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Insect cuticular proteins and their role in transmission of phytoviruses

Maëlle Deshoux, Baptiste Monsion¹ and Marilyne Uzest



Many viruses of agricultural importance are transmitted to host plants via insect vectors. Characterizing virus–vector interactions at the molecular level is essential if we are to fully understand the transmission mechanisms involved and develop new strategies to control viral spread. Hitherto, insect proteins involved in virus transmission have been characterized only poorly. Recent advances in this topic, however, have significantly filled this knowledge gap. Among the vector molecules identified, cuticular proteins have emerged as key molecules for plant virus transmission, regardless of transmission mode or vector considered. Here, we review recent evidence highlighting that the CPR family, and particularly RR-1 proteins, undoubtedly deserves special attention.

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Introduction

To ensure sustainability in the environment, phytoviruses must overcome two major constraints: their hosts are immobile, and the plant cell wall represents a physical barrier that viruses have to cross before they can replicate and spread in plant tissues. Most plant viruses are transmitted horizontally and re-transported by plant-feeding organisms (vectors) that are able to move from plant to plant [1,2]. The most frequent vectors of plant viruses are hemipteran and thysanopteran insects with piercing-sucking mouthparts, including aphids, whiteflies, leafhoppers, planthoppers, and thrips [2]. Virus–vector interactions, which are sophisticated and highly specific [3], can be classified into two main categories (for a review see [4]). Noncirculative viruses are reversibly attached to the cuticle of the insect mouthparts, in the stylets or foregut

of their vectors [4–6], during their journey from one plant to another. Circulative viruses are ‘internalized’ in the vector body, and must cross the gut barrier to reach the hemolymph and/or other tissues. Ultimately, viruses reach the salivary glands, and are injected, together with vector saliva, into new host plants. A few virus species have been shown to replicate within their vectors during their journey and have been classified in the circulative propagative subcategory [7]. Numerous studies have focused on elucidating the mechanisms underlying vector transmission [3,5,6,8], and viral determinants have been well characterized for most plant virus species studied. These include structural proteins, membrane viral glycoproteins, or non-structural virus-encoded proteins [4,9,10]. However, an extensive study of virus–vector interactions at the molecular level is still challenged by the difficulty of identifying vector partners and validating their role in virus transmission [11^{••},12^{••},13^{••},14^{••},15^{••}].

Nonetheless, considerable efforts have been made to develop complementary approaches and high-throughput methods to help identify vector proteins involved in virus–vector interactions. Among these interacting molecules are several cuticular proteins (CuPs) [13^{••},15^{••},16,17,18^{••},19–26]. CuPs are chitin-binding proteins that contribute to cuticle structural integrity, and reflect its diversity and mechanical properties [27–29]. CuPs have been classified into 14 families [30[•],31,32], the most abundant by far being the CPR family comprising proteins with a Rebers and Riddiford (RR) consensus [33]. The RR family is divided into three subfamilies, RR-1, RR-2, RR-3 [30[•]], to which most identified virus-interacting proteins can be assigned (Table 1). Their role as a key partner of both noncirculative and circulative plant viruses was hitherto unforeseen. Here, we review striking advances in the characterization of CuP–virus interactions that have brought novel insights to the field of vector transmission of plant viruses.

Role of cuticular proteins in noncirculative plant virus transmission

Noncirculative viruses bind reversibly to specific retention sites on the cuticle of the feeding apparatus. Therefore, virus-interacting molecules should be cuticular compounds that fulfill the role of virus receptors. To date, receptors of foregut-borne viruses have been poorly characterized [34,35], and no CuP has been shown to be involved in their retention or transmission. The great majority of noncirculative viruses, among which are members of the families *Potyviridae*, *Bromoviridae* and

Table 1

Cuticular proteins (CuPs) identified in the virus–insect vector interaction studies presented in this review

