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Abstract:

The purpose of the article is to implement a holistic concept namely Quality by Design (QbD) approach for assessment of deep frying of potatoes chips. Critical quality attributes (CQAs), critical process parameters (CPPs) and quality target parameters (QTPs) were identified and measured all along the chips processing chain in 98 independent experiments. Temperature, time and oil quality usually used in the food industry were applied. Multilinear regression (MLR) was conducted to identify the variables (CQAs and CPPs) that could explain variation of the QTPs. An aggregation of significant QTPs was also performed in order to determine a single value that could express final products quality coupled to MLR analysis. It was possible to identify the main CQAs and CPPs that can explain the variation of some QTPs (colour a*, "flavour roast" sensory attribute, pentylfuran content and acrylamide content) as well as aggregated data.

Keywords: Quality by Design, Potatoe Chips, Deep frying, Multilinear regressions, temperature

1. INTRODUCTION

Food consumer and retailer expectations are incessantly increasing, market requires safe and nutritious food that looks appetizing, tastes good, at an affordable price and with a minimal environmental impact. To achieve consistency in all the product properties the process conditions (path to endpoint or process signature) must also be kept under statistical control [Kourti, 2006]. However, food materials are complex biological matrices, and the variability introduced by the sequence of unit operations in food processing directly influences the compositional and sensorial properties as well as the safety and the shelf-life of the final food products. To reduce this variability, the strategies based on Quality Assurance can be quite effective but are expensive and not flawless (Chen et al., 2011).

Therefore, the food producers must frequently manage poor repeatability of food quality attributes and batch failures; unsuitable or noncompliant batches must be discarded or reworked with high additional costs. To overcome these problems, the food industry is trying to shift to a novel holistic concept, the Quality by Design (QbD), which initially has been implemented by the pharmaceutical industry in 2004 by the United States Food and Drug Administration (FDA, 2004; Bakeev, 2010; van den Berg et al. 2013; Tajmmal Munir et al. 2015). The QbD hypothesis is that the quality of the food products should be incorporated during their development by precisely designing and controlling the process, and not by post-production quality testing (Rathore & Kapoor, 2017). Adoption of such innovative process concept can also give a broader view of the parameters to be optimized to ensure safe and high-quality food products (Cullen et al. 2014).

Examples of QbD applications in the food industry are increasing, even if examples of real industrial during-production monitoring are rare in the scientific literature because it might reveal confidential product and process information. In many cases there is, however, a clear

need to bridge the gap between the many promising scientific reports and actual use of these methods in the food industry (van den Berg et al. 2013; Panikuttira & O'Donnell 2018).

Among the industrial food processes, deep-frying is a common, but complex, multifunctional unit operation for fast dewatering, texturing or cooking foods, which simultaneously involves heat and mass transfer. One of the most widespread fried products are the potato chips, whose production embraces different steps, such as washing and peeling of raw materials, slicing, blanching and dewatering, etc. Deep frying is considered the more critical step, because the quality and safety of the final fried products are influenced by many factors, such as the nature and composition of fried materials, the combination of processing time and temperature, the heating profile, the oxidation status of frying oil, etc. (Rojo & Perkins, 1987; Vitrac et al., 2003; González-Martínez et al., 2004; Chatzilazarou et al., 2006; Romani et al., 2009; Kalogianni et al., 2010; Zhang et al., 2012; Kalogianni et al. 2017).

The main objective of the present study is to establish a Quality by Design approach in order to identify main quality parameters of the final products related with safety, taste and colour and to identified the useful quality and process parameters that can explain variation during production of deep-fried potatoes "chips". Another objective is the evaluation of suitable data aggregation strategies that could predict the quality and safety parameters of the final product.

2. MATERIAL AND METHODS

2.1 Fresh potatoes and frying oils

Homogenous 30 kg batches of potatoes (cultivar Agria) suitable for deep frying (Yang et al., 2016) were provided by Frufesc (Disbesa Grup, Barcelona, Spain) during a period of 5 month (from October to February). Each batch was used to carry out five frying experiments during

78 the same working day. The potato batch was randomly divided in 5 aliquots of 5 kg each,
79 which were processed sequentially along the same working day. Commercial fresh and
80 exhaust sunflower oil, commonly used in the industry were both provided by an industrial
81 manufacturer of potatoes chips (Grupo Siro, Palencia, Spain).

82

83 2.1 Frying equipment

84 The frying process was carried out with a continuous fryer model Frymatic24 (Nilma S.p.a.,
85 Parma, Italy), with a maximum capacity of 40 kg/h, and equipped with an original Distribute
86 Temperature Sensor (DTS) made by the Institute of Photonic Sciences (ICFO, Casteldefelds,
87 Spain). The DTS probes were based on Fibre Bragg Gratings (FBG) written in two optical
88 fibres. Each of the two probes consisted of five single FBGs, equi-spaced (15 cm) on the same
89 optical fibre, protected by a stainless tube and connected only at one end, on an armoured
90 patch-cord terminated with a FC/UPC connector. Therefore DTS probes recorded
91 simultaneously, each second, oil temperature in ten points along the frying tank (Figure 1).
92 Temperature values recorded by the two probes in the same position along the tank were
93 aggregated to define five temperature zones called E, M1, M2, M3 and Ex, where "E" zone
94 corresponded to the entrance of the potatoes in the frying tank, and the "Ex" zone
95 corresponded to the exit (Figure 1). Oil temperature was measured before starting (TO_{av}) and
96 during ($TC°_{E}$, $TC°_{M1}$, $TC°_{M2}$, $TC°_{M3}$ and $TC°_{Ex}$) frying process. The average temperature of
97 the oil ($TC°_{av}$) was also calculated as the average of all the values recorded in the five zones at
98 the same time.

