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Limited genetic structure and demographic expansion of the *Brassicogethes aeneus* populations in France and in Europe

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Abstract

BACKGROUND: The pollen beetles, *Brassicogethes aeneus* (Fabricius, 1775), is one of the most significant pests of oilseed rape. To shed light on past and present pollen beetle demography (dispersal, population size), twelve microsatellite markers were developed and population genetic diversity and structure were analyzed at different spatial scales in France and in Europe from 433 individuals collected in 18 winter oilseed rape fields.

RESULTS: The genetic differentiation among the population samples was low but significant between the Estonian sample and the rest of Europe. Isolation by distance was only significant at the European scale. The genetic variability was similar among the 18 population samples. Demographic inferences suggested a recent expansion of *B. aeneus* population size over Europe, possibly corresponding to an increase of oilseed rape crop's area during the last decades.

CONCLUSION: Current population size and dispersal are not straightforward to estimate from the distribution of genetic variability in *B. aeneus* over Europe because of the complexity of the demographic history of the pests. Nevertheless, as gene flow was important enough to prevent strong genetic differentiation at large geographical scales, the management of pollen beetle populations should likely be thought at the European continental level.

Keywords: *Brassicogethes aeneus*, genetic diversity, genetic structure, microsatellites, oilseed rape, pollen beetles

1 Introduction

Population genetic studies on crop pests provide information on the spatial scales at which population structure and gene flow occurs. Such information can help defining spatially relevant strategies of pests control¹. In addition, genetic diversity contains information on past and present demography that could be useful to characterize the demographic history of crop pests.

The pollen beetle (*Brassicogethes aeneus*, Fabricius, 1775), formally named *Meligethes aeneus* (Fabricius, 1775) is one of the major pests of oilseed rape (OSR) crop in Europe². After their emergence from overwintering areas in early spring, adults migrate to OSR fields to feed on pollen and oviposit in buds, thereby inflicting severe yield losses³. Larvae also feed on OSR flowers before dropping down on the ground to pupate². The new generation emerges in early summer (June) and seeks for overwintering sites outside the crop, mainly in woodland habitat. Consequently, the pollen beetle dispersal is split in two phases, one in spring and one in summer⁴, between crops and semi-natural habitats⁵, suggesting important gene flows at the scale of a few kilometers.

In a study based on mitochondrial DNA, five phylogenetic groups of pollen beetles were identified in Europe: a first one in England and Wales, a second one in Scotland, a third one in South Eastern Europe (Italy, Romania and Greece), a fourth one in North Western Europe (France, Germany, Belgium), and a fifth in the Baltic and Scandinavian countries⁶. Studies based on amplified fragment length polymorphism (AFLP) suggested a high level of gene flow between pollen beetle populations in Sweden⁷, but confirmed genetic structure among European pollen beetle populations⁸. However, the study of contemporary processes affecting genetic variation and genetic structure requires more variable genetic markers⁹. Microsatellite markers are highly polymorphic with an estimated average mutation rate of roughly $\mu = 5 \times 10^{-4,10}$ compared to an average of roughly $\mu = 6 \times 10^{-8}$ for mitochondrial DNA. In addition, microsatellite are co-dominant genetic markers, abundant throughout the genome and cost effective to develop and screen. These properties have made microsatellite markers very popular in population genetics studies, as they allow the investigation of spatial genetic structure and demographic changes at small geographic and temporal scales¹¹.

OSR cropped area in Europe has strongly fluctuated in the recent history, which may have affected the demography of the pollen beetle populations. OSR was first cropped during the middle age in Flanders and was the main source of vegetal oil until the middle of the nineteenth century in Europe¹². In France, OSR cropped area peaked up to 200 kha in 1862,

before coming down to 11 kha in 1939 because of the progressive replacement of OSR oil by peanut oil from the colonies¹². Oleaginous crops in France raised up again after the Second World War: 391 kha in 1970, 460 kha in 1982, and 1600 kha in 2012^{12,13}. In Europe, the expansion of the OSR cropped area likely followed a similar trend but might have been even more sudden. For example in Finland, OSR cropped area raised from 10 kha in 1973 to 70 kha in 1993¹⁴. Following the expansion of OSR cropped area over the last 60 years in Europe, pollen beetle population size would have been expected to increase. However, OSR crops were treated using large amounts of pesticides in most European countries since 1950¹⁵, which might have inversely reduced pollen beetle population size. As a consequence of these treatments, the pollen beetle has developed resistance to several insecticides^{16,17}, and resistance to the pyrethroids have spread throughout Europe since 2000¹⁶, which may have reduced the impact of the pesticides on variations in pollen beetle population sizes.

The aim of this paper is to report analyzes of the genetic variation and structure in pollen beetle populations at various spatial scales in Europe using microsatellite markers. First, we developed microsatellite markers in *B. aeneus* and cross amplification on another OSR pest, *Brassicogethes viridescens*. Second, genetic structure among pollen beetle populations from 18 OSR fields was assessed using the newly designed microsatellite markers. Third, temporal change in pollen beetle population effective sizes was investigated to test if it increased with OSR cropped area in the recent history.

2 Material and Methods

2.1 Study sites and pollen beetle sampling

Pollen beetle adults were collected in 18 fields organized at three nested spatial scales: six OSR fields at a fine spatial scale in the Eure department in France (Fig. 1a), seven fields from various other regions in France (Fig. 1b) and five fields from five other European countries (Fig. 1c). Hereafter, each sampled field was considered as a different pollen beetle sampling site. Pollen beetle were collected by beating in one point of each field. Collection took place in spring 2015 with the exception of the sampling sites from Switzerland (spring 2014) and of the six French sampling sites from Eure (spring 2016). All individuals were identified at the species level comparing their meta-femur¹⁸. *B. viridescens* as a spine on the meta-femur whereas the meta-femur of *B.aeneus* is smooth. Most pollen beetles were identified as *B. aeneus* (99%), but several *B. viridescens* individuals were collected at location 12 (70% were *B. aeneus* and 30% were *B. viridescens*). Hereafter, we only used the *B. aeneus* individuals unless otherwise specified.

2.2 DNA extraction

DNA extractions were performed in 96-well format plates. First, pollen beetle tissues were ground in 50 μ L of H₂O using a 2 mm steel bead using a 1600 MiniG (Spex® SamplePrep) homogenizer at 1500 strokes/min for 30 seconds. Second, tissues were digested at 56°C for 14 hours in a 100 μ L solution including 10% Chelex® 100 (Biorad) and 3% proteinase K using a Mastercycler thermocycler (Eppendorf). The tissue digestion was stopped by a final thermocycler step of 30 min at 98°C. Finally, the supernatant of this solution was used as DNA template for the PCR reactions.

2.3 Development of microsatellite markers

An enriched DNA library with microsatellite sequences was produced following Malausa et al.,¹⁹ on 9 pollen beetles from three different oil seed rape fields in France. The DNA library was produced by GenoScreen (Lille, France) using Roche 454 GS-FLX Titanium pyrosequencing. A total of 255 microsatellite markers were identified by their number of repetitions out of the 93,675 DNA sequences obtained (0.3%). The 48 longest microsatellite sequences were selected and a primer pair was chosen for each of them using Primer3²⁰. Each forward primer was tailed with a labelled M13 sequence²¹. PCR amplifications were carried out independently for each microsatellite locus in a 12 μ L reaction volume containing 1X GoTaq® Flexi Buffer, 1.5 mM MgCl₂, and 0.1 mg/ml Bovine Serum Albumin, 200 μ M of each dNTPs, 0.4 μ M of reverse primer, 0.4 μ M of labelled M13 forward primer, 0.04 μ M of M13-tailed forward primer, 1 unit of GoTaq® Flexi DNA Polymerase (Promega) and 2 μ L of DNA template. The PCR conditions were: 5 min at 95°C followed by 30 cycles at 95°C for 30 s, 54°C for 45 s, and 72°C for 45 s, followed by 10 cycles at 94°C for 30 s, 50°C for 45 s, and 72°C for 45 s with a final extension step at 72°C for 10 min.

