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SUMMARY

The retrieval of odor-related memories shapes animal behavior. The anterior piriform cortex (aPC) is the largest part of the olfactory cortex and it plays important roles in olfactory processing and memory. However, it is still unclear whether specific cellular mechanisms in the aPC control olfactory memory depending on the appetitive or aversive nature of the stimuli involved. Cannabinoid-type 1 (CB1) receptors are present in the aPC (aPC-CB1), but their potential impact on olfactory memory was never explored. Here, we used a combination of behavioral, genetic, anatomical and electrophysiological approaches to characterize the functions of aPC-CB1 receptors in the regulation of appetitive and aversive olfactory memory. Pharmacological blockade or genetic deletion of aPC-CB1 receptors specifically impaired the retrieval of conditioned odor preference (COP). Interestingly, expression of conditioned odor aversion (COA) was unaffected by local CB1 receptor blockade, indicating that the role of aPC endocannabinoid signaling is selective for retrieval of appetitive memory. Anatomical investigations revealed that CB1 receptors are highly expressed on aPC GABAergic interneurons, and *ex vivo* electrophysiological recordings showed that their pharmacological activation reduces miniature inhibitory currents (mIPSCs) onto aPC semilunar (SL), but not pyramidal principal neurons. COP retrieval, but not COA, was associated with a specific CB1 receptor-dependent decrease of mIPSCs in SL cells. Altogether, these data indicate that aPC-CB1 receptor-dependent mechanisms physiologically control the retrieval of olfactory memory depending on odor valence and engaging modulation of local inhibitory transmission.

Keywords: anterior piriform cortex; CB1 receptors; conditioned odor preference; conditioned odor aversion; neuroanatomy; miniature inhibitory currents (mIPSCs); semilunar neurons; pyramidal neurons.

INTRODUCTION

Chemosensory information is crucial for the survival of humans and other animals. For example, a large part of animal behavior including control of emotional states, food intake and social interactions relies on the capacity to perceive odor information and to retrieve its potential meaning based on previous experiences [1,2]. Olfactory perception starts when odorant molecules, travelling through orthonasal or retronasal pathways, reach olfactory receptors on sensory neurons located in the olfactory epithelium [3]. These neurons project to the olfactory bulb that in turn transmits the signal to other brain regions, including the anterior piriform cortex (aPC) [4,5], which plays a key role in olfactory processing and memory [6–8]. However, the specific cellular mechanisms governing odor information storage and retrieval in the aPC are still unclear.

Cannabinoid type-1 (CB1) receptors together with their endogenous ligands (endocannabinoids) form the core of the so-called endocannabinoid system (ECS) in the brain [9], which is an important modulator of many functions, including learning and memory [10,11]. Activation of presynaptic CB1 receptors is well-known to physiologically control the release of several neurotransmitters in many brain regions [12,13]. CB1 receptors are present in different olfactory structures [14], where they can modulate olfactory processes [15–19]. However, little is known about the specific impact of CB1 receptor signaling in olfactory brain structures on odor-dependent memory functions.

Considering that the aPC is an important region for olfactory memory [6–8,20–22], we hypothesized that CB1 receptors in the aPC (aPC-CB1) could modulate odor-related memory processes. Our data show that aPC-CB1 receptors are specifically

required for the expression of appetitive, but not aversive, olfactory memory, and they are involved in the direct control of the associated modulation of local inhibitory circuits. Altogether, these results indicate that the physiological activation of CB1 receptors in the aPC exerts a fine-tuned regulation of olfactory circuits and functionally discriminates the retrieval of positively- and negatively-motivated olfactory memories.

RESULTS

CB1 receptors in the aPC are necessary for the retrieval of conditioned odor preference

To investigate the potential impact of CB1 receptor signaling in odor-related memory, we set up a behavioral protocol to assess conditioned odor preference (COP) in mice (see STAR Methods, Figures 1A and S1A,B) [23,24]. Using this protocol, mice displayed a reliable preference for the odor-scented solution previously associated to sucrose (C+) as compared to the other one (C-), revealing the formation of COP (Figure S1C), regardless of the odor used as C+ (Figures S1D,E).

We next investigated the role of CB1 receptors in the aPC (aPC-CB1) during COP. Specific deletion of the *CB1* gene in the aPC (aPC-*CB1*-KO, see STAR Methods, Figures 1B,C and S1F-I) [25–27] abolished the preference for the C+ solution during test (Figure 1D), without altering sucrose preference upon training (Figure S1J), nor total liquid intake (Figure S1K), and independently from the anatomical extension of the CB1 deletion (Figure S1L). These results indicate that COP requires aPC-CB1 receptors. To determine the specific role of CB1 receptor signaling in the different phases of the COP protocol, we acutely injected the CB1 receptor antagonist AM251 into the aPC (4µg/0.5µl per side; Figures S2A-E) prior to each odor-sucrose pairing

or before the retrieval test (Figure 1A). Neither consumption during training, nor COP performance were affected by aPC-CB1 blockade before each pairing (AM pairings; Figures 1D, S1J,K and S2F). Conversely, AM251 acutely injected into the aPC prior to the retrieval test abolished COP, without altering total liquid consumption (AM test; Figures 1D, S1J,K and S2F). Acute blockade of aPC-CB1 receptors might permanently impair COP retrieval. On the other hand, longer training might render COP retrieval independent of aPC-CB1 receptors. To simultaneously test for these possibilities, animals previously treated with AM251 or vehicle received 4 additional odor-sucrose pairings (see STAR Methods) and were injected with vehicle ["Veh(AM) test"] or AM251 ["AM(Veh) test"] before the second retrieval test, respectively. In these conditions, vehicle-treated mice displayed clear COP, whereas AM251 blocked this behavior (Figures 1E and S2G). aPC-CB1 receptors might control expression of preference independently of previous learning. However, aPC-CB1 blockade did not impair innate sucrose preference (Figures S3A-C).

Altogether, these results indicate that endogenous activation of aPC-CB1 is specifically required during retrieval of COP, without affecting innate responses to attractive stimuli.

CB1 receptors in the aPC are not involved in the retrieval of conditioned odor aversion

We next asked whether aPC-CB1 receptors are also involved in the retrieval of conditioned odor aversion (COA) induced by lithium chloride (LiCl) injections (Figure 2A, COA-lithium, COA-L) [25,28–30]. Notably, AM251 acutely injected into the aPC did not impair COA-L expression (Figures 2A,B, S2A-F and S3D), suggesting that aPC-CB1 receptors are dispensable for the retrieval of negatively motivated olfactory memory. However, the differential effects of CB1 receptor blockade between COP

and COA-L could be due to the different types of associations involved (sensory-sensory *versus* sensory-gastric). Using a sensory-sensory COA protocol, where sucrose was substituted by the aversive taste quinine (COA-Q; Figures 2C and S3E) [31,32], mice treated with vehicle or the CB1 receptor antagonist before the retrieval test displayed the same avoidance towards the conditioned odor (Figures 2D, S2F and S3F).

Altogether, these data indicate that aPC-CB1 receptor signaling is necessary for the retrieval of COP but not of COA, suggesting that expression of acquired odor choices rely on different mechanisms depending on the valence of the unconditioned stimulus.

CB1 receptors are highly expressed in GABAergic interneurons in the aPC

As previously reported [14,33], fluorescent immunohistochemistry revealed that CB1 receptor protein is highly expressed in layer II of the aPC (Figure 3A), where the aPC principal neurons are mainly localized [34,35]. To detail the cellular distribution of CB1 receptors, we analyzed aPC tissues from conditional mutant mice carrying exclusive expression of the protein in identified specific cell types (Rescue mice) [33,36,37]. A similar pattern of CB1 receptor immunoreactivity was observed across aPC tissues from WT mice and global CB1-Rescue mice (Figure 3A) [33,37], but also from mice with specific re-expression in GABAergic neurons (GABA-CB1 Rescue; Figure 3A) [33,36]. In contrast, the immunoreactivity was extremely low in mice re-expressing the receptor only in cortical glutamatergic neurons (Glu-CB1 Rescue; Figure 3A) [37] and, as expected, was undetectable in mice where CB1 receptor expression is absent (CB1 Stop; Figure 3A) [33,36,37]. To better characterize the expression of CB1 receptors at the synaptic level, we next used immunogold electron microscopy (Figure 3B). As expected, CB1 receptor immunogold particles were

specifically present in different amounts at many cellular locations (Figures 3B and S4). Amongst CB1 receptor particles located at terminals (1856 over 3409 total counted in WT; Figures 3B and S4), approximately 88% and 12% were at symmetric (presumably inhibitory) and asymmetric synapses (presumably excitatory), respectively (Figures 3B,C). Moreover, whereas only 23% of excitatory terminals were labeled with CB1 receptor immunogold particles, this percentage was as high as 87% in inhibitory ones (Figures 3B,D). As expected, only background staining was detected in sections from *CB1*-KO mice (18 particles at terminals; Figures 3B,D and S4).

To identify the specific topographical distribution of CB1-expressing cells within the aPC, we used double Fluorescent *In Situ* Hybridization (D-FISH) to label the mRNAs of CB1 receptor and of glutamic acid decarboxylase 65KDa (GAD), a marker of GABAergic neurons (Figure 4A). Consistent with previous studies [38], the majority of GAD-expressing cells were observed in deep layer III (Figures 4A,B). CB1 mRNA was also highly expressed in layer III, with scattered positive cells in layer I (Figures 4A,C). Accordingly, counting of positive cells revealed that a high proportion of GAD+ neurons contained also CB1 mRNA (63%; Figures 4A,D), following the distribution of GAD across layers (Figures 4A,B,E). Similarly to other cortical regions such as the hippocampus [39], CB1 mRNA was expressed at very different levels across CB1-positive aPC cells. Whereas a majority of cells expressed low-to-moderate amounts of the transcript, scattered cells contained very high levels of CB1 mRNA, especially in layers II and III (Figure 4A). Interestingly, virtually 100% of these high CB1-expressing neurons co-expressed GAD mRNA (Figure 4F). Conversely, low CB1-expressing neurons were virtually all identified as GAD-positive in layer I, but this proportion was reduced in layers II and III (Figure 4F).

Altogether, these data indicate that the large majority of aPC-CB1 receptor protein is expressed in terminals of local GABAergic neurons, with layer-specific topographical distribution.

