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1 **IL-17c is involved in olfactory mucosa responses to**
2 **Poly(I:C) mimicking virus presence**

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10

11 **Keywords:** interleukin-17c, neuromodulation, neurovirology, Poly(I:C), ATP,
12 neuroimmune interaction, olfactory mucosa, olfaction

13

14 **Abbreviations:**

15 OEC, olfactory ensheathing cells; OM, olfactory mucosa; OSN, olfactory sensory
16 neuron; CNS, central nervous system; IL-17c, interleukin 17c; IL-17c, mouse
17 recombinant interleukin 17c; IL-17ra, interleukin 17 receptor A; IL-17re, interleukin 17
18 receptor E; Poly(I:C), polyinosinic–polycytidylic; PPADS, pyridoxalphosphate-6-
19 azophenyl-2,4'-disulfonic acid

1 **Abstract**

2 At the interface of the environment and the nervous system, the olfactory mucosa
3 (OM) is a privileged pathway for environmental toxicants and pathogens towards the
4 central nervous system. The OM is known to produce antimicrobial and
5 immunological components but the mechanisms of action of the immune system on
6 the OM remain poorly explored. IL-17c is a potent mediator of respiratory epithelial
7 innate immune responses, whose receptors are highly expressed in the OM of mice.
8 We first characterized the presence of the IL-17c and its receptors in the OM. While
9 IL-17c was weakly expressed in the control condition, it was strongly expressed *in*
10 *vivo* after intranasal administration of polyinosinic–polycytidylic (Poly I:C), a Toll Like
11 Receptor 3 agonist, mimicking a viral infection. Using calcium imaging and
12 electrophysiological recordings, we found that IL-17c can effectively activate OM cells
13 through the release of ATP. In the longer term, intranasal chronic instillations of IL-
14 17c increased the cellular dynamics of the epithelium and promoted immune cells
15 infiltrations. Finally, IL-17c decreased cell death induced by Poly(I:C) in an OM
16 primary culture. The OM is thus a tissue highly responsive to immune mediators,
17 proving its central role as a barrier against airway pathogens.

1 Introduction

2 The olfactory system is composed of a peripheral part: the olfactory mucosa (OM)
3 which represents the sensory organ of detection of odorant molecules within the
4 nasal cavity; and a central part including the olfactory bulb (OB) and other brain
5 structures dedicated to the processing and encoding of olfactory information. Within
6 the OM, olfactory sensory neurons (OSNs) are directly exposed to a variety of
7 potentially harmful environmental agents including viruses, bacteria and
8 environmental chemicals. The axons of the olfactory nerve connect the epithelium of
9 the OM to the central nervous system at the OB *via* the lamina propria and penetrate
10 the skull at the cribriform plate level. Some microorganisms exploit this olfactory
11 pathway and access the olfactory bulb where they can spread to other brain regions
12 like hypothalamus and cortical areas as shown in animal models of infection (Dando
13 et al., 2014; Pägelow et al., 2018). However, infections of the central nervous system
14 by this pathway remain relatively rare in non-immunocompromised subjects,
15 demonstrating the effectiveness of barriers against pathogens within the nasal cavity.
16 The mucocilliary clearance system within the nasal cavity is well described and acts
17 as a very effective innate defence mechanism to eliminate incoming particles and
18 microorganisms from inhaled air (Andersen et al., 1971; Antunes, and Cohen, 2007).
19 The prevention of dissemination of pathogens into deeper tissue also rely on nasal-
20 associated lymphoid tissue (NALT), which can trigger an immune response (Gänger,
21 and Schindowski, 2018). However, very few local defence mechanisms are described
22 and the mechanisms by which immune components act in the OM are not well
23 characterized.

24 The interleukin-17 (IL-17) family is composed of six members (IL-17a, IL-17b, IL-17c,
25 IL-17d, IL-17e, and IL-17f; (Gu et al., 2013)) that participate in both acute and chronic

1 inflammatory responses. To date, IL-17c has been identified *in vitro* as an epithelial
2 cell-derived cytokine that enhances mucosal host defence responses in a unique
3 autocrine/paracrine manner (Kusagaya et al., 2013). IL-17c is implicated in the
4 protection against microbial infection and in the pathogenesis of autoimmune disease
5 including multiple sclerosis (Chang et al., 2011) and act as a mediator of respiratory
6 epithelium innate immune response (Pfeifer et al., 2013). Furthermore, a recent study
7 shows that infection of the peripheral nervous system by herpes simplex virus
8 induces the production of IL-17c (Peng et al., 2017). In this study, the authors
9 observed that IL-17c acts as a neurotrophic factor promoting neurite growth of human
10 foetal dorsal root ganglia neurons and provides a survival signal in mouse primary
11 cortical neurons. Transcriptomic studies show a strong expression of IL-17c
12 receptors (IL-17re, IL-17ra) and a potential expression of IL-17c in the OM (Ibarra-
13 Soria et al., 2014; Kanageswaran et al., 2015). Thus, we hypothesized that IL-17c
14 play a role in the immune response of the olfactory epithelium to viral infection and
15 that it may impact the sensory process in the OM by modulating the functioning of
16 OSNs in the mouse.

17 To explore the role of IL-17c in the OM, we first characterized the presence of the IL-
18 17c system in the nasal cavity of CD1 mice by immunohistochemistry and *in situ*
19 hybridization. While IL-17c was weakly expressed in the control condition, its level
20 was increased under a viral-like context using polyinosinic-polycytidylic (Poly I:C), a
21 Toll Like Receptor 3 (TLR3) agonist. To decipher the impact of IL-17c on olfactory
22 mucosa, we combined electrophysiological, molecular biology and
23 immunohistochemistry approaches using primary culture, and *ex vivo* and *in vivo*
24 experiments.

1 **Materials and methods**

2 **Animals and intranasal instillations**

3 CD1 male and female mice (*Mus musculus*) were obtained from multiparous females,
4 and housed in our local animal care facilities (Unité Expérimentale Animalerie
5 Rongeurs (UEAR), Jouy-en-Josas, France) in 12 h light, 12 h dark cycles. Mothers
6 were fed standard chow *ad libitum* and had free access to tap water. All animal
7 experiments were conducted in accordance with the European Council Directive
8 2010/63/UE and our study was agreed upon by our Local Ethics Committee
9 (APAPHIS#4589). All efforts were made to minimize the number and suffering of
10 mice used. N.M, B.B, P.C holds the Individual Authorization for Performing
11 Experiments in Animals, including the animals experiments conducted in the present
12 study, provided by Préfecture des Yvelines (France), according to French and
13 European laws (agreements #78-154).

14 Intranasal delivery of Poly(I:C), of mouse recombinant IL-17c (IL-17c : 500ng/mL, 10
15 μ L per nostril) or of vehicle solution were performed in awake mice daily using a
16 specialized intranasal grip adapted from Hanson et al. (2013). Mice received
17 intranasal administration of 20 μ g Poly(I:C) sodium salt (P 4287/10 Tocris
18 Bioscience, UK) dissolved in 10 μ L of sterile saline into each nostril. The dose of
19 Poly(I:C) used was determined according to a previous study measuring the effect of
20 Poly(I:C) on the OM (Kanaya et al., 2014).

21

22 **Immunohistochemistry and OM dynamics quantification**

23 The immunohistochemistry from OM tissue sections was performed as described
24 previously (François et al., 2016). The nasal septum and turbinates were removed as
25 a block and post-fixed overnight at 4°C in 4% paraformaldehyde PBS. Blocks were

1 cryoprotected with sucrose (30%) and cryo-sectioned sagittally (14 μ m thickness).
2 Sections were stored at -80°C until use. PCNA staining requires an antigen retrieval
3 in a citrate buffer (pH=6) at 95°C for 30 min. Non-specific staining was blocked by
4 incubation in PBS containing 2% bovine serum albumin and 0.3% Triton X-100. The
5 sections were then incubated overnight with primary antibodies directed against
6 PCNA (1 : 200; mouse monoclonal PC10. GeneTex; Tebu-bio SAS, France) ;
7 cleaved caspase 3 (1: 400; rabbit polyclonal, Cell Signaling; Ozyme, France) ;
8 ionized calcium-binding adapter molecule 1 (1 : 1000; goat polyclonal; Abcam,
9 France) ; or interleukin-17 receptor A (1 : 200; goat polyclonal, R&D systems;
10 Biotechne, France). Fluorescence staining was performed using Alexa-Fluor-488-
11 conjugated goat (1: 1000; Molecular Probes; Invitrogen, France) or CyTM3-conjugated
12 AffiniPure donkey secondary antibodies (1 : 500; Jackson ImmunoResearch;
13 France). Sections were finally stained with 2 μ g/mL Hoechst 33342 for 10 min and
14 mounted in Vectashield after extensive wash in PBS.

