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**Novel insights into the diet of the Pyrenean desman,
 Galemys pyrenaicus, using next-generation sequencing
 molecular analyses**

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Keywords:	Foraging habitat, scat analysis, semi-aquatic mammal, spatial structure

1 **Novel insights into the diet of the Pyrenean desman, *Galemys pyrenaicus*, using next-**
2 **generation sequencing molecular analyses**

3 **Running header:** Genetic analyses of Pyrenean desman diet

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26 To date, the Pyrenean desman, a threatened, semi-aquatic mammal, is considered a specialist
27 predator feeding on aquatic benthic invertebrates. This knowledge comes from visual
28 identification of prey in scat or gut contents, often based on a limited number of samples and
29 locations. In this study, diet analyses using next-generation sequencing methods were
30 combined with an extensive survey to explore the Pyrenean desman summer diet across the
31 French Pyrenees. This study thus provides an unprecedented level of detail on Pyrenean
32 desman trophic ecology. Our results highlighted an extremely diverse diet containing a high
33 proportion of rare prey and substantial consumption of terrestrial prey, which suggests a more
34 generalist diet than previously understood. Three diet groups were identified, with significant
35 differences in prey composition. These differences were, however, not related to geographic
36 location, but rather to contrasted local environmental variables. These results suggest that the
37 highlighted spatial dietary variation is likely induced by local abiotic parameters that affect
38 prey availability or foraging habitat use.

39

40 **Keywords:** foraging habitat, scat analysis, semi-aquatic mammal, spatial structure

41 Enhancing knowledge of how species interact with their environment and with other
42 organisms is required to evaluate species vulnerability (Rodrigues et al. 2006). Biotic
43 interactions such as competition, predation, and trophic resource availability are known to
44 play a central role in the distribution of species and their abundance (Boulangeat et al. 2012;
45 Kissling et al. 2012), and to influence their responses to changing environments at different
46 scales (Pearson & Dawson, 2003; Araújo & Luoto, 2007; Wisz et al. 2013; Belmaker et al.
47 2015). For instance, the decline of the Iberian lynx, *Lynx pardinus*, the world's most
48 threatened felid, was reported to be caused by the decline of its staple prey, the European
49 rabbit, *Oryctolagus cuniculus* (Real et al. 2009), due to changes in environment and diseases
50 (e.g., myxomatosis). In studies focused on one single species, the biotic component has often
51 been overlooked due to sparse data and the difficulty of quantifying and incorporating
52 complex relationships between several organisms (Soberón & Peterson, 2005). This is
53 particularly true when the species of interest is rare and elusive, and thus difficult to study.

54 The Pyrenean desman, *Galemys pyrenaicus* (E. Geoffroy Saint-Hilaire, 1811), is a small,
55 semi-aquatic mammal (Talpidae) endemic to the Pyrenees Mountains (France, Spain, and
56 Andorra) and the Iberian Peninsula (northern and central Spain, northern Portugal). The
57 species is listed as vulnerable by the IUCN (Fernandes et al. 2008), and is legally protected in
58 the four countries encompassing its distribution area. For a long time, the Pyrenean desman
59 remained relatively unstudied, while also suffering from a substantial decline across its range
60 (Fernandes et al. 2008; Gisbert & García-Perea, 2014; Charbonnel et al. 2016). Although
61 recent studies have improved general knowledge about this species (e.g., habitat: Charbonnel
62 et al. 2015 and Biffi et al. 2016; space use: Melero et al. 2012, 2014; population genetics:
63 Gillet et al. 2015, 2016), a lack of information on biotic interactions, both with predators and
64 trophic resources, remains to be supplemented.

65 To study the diet of small mammals, morphological identification of prey remains has been
66 widely used (e.g., Castién & Gosálbez, 1999; Churchfield & Rychlik, 2006). With these
67 methods, the Pyrenean desman was considered a specialist predator that fed on benthic
68 invertebrates in streams (mainly Trichoptera, Ephemeroptera, Plecoptera or Diptera, e.g.,
69 Santamarina & Guitian, 1988; Santamarina, 1993; Bertrand, 1994; Castién & Gosálbez,
70 1995).

71 Even though traditional methods enable the identification of a wide variety of prey, they are
72 becoming less and less popular because (i) they generally involve the destruction of animals
73 to analyse gut contents, (ii) they provide degraded prey remains that cannot be identified with
74 precision (rarely beyond the family level), (iii) they underestimate the consumption of prey
75 that are thoroughly masticated (e.g., arthropods consumed by bats) and soft-bodied prey (e.g.,
76 molluscs and earthworms) that leave no hard remains after digestion, and (iv) they are time-
77 consuming and require expert knowledge of the range of potential prey. The recent
78 development of molecular analyses based on barcoding of faecal DNA fragments and next-
79 generation sequencing has overcome these difficulties (see a review by Pompanon et al.
80 2012).

81 In that context, the first aim of this study was to improve the knowledge of the trophic
82 ecology of the Pyrenean desman by combining next-generation molecular analyses and an
83 extensive sampling covering the species range over the whole French Pyrenees. Gillet (2015)
84 recently identified three Pyrenean desman genetic populations using contrasted habitats (Biffi
85 et al., 2016) in the three main hydrographic regions of the area. The second aim of the study
86 was to compare the diet of the Pyrenean desman in those three regions. We hypothesized that

87 the diet would vary according to the three hydrographic regions of the French Pyrenees,
88 which differ in terms of catchment surface, climate, elevation, and land cover.

89 MATERIALS AND METHODS

90 *Study area.*— The Pyrenees Mountains are located in southwestern Europe and are a
91 natural barrier between France and Spain. This study focused on the French part of the
92 Pyrenees (W1°400–E3°100, N43°080–N42°230; Fig. 1), which extends for about 400 km
93 from the Bay of Biscay to the Mediterranean Sea and has a maximum elevation of 3,298 m.
94 The stream network can be divided into three main hydrographic regions: (1) the western
95 streams that flow mostly westward to the Atlantic Ocean (coastal streams and Adour
96 catchment; 9,412 km² in the Pyrenees), (2) the central streams that flow mainly northward and
97 form the upstream part of the Garonne river catchment (7,702 km² in the Pyrenees), and (3)
98 the eastern streams that flow to the Mediterranean Sea (Aude, Tech and Têt catchments; 6,773
99 km² in the Pyrenees).

100 *Faeces collection.*— A national survey was conducted from 2011 to 2013 within the
101 framework of the Conservation Action Plan for the Pyrenean desman (Némoz et al. 2011).
102 Given the elusive behaviour of this species, searches for indirect signs (i.e., faeces) were
103 conducted in 1,330 sites covering the entire French Pyrenees (Fig. 1A). Skilled observers
104 meticulously inspected each emergent item (i.e., rock, tree root, and branch) along 500 m
105 riverbed transects (Castién & Gosálbez, 1992; Bertrand, 1994; Aymerich & Gosálbez, 2002;
106 Charbonnel et al. 2014, 2015; Biffi et al. 2016). From the 1,330 original sites, 989 faeces
107 suspected of being left by a Pyrenean desman, based on their colour, size, smell, and position,
108 were collected in 958 sites and preserved in absolute ethanol for further molecular analyses.

109 *Molecular analyses.*— DNA was extracted from faeces using the Stool Mini Kit
110 (Qiagen Inc., Hilden, Germany), following the manufacturer's instructions. PCR
111 amplifications and Ion Torrent PGM sequencing (Life Technologies) were duplicated on the
112 989 faecal samples following the tagging and multiplexing method developed by Galan et al.
113 (2012). Briefly, a 133 bp mini-barcode of the cytochrome oxidase I gene (COI) was
114 amplified for each sample using a modified forward primer LepF1 (Hebert et al. 2004): 5'-
115 CCATCTCATCCCTGCGTGTCTCCGACTCAGNNNNNNNATTCHACDAAYCAYAARG
116 AYATYGG-3', and a modified reverse primer EPT-long-univR (Hajibabaeiet al. 2011): 5'-
117 CCTCTCTATGGGCAGTCGGTGATNNNNNNNACTATAAAARAAAATYTDAYAAA
118 DGCRTG-3'. The 5' parts of the primers were modified by the addition of individual-specific
119 MIDs (Multiplex IDentifiers NNNNNNN), consisting of a short 7 bp sequence and adaptors
120 required for emPCR and Ion Torrent sequencing. By using a combination of different forward
121 and reverse MIDs sequences, several hundred samples can be multiplexed in the same
122 sequencing run, and the sequences can be recognized after sequencing, when all the PCR
123 products from the different samples are mixed together (Gillet et al., 2015)

124 PCRs were carried out in a 10 µl reaction volume using 5 µl of 2x QIAGEN Multiplex Kit
125 (Qiagen, Hilden, Germany), 0.5 µM of each primer (LepF1 and EPT-long-univR,
126 concentrated at 10µM), and 1 µl of DNA extract. The PCR conditions consisted of an initial
127 denaturation step at 95 °C for 15 min, followed by 40 cycles of denaturation at 94 °C for 30 s,
128 annealing at 45 °C for 45 s, and extension at 72 °C for 30 s, followed by a final extension step
129 at 72 °C for 10 min. After pooling all PCR products at 5 ng/µL, the amplicon libraries were
130 sequenced by the company Genes Diffusion (Douai, France) on an Ion Torrent PGM system
131 using an Ion 316 Chip version 2 (LifeTechnologies).