CB1 receptors control inhibitory transmission in the aPC

To start addressing the so-far unexplored cannabinoid-dependent control of inhibitory transmission in the aPC, we first determined by *ex vivo* patch-clamp electrophysiological recordings the impact of CB1 receptor activation on GABAergic neurotransmission impinging onto specific populations of principal cells of the aPC (semilunar-like neurons, SL, and pyramidal-like neurons, PNs; Figures S5A-B and see STAR Methods). These cells receive many inputs from the olfactory bulb and other brain regions, project to other olfactory cortical areas [35,40,41] and are extensively innervated by local inhibitory interneurons [38,42,43]. Thus, we reasoned that selective modulation of GABAergic inputs onto SL cells and/or PNs might represent a way through which CB1 receptors rapidly regulate principal cell activity and consequently odor processing. To address this possibility, we recorded miniature inhibitory post-synaptic currents (mIPSCs) representing the global inhibitory inputs of principal cells. These events occur at a frequency of 2.06 ± 0.23 Hz and an amplitude of 76.29 ± 4.41 pA (Vehicle; Figures 5A-C) in SL cells, and at a frequency of 2.50 ± 0.39 Hz and an amplitude of 64.06 ± 4.27 pA in PNs of naïve animals (Vehicle; Figures 5D-F). Similarly to what observed in the hippocampus [44], the application of the CB1 receptor agonist WIN 55,212-2 (WIN, 5μM) significantly reduced the frequency of mIPSCs in SL cells, with only a slight impact, if any, on their amplitude (Figures 5A-C), suggesting a presynaptic inhibitory effect. This decrease was fully reversed by the application of the CB1 receptor antagonist AM251 (4μM; WIN+AM251; Figures 5A-C). Conversely and surprisingly, no such effects were

observed in PNs (Figures 5D-F), indicating a cell-type specific impact of CB1 receptor-dependent control of inhibitory currents in naïve animals.

In summary, CB1 receptors are highly expressed in GABAergic interneurons of the aPC, and their activation results in the modulation of inhibitory inputs onto SL principal neurons.

COP retrieval reduces mIPSCs in the aPC *via* presynaptic CB1 receptors

We next hypothesized that retrieval of COP might be associated with aPC-CB1 receptor-dependent modulation of principal cells' mIPSCs. A significant reduction of mIPSCs frequency (~26%) was found in SL cells from mice sacrificed during COP retrieval, as compared to a control group exposed to the same number of only water-drinking sessions (Water; Figures 6A,B). SL cells from mice receiving the same number of “training” sessions, but without the presence of sucrose (Sucrose free) or odor (Odor free) did not display any reduction of mIPSCs frequency (Figure 6B). Considering that aPC-CB1 receptors regulate COP but not COA retrieval (Figure 1 and Figure 2), we next evaluated the effect of COA-Q retrieval and we found that this condition did not affect mIPSCs frequency (Figure 6B). Notably, no differences in amplitudes were observed across the groups (Figure S6A). Altogether, these results indicate that COP retrieval, but not exposure to odor or sucrose alone or COA retrieval, is associated with a specific reduction of presynaptic inhibitory transmission onto SL neurons in the aPC.

As local blockade, the systemic injection of the CB1 receptor antagonist Rimonabant (Rim, 1 mg/kg) impaired COP retrieval (Figures 6C and S6B), independently of total liquid consumption (Figures S6C-F). As expected, systemic injection of vehicle before COP retrieval did not alter the associated reduction of mIPSCs frequency in SL cells

(Figures 6D,E; $p>0.8$, as compared to Figure 6B). Conversely, systemic administration of Rim abolished this decrease up to levels undistinguishable from control mice (Figures 6D,E; $p>0.8$, as compared to Figure 6B), with no effect on amplitude (Figure S6G). Notably, mIPSC frequencies, but not amplitudes, of individual animals were inversely correlated with the COP retrieval performance (Figure 6F and data not shown), suggesting that the level of presynaptic inhibition of SL cells is linked to the behavioral retrieval of COP. Next, we examined the impact of COP retrieval on mIPSCs of PNs.

Similarly to SL neurons, the frequency of mIPSCs was reduced (~33%) in PNs of animals undergoing COP retrieval, as compared to water control mice (Figures 6G-H) with no change in amplitude (Figure S6H). A slight non-significant increase of mIPSCs amplitude was observed in PNs from mice receiving systemic injection of Rim prior COP retrieval (Figure S6H). However, this treatment was not able to reverse the COP retrieval-associated reduction of mIPSCs frequency in PNs (Figures 6G-H). No correlations between COP retrieval and the levels of mIPSC frequencies or amplitudes of PNs in individual mice were observed (Figure 6I and data not shown).

Altogether, these results indicate that COP retrieval is associated to a reduction of inhibitory inputs on both SL cells and PNs. However, presynaptic CB1 receptors appear to regulate inhibitory transmission in the aPC in a cell type-specific manner, thereby providing an unforeseen fine-tuned modulation of olfactory memory circuits, likely contributing to appropriate behavioral responses.

Acute blockade of CB1 receptors affects COP retrieval through GABAergic neurons

We then addressed the potential involvement of CB1 receptors expressed in inhibitory neurons (GABAergic CB1) in COP retrieval. Surprisingly, mice lacking *CB1* gene expression from GABAergic neurons (GABA-*CB1*-KO) [27,45] did not display any alteration of COP retrieval as compared to control wild-type littermates (GABA-*CB1*-WT; Figure S6I). This negative result might suggest that CB1 receptors in GABAergic neurons are not necessary for acute COP retrieval. However, GABA-*CB1*-KO mice carry a deletion of CB1 receptors in all GABAergic cells of the whole forebrain, starting from early developmental stages [27,46]. Such diffuse and long-lasting absence of CB1 receptor signaling might induce opposing effects in different brain regions and/or stimulate developmental compensatory phenomena, whose general mechanisms recently started to be elucidated [47,48]. If occurring in GABA-*CB1*-KO mice, these mechanisms might mask the specific acute role of CB1 receptors signaling in GABAergic neurons in COP retrieval during adulthood. Considering this possibility, we adopted an alternative strategy to investigate the specific role of CB1 receptors in GABAergic neurons, by testing whether the blockade of COP retrieval by the acute administration of Rim was still effective in GABA-*CB1*-KO mice. A systemic acute injection of the drug blocked COP retrieval in GABA-*CB1*-WT, but it failed to impair this behavior in GABA-*CB1*-KO mice (Figures 6J,K and S6J-L), indicating that CB1 receptors in GABAergic neurons are required for the COP retrieval-disrupting effect of the acute pharmacological blockade of CB1 receptor signaling.

DISCUSSION

In this study, we functionally characterized the presence and the role of CB1 receptors in the aPC. We found that these receptors are specifically involved in the retrieval of appetitive, but not aversive, olfactory memory and in the associated modulation of inhibitory transmission onto specific aPC principal cells. Moreover, our data show that the retrieval impairment of appetitive olfactory memory induced by CB1 antagonism requires CB1 receptors expressed in inhibitory neurons.

aPC-CB1 receptors are necessary for the retrieval of appetitive but not aversive olfactory memory

In other brain structures, CB1 receptors have been reported to play crucial roles in different phases of learning and memory processes [10,11]. Our data reveal that the endogenous activation of aPC-CB1 receptors is necessary for COP retrieval, but it is dispensable for its acquisition, thereby enlarging the spectrum of CB1 receptor involvement in different phases of learning and memory. Interestingly, it was recently found that a small enhancement of hippocampal GABAergic inhibition blocked cell firing and memory retrieval but left memory encoding intact [49]. This nicely parallels our results showing that a small, but CB1 receptor- and cell type-dependent reduction in GABAergic inhibition in the aPC, is associated with effective COP memory retrieval. We here propose that CB1 receptors on GABAergic axon terminals might be essential to reduce inhibition onto SL cells, thereby permitting cell firing in the ensemble population to retrieve the COP memory trace. In this context, it is interesting to note that COP retrieval is also associated with a similar reduction of inhibitory drive onto PNs, but in a CB1 receptor-independent manner. This suggests

that cell type-specific mechanisms are involved in the coordinated regulation of the activity of distinct aPC principal neurons during retrieval of olfactory memory.

Our data show that the retrieval of aversive olfactory memory is independent of aPC-CB1 receptor signaling. These intriguing results might be explained by two possibilities: either COA depends on aPC but it does not involve CB1 receptor signaling, or COA does not depend on aPC. There is currently no clear answer to this question, but some pieces of evidence seem to indicate a certain level of specialization of the aPC for positively motivated olfactory memory. Optogenetic or chemogenetic manipulations of selected neurons in the PC can modulate both aversive and appetitive behavioral responses [50,51]. Despite the fact that specific aPC activity was observed during COA retrieval [52], olfactory cues associated with sucrose activate more aPC neurons than odors associated with quinine [53] and aPC lesions impair appetitive but not aversive odor-related memory [21]. In addition, CB1 receptor signaling can mediate aversive olfactory memory in other brain regions. For instance, Laviolette and Grace (2006) showed that CB1 receptors in the medial prefrontal cortex are required for odor-dependent fear conditioning [54], and we recently demonstrated that deletion of the *CB1* gene specifically in medial habenular neurons selectively abolishes COA, but not COP [30]. Moreover, it has been shown that the basolateral nucleus of the amygdala (BLA), which is essential for COA [31,55], is more strongly connected with the posterior PC (pPC) than with the aPC [56], and BLA-pPC interactions are important for aversive odor conditioning [57]. Conversely, the aPC is more densely connected than the pPC to the olfactory tubercle, which, by receiving intense dopaminergic inputs, might be specifically involved in the processing of reward-related information [58]. Altogether, this suggests a potential double dissociation in the roles of aPC and pPC in COP and

COA, with the aPC being somehow specialized in processing positive acquired values of odors and the pPC more involved in aversive odor memory.

CB1 receptors are highly expressed in aPC GABAergic interneurons and regulate local inhibitory neurotransmission

Our immunohistochemical, D-FISH and electron microscopy data show that CB1 receptors are found in a high proportion of GABAergic neurons located in the three layers of the aPC and that they are strongly expressed at inhibitory synaptic terminals. Moreover, pharmacological activation of aPC-CB1 receptors decreases miniature inhibitory currents frequency specifically in SL cells. Similarly to the hippocampus [39], cells expressing high levels of CB1 mRNA are exclusively GABAergic interneurons. Recent evidence points to the presence of long-range inhibitory neurons as a novel neuroanatomical and functional feature in cortical areas [59]. These putative long-range inputs to the aPC might contain CB1 receptors, but our viral manipulations exclude their participation in the CB1 receptor-dependent retrieval of COP. Interestingly, depending on the layer, a portion of low CB1-expressing cells do not co-express GAD mRNA and are presumably glutamatergic neurons. Indeed, CB1 receptor protein is abundantly present in the main olfactory bulb at terminals of glutamatergic centrifugal fibers coming from principal neurons of the anterior olfactory nucleus and aPC [18]. Importantly, as these "glutamatergic" CB1 receptors play a key role in the control of olfactory perception and food intake [18], we cannot fully exclude that alterations in olfactory perception might participate in the phenotype of aPC-*CB1*-KO mice. However, intra-aPC pharmacological manipulations indicate that local CB1 receptor signaling is necessary for COP retrieval, but it is dispensable for expression of COA. Therefore, any putative impairment of olfactory perception induced by deletion of the *CB1* gene in projecting

glutamatergic neurons of the aPC is unlikely to be responsible for the phenotype of aPC-*CB1*-KO mice. Nevertheless, our electron microscopy immunogold results indicate that a small proportion of CB1 receptors are specifically present at glutamatergic terminals within the aPC. Moreover, a consistent portion of CB1 receptors appears to be located outside of terminals. Future studies will investigate the origin and the potential roles of these aPC subpopulations of CB1 receptors, which might still have functional significance.

