

### Active biomonitoring for assessment of chemical contamination and toxicity of aquatic environments

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# Active biomonitoring for assessment of chemical contamination and toxicity of aquatic environments



# Presentation of in situ bioassays by caging gammarids

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### **CONTEXT & CHALLENGE**

Standard approaches used to monitor the quality of continental surface water

- ❖ Chemical analysis of water → Toxicity? Bioavailability of chemical substances? ...
- ❖ Ecological indicators → Sources of pollution ? (habitat, chemicals, ...), ...

Need of complementary approaches like ecotoxicological assays to monitor contamination and toxicity of aquatic environments

### **OBJECTIVES**

### Development of ACTIVE BIOMONITORING with a freshwater crustacean

- 1. Method for in situ bioassays with Gammarus fossarum
- 2. Method for endpoints interpretation
- 3. Applications for field monitoring

### 1. METHOD for IN SITU BIOASSAYS with Gammarus fossarum



One natural source of organisms = Reference population (GAMMAREF®)



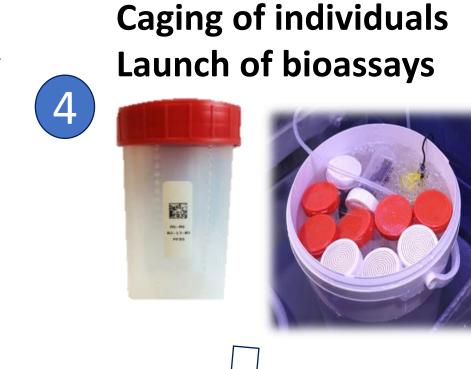
**Acclimatization** Laboratory controlled conditions during 2 weeks

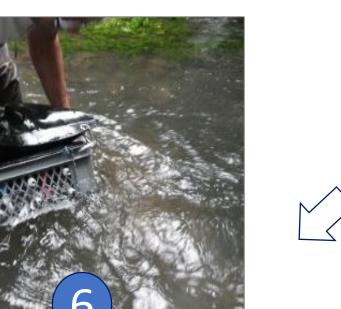
**Transport** 

to laboratory



**Calibration of** individuals: size, sex, reproductive status, ...





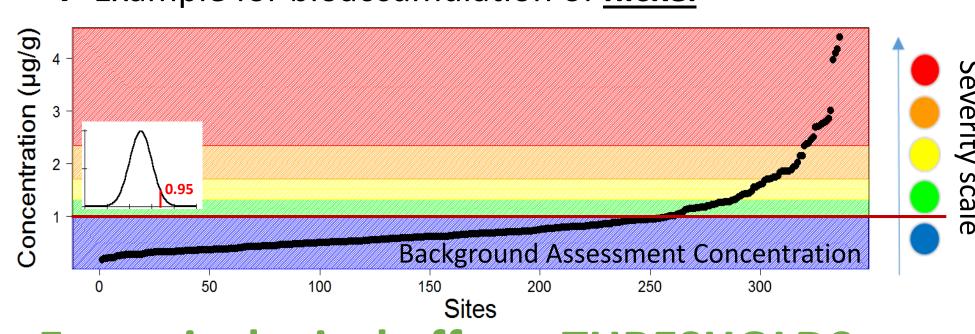
Field exposure during 1 or 3 weeks with continuous

### a. Bioaccumulation BIOTA EQS CONFORMITY tracking of 15 substances for conformity to the WFD

b. Bioaccumulation THRESHOLDS

defined from a large scale field experiments (database)

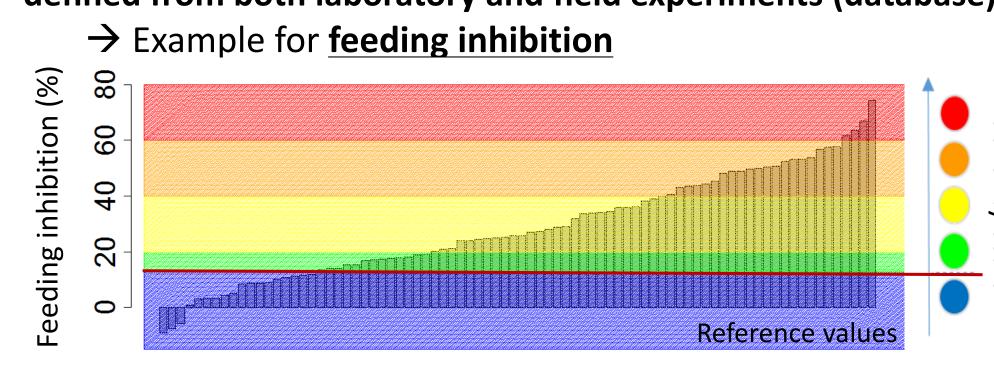
→ Example for bioaccumulation of <u>nickel</u>



