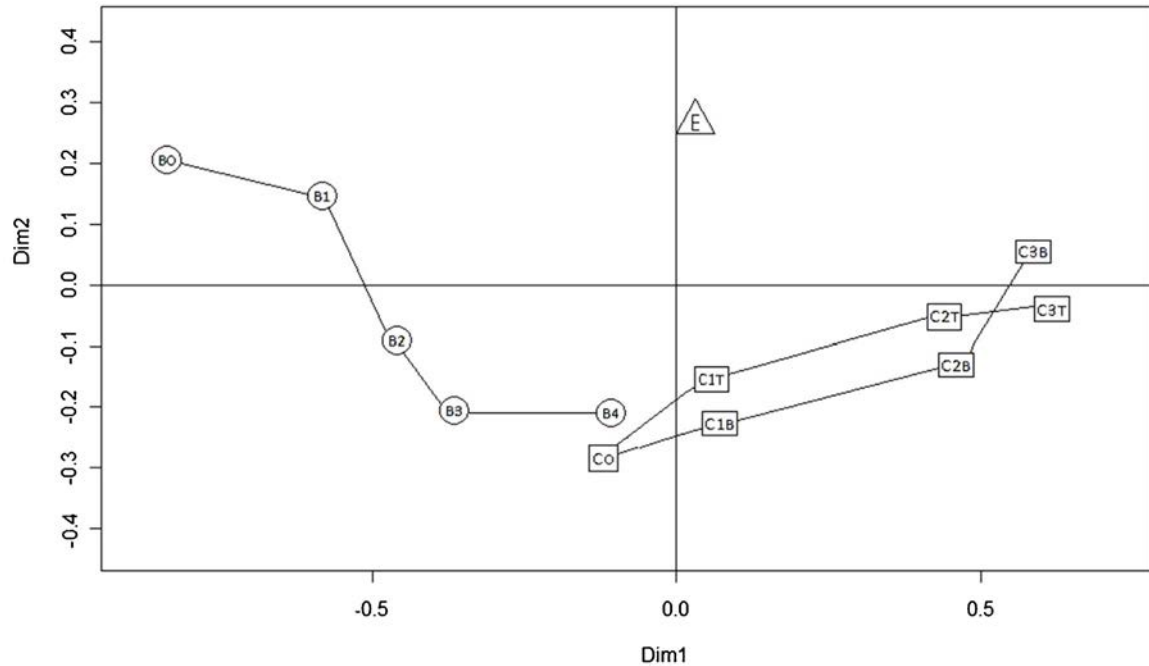


Fig. 6. Non-metric multidimensional scaling (NMDS) plot showing the evolution of the microbial community structure during batch enrichment (circles; B0-B4) and subsequently in the continuous upflow reactor (squares; C0-C3X). Sampling position regarding the carrier is identified with the letter "B" for bottom or "T" for top. For comparison, the external sample coming from USDA-ARS is also included (triangle; E).