132 In addition, a customised database of COI sequences was built from 84 typical invertebrate
133 species from the study area collected in some Pyrenean rivers and identified by local
134 entomologist experts from the Conservatoire d'Espaces Naturels Midi-Pyrénées (CEN-MP)
135 and the EcoLab laboratory of Toulouse. A 710 bp fragment of COI was amplified in these
136 samples with universal primers LCO1490 and HCO2198 (Folmer et al. 1994), following the
137 PCR conditions reported in Folmer et al. (1994).

138 The sequences were sorted using SESAME barcode software (SEquence Sorter & AMplicon
139 Explorer; Piry et al. 2012). These sequences were compared with sequences available in the
140 customized and BOLD databases (Ratnasingham & Hebert, 2007). Sequences that had a
141 unique best-hit with an identity score greater than or equal to 98% were considered to be
142 positive matches and allowed identification of the predator producing the faeces as well as the
143 prey contained in them.

144 Taxa were validated as possibly occurring in France and in the Pyrenees region using the
145 French National Inventory of Natural Heritage (Muséum national d'Histoire naturelle, 2003-
146 2017), the French Office for Insects and their Environment (OPIE-Benthos, 2017) online
147 databases, and the help of local experts. Taxa identified as endemic to other parts of the world
148 were kept in the analysis and are designated by an asterisk hereafter (*), as they correspond
149 more likely to a genetically similar taxon present in the Pyrenees but not available in the
150 reference databases.

151 The frequency of occurrence (i.e., the number of faeces containing the taxon divided by the
152 total number of Pyrenean desman faeces) in the Pyrenean desman diet was then calculated for
153 each order, family, and genus, and for the different types of prey's modes of life (i.e.,
154 exclusively aquatic, exclusively terrestrial, or with aquatic and terrestrial stages).

155 *Study of the summer Pyrenean desman diet.*— To investigate a potential spatial
156 structure in the Pyrenean desman diet, 287 Pyrenean desman faeces collected at 115 sampling
157 sites were kept for statistical analysis. This strong reduction in the number of faeces compared
158 to the initial pool of faeces (n=989) was due to drastic selection following three criteria: (i)
159 the predator producing the faeces has been identified by molecular analyses, (ii) the prey
160 contained in the faeces have been identified by molecular analyses, too, and (iii) the faeces
161 were collected during summer (i.e., June to September) in order to prevent potential strong
162 seasonal variation. The summer season was chosen as most samplings were conducted during
163 this low-flow period between 2011 and 2013. Prey occurrences were not different between the
164 three summers of sampling, as revealed by a non-parametric permutation-based multivariate
165 analysis of variance (PERMANOVA, Anderson, 2001), which was not significant ($p = 0.12$).
166 To correct for potential sampling bias (i.e., more than one faeces collected per site), all the
167 faeces collected in a single site were pooled. Consequently, a taxon was assumed to be
168 present in the Pyrenean desman diet of a site if it was found in at least 50% of the faeces
169 collected for this site. All prey taxa were kept for subsequent analysis.

170 *Cluster Analysis And Identification Of Summer Diet Groups* - The binary matrix (i.e.,
171 presence/absence of prey genera at each site) was converted to a distance matrix calculated
172 with the Sørensen similarity coefficient, which is the equivalent of Bray-Curtis distance but
173 for presence-absence instead of abundance data (Borcard et al., 2011). Then, a hierarchical
174 ascendant clustering was computed with Ward's algorithm to build a dendrogram representing
175 the distance between each pair of sites according to their similarity in the summer Pyrenean
176 desman diet (i.e., prey taxa genetically identified in faeces). The number of diet groups was
177 chosen according to the dendrogram, so that it increases the variation between groups and

178 decreases the variation within groups while keeping a relatively balanced number of sites
179 within groups.

180 The mean composition of prey communities within sites was compared among the three diet
181 groups using a PERMANOVA (Anderson, 2001). As PERMANOVA may be sensitive to
182 within-group effects, PERMDISP (Anderson et al. 2006), a permutation dispersion analysis,
183 was then used to test for differences in within-group dissimilarity (i.e., the variability of diet
184 composition within the groups as the mean distance of diet compositions to their group
185 centroid). When the PERMDISP test was significant, a pairwise Tukey HSD test was run to
186 examine which of the diet groups had higher dispersion. The Sørensen matrix was used as the
187 measure of similarity in both PERMANOVA and PERMDISP.

188 In order to identify the prey taxa specific to each diet group, an indicator value (IndVal)
189 analysis was performed (Dufrêne & Legendre, 1997; Cáceres & Legendre, 2009) using the
190 occurrence matrix of the summer prey taxa and the classification of sites in diet groups
191 obtained through the hierarchical clustering. The IndVal is the product of the specificity (i.e.,
192 the probability that the survey site belongs to the target site group, given the fact that the
193 species has been found) and the fidelity (i.e., the probability of finding the species in sites
194 belonging to the site group) of species for each cluster. A taxon was only able to be a
195 potential indicator of one diet group, as group combinations were not allowed. The IndVal
196 statistical significance was tested using a permutation test (9,999 permutations) for each prey
197 taxon.

198 Finally, a chi-squared test was performed to test whether the assignment of sites to the three
199 diet groups according to prey community composition found in faeces was dependent on their
200 location in the three hydrographic regions of the French Pyrenees.

223 The proportion of items obtained twice in the PCR duplicates for the same DNA extract was
224 computed as the percentage of repeatability. Repeatability of the results reached 99% when
225 only host species were considered, 83% when only prey species were considered, and 87%
226 when all species were considered.

227 *Overall diversity of prey contained in the Pyrenean desman faeces.*— The 383
228 Pyrenean desman faeces collected between 2011 and 2013 contained prey belonging to 11
229 classes, 30 orders (Table 2), 91 families, and 156 genera (Supplementary Data S1). A mean of
230 5.8 ± 2.0 genera were present per faeces (5.1 ± 1.9 and 3.7 ± 1.2 for family and order,
231 respectively). Among the 156 genera, 100 were confirmed to be present in the Pyrenees, 31 in
232 France, and 21 were endemic of other parts of the World (e.g., New Zealand, Australia) and
233 were thus misidentified by genetic databases. The presence in the Pyrenees of the four
234 additional genera remains unknown. For one taxon, databases were not able to discriminate
235 between two macroinvertebrate genera and families (designated hereafter as
236 “*Perlodes_Epeorus**” at the genus level).

237 The diet of the Pyrenean desman was mainly composed of Insecta and Malacostraca, which
238 were present in 99.7% and 18.0% of the 383 faeces respectively (Table 2). Other classes were
239 present in less than 4% of the Pyrenean desman faeces (e.g., Diplopoda, Lissamphibia,
240 Arachnida, Clitellata, Gastropoda). Only 7 orders (23.3%), 14 families (15.4%), and 14
241 genera (9.0%) had a frequency of occurrence in the faeces above 10%, meaning that the
242 majority of prey taxa were found in less than 10% of the faeces. The most frequent order was
243 Ephemeroptera (86.7%), which included the most frequent family, Heptageniidae (59.0%),
244 and the most frequent genus, *Baetis* (Baetidae; 56.4%; Supplementary Data S1).

245 Ephemeroptera, Plecoptera, and Trichoptera (EPT) together represented on average 79.1% of
246 the identified genera in each faeces. However, when considering the total number of identified
247 taxa (TIT) in the overall Pyrenean desman diet, EPT orders represented only 28.2% of
248 identified prey. Exclusively aquatic items represented 4.1% of prey vs. 7.7% of TIT, while
249 exclusively terrestrial prey represented 7.7% of prey (vs. 36.5% TIT). Taxa with at least one
250 aquatic stage and one terrestrial stage represented 88.3% of prey (vs. 55.8% TIT).