15 Immunohistochemistry was performed on median transversal sections of the OM. For
16 all sections, we took six to eight images situated in caudal, medial or rostral area
17 from the base of the septum to cover the variability of distribution of apoptosis across
18 the OM (Vedin et al., 2009).

19 Images were taken at x100 magnification using a DMBR Leica microscope equipped
20 with an Olympus DP-50 CCD camera using CellF software (Olympus Soft Imaging
21 Solutions GmbH, OSIS, Münster, Germany), except for cleaved caspase 3 images
22 acquired with an Olympus IX71 inverted microscope equipped with an Orca ER
23 Hamamatsu cooled CCD camera (Hamamatsu Photonics France, Massy, France).

24 Images were quantified using ImageJ (Rasband, W.S., ImageJ, U. S. National
25 Institutes of Health, Bethesda, Maryland, USA, <http://imagej.nih.gov/ij/>, 1997–2012)

1 to threshold specific (François et al., 2016) cleaved caspase 3 and PCNA staining.
2 The same threshold was applied for all images arising from the same experiment.
3 The area of the olfactory epithelium (OE) was measured from the Hoechst staining
4 for cleaved caspase 3 and PCNA signal quantification. Iba1+ cells were counted
5 using the detection particle tool from ImageJ and expressed relative to the OM area
6 as described above.

7

8 **Hematoxylin and eosin (H&E) staining**

9 After rehydration, slides were stained in Hematoxylin (H-3404 Vector Laboratories)
10 for 30 s and washed with water. Slides were then transferred in an acid-alcohol
11 solution (0.5% HCl in 70% Ethanol) for 10 s and washed in water, then immersed in
12 Eosin (Sigma HT110316) for 15 s and washed again in water. Finally, slides were
13 gradually dehydrated and mounted in Eukitt (Sigma 25608-33-7). Slides were
14 digitized using NanoZoomer C9600-01 (Hamamatsu).

15

16 **In situ hybridization**

17 The *in situ* hybridization from OM tissue sections was performed with slight
18 modifications from our previously described protocols (François et al., 2013; Le
19 Bourhis et al., 2014). Briefly, the expression of IL-17c and IL-17re were assessed
20 using digoxigenin (DIG)-labelled RNA probes transcribed *in vitro* (DIG RNA labelling
21 kit from Roche) from target sequences subcloned into a pGEM-T plasmid (Promega)
22 from GenBank (IL-17C nucleotide 242–615, NM_145834.4; IL-17re nucleotide 1152-
23 1626, NM_145826). After rehydration, sections were treated with proteinase K (10
24 µg/mL) for 10 min at 37°C, washed twice in PBS and incubated in post-fixation
25 solution (4% PFA) for 5 min at 4°C. Sections were then washed in PBS for 10 min at

1 37°C and incubated in acetylation solution (0.1 M triethanolamine and 0.25%
2 anhydrous acetic acid) for 10 min. The digoxigenin-labeled probes (0.1 µg/µl) were
3 denatured beforehand at 65°C for 5 min then added to the hybridization mix (50%
4 formamide; 2.5% Dextran; 2×SSC; 5x Denhart; 50µg/ml yeast tRNA; 250µg/ml
5 salmon sperm, and 4 mM EDTA) and hybridized overnight at 55°C. Sections were
6 then sequentially washed in 2×SSC (2 ×10 min at 45°C), incubated with RNase
7 (20µg/mL), washed in 2×SSC (4x5 min at 45°C) and washed in 2×SSC – 30%
8 Formamide (1h at 60°C). Hybridized probes were revealed with NBT-BCIP as
9 substrate in detection buffer and the reaction was stopped by rinsing sections in PBS.

10

11 **Olfactory mucosa primary cultures**

12 We used a long-term primary culture of mouse OM cells based on a rat OM primary
13 culture described previously (Gouadon et al., 2010). For each experiment, two to
14 three CD1 new-born mice (P4 to P8) were sacrificed by decapitation. The entire OM
15 from turbinates and septum was dissected. After digestion and centrifugation, cells
16 were suspended in sterile HEPES-buffered Dulbecco's Modified Eagle
17 Medium/HamF12 based medium (DMEM/Ham F12; Eurobio, Les Ulis, France) and
18 10% new-born calf serum (NCS; Eurobio, Les Ulis, France). Cells were counted and
19 seeded at approximately 700 cells/mm² on glass coverslips previously coated with
20 poly-L-lysine. Cells were grown at 37°C under 5% CO₂ in the DMEM/HamF12
21 medium with 10% NCS. Cultures were then treated with one of the following
22 components: Poly(I:C) at 1mg/mL or IL-17c at 200ng/mL.

23

24

1 **Assessment of cellular death by lactate dehydrogenase release**

2 The cell death in the primary culture was assessed by quantifying the release of LDH
3 in the culture medium, as previously described (Laziz et al., 2011). Briefly, we
4 measured activities of both the extracellular and the intracellular LDH, respectively
5 from the bathing culture medium and from the primary cultured adherent cells
6 extemporaneously lysed with PBT (PBS 0.5% Triton X-100). The LDH concentration
7 was assayed by the rate of NADH absorbance decrease at 355 nm, using a
8 microplate reader (Tristar LB 941, Berthold technology).

9

10 **OM slices preparation and calcium imaging**

11 OM slices were prepared from CD1 mouse pups (P5-10) with adaptation from our
12 previous protocol based on rat (Le Bourhis et al., 2014). After the removal of the
13 lower jaw and teeth, the upper skull bone was embedded in 4% low-gelling-
14 temperature type VIIa agarose and placed in ice-cold artificial cerebrospinal fluid
15 (ACSF) solution containing (in mM): 124 NaCl, 5 KCl, 1.25 NaH₂PO₄, 2 MgSO₄, 2
16 CaCl₂, 26 NaHCO₃ and 10 glucose, saturated with 95% O₂ and 5% CO₂ at pH 7.4
17 and ~300 mOsmol/L. Coronal slices (200 μm) were cut using a vibratome (VT1200S;
18 Leica Microsystems, LEICA Microsystemes SAS, Nanterre, France), and kept
19 immersed in oxygenated ACSF at room temperature until use. IL-17c and ATP
20 impact on intracellular Ca²⁺ dynamics in the OM were monitored using the calcium-
21 sensitive dye Oregon green BAPTA-1 (2 mM stock solution in dimethyl sulphoxide),
22 mixed 1 : 1 with 20% Pluronic F-127 (Molecular Probes). After a loading and a
23 washing-out period, slices were transferred into a home-made recording chamber
24 under a 209 long-range water immersion objective (XLUMPLFL20XW, NA 0.95) of an
25 upright microscope (BX51WI, Olympus, Rungis, France), equipped with differential

1 interference contrast and an infrared video camera. The chamber was continuously
2 gravity perfused with oxygenated ACSF at a rate of 2.5–3 mL/min except when slices
3 were stimulated. Slices were stimulated for 15 seconds with a glass pipette (1M Ω)
4 filled with 10⁻⁴M ATP or 500 ng/mL IL-17c under pressure. The pipette was placed at
5 a distance of ~170 μ m from the apical part of the slice. For off-line analysis of calcium
6 transients, images were background subtracted and changes in fluorescence were
7 calculated relative to the averaged baseline (from 30 s before stimulation) and
8 expressed as $\% \Delta F/F$ ($((F - F_{baseline}) / F_{baseline}) \times 100$) using Xcellence RT software. The
9 general nonspecific purinergic receptor antagonists suramin (100 μ M) and
10 pyridoxalphosphate-6-azophenyl-2,4'-disulfonic acid (PPADS – 25 μ M) were used to
11 confirm that purinergic and IL-17c-evoked Ca²⁺ transients were mediated *via*
12 purinergic receptors. Caffeine (50mM) was used as a control to assess the
13 functionality of OM slices under antagonists treatments (Hegg, and Lucero, 2006).