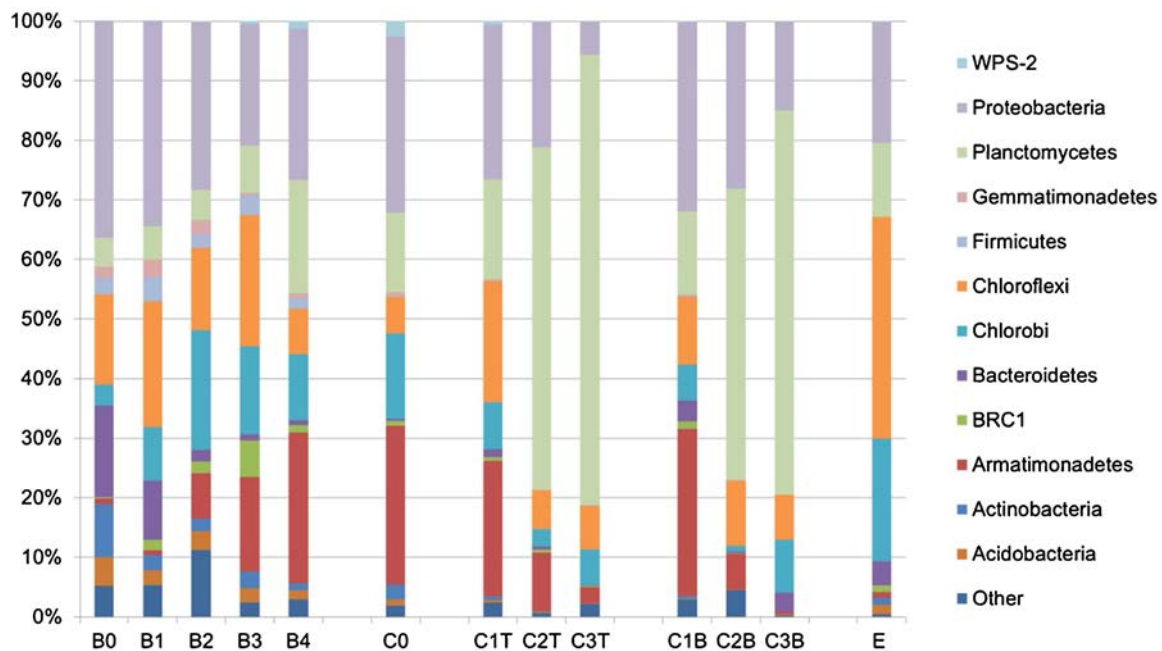


Fig. 7. Changes in the microbial community structure at the phylum level during the enrichment in batch mode (B0-B4) and subsequently in the continuous upflow reactor (C0-C3X). Sampling position regarding the carrier is identified with the letter "B" for bottom or "T" for top. For comparison, the external sample coming from USDA-ARS is also included (E). Results are expressed as relative abundances (% of the total number of sequences).

teroidetes, mainly found in the seeding sludge as bacteria related to the family *Saprosiraceae* (which are known as aerobic and denitrifying chemoorganotrophs). *Chloroflexi*, which is composed of filamentous bacteria frequently present in activated sludge, was almost exclusively represented by genera belonging to the class *Anaerolineae*. The relative abundance of such class decreased from 12% to 7% at the end of the batch enrichment (B4). Similar abundance was found at the end of the continuous culture (C3X). *Chloroflexi* is usually present in anammox reactors and it has been suggested that they can grow using materials derived from decaying anammox bacterial cells (Kindaichi et al., 2012). In the external sample coming from the USDA-ARS anammox reactor such phylum reached a relative abundance as high as 37%. Relative abundances for *Actinobacteria* and *Acidobacteria* declined rapidly during the batch enrichment reaching final values below 2% (B4). Such trend continued during the continuous stage and final percentages below 0.2% (C3X) were obtained. The same behavior was observed for the phyla *Firmicutes* and *Gemmatimonadetes* that were poorly represented in the initial sludge (3% and 2%, respectively, in B0).

Conversely, the batch stage promoted an increase in the relative abundance of *Planctomycetes* from 5% (B0) to 19% (B4) that further pursued in the continuous stage from 13% (C0) to about 70% (C3X). Interestingly, this phylum was initially represented in the seeding sludge by the family *Pirellulaceae*, and anammox-like microorganisms were undetectable during the first 3 months of enrichment. Throughout incubation, the relative abundance of *Pirellulaceae* declined in favor of the anammox family *Brocadia*. The percentages for this last family reached 17% of the total sequences for B4 and 57–75% for C3X. The enriched species was *Ca. Brocadia sinica* which was the only anammox microorganism detected during the whole culture period except in one sample. Those experimental conditions applied for conducting the culture strongly prompted the selection of this species, that was certainly responsible for the observed anammox metabolism, and which has been proposed as an *r*-strategist (Ali and Okabe, 2015; Oshiki et al., 2011). The other aforementioned anammox species was identified as *Ca. B. fulgida*, and it was observed only in the sample C3B (relative abundance only was 0.5% of the total anammox sequences at this time). A much lower relative abundance of anammox-related sequences was identified for the external sample coming from the USDA-ARS anammox reactor (only 12% of the total microbial sequences). Here, the dominant species was *Ca. B. fulgida* that accounted for the 97% of the total anammox sequences, and to a lesser extent *Ca. B. caroliniensis* and *Ca. B. sinica* (accounting for 2% and 1% of the total anammox sequences, respectively).