We tested PCR amplification at these 48 microsatellite loci on six *B. aeneus* individuals with labelled M13 forward primers. For the loci that amplified, we verified their variability for population genetic studies on 20 additional *B. aeneus* individuals from location 1 (Fig. 1). For each microsatellite locus, we calculated the number of alleles, the observed heterozygosities (H_o) and gene diversities (H_E). We estimated the frequency of null alleles following Brookfield (1996). We tested deviations to Hardy-Weinberg equilibrium (HWE) using Genepop, version 4.2.2²². We selected the polymorphic loci with less than 10% of null allele and at HWE.

The selected microsatellite loci were combined in PCR multiplex, labelling each forward primer with a fluorescent dye at their 5'-end (Table S1), either 6-FAM (6-carboxyfluorescein),

or *HEX* (hexachloro-fluoresceine), or *TAMRA* (carboxy-tetramethyl-rhodamine), or *ATTO 565* (Rhodamine dyesclass). These loci were also tested on nine *B. viridescens* individuals from location 12 to verify interspecific cross-amplification.

2.4 Microsatellite analysis

A total of 433 pollen beetle individuals from the 18 OSR sample sites were genotyped at the selected microsatellite loci (Table 1). PCR amplification was carried out with a Mastercycler thermocycler (Eppendorf) in a 10 µl reaction volume containing 5 µl of master-mix (QIAGEN), 2 µl of primer-mix (primers concentration ranging from 0.13 to 2.50 µM, Table S1), and 2 µl of DNA template. The PCR conditions were: 15 minutes at 95°C followed by 35 cycles at 94°C for 30 s, 54°C for 90 s, and 72°C for 1 min with a final extension step at 72°C for 20 min.

Each PCR multiplex product was diluted in 40 µl H₂O. Two microliters of this dilution was added to 7.8 µl of HiDi formamide, and 0.2 µl GeneScan™- 600 LIZ® Size standard (Applied Biosystems). This was injected on an ABI 3730xl DNA Analyzer using POP7 polymer (Applied Biosystems). Genotypes were scored using GeneMapper®, version 4.1 (Applied Biosystems).

2.5. Statistical analysis

2.5.1. Genetic diversity

For each sample site, deviation from Hardy-Weinberg equilibrium (HWE) for each locus and linkage equilibria for each locus pair were tested with GENEPOP version 4.2.2²². The proportions of heterozygous genotypes (H_O) and unbiased estimate of gene diversities (H_E) were calculated at each locus for each sample site according to Nei²³ using GENEPOP. Allelic richness (A_r) in each sample site was estimated using a rarefaction method, using the HP-RARE program²⁴, which was parameterized with the smallest population sample size at site 8 ($n=16$) as the reference. Allelic richness, observed heterozygosities, and gene diversities were compared between population samples using Mann-Whitney-Wilcoxon tests.

2.5.2. Population genetic structure at multiple scales

First, to explore the number of homogeneous genetic clusters (K) within our dataset, population genetic structure across all sample sites was assessed using Bayesian clustering in STRUCTURE version 2.2²⁵. For each model with values of K from 1 to 20, we performed 10 runs of 2,000,000 iterations after a 'burn-in' period of 500,000 iterations. Individual assignments were computed assuming admixture among the K groups and non-equal

contribution of the K sources to the admixture. As recommended by Wang²⁶, the geographical locations of the sampled individuals was used as prior in the simulations. Results were summarized using the standard pipeline on the CLUMPAK web server²⁷. The most likely number of clusters (K) was explored using the estimated logarithm of likelihood (LnP(D)) and the ΔK method²⁸ that finds the point of greatest change in the distribution of LnP(D) with STRUCTURE HARVESTER version 0.6.92²⁹. Each individual was assigned to the cluster for which the membership coefficients Q was the highest²⁵.

Second, genotypic differentiation between population samples was calculated (F_{ST}) and tested using GENEPOP and isolation-by-distance (IBD) was analysed by regressing pairwise estimates of $F_{ST} / (1 - F_{ST})$ against the logarithm of geographical distance between sample sites³⁰. We also tested the groups found with STRUCTURE. IBD was tested using a Mantel test (1000 permutations) at different spatial scales using *i*) all the 18 sample sites, *ii*) 17 sample sites excluding the Estonian sample site, *iii*) the 13 sample sites located in France and *iv*) the six Eure sample sites (Fig. 1). The inverse of the IBD slope was used to estimate the genetic neighborhood size $D\sigma^2$ ³¹, a composite parameter that describes the strength of isolation by distance³², where D is the effective density of adults, and σ^2 is the mean squared parent-offspring dispersal distance.

2.5.3. Past demographic changes

Gene diversity (H_E) at a locus within a population can differ from the gene diversity expected at mutation-drift equilibrium for the same number of alleles (H_{eq}) due to demographic changes³³. In order to test if there has been a recent population expansion in Europe (i.e. $H_E < H_{eq}$), we have estimated H_{eq} using two contrasted models of mutation for the microsatellite markers, the stepwise (SMM) and the two-phase (TPM) mutation models with BOTTLENECK version 1.2³³. Under the TPM, H_{eq} was computed using 50% single-step mutations, 50% multiple-step mutations and a variance among multiple steps of 12. Difference between H_E and H_{eq} was tested for each sampling site separately, for pooled samples from Eure, for pooled samples from France and for all the samples pooled using the two tailed Wilcoxon's signed-rank test.

Population demographic history was further explored using the OnePopVarSize model implemented in the MIGRAINE software, which uses coalescent simulations under a maximum likelihood framework³⁴. The OnePopVarSize model infers parameters of a continuous past variation in population size for a single population using a generalized stepwise mutation model (GSM) adapted for microsatellite loci. MIGRAINE infers point estimates and 95% confidence intervals (95% CI) for up to five parameters: $pGSM$, the

parameter of the geometric distribution describing the number of repeats removed or added to the ancestral state for each mutation; $2N\mu$, the current population size scaled by the mutation rate per generation and locus; $2N_{anc}\mu$, the scaled ancestral population size; and $D_g\mu$, the duration of the population size change scaled by mutation rate. An additional composite parameter, N_{ratio} , is inferred to better characterize the strength of the change between current and ancestral population sizes ($N_{ratio} = N/N_{anc}$). Past changes in population size are thus significant when the N_{ratio} 95% confidence interval does not include one. We fixed initial large ranges for all parameter value since MIGRAINE can automatically expand or reduce the parameter range explored between different iterations. We inferred past changes in population size with a dataset pooling all the genotyped individuals from the 13 French sites and using the PACanc algorithm to estimate likelihood. Preliminary runs were performed using 2,000 points, 5,000 trees, and 5 iterations. Next, final runs were performed using 6,000 points, 20,000 trees, and 3-5 iterations with narrow parameter ranges deduced from the preliminary runs. We used a mutation rate of 5×10^{-4} per locus per generation for all microsatellite loci¹⁰ to convert scaled parameter estimates into biological reliable ones (i.e. N , N_{anc} , and D_g).

3 Results

3.1 Design and selection of microsatellite markers

The DNA library produced using Roche 454 GS-FLX Titanium pyrosequencing provided 93,675 sequences among which 255 were available to design microsatellite markers. Out of the 48 markers designed, 23 amplified on the six pollen beetle individuals tested and were selected for amplification on a reference sampling site of 24 individuals from location 11 (Figure 1). Among these 23 loci, twelve were finally used to study the genetic of pollen beetle populations. First, we discarded two loci that were monomorphic (*Ma-DMBX2*, *Ma-ERC7C1*). Second, we discarded nine additional loci that were not at HWE and that showed proportions of null alleles over 10 % in the reference sampling site (*Ma-C6CAE*, *Ma-C5QWM*, *Ma-D904J*, *Ma-DCGTK*, *Ma-EJCJJ*, *Ma-EL5EB*, *Ma-C2D1M*, *Ma-DJDOQ*, *Ma-EGYLO*).

At these twelve remaining loci, the number of alleles ranged from three to nine (mean = 6.00), the gene diversity (H_E) ranged from 0.17 to 0.85 (mean = 0.63) and proportion of heterozygotes (H_O) ranged from 0.18 to 0.87 (mean = 0.60) (Table 1). The proportions of null allele did not exceed 0.09 (mean = 0.03). Among these twelve *B. aeneus* microsatellite loci, nine markers successfully cross-amplified in *B. viridescens*, of which seven were observed

polymorphic (Table S2). Furthermore, four of these loci had alleles observed in *B. viridescens* but not in *B. aeneus* (*Ma-C4QRG*, *Ma-D3QFM*, *Ma-DDEYS*, *Ma-DQM5T*).