14

15 **Electro-olfactogram (EOG) recording**

16 EOG recordings were performed on the OM of 2 month old mice in an opened nasal
17 cavity configuration as described earlier (François et al., 2016). The hemi-head was
18 placed in a recording chamber under an upright Olympus SZ51 stereo microscope
19 (Olympus, Rungis, France) equipped with a low magnification objective (0.8 to 4x)
20 and two MX-160 micromanipulators (Siskiyou, Inc., Grants Pass, OR, USA). The
21 hemi-head was kept under a constant flow of humidified filtered air (~1000 ml/min)
22 delivered through a 9 mm glass tube. This tube was positioned 2 cm from the
23 epithelial surface. Odour stimulations were performed by blowing air puffs (200 ms,
24 200 ml/min) through an exchangeable Pasteur pipette enclosed in the glass tube
25 containing a filter paper impregnated with 20 μ L of odorant (Sigma Aldrich, Saint-

1 Quentin Fallavier, France). Recordings were made with glass micropipettes of 4–
2 5MΩ. Simultaneous EOG recordings of the responses evoked by heptanal (1:1000 in
3 mineral oil) were recorded until their amplitudes stabilized, (i.e. at least 3 successive
4 responses displaying the same amplitude), and the last response was selected as
5 the reference. We focused on heptanal as [it gives the best results in our recordings in](#)
6 [mice, with signal ranging from 5 to 25 mV according to the dose used \(François et al.](#)
7 [2016\)](#). One endoturbinat was treated by local application of the mucosal saline
8 solution (45 mM KCl, 20 mM KC₂H₃O₂, 55 mM NaCH₃SO₄, 1 mM MgSO₄, 5 mM
9 CaCl₂, 10 mM HEPES, 11 mM glucose, 50 mM mannitol, pH 7.4, 350 mOsm
10 adjusted with mannitol (Negroni et al., 2012)), while the other endoturbinat received
11 the tested molecule (1µg/mL IL-17c (n=8)) through constant diffusion from a glass
12 micropipette of ~10µM diameter. Control and treated endoturbinates (II_b and IV) were
13 systematically alternated from one mouse hemi-head to the other. Responses were
14 recorded on both endoturbinates simultaneously every 3 min, for 30 min following
15 local treatment. Analyses were performed off-line using Clampfit 9.2 (Axon
16 Instruments). Peak amplitude following the drug application was normalized to peak
17 amplitude of the control response prior to the treatment. Results are presented as
18 mean ± standard error of the mean (SEM).

19

20 **qPCR and molecular biology**

21 Total RNA was extracted from OM primary culture cells using the Trizol method then
22 treated with DNase I. OligodT first strand cDNA were synthesized from 5 µg total
23 RNA by the Superscript II reverse transcriptase (Invitrogen) following the
24 manufacturers recommendations. cDNA templates (5 µL) were added to a 15 µl
25 reaction mixture containing 200 nM primers (sequences in **Table 1**) and SYBR Green

1 GoTaq® qPCR Master Mix (Promega, Charbonnieres, France). The expression
2 levels of target genes were measured using the CFX Connect qPCR platform
3 (BioRad, Hercules, CA, USA). A dissociation curve was carried out at the end of the
4 PCR cycle to verify the efficiency of the primers to produce a single and specific PCR
5 amplification. Quantification was achieved using the $\Delta\Delta C_t$ method. Standard controls
6 of specificity and efficiency of the qPCR assays were performed. The mRNA
7 expression was normalized to the expression level of β -actin and an efficiency
8 corrective factor was applied for each primer pair (François et al., 2016).

9 **Statistical analysis**

10 Data are expressed as mean \pm standard error of the mean (SEM). Regarding the
11 immunohistochemistry quantification, the investigator performing the quantification
12 was blind to the treatment group during analysis. Statistical significance was
13 assessed using non-parametric Mann-Whitney tests or one-way or two-way ANOVA
14 followed by Bonferroni multiple comparison post-hoc tests (GraphPad Prism
15 software, GraphPad software Inc., CA, USA). A probability value of $P \leq 0.05$ was set
16 for significant differences.

17

1 Results

2 Expression and functionality of IL-17c receptors in the OM

3 To specify the distribution of IL-17c receptors (IL-17re and IL-17ra), we first
4 performed *in situ* hybridization of IL-17re and immunofluorescent staining of IL-17ra,
5 the specific receptor subunits for IL-17c (Song et al., 2011). IL-17re expression was
6 found on the apical part of the sustentacular cells (SCs) and in axon bundles (Fig.
7 **1A**). We also detected IL-17ra expression in the same part of the OM (Fig. **1B**).
8 These data show that in the OM, the axon bundles of OSNs and SCs may be
9 responsive to IL-17c.

10 We next examined the functionality of the IL-17c system in neonatal mice using
11 calcium imaging on OM slices. Transient application of mouse recombinant IL-17c
12 (IL-17c – 500ng/mL) evoked robust calcium rises in SCs body layer (Fig. **2A**). To
13 compare the IL-17c evoked responses in SCs to the effect of a known SC modulator,
14 we used ATP to generate a calcium increase in the SCs, as described previously
15 (Hegg et al. 2003; Fig. **2B**). Given the similar response patterns observed with ATP
16 and IL-17c (Fig. **2C**), we hypothesized that stimulation with IL-17c might induce ATP
17 release. Therefore, we preincubated slices with purinergic receptors antagonists
18 (PRA - suramin 100 μ M and PPADS 25 μ M ; (Hegg et al. 2003)) and then performed
19 stimulations with ATP or IL-17c (Fig. **2B₃, A₃**). As expected, slices incubated with the
20 purinergic receptors antagonists no longer respond to stimulation with ATP (Fig.
21 **2B₃**). The preincubation of slices with PRA also specifically blocked the calcium
22 increase mediated by IL-17c. The responsiveness of the OM slices after PRA
23 treatment was assessed with 5 mM caffeine inducing calcium level rises in OM slices
24 independently of the purinergic pathway (Hegg et al., 2009). As expected, caffeine
25 elicited an increase in intracellular calcium in both OSNs and sustentacular cells

1 independently of the presence of purinergic antagonists (**Supplementary Data**
2 **Figure 1**). These results show that acute application of IL-17c is likely to induce the
3 ATP release by olfactory mucosa cells, which in turn increase intracellular calcium
4 level.

5 We next explored if this process was still present in adult mice. As OM slices can
6 only be performed on very young mice due to the solidity of the cranial bones,
7 another technique was needed. We therefore used electro-olfactogram (EOG)
8 recordings as a proxy to validate the functionality of the IL-17c system. EOG
9 recordings reflect the responses of the OM to odorant stimulation (Scott, and Scott-
10 Johnson, 2002), and ATP application on mice OM decreases odour sensitivity (Hegg
11 et al. 2003; Hegg et Lucero 2006). If IL-17c induces the release of ATP from OM cells
12 in adult mice, it should impact the odorant responses of the OM similarly as ATP
13 application. Simultaneous EOG recordings of the responses evoked by heptanal
14 were obtained from the endoturbinates IIb and IV (Fig. **3A**). One endoturbinate was
15 treated by a local application of mucus solution (MS), while the other endoturbinate
16 received the same MS containing IL-17c at 1µg/mL (Fig. **3A, B**). The evolution over
17 time of the amplitude was significantly decreased following treatment with IL-17c
18 compared to saline (Two-way ANOVA; $F_{(1,144)} = 5.66$; $P = 0.031$ Fig. **3C, D**). This
19 decrease is consistent with a release of ATP induced by the presence of IL-17c.

20 Overall, our results show that the IL-17c system is functional on *ex vivo* preparations
21 of the OM in neonatal and adult mice. They further suggest that IL-17c modulates
22 calcium signalling indirectly *via* the release of ATP.

1 **IL-17c increases cellular dynamics of the OM**

2 After highlighting the functionality of the IL-17c in an *ex vivo* system, we decided to
3 explore the impact of IL-17c on the OM *in vivo*. We thus examined whether the
4 cellular dynamics of the OM was altered in mice intranasally instilled with IL-17c. As
5 cellular dynamic of the olfactory epithelium is region dependent (Vedin et al., 2009),
6 we studied the distribution of apoptosis, proliferation, and Iba1+ immune cells
7 infiltration across the olfactory mucosa. In order to explore the effect of acute and
8 chronic treatment with IL-17c, we performed intranasal instillations of IL-17c for one
9 day and one week, respectively.

10 We first examined the effect of a single IL-17c administration on the OM dynamics.
11 24h after this nasal instillation, we did not observe any significant modification of the
12 cellular dynamic of the epithelium in terms of apoptosis (Mann-Whitney; $P = 0.519$; P
13 $= 0.774$; $P = 0.628$ in caudal, medial and rostral area respectively), cell proliferation
14 ($P = 0.533$; $P = 0.731$; $P = 0.295$ in caudal, medial and rostral area respectively)
15 and immune cell infiltrations ($P = 0.534$; $P = 0.366$; $P = 0.234$ in caudal, medial and
16 rostral area respectively) (**Supplementary Figure 2**). However, we observed a
17 significant increase of cleaved caspase 3 signal, (major component of the apoptosis
18 signalling pathway) in one week treated IL-17c mice compared to the untreated
19 control group. This increase was present in medial and rostral areas of the OM (Fig.
20 **4A₁**; Mann Whitney; $P = 0.022$ and 0.014 , respectively). We next examined the OM
21 proliferation rate by immunohistochemistry raised against PCNA (a scaffold protein
22 involved in DNA replication; (François et al., 2016)) and also observed a significant
23 increase of PCNA staining in the rostral area (Fig. **4B₁**; Mann Whitney; $P = 0.002$).
24 We finally examined the immune Iba1+ cell infiltration (specific marker of
25 monocyte/macrophage lineage, (Hasegawa-Ishii et al., 2017) in the OM. Treatment

1 with IL-17c led to an increase of immune cells infiltration in the OM in its medial and
2 rostral areas (Fig. **4C₁**; Mann Whitney; both P = 0.035).