251 *Variation of summer diet across the French Pyrenees.*— When considering the
252 summer months (i.e., June to September), analyses were limited to 287 faeces collected from
253 115 sites. From these faeces, 91 prey genera were kept to study possible spatial variation in
254 the summer Pyrenean desman diet. Hierarchical ascendant clustering made it possible to
255 identify three distinct diet groups of sites (39, 48, and 28 sites, respectively) based on prey
256 assemblage (Fig. 2). There was no significant difference ($X^2 = 4.05$, d.f. = 2, $p = 0.13$)
257 between the number of samples collected per site and its classification by cluster analysis
258 (i.e., diet group), excluding potential bias due to diet profiles merging. Prey composition was
259 closer between Groups 1 and 2 than with Group 3. Within-site prey composition differed
260 significantly among groups (PERMANOVA: $F = 6.83$, d.f. = 2, $p < 0.01$; Fig. 3). The
261 dissimilarity of prey assemblages among sites similarly differed by diet group (PERMDISP: F
262 = 3.28, d.f. = 2, $p < 0.01$), and was driven by slightly lower among-site variability in Group 3
263 than in Group 2 (Tukey HSD: $p = 0.045$). There was no significant difference in among-site
264 variability between Groups 1 and 2, and Groups 1 and 3 (Tukey HSD: $p > 0.05$).

265 Diet Group 3 exhibited the least diverse prey assemblage, with 45 different invertebrate
266 genera consumed by the Pyrenean desman, versus 51 in Group 1 and 57 in Group 2. Only 21
267 genera were consumed in all three groups. Eleven significant indicator taxa were identified

268 for the three summer diet groups of the Pyrenean desman ($p < 0.05$, Table 3, Fig. 3). Group 2
269 was characterised by three Ephemeroptera taxa, whereas the indicator taxa of Group 1 and
270 Group 3 were more diverse (Trichoptera, Plecoptera, Diptera and Ephemeroptera for Group 1
271 and Amphipoda, Plecoptera and Diptera for Group 3, Table 3).

272 The differences in diet composition were not significantly related to the geographical location
273 of sites within the three hydrographic regions of the French Pyrenees ($X^2 = 5.93$, d.f. = 4, $p =$
274 0.20; Fig. 1B).

275 Linear discriminant analysis computed two significant functions, F1 and F2 (Monte-Carlo
276 test: $p = 0.001$), accounting for 60.8% and 39.2% of the variability between the three diet
277 groups, respectively (Fig. 4). The first function, F1, separated Group 3 from the two other
278 groups, whereas the second function, F2, separated Group 2 from Groups 1 and 3. The
279 environmental variables discriminating the three diet groups were (i) SHELTER (i.e., proxy
280 for riverbed heterogeneity) and SLO (i.e., slope of river section), which were positively
281 correlated with F1; (ii) WOOD (i.e., proportion of bankside with shrubby-woody vegetation),
282 which was negatively correlated with F1; (iii) HUM_IMP (i.e., proxy for human impacts),
283 which was positively correlated with F2; and (iv) SHEET (i.e., proportion of the stretch with
284 non-turbulent fast water units of shallow water that flows uniformly over smooth bedrock)
285 and TRI (i.e., number of tributaries), which were negatively correlated with F2 (Fig. 4). This
286 suggests that on average, the sites in Group 3 exhibited lower slopes, less heterogeneous
287 riverbeds, and wooded riverbanks (Table 4). Group 2 clustered sites with low human impact
288 along the near floodplain. The sites in Group 1 showed intermediate local environmental
289 conditions (Table 4).

290

DISCUSSION

291 *Next-generation sequencing as a powerful tool for studying the diet of the Pyrenean*
292 *desman.*— The amplification of a COI mini-barcode successfully provided the identification
293 of the Pyrenean desman and its prey species, as well as several other host species, in 57% of
294 faecal samples. This confirms the relevance of using such a genetic marker with next-
295 generation sequencing methods in diet assessments (Pompanon et al. 2012; Piñol et al. 2014;
296 Gillet et al. 2015) without the need for predator-specific blocking probes. The remaining 43%
297 of the samples could not be correctly assigned. According to McInnes et al. (2017), results are
298 highly dependent on faeces freshness and size. In addition, small faeces such as those of the
299 Pyrenean desman (10 to 15 mm long) limit the amount of DNA they contain, but also affect
300 the reproducibility of the extraction step, since the totality of the faeces has to be used.
301 Moreover, DNA is rapidly degraded by contact with water and UV radiation (Lindahl, 1993).

302 In spite of these shortcomings, molecular analysis allowed the identification of 156 different
303 invertebrate genera across the French Pyrenees. These genera belonged to 30 orders and 91
304 families, of which almost 70 were identified as Pyrenean desman prey for the first time. Using
305 traditional methods of faeces analysis, Bertrand (1994) had identified only 20 families from
306 an extensive sampling of 521 faeces collected in two small French catchments. In Northern
307 Spain, Castién & Gosálbez (1995) trapped 49 desmans throughout the year and were able to
308 identify only 11 orders as the lowest taxonomic level. The present study is therefore the most
309 extensive ever done on the diet of this species in terms of number of samples, geographic
310 coverage, and taxonomic resolution altogether. The high number of newly identified prey
311 emphasizes the efficiency of molecular analysis in detecting taxa that cannot be identified in
312 faeces through morphological analysis, and highlights the Pyrenean desman's capacity to
313 adapt to its trophic environment. Nevertheless, some taxa identified, such as Collembola,
314 Eurotatoria, and Eutardigrada, are unlikely to be direct prey of the Pyrenean desman, as they

315 are part of the soil microfauna or aquatic zooplankton. Other taxa, such as *Sargus* and small
316 Coleoptera, may develop at the larval stage or feed on scat, and may have thus been collected
317 with the faeces. This contributes to the debate about the high sensitivity of next-generation
318 sequencing methods and the detection of secondary predation (Sheppard et al. 2005).
319 Molecular diet analyses are also limited by the deficiencies in the reference databases: a taxon
320 can be identified at the species (or genus) level only if it has already been sequenced
321 (Pompanon et al. 2012). In our study, 21 prey species were endemic of other parts of the
322 world, and were thus misidentified. This issue stresses the importance of encouraging
323 inventories of genetic biodiversity. Recently, the usage of COI markers led to some concerns
324 in metabarcoding studies (see Deagle et al. 2014) using environmental DNA or bulk
325 biodiversity sample. However, in this study, we used only samples which belonged to
326 previously identified species. Some taxa could still not have been discovered but as we
327 specifically designed the primers and, in sight of the number of genera found among prey, we
328 are confident that the number of missing taxa is very low.

329 *A large diversity of prey in the Pyrenean desman diet.*—So far, the Pyrenean desman
330 has been described as a specialist predator targeting prey in an aquatic environment (e.g.,
331 Bertrand, 1994; Castién & Gosálbez, 1995). In this study, we confirmed that dietary
332 preferences seem to be directed toward Ephemeroptera, Plecoptera, and Trichoptera.
333 However, the wide variety of prey identified suggests a more generalist diet for the Pyrenean
334 desman. First, 91% of taxa were identified in less than 10% of the sites, representing a very
335 high proportion of infrequent prey. Second, the most frequent prey (e.g., *Baetis*,
336 *Protonemura*, and *Rhithrogena*) are among the most abundant aquatic macroinvertebrates in
337 the Pyrenees (e.g., Brown et al. 2006; Finn et al. 2013). Third, strictly terrestrial prey
338 represent about 8% of the prey eaten in each faeces and more than 35% of all identified prey

339 taxa. This substantial percentage of terrestrial invertebrate consumption may result from (i)
340 active hunting of terrestrial prey, (ii) opportunistic feeding on terrestrial prey while moving on
341 the banks, and/or (iii) consumption of drowned terrestrial invertebrates. Other aquatic (e.g.,
342 brown trout *Salmo trutta*) and semi-aquatic (e.g., European otter *Lutra lutra*) species are
343 known to rely to some extent on a pool of alternative prey including terrestrial subsidies to
344 fulfil their energetic needs (Clavero et al. 2003; Evangelista et al. 2014; Milardi et al. 2016).
345 This diversification of diet may be linked to aquatic stressors (e.g., pollution) or seasonal
346 effects (e.g., variation of climatic conditions and water flow) that limit or modify *in situ*
347 aquatic communities (Clavero et al. 2003; Kraus et al. 2016; Milardi et al. 2016). The highly
348 diverse summer diet of the Pyrenean desman could thus be considered here, as for the otter, a
349 response to summer drought conditions (Ruiz-Olmo et al. 2001). Indeed, during the summer
350 period, most aquatic insects have already emerged in mountain streams (Füreder et al. 2004)
351 or occur in small-size life stages, especially Ephemeroptera, Plecoptera and Trichoptera,
352 which induces a potential diversification of the Pyrenean desman prey diet towards an
353 increasing number of alternative aquatic and terrestrial food items. Its diet would be less
354 diverse and with less abundant terrestrial prey during the other seasons, but this would require
355 confirmation through a seasonal survey. This temporal survey should be combined with
356 standardized sampling of both terrestrial and aquatic potential prey on river banks and in
357 streams to get a quantitative assessment of the trophic resources available for the Pyrenean
358 desman and therefore better understand its feeding behaviour.