Virus species/genus	Vector – species	Transmission mode	CuPs identifier ^a – other name	Protein family ^b (subfamily)	Approaches	Reference
ZYMV/ <i>Potyvirus</i>	Aphid – <i>M. persicae</i>	Noncirculative	AAO63549	CPR (RR-2)	Urea extraction of aphid CuPs, 1-D & 2-D gel electrophoresis, Far-western blot, MS analyses	[17]
			AAL29466 AAZ20451 AAZ20447	CPR (RR-2) CPR (RR-1) CPR (RR-2)		
CaMV/ <i>Caulimovirus</i>	Aphid	Noncirculative	ND	ND	Biochemical characterization, Stylet immunolabeling	[40*]
CaMV/ <i>Caulimovirus</i>	Aphid – <i>A. pisum</i> , <i>M. persicae</i>	Noncirculative	MG188739 – <i>Stylin-01</i>	CPR (RR-1)	Stylet immunolabeling, Colocalization, <i>in vitro</i> competition assays, RNAi	[15**]
			MG188741 – <i>Stylin-01</i>	CPR (RR-1)		
CMV/ <i>Cucumovirus</i>	Aphid – <i>M. persicae</i>	Noncirculative	DQ108938 – <i>Mpcp4</i>	CPR (RR-1)	YTH	[22]
CMV/ <i>Cucumovirus</i>	Aphid – <i>A. pisum</i>	Noncirculative	ND	CPR (RR-2)	Peptide array (RR-2 proteins)	[25]
TuYV ^c / <i>Polerovirus</i>	Aphid – <i>M. persicae</i>	Circulative	ND ^d	ND	Whole cell lysate (aphids), 1-D & 2-D gel electrophoresis, Far-western blot & MS analyses	[16]
CYDV-RPV/ <i>Polerovirus</i>	Aphid – <i>S. graminum</i>	Circulative	gi:193647865	CPR (RR-2)	Genetics coupled to 2-D-DIGE & MS analyses	[18**]
			gi:193706873 gi:193647875 gi:193582403	ND ^d CPR (RR-2) CPR (RR-2)		
BYDV-GPV/ <i>Luteoviridae</i> ^e	Aphid – <i>R. padi</i>	Circulative	gi:288558725 – <i>Cp62 precursor</i> NP_001156154.1 – <i>Cp5 precursor</i>	CPR (RR-2) CPR (RR-2)	iTRAQ & MS analyses	[20]
RSV/ <i>Tenuivirus</i>	Planthopper – <i>L. striatellus</i>	Circulative-Propagative	KC485263 – <i>CPR1</i>	CPR (RR-1)	YTH, Chemiluminescent co-IP, Colocalization, GST pull-down, RNAi	[13**]
RSV/ <i>Tenuivirus</i>	Planthopper – <i>L. striatellus</i>	Circulative-Propagative	XM_014390248.1 – <i>Cuticle protein A3A like</i> JAS02196.1	CPR (RR-2) Tweedle	YTH	[26]

^a Given accession numbers from original studies.

^b Classified using CutProtFam-Pred (<http://aias.biol.uoa.gr/CutProtFam-Pred/>).

^c Formerly BWYV-FL1 (beet western yellows virus).

^d published accession number does not correspond to a CuP according to databases and CutProtFam-Pred.

^e Unassigned member in the family Luteoviridae. CuP: cuticular protein; ND: not determined; MS: mass spectrometry; RNAi: RNA interference; DIGE: difference gel electrophoresis; iTRAQ: isobaric tags for relative and absolute quantification; co-IP: co-immunoprecipitation; YTH: yeast two-hybrid. ZYMV: zucchini mosaic virus; CaMV: cauliflower mosaic virus; CMV: cucumber mosaic virus; TuYV: turnip yellows virus; CYDV: cereal yellow dwarf virus; BYDV: barley yellow dwarf virus, RSV: rice stripe virus.

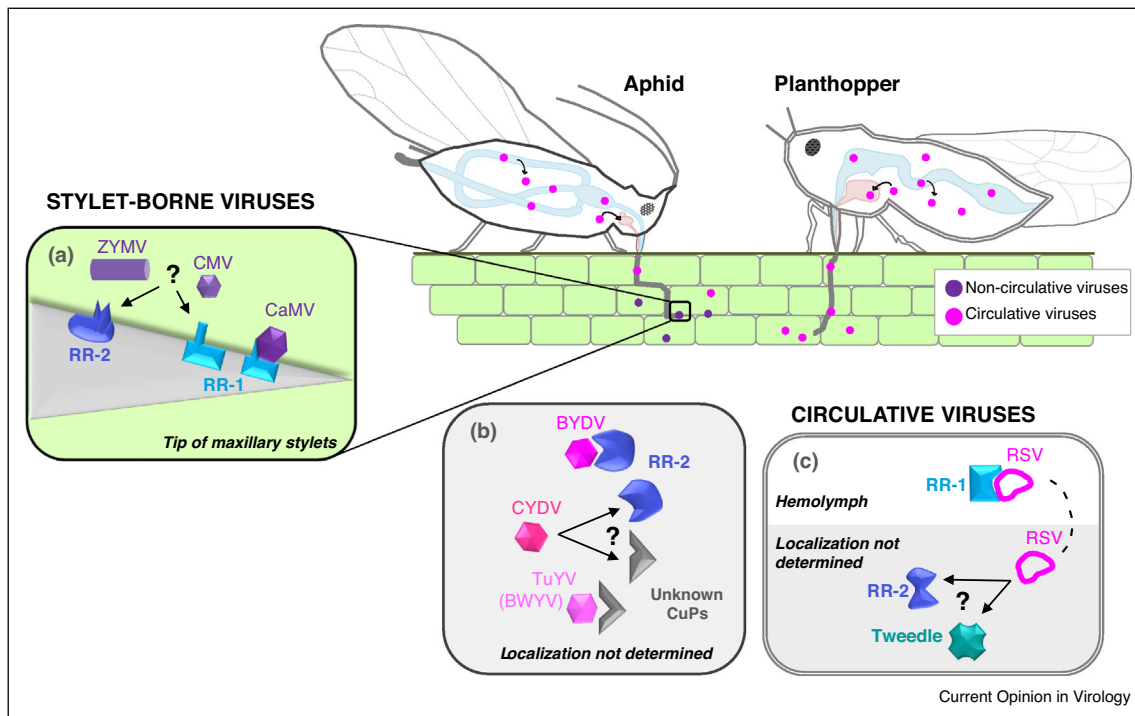
Caulimoviridae, is retained in the stylets [36–38], whose composition is poorly characterized [39]. Using *in vitro* interaction assays on dissected stylets, Uzest *et al.* [38] demonstrated that the receptor of the cauliflower mosaic virus (CaMV) was a CuP located at the fused food/salivary common canal of aphid maxillary stylets, on the acrostyle — an organ discovered later [40*] in an area described to harbor receptors of potyviruses and cucumoviruses [37,41]. However, binding to the acrostyle has been demonstrated only for a caulimovirus, and direct evidence of virus retention within the common canal is still lacking for other viruses.