COP retrieval is associated with CB1 receptor-dependent modulation of inhibitory transmission on specific aPC principal cells

An approximate 30% decrease of mIPSCs frequency recorded in both aPC SL cells and PNs was observed in slices from mice undergoing COP retrieval. In PNs, CB1 receptor antagonism did not affect COP-dependent frequency decrease. Conversely, the same treatment fully reversed mIPSCs frequency in SL cells up to the same levels of mice exposed to water alone, odor alone, sucrose alone or COA. Together with the fact that mIPSCs amplitude was not affected and that frequency values were inversely correlated with behavioral performance, these results indicate that COP retrieval is likely associated with presynaptic reduction of inhibitory transmission onto aPC SL cells. This idea is reinforced by the fact that the COP retrieval impairment under pharmacological CB1 receptor blockade is absent in mice lacking CB1 receptors from forebrain GABAergic neurons.

Hence, these results suggest that aPC-CB1 receptors control the behavioral responses induced by appetitive olfactory memory by regulating cell type-specific inhibitory transmission. More generally, they imply a dissociation between the roles of SL cells and PNs in the processing of olfactory information that will be very interesting to study in deeper details.

Interestingly, inhibitory circuits within the aPC have been shown to be strongly recruited in olfactory-dependent processes [43,60–62]. For instance, *in vivo* odor exposure widely activates GABAergic interneurons in the aPC [43], potentially participating in the processing of odors and their meaning [63]. In this context, the spatially restricted functions of CB1 receptors (i.e. on SL cells but not on PNs) suggest that CB1 receptor-dependent processes selectively tune the excitability of specific aPC principal neurons during COP retrieval. These processes would, in turn, refine the response to positively conditioned odor stimulations, eventually allowing the precise "funneling" of behavior towards preference responses.

In conclusion, this study provides a first characterization of the functional role of CB1 receptor signaling in aPC circuitry and related behaviors, thereby contributing to a better understanding of how the aPC participates in specific memory functions.

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AUTHOR CONTRIBUTIONS

G.T., G.F., and G.M. designed research; G.T., M.J., S.A., A.C., L.B., N.P., E.S-G., performed research; G.T., A.B-G., F.M., P.G., G.F. and G.M. supervised research; G.T.,S.A. and I.B-D.R analyzed data; G.T., G.F., and G.M. wrote the manuscript. All authors edited and approved the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

MAIN-TEXT FIGURE LEGENDS

Figure 1. CB1 receptors in the aPC are necessary for retrieval of odor preference

(A) Schematic protocol used for conditioned odor preference (COP). During conditioning, the two odor-scented solutions (O1 and O2) are associated to the absence or presence of sucrose, becoming neutral (C-) or conditioned (C+) stimuli, respectively. Blue arrows, aPC infusions before pairings; red arrow, aPC infusion before test for pharmacological experiments. **(B)** Representative images of Fluorescent *In Situ* Hybridization against CB1 mRNA (red), showing the virally induced deletion of CB1 receptors in the aPC of *CB1*-Flox mice locally injected with

AAV-GFP or AAV-CRE. Scale bar, 200µm. **(C)** Quantification of fluorescence intensity of CB1 mRNA in the insular cortex, the aPC and the lateral olfactory tubercle from sections where maximal deletion of CB1 expression was observed in aPC-*CB1*-KO and from equally located sections in control mice (n=7-10). **(D)** Consumption of C+ and C- odor-scented solutions in control mice (Control, n=36), mice carrying deletion of aPC-CB1 receptors (aPC-*CB1*-KO, n=10), mice receiving aPC infusions of the CB1 receptor antagonist AM251 (4µg/0.5µl per side) before each odor-sucrose pairing (AM pairings, n=8), or before the COP retrieval test (AM test, n=12). **(E)** Consumption of C+ and C- solutions during a second COP retrieval test performed after retraining (see STAR Methods). Mice previously infused with AM were infused with vehicle before the second test ["Veh(AM) test", n=12] and those previously infused with vehicle were now infused with AM ["AM(Veh) test", n=13]. **, $p<0.01$, ***, $p<0.001$; ****, $p<0.0001$; ns, not significant. For statistical details, see Tables S1-S3. For supplemental information, see Figures S1 and S2.

Figure 2. CB1 receptors in the aPC are not involved in the retrieval of odor aversion

(A) Schematic representation of the protocol used for LiCl-induced conditioned odor aversion (COA-L). **(B)** Consumption of the odor-scented solutions (C+ and C-) during test of COA-L in mice receiving aPC infusions of the CB1 receptor antagonist AM251 (4µg/0.5µl per side, n=11) or vehicle (n=13). **(C)** Schematic representation of the protocol used for quinine-induced conditioned odor aversion (COA-Q). **(D)** Consumption of the odor-scented solution during test of COA-Q in mice receiving aPC infusions of AM251 (n=12) or vehicle (n=11). C+, odor-scented solutions previously paired with LiCl injections **(B)** or quinine **(D)**. C-, odor-scented solutions paired with saline injections **(B)** or water **(D)**. Red arrows, time of intra-aPC infusions. ****, $p<0.0001$ general solution effect. For statistical details, see Tables S1-S3. For supplemental information, see Figures S2 and S3.

Figure 3. CB1 receptors are highly expressed in GABAergic interneurons in the aPC

(A) Representative coronal brain sections showing immunostaining of CB1 receptors in the aPC of wild-type (WT), CB1 Rescue, GABA-CB1 Rescue, Glu-CB1 Rescue and CB1 Stop mice. Dotted lines delimitate the different cortical layers (I, II and III). Scale bars, 100µm. **(B)** Electron microscopy micrographs of immunogold staining for CB1 receptors in the aPC of CB1 wild-type (*CB1*-WT) and knockout mice (*CB1*-KO). Black arrowheads, synapses; sp, spines; den, dendrites; red areas, presumably inhibitory terminals and preterminals; green areas, presumably excitatory terminals; purple areas, mitochondria; red arrows, CB1 receptors on inhibitory terminals and pre-terminals; green arrows, CB1 receptors on excitatory terminals; purple arrows, CB1 receptors on mitochondria. Scale bars, 1µm. **(C)** Proportion of CB1 receptor immunoparticles on inhibitory and excitatory terminals over total CB1 labeling on terminals (100 %). **(D)** Percentage of CB1 receptor-labeled inhibitory and excitatory terminals in *CB1*-WT and *CB1*-KO. ****, $p<0.0001$. For statistical details, see Tables S1-S3. For supplemental information, see Figure S4.

Figure 4. Topographic distribution of CB1 receptor-positive GABAergic interneurons in the aPC

(A) Representative images showing double Fluorescent *In Situ* Hybridization (D-FISH) of GAD 65kDa mRNA (GAD, red) and CB1 mRNA (green) in the aPC. Lower panels, higher magnifications of the boxed in the top panels. Yellow arrows, GAD-positive cells that do not express CB1; blue arrows, GAD-positive cells containing low levels of CB1 mRNA; white arrows, GAD-positive cells containing high levels of CB1 mRNA. Lines in top panels delimitate the different cortical layers (I, II and III). Scale bar, 100µm (top) and 50µm (bottom). **(B-E)** Pie-charts representing percentage distribution of **(B)** cells expressing GAD mRNA in different layers, **(C)** cells expressing CB1 mRNA in different layers, **(D)** GAD-positive cells expressing or not CB1 mRNA, and **(E)** cells expressing both GAD and CB1 mRNAs in different layers. Total numbers of counted cells are below each chart (n=118 sections from 4 animals). **(F)** Percentages of GAD mRNA expression in total high- (green) and low-CB1 expressing cells (light green) in the different layers of the aPC.

Figure 5. CB1 receptors control inhibitory transmission in SL cells of the aPC

(A) Representative traces of miniature inhibitory postsynaptic currents (mIPSCs) recorded in aPC semilunar-like neurons (SL) under different sequential treatments: Vehicle; WIN, CB1 receptor agonist WIN55,212-2 (5µM); WIN + AM251, WIN together with the CB1 receptor antagonist AM251 (4µM). **(B, C)** Quantifications of mIPSCs frequency **(B)** and amplitude **(C)** recorded in SL cells under Vehicle, WIN and WIN+AM251 treatments (n=10 cells from 4 mice). **(D)** Representative traces of mIPSCs recorded in aPC pyramidal-like neurons (PNs) under the same sequential treatments as in **(A-C)**. **(E, F)** Quantifications of mIPSCs frequency **(E)** and amplitude **(F)** recorded in PNs under the different treatments (n=9 cells from 5 mice). *, $p<0.05$; ns, not significant (vs Vehicle). For statistical details, see Table S1. For supplemental information, see Figure S5.

Figure 6. Involvement of GABAergic transmission in the CB1 receptor-dependent control of COP retrieval

(A) Representative traces of mIPSCs recorded in SL neurons in the aPC, from mice sacrificed during control water consumption (Water) or COP retrieval test (COP). **(B)** Quantifications of mIPSCs frequency in SL neurons of mice sacrificed during water consumption (Water, n=44 cells from 10 animals), COP retrieval test (COP, n=30 cells from 8 animals), exposure to odor-scented solutions without sucrose (Sucrose free, n=17 cells from 4 animals), exposure to sucrose solution without odors (Odor free, n= 27 cells from 4 animals), or COA retrieval test (COA, n= 24 cells from 4 animals). **(C)** Consumption of C+ and C- during COP retrieval test after administration of Vehicle or the CB1 receptor antagonist Rimonabant (IP, Rim, 1 mg/kg). **(D-F)** Effect of IP injections of Vehicle or Rim on mIPSCs of SL cells from mice sacrificed during COP retrieval test. **(D)** Representative traces. **(E)** Quantification of mIPSCs frequency (Vehicle, n=30 cells from 5 animals; Rim, n=32 cells from 5 animals). **(F)** Correlation between the average of mIPSC frequencies in SL neurons of individual animals and their COP retrieval performances expressed as Preference index (COP vehicle, n=5; COP Rim, n=5). **(G-I)** Effect of IP injections of Vehicle or Rim on mIPSCs of PNs in mice sacrificed during water consumption (Water vehicle) or COP retrieval test (COP vehicle and COP Rim). **(G)** Representative traces. **(H)** Quantification of mIPSCs frequency (Water vehicle, n=21 from 3 animals; COP vehicle, n=26 from 4 animals; COP Rim, n=27 from 5 animals). **(I)** Correlation between the average of mIPSC frequencies in PNs from individual animals and their COP retrieval performances, expressed as Preference index (COP vehicle, n=4; COP Rim, n=5). **(J-K)** Consumption of the C+ and C- odor-scented solutions in **(J)** GABA-*CB1*-WT (n=31) and in **(K)** GABA-*CB1*-KO mice (n=23)

receiving IP injections of either vehicle or Rim before a COP test (see STAR Methods). *, $p<0.05$; **, $p<0.01$; ***, $p<0.001$; ns, not significant. For statistical details, see Tables S1, S3. For supplemental information, see Figures S5 and S6.