359 *Spatial variation in the Pyrenean desman diet and the influence of the environment.*—

360 While accounting only for the most common prey consumed during the summer by the
361 Pyrenean desman, three different diets were identified in the French Pyrenees according to
362 prey composition and indicator prey taxa. There was no major regional influence from the

363 three main hydrographic regions of the French Pyrenees, despite contrasted environmental
364 conditions from the western wet Atlantic area to the eastern dry Mediterranean. This result
365 does not support our assumption that the diet of the Pyrenean desman would be different
366 according to the genetic populations identified by Gillet (2015) or to the different habitat use
367 reported by Biffi et al. (2016).

368 However, sites grouped within the three Pyrenean desman diets exhibited differences in
369 environmental variables, suggesting some influence at the site scale. Group 2 diet sites
370 exhibited conditions typical of upstream parts of river basins with higher reach slope and
371 lower impact from human activities (i.e., urbanization). At the opposite end, the Group 3 diet
372 clustered lower altitude sites (e.g., low slope of river stretches) with quite high levels of
373 human impact on the near floodplain. These sites also included more homogeneous riverbeds.

374 First, the difference in prey composition among groups could be explained by the local
375 availability of prey, which is dependent on fine-scale environmental conditions. The
376 abundance and richness of aquatic macroinvertebrates are indeed directly dependent on local
377 habitat conditions, such as (i) human-induced pollution (e.g., near agricultural or urban areas),
378 impacts water quality (e.g., Johnson et al. 2013; Pallottini, et al. 2016), (ii) heterogeneity of
379 substrate types and emerging items, which provides different types of microhabitat in streams
380 (Reid et al. 2010), (iii) water current and oxygenation (Tachet et al., 2000; Usseglio-Polatera
381 et al. 2000), which are influenced by the slope of river reaches, and (iv) small-scale climate
382 variables, which can modify the period of invertebrate emergence and their availability in
383 streams (e.g., Füreder et al. 2004). Such diet adaptation to site-scale differences in resource
384 availability would suggest an adaptive foraging strategy for the Pyrenean desman.

385 Secondly, the availability of foraging habitats of the Pyrenean desman could be constrained
386 by local physical features. The species would thus feed on the fauna that is present in the
387 microhabitats of streams it can access. For instance, in sites of diet Group 3 where riverbeds
388 are the most homogeneous, the Pyrenean desman may forage on the littoral margins, while it
389 may have access to a more diverse choice of microhabitats and prey in sites of diet Groups 1
390 and 2, where riverbeds are more heterogeneous. This assumption is corroborated by the
391 indicator species identified for each diet. Indicator species of Group 3, such as *Gammarus* and
392 *Protonemura*, can indeed be found in zones of plant and organic debris accumulation (i.e.,
393 litter) and dense root hairs, which are typical of low-slope areas along riverbanks (Tachet et
394 al.2000; Usseglio-Polatera et al. 2000). Indicator species of Group 2 are Ephemeroptera taxa
395 sensitive to water quality. This is consistent with the apparent less disturbed environmental
396 conditions of these sites, which also provide higher water velocity (i.e., higher slopes) in more
397 open areas (i.e., higher altitude). These local conditions make possible the development of
398 biofilms, which are important food resources for scraper taxa such as *Ecdyonurus*. The
399 intermediate environmental conditions of Group 1 likely induced a higher diversity of
400 indicator taxa, which may reflect a more heterogeneous set of habitats.

401 *Conclusion and perspectives.*— This study suggests that the Pyrenean desman would
402 adopt a more generalist foraging strategy than previously reported. Even if a wider dietary
403 niche is supposed to increase the chance of adaptation to altered environments (Murgatroyd et
404 al. 2016), food availability remains an important issue for species conservation. Many
405 disturbances to freshwater environments may result in the general decline of abundance and
406 richness of aquatic invertebrate communities (Paul & Meyer, 2001) and have thus detrimental
407 consequences for the Pyrenean desman. In addition, many other threats affecting its nesting
408 (on river banks) and foraging (in river beds) habitats as well as population viability, such as

409 predation and mortality induced by human activities, should be limited as much as possible in
410 order to protect the Pyrenean desman. In that context, further studies will be needed to (i)
411 investigate the extent to which the diet and spatial distribution of the Pyrenean desman are
412 influenced by prey availability and other biotic interactions, and (ii) get a thorough knowledge
413 of its foraging areas within streams with the constraint that real-time monitoring and/or
414 radiotracking of this elusive species are difficult and costly to implement.

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431 Midi-Pyrenees and coordinated by the CEN MP.

432

SUPPLEMENTARY DATA

433 **Supplementary Data S1.**— Complete list of taxa identified as Pyrenean desman prey by
434 molecular analysis of 383 faeces collected in the French Pyrenees. Frequencies of occurrence
435 of each prey (FO: % of faeces with taxa) are displayed. * indicates misidentified taxa whose
436 distribution areas exclude the Pyrenees and France. Habitat type of taxa is given (aq:
437 exclusively aquatic; aq/te: with aquatic and terrestrial stages; te: exclusively terrestrial).

438

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607

FIGURES LEGENDS

608 **Fig. 1.**— Location of the study area in France. The three hydrographic regions of interest are
609 shown in light grey (western region: coastal streams and Adour catchment), medium grey
610 (central region: Garonne river catchment) and dark grey (eastern region: Aude, Tech and Têt
611 catchments). A) Location of all sampling sites (dark dots). B) Location of sampling sites
612 according to the three summer diet groups for the Pyrenean desman.

613 **Fig. 2.**— Hierarchical clustering dendrogram of Sørensen distances between the sampling
614 sites according to their similarity in the summer Pyrenean desman diet (i.e., 91 prey taxa
615 genetically identified in faeces). Three diet groups are identified.

616 **Fig. 3.**— Prey composition of the three groups of sampling sites based on the Pyrenean
617 desman summer diet composition (% of sites with prey). The diet groups were determined
618 from a hierarchical ascendant clustering based on a Sørensen distance matrix of presence-
619 absence of prey genera per site (see Fig. 1). Prey are sorted by decreasing proportion in the
620 first group of sites. * indicates prey taxa which were misidentified by the genetic databases.
621 Indicator taxa for diet groups are identified in bold and underlined characters (see Table 3 for
622 details). Habitat type of taxa is given (black triangles: exclusively aquatic; black rectangles:
623 exclusively terrestrial; no symbol: with aquatic and terrestrial stages).

624 **Fig. 4.**— Output of the linear discriminant analysis. The projection onto the first two linear
625 discriminants is displayed: A) 12 environmental variables; B) sites clustered by diet groups.
626 See Table 1 for variables description.

627

TABLES

628 **Table 1.**— Environmental variables (Biffi et al., 2016; Charbonnel et al., 2016) expected to
 629 influence the prey composition of the Pyrenean desman diet groups in the French Pyrenees.