3.2. Genetic variability in pollen beetle populations

In total, 433 individuals from 18 OSR fields in Europe were genotyped at the 12 microsatellite loci. The number of alleles per sampling site and per locus ranged from 4.8 to 6.5. The only significant linkage disequilibrium was observed between loci *Ma-EL7YR* and *Ma-DCH30* in one sampling site (location 5) deemed insufficient to discard one of them.

Multi-locus tests did not reveal any significant departure from HWE in any of the 18 sampling sites. Observed heterozygosities (H_O) ranged from 0.54 to 0.67, with a mean value of 0.60 across all loci and sampling sites. Similarly, gene diversities (H_E) ranged from 0.57 to 0.64 with a mean value of 0.62 across all loci and sampling sites. The mean allelic richness (A_r) ranged from 4.4 to 5.9 with a mean value of 5.2 across all loci and sampling sites (Table 2). No difference in polymorphism was detected between the 18 sampling sites using H_E , H_O or A_r ($p > 0.05$).

3.3. Genetic structure at multiple scales

The most likely value of K from the STRUCTURE analysis based on the ΔK method²⁸ method or on $\text{LnP}(K)$ was three (Table S3). For 404 out of the 433 individuals, the estimated membership coefficients Q in each cluster, was the highest in the first cluster (343 individuals had $Q > 0.5$ in this first cluster). All 24 individuals from Estonia had the highest Q value in the second cluster (all with $Q > 0.5$). None of the 433 individuals had a Q value above 0.5 in the third cluster, but four individuals from Switzerland had their highest Q value in this third cluster (Fig. 2).

Out of the 153 pairwise F_{ST} values calculated between sample sites only 31 were associated to significant genetic differentiation ($p < 0.05$), all including at least one easternmost sample site (i.e. samples from Estonia, Poland or Sweden, Table S4). Consistently with the STRUCTURE results, all pairwise F_{ST} values including the Estonian sample site significantly differed from zero ($p < 0.001$), suggesting that pollen beetle from Estonia might be an outlier sampling site in comparison with the other genotyped sampling sites. Global F_{ST} over the Eure sampling sites and over the French sampling sites did not significantly differ from 0 (respectively, Fischer's exact test, $p = 0.79$ and $p = 0.18$). Global F_{ST} over the European countries (grouping all individuals from France in one sampling sites) was 0.0082 ($p < 0.001$) or 0.0041 ($p = 0.001$), respectively with or without the Estonian sampling site. F_{ST} between all populations and Estonian population samples was 0.0218 ($p < 0.001$).

Mantel's test for isolation by distance revealed significant positive correlations between genetic differentiation and geographical distance when all sampling sites were considered (Fig. 3; $F_{ST} / (1 - F_{ST}) = -0.0125 + 0.0029 \cdot \ln(\text{km})$; $p = 0.001$). Isolation by distance was also significant when excluding the Estonian sampling sites ($F_{ST} / (1 - F_{ST}) = -0.0058 + 0.0014 \cdot \ln(\text{km})$, $p = 0.01$), but not when only the French or the Eure sampling sites were considered ($F_{ST} / (1 - F_{ST}) = -0.0030 + 0.0008 \cdot \ln(\text{km})$, $p = 0.20$ and $F_{ST} / (1 - F_{ST}) = -0.0040 + 0.0012 \cdot \ln(\text{km})$, $p = 0.23$ respectively). Neighborhood size was not calculated on the full dataset as STRUCTURE analysis revealed that the Estonian sample site was isolated from the rest of Europe. The estimate of genetic neighborhood size $D\sigma^2$ was 57 individuals based on the European dataset excluding the Estonian sample site and 100 for all France samples pooled.

3.4 Past demographic changes

Each sampling sites were analyzed separately with BOTTLENECK. In any case, the mean gene diversities (H_E) was lower than the gene diversities (H_{eq}) expected for the same number of alleles at mutation drift equilibrium with both TPM and SMM mutation models in agreement with signals of demographical expansions. The deficit of gene diversity H_E in comparison with H_{eq} was significant in nine out of the 18 sampling sites with SMM but was not significant for the TPM (Table 2). The deficit of gene diversity was significant under SMM for the pooled sampling sites from Eure, France and overall. No significant excess of gene diversity H_E in comparison with H_{eq} was detected with the TPM model.

To shed light on the history of past demographic changes in *B. aeneus* populations, a MIGRAINE analysis was performed on a dataset pooling all the genotyped individuals from the 13 French sites among which no genetic structure was detected. Outputs from the MIGRAINE analysis were consistent with those from BOTTLENECK analyses supporting a recent population expansion. The scaled current population size ($2N\mu$: 15.9, 95% CI: [4.7-2,715]) was larger than the scaled ancestral population size ($2N_{anc}\mu$: 2.7, 95% CI: [1.7-632]), and N_{ratio} confidence interval excluded one, despite being relatively large (5.9, 95% CI: [1.2-41.2]). $D_g\mu$ was estimated to 0.02 (95% CI: [0.003-0.44]). Based on an average mutation rate of 5×10^{-4} , population expansion was estimated to begin 40 generations ago (95% CI = [5.4-1,740]) while current and ancestral population sizes were 7,970 (95% CI: [2,370-1,357,500]) and 2,930 (95% CI: [607-20,630]), respectively. Note that $pGSM$ was estimated to be 0.375, which suggest that mutation on those markers are often multistep rather than single step, and thus do not fit a SMM.

4 Discussion

We have developed a set of twelve microsatellite markers for *B. aeneus*, a coleopteran pest for which no such markers were previously available. Using this set of highly polymorphic genetic markers we genotyped 433 *B. aeneus* individuals collected in 18 different OSR fields in France and Europe. These 12 microsatellite loci were selected among 48 markers designed from an enriched DNA library using a high-throughput 454 pyrosequencing approach. The proportion of microsatellite loci selected to study *B. aeneus* population genetics was relatively low in comparison with the number of markers designed, but similar to the proportion of microsatellite loci selected for population genetic study with the same method in other coleopteran species such as the ground beetles *Carabus nemoralis*³⁵ and *Poecilus cupreus*³⁶ or the invasive western corn rootworm *Diabrotica virgifera virgifera*³⁷. Interestingly, nine out of the twelve selected microsatellite loci in *B. aeneus* amplified and seven were polymorphic in *B. viridescens*, another closely related OSR pest³⁸. These loci are likely good candidates for population genetic studies in other *Brassicogethes* species, though additional evaluation of their polymorphism and neutrality is necessary³⁹. In addition, four loci had alleles detected in *B. viridescens* that were absent in *B. aeneus*, which might be used to differentiate larvae of these two sympatric species in the OSR crops⁴⁰.

4.1. Lack of population genetic structure

Overall, the *B. aeneus* genetic structure based on microsatellites markers was low at the scale of our study area. The STRUCTURE analysis clearly supported an isolation of the Estonian samples from the rest of Europe. Genetic differentiation between all populations pairs including the Estonian sample were significant but F_{ST} values were always lower than 0.05, in agreement with little genetic differentiation⁴¹. The number of genetic clusters estimated with STRUCTURE in our dataset was three. It has been suggested that the STRUCTURE algorithm may detect non-existent clusters when geographical sampling is clumped along an IBD cline⁴² and IBD was significant at the European scale when excluding the Estonian sample site. The STRUCTURE plots was thus consistent with an overestimation of the genetic structure due to IBD as ancestry proportion of individuals assigned to the third cluster increased with eastern longitude in Austria, Poland, Sweden and Estonia. The detection of three clusters with STRUCTURE may also correspond to post glacial admixtures between *B. aeneus* populations that had differentiated in different refuges in southern Europe during the last glaciations^{43,44}. Although, in general, the mitochondrial polymorphism does not inform a lot on population demography because of recurrent adaptive evolution⁴⁵, it is very suitable to

reveal population structure because of absence of recombination⁴³. Ouvrard et al.⁶ analyzed mitochondrial DNA sequences (COI) from number of pollen beetles collected over the European continent. Interestingly, they also delimited three groups of pollen beetle populations in the Baltic and Scandinavia, in western and central Europe and in south-eastern Europe, which roughly overlaps with the genetic structure we observed with the microsatellite loci in this study. Kazachkova (2008)⁸ found an even stronger genetic structure between European populations using AFLP markers. Differences between studies in the degree of population genetic structure are likely to mainly correspond to differences in the markers used, notably because of the dominance of AFLP markers, and to differences in the geographical distributions of the analyzed population samples. At a smaller spatial scale, genetic differentiation was low and not significant among the samples from the French department Eure ($F_{ST} = 0.001$) and among the French samples ($F_{ST} = 0.003$) in agreement with the lack of population genetic structure observed in Sweden with the AFLP⁷.