Concerning other phyla less represented in the seeding sludge, the batch stage promoted *Chlorobi*, with relative abundances rising from 4% to 20% in 2 months, but it was followed by a decrease and stagnation at the end of the first stage and during the continuous stage (8%). Those genera enriched during the batch stage mainly belonged to the photoautotrophic ranks C20 and OPB56 which subsequently disappeared during the continuous stage. Within the upflow reactor, *Chlorobi* was mostly represented by the non-phototrophic chemoheterotrophic family *Ignavibacteriaceae* (6% of the total sequences, 100% similarity with uncultured clone KU000307). It is suggested that bacteria belonging to such family uses organic matter coming from other cells forming the biofilm. This is also supported by its high relative abundance in the USDA-ARS sample (19% of the total sequences). Finally, the phylum *Armatimonadetes* showed a significant enrichment during the batch stage with a relative abundance increasing from 1% (B0) to 25% (B4). Conversely, the continuous stage promoted its disappearance with relative abundances decreasing from 27% (C0) to 2% (C3X). Recently identified organisms related to *Armatimonadetes* have been found in different natural environments and freshwater sediments (Tamaki et al., 2011; Yin et al., 2015).

The highly specific conditions applied to the biomass during this study (i.e., mild temperature, lack of oxygen, absence of organic compounds in the feeding, simultaneous exposure to ammonium and nitrite, and prevention to light exposure, among others) represent a significant shift in comparison to the environmental conditions existing in the inoculum's sampling site (municipal WWTP), and governed the evolution and selection of the microbial community. As a consequence, the microbial community of the seeding activated sludge underwent strong composition rearrangement. Initially primarily composed by *Proteobacteria*, *Bacteroidetes*, *Chloroflexi*, and *Actinobacteria*, with only 5% of *Planctomycetes* and any detectable anammox microbial group, it resulted in a biomass culture with ~70% relative abundance of the anammox species *Ca. B. sinica*, as well as up to 2–11% of *Rhodocyclaceae* (class β -*Proteobacteria*), 7–8% of *Chloroflexi* (dominated by family *Anaerolineae*), and 6–9% of *Chlorobi* (dominated by family *Ignavibacteriaceae*). According to these findings, the particular conditions applied in our system favored the selection of not only anammox bacteria but also of other bacterial groups without anammox activity that could be involved either in competitive or symbiotic microbial relationships (Connan et al., 2016; Wang et al., 2016b). Such final microbial community composition is somehow different from the microbial community of the sample from the USDA-ARS anammox reactor which was primarily composed by *Chloroflexi* (37%), *Chlorobi* (21%), *Proteobacteria* (20%), and *Planctomycetes* (12%). The same families *Rhodocyclaceae* and *Ignavibacteriaceae* but different anammox *Ca. Brocadia* species were identified.

Indeed, high selective pressure through restrictive environmental trophic conditions has been widely reported to decrease microbial diversity and to favor the appearance of dominant species fitting the metabolic requirements, as expected in this study for the anammox bacteria (Connan et al., 2016; Costa et al., 2014; Isanta et al., 2015). On the other hand, when comparing the diversity indices for the anammox biomass here produced with those of the anammox biomass coming from the USDA-ARS anammox reactor, higher values were clearly found for the external sample (Table 2) although both biomasses were obtained from a continuous upflow reactor running during a long period (>1 year) under very similar stable conditions. In this regard, some authors (Costa et al., 2014; Date et al., 2009) have reported that despite the long-term application of similar operational conditions in anammox reactors, differences may still exist in the microbial communities according to the biomass seeding source. The particular procedure followed for initial biomass enrichment, which was different in both cases (Connan et al., 2016; Vanotti et al., 2011), may also impact on the microbial community. The hydraulic performance of both reactors (affecting substrate concentration and biomass gradients within the reactor) and also the different conditions applied for storing the DNA samples are other factors that may play a role in such results.