STAR METHODS

KEY RESOURCES TABLE

LEAD CONTACT AND MATERIALS AVAILABILITY

This study did not generate new unique reagents. Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Giovanni Marsicano (giovanni.marsicano@inserm.fr).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

All experimental procedures were approved by the local Committee on Animal Health and Care of Bordeaux and the French Ministry of Agriculture and Forestry (authorization number A33063098) and Committee of Ethics for Animal Welfare of the University of the Basque Country (CEEA/408/2015/Grandes Moreno, CEIAB/ 213/2015/Grandes Moreno). Two to three months-old naive male *CB1-flox* [mice carrying the "floxed" *CB1* gene (*CB1* f/f)] were used [26,27,64]. Rescue, stop and knockout lines were generated as described [33,36,37,64]. Briefly, Stop-CB1 mouse line was produced by silencing the endogenous *CB1* gene with a *loxP*-flanked stop cassette in the 5′ UTR of the CB1 receptor start codon. To rescue the expression of the CB_1 receptor, Stop-CB1 line was crossed with a Cre-deleter mouse line. Conditional rescue mice were obtained by crossing Stop-CB_1 mice with Dlx5/6-CRE mice (gene expressed in differentiating GABAergic neurons) allowing the expression of CB1 in GABAergic neurons, named as "GABA-CB1 rescue", and with Nex-CRE mice (gene expressed in cortical glutamatergic neurons) allowing the expression of CB1 in cortical glutamatergic neurons, named as "Glu-CB1 rescue". Total CB1 receptor knockout (*CB1*-KO) mice and conditional knockout animals lacking CB1 receptor in forebrain GABAergic Dlx5/6 positive neurons (GABA-*CB1*-KO) were

obtained as described [27,45,64]. All behavioral experiments were performed during the light phase (from 9am to 1pm) and animals were kept in individual cages under a 12h light/dark cycle (lights on 7 am) and were maintained under standard conditions with food and water *ad libitum* prior undergoing behavioral procedures. At least three animals from each genotype or experimental group were used for immunohistochemistry, fluorescent *in situ* hybridization and electrophysiology recordings.

METHOD DETAILS

Behavioral procedures

Conditioned Odor Preference (COP)

Mice were water deprived during the whole protocol. During three consecutive days, animals had 1-hour access to two bottles of water. Over the following 4 days, animals received simultaneously (1-hour access) one bottle with an odor-scented solution, either banana (isoamyl acetate, 0.05%) or almond (benzaldehyde, 0.01%) diluted in water and one bottle with a different odor-scented solution (either almond or banana) mixed with the sweet taste sucrose (0.15M, 5%). Concentrations of banana (0.05%) and almond (0.01%) solutions were chosen to be equally consumed when diluted in water and prior to any associations with other stimuli [25,28,29]. Moreover, these almond- and banana-scented solutions were chosen to specifically served as odor cue based on previous studies indicating anosmic animals were unable to reliably detect almond- or banana-scented water (at higher concentrations than the ones used here), whereas they performed as well as control for taste detection [23,24]. This provides evidence that these aqueous banana and almond compounds did not confer any behaviorally detected gustatory sensation to the drinking solution.

During COP training, the odor-scented solution present in water was named odor 1 (O1) and the other odor-scented solution associated with sucrose was named odor 2+Sucrose (O2+Sucrose). Half of the mice received banana-sucrose and the other half almond-sucrose. No differences were observed between either conditions in all the experiments performed. The position of the bottles was changed every day. After this training, a preference test was performed using a 1-hour two bottles choice: each bottle was presented with an odor-alone solution (almond versus banana diluted in water without sucrose). Subjects showing COP will drink more liquid in the bottle with the odor previously associated with sucrose (C+) than in the other bottle (C-).

In order to test the impact of CB1 receptor blockade in some experiments (Figures 1E and 6J,K), mice injected with either the CB1 receptor antagonist (either AM251 in aPC or Rimonabant IP) or vehicle prior to the first COP test received 4 additional odor-sucrose pairings and were injected with the other treatment (either vehicle or CB1 receptor antagonist) before a second COP test.

Sucrose Preference

All subjects underwent 3 days habituation to water followed by 3 days with two bottles containing either water or sucrose. Finally, we evaluated the effect of aPC injection of AM251, or its vehicle, on their preference for sucrose over water.

Conditioned Odor Aversion (COA)

<u>COA induced by gastric malaise</u>

COA using gastric malaise was adapted from previous studies [25,28–30]. Mice followed the same habituation phase as described above. The conditioning phase consisted in 4 days. On days 4 and 6 the subjects received 1-hour access to odorized water (banana or almond) followed by an injection of Saline immediately

after the session (O1 + sal.). On Days 5 and 7 subjects received 1-hour access to the other odor (almond or banana) that they did not receive on Days 4 and 6, followed by an injection of lithium chloride (LiCl, 0.3 M, 1% b.w.; Sigma-Aldrich; O2 + LiCl) immediately after the session. The different odors were counterbalanced between each group. After this conditioning, the subjects were given a recovery day during which they received water dispensed in two bottles during 1 hour. The following day, a preference test was performed using a 1-hour two bottles choice: each bottle was presented with an odor (almond versus banana). During the test, subjects showing COA will drink less liquid in the bottle with the odor previously associated with LiCl (C+) than in the other bottle (C-).

COA induced by quinine

The COA with quinine followed the same procedure as the COP by replacing the sucrose by 0.1mM of quinine (Sigma-Aldrich). During the test, subjects showing COA will drink less liquid in the bottle with the odor previously associated with quinine (C+) than in the other bottle (C-). An index of preference or aversion was calculated for COP or COA experiments as the following:

$$\frac{\boldsymbol{Liquid\ intake\ of\ (C+) - liquid\ intake\ of\ (C-)}}{\boldsymbol{Total\ liquid\ intake}}$$

Surgery

Mice were anesthetized by IP injection of a mixture of ketamine (100mg/kg, Imalgene 500®) and xylazine (10mg/kg, Rompun®) or with isoflurane. Then, animals were placed into a stereotaxic apparatus (Model 900, Kopf instruments, CA, USA) with a mouse adaptor and lateral ear bars. For local deletion of CB1 receptors [25,27] in the aPC, CB1 flox mice were injected with an AAV-cag-CRE or its control AAV-cag-GFP

(mixed serotype AAV1/AAV2, 10^{10} Vg/ml) into the aPC (250µl per side, 125µl/min) with the following coordinates according to Paxinos and Franklin's mouse brain atlas [65]: AP +1.6, L ± 2.5, DV -4.8. For each animal receiving AAV-cag-CRE, CB1 deletion was verified by Fluorescent *In Situ* Hybridization against CB1 mRNA. To control that recombination did not involve the posterior PC or the anterior olfactory nucleus, data were obtained by averaging CB1 mRNA fluorescence intensity from 2/4 slices for each level in the antero-posterior axis (Figures S1H,I). In order to check region specificity across the medio-lateral axis, data were obtained from brain regions (insular cortex, aPC and lateral olfactory tubercle) bilaterally in the section for each mouse where maximal deletion of CB1 mRNA was observed (Figure 1C). Corresponding sections were quantified in both antero-posterior and medio-lateral axes in control mice injected with AAV-cag-GFP.

For local pharmacology experiments, mice were bilaterally implanted with 3.5mm stainless steel guide cannulae (Bilaney, UK) targeting the aPC with the following coordinates [65]: AP +1.6, L ± 2.5, DV -4.5. Guide cannulae were secured in place with dental cement. Mice were allowed to recover for 2 weeks in individual cages before the beginning of the experiments.

The placement of aPC cannulae was determined by injection of 2% pontamine sky blue solution (0.5µl per side).

Drugs

For *in vitro* patch-clamp experiment, WIN 55,212-2 (5µM) (Tocris Bioscience) and AM251 (4µM) (Tocris Bioscience) were prepared in Dimethyl Sulfoxide (DMSO) and applied for 10min.

For behavioral experiments, AM251 was dissolved in a mixture of 10% Cremophor-EL, 10% DMSO and 80% saline (NaCl 0.9%). AM251 (4µg/0.5µl per side) or its vehicle was injected bilaterally in the aPC using silicone tubing connected to a peristaltic pump (PHD 22/2000 Syringe Pump Infusion, Harvard Apparatus, Massachusetts, USA, flow rate: 0.5µl/min). Rimonabant (Cayman Chemical) was dissolved in a mixture of 1.25% Tween20, 1.25% DMSO and 97.5% saline (NaCl 0.9%). Rimonabant (1 mg/kg) or its vehicle was injected intraperitoneally (IP) in a volume of 10 ml/kg.

Mice injected with AM251 (4µg/0.5µl per side) or Rimonabant (1mg/kg) were left in their home cage 10 min or 30min before bottles presentation, respectively. In order to habituate animals to receive aPC infusion and systemic injection, animals were injected with a saline solution (NaCl 0.9%) in the same manner during the two previous days. Mice receiving local aPC infusion were kept awake and maintained by the tail during the injection.

Immunohistochemistry

Mice were anesthetized with pentobarbital (Exagon, 400 mg/kg body weight), transcardially perfused with phosphate-buffered solution (PBS 0.1M, pH 7.4) before being fixed with 4% formaldehyde prepared at 4°C. Serial coronal sections were cut at 40µm and collected in PBS at room temperature (RT). Sections were permeabilized in a blocking solution of 10% donkey serum, 0.3% Triton X-100 and 0.02% sodium azide in PBS for 1 hour at RT. Free-floating sections were incubated with a goat polyclonal antibody against C-terminal sequence of the mouse CB1 receptor (1:2000, Frontier Science Co.) for 48h at 4°C. After several washes, slices were incubated for 2 hours with a secondary anti-goat antibody conjugated to Alexa 488 (1:500, Fisher Scientific) and then washed in PBS at RT. Finally, sections were

incubated with DAPI (1:20 000, Fisher Scientific) for 5 minutes before being washed, mounted and coverslipped. The fluorescence was visualized with an epifluorescence Leica DM6000 microscope.