4.2. *B. aeneus* demographic changes in recent history

Consistent with the lack of genetic structure, genetic diversity was similar across the different European sampling sites suggesting a common demographic history of the pollen beetles across Europe⁴⁶. Temporal variations of the effective population size were expected given the variations in OSR cropped surfaces in Europe as pollen beetles need oilseed rape buds for their oviposition². The OSR cultivated surfaces are in the 2010's at an all-time high. OSR is recent on the evolutionary time scale (1,000 years or generations of pollen beetles), and the cultivated surfaces went up 4, 145, or 8 folds in respectively 40, 80 or 150 years in France (figures likely to be generalizable at the European scale). It was reasonable to expect an increase of *B. aeneus* population not only with the late increase of OSR surfaces but even more drastically since the middle age with the apparition of OSR crops¹⁴. The hypothesis of a demographic expansion of *B. aeneus* population was confirmed by the results of both the BOTTLENECK and MIGRAINE analyses. Point estimates of the demographic parameters computed with MIGRAINE supported a limited expansion of the pollen beetle populations of about 6 folds in the last 40 years. However, we should take into consideration large CI's that encompass very high current population size and much older start of expansion that might result from successive expansion and contraction of *B. aeneus* population in the recent history. For example, before insecticide resistance became widespread in the 2000s¹⁶, pesticides might have limited the expansion of pollen beetle populations, and increased population genetic structuration⁴⁷. Nevertheless, Kazachkova et al.⁷ did not found any effect

of pesticides on *B. aeneus* population structure, which can be understood if reduction in population size due to insecticide uses was no greater than increase in population size because of larger availability of OSR resources.

MIGRAINE estimates of the current *B. aeneus* effective population size in France ranged between 2,370 and 1,357,500 individuals in France, which correspond to approximately 1.5 to 850 individuals per kha of OSR crop¹³. These genetic estimates of the pollen beetle density may seem high, but remain extremely low in comparison with the ecological density of pollen beetles reported in the literature, which are about 2.5 adult individuals per plant which corresponds to having more adults in an OSR field than the maximum genetic estimate of the population size for France⁴⁸⁻⁵⁰. Such discrepancies between genetic and ecological estimates of the pollen density may be induced by huge variances of reproductive success between pollen beetle individuals and drastic changes of pollen beetle densities between years and between OSR crop production regions. Genetic neighborhood sizes estimated based on isolation by distance patterns ranged between 50 to 100 individuals and were quite high in comparison with neighborhood sizes reported in insect taxa³⁰. However, this number of individuals is still extremely small given the number of individuals per field and the known dispersal over several kilometers of the adults⁵¹.

5 Conclusion

The high variability observed in the 12 developed microsatellite loci indicates they are useful for measuring genetic patterns in *B. aeneus* populations. We confirmed the weak genetic structure of pollen beetle populations in Europe and the absence of genetic structure in France. Pollen beetle population were mainly structured by isolation by distance at the European continental scale. We also found the signature of an expansion of the *B. aeneus* population, which likely correspond to the OSR apparition in the middle age and the rapid expansion of the OSR cropping area in Europe since the Second World War. In any cases, given the unity of the population at the European scale and the important dispersal according to ecological studies, some gene flow likely occurs at the continental scale, implying that the managements of pollen beetle populations in OSR fields and evolution of insecticide resistances, including area-wide management of *B. aeneus* habitats, OSR in crop rotation, and insecticide interdictions, should be considered at the European continental scale. Finally, this case study shows how inferring current demographical characteristics based on molecular markers is challenging for insect pests because of their huge population sizes and demographic fluctuations in the recent past.

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References

- 1 Zimmer CT, Maiwald F, Schorn C, Bass C, Ott M-C, and Nauen R, A *de novo* transcriptome of European pollen beetle populations and its analysis, with special reference to insecticide action and resistance: Next-generation sequencing of the pollen beetle, *Insect Mol Biol* **23**:511–526 (2014).
- 2 Williams IH, ed., *Biocontrol-Based Integrated Management of Oilseed Rape Pests*, Springer Netherlands, Dordrecht (2010).
- 3 Gagic V, Riggi LG, Ekbom B, Malsher G, Rusch A, and Bommarco R, Interactive effects of pests increase seed yield, *Ecol Evol* **6**:2149–2157 (2016).
- 4 Mauchline AL, Cook SM, Powell W, Chapman JW, and Osborne JL, Migratory flight behaviour of the pollen beetle *Meligethes aeneus*, *Pest Manag Sci* (2017).
- 5 Taimr L, Sedivy J, Bergmannova E, and Hanker I, Further experience obtained in studies on dispersal flights of *Meligethes aeneus* F., marked with P23 (coleoptera), *Acta Entomol Bohemoslovica* **64**:325–332 (1967).
- 6 Ouvrard P, Hicks DM, Mouland M, Nicholls JA, Baldock KCR, Goddard MA, *et al.*, Molecular taxonomic analysis of the plant associations of adult pollen beetles (Nitidulidae: Meligethinae), and the population structure of *Brassicogethes aeneus*, *Genome* **59**:1101–1116 (2016).
- 7 Kazachkova N, Meijer J, and Ekbom B, Genetic diversity in pollen beetles (*Meligethes aeneus*) in Sweden: role of spatial, temporal and insecticide resistance factors, *Agric For Entomol* **9**:259–269 (2007).
- 8 Kazachkova N, Meijer J, and Ekbom B, Genetic diversity in European pollen beetle, *Meligethes aeneus* (Coleoptera: Nitidulidae), populations assessed using AFLP analysis, *Eur J Entomol* **105**:807–814 (2008).