Dombrovsky *et al.* [17] were the first to identify cuticular partners of a noncirculative virus, the zucchini yellow mosaic virus (ZYMV) — a potyvirus that interacts with its aphid vectors through a viral-encoded protein (helper component, HC-Pro [42]). A far-western blot approach allowed detection of a few spots that specifically interacted with wt ZYMV-HC-Pro combinations, but not with a transmission-defective mutant. Out of nine spots micro-sequenced, four were identified as CuPs: one RR-1 and three RR-2 proteins (Table 1). At the time, it was not possible to ascertain the presence of any of these CuPs in

the stylets, nor to confirm the biological relevance of these results. However, since then, using immunodetection approaches, some peptides have been identified at the tip of maxillary stylets [15**,25,40*], among which the peptide PepS was found to be present in two CuPs identified in this study (AAO63549.1 and AAL29466.1), reinstating them as prime candidate receptors of ZYMV (Figure 1). Further investigations should determine if ZYMV is retained in the common canal, and evaluate the role of these RR-2 proteins in ZYMV transmission.

Two independent studies on cucumber mosaic virus (CMV) receptor candidates were published in 2017 [22,25]. This cucumovirus, transmitted by aphids, interacts directly with vector receptors through its coat protein [43,44]. In the first study, using a yeast-two-hybrid (YTH) system, Liang and Gao [22] reported interaction between the coat protein of CMV, and one of four reported *Myzus persicae* CuPs [45], the RR-1 protein Mpcp4 (Table 1, Figure 1). Whether this interaction in yeast reflects a true binding of CMV to Mpcp4 within vector stylets remains to be demonstrated, and the role of this protein in CMV transmission has not yet been assessed; however, the very recent identification of

Figure 1



Plant virus–cuticular protein (CuP) interactions. (a) Interaction of noncirculative viruses with cuticular proteins at the tip of aphid maxillary stylets: cauliflower mosaic virus (CaMV) with RR-1 protein via its helper protein P2 [15**], cucumber mosaic virus (CMV) with RR-1 and/or RR-2 proteins [22,25], zucchini yellow mosaic virus (ZYMV) via its helper protein HC-Pro with RR-1 and/or RR-2 proteins [17]. (b) Interaction of barley yellow dwarf virus (BYDV), cereal yellow dwarf virus (CYDV) and turnip yellow virus (TuYV), three circulative viruses, with RR-2 and/or unknown cuticular proteins within their aphid-vector body [16,18**,20]. (c) The nucleocapsid protein of the circulative rice stripe virus (RSV) binds an RR-1 protein in the hemolymph of its planthopper vector [13**], and may also interact with another RR-2 and a Tweedle cuticular protein [26].