Immunocytochemistry for electron microscopy

For detailed methodological procedure see [66]. Coronal anterior Piriform Cortex (aPC) vibrosections were cut at 50 µm and collected in 0.1 M phosphate buffer (pH 7.4) at RT. Sections were preincubated in a blocking solution of 10% BSA, 0.1% sodium azide, and 0.02% saponin prepared in 1X Tris-HCl-buffered saline, pH 7.4, for 30 minutes at RT. A pre-embedding silver-intensified immunogold method was used for localization of the CB1 receptor protein. Briefly, aPC sections were incubated with the primary goat polyclonal anti-CB_1 receptor antibody (2 µg/ml Frontier Sciences Institute; goat polyclonal) in 10% BSA/Tris-HCl-buffered saline containing 0.1% sodium azide and 0.004% saponin on a shaker for 48h at 4ºC. After several washes in 1% BSA/Tris-HCl-buffered saline, tissue sections were incubated with a secondary 1.4-nm gold-labeled rabbit anti-goat Immunoglobulin G (Fab fragment; 1:100; Nanoprobes Inc.) in 1% BSA/Tris-HCl-buffered saline with 0.004% saponin on a shaker for 4 hours at RT. Sections were washed in 1% BSA/ Tris-HCl-buffered saline overnight at 4ºC and postfixed in 1% glutaraldehyde in Tris-HCl-buffered saline for 10 minutes at RT. After several washes in double-distilled water, gold particles were silver intensified with an HQ Silver kit (Nanoprobes Inc.) for approximately 12 minutes in the dark and then washed in double-distilled water first, and in a 0.1M phosphate buffer, pH 7.4 later. Stained sections were osmicated (1% osmium tetroxide, in 0.1 M phosphate buffer, pH 7.4, 20 minutes), dehydrated in graded alcohols to propylene oxide, and plastic-embedded in Epon resin 812. Ultrathin sections of 50 nm were collected on nickel mesh grids, stained with 2.5%

lead citrate for 20 minutes, and examined in a JEOL JEM 1400 Plus electron microscope. Tissue preparations were photographed by using a digital camera coupled to the electron microscope. Adjustments in contrast and brightness were made to the figures in Adobe Photoshop (Adobe Systems, San Jose, CA).

Fluorescent *in situ* hybridization

The procedure was performed as described [18,39]. Briefly, mice were sacrificed by cervical dislocation. Their brains were extracted, frozen on dry ice and stored at –80°C until sectioning in a cryostat (14 µm, Microm HM 500M, Microm Microtech). Fluorescein (FITC)-labeled riboprobes against mouse CB1 receptor and digoxigenin (DIG)-labeled riboprobes against mouse GAD65 were prepared as described [39]. After hybridization overnight at 60°C with the mixture of probes, the slides were washed with different stringency wash buffers at 65°C. Then, the slides were blocked with a blocking buffer prepared according to the manufacturer's protocol. Anti-DIG or anti-FITC antibodies conjugated to horseradish peroxidase (HRP) (Roche; 1 :2000) were applied 2 hours at RT or overnight at 4°C to detect respectively GAD65-DIG or CB1-FITC probes. Probes hybridization was revealed by a tyramide signal amplification (TSA) reaction using Cyanine 3-labeled tyramide (Perkin Elmer; 1:100 for 10 minutes) to detect GAD65 signal or FITC-conjugated tyramide (Perkin Elmer; 1:80 for 12 minutes) to amplify the signal of CB1. The slides were incubated in 4',6-diamidino-2-phenylindole (DAPI; 1:20 000; FISHER Scientific) before being washed, coverslipped and visualized with an epifluorescence Leica DM6000 microscope.

Electrophysiology

All the animals were sacrificed by dislocation during the light phase (9am to 12am). The brains were quickly removed and immerged in ice-cold oxygenated cutting

solution containing in mM: 180 Sucrose, 26 $NaHCO_3$, 12 $MgCl_2$, 11 Glucose, 2.5 KCl, 1.25 NaH_2PO_4, 0.2 $CaCl_2$, oxygenated with 95% O2/5% CO_2 ≈ 300mOsm. Coronal aPC slices (300µm thick) were obtained using a vibratome (VT1200S, Leica) and transferred for 30min into a 34°C bath of oxygenated ACSF containing in mM: 123 NaCl, 26 $NaHCO_3$, 11 Glucose, 2.5 KCl, 2.5 $CaCl_2$, 1.3 $MgCl_2$, 1.25 NaH_2PO_4 ≈ 305 mOsm. After a minimum of 30min recovery at RT (22-25°C), slices were transferred to a recording chamber in ACSF at 32°C. Recordings were performed using a Multiclamp 700B amplifier (Molecular devices) in principal glutamatergic neurons clamped with glass pipettes (3-5 MΩ) filled with an internal solution containing in mM: 130 KCl, 10 HEPES, 1 EGTA, 2 $MgCl_2$, 0.3 $CaCl_2$, 7 Phosphocreatin, 3 Mg-ATP, 0.3 Na-GTP; pH=7.2; 290mOsm. These cells were identified based on their morphology and somatic location using a contrast microscope (axio examiner.A1, Zeiss) and through electrical properties by measuring their resting potential and their excitability in current-clamp mode after 300ms steps of current injections from -50 to 300pA with steps of 25pA [34,35,51]. Neurons with large apical dendrites, soma located in the upper half of layer II, resting potential of around -70mV, input resistance of around 200-300 MΩ and displaying regular spiking were considered as semilunar-like cells (SL, see Figures S5A-B). Instead, neurons with large basal dendrites, soma located in the lower half of layer II and upper part of layer III, resting potential around -75mV, input resistance of around 100-150 MΩ and showing initial burst firing were classified as pyramidal-like neurons (PN, see Figures S5A-B). Given the complex layered structure of the aPC, miniature inhibitory post-synaptic currents (mIPSCs), known to be regulated by CB1 receptors [44], were specifically chosen to avoid restricting the study of inhibitory inputs coming from a specific layer where the stimulating electrode would have been placed. mIPSCs were obtained in voltage clamp mode in presence

of NMDA and AMPA/Kainate receptor antagonists (50µM D-APV and 10µM NBQX) and of the voltage-gated sodium channels blocker, tetrodotoxin (1µM TTX). Vehicle (DMSO) was applied before starting the recording and CB1 agonist (WIN 5µM) and antagonist (AM251 4µM) were applied for 10min successively. For experiments performed after behavior, animals underwent the two bottles choice test for 15min and were sacrificed 5min later. mIPSCs were collected in the same manner as for naïve animals, for 5 min in presence of vehicle (DMSO).

QUANTIFICATION AND STATISTICAL ANALYSIS

Behavioral data

For all the experiments, data are presented as absolute liquid intake. Considering the variability of liquid consumption during the two first days of pairings (likely due to the random choice of a bottle at day 1 and the confusion that might appear because of the inverted position of the bottles at day 2), only the last two days of the learning phase showing a reliable preference/aversion behavior were presented for each experiments.

Numerical evaluation for electron microscopy

Semi-quantitative analysis of CB1 receptor presence in excitatory or inhibitory terminals was done in aPC layers I and II of *CB1*-WT and *CB1*-KO according to our published procedure [66]. Total analyzed area was more than 2200 μm^2 per genotype (n=3).

Numerical evaluation for FISH

Cells expressing mRNAs were quantified in the three layers of the aPC. Because CB1 mRNA level is variable, CB1 positive cells were classified according to the level

of transcript visualized by the intensity of fluorescence [39]. “High-CB1” cells were considered to be round-shaped and intense staining covering the entire nucleus whereas “Low-CB1” cells were defined with discontinuous shape and lowest intensity of fluorescence allowing the discrimination of grains of staining. Numerical evaluation of the double FISH was performed manually in 118 sections from 4 animals, by evaluating the coexpression of CB1-positive cells with GAD 65 marker.

Electrophysiology

Electrophysiological data were filtered at 4kHz by a Digidata 1440A (Molecular devices) and they were collected during the last 5min of recording in each condition. Electrical properties were analyzed with Clampfit and mIPSCs were analyzed using Axograph software.

Statistics

Electrophysiological and behavioral data were analyzed with Prism Software (GraphPad). Repeated or unpaired statistical analyses were obtained with Student’s t-test,ANOVA (one-way or two way), mixed effects analysis and linear regression to compare two or multiple groups and for correlation where appropriate. When ANOVA provided significant main factor effects or significant interactions, Dunnett or Sidak post-hoc analyses were performed as appropriate. Statistical details are presented in tables S1, S2 and S3. Significance was set at $p < 0.05$ and data are expressed as mean ± SEM.

DATA AND CODE AVAILABILITY

This study did not generate datasets/code. Further data information are available upon request by contacting the Lead Contact, Giovanni Marsicano

(giovanni.marsicano@inserm.fr).