- 9 Wang IJ, Recognizing the temporal distinctions between landscape genetics and
phylogeography, *Mol Ecol* **19**:2605–2608 (2010).
- 10 Goldstein D and Schlotterer C, *Microsatellites: evolution and applications*, Oxf Univ
Press Oxf (1999).
- 11 Storfer A, Murphy MA, Evans JS, Goldberg CS, Robinson S, Spear SF, *et al.*, Putting the
“landscape” in landscape genetics, *Heredity* **98**:128–142 (2007).
- 12 Doré C, *Histoire et amélioration de cinquante plantes cultivées*, Institut national de la
recherche agronomique, Paris (2006).
- 13 Desrier M, *L’agriculture française depuis cinquante ans : des petites exploitations
familiales aux droits à paiement unique.*, Agric Nouv Defis, 2007.
<http://agreste.agriculture.gouv.fr/IMG/pdf/AGRIFRA07c-2.pdf>.
- 14 Hokkanen HM, The making of a pest: recruitment of *Meligethes aeneus* onto oilseed
Brassicas, *Entomol Exp Appl* **95**:141–149 (2000).
- 15 Węgorzek P, Mrówczyński M, and Zamojska J, Resistance of pollen beetle (*Meligethes
aeneus* F.) to selected active substances of insecticides in Poland, *J Plant Prot Res*
49:119–128 (2009).
- 16 Hansen LM, Insecticide-resistant pollen beetles (*Meligethes aeneus* F) found in
Danish oilseed rape (*Brassica napus* L) fields, *Pest Manag Sci* **59**:1057–1059 (2003).
- 17 Heimbach U and Müller A, Incidence of pyrethroid-resistant oilseed rape pests in
Germany: Pyrethroid-resistant oilseed rape pests in Germany, *Pest Manag Sci*
69:209–216 (2013).
- 18 Audisio P, *Coleoptera : Nitidulidae-Kateretidae*, Calderini Bologna (1993).
- 19 Malausa T, Gilles A, Meglécz E, Blanquart H, Duthoy S, Costedoat C, *et al.*, High-
throughput microsatellite isolation through 454 GS-FLX Titanium pyrosequencing of
enriched DNA libraries, *Mol Ecol Resour* **11**:638–644 (2011).
- 20 Rozen S and Skaletsky H, Primer3 on the WWW for General Users and for Biological
Programmers, *Bioinforma Methods Protoc*:365–386 (1999).
- 21 Schuelke M, An economic method for the fluorescent labeling of PCR fragments, *Nat
Biotechnol* **18**:233–234 (2000).
- 22 Rousset F, genepop’007: a complete re-implementation of the genepop software for
Windows and Linux, *Mol Ecol Resour* **8**:103–106 (2008).
- 23 Nei M, Estimation of average heterozygosity and genetic distance from a small
number of individuals, *Genetics* **89**:145–163 (1978).
- 24 Kalinowski ST, hp-rare 1.0: a computer program for performing rarefaction on
measures of allelic richness, *Mol Ecol Notes* **5**:187–189 (2005).
- 25 Pritchard JK, Stephens M, and Donnelly P, Inference of population structure using
multilocus genotype data, *Genetics* **155**:945–959 (2000).
- 26 Wang J, The computer program Structure for assigning individuals to populations:
easy to use but easier to misuse, *Mol Ecol Resour* (2016).
- 27 Kopelman NM, Mayzel J, Jakobsson M, Rosenberg NA, and Mayrose I, CLUMPAK : a
program for identifying clustering modes and packaging population structure
inferences across K, *Mol Ecol Resour* **15**:1179–1191 (2015).
- 28 Evanno G, Regnaut S, and Goudet J, Detecting the number of clusters of individuals
using the software structure: a simulation study, *Mol Ecol* **14**:2611–2620 (2005).
- 29 Earl DA and vonHoldt BM, STRUCTURE HARVESTER: a website and program for
visualizing STRUCTURE output and implementing the Evanno method, *Conserv Genet
Resour* **4**:359–361 (2012).

- 30 Rousset F, Genetic differentiation and estimation of gene flow from F-statistics under
isolation by distance, *Genetics* **145**:1219–1228 (1997).
- 31 Wright S, Isolation by distance under diverse systems of mating, *Genetics* **31**:39–59
(1946).
- 32 Guillot G, Leblois R, Coulon A, and Frantz AC, Statistical methods in spatial genetics,
Mol Ecol **18**:4734–4756 (2009).
- 33 Cornuet JM and Luikart G, Description and power analysis of two tests for detecting
recent population bottlenecks from allele frequency data, *Genetics* **144**:2001–2014
(1996).
- 34 Leblois R, Pudlo P, Néron J, Bertaux F, Reddy Beeravolu C, Vitalis R, *et al.*, Maximum-
Likelihood Inference of Population Size Contractions from Microsatellite Data, *Mol
Biol Evol* **31**:2805–2823 (2014).
- 35 Brouat C, Mondor-Genson G, Audiot P, Sennedot F, Lesobre L, and Rasplus J-Y,
Isolation and characterization of microsatellite loci in the ground beetle *Carabus
nemoralis* (Coleoptera, Carabidae), *Mol Ecol Resour* **2**:119–120 (2002).
- 36 Marrec R, Ruault S, Ribout C, Plantegenest M, Gauffre B, and others, Isolation and
characterization of eleven polymorphic microsatellite markers from the beneficial
carabid beetle, *Poecilus cupreus* (Coleoptera: Carabidae), and genetic structuring
among three populations from western France, *Eur J Entomol* **111**:726–730 (2014).
- 37 Kim KS and Sappington TW, Polymorphic microsatellite loci from the western corn
rootworm (Insecta: Coleoptera: Chrysomelidae) and cross-amplification with other
Diabrotica spp, *Mol Ecol Notes* **5**:115–117 (2005).
- 38 Mason PG, Olfert O, Sluchinski L, Weiss RM, Boudreault C, Grossrieder M, *et al.*,
Actual and potential distribution of an invasive canola pest, *Meligethes viridescens*
(Coleoptera: Nitidulidae), in Canada, *Can Entomol* **135**:405–413 (2003).
- 39 Tereba A and Niemczyk M, Variability and Cross-Species Amplification of
Microsatellite Loci in *Melolontha melolontha* and *Melolontha hippocastani*
(Coleoptera, Scarabaeidae), *Entomol News* **127**:192–197 (2017).
- 40 Audisio P, De Biase A, Romanelli P, Cristina Angelici M, Ketmaier V, and De Matthaeis
E, Molecular re-examination of the taxonomy of the *Meligethes viridescens* species
complex (Coleoptera: Nitidulidae), *Biochem Syst Ecol* **28**:1–13 (2000).
- 41 Hartl DL, Clark A., and Clark A., Principles of population genetics, Sunderland Sinauer
Assoc **116** (1997).
- 42 Frantz AC, Cellina S, Krier A, Schley L, and Burke T, Using spatial Bayesian methods to
determine the genetic structure of a continuously distributed population: clusters or
isolation by distance?, *J Appl Ecol* **46**:493–505 (2009).
- 43 Franck P, Garnery L, Celebrano G, Solignac M, and Cornuet J-M, Hybrid origins of
honeybees from Italy (*Apis mellifera ligustica*) and Sicily (*A. m. sicula*), *Mol Ecol*
9:907–921 (2000).
- 44 Dennis RLH, Williams WR, and Shreeve TG, A multivariate approach to the
determination of faunal structures among European butterfly species (*Lepidoptera*:
Rhopalocera), *Zool J Linn Soc* **101**:1–49 (1991).
- 45 Bazin E, Population Size Does Not Influence Mitochondrial Genetic Diversity in
Animals, *Science* **312**:570–572 (2006).
- 46 Frankham R, Relationship of genetic variation to population size in wildlife, *Conserv
Biol* **10**:1500–1508 (1996).

- 47 Giles B. and Goudet J, A case study of genetic structure in a plant metapopulation
differentiation., *Metapopulation Biology: Ecology, Genetics, and Evolution*, Hanski I,
Gilpin ME (1997).
- 48 Kaasik R, Kovács G, Mölder J, Treier K, Vaino L, and Veromann E, The impact of semi-
natural habitats on the abundance of pollen beetle adults on winter oilseed rape
fields, *IOBC-WPRS Bull* **104**:85–89 (2014).
- 49 Rusch A, Valantin-Morison M, Sarthou JP, and Roger-Estrade J, Effect of crop
management and landscape context on insect pest populations and crop damage,
Agric Ecosyst Environ **166**:118–125 (2013).
- 50 Schneider G, Krauss J, Riedinger V, Holzschuh A, and Steffan-Dewenter I, Biological
pest control and yields depend on spatial and temporal crop cover dynamics, ed. by
Manning P, *J Appl Ecol* **52**:1283–1292 (2015).
- 51 Juhel A., Barbu C, Franck P, Valantin-Morison M, Roger-Estrade J, Butier A, *et al.*,
Characterization of the pollen beetle, *Brassicogethes aeneus*, dispersal from
woodlands to winter oilseed rape fields, *PLoS ONE*.

Figure Legends

Fig. 1: Geographical positions of the 18 *B. aeneus* population samples. a) Population samples from the Eure department. b) Population samples from France. c) Population samples from other countries in Europe. Details on the population locations are in Table 2.

Fig. 2: Summary of the results based on the STRUCTURE analysis for $K = 2$ and $K = 3$: plots indicate proportional assignment of individuals (bars) to the colour-coded genetic clusters. The population of origin is indicated on the x-axis.

Fig. 3: Isolation by distance among European populations of *Brassicogethes aeneus*. Pairwise comparison between the Eure population samples (Squares), between the French population samples (circles), between population samples from different countries (triangles), between populations including the Estonian population (diamonds). Solid line: all populations, $F_{ST} / (1 - F_{ST}) = -0.0125 + 0.0029 * \ln(\text{km})$ ($p < 0.001$). Long dash line: all populations except the Estonian sample, $F_{ST} / (1 - F_{ST}) = -0.0058 + 0.0014 * \ln(\text{km})$, ($p < 0.01$). Dashed line: French populations, ($p > 0.05$). Dotted line: Eure populations ($p > 0.05$).