630

Scale	Category	Code	Description
Site	Riverbed	SHELTER	Proportion (%) of the heterogeneity of shelters and substrate (i.e. presence of emerging items and cavities, diversity of substrate types and banks components) as a proxy for riverbed heterogeneity
		CLOG	Proportion (%) of the surface of streambed rocks covered by fine sediment (silt and mud)
		SHEET	Proportion (%) of the stretch with non-turbulent fast water units of shallow water that flows uniformly over smooth bedrock
		RUN	Proportion (%) of the stretch with non-turbulent fast water units of shallow gradient with relatively fine substrata ranging from sand to cobble
		RIFFLE	Proportion (%) of the stream stretch with turbulent fast water units with rapid and shallow flow with steep water surface gradient
	Riverbanks	BANKS	Shannon's diversity index derived from the proportion (%) of banks type components along the stretch (DIRT, ROOTS, ROCKS)
		WOOD	Proportion (%) of bankside with shrubby-woody vegetation
	Floodplain	HUM_IMP	Proportion (%) of urbanized land within the stream stretch floodplain as a proxy for human impacts
Reach	Climate	TEM	Atmospheric mean annual temperature (°C)
	Hydrology	FLO	Mean monthly stream flow (m ³ /s)
	Hydrography	SLO	Mean slope of the river section
		TRI	Number of tributaries in the focal section and in its adjacent upstream and downstream sections

631 **Table 2.**— Prey taxa identified with positive matches ($\geq 98\%$) from 383 faeces of Pyrenean
 632 desman collected in the French Pyrenees. Frequencies of occurrence (FO: % of faeces
 633 containing the prey) are displayed. See **Supplementary Data S1** for the complete list of taxa
 634 at a finer taxonomic resolution (family and genus). * indicates misidentified taxa by genetic
 635 databases (see Material and Methods).

Class	FO (%)	Order	FO (%)		
Insecta	99.7	Ephemeroptera	86.7		
		Plecoptera	84.9		
		Trichoptera	64		
		Diptera	45.7		
		Plecoptera_Ephemeroptera*	30.8		
		Coleoptera	14.1		
		Orthoptera	6.5		
		Neuroptera	1.8		
		Hemiptera	1.3		
		Lepidoptera	1.0		
		Odonata	0.5		
		Hymenoptera	0.3		
		Malacostraca	18	Amphipoda	16.7
				Isopoda	1.6
Decapoda	0.3				
Diplopoda	3.7	Glomerida	2.4		
		Polydesmida	0.8		
		Julida	0.5		
		Chordeumatida	0.3		
Lissamphibia	2.9	Urodeles	2.9		
Arachnida	2.4	Araneae	0.8		
		Opiliones	0.5		
		Sarcoptiformes	0.5		
		Trombidiformes	0.5		
		Haplotaxida	0.5		
Clitellata	0.5	Haplotaxida	0.5		
Collembola	0.3	Entomobryomorpha	0.3		
Eurotatoria	0.3	Bdelloida	0.3		
Eutardigrada	0.3	Parachela	0.3		
Gastropoda	0.3	Littorinimorpha	0.3		
Unidentified fish			0.5		

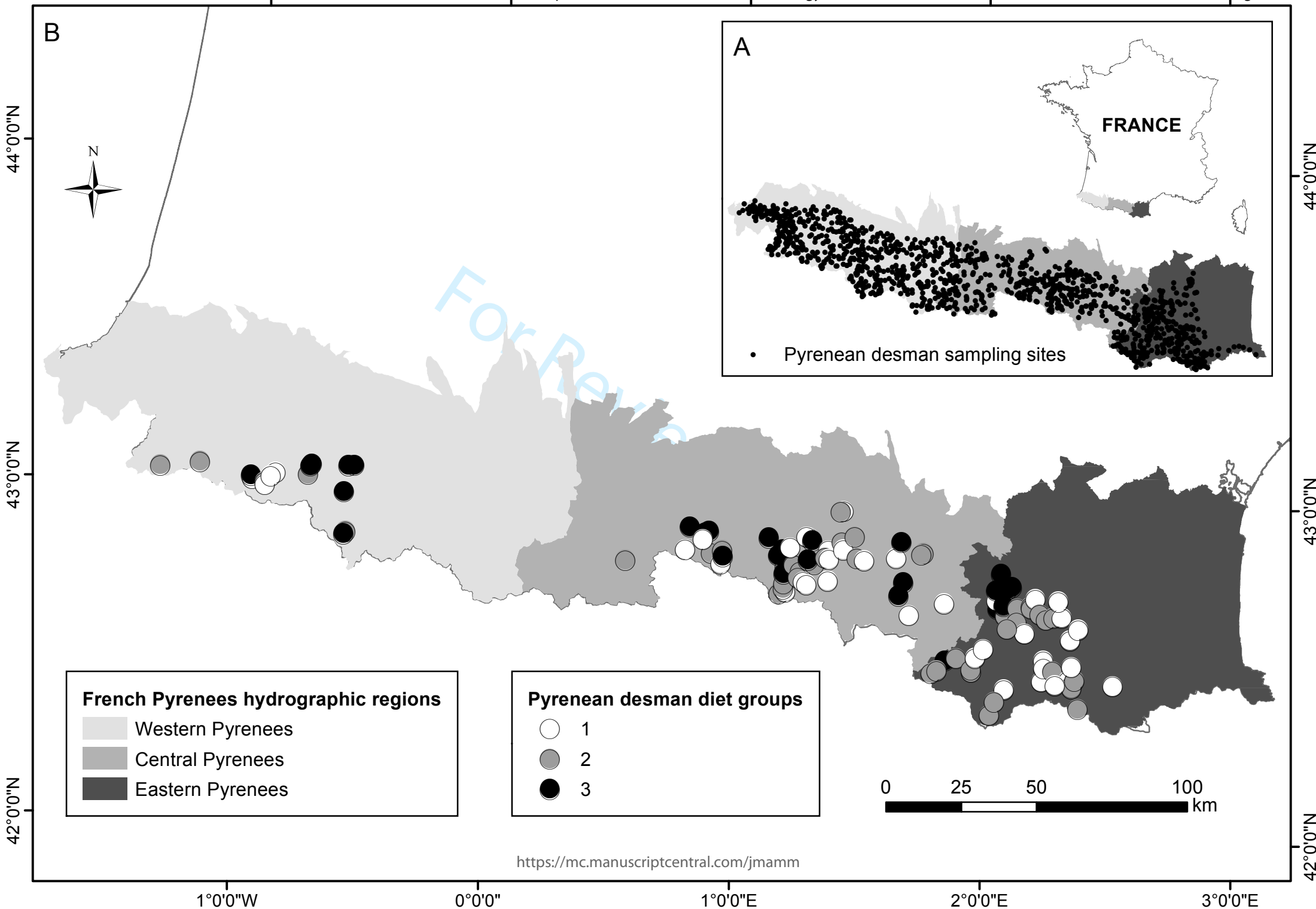
636 **Table 3.**— Significant indicator prey taxa ($p < 0.05$) of the three summer diet groups of the
 637 Pyrenean desman in the French Pyrenees identified by an IndVal analysis.

Groups	Indicator taxa		IndVal index	$p < 0.05$
	Order	Genus		
1	Trichoptera	<i>Hydropsyche</i>	0.72	0.0001
	Plecoptera	<i>Perlodes</i>	0.62	0.0001
	Diptera	<i>Allotrichoma</i>	0.36	0.0035
	Plecoptera	<i>Halticoperla*</i>	0.35	0.0289
	Ephemeroptera	<i>Epeorus</i>	0.32	0.0488
2	Ephemeroptera	<i>Ecdyonurus</i>	0.73	0.0001
	Ephemeroptera	<i>Rhithrogena</i>	0.58	0.0008
	Ephemeroptera	<i>Ephemerella</i>	0.54	0.02
3	Amphipoda	<i>Gammarus</i>	0.62	0.0001
	Plecoptera	<i>Protonemura</i>	0.61	0.0003
	Diptera	<i>Simulium</i>	0.45	0.0023