4. Conclusions

A combined batch-continuous procedure for the mass culture of anammox biomass was described in detail. In the first stage, successful batch enrichment of anammox bacteria from activated sludge was achieved under a progressively increasing nitrite load (i.e., final measured NCR was 560 ± 11 mgN/(Ld) after 120 days). Subsequently, and under an appropriate feeding regime, biomass culture was accomplished in the second stage consisting on a continuous upflow reactor packed with a fabric material to promote the attached growth of the biomass. A sustained NCR of 1183 ± 100 mgN/(Ld) was reached according to the NLR applied. The upflow reactor was operated without interruption during

400 days. The overall procedure resulted in the selection of a microbial community with up to ~70% of the anammox species *Ca. B. sinica*. Other significant coexisting microbial groups belonged to the family *Rhodocyclaceae* (class β -*Proteobacteria*) and the phyla *Chloroflexi* and *Chlorobi*.

Conflict of interest

The authors have declared no conflict of interest.

Acknowledgements

This work was supported by Rennes Métropole [Allocation d'Installation Scientifique (AIS) ref. 14C0509]. The authors are grateful to Chrystelle Bureau for technical assistance with microbial analyses and to the master's student Marina Moya-Espinosa for help with the operation of the upflow reactor. The external USDA-ARS sample was provided to Irstea through the Material Transfer Agreement (MTA) no. 10598.