Mcp4 in the acrostyle (named Stylin-01 by Webster and co-workers [15**], see below) supports this CuP as a CMV receptor candidate. In the second study, a peptide array approach, based on results showing the presence of RR-2 proteins in the acrostyle [40*], was developed to characterize CuP-virus interactions [25]. Two consensus sequences were deduced from the pattern of hybridization of CMV onto the array, one within the RR-2 chitin-binding domain, the other being a sequence frequently found in RR-2 proteins, likely exposed on the acrostyle surface [25]. These two studies revealed that CMV interacts with both RR-1 proteins in yeast, and with RR-2 peptides *in vitro* (Table 1, Figure 1). As these two types of CuPs are part of the acrostyle [15**], we hypothesize that they both play a role in virus retention. However, as already mentioned for ZYMV, binding of CMV in the common canal lacks direct evidence [37], and additional biochemical and functional validation are needed to determine the role of these CuPs in CMV transmission.

A recent study has made great strides towards noncirculative virus receptor identification [15**]. By immunolabeling with a series of antibodies specific to annotated CuPs from the CPR family, two highly homologous RR-1 proteins, Stylin-01 and Stylin-02, were detected at the tip of aphid stylets [15**]. A peptide corresponding to the C-terminus of Stylin-01 and Stylin-02 was shown to evenly cover the surface of the acrostyle, and to overlap with CaMV retention sites. Two series of experiments support Stylin-01 as a receptor candidate for CaMV (Table 1, Figure 1): firstly, CaMV helper protein P2 and the antibody targeting the surface-exposed peptide compete for binding to the acrostyle, and secondly, silencing Stylin-01 in the vector species *M. persicae* via RNA-mediated interference resulted in a 40% decrease in CaMV transmission efficiency.

To date, the only CuPs identified in insect stylets are proteins from the CPR family [15**,25,40*]. Both RR-1 and RR-2 proteins have been detected at the tip of maxillary stylets, displaying domains at the interface in direct contact with contaminated phloem sap. And both are involved in virus–vector interactions. One of the RR-1 proteins is already clearly involved in CaMV transmission, and some results suggest that it might also be important for the transmission of cucumoviruses. However, current knowledge does not distinguish if only a single CPR protein is required for binding all noncirculative viruses, which includes hundreds of virus species, or if several CuPs can act as receptors of noncirculative viruses within insect stylets.

Role of cuticular proteins in circulative plant virus transmission

Since circulative transmission entails crossing several vector tissues and travelling within the insect body, various vector molecules are likely to interact with

circulative viruses throughout their journey to, firstly, promote virus entry into, and release from, insect tissues, secondly, facilitate virus movement, thirdly, protect viruses from degradation within the insect hemolymph, and lastly, play a role in virus replication in the case of propagative viruses. In addition to classical methods such as YTH screenings, current attempts to characterize insect partners of circulative viruses often rely on the development of high-throughput approaches such as proteomic and transcriptomic analyses able to detect changes in protein abundance or genes differentially expressed between viruliferous or healthy insects. Generally, these studies generate listings with many protein candidates, including some CuPs [13**,18**,19–21,23,26,46–48]. These listings are important resources of potential virus putative partners. However, the major issue will be to confirm virus–CuP interactions and to validate the role of these candidates in virus transmission.

In a pioneering study, Seddas *et al.* [16] searched for aphid proteins that interact with a polerovirus, the beet western yellows virus (BWYV, currently known as turnip yellows virus or TuYV [49]), and that could be involved in virus transmission. Using wt viruses or derived non-transmissible mutants as overlays, a few aphid proteins of *M. persicae* were revealed by far-western blot analyses, and a CuP of 33 kDa was identified from 2-dimensional gel electrophoresis and mass spectrometry analysis (Table 1). This CuP was shown to interact strongly with wt BWYV, but not with the mutant lacking the readthrough (RT) domain, suggesting an interaction through the RT protein (Figure 1)—a protein strictly required for aphid transmission [50] and shown to play a role in virus movement [51]. At that time, as it was unexpected to find an interaction between a circulative virus and a CuP, the authors concluded that this aphid protein might not play a role in the BWYV transmission mechanism. However, a plausible involvement of CuPs in polerovirus translocation in the gut was later suggested with the identification of several aphid CuPs that correlated with a transmission efficient phenotype and cereal yellow dwarf virus-RPV (CYDV-RPV) movement through the insect gut [18**]. Amongst the aphid proteins predicted to facilitate virus entry at the gut barrier, two CuPs were detected: one RR-2 and CPG12 (gi:193647865 and gi:193706873, respectively) (Table 1, Figure 1). Additionally, two other RR-2 CuPs (gi:193647875, gi:193582403) were upregulated both in competent and hindgut-refractive genotypes (Table 1), precluding any clear conclusion on their role in virus translocation [18**]. In a third recent study [20], interactions between barley yellow dwarf virus BYDV-GPV, a member of the *Luteoviridae* family, and CuPs from its aphid vector *Rhopalosiphum padi* were also reported. The two insect partners, which are RR-2 proteins, were identified using two approaches: firstly, differential proteomics analyses between healthy and viruliferous *R. padi* (Table 1, Figure 1), and secondly, YTH using viral RT

protein as bait (Table 1, Figure 1). Interestingly, this second RR-2 protein, called *cuticular protein 5 precursor*, is highly homologous with CYDV-RPV partners (gi:193582403, gi:193647865, [18**]). Although additional experiments are required to determine whether CuPs play a role in luteovirids transmission by facilitating virus entry at the gut level or at another step of the transmission process, taken together, these results stress the importance of characterizing luteovirids–CuPs interactions in future work.