2. METHOD for ENDPOINTS INTERPRETATION

### c. Ecotoxicological effects THRESHOLDS

defined from both laboratory and field experiments (database)



## **ENDPOINTS**

- \*Bioaccumulation : bioavailable concentration of micropollutants (metals and organic compounds)
- **Ecotoxicological effects**: survival, feeding, AChE, reproduction and endocrine disruption

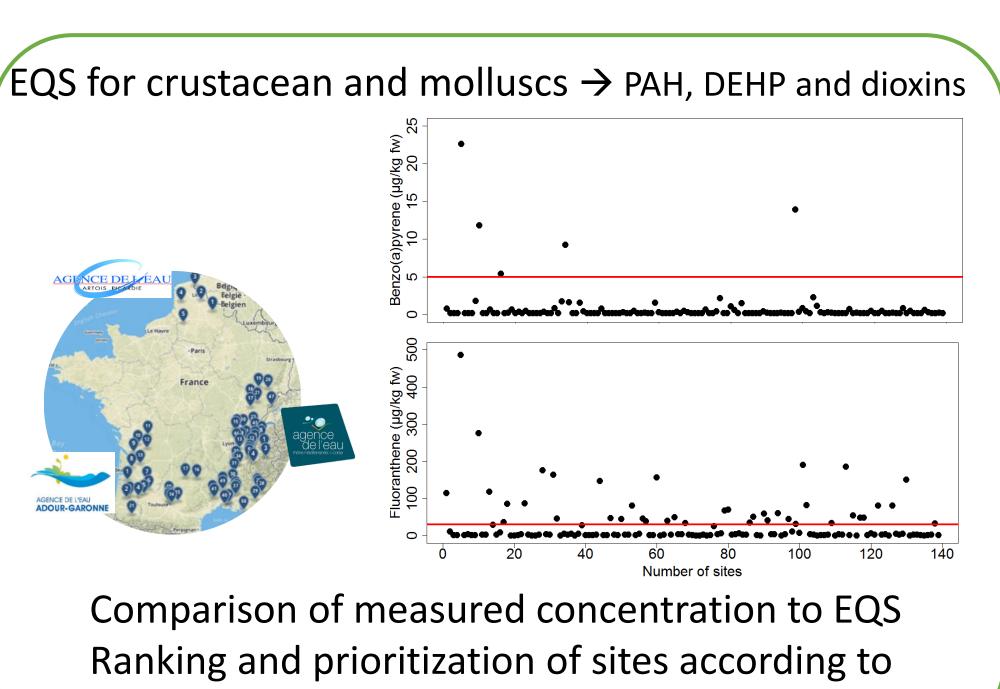
# temperature control

## 3. APPLICATIONS for FIELD MONITORING

**Transport** 

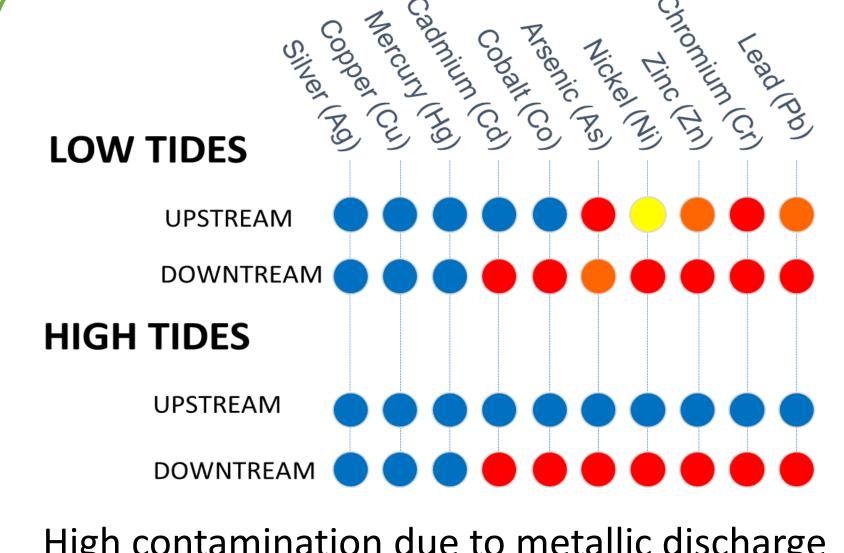
to field stations

### a. LARGE SCALE DEPLOYMENT IN WATER AGENCIES



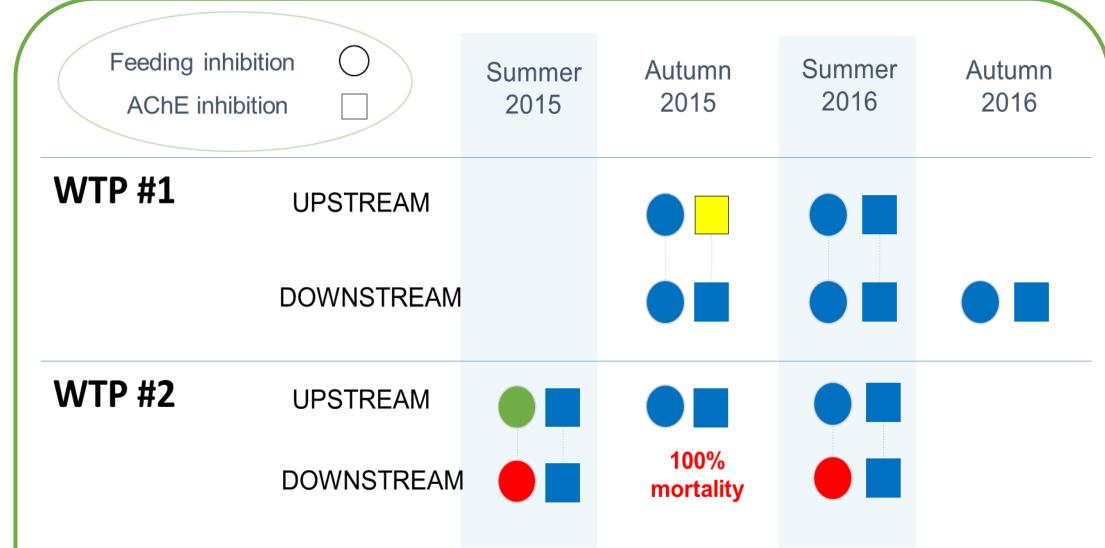
contamination

### b. INDUSTRIAL DISCHARGE



High contamination due to metallic discharge Temporal variation (dilution due to outflow) Dilution after confluence (data not shown)

### c. WASTEWATER TREATMENT PLANTS



WTP#1: no toxic impact; upstream neurotoxicity (AChE) WTP#2: toxic impact, especially in autumn (strong low-water period)

## **CONCLUSION**

### In situ bioassays: relevant and complementary approach for biomonitoring

Selected organisms (versus passive monitoring) to control biological confounding factors Realistic and integrative exposure (versus laboratory exposure)

Proposition of reference values integrating effects of environmental confounding factors Spatial and/or Temporal gradients with a « one-week » resolution scale

Operational for large scale deployment

For more information