References

- Ali, M., Okabe, S., 2015. Anammox-based technologies for nitrogen removal: advances in process start-up and remaining issues. *Chemosphere* 141, 144–153, <http://dx.doi.org/10.1016/j.chemosphere.2015.06.094>.
- Bae, H., Park, K.-S., Chung, Y.-C., Jung, J.-Y., 2010. Distribution of anammox bacteria in domestic WWTPs and their enrichments evaluated by real-time quantitative PCR. *Process Biochem.* 45, 323–334, <http://dx.doi.org/10.1016/j.procbio.2009.10.004>.
- Bae, H., Choi, M., Lee, C., Chung, Y.-C., Yoo, Y.J., Lee, S., 2015. Enrichment of ANAMMOX bacteria from conventional activated sludge entrapped in poly(vinyl alcohol)/sodium alginate gel. *Chem. Eng. J.* 281, 531–540, <http://dx.doi.org/10.1016/j.cej.2015.06.111>.
- Bertrand, J.-C., Caumette, P., Lebaron, P., Matheron, R., Normand, P., 2011. *Écologie Microbienne: Microbiologie des Milieux Naturels et Anthropisés*. Presses universitaires de Pau et des Pays de l'Adour, Pau, France (In French).
- Caporaso, J.G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F.D., Costello, E.K., Fierer, N., Gonzalez Peña, A., Goodrich, J.K., Gordon, J.I., Huttley, G.A., Kelley, S.T., Knights, D., Koenig, J.E., Ley, R.E., Lozupone, C.A., McDonald, D., Muegge, B.D., Pirrung, M., Reeder, J., Sevinsky, J.R., Turnbaugh, P.J., Walters, W.A., Widmann, J., Yatsunenko, T., Zaneveld, J., Knight, R., 2010. QIIME allows analysis of high-throughput community sequencing data. *Nat. Methods* 7, 335–336, <http://dx.doi.org/10.1038/nmeth.f303>.
- Carvajal-Arroyo, J.M., Sun, W., Sierra-Alvarez, R., Field, J.A., 2013. Inhibition of anaerobic ammonium oxidizing (anammox) enrichment cultures by substrates, metabolites and common wastewater constituents. *Chemosphere* 91, 22–27, <http://dx.doi.org/10.1016/j.chemosphere.2012.11.025>.
- Casagrande, C.G., Kunz, A., Soares, H.M., de Prá, M.C., Neto, G.F.S., 2011. Establishment of anammox process in sludge samples collected from swine wastewater treatment system. *Eng. Agr. Jaboticabal* 31, 1170–1178, <http://dx.doi.org/10.1590/S0100-69162011000600014>.
- Connan, R., Dabert, P., Khalil, H., Bridoux, G., Béline, F., Magrì, A., 2016. Batch enrichment of anammox bacteria and study of the underlying microbial community dynamics. *Chem. Eng. J.* 297, 217–228, <http://dx.doi.org/10.1016/j.cej.2016.03.154>.
- Costa, M.C.M.S., Carvalho, L., Leal, C.D., Dias, M.F., Martins, K.L., Garcia, G.B., Mancuelo, I.D., Hipólito, T., MacConell, E.F.A., Okada, D., Etchebehere, C., Chernicharo, C.A.L., Araujo, J.C., 2014. Impact of inocula and operating conditions on the microbial community structure of two anammox reactors. *Environ. Technol.* 35, 1811–1822, <http://dx.doi.org/10.1080/09593330.2014.883432>.
- Date, Y., Isaka, K., Ikuta, H., Sumino, T., Kaneko, N., Yoshie, S., Tsuneda, S., Inamori, Y., 2009. Microbial diversity of anammox bacteria enriched from different types of seed sludge in an anaerobic continuous-feeding cultivation reactor. *J. Biosci. Bioeng.* 107, 281–286, <http://dx.doi.org/10.1016/j.jbiosc.2008.11.015>.
- de Almeida, N.M., Maalcke, W.J., Keltjens, J.T., Jetten, M.S.M., Kartal, B., 2011. Proteins and protein complexes involved in the biochemical reactions of anaerobic ammonium-oxidizing bacteria. *Biochem. Soc. Trans.* 39, 303–308, <http://dx.doi.org/10.1042/BST0390303>.