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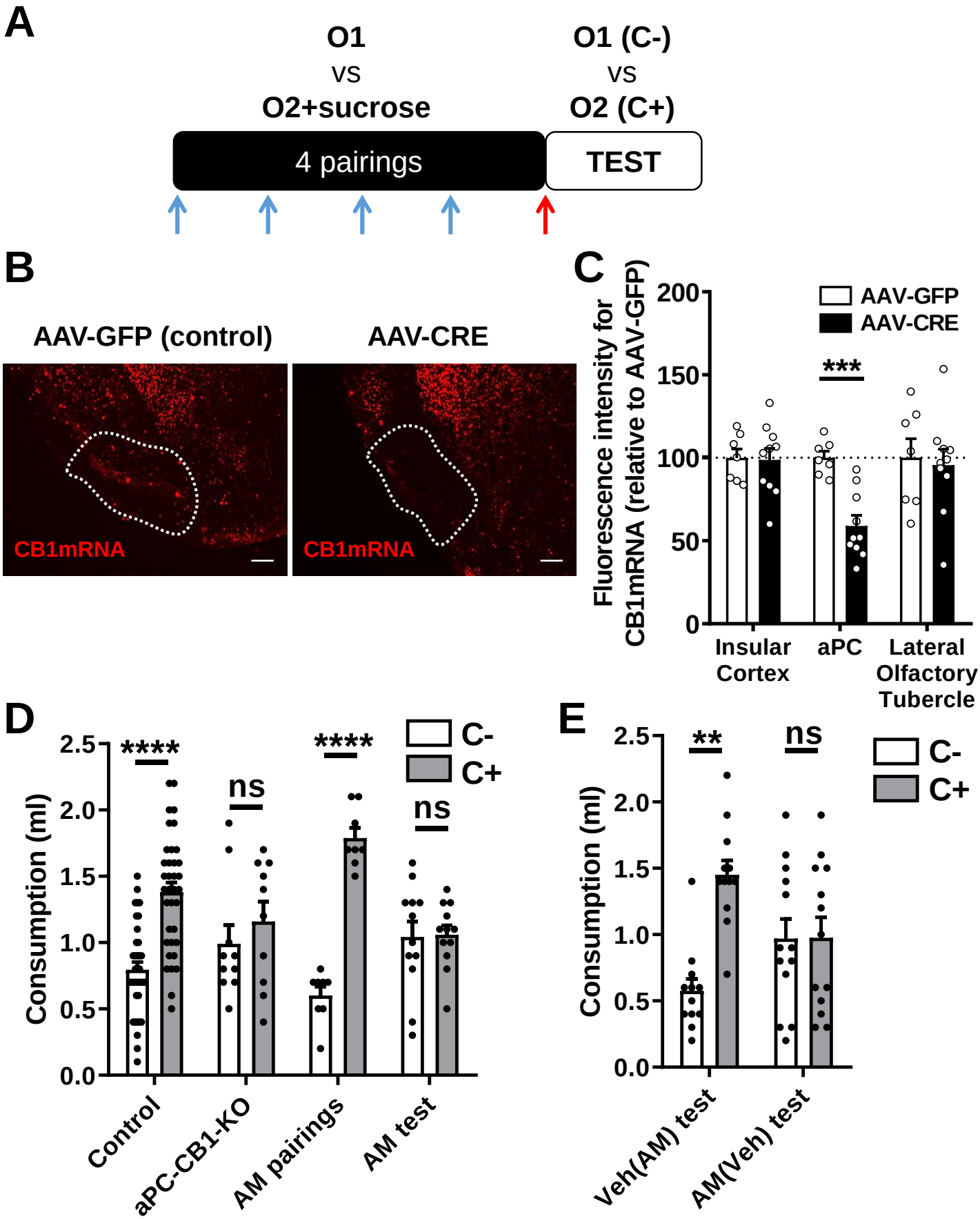
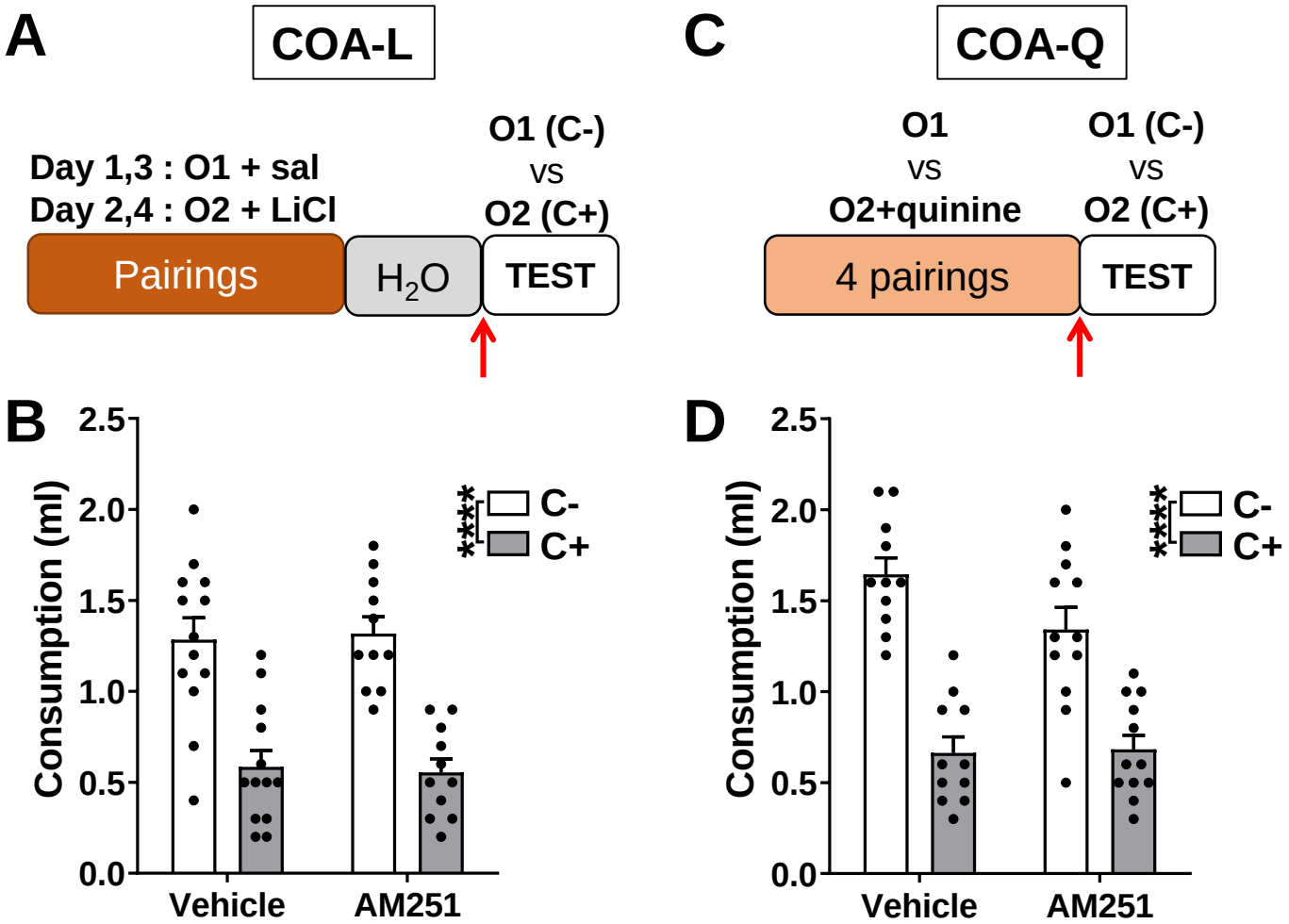
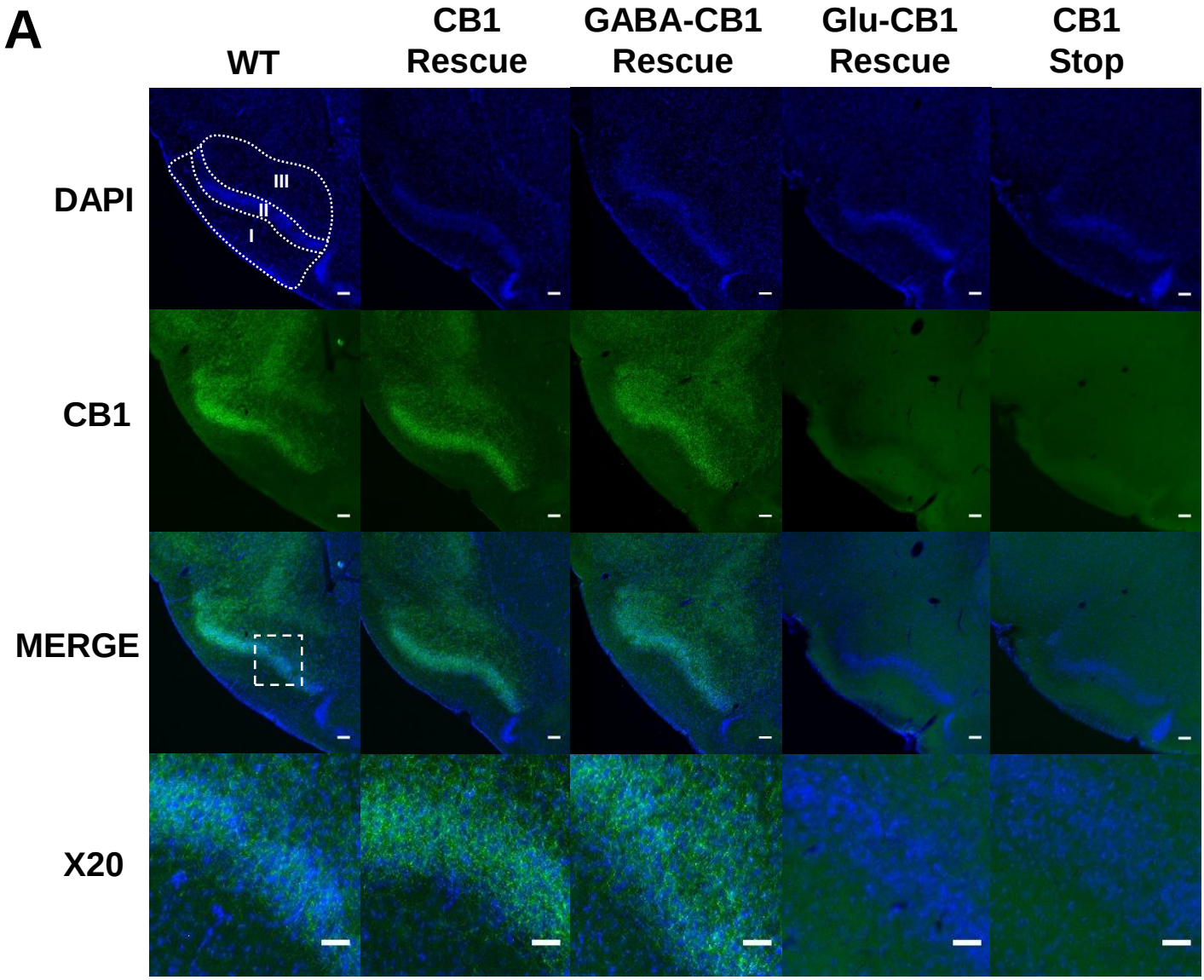
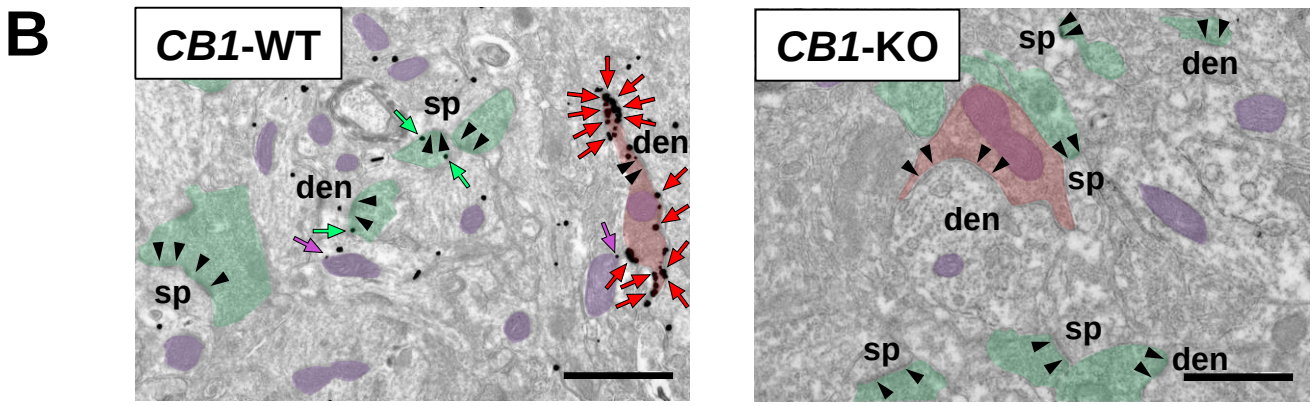
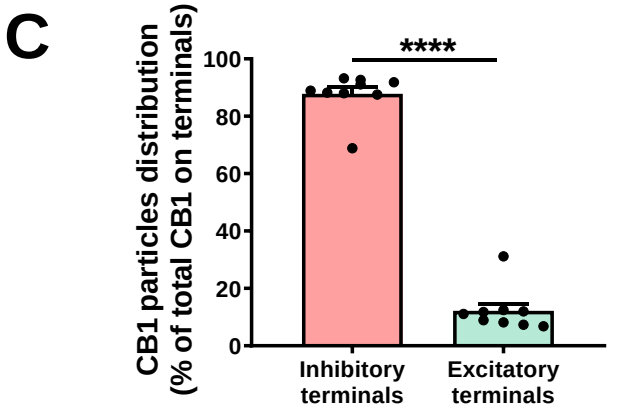
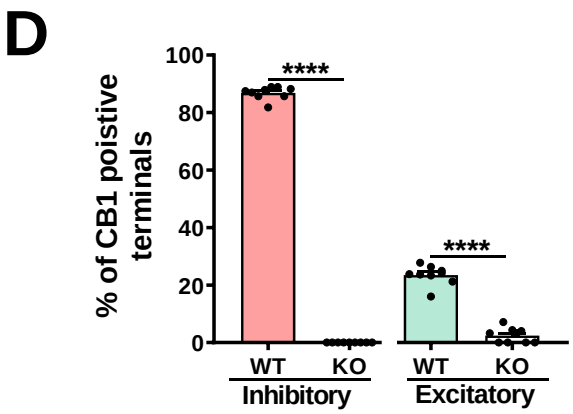