Tables

Table 1: Characterization of the genetic variation at 21 polymorphic microsatellite loci based on 24 *Brassicogethes aeneus* samples collected in an oil seed rape (*Brassica napus*) field at location 1. The name of each locus was underlined. Accession Number (Acc. No.), sequences

of the forward (F) and reverse (R) primers, motif and number of repeat in the sequenced microsatellite allele, and size of the sequenced microsatellite allele were reported for each locus. Number of alleles (A), range of allele sizes, estimates of the proportion of null alleles, of observed (H_O) and expected (H_E) proportion of heterozygotes were reported for each locus in the *B. aeneus* population sample from location 1.

Locus								
Acc. No.	Primer sequences (5'-3')	Repeat motif	Size (bp)	A	Size range	Null alleles	H_E	H_O
<u>Ma-CEALO</u>	F: TTTCATTAAGCAACCTGTCG	(TC) ₆	222	3	206-222	0.00	0.17	0.18
MF321854	R: GTGAGAGTAAGTTAAAGGCG							
<u>Ma-D3QFM</u>	F: GGAGCACGTAGCAGGAC	(AC) ₈	103	6	93-109	0.00	0.44	0.43
MF321855	R: GCTGCTGCGTAATTATAGTG							
<u>Ma-DCH30</u>	F: CGCAGATCTAAATTCGTGTG	(AC) ₇	117	5	111-125	0.00	0.72	0.87
MF321862	R: GTATAGCGAAACAACAAGTGC							
<u>Ma-DDEYS</u>	F: GTACACCGAGAGGCTTTGTC	(CA) ₇	128	5	122-134	0.00	0.62	0.67
MF321863	R: GACTGTTCGGCTAGTTTTTATG							
<u>Ma-EL7YR</u>	F: CAGGATGATTTTCAGTGGAG	(GGA) ₉	193	4	184-193	0.00	0.58	0.71
MF321871	R: CGGAAGAGTTGTTTTGTATG							
<u>Ma-ESPVO</u>	F: GTTAGGATATGAATGTTTCTGC	(CA) ₈	134	9	130-160	0.00	0.79	0.86
MF321873	R: CATATGCGACATCGTTGG							
<u>Ma-ESVIZ</u>	F: GTCAGTTGTTTGGCTTATTG	(GAA) ₇	110	7	104-116	0.00	0.53	0.50
MF321874	R: CAGGAACTCGAACAAAGC							
<u>Ma-EPL2N</u>	F: CCGACTTATCAGGTGTATGG	(GGA) ₁₁	171	8	177-198	0.03	0.77	0.70
MF321872	R: CGTTCGACGTTGTGTTACC							
<u>Ma-DOM5T</u>	F: GGGAGAGTGATGTACCTTTG	(TG) ₉	153	5	143-153	0.07	0.65	0.52
MF321866	R: GGCAAGATAACTCAGATCC							
<u>Ma-EB7XX</u>	F: CTTACTCGCTCGTCCTATATC	(AC) ₇	238	8	218-232	0.08	0.85	0.70
MF321867	R: CACCTATCGTGCAGATCAC							
<u>Ma-C4QRG</u>	F: AAGAGTATAAGTCGTCGAGC	(CT) ₆	305	3	301-305	0.08	0.65	0.50
MF321857	R: AGTGGATGTAGAGAAATAGTGG							
<u>Ma-DM3QY</u>	F: CATGTAAGCTATTTTGGGACG	(AC) ₁₂	195	7	183-197	0.09	0.77	0.61
MF321865	R: CTATTTGCTTTGCTTGGATGC							
<u>Ma-C6CAE</u>	F: CCTCTACGTCATGGTATGG	(GT) ₇	126	4	121-129	0.23	0.49	0.17
MF321859	R: CCAAGATTAGGTCCACTCG							
<u>Ma-EGYLO</u>	F: TCAAGTCTGACAACCAAAAG	(GA) ₇	172	2	168-190	0.15	0.41	0.22
MF321868	R: GTTTCTGATTCGTTCTTGTC							
<u>Ma-C5QWM</u>	F: TATTATGCTCCACCATTAGG	(AC) ₈	257	4	250-256	0.22	0.67	0.31
MF321858	R: CCTTCAACTGTAATCAAAGC							
<u>Ma-D904J</u>	F: GGAGGGTCAGAAGAGTTTG	(AAG) ₇	143	6	123-139	0.21	0.71	0.36

MF321860	R: GTGTACGTGTTATAAGGCTGTG								
<i>Ma-DCGTK</i>	F: ATCGTAGCCATCTATTGAGC	(CTT) ₁₂	162	9	159-180	0.17	0.62	0.36	
MF321861	R: TGGACCTCTTGGTATATTGG								
<i>Ma-EJCJJ</i>	F: CCGACAAGTGCATTACG	(AG) ₈	176	5	154-170	0.21	0.70	0.35	
MF321869	R: CAATAGACACCATCAATTAGG								
<i>Ma-EL5EB</i>	F: CATAGTCTGTAAAGCATGTTG	(TC) ₇	295	3	295-311	0.28	0.51	0.10	
MF321870	R: CAAGAAACTTGACACTTAAATC								
<i>Ma-C2D1M</i>	F: CCAAGAAAGGGAACAGG	(AG) ₈	313	3	282-313	0.18	0.46	0.18	
MF321856	R: CAGTAACCATAGCTCGACAC								
<i>Ma-DJDOQ</i>	F: ACGTTTAATTAGTTGGTTGG	(GA) ₁₃	119	3	153-163	0.24	0.43	0.11	
MF321864	R: TATTGACGAGCTAATTTTGG								

Table 2: Mean genetic variability (\pm standard deviation) over 12 microsatellite loci in 18 *B. aeneus* population samples. Latitude (Lat) and longitude (Long) of the location of the population sample, number of individuals genotyped per population sample (N), allelic richness (A_r) computed for $n=16$, observed proportion of heterozygous genotypes (H_o), observed gene diversity (H_E) and expected gene diversity (H_{eq}) at the mutation-drift equilibrium under the Two-Phase mutations (TPM). Significantly lower H_E values than H_{eq} under the SMM were underlined.