- Edgar, R.C., 2013. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nat. Methods* 10, 996–998, <http://dx.doi.org/10.1038/nmeth.2604>.
- Egli, K., Bosshard, F., Werlen, C., Lais, P., Siegrist, H., Zehnder, A.J.B., van der Meer, J.R., 2003. Microbial composition and structure of a rotating biological contactor biofilm treating ammonium-rich wastewater without organic carbon. *Microb. Ecol.* 45, 419–432, <http://dx.doi.org/10.1007/s00248-002-2037-5>.
- Fogler, H.S., 2006. *Elements of Chemical Reaction Engineering*, fourth ed. Pearson Education Inc., Upper Saddle River, NJ, USA.
- Furukawa, K., Rouse, J.D., Yoshida, N., Hatanaka, H., 2003. Mass cultivation of anaerobic ammonium-oxidizing sludge using a novel nonwoven biomass carrier. *J. Chem. Eng. Jpn.* 36, 1163–1169, <http://dx.doi.org/10.1252/jcej.36.1163>.
- Hu, M., Wang, X., Wen, X., Xia, Y., 2012. Microbial community structures in different wastewater treatment plants as revealed by 454-pyrosequencing analysis. *Bioresour. Technol.* 117, 72–79, <http://dx.doi.org/10.1016/j.biortech.2012.04.061>.
- Ibrahim, M., Yusof, N., Yusoff, M.Z.M., Hassan, M.A., 2016. Enrichment of anaerobic ammonium oxidation (anammox) bacteria for short start-up of the anammox process: a review. *Desalin. Water Treat.* 57, 13958–13978, <http://dx.doi.org/10.1080/19443994.2015.1063009>.
- Isanta, E., Bezerra, T., Fernández, I., Suárez-Ojeda, M.E., Pérez, J., Carrera, J., 2015. Microbial community shifts on an anammox reactor after a temperature shock using 454-pyrosequencing analysis. *Bioresour. Technol.* 181, 207–213, <http://dx.doi.org/10.1016/j.biortech.2015.01.064>.
- Kartal, B., Rattray, J., van Niftrik, L.A., van de Vossenberg, J., Schmid, M.C., Webb, R.I., Schouten, S., Fuerst, J.A., Damsté, J.S., Jetten, M.S.M., Strous, M., 2007. *Candidatus* "Anammoxoglobus propionicus" a new propionate oxidizing species of anaerobic ammonium oxidizing bacteria. *Syst. Appl. Microbiol.* 30, 39–49, <http://dx.doi.org/10.1016/j.syapm.2006.03.004>.
- Kartal, B., Maalcke, W.J., de Almeida, N.M., Cirpus, I., Gloerich, J., Geerts, W., Op den Camp, H.J.M., Harhangi, H.R., Janssen-Megens, E.M., Francoijs, K.-J., Stunnenberg, H.G., Keltjens, J.T., Jetten, M.S.M., Strous, M., 2011. Molecular mechanism of anaerobic ammonium oxidation. *Nature* 479, 127–132, <http://dx.doi.org/10.1038/nature10453>.
- Khranenkov, S.V., Kozlov, M.N., Kevbrina, M.V., Dorofeev, A.G., Kazakova, E.A., Grachev, V.A., Kuznetsov, B.B., Polyakov, D.Y., Nikolaev, Y.A., 2013. A novel bacterium carrying out anaerobic ammonium oxidation in a reactor for biological treatment of the filtrate of wastewater fermented sludge. *Microbiology* 82, 628–636, <http://dx.doi.org/10.1134/S002626171305007X>.
- Kindaichi, T., Yuri, S., Ozaki, N., Ohashi, A., 2012. Ecophysiological role and function of uncultured *Chloroflexi* in an anammox reactor. *Water Sci. Technol.* 66, 2556–2561, <http://dx.doi.org/10.2166/wst.2012.479>.
- Kuyper, M.M.M., Sliemers, A.O., Lavik, G., Schmid, M., Jørgensen, B.B., Kuenen, J.G., Damsté, J.S.S., Strous, M., Jetten, M.S.M., 2003. Anaerobic ammonium oxidation by anammox bacteria in the Black Sea. *Nature* 422, 608–611, <http://dx.doi.org/10.1038/nature01472>.