3 Overall, these results indicate that the cellular turn-over in the OM is increased in IL-
4 17c treated mice and that several days of IL-17c treatment trigger an immune cell
5 infiltration in the OM.

6

7 **IL-17c is induced *in vivo* in the OM following Poly(I:C) stimulation mimicking** 8 **virus presence**

9 Under physiological conditions, IL-17c is expressed at a very low level in the OM
10 according to our *in situ hybridization* and qPCR experiments. As the receptors to IL-
11 17c are strongly expressed in the OM, we next hypothesized that IL-17c may be
12 released under specific situations like inflammation following bacterial or viral stimuli
13 as shown for respiratory epithelium cells *in vitro* (Pfeifer et al., 2013).
14 Polyinosinic:polycytidylic acid (Poly(I:C) is structurally similar to double-stranded RNA
15 present in part of the RNA virus family and is used experimentally to model viral
16 infections *in vivo* (Kanaya et al., 2014). Furthermore, Poly(I:C) is a potent inducer of
17 IL-17c in primary airway epithelial cells according to *in vitro* studies (Kusagaya et al.,
18 2013; Pfeifer et al., 2013). We performed intranasal administration of Poly(I:C)
19 similarly as a previous study on the innate immune responses of the OM (Kanaya et
20 al. 2014 - Fig. **5**). After a 3-day treatment, we observed degenerative changes of the
21 olfactory area treated with Poly(I:C) (Fig. **5C**). Degeneration was characterized by
22 detachment of the neuroepithelium from the basal lamina (Fig. **5C₃**) and was absent
23 in the contralateral, non-treated OM (Fig. **5C₂**). Intranasal instillations of Poly(I:C)
24 also induced a strong expression of IL-17c in the whole olfactory epithelium, the
25 nasal-associated lymphoid tissue (NALT) and particularly in the Steno's gland (also

1 known as lateral nasal gland ; Fig. **5D**). The IL-17c expression followed the time
2 course of Poly(I:C) treatment and decreased after the end of Poly(I:C) nasal
3 instillations at D7 (One way ANOVA analysis followed by Bonferroni's Multiple
4 Comparison Test; $P < 0.001$; Fig. **5B**). Together, these results indicate that IL-17c is
5 mainly expressed locally *in vivo* in the OM, NALT and lateral nasal gland under a
6 Poly(I:C) challenge.

7

8 **Intranasal instillation of IL-17C results in a decrease of expression of tight** 9 **junction related proteins**

10 Nasal administration of Poly(I:C) led concomitantly to a disturbance of OM integrity
11 and to an increase in IL-17c expression. As IL-17c plays a critical role in intestinal
12 mucosal barrier integrity (Reynolds et al., 2012), we evaluated a potential
13 involvement of IL-17c in OM barrier stability. As we previously observed that ATP
14 was likely released from OM cells in the presence of IL-17c (Fig. **2**), we also
15 evaluated if ATP could also be involved in this process. The observed shedding of
16 the OM neuroepithelium (Fig. **5C₃**) in mice treated with Poly(I:C), was absent in mice
17 treated with vehicle (Fig. **5C₂**), IL-17c (Fig. **5C₄**) or ATP (Fig. **5C₅**). We thus
18 compared the effects of Poly(I:C), IL-17c, and ATP delivered intranasally on the
19 expression of genes related to barrier integrity. We measured the expression of
20 genes related to tight junction in the OM (claudin 1, claudin 11 and tight junction
21 protein 1 – TJP1 (Steinke et al., 2008; Krishan et al., 2014)). As expected (Kanaya et
22 al., 2014; Huang et al., 2016; Gon et al., 2016), the expression of genes related to
23 tight junction formation was reduced in olfactory mucosa of Poly(I:C) treated-mice:
24 claudin 1 (Fig. **6A₁**; Mann Whitney; $P = 0.023$), claudin 11 (Fig. **6A₂**; $P = 0.028$) and
25 TJP1 (Fig. **6A₃**; $P = 0.006$). With regards to IL-17c, intranasal treatment also resulted

1 in a decrease of expression of claudin 1 (Fig. **6A₁**; P = 0.007), claudin 11 (Fig. **6A₂**; P
2 = 0.003) and TJP1 (Fig. **6A₃**; P = 0.046). Interestingly, the expression of various tight
3 junction proteins remained unchanged between vehicle and ATP-treated mice (Fig.
4 **6A** - claudin 1, P = 0.681; claudin 11, P = 0.147 and TJP1, P = 0.244).

5 Overall, our results suggest that IL-17c decreases the permeability of the
6 neuroepithelium as indicated by changes in the tight junction genes expression.

7

8 **Pretreatment and cotreatment with IL-17c modulate cell death induced by** 9 **Poly(I:C) treatment of OM primary culture**

10 IL-17c is a neurotrophic cytokine that protects peripheral nerve systems during HSV
11 reactivation *in vitro* (Peng et al., 2017). We explored its protective role in a primary
12 culture model. We selected the *in vitro* model for its reliability, as compared to the
13 variability observed *in vivo* and to avoid the endogenous production of IL-17c. We
14 tested if pretreatment with IL-17c could improve the cellular survival of a primary
15 culture of OM during a Poly(I:C) challenge. Seven days after *in vitro* development of
16 the culture, primary culture of the OM were treated with Poly(I:C) alone or with IL-17c
17 (Fig. **7A**). The level of cellular death was then evaluated by the LDH released from
18 dying cells one week after the treatments. The level of cellular death was increased
19 following Poly(I:C) treatment (Fig. **7B**; Mann Whitney; P < 0.001) while the treatment
20 with IL-17c alone did not affect LDH release (Fig. **7B**; P = 0.611). Pretreatment or co-
21 treatment with IL-17c of cell treated with Poly(I:C) led to a significant decrease of
22 LDH release (Fig **7B**; P < 0.001).

23

1 Discussion

2 Due to its position at the interface between environment and central nervous system,
3 the olfactory mucosa is an immunologically sensitive site. While OM studies are
4 focused on odorant detection, it is also a place of synthesis and secretion of
5 antimicrobial and immunological compounds (Mellert et al., 1992). However, very few
6 defence mechanisms are described and the interaction of the immunity actors within
7 the OM remains to be explored (Chen et al., 2017). Similarly, there is an important
8 body of literature for peripheral modulation of odorant detection (Lucero, 2013) but
9 very few papers focus on the potent neuromodulator effects of immune compounds
10 despite the growing evidence on neuromodulatory properties of cytokines (Vezzani,
11 and Viviani, 2015). Previous findings on primary normal human bronchial epithelial
12 cells suggest a pivotal role of IL-17c in the respiratory epithelium (Kusagaya et al.,
13 2013; Pfeifer et al., 2013). Indeed, in these studies, Poly(I:C) treatment induced IL-
14 17c expression *via* the Toll-like receptor 3, and IL-17c enhanced mucosal host
15 defence responses *via* a unique autocrine/paracrine manner. In our study, we
16 explored IL-17c presence, production and functionality on the olfactory mucosa.
17 We first report that IL-17c receptors are expressed in the OM by *in situ* hybridization
18 and immunohistochemical approaches (**Fig. 1**). The distribution patterns of IL-17re
19 and IL-17ra show the presence of receptors in SCs and in axonal bundles of OSNs.
20 The expression of IL-17c receptors in axonal parts of neurons is consistent with a
21 previous study showing that IL-17c receptors are present in sensory neurons from
22 human foetal dorsal root ganglia (Peng et al., 2017). Together these results highlight
23 the expression of IL-17c receptors in two immunologically sensitive sites as
24 sustentacular cells are directly exposed to airborne substances (Makino et al., 2009;

1 Kanaya et al., 2014) and axonal bundles connect the environment to the central
2 nervous system.

3 The expression of IL-17c was reported in transcriptomic studies (Ibarra-Soria et al.,
4 2014; Kanageswaran et al., 2015). Our qPCR and *in situ hybridization* results on
5 control animals indicate that the basal level of IL-17c expression in the Steno's gland
6 and in the OM is very low. However, its expression was induced by *in vivo* intranasal
7 instillation of Poly(I:C) in a time-dependent manner. The IL-17c expression follows
8 the time course of Poly(I:C) treatment and decreases after the end of the treatment.
9 IL-17c was mainly expressed in lateral nasal glands (LNG – also known as Steno's
10 gland; **Fig. 5**), which are described as an important site of secretion of mucus
11 components within the nasal cavity (Badonnel et al., 2009). The LNG, even if poorly
12 described in the literature, are believed to act as an immune barrier through secretion
13 of immunoglobulin A (Mellert et al., 1992) and their expression of IL-17c supports a
14 role in immunity for this gland. IL-17c is also induced by Poly(I:C) treatment *in vivo*.
15 As we observed that IL-17c is active in the OM, our results are in agreement with
16 previous evidences of autocrine and paracrine action of IL-17c (Kusagaya et al.,
17 2013).