Site	Location	Latitude	Longitude	n	A_r	H_o	H_E	H_{eq} (TPM)	H_{eq} (SMM)
1	Eure 1	48.864	1.243	25	5.40 (± 1.75)	0.67 (± 0.21)	<u>0.62 (± 0.18)</u>	0.65 (± 0.11)	0.72 (± 0.07)
2	Eure 2	48.923	1.213	22	5.06 (± 1.59)	0.61 (± 0.17)	<u>0.60 (± 0.20)</u>	0.63 (± 0.10)	0.70 (± 0.08)
3	Eure 3	48.945	1.465	24	5.63 (± 2.35)	0.63 (± 0.16)	0.63 (± 0.18)	0.66 (± 0.17)	0.71 (± 0.08)
4	Eure 4	48.973	1.421	23	5.43 (± 1.83)	0.57 (± 0.16)	0.58 (± 0.22)	0.66 (± 0.11)	0.71 (± 0.07)
5	Eure 5	48.986	1.428	23	5.07 (± 1.74)	0.62 (± 0.16)	0.58 (± 0.19)	0.64 (± 0.13)	0.69 (± 0.08)
6	Eure 6	48.972	1.448	24	5.23 (± 1.78)	0.59 (± 0.14)	<u>0.54 (± 0.17)</u>	0.63 (± 0.15)	0.69 (± 0.08)
	Eure Total	-	-	138	5.30 (± 1.66)	0.63 (± 0.17)	<u>0.63 (± 0.17)</u>	0.65 (± 0.11)	0.75 (± 0.09)
7	Eure&Loire	48.299	1.245	26	5.01 (± 1.35)	0.61 (± 0.15)	<u>0.60 (± 0.18)</u>	0.64 (± 0.11)	0.69 (± 0.08)
8	Cher	47.217	2.199	16	5.50 (± 2.43)	0.61 (± 0.22)	0.63 (± 0.23)	0.64 (± 0.17)	0.69 (± 0.09)
9	Aube	48.259	3.845	30	5.08 (± 1.98)	0.58 (± 0.21)	<u>0.55 (± 0.21)</u>	0.62 (± 0.14)	0.69 (± 0.10)
10	Maine&Loire	47.664	-0.789	25	5.42 (± 2.03)	0.63 (± 0.17)	0.65 (± 0.19)	0.64 (± 0.14)	0.70 (± 0.08)
11	Ile&Vilaine	48.112	-1.776	45	5.24 (± 1.83)	0.63 (± 0.18)	<u>0.61 (± 0.20)</u>	0.64 (± 0.12)	0.72 (± 0.07)
12	Haute-Garonne	43.408	1.646	24	5.44 (± 1.56)	0.63 (± 0.19)	0.59 (± 0.18)	0.67 (± 0.09)	0.73 (± 0.07)
13	Lot&Garonne	44.180	0.530	24	5.50 (± 1.61)	0.63 (± 0.20)	<u>0.66 (± 0.15)</u>	0.67 (± 0.10)	0.73 (± 0.07)
	France Total	-	-	312	5.32 (± 1.67)	0.63 (± 0.21)	<u>0.63 (± 0.18)</u>	0.68 (± 0.07)	0.78 (± 0.05)
14	Switzerland	47.431	8.519	28	5.97 (± 2.19)	0.63 (± 0.22)	<u>0.64 (± 0.20)</u>	0.68 (± 0.16)	0.69 (± 0.11)
15	Austria	48.208	16.374	30	5.65 (± 2.43)	0.63 (± 0.23)	0.59 (± 0.22)	0.65 (± 0.18)	0.69 (± 0.12)
16	Poland	52.187	18.809	24	5.22 (± 1.93)	0.63 (± 0.24)	0.60 (± 0.18)	0.63 (± 0.14)	0.69 (± 0.13)
17	Sweden	55.658	13.083	21	5.18 (± 1.53)	0.63 (± 0.25)	0.57 (± 0.17)	0.63 (± 0.15)	0.69 (± 0.14)
18	Estonia	58.343	26.527	24	4.43 (± 1.54)	0.63 (± 0.26)	0.59 (± 0.22)	0.59 (± 0.15)	0.69 (± 0.15)
	Europe Total	-	-	433	5.38 (± 1.79)	0.63 (± 0.27)	<u>0.63 (± 0.18)</u>	0.68 (± 0.08)	0.79 (± 0.06)

Figures

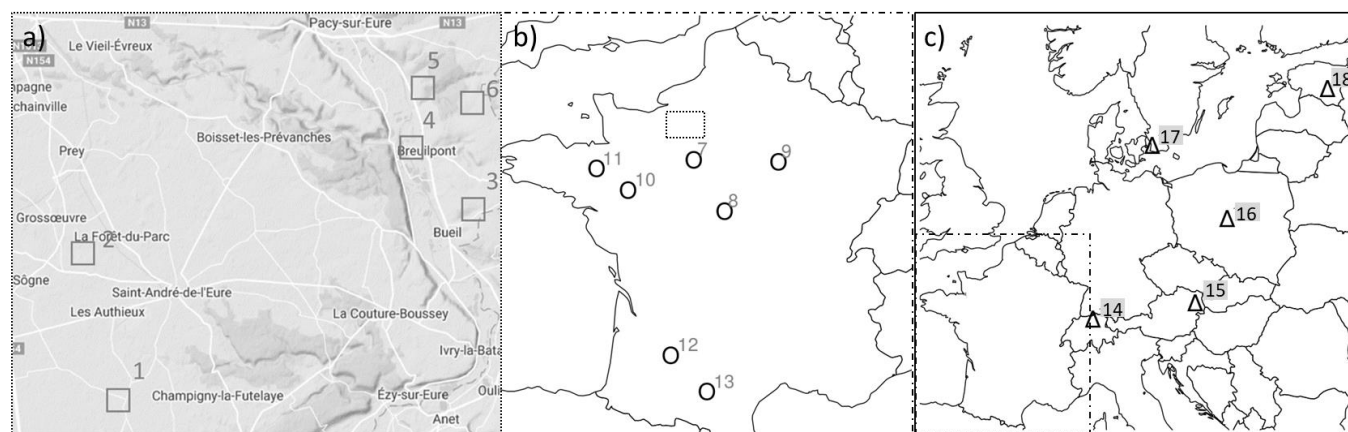
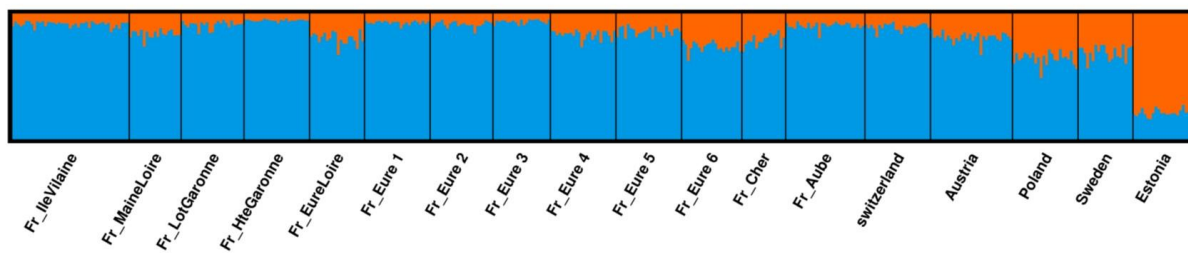
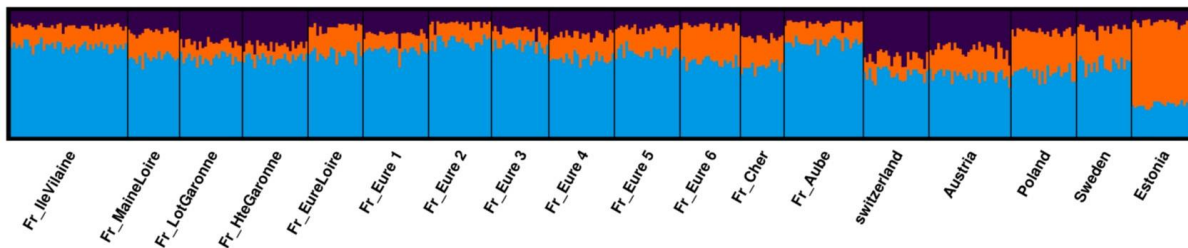