A role for a CuP in circulative virus transmission was formally demonstrated for rice stripe virus (RSV) [13**]—a tenuivirus transmitted mostly by the planthopper *Laodelphax striatellus* in a propagative manner [52]. The authors provided the only direct experimental evidence thus far of an interaction between a CuP and a circulative virus within an insect vector. CPR1, an RR-1 protein, was identified by YTH screening of a cDNA library using the viral nucleocapsid protein pc3 (N) as bait (Table 1). A strong interaction between CPR1 and pc3 proteins was confirmed *in vivo* by a co-immunoprecipitation assay. Moreover, the two proteins co-localized in insect cell culture, and in hemocytes isolated from the hemolymph of viruliferous insects (Figure 1), whereas silencing CPR1 in *L. striatellus* resulted in a 57% decrease in RSV transmission capacity. The authors proposed that CPR1 could bind virus particles in the hemolymph and hence assist viral movement towards the salivary glands [13**]. Two additional CuPs were shown to interact with RSV nucleocapsid protein N (pc3) in another, YTH-independent, screening [26]. These CuPs belong to the CPR (RR-2) and Tweedle protein families. It will be interesting to determine if RSV can interact with several CuPs within its vector, and if several CuPs are required for successful transmission. However, interactions of these novel candidates have not been confirmed in the vector, and their role in RSV transmission remain to be investigated thoroughly [26].

Role of cuticular proteins in arbovirus transmission

Interestingly, CuPs have also been reported to potentially interact with arboviruses, but the precise role of CuPs in arboviruses infection is not well documented. A deregulation of CuP gene expression in insects upon infection with animal viruses has been reported [53,54**,55]. In a more extended study, Colpitts *et al.* [54**] characterized a mosquito-borne flavivirus–CuP interaction. The authors investigated the role of a RR-2 pupal CuP, whose transcripts were downregulated in *Aedes aegypti* pupae upon infection by West Nile (WNV), dengue (DENV) or yellow fever (YFV) viruses. Overexpression of this CuP gene in mosquito cells or in live mosquitoes inhibited WNV infection. Furthermore, this pupal CuP interacted with the envelope of WNV, DENV and the capsid of YFV, which might impede viral entry to host cells. More

importantly, pre-incubation of WNV with pupal CuP prevented lethal WNV encephalitis in mice. This work highlights once again the key role of a CuP from the CPR family in virus transmission through a direct virus–vector interaction. However, in contrast to vectored plant viruses for which CuPs have been shown to promote virus transmission, chitin-binding proteins were instead proposed to protect the insect from viral infection by preventing virus–receptor interactions, or by strengthening natural barriers to pathogen infection.

Conclusions

Cuticular proteins now feature prominently on the short-list of insect molecules demonstrated to play a key role in plant virus transmission [12**,13**,14**,15**]. Recent literature has revealed that CuPs—the most obvious candidates as receptors for noncirculative viruses—also interact with plant circulative viruses. Current knowledge indicates that insect CuPs may facilitate their entry at the gut level, and assist virus particles in the hemolymph.

Almost all the CuPs identified so far in plant virus–vector interaction studies belong to the large CPR family. More precisely, while the role of RR-2 proteins in virus transmission remains to be determined, RR-1 proteins are definitely associated with circulative and noncirculative virus transmission. Future research should help define whether additional CuPs also participate in the transmission process.

The data presented in this review highlight the interest in strengthening further efforts to characterize insect CuPs and their interactions with plant viruses. In the future, we can count on growing interest from virologists to characterize CuPs, as novel candidates in the search for innovative viral control strategies.

Conflict of interest statement

Nothing declared.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as

- of special interest
- of outstanding interest

1. Thresh JM: **The long-range dispersal of plant viruses by arthropod vectors.** *Philos Trans R Soc Lond* 1983, **302**:497–528.
2. Bragard C, Caciagli P, Lemaire O, Lopez-Moya JJ, MacFarlane S, Peters D, Susi P, Torrance L: **Status and prospects of plant virus control through interference with vector transmission.** *Annu Rev Phytopathol* 2013, **51**:177–201.