18 Functionally, we observed that IL-17c modulates the calcium levels within seconds of
19 application *ex vivo*. A transduction pathway of cytokines linked to calcium level
20 increase is rather unusual but has been observed with IL-1 β in several models of
21 primary culture (cortical astrocytes, hypothalamus neurons and area postrema cells -
22 Holliday et Gruol 1993; De et al. 2002; Wuchert et al. 2009). We observed a very
23 similar cellular pattern of activation between IL-17c and ATP used as a positive
24 control. We hypothesized that the Ca²⁺ level rise induced by IL-17c application could
25 be indirect through the release of ATP. Such mechanism has already been observed

1 with IL-1 β inducing ATP release in rat hippocampal slices (Sperlágh et al., 2004). In
2 our study, IL-17c responses were effectively blocked by purinergic receptors
3 antagonists supporting a similar mechanism of action for IL-17c (**Fig. 2**). As these
4 experiments were performed on slices from mice pups, we wanted to confirm it in
5 adult mice. Unfortunately, the robustness of adult mice bones prevent an approach
6 using OM slices. ATP has been previously characterized for its ability to reduce the
7 odour-induced responses in mice (Hegg et al., 2003; Lucero, 2013). If IL-17c leads to
8 a release of ATP in adult mice, its application in the OM should also decrease the
9 response to odourant stimulations. Our EOG results show that IL-17c decrease the
10 amplitude of odour induced response in the OM. Overall, our results are consistent
11 with an IL-17c-induced ATP release in mice.

12 We next examined how exogenous murine recombinant IL-17c nasal instillation
13 impacts the cellular turn-over of OM. A single instillation of IL-17c had no effect in
14 terms of apoptosis, cell proliferation or Iba1+ cell infiltrations. However, we observed
15 that repeated administration of IL-17c promoted both apoptosis and proliferation,
16 suggesting a role in the cellular dynamics of the OM (**Fig. 4**). In addition to the
17 cellular dynamic modulation, we also observed that IL-17c led to
18 macrophage/microglia-like cell infiltration in the olfactory mucosa. Similarly, IL-17c
19 was shown to promote the recruitment of immune cells around smooth muscle cells
20 in mice (Butcher et al., 2016). Therefore, IL-17c may act *in vivo* as a chemotactic
21 compound in the OM. The infiltration of Iba1+ cells may explain the increase in
22 cellular dynamics of the OM in our chronic model. Indeed, recruitment of
23 macrophages will lead to the establishment of an inflammatory state modulating
24 neuronal survival and basal cell proliferation (Borders et al., 2007a; Borders et al.,
25 2007b). Additionally, as ATP promotes cell proliferation in the OM (Jia et al., 2011),

1 the OM cellular proliferation induced by IL-17c may rely in part, on the ATP release
2 induced by IL-17c.

3 Poly(I:C) nasal instillation led to morphological changes of the OM (**Fig. 5**). This
4 nasal instillation also increased the IL-17c expression level and we examined the
5 importance of IL-17c for the OM morphological changes induced by Poly(I:C)
6 treatment. Similarly as Poly(I:C), IL-17c instillation led to a decrease of the
7 expression of genes related to the neuroepithelium barrier but without alteration of
8 the epithelium morphology (**Fig. 5, 6**). Furthermore, this effect did not rely on ATP
9 release induced by IL-17c treatment as nasal instillation of ATP did not modify tight
10 junction protein expression. This last result shows that IL-17c impacts the OM
11 physiology not only through ATP release. Our results of the effect of IL-17c on the
12 OM barrier integrity contrast with its impact on colonic epithelial cells where it
13 promotes tight junction genes expression (Reynolds et al., 2012). However, cytokines
14 can increase or decrease the expression of tight junction proteins depending on the
15 organ (Capaldo, and Nusrat, 2009).

16 A recent study highlighted a protective role of IL-17c in the peripheral nervous system
17 (Peng et al., 2017). We choose to explore a similar role of IL-17c during a Poly(I:C)
18 challenge in the OM. As IL-17c was induced by Poly(I:C) *in vivo*, we used a primary
19 culture of OM cells allowing a control of the extracellular medium and thus of IL-17c
20 concentration. In this culture, while Poly(I:C) increased the level of cellular death, co
21 and pre-treatment with IL-17c improved the survival of OM cells (**Fig. 7**). This last
22 result contrasts with the *in vivo* treatment where IL-17c nasal instillation led to an
23 increase in apoptosis level. The simplest explanation for this discrepancy would be
24 that the up regulation of cell turnover observed *in vivo* is triggered by the
25 macrophage/monocyte infiltration induced by the administration of IL-17c. As those

1 cells are not present in the primary culture, we could observe an anti-apoptotic effect
2 of IL-17c.

3 Overall, our results show that IL-17c can increase the cellular dynamic of the OM
4 toward regeneration. Such action would help to counterbalance the massive
5 damages observed after viral infection of the olfactory mucosa leading to post viral
6 olfactory disorders (Kanaya et al., 2014). The olfactory bulb is often considered as
7 the first barrier against airborne pathogens entry to the central nervous system
8 (Durrant et al., 2016). As several other cytokines receptors are expressed in the OM
9 (Ibarra-Soria et al., 2014; Saraiva et al., 2016), our results indicate that before the
10 olfactory bulb line of defences, the OM could be an effective barrier against airborne
11 pathogens.

12

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6 **Conflict of interests:**

7 The authors declare no conflict of interests.

8 **Author contributions:**

9 BB and NM designed, performed experiments, analysed data and wrote the paper,

10 AD performed experiments and analysed the data, CLPS and AS performed

11 experiments; PC designed and performed experiments.

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1 **Figure 1: Expression of IL-17c receptors in the mouse olfactory mucosa.** (A) In
2 situ hybridization of IL-17re and (B) immunohistochemistry of IL-17ra indicate that
3 both IL-17c receptors subunits are mainly expressed in the apical part of the olfactory
4 epithelium corresponding to sustentacular cells and in axon bundles of the lamina
5 propria.

1 **Figure 2: IL-17c mediates a rise in calcium in sustentacular cells and in**
2 **neurons through the release of ATP.** Representative calcium wave spreading
3 before (**A₁**, **B₁**) and after IL-17c (**A₂**) or ATP (**B₂**). Stimulation of slices preincubated in
4 presence of purinergic receptors antagonists (“PRA” -suramin (100μM), PPADS
5 (25μM - **A₃**; **B₃**) or not (**A₂**, **B₂**)). (**C**) Average time courses of the effect of ATP (10⁻⁴
6 M), IL-17c (500ng/mL) on the calcium signal measured in the ROI containing the SCs
7 layer. Drug application starts at the black arrow and lasts 30s ($\Delta F/F \pm SEM$; n=5 per
8 condition).

1 **Figure 3: Amplitude of olfactory response is diminished by IL-17c treatment.**
2 (A) Schematic diagram of the experimental preparation allowing the simultaneous
3 recording of two OM areas treated with saline and IL-17c (1 μ g/mL) respectively. EOG
4 recordings were performed on endoturbinates IIb and IV located centrally in the nasal
5 cavity. (B) Time course of recordings and treatments (C) Effects of treatment on the
6 time course of normalized EOG amplitudes in response to heptanal (1:1000 in
7 mineral oil). Mean \pm SEM values are expressed as % of control EOG recordings prior
8 to treatment (n=9). Two way ANOVA (*P<0.05). (D) Typical normalized EOG
9 responses to heptanal (1:1000 in mineral oil) before and 20 minutes after treatment.