### Wide range of applications for public managers and industrials

REGULATION -> Compliance to EQS in biota (WFD, 2013) Spatial and temporal comparison of stations into monitoring networks Impact studies of industrials and hydraulic structures Assessment of WTP treatment efficiency

[1] JP. Besse, O. Geffard, M. Coquery. Relevance and applicability of active biomonitoring in continental waters under the Water Framework Directive. Trends in Analytical Chemistry, Vol. 36, 2012 [2] M. Carere, V. Dulio, G. Hank. A. Hebert. Guidance for sediment and biota monitoring under the Common Implementation Strategy for the Water Framework Directive. Trends in Analytical Chemistry, Vol. 36, 2012. [3] P.Y. Kunz, C. Kienle, A. Gerhardt. Gammarus spp. in aquatic ecotoxicology and water quality assessment: Toward integrated multilevel tests. (2010) Reviews of Environmental Contamination and Toxicology, 205, pp.

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water quality biomonitoring. Water Research. 2011, 45, 6417–6429

of Biodiversity

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### **REFERENCES**

1-76. [4] DIRECTIVE 2013/39/EU OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL of 12 August 2013 amending Directives 2000/60/EC and 2008/105/EC as regards priority substances in the field of water policy. Official

[5] Coulaud, R.; Geffard, O.; Xuereb, B.; Lacaze, E.; Quéau, H.; Garric, J.; Charles, S.; Chaumot, A. In situ feeding assay with Gammarus fossarum (Crustacea): Modelling the influence of confounding factors to improve