3. Dietzgen RG, Mann KS, Johnson KN: **Plant virus–insect vector interactions: current and potential future research directions.** *Viruses* 2016, **8** pii:E303.
 4. Blanc S, Drucker M, Uzest M: **Localizing viruses in their insect vectors.** *Annu Rev Phytopathol* 2014, **52**:403-425.
 5. Whitfield AE, Falk BW, Rotenberg D: **Insect vector-mediated transmission of plant viruses.** *Virology* 2015, **479–480**:278-289.
 6. Ng JC, Zhou JS: **Insect vector–plant virus interactions associated with non-circulative, semi-persistent transmission: current perspectives and future challenges.** *Curr Opin Virol* 2015, **15**:48-55.
 7. Hogenhout SA, Ammar el D, Whitfield AE, Redinbaugh MG: **Insect vector interactions with persistently transmitted viruses.** *Annu Rev Phytopathol* 2008, **46**:327-359.
 8. Jia D, Chen Q, Mao Q, Zhang X, Wu W, Chen H, Yu X, Wang Z, Wei T: **Vector mediated transmission of persistently transmitted plant viruses.** *Curr Opin Virol* 2018, **28**:127-132.
 9. Ziegler-Graff V, Brault V: **Role of vector-transmission proteins.** *Methods Mol Biol* 2008, **451**:81-96.
 10. Brault V, van den Heuvel J, Verbeek M, Ziegler-Graff V, Reutenauer A, Herrbach E, Garaud J, Guillely H, Richards K, Jonard G: **Aphid transmission of beet western yellows luteovirus requires the minor capsid read-through protein P74.** *EMBO J* 1995, **14**:650-659.
 11. Liu S, Sivakumar S, Sparks WO, Miller WA, Bonning BC: **A peptide that binds the pea aphid gut impedes entry of Pea enation mosaic virus into the aphid hemocoel.** *Virology* 2010, **401**:107-116.
- These papers identified the first virus receptor in a vector.
12. Linz LB, Liu S, Chougule NP, Bonning BC: **In vitro evidence supports membrane alanyl aminopeptidase N as a receptor for a plant virus in the pea aphid vector.** *J Virol* 2015, **89**:11203-11212.
- See annotation to Ref [11**].
13. Liu W, Gray S, Huo Y, Li L, Wei T, Wang X: **Proteomic analysis of interaction between a plant virus and its vector insect reveals new functions of hemipteran cuticular protein.** *Mol Cell Proteomics* 2015, **14**:2229-2242.
- First cuticular protein shown to interact with a circulative virus in its insect vector, and shown to play a key role in transmission.
14. Mulot M, Monsion B, Boissinot S, Rastegar M, Meyer S, Bochet N, Brault V: **Transmission of Turnip yellows virus by *Myzus persicae* is reduced by feeding aphids on double-stranded RNA targeting the ephrin receptor protein.** *Front Microbiol* 2018, **9**:457.
- This study identified an aphid protein involved in circulative virus transmission.
15. Webster CG, Pichon E, van Munster M, Monsion B, Deshoux M, Gargani D, Calevro F, Jimenez J, Moreno A, Krenz B *et al.*: **Identification of plant virus receptor candidates in the stylets of their aphid vectors.** *J Virol* 2018 <http://dx.doi.org/10.1128/JVI.00432-18>. pii: JVI.00432-18. [Epub ahead of print].
- First cuticular protein identified in aphid stylets, shown to play a key role in the transmission of a noncirculative virus, the CaMV.
16. Seddas P, Boissinot S, Strub JM, Van Dorsselaer A, Van Regenmortel MH, Pattus F: **Rack-1, GAPDH3, and actin: proteins of *Myzus persicae* potentially involved in the transcytosis of beet western yellows virus particles in the aphid.** *Virology* 2004, **325**:399-412.
 17. Dombrovsky A, Gollop N, Chen S, Chejanovsky N, Raccach B: **In vitro association between the helper component-proteinase of zucchini yellow mosaic virus and cuticle proteins of *Myzus persicae*.** *J Gen Virol* 2007, **88**:1602-1610.
 18. Cilia M, Tamborindeguy C, Fish T, Howe K, Thannhauser TW, Gray S: **Genetics coupled to quantitative intact proteomics links heritable aphid and endosymbiont protein expression to circulative polerovirus transmission.** *J Virol* 2011, **85**:2148-2166.
- This study combines an original genetic approach to quantitative proteomics analyses to identify virus-vector interacting partners involved in circulative transmission.