1 **Figure 4: Cellular dynamics of the OM is enhanced in mice intranasally instilled**
2 **with IL-17c.** The cellular dynamic of the OM was evaluated by quantifying the areas
3 with cleaved caspase 3 (C3C; **(A)**), proliferating cell nuclear antigen (PCNA; **(B)**) and
4 the number of cell stained against ionized calcium-binding adapter molecule 1
5 (Iba1+; **(C)**), taken as indices of apoptosis, proliferation and infiltration of
6 monocyte/macrophage cells, respectively. Results were expressed as mean of the
7 C3C and PCNA signal area and Iba1+ cells counts in the OM in each region of
8 interest (n=6 vehicle-treated mice / n=7 IL-17c (500ng/mL) treated mice).
9 Representative images of **(A2-3)** C3C staining with OSN ongoing apoptosis, **(B2-3)**
10 PCNA staining mainly present in basal cell and **(C2-3)** Iba1+ staining.

1 **Figure 5: Expression of IL-17c in a pseudo-viral context induced by Poly(I:C)**
2 **(A)** Experimental protocol for intranasal administration of Poly(I:C) (20 µg Poly(I:C)
3 sodium salt dissolved in 10 µL of sterile saline into each nostril) or vehicle. Mice
4 received a maximum of five intranasal administrations of Poly(I:C) or vehicle solution,
5 once every 24 h, and were sacrificed 3, 5 or 7 days after the first administration (n =
6 8,8,6 respectively). Control mice were also sacrificed at 3, 5 and 7 days after the first
7 vehicle administration (n=9). **(B)** Dot plot of the mRNA relative level of IL-17c
8 concentration for individual mice expressed in the OM. The expression level was
9 normalized to β-actin and is presented as mean ± SEM. Different letters represent
10 significantly different groups. **(C)** Hematoxylin *and* eosin staining of representative
11 OMs from a mouse treated unilaterally with Poly(I:C). Panels show details of olfactory
12 mucosa of mice nasally instilled with vehicle (**C₂**), Poly(I:C) (**C₃**), IL-17c (500ng/mL
13 **C₄**) or ATP (10⁻⁴M) (**C₅**). **(D)** Representative image with light microscopy of IL-17c
14 mRNA expression in a D-5 treated mouse visualized by *in situ* hybridization with
15 sense and antisense probe. Panels on the right show details of Steno's gland (**D₂**,
16 **D₃**), olfactory mucosa (**D₄**, **D₅**) and nasal associated lymphoid tissue (**D₆**, **D₇**). The
17 zone with the sense probe as a control is on the left (**D₂**, **D₄**, **D₆**).

1 **Figure 6: Intranasal instillation of IL-17C results in a decrease in expression of**
2 **tight junction related proteins.** Mice accustomed to receive intranasal instillations
3 receive a three day treatment of vehicle (Ctrl), Poly(I:C) (2g/L), IL-17C (500ng/mL) or
4 ATP (10^{-4} M); n=13, 9, 16, 13 respectively. The mRNA level of tight junction related
5 proteins (claudin-1 **(A)**), claudin-11 **(B)**) or tight junction protein-1 **(C)**) was further
6 evaluated by quantification on cDNAs from olfactory mucosa by real time RT-PCR.
7 Their expression levels were normalized to that of β -actin and are given as
8 mean \pm SEM. Different letters represent significantly different groups.

1 **Figure 7: Anti-cytotoxic effect of IL-17c in a primary culture of OM during a**
2 **Poly(I:C) challenge. (A)** Experimental design. The OM primary culture was grown
3 for 7 days before supplementation with culture medium alone (control) or containing
4 Poly(I:C) (1mg/mL) and/or IL-17c (200ng/mL). The impact of these various
5 treatments on cell death in the OM primary culture was quantified by the percentage
6 of LDH released in the extracellular medium. **(B)** Dot plot representing the relative
7 LDH released in presence of various treatments including co-treatment (“Co”
8 Poly(I:C) - IL-17c) and pre-treatment with IL-17c (“Pre”). The relative LDH release
9 was estimated by measuring the cellular LDH content and expressed as a relative
10 value to the control condition. Data are presented as mean \pm SEM (n=16 per
11 condition). Columns with different letters differ significantly.

1 **Table 1. Primers used for qPCR reactions**

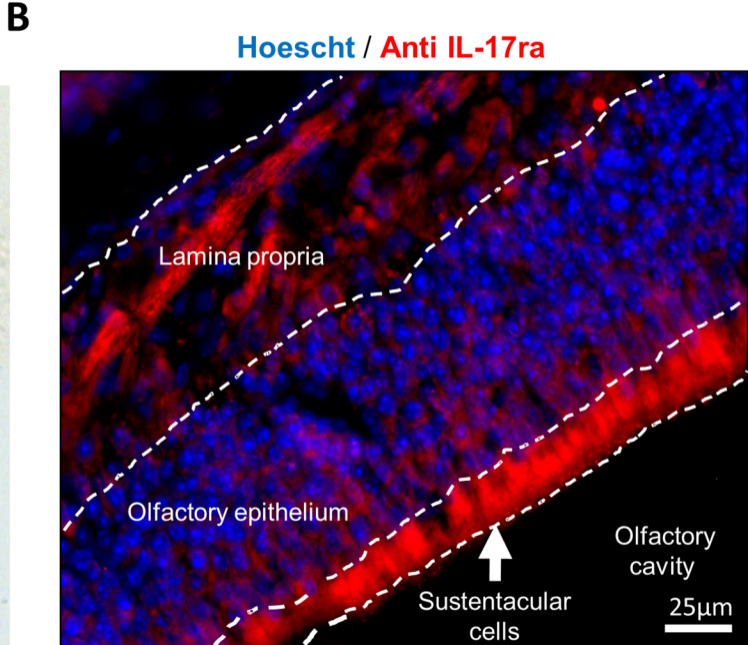
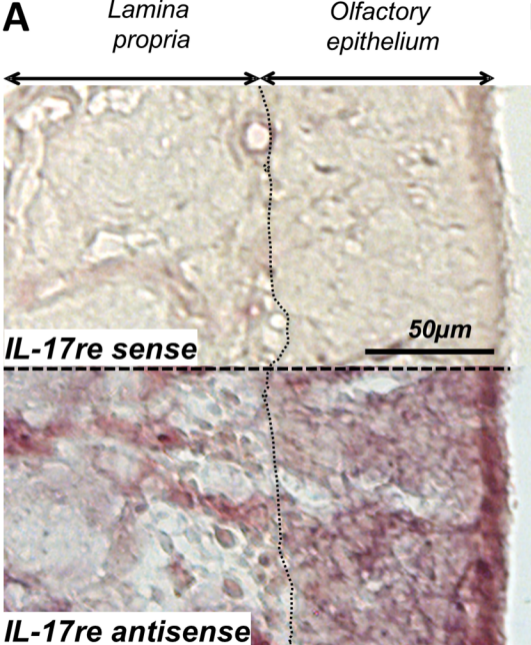
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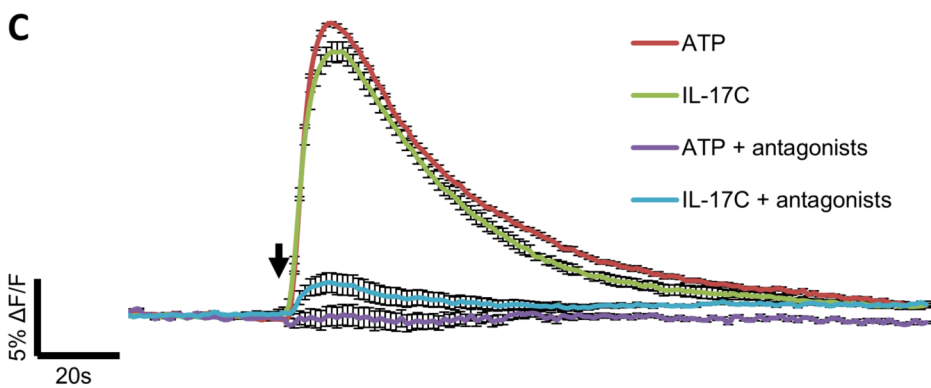
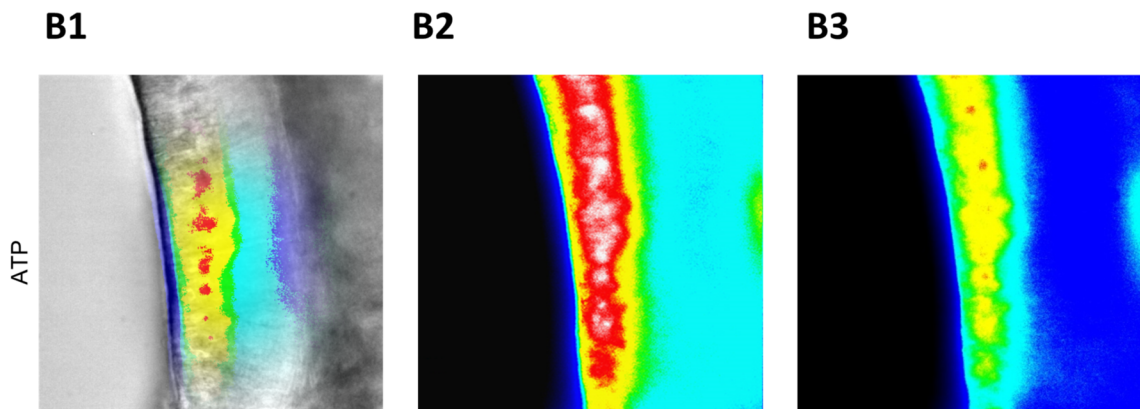
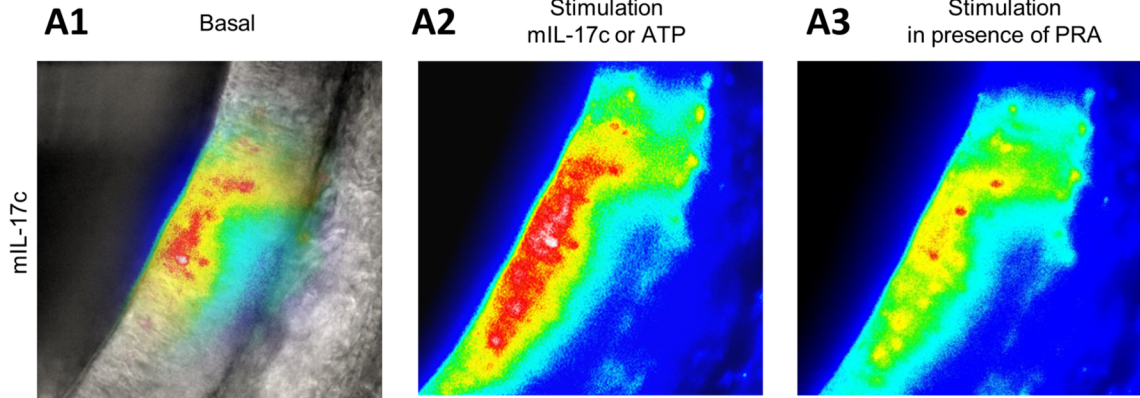
	GenBank Accession number	Sequences primers (5'>3')	Tm (°C)
β- actin	NM_007393.5	GACCCAGATCATGTTTGAGACCTT	60.57
		CACAGCCTGGATGGCTACGT	62.24
Claudin-1	NM_016674.4	ACTGTGGATGTCCTGCGTTT	59.89
		CCCCAGCAGGATGCCAATTA	60.11
Claudin-11	NM_008770.3	ACATCCTCATCCTTCCAGGCTA	60.36
		ATGCAGGGGAGAACTGTCAA	58.93
Tight Junction Protein 1	NM_009386	GGAGATGTTTATGCGGACGG	58.78
		CCATTGCTGTGCTCTTAGCG	59.62
IL-17c	NM_145834.4	CTGGAAGCTGACACTCACGA	59.68
		CCACGACACAAGCATTCTGC	60.11