- Lotti, T., Kleerebezem, R., Lubello, C., van Loosdrecht, M.C.M., 2014. Physiological and kinetic characterization of a suspended cell anammox culture. *Water Res.* 60, 1–14, <http://dx.doi.org/10.1016/j.watres.2014.04.017>.
- Lotti, T., Kleerebezem, R., Abelleira-Pereira, J.M., Abbas, B., van Loosdrecht, M.C.M., 2015. Faster through training: the anammox case. *Water Res.* 81, 261–268, <http://dx.doi.org/10.1016/j.watres.2015.06.001>.
- Ma, B., Wang, S., Cao, S., Miao, Y., Jia, F., Du, R., Peng, Y., 2016. Biological nitrogen removal from sewage via anammox: recent advances. *Bioresour. Technol.* 200, 981–990, <http://dx.doi.org/10.1016/j.biortech.2015.10.074>.
- Magrì, A., Vanotti, M.B., Szögi, A.A., 2012a. Anammox sludge immobilized in polyvinyl alcohol (PVA) cryogel carriers. *Bioresour. Technol.* 114, 231–240, <http://dx.doi.org/10.1016/j.biortech.2012.03.077>.
- Magrì, A., Vanotti, M.B., Szögi, A.A., Cantrell, K.B., 2012b. Partial nitrification of swine wastewater in view of its coupling with the anammox process. *J. Environ. Qual.* 41, 1989–2000, <http://dx.doi.org/10.2134/jeq2012.0092>.
- Magrì, A., Béline, F., Dabert, P., 2013. Feasibility and interest of the anammox process as treatment alternative for anaerobic digester supernatants in manure processing – an overview. *J. Environ. Manage.* 131, 170–184, <http://dx.doi.org/10.1016/j.jenvman.2013.09.021>.
- Magurran, A.E., 2004. *Measuring Biological Diversity*. Blackwell Science Ltd., UK.
- Okabe, S., Oshiki, M., Takahashi, Y., Satoh, H., 2011. N₂O emission from a partial nitrification-anammox process and identification of a key biological process of N₂O emission from anammox granules. *Water Res.* 45, 6461–6470, <http://dx.doi.org/10.1016/j.watres.2011.09.040>.
- Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Wagner, H., 2016. *Package vegan, community ecology package, ver. 2.3–4*, <https://cran.r-project.org/web/packages/vegan/vegan.pdf>. (Accessed 28 March 2016).
- Oshiki, M., Shimokawa, M., Fujii, N., Satoh, H., Okabe, S., 2011. Physiological characteristics of the anaerobic ammonium-oxidizing bacterium '*Candidatus* Brocadia sinica'. *Microbiology* 157, 1706–1713, <http://dx.doi.org/10.1099/mic.0.048595-0>.
- Pereira, A.D., Leal, C.D., Dias, M.F., Etchebehere, C., Chernicharo, C.A.L., de Araújo, J.C., 2014. Effect of phenol on the nitrogen removal performance and microbial community structure and composition of an anammox reactor. *Bioresour. Technol.* 166, 103–111, <http://dx.doi.org/10.1016/j.biortech.2014.05.043>.
- Poirier, S., Bize, A., Bureau, C., Bouchez, T., Chapleur, O., 2016a. Community shifts within anaerobic digestion microbiota during phenol inhibition: towards early warning microbial indicators? *Water Res.* 100, 296–305, <http://dx.doi.org/10.1016/j.watres.2016.05.041>.
- Poirier, S., Quémener, E.D.-L., Madigou, C., Bouchez, T., Chapleur, O., 2016b. Anaerobic digestion of biowaste under extreme ammonia concentration: identification of key microbial phylotypes. *Bioresour. Technol.* 207, 92–101, <http://dx.doi.org/10.1016/j.biortech.2016.01.124>.
- Quan, Z.-X., Rhee, S.-K., Zuo, J.-E., Yang, Y., Bae, J.-W., Park, J.R., Lee, S.-T., Park, Y.-H., 2008. Diversity of ammonium-oxidizing bacteria in a granular sludge anaerobic ammonium-oxidizing (anammox) reactor. *Environ. Microbiol.* 10, 3130–3139, <http://dx.doi.org/10.1111/j.1462-2920.2008.01642.x>.