Fig. 1

K = 2



K = 3

**Fig. 2**

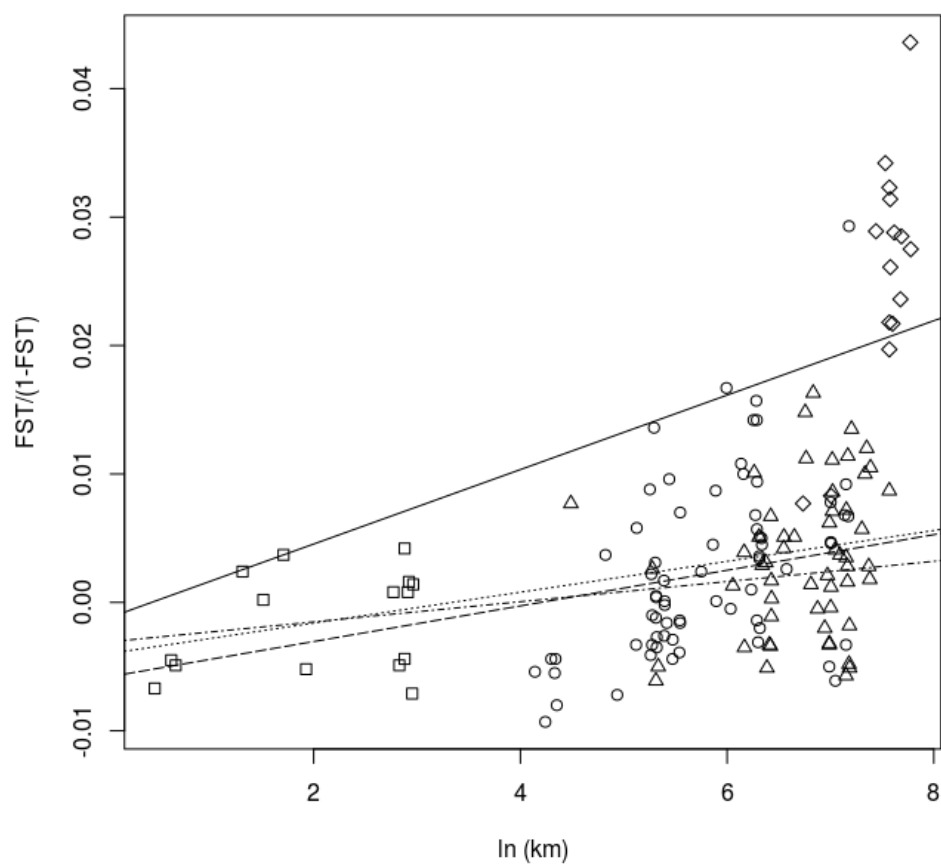


Fig. 3

Supporting Information

Table S1: Polymorphic microsatellite loci selected in this study with reference to the labelled dye of the forward primer and the molar concentration (C) of each locus each multiplex. 6-FAM (6-carboxyfluorescein), HEX (hexachloro-fluoresceine), TAMRA (carboxy-tetramethyl-rhodamine), ATTO 565 (Rhodamine dyesclass).

Locus ID	Dye	C (nM)
Multiplex 1		
<i>Ma-D3QFM</i>	6-FAM	0.07 μ M
<i>Ma-ESPVQ</i>	Tamra	0.50 μ M
<i>Ma-DM3QY</i>	Hex	0.20 μ M
<i>Ma-C4QRG</i>	Hex	0.15 μ M
<i>Ma-ESVIZ</i>	Atto-565	0.15 μ M
<i>Ma-EPL2N</i>	Atto-565	0.30 μ M
Multiplex 2		
<i>Ma-DDEYS</i>	6-FAM	0.05 μ M
<i>Ma-DCH30</i>	Tamra	0.25 μ M
<i>Ma-EL7YR</i>	Tamra	0.50 μ M
<i>Ma-DQM5T</i>	Hex	0.15 μ M
<i>Ma-EB7XX</i>	Hex	0.10 μ M
<i>Ma-CEALQ</i>	Atto-565	0.15 μ M

Table S2: Characterization of genetic variation in eight *Brassicogethes viridescens* individuals in twelve *B. aeneus* microsatellite. *B. viridescens* individuals were collected in a seed rape field at site14. Sizes of the PCR products, number of alleles per locus and allele specific of *B. viridescens* but not *B. aeneus* were reported.

Locus	Number of alleles	Size range	Alleles only present in <i>B. viridescens</i>
<i>Ma-C4QRG</i>	2	313-315	313, 315
<i>Ma-CEALQ</i>	2	222-224	/
<i>Ma-EL7YR</i>	1	175	/
<i>Ma-EB7XX</i>	/	/	/
<i>Ma-D3QFM</i>	1	91	91
<i>Ma-DCH30</i>	/	/	/
<i>Ma-DDEYS</i>	3	110-116	110, 112
<i>Ma-DM3QY</i>	/	/	/
<i>Ma-DQM5T</i>	5	151-161	155, 157
<i>Ma-EPL2N</i>	3	161-170	/
<i>Ma-ESPVQ</i>	2	132-134	/
<i>Ma-ESVIZ</i>	2	113-116	/

Table S3: Summary of the STRUCTURE analysis performed on the full dataset for a range K ranging between 1 and 20. The highest ΔK value and the two highest means $\text{LnP}(K)$ are in bold font. The posterior probabilities of $K=2$ and $K=3$ are 0.001 and 0.999 respectively.

K	Reps	Mean $\text{LnP}(K)$	Stddev $\text{LnP}(K)$	$\text{Ln}'(K)$	$ \text{Ln}''(K) $	ΔK
1	10	-14021.40	0.00	—	—	—
2	10	-13995.95	13.66	25.45	18.33	1.34
3	10	-13988.83	9.82	7.12	101.91	10.38
4	10	-14083.62	39.55	-94.79	65.75	1.66
5	10	-14112.66	47.01	-29.04	41.65	0.89
6	10	-14183.35	61.23	-70.69	26.97	0.44
7	10	-14227.07	71.71	-43.72	50.82	0.71
8	10	-14321.61	74.66	-94.54	145.18	1.94
9	10	-14270.97	84.94	50.64	35.55	0.42
10	10	-14255.88	81.65	15.09	61.35	0.75
11	10	-14302.14	122.57	-46.26	56.69	0.46
12	10	-14291.71	96.92	10.43	6.42	0.07
13	10	-14274.86	88.99	16.85	67.53	0.76
14	10	-14325.54	135.01	-50.68	81.85	0.61
15	10	-14294.37	143.59	31.17	1.07	0.01
16	10	-14264.27	87.80	30.10	95.09	1.08
17	10	-14329.26	156.77	-64.99	110.42	0.70
18	10	-14283.83	95.97	45.43	36.06	0.38
19	10	-14274.46	85.56	9.37	40.50	0.47
20	10	-14224.59	119.09	49.87	—	—

Table S4. Pairwise F_{ST} estimates (left part) and significant F_{ST} values (right part) between 18 population samples of *Brassicogethes aeneus* (for details on locations and code see Table 2). Bold character indicate significant F_{ST} values.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
1	/	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	*	***	NS	***
2	0.0077	/	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	***
3	0.0030	0.0010	/	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	*	NS	***
4	0.0099	0.0164	0.0037	/	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	***	*	***
5	-0.0016	-0.0033	-0.0031	0.0107	/	NS	NS	NS	NS	NS	NS	NS	NS	NS	*	NS	NS	***
6	-0.0044	0.0031	-0.0033	0.0100	-0.0054	/	NS	NS	NS	NS	NS	NS	NS	NS	*	NS	*	***
7	-0.0029	-0.0035	-0.0035	0.0067	-0.0094	-0.0053	/	NS	NS	NS	NS	NS	NS	NS	NS	***	NS	***
8	0.0070	-0.0026	0.0066	0.0154	-0.0044	0.0042	0.0008	/	NS	NS	NS	NS	NS	NS	NS	***	NS	***
9	-0.0039	0.0017	0.0017	0.0140	-0.0055	0.0016	0.0008	0.0002	/	NS	NS	NS	NS	NS	NS	*	NS	***
10	-0.0014	0.0001	0.0003	0.0093	-0.0081	-0.0071	-0.0049	0.0037	-0.0067	/	NS	NS	NS	NS	NS	NS	NS	***
11	-0.0016	-0.0002	-0.0011	0.0057	-0.0044	0.0014	-0.0044	0.0023	-0.0045	-0.0049	/	NS	NS	NS	NS	NS	NS	***
12	0.0024	0.0095	0.0013	0.0001	-0.0073	0.0026	-0.0013	0.0134	0.0005	-0.0027	-0.0061	/	NS	NS	NS	NS	NS	***
13	-0.0005	0.0045	0.0044	0.0140	-0.0042	0.0004	-0.0050	0.0088	-0.0033	-0.0010	0.0022	0.0058	/	NS	NS	*	NS	***
14	0.0051	0.0051	0.0042	0.0026	0.0036	0.0050	0.0029	0.0051	0.0051	-0.0020	0.0034	-0.0036	0.0087	/	NS	***	NS	***
15	0.0134	0.0072	0.0067	0.0113	0.0085	0.0070	0.0110	0.0077	0.0082	0.0012	0.0046	0.0021	0.0161	-0.0051	/	NS	*	***
16	0.0099	0.0057	0.0018	0.0104	-0.0052	0.0016	0.0028	0.0091	0.0035	-0.0057	-0.0033	-0.0049	0.0041	0.0014	0.0039	/	NS	***
17	-0.0018	0.0066	0.0028	0.0118	-0.0061	0.0046	-0.0004	0.0062	-0.0032	-0.0034	-0.0051	0.0037	-0.0020	-0.0005	0.0146	-0.0014	/	***
18	0.0277	0.0230	0.0267	0.0418	0.0212	0.0254	0.0305	0.0313	0.0214	0.0194	0.0087	0.0280	0.0331	0.0281	0.0285	0.0077	0.0110	/