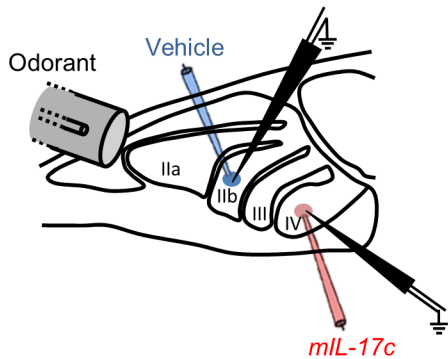
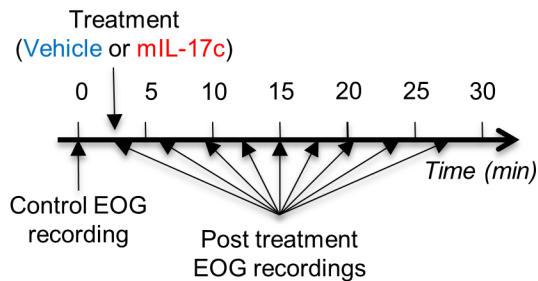
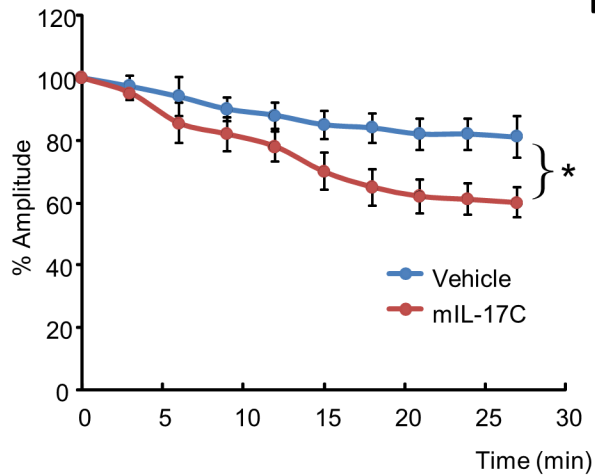
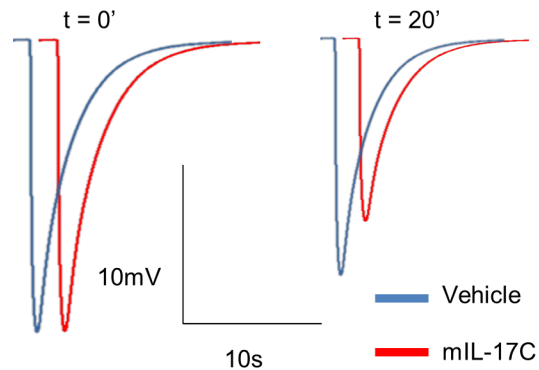
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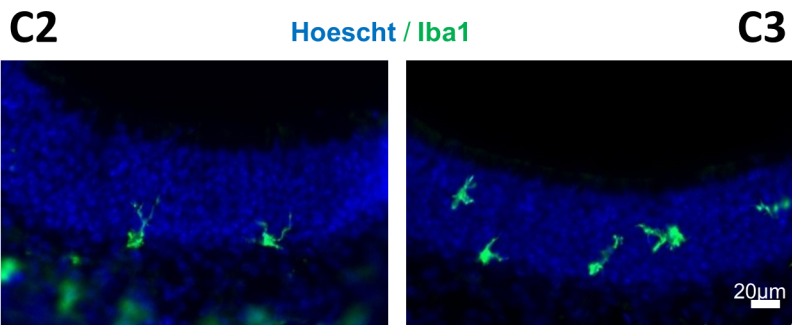
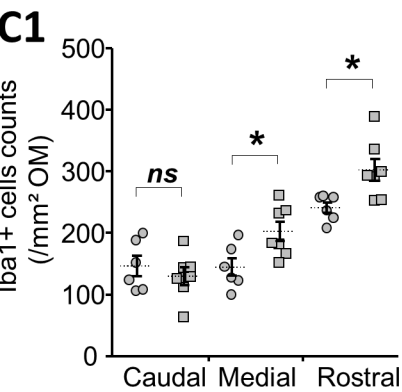
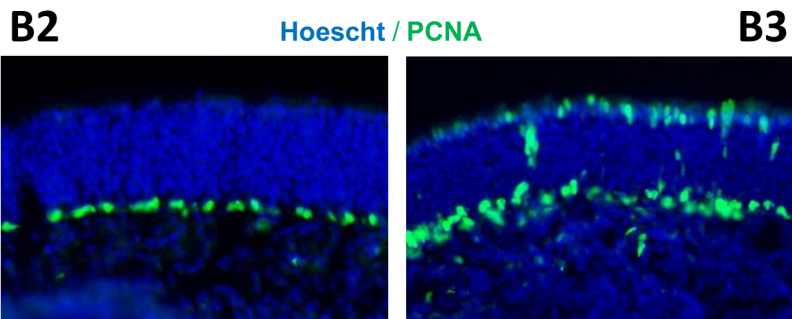
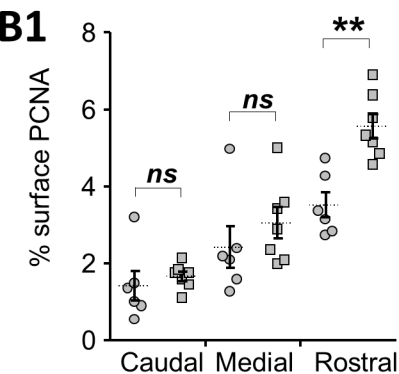
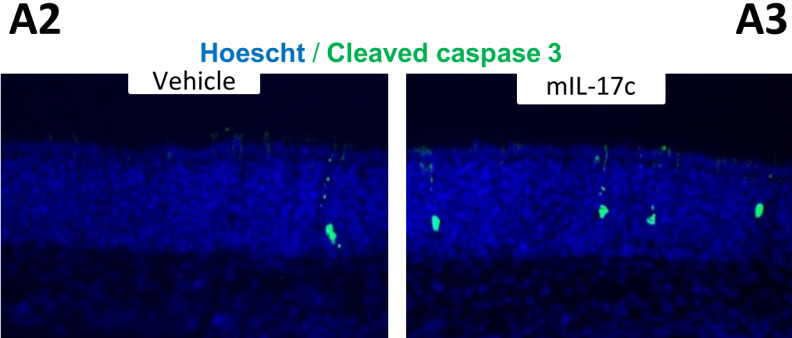
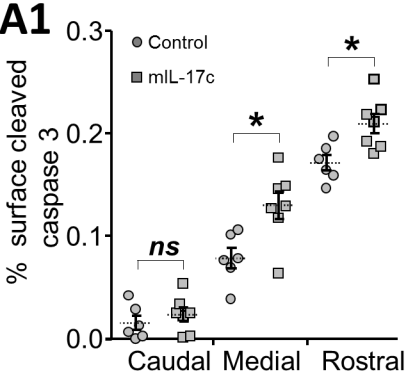
1 **Supplementary Figure 1: Caffeine mediates a rise in calcium rise in**
2 **sustentacular cells and in neurons in the presence of purinergic antagonists**
3 **receptors.** Representative calcium wave spreading with caffeine (50nM) before
4 (caffeine) or after (caffeine + antagonists) the incubation with purinergic antagonist
5 receptors. Average time courses of the effect of caffeine on the calcium signal
6 measured in the ROI containing the SCs/OSNs layer. Drug application starts at the
7 black arrow and lasts 30s ($\Delta F/F \pm SEM$; n=4).