19. Mar T, Liu W, Wang X: **Proteomic analysis of interaction between P7-1 of Southern rice black-streaked dwarf virus and the insect vector reveals diverse insect proteins involved in successful transmission.** *J Proteomics* 2014, **102**:83-97.
 20. Wang H, Wu K, Liu Y, Wu Y, Wang X: **Integrative proteomics to understand the transmission mechanism of Barley yellow dwarf virus-GPV by its insect vector *Rhopalosiphum padi*.** *Sci Rep* 2015, **5**:10971.
 21. Xu D, Zhong T, Feng W, Zhou G: **Tolerance and responsive gene expression of *Sogatella furcifera* under extreme temperature stresses are altered by its vectored plant virus.** *Sci Rep* 2016, **6**:31521.
 22. Liang Y, Gao XW: **The cuticle protein gene MPCP4 of *Myzus persicae* (Homoptera: Aphididae) plays a critical role in cucumber mosaic virus acquisition.** *J Econ Entomol* 2017, **110**:848-853.
 23. Schneeweis DJ, Whitfield AE, Rotenberg D: **Thrips developmental stage-specific transcriptome response to tomato spotted wilt virus during the virus infection cycle in *Frankliniella occidentalis*, the primary vector.** *Virology* 2017, **500**:226-237.
 24. Shrestha A, Champagne DE, Culbreath AK, Rotenberg D, Whitfield AE, Srinivasan R: **Transcriptome changes associated with Tomato spotted wilt virus infection in various life stages of its thrips vector, *Frankliniella fusca* (Hinds).** *J Gen Virol* 2017, **98**:2156-2170.
 25. Webster CG, Thillier M, Piroles E, Cayrol B, Blanc S, Uzest M: **Proteomic composition of the acrostyle: Novel approaches to identify cuticular proteins involved in virus-insect interactions.** *Insect Sci* 2017, **24**:990-1002.
 26. Li S, Li X, Zhou Y: **Ribosomal protein L18 is an essential factor that promote rice stripe virus accumulation in small brown planthopper.** *Virus Res* 2018, **247**:15-20.
 27. Willis JH: **Cuticular proteins: the neglected component.** *Arch Insect Biochem Physiol* 1987, **6**:203-215.
 28. Andersen SO, Hojrup P, Roepstorff P: **Insect cuticular proteins.** *Insect Biochem Mol Biol* 1995, **25**:153-176.
 29. Willis JH: **Cuticular proteins in insects and crustaceans.** *Am Zool* 1999, **39**:600-609.
 30. Willis JH: **Structural cuticular proteins from arthropods: annotation, nomenclature, and sequence characteristics in the genomics era.** *Insect Biochem Mol Biol* 2010, **40**:189-204.
- Comprehensive review on cuticular proteins of arthropods.
31. Ioannidou ZS, Theodoropoulou MC, Papandreou NC, Willis JH, Hamodrakas SJ: **CutProtFam-Pred: detection and classification of putative structural cuticular proteins from sequence alone, based on profile hidden Markov models.** *Insect Biochem Mol Biol* 2014, **52**:51-59.
 32. Pan PL, Ye YX, Lou YH, Lu JB, Cheng C, Shen Y, Moussian B, Zhang CX: **A comprehensive omics analysis and functional survey of cuticular proteins in the brown planthopper.** *Proc Natl Acad Sci U S A* 2018, **115**:5175-5180.
 33. Rebers JE, Riddiford LM: **Structure and expression of a *Manduca sexta* larval cuticle gene homologous to *Drosophila* cuticle genes.** *J Mol Biol* 1988, **203**:411-423.
 34. Chen AY, Walker GP, Carter D, Ng JC: **A virus capsid component mediates virion retention and transmission by its insect vector.** *Proc Natl Acad Sci U S A* 2011, **108**:16777-16782.
 35. Killiny N, Harper SJ, Alfaress S, El Mohtar C, Dawson WO: **Minor coat and heat shock proteins are involved in the binding of citrus tristeza virus to the foregut of its aphid vector, *Toxoptera citricida*.** *Appl Environ Microbiol* 2016, **82**:6294-6302.
 36. Wang RY, Ammuar ED, Thornbury DW, Lopez-Moya JJ, Pirone TP: **Loss of potyvirus transmissibility and helper-component activity correlate with non-retention of virions in aphid stylets.** *J Gen Virol* 1996, **77**:861-867.
 37. Martin B, Collar JL, Tjallingii WF, Fereres A: **Intracellular ingestion and salivation by aphids may cause the acquisition and inoculation of non-persistently transmitted plant viruses.** *J. Gen. Virol.* 1997, **78**:2701-2705.