- Quast, C., Pruesse, E., Yilmaz, P., Gerken, J., Schweer, T., Yarza, P., Peplies, J., Glöckner, F.O., 2013. The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic Acids Res.* 41, D590–D596, <http://dx.doi.org/10.1093/nar/gks1219>.
- Schloss, P.D., Westcott, S.L., Ryabin, T., Hall, J.R., Hartmann, M., Hollister, E.B., Lesniewski, R.A., Oakley, B.B., Parks, D.H., Robinson, C.J., Sahl, J.W., Stres, B., Thallinger, G.G., Van Horn, D.J., Weber, C.F., 2009. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl. Environ. Microbiol.* 75, 7537–7541, <http://dx.doi.org/10.1128/AEM.01541-09>.
- Schmid, M., Twachtmann, U., Klein, M., Strous, M., Juretschko, S., Jetten, M., Metzger, J.W., Schleifer, K.-H., Wagner, M., 2000. Molecular evidence for genus level diversity of bacteria capable of catalyzing anaerobic ammonium oxidation. *Syst. Appl. Microbiol.* 23, 93–106, [http://dx.doi.org/10.1016/S0723-2020\(00\)80050-8](http://dx.doi.org/10.1016/S0723-2020(00)80050-8).
- Siegrist, H., Salzgeber, D., Eugster, J., Joss, A., 2008. Anammox brings WWTP closer to energy autarky due to increased biogas production and reduced aeration energy for N-removal. *Water Sci. Technol.* 57, 383–388, <http://dx.doi.org/10.2166/wst.2008.048>.
- Strous, M., Heijnen, J.J., Kuenen, J.G., Jetten, M.S.M., 1998. The sequencing batch reactor as a powerful tool for the study of slowly growing anaerobic ammonium-oxidizing microorganisms. *Appl. Microbiol. Biotechnol.* 50, 589–596, <http://dx.doi.org/10.1007/s002530051340>.
- Strous, M., Fuerst, J.A., Kramer, E.H.M., Logemann, S., Muyzer, G., van de Pas-Schoonen, K.T., Webb, R., Kuenen, J.G., Jetten, M.S.M., 1999. Missing lithotroph identified as new planctomycete. *Nature* 400, 446–449, <http://dx.doi.org/10.1038/22749>.
- Tamaki, H., Tanaka, Y., Matsuzawa, H., Muramatsu, M., Meng, X.-Y., Hanada, S., Mori, K., Kamagata, Y., 2011. *Armatimonas rosea* gen. nov., sp. nov., of a novel bacterial phylum, *Armatimonadetes* phyl. nov., formally called the candidate phylum OP10. *Int. J. Syst. Evol. Microbiol.* 61, 1442–1447, <http://dx.doi.org/10.1099/ijs.0.025643-0>.
- Tsushima, I., Ogasawara, Y., Kandaichi, T., Satoh, H., Okabe, S., 2007. Development of high-rate anaerobic ammonium-oxidizing (anammox) biofilm reactors. *Water Res.* 41, 1623–1634, <http://dx.doi.org/10.1016/j.watres.2007.01.050>.
- Uyanik, S., Bekmezci, O.K., Yurtsever, A., 2011. Strategies for successful ANAMMOX enrichment at laboratory scale. *Clean-Soil Air Water* 39, 653–657, <http://dx.doi.org/10.1002/clen.201000591>.
- Van Hulle, S.W.H., Vandeweyer, H.J.P., Meesschaert, B.D., Vanrolleghem, P.A., Dejana, P., Dumoulin, A., 2010. Engineering aspects and practical application of autotrophic nitrogen removal from nitrogen rich streams. *Chem. Eng. J.* 162, 1–20, <http://dx.doi.org/10.1016/j.cej.2010.05.037>.
- van Niftrik, L., Jetten, M.S.M., 2012. Anaerobic ammonium-oxidizing bacteria: unique microorganisms with exceptional properties. *Microbiol. Mol. Biol. Rev.* 76, 585–596, <http://dx.doi.org/10.1128/MMBR.05025-11>.
- Vanotti, M., Fujii, T., Szögi, A., Rothrock, M., Garcia, M.C., Kunz, A., Magrí, A., Furukawa, K., 2011. Experiences with anammox in the USA: isolation, preservation and treatment performance of *Brocadia caroliniensis*. In: *Proc. 1st International Anammox Symposium (IANAS 2011)*, Kumamoto University, Japan, pp. 99–106.
- Venables, W.N., Smith, D.M., The R Core Team, 2016. *An Introduction to R, Notes on R: a Programming Environment for Data Analysis and Graphics*. Ver. 3.2.4, <https://cran.r-project.org/doc/manuals/r-release/R-intro.pdf>. (Accessed 28 March 2016).
- Wang, Y., Qian, P.-Y., 2009. Conservative fragments in bacterial 16S rRNA genes and primer design for 16S ribosomal DNA amplicons in metagenomic studies. *PLoS One* 4, e7401, <http://dx.doi.org/10.1371/journal.pone.0007401>.
- Wang, T., Zhang, H., Yang, F., Liu, S., Fu, Z., Chen, H., 2009. Start-up of the anammox process from the conventional activated sludge in a membrane bioreactor. *Bioresour. Technol.* 100, 2501–2506, <http://dx.doi.org/10.1016/j.biortech.2008.12.011>.
- Wang, T., Zhang, H., Yang, F., 2016a. Performance of Anammox process and low-oxygen adaptability of Anammox biofilms in a FBR with small ring non-woven carriers. *Ecol. Eng.* 86, 126–134, <http://dx.doi.org/10.1016/j.ecoleng.2015.11.025>.
- Wang, Y., Hu, X., Jiang, B., Song, Z., Ma, Y., 2016b. Symbiotic relationship analysis of predominant bacteria in a lab-scale anammox UASB bioreactor. *Environ. Sci. Pollut. Res.* 23, 7615–7626, <http://dx.doi.org/10.1007/s11356-015-6016-z>.
- Xiong, L., Wang, Y.-Y., Tang, C.-J., Chai, L.-Y., Xu, K.-Q., Song, Y.-X., Ali, M., Zheng, P., 2013. Start-up characteristics of a granule-based anammox UASB reactor seeded with anaerobic granular sludge. *Biomed Res. Int.*, <http://dx.doi.org/10.1155/2013/396487> (Article ID 396487).
- Yasuda, T., Waki, M., Yoshinaga, I., Amano, T., Suzuki, K., Tanaka, Y., Yamagishi, T., Suwa, Y., 2011. Evidence of exponential growth of an anammox population in an anaerobic batch culture. *Microbes Environ.* 26, 266–269, <http://dx.doi.org/10.1264/jisme2.ME10181>.
- Yin, H., Niu, J., Ren, Y., Cong, J., Zhang, X., Fan, F., Xiao, Y., Zhang, X., Deng, J., Xie, M., He, Z., Zhou, J., Liang, Y., Liu, X., 2015. An integrated insight into the response of sedimentary microbial communities to heavy metal contamination. *Sci. Rep.* 5, 14266, <http://dx.doi.org/10.1038/srep14266>.
- Zhang, L., Liu, M., Zhang, S., Yang, Y., Peng, Y., 2015. Integrated fixed-biofilm activated sludge reactor as a powerful tool to enrich anammox biofilm and granular sludge. *Chemosphere* 140, 114–118, <http://dx.doi.org/10.1016/j.chemosphere.2015.02.001>.