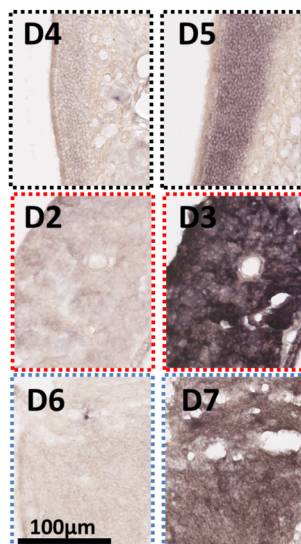
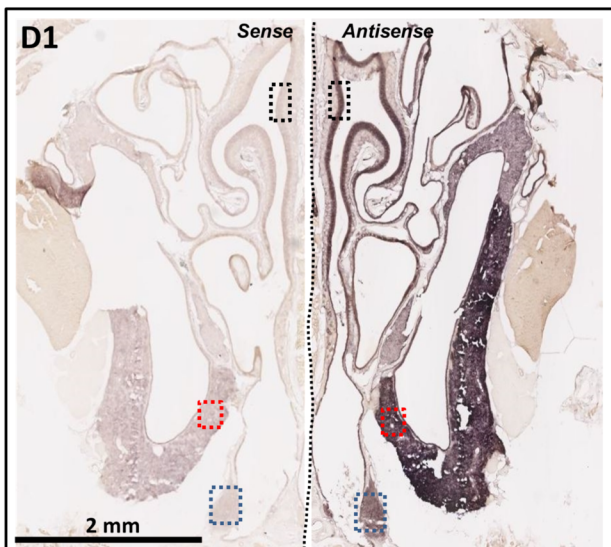
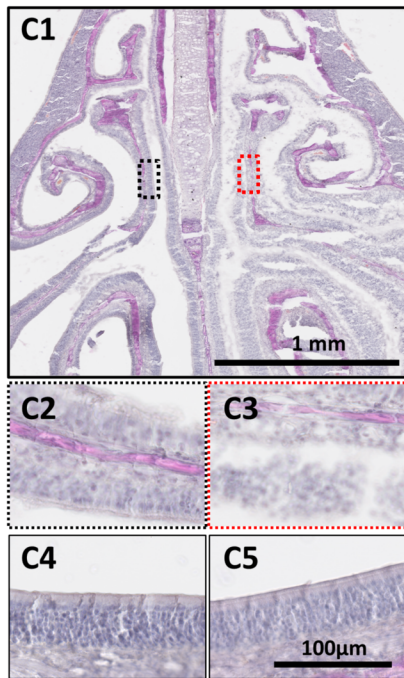
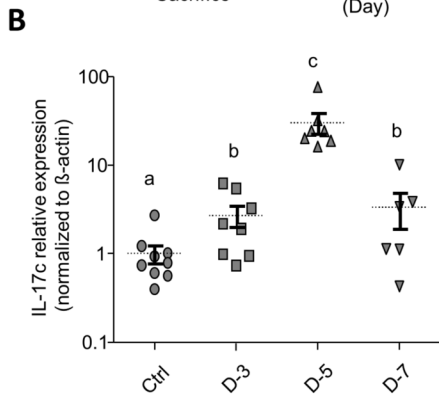
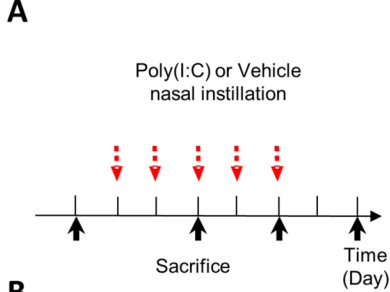
1 **Supplementary Figure 2: Cellular dynamics of the OM is not altered 24h after**
2 **unique nasal instillation of IL-17c.** The cellular dynamic of the OM was evaluated
3 by quantifying the areas with cleaved caspase 3 (C3C; **(A)**), proliferating cell nuclear
4 antigen (PCNA; **(B)**) and the number of cell stained against ionized calcium-binding
5 adapter molecule 1 (Iba1+; **(C)**), taken as indices of apoptosis, proliferation and
6 infiltration of monocyte/macrophage cells, respectively. Results were expressed as
7 mean of the C3C and PCNA signal area and Iba1+ cells counts in the OM in each
8 region of interest (n=6 vehicle-treated mice / n=7 IL-17c (500ng/mL) treated mice).

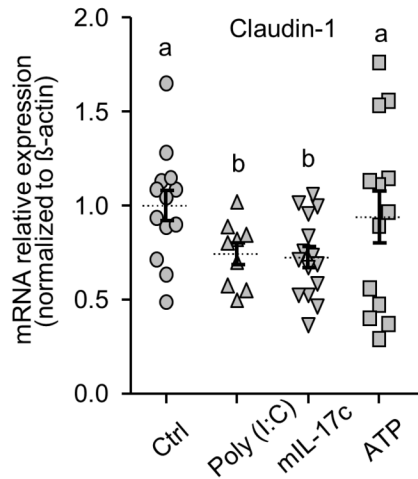
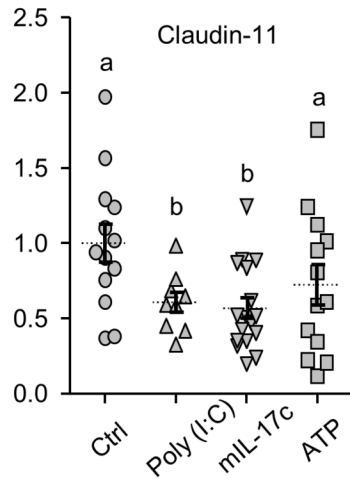
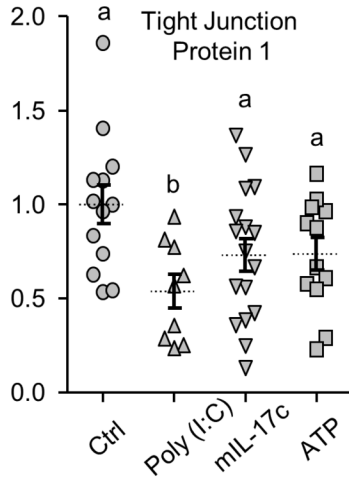


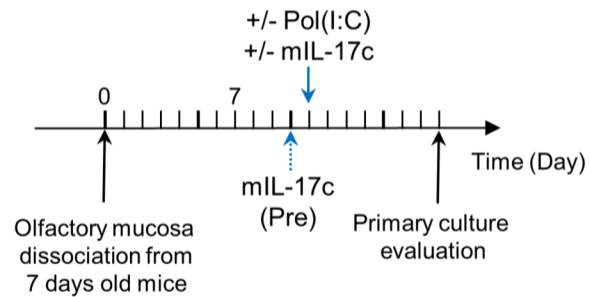


A**B****C****D**





A**B****C**

A**B**