38. Uzeit M, Gargani D, Drucker M, Hebrard E, Garzo E, Candresse T, Fereres A, Blanc S: **A protein key to plant virus transmission at the tip of the insect vector stylet**. *Proc Natl Acad Sci U S A* 2007, **104**:17959-17964.
39. Parrish WB: **The origin, morphology, and innervation of aphid stylets (Homoptera)**. *Ann Entomol Soc Am* 1967, **60**:273-276.
40. Uzeit M, Gargani D, Dombrovsky A, Cazevielle C, Cot D, Blanc S:
 • **The “acrostyle”: a newly described anatomical structure in aphid stylets**. *Arthropod Struct Dev* 2010, **39**:221-229.
 This study reports the discovery of an organ in aphid stylets that harbors the receptor of CaMV and likely those of other a non-circulative virus.
41. Powell G: **Intracellular salivation is the aphid activity associated with inoculation of non-persistently transmitted viruses**. *J Gen Virol* 2005, **86**:469-472.
42. Gal-On A: **Zucchini yellow mosaic virus: insect transmission and pathogenicity – the tails of two proteins**. *Mol Plant Pathol* 2007, **8**:139-150.
43. Gera A, Loebenstein G, Raccach B: **Protein coats of two strains of cucumber mosaic virus affect transmission of *Aphis gossypii***. *Phytopathology* 1979, **69**:369-399.
44. Chen B, Francki RIB: **Cucumovirus transmission by the aphid *Myzus persicae* is determined solely by the viral coat protein**. *J Gen Virol* 1990, **71**:939-944.
45. Dombrovsky A, Sobolev I, Chejanovsky N, Raccach B: **Characterization of RR-1 and RR-2 cuticular proteins from *Myzus persicae***. *Comp Biochem Physiol B Biochem Mol Biol* 2007, **146**:256-264.
46. Brault V, Tanguy S, Reinbold C, Le Trionnaire G, Arneodo J, Jaubert-Possamai S, Guernec G, Tagu D: **Transcriptomic analysis of intestinal genes following acquisition of pea enation mosaic virus by the pea aphid *Acyrtosiphon pisum***. *J Gen Virol* 2010, **91**:802-808.
47. Badillo-Vargas IE, Rotenberg D, Schneeweis DJ, Hiromasa Y, Tomich JM, Whitfield AE: **Proteomic analysis of *Frankliniella occidentalis* and differentially expressed proteins in response to tomato spotted wilt virus infection**. *J Virol* 2012, **86**:8793-8809.
48. Kaur N, Chen W, Zheng Y, Hasegawa DK, Ling KS, Fei Z, Wintermantel WM: **Transcriptome analysis of the whitefly, *Bemisia tabaci* MEAM1 during feeding on tomato infected with the crinivirus, Tomato chlorosis virus, identifies a temporal shift in gene expression and differential regulation of novel orphan genes**. *BMC Genomics* 2017, **18**:370.
49. Stevens M, Freeman B, Liu HY, Herrbach E, Lemaire O: **Beet poleroviruses: close friends or distant relatives?** *Mol Plant Pathol* 2005, **6**:1-9.
50. Brault V, Uzeit M, Monsion B, Jacquot E, Blanc S: **Aphids as transport devices for plant viruses**. *C R Biol* 2010, **333**:524-538.
51. Mutterer JD, Stussi-Garaud C, Michler P, Richards KE, Jonard G, Ziegler-Graff V: **Role of the beet western yellows virus readthrough protein in virus movement in *Nicotiana clevelandii***. *J Gen Virol* 1999, **80**(Pt 10):2771-2778.
52. Falk BW, Tsai JH: **Biology and molecular biology of viruses in the genus *Tenuivirus***. *Annu Rev Phytopathol* 1998, **36**:139-163.
53. Sanders HR, Foy BD, Evans AM, Ross LS, Beaty BJ, Olson KE, Gill SS: **Sindbis virus induces transport processes and alters expression of innate immunity pathway genes in the midgut of the disease vector, *Aedes aegypti***. *Insect Biochem Mol Biol* 2005, **35**:1293-1307.
54. Colpitts TM, Cox J, Vanlandingham DL, Feitosa FM, Cheng G, Kurscheid S, Wang P, Krishnan MN, Higgs S, Fikrig E: **Alterations in the *Aedes aegypti* transcriptome during infection with West Nile, dengue and yellow fever viruses**. *PLoS Pathog* 2011, **7**: e1002189.
 This paper demonstrates that a cuticular protein interacts with flavivirus-mosquito-borne viruses.
55. Behura SK, Gomez-Machorro C, Harker BW, deBruyn B, Lovin DD, Hemme RR, Mori A, Romero-Severson J, Severson DW: **Global cross-talk of genes of the mosquito *Aedes aegypti* in response to dengue virus infection**. *PLoS Negl Trop Dis* 2011, **5**:e1385.