A
O1
vs
O2+sucrose
O1 (C-)
vs
O2 (C+)
4 pairings
TEST
B
AAV-GFP (control)
AAV-CRE
CB1mRNA
CB1mRNA
C
Fluorescence intensity for CB1mRNA (relative to AAV-GFP)
AAV-GFP
AAV-CRE
***
Insular Cortex
aPC
Lateral Olfactory Tubercle
D
Consumption (ml)
C-
C+
****
ns
****
ns
Control
aPC-CB1-KO
AM pairings
AM test
E
Consumption (ml)
C-
C+
**
ns
Veh(AM) test
AM(Veh) test

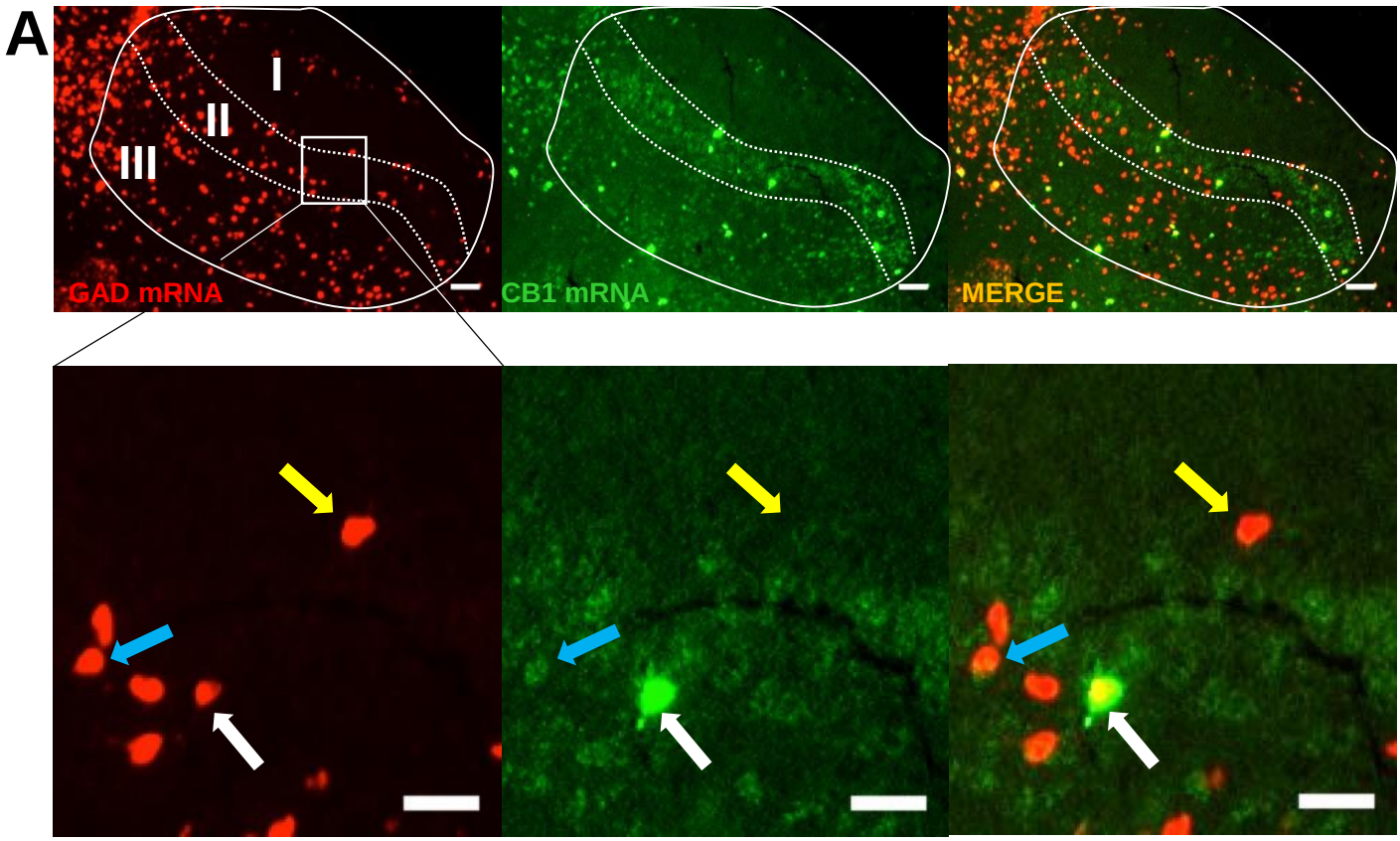

A
COA-L
Day 1,3 : O1 + sal
Day 2,4 : O2 + LiCl
O1 (C-)
vs
O2 (C+)
Pairings
H2O
TEST
C
COA-Q
O1
vs
O2+quinine
O1 (C-)
vs
O2 (C+)
4 pairings
TEST
B
D
Consumption (ml)
2.5
2.0
1.5
1.0
0.5
0.0
****
C-
C+
Vehicle
AM251

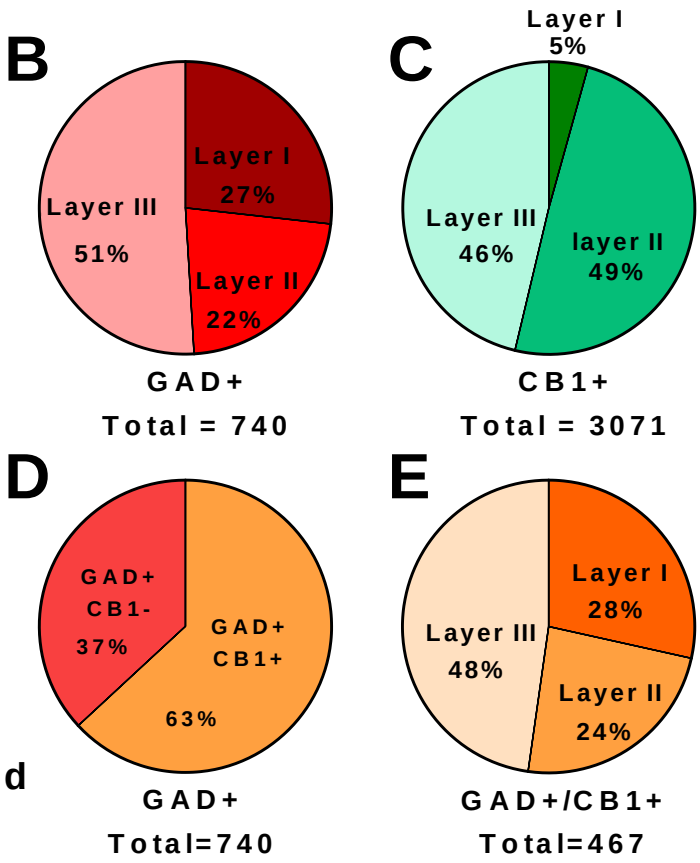

A
WT
CB1 Rescue
GABA-CB1 Rescue
Glu-CB1 Rescue
CB1 Stop
DAPI
CB1
MERGE
X20
III
II
I