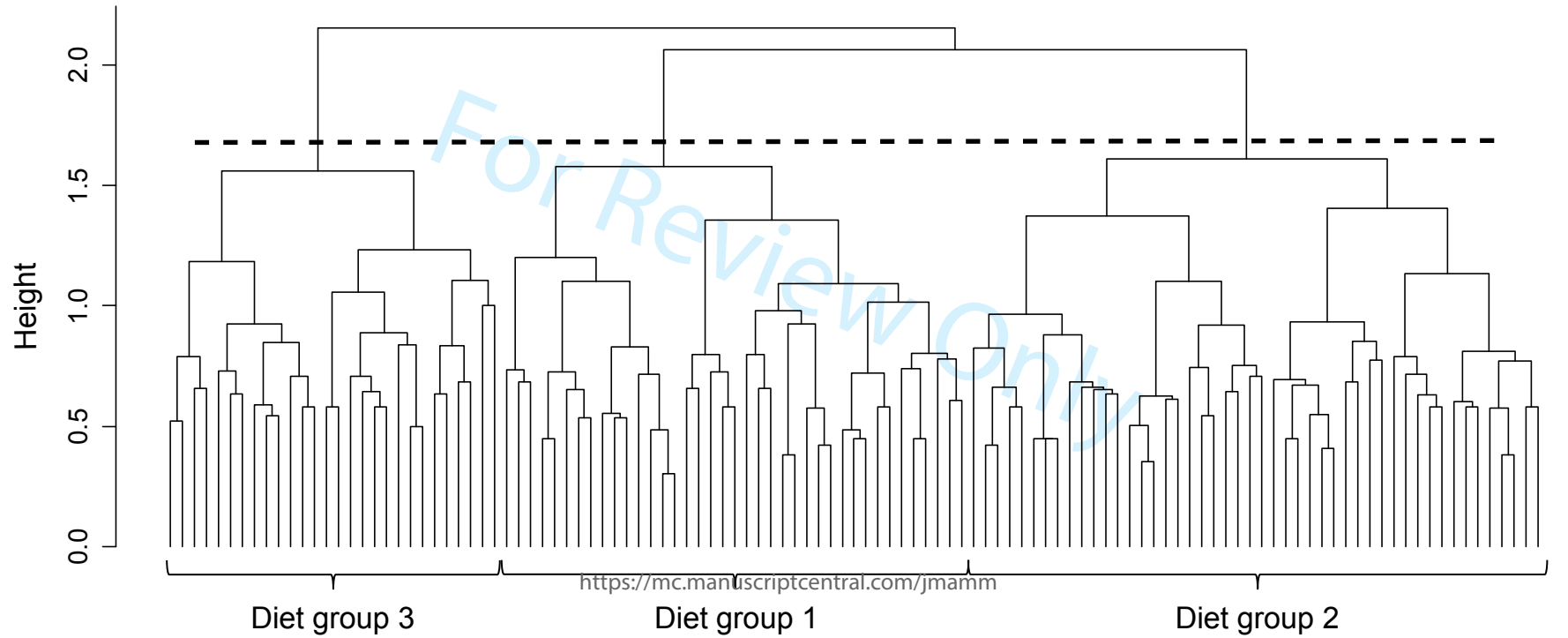
638 **Table 4.**— Mean and standard deviation (%) of the 12 environmental variables for the three
 639 summer diet groups of the Pyrenean desman in the French Pyrenees. See Table 1 for variables
 640 description.

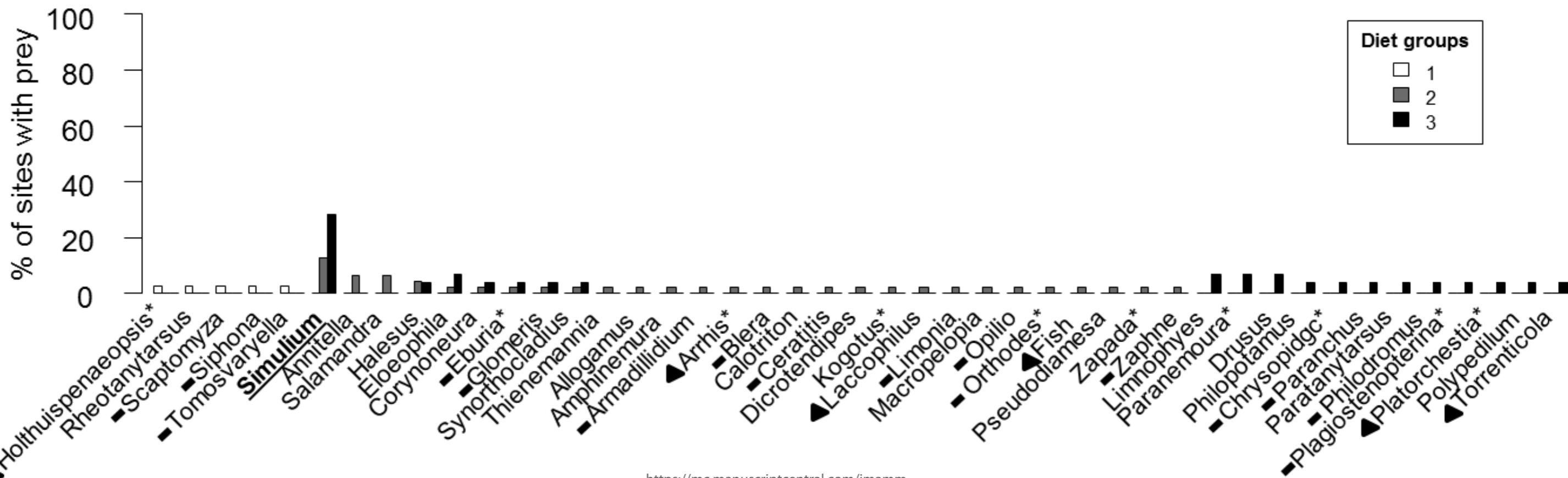
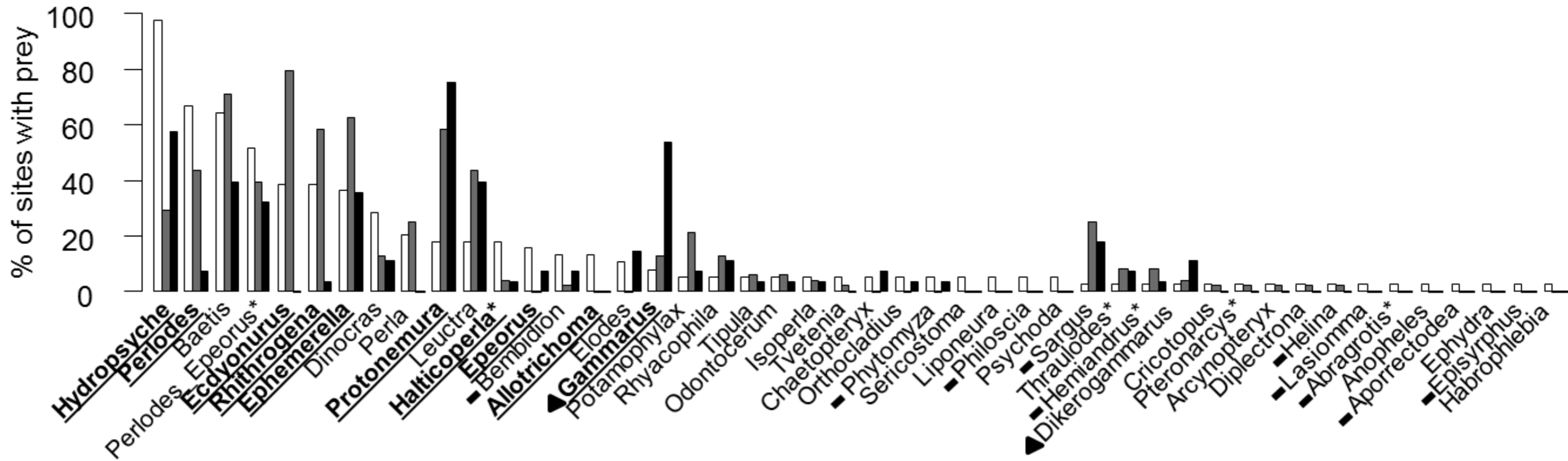
Environmental variables	Groups		
	1	2	3
SHELTER	74.42 ± 18.33	67.79 ± 17.16	59.42 ± 21.76
CLOG	11.89 ± 12.61	15.43 ± 16.79	19.71 ± 16.23
SHEET	4.81 ± 6.20	6.60 ± 11.08	4.64 ± 5.12
RUN	23.39 ± 16.04	20.75 ± 11.91	21.24 ± 12.77
RIFFLE	35.77 ± 23.52	37.26 ± 18.82	44.43 ± 22.60
BANKS	0.35 ± 0.25	0.47 ± 0.23	0.52 ± 0.28
WOOD	50.50 ± 23.03	44.51 ± 17.63	53.47 ± 17.74
HUM_IMP	28.79 ± 29.69	9.29 ± 17.49	19.79 ± 20.64
TEM	9.70 ± 1.93	8.94 ± 1.73	10.34 ± 1.48
FLO	1.32 ± 1.87	0.68 ± 0.83	1.27 ± 1.64
SLO	8.78 ± 8.35	10.79 ± 8.91	8.07 ± 10.54
TRI	3.31 ± 2.13	3.42 ± 1.93	2.68 ± 1.19

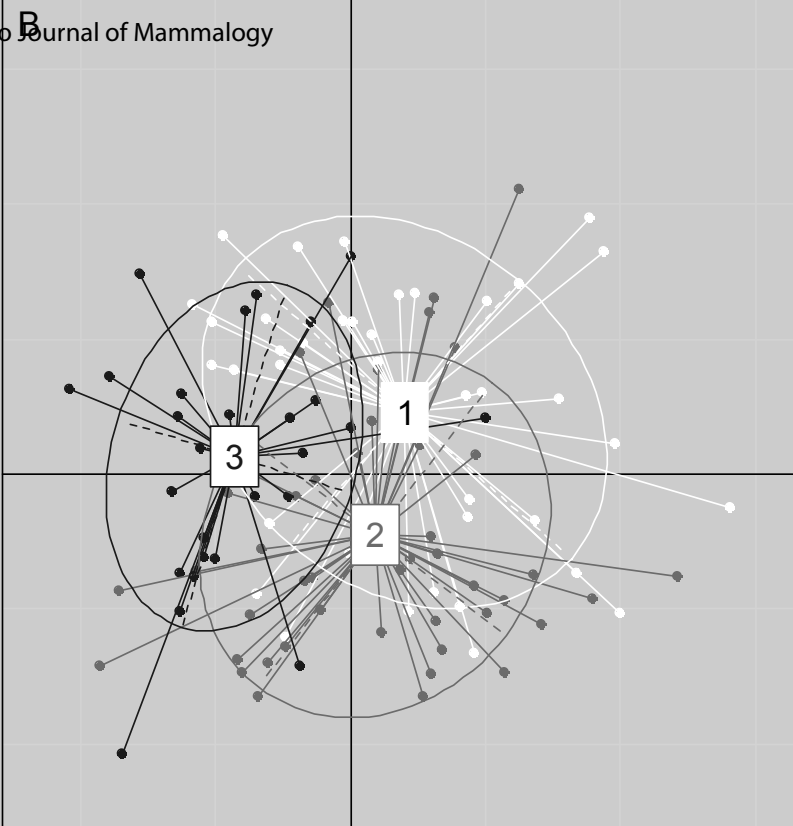
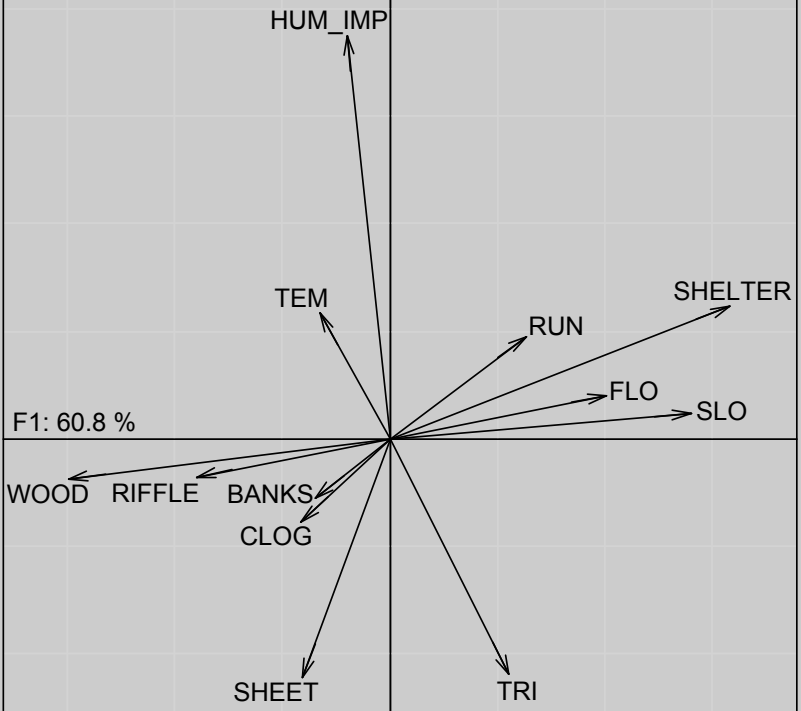
641



Cluster Dendrogram







Supplementary Data S1.— Complete list of taxa identified as Pyrenean desman prey by molecular analysis of 383 faeces collected in the French Pyrenees. Frequencies of occurrence of each prey (FO: % of faeces with taxa) are displayed. * indicates misidentified taxa whose distribution areas exclude the Pyrenees and France. Habitat type of taxa is given (aq: exclusively aquatic; aq/te: with aquatic and terrestrial stages; te: exclusively terrestrial).

Class	FO (%)	Order	FO (%)	Family	FO (%)	Genus	Habitat	FO (%)
Insecta	99.7	Ephemeroptera	86.7	Heptageniidae	59.0	<i>Rhithrogena</i>	aq/te	36.6
						<i>Ecdyonurus</i>	aq/te	35.3
						<i>Epeorus</i>	aq/te	9.1
						<i>Electrogena</i>	aq/te	0.5
				Baetidae	56.7	<i>Baetis</i>	aq/te	56.4
						<i>Acentrella</i>	aq/te	0.3
						<i>Ephemerella</i>	aq/te	37.1
				Ephemerellidae	37.1	<i>Thraulodes*</i>	aq/te	3.9
						<i>Habroleptoides</i>	aq/te	0.5
						<i>Habrophlebia</i>	aq/te	0.3
				Nemouridae	49.4	<i>Protonemura</i>	aq/te	47.3
						<i>Nemoura</i>	aq/te	1.8
						<i>Paranemoura*</i>	aq/te	1.3
						<i>Amphinemura</i>	aq/te	0.3
						<i>Zapada*</i>	aq/te	0.3
		Perlodidae	32.6	<i>Perlodes</i>	aq/te	30.6		
				<i>Isoperla</i>	aq/te	3.1		
				<i>Arcynopteryx</i>	aq/te	0.5		
		Plecoptera	84.9	Leuctridae	29.8	<i>Kogotus*</i>	aq/te	0.5
						<i>Leuctra</i>	aq/te	29.5
				Perlidae	25.9	<i>Perlomyia*</i>	aq/te	0.3
						<i>Perla</i>	aq/te	15.7
				Notonemouridae	5.5	<i>Dinocras</i>	aq/te	15.1
						<i>Halticoperla*</i>	aq/te	5.2
				Pteronarcyidae	5.2	<i>Spaniocerca*</i>	aq/te	0.3
						<i>Pteronarcys*</i>	aq/te	5.2
				Taeniopterygidae	2.6	<i>Brachyptera</i>	aq/te	2.1
<i>Taeniopteryx</i>	aq/te					0.5		
Trichoptera	64.0	Hydropsychidae	46.7	<i>Hydropsyche</i>	aq/te	46.2		
				<i>Diplectrona</i>	aq/te	1.3		
		Limnephilidae	19.1	<i>Potamophylax</i>	aq/te	8.1		
				<i>Chaetopteryx</i>	aq/te	3.4		
				<i>Allogamus</i>	aq/te	2.6		
				<i>Halesus</i>	aq/te	2.4		