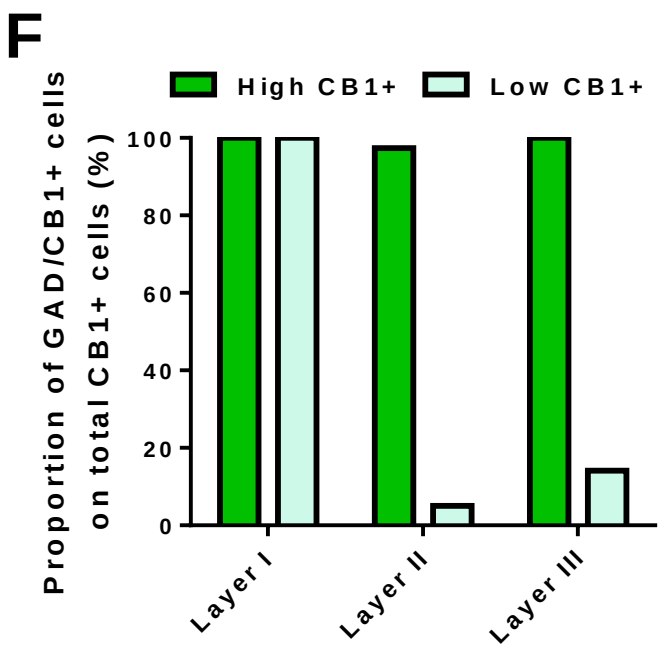

B
CB1-WT
sp
den
sp
den
CB1-KO
sp
den
sp
den
sp
sp
den

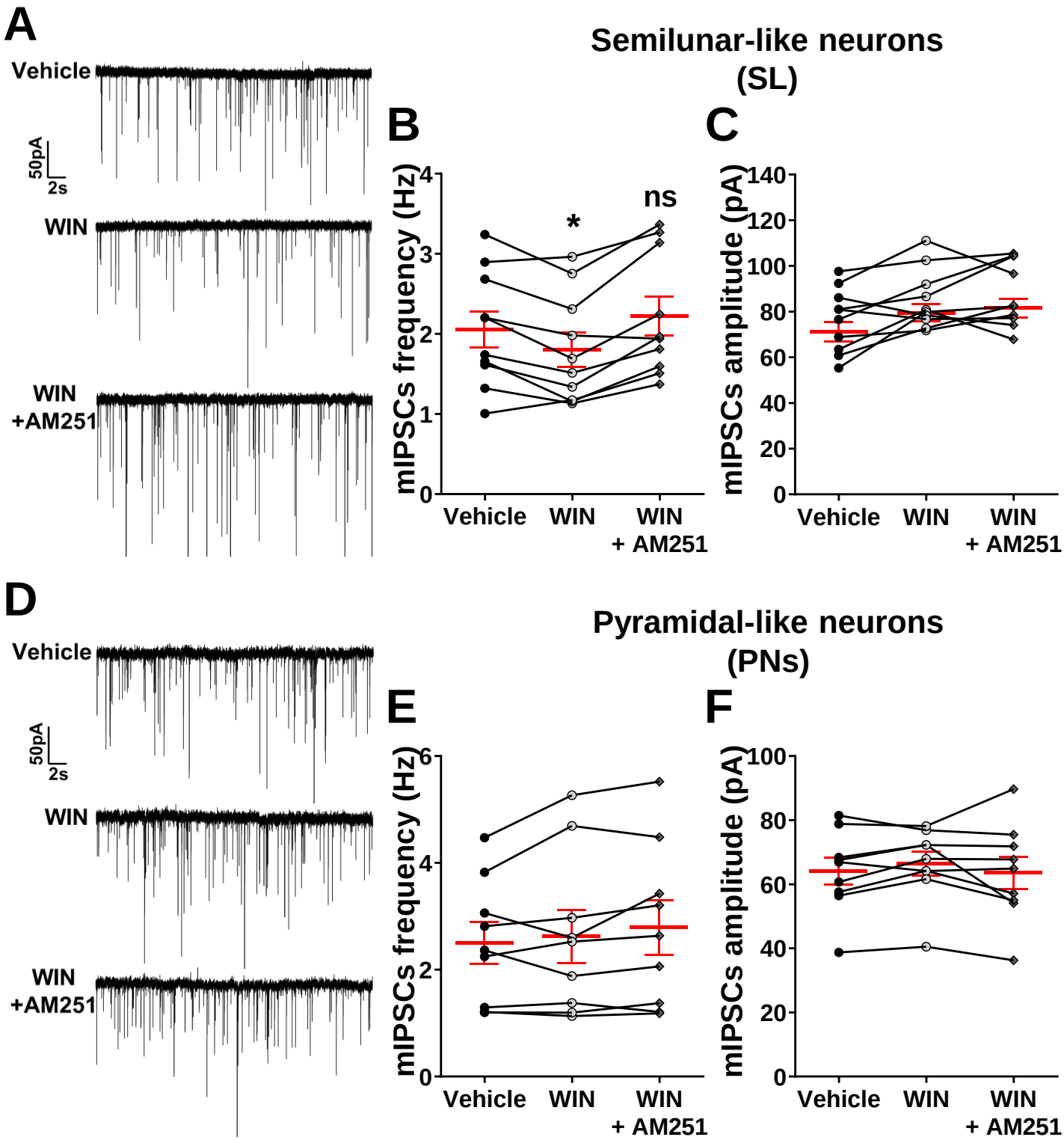

C
CB1 particles distribution (% of total CB1 on terminals)
****
100
80
60
40
20
0
Inhibitory terminals
Excitatory terminals

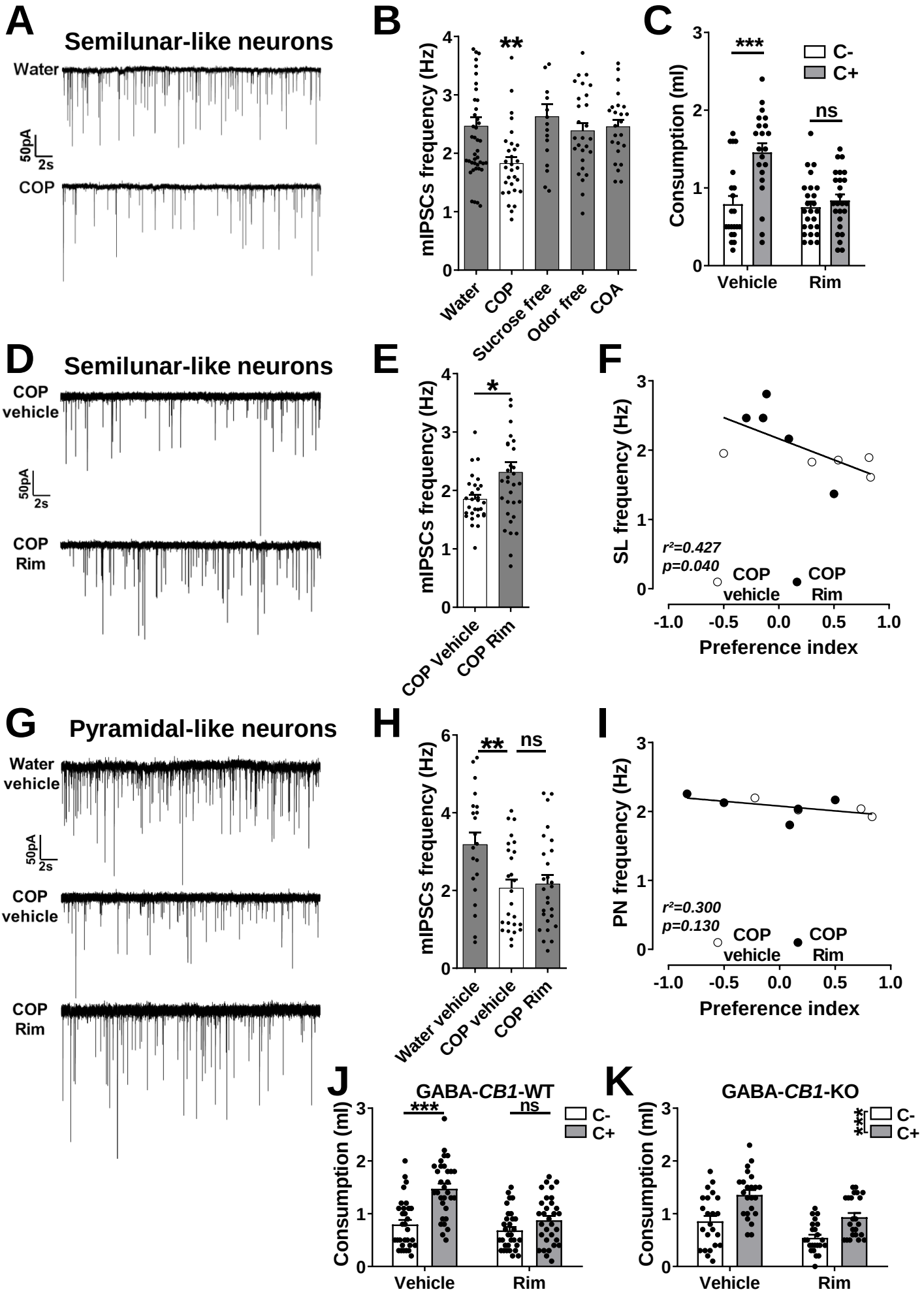

D
% of CB1 poistive terminals
****
****
100
80
60
40
20
0
WT
KO
WT
KO
Inhibitory
Excitatory


A
I
II
III
GAD mRNA
CB1 mRNA
MERGE


B
Layer I
27%
Layer II
22%
Layer III
51%
GAD+
Total = 740
C
Layer I
5%
layer II
49%
Layer III
46%
CB1+
Total = 3071
D
GAD+
CB1-
37%
GAD+
CB1+
63%
d
GAD+
Total=740
E
Layer I
28%
Layer II
24%
Layer III
48%
GAD+/CB1+
Total=467


F
High CB1+
Low CB1+
Proportion of GAD/CB1+ cells on total CB1+ cells (%)
100
80
60
40
20
0
Layer I
Layer II
Layer III


A
Vehicle
50pA
2s
WIN
WIN
+AM251
Semilunar-like neurons
(SL)
B
mIPSCs frequency (Hz)
0
1
2
3
4
*
ns
Vehicle
WIN
WIN
+ AM251
C
mIPSCs amplitude (pA)
0
20
40
60
80
100
120
140
Vehicle
WIN
WIN
+ AM251
D
Vehicle
50pA
2s
WIN
WIN
+AM251
Pyramidal-like neurons
(PNs)
E
mIPSCs frequency (Hz)
0
2
4
6
Vehicle
WIN
WIN
+ AM251
F
mIPSCs amplitude (pA)
0
20
40
60
80
100
Vehicle
WIN
WIN
+ AM251


A
Semilunar-like neurons
Water
50pA
2s
COP
B
mIPSCs frequency (Hz)
**
Water
COP
Sucrose free
Odor free
COA
C
Consumption (ml)
***
C-
C+
ns
Vehicle
Rim
D
Semilunar-like neurons
COP vehicle
50pA
2s
COP Rim
E
mIPSCs frequency (Hz)
*
COP Vehicle
COP Rim
F
SL frequency (Hz)
r²=0.427
p=0.040
COP vehicle
COP Rim
Preference index
G
Pyramidal-like neurons
Water vehicle
50pA
2s
COP vehicle
COP Rim
H
mIPSCs frequency (Hz)
**
ns
Water vehicle
COP vehicle
COP Rim
I
PN frequency (Hz)
r²=0.300
p=0.130
COP vehicle
COP Rim
Preference index
J
GABA-CB1-WT
***
ns
C-
C+
Consumption (ml)
Vehicle
Rim
K
GABA-CB1-KO
***
C-
C+
Consumption (ml)
Vehicle
Rim