		<i>Annitella</i>	aq/te	2.1		
		<i>Drusus</i>	aq/te	1.8		
Rhyacophilidae	6.5	<i>Rhyacophila</i>	aq/te	6.5		
Odontoceridae	3.4	<i>Odontocerum</i>	aq/te	3.4		
Sericostomatidae	2.1	<i>Sericostoma</i>	aq/te	2.1		
Philopotamidae	1.3	<i>Philopotamus</i>	aq/te	1.0		
		<i>Wormaldia</i>	aq/te	0.3		
Goeridae	0.3	<i>Goeracea</i> *	aq/te	0.3		
Uenoidae	0.3	<i>Thremma</i>	aq/te	0.3		
Simuliidae	15.4	<i>Simulium</i>	aq/te	14.9		
		<i>Stegopterna</i> *	aq/te	0.5		
		<i>Limnophyes</i>	aq/te	2.4		
		<i>Orthocladius</i>	aq/te	2.4		
		<i>Synorthocladius</i>	aq/te	1.6		
		<i>Cricotopus</i>	aq/te	1.0		
		<i>Tvetenia</i>	aq/te	1.0		
		<i>Brillia</i>	aq/te	0.8		
		<i>Corynoneura</i>	aq/te	0.8		
		<i>Polypedilum</i>	aq/te	0.8		
		<i>Thienemannia</i>	aq/te	0.8		
		<i>Diamesa</i>	aq/te	0.5		
		<i>Dicrotendipes</i>	aq/te	0.5		
		Chironomidae	14.6	<i>Microspectra</i>	aq/te	0.5
				<i>Smittia</i>	aq/te	0.5
				<i>Cardiocladius</i>	aq/te	0.3
<i>Conchapelopia</i>	aq/te			0.3		
<i>Macropelopia</i>	aq/te			0.3		
<i>Microtendipes</i>	aq/te			0.3		
<i>Parametriocnemus</i>	aq/te			0.3		
<i>Paratanytarsus</i>	aq/te			0.3		
<i>Paratrachocladius</i>	aq/te			0.3		
<i>Pseudodiamesa</i>	aq/te			0.3		
<i>Rheotanytarsus</i>	aq/te	0.3				
<i>Zavreliomyia</i>	aq/te	0.3				
Stratiomyidae	10.7	<i>Sargus</i>	te	10.7		
Tipulidae	4.7	<i>Tipula</i>	aq/te	4.7		
		<i>Allotrichoma</i>	aq/te	2.6		
		<i>Ephydra</i>	aq/te	0.8		
Ephydriidae	3.7	<i>Scatophila</i>	aq/te	0.3		
		<i>Lasiomma</i>	te	1.3		
Anthomyiidae	2.6	<i>Zaphne</i>	te	1.3		
		<i>Psychoda</i>	aq/te	2.6		
Psychodidae	2.6	<i>Psychoda</i>	aq/te	2.6		
Pediidae	1.8	<i>Dicranota</i>	aq/te	1.6		
		<i>Pedicia</i>	aq/te	0.3		
Agromyzidae	1.3	<i>Phytomyza</i>	te	1.3		
Limoniidae	1.3	<i>Eloeophila</i>	aq/te	1.0		

Diptera 45.7

				<i>Limonia</i>	te	0.3		
		Calliphoridae	0.8	<i>Lucilia</i>	te	0.8		
		Muscidae	0.8	<i>Helina</i>	te	0.5		
				<i>Thricops</i>	te	0.3		
		Platystomatidae	0.8	<i>Plagiostenoptera</i> *	te	0.8		
		Sciomyzidae	0.8	<i>Pteromicra</i>	aq/te	0.8		
				<i>Blera</i>	te	0.3		
				<i>Episyrphus</i>	te	0.3		
		Syrphidae	0.9	<i>Spilomyia</i>	te	0.3		
				<i>Euthera</i> *	te	0.3		
				<i>Siphona</i>	te	0.3		
		Tachinidae	0.8	<i>Tachinid</i> *	te	0.3		
				<i>Anastrepha</i> *	te	0.5		
		Tephritidae	0.8	<i>Ceratitis</i>	te	0.3		
		Blephariceridae	0.5	<i>Liponeura</i>	aq/te	0.5		
		Drosophilidae	0.5	<i>Scaptomyza</i>	te	0.5		
		Anisopodidae	0.3	<i>Sylvicola</i>	te	0.3		
		Athericidae	0.3	<i>Atherix</i>	aq/te	0.3		
		Chloropidae	0.3	<i>Epichlorops</i>	te	0.3		
		Culicidae	0.3	<i>Anopheles</i>	aq/te	0.3		
		Hybotidae	0.3	<i>Platypalpus</i>	te	0.3		
		Pipunculidae	0.3	<i>Tomosvaryella</i>	te	0.3		
		Sarcophagidae	0.3	<i>Sarcophaga</i>	te	0.3		
		Tabanidae	0.3	<i>Haematopota</i>	te	0.3		
Plecoptera_	30.8	Perlodidae_	30.8	<i>Perlodes_Epeorus</i> *	aq/te	30.8		
Ephemeroptera		Heptageniidae						
		Carabidae	7.1	<i>Bembidion</i>	te	6.5		
				<i>Abax</i>	te	0.3		
				<i>Paranchus</i>	te	0.3		
Coleoptera	14.1	Scirtidae	6.3	<i>Elodes</i>	aq/te	6.3		
		Cerambycidae	0.5	<i>Eburia</i> *	te	0.5		
		Dryopidae	0.3	<i>Dryops</i>	aq/te	0.3		
		Dytiscidae	0.3	<i>Laccophilus</i>	aq	0.3		
		Staphylinidae	0.3	<i>Lesteva</i>	te	0.3		
Orthoptera	6.5	Anostomatidae	6.5	<i>Hemiandrus</i> *	te	6.5		
Neuroptera	1.8	Chrysopidae	1.8	<i>Chrysopidgc</i> *	te	1.8		
Hemiptera	1.3	Aphididae	0.5	<i>Rhopalosiphum</i>	te	0.5		
		Pentatomidae	0.5	<i>Pentatoma</i>	te	0.5		
		Cercopidae	0.3	<i>Philaenus</i>	te	0.3		
Lepidoptera	1.0	Geometridae	0.3	<i>Lycia</i>	te	0.3		
		Noctuidae	0.8	<i>Abragrotis</i> *	te	0.3		
				<i>Lacinipolia</i> *	te	0.3		
				<i>Orthodes</i> *	te	0.3		
Odonata	0.5	Calopterygidae	0.3	<i>Calopteryx</i>	aq/te	0.3		
		Gomphidae	0.3	<i>Ophiogomphus</i>	aq/te	0.3		
Hymenoptera	0.3	Formicidae	0.3	<i>Lasius</i>	te	0.3		
Malacostraca	18.0	Amphipoda	16.7	Gammaridae	16.5	<i>Gammarus</i>	aq	14.6

					<i>Dikerogammarus</i>	aq	2.6
				Oedicerotidae	0.5	<i>Arrhis</i> *	aq 0.5
				Talitridae	0.3	<i>Platorchestia</i> *	aq 0.3
				Philosciidae	0.5	<i>Philoscia</i>	te 0.5
				Armadillidiidae	0.3	<i>Armadillidium</i>	te 0.3
				Oniscidae	0.3	<i>Oniscus</i>	te 0.3
				Porcellionidae	0.3	<i>Porcellio</i>	te 0.3
				Scyphacidae	0.3	<i>Haloniscus</i> *	aq 0.3
				Decapoda	0.3	Penaeidae	0.3
						<i>Holthuispenaeopsis</i> *	aq 0.3
				Glomerida	2.4	Glomeridae	2.1
						<i>Glomeris</i>	te 2.1
						Glomeridellidae	0.3
						<i>Glomeridella</i>	te 0.3
				Polydesmida	0.8	Polydesmidae	0.8
						<i>Polydesmus</i>	te 0.8
				Julida	0.5	Julidae	0.5
						<i>Allajulus</i>	te 0.3
						<i>Pteridoiulus</i> *	te 0.3
				Chordeumatida	0.3	Craspedosomatidae	0.3
						<i>Ochogona</i>	te 0.3
				Lissamphibia	2.9	Urodeles	2.9
						Salamandridae	2.9
						<i>Calotriton</i>	aq/te 1.8
						<i>Salamandra</i>	aq/te 1.0
				Araneae	0.8	Philodromidae	0.5
						<i>Philodromus</i>	te 0.5
						Tetragnathidae	0.3
						<i>Metellina</i>	te 0.3
				Opiliones	0.5	Phalangiidae	0.5
						<i>Opilio</i>	te 0.5
				Sarcoptiformes	0.5	Ceratozetidae	0.3
						<i>Fuscozetes</i>	te 0.3
						Oribatulidae	0.3
						<i>Zygoribatula</i>	te 0.3
				Trombidiformes	0.5	Torrenticolidae	0.5
						<i>Torrenticola</i>	aq 0.5
				Clitellata	0.5	Haplotaxida	0.5
						Lumbricidae	0.3
						<i>Aporrectodea</i>	te 0.3
						Megascolecidae	0.3
						<i>Metaphire</i>	te 0.3
				Collembola	0.3	Entomobryomorpha	0.3
						Tomoceridae	0.3
						<i>Tomocerus</i>	te 0.3
				Eurotatoria	0.3	Bdelloida	0.3
						Habrotrochidae	0.3
						<i>Habrotrocha</i>	aq 0.3
				Eutardigrada	0.3	Parachela	0.3
						Isohypsibiidae	0.3
						<i>Thulinus</i> *	aq 0.3
				Gastropoda	0.3	Littorinimorpha	0.3
						Tateidae	0.3
						<i>Potamopyrgus</i>	aq 0.3
				Unidentified fish			aq 0.5