99

100 2.2 Frying experiments

A specific design of experiment (DoE) was defined, based on 65 independent frying experiments for the calibration set and 33 independent frying experiments for the validation set. Independent variables considered in the DoE were: *i*) frying temperature (ranging from 150 °C to 175 °C; n = 5 levels), *ii*) time of frying (ranging from 150 to 180 seconds, n= 5 levels) and *iii*) oil quality (ranging from 100% fresh oil to 100% exhaust oil defined as used oil with a level of total polar material above 12%, n= 5 levels).

For all the frying experiments the same protocol was followed, which included: *i*) washing of the fresh potatoes with cold water and peeling (potato peeler M5, Sammic S.L., Azkoita - Spain) *ii*) immersion of peeled potatoes in cold-water, *iii*) slicing (Robot Coupe CL50 with a 1 mm disk, Dijon, France), and i*v*) final washing with cold water (5 °C).

Oil temperature and time of frying were precisely adjusted to the DoE by the controller of the continuous fryer. The frying tank was filled with 100 L of sunflower oil and oil quality was modified by mixing fresh with exhaust sunflower oil in established proportions according to the DoE. When oil reached the target temperature, a batch of about 4 kg sliced potatoes was loaded in the fryer.

2.4 Process monitoring and sampling

For each one of the 98 independents frying experiments (Calibration and Validation sets), nine CQAs of the raw material and nineteen CQAs (related to oil quality), were monitored during the frying process in addition to three critical process parameters (CPPs). Every day, before starting the frying experiments, ten potatoes were randomly selected from the potato batch, in order to assess the CQAs of the raw material before frying. Each sampled potato was cut in two halves; the first one was used to immediately measure the colour, the second one

was divided in five aliquots, which were separately packed in multilayer PP-aluminium bags and immediately stored at -80 ºC.

Oil samples were taken during each frying process with a stainless spoon; samples were immediately transferred in a 100 mL aluminium bottle (ISO Al 99.5; Bürkle, Bad Bellingen, Germany), refrigerated with liquid nitrogen and stored at -80 ºC for chemical analyses.

After processing, and taking out the first kg of sliced potatoes to stabilize the fryer, an aliquot of chips was taken for each one of the frying experiments, then packaged in multilayer PP-aluminium bags and immediately stored at -80 ºC for analysis of twelve QTPs (Quality target Parameters), including both chemical and sensorial parameters related with quality and safety. Average, standard deviation, maximum and minimum of all parameters (CQAs and CPPs) for calibration and validation sets are presented in table 1, while QTPs are presented in table 2.

2.4.1 Colour measurement

Instrumental colour parameters in fresh potatoes samples, before frying, were measured with a Konica Minolta chromameter Model CR-400 HS (Minolta, Tokyo, Japan) with an aperture of 8 mm. In potatoes chips, after frying, a Konica Minolta chromameter Model CR-410 HS (Minolta, Tokyo, Japan) with an aperture of 50 mm was used. In both cases, the equipment was set up for illuminate D65 (2º observer angle) and calibrated using a standard white reflector plate. On the Model CR-400 HS, 5 points were measured for each samples while for the Model CR-410 HS, 3 measurements were taken in succession on a batch of chips. Readings were obtained applying the standard CIE 1976 L^*, a^* and b^* (1976) colour system space.

2.4.2 Total Soluble Solids Content

147 Total Soluble Solids (TSS) content in fresh potatoes was determined by using a Quick Brix
148 TM90 (Mettler Toledo GmbH, Giessen, Germany). Potatoes samples were smashed, and one
149 drop placed on the refractometer glass, measurements were done in triplicate.

150 *2.4.3 Sugars Content*

151 Sucrose, Glucose and Fructose content in fresh potatoes were quantified by HPLC-RI
152 following the method of Folgado et al., (2014). Briefly, fresh potato samples (4 grams) were
153 homogenised and extracted two times with cold (-20 ºC) ethanol 95%. After centrifugation, an
154 aliquot of the ethanolic fractions was evaporated with N_2, re-dissolved in 0.5 mL of ultrapure
155 water, membrane filtered (pore size 0.2 μm) and injected in the HPLC system (20 μL).
156 Chromatographic separation was carried out with a binary pump 515 equipped with a 2414
157 Refractive Index detector (Waters, Milford MA, USA) and an Aminex HPX-87C 300 x 7.8
158 mm column (Bio-Rad, CA, USA) thermostated at 80 ºC. Isocratic elution was carried out with
159 ultrapure MilliQ® water (Merck KGaA, Darmstadt, Germany) at a flow of 0.6 mL/min., and
160 quantification was made with an external calibration curve.

161

162 *2.4.4 Oil oxidation parameters*

163 Total Polar Material (TPM) in oil was quantified during frying with a cooking oil tester mod.
164 270 (Testo, Lenzkirch, Germany). Results were express in percentage (%) of Total Polar
165 Material. Data was collected in triplicate during each frying process. Peroxide Index, Acidity
166 Index and p-anisidine value in frying oil were assessed with a FoodLab Fat system (CDR s.r.l,
167 Florence – Italy) following the protocols and the reactants provided by the fabricant.

168

169 *2.4.5. Fatty acids profile*

Fatty acids profile in frying oil was analysed according to Mach et al. (2006). Fatty acid methyl esters (FAMEs) were obtained by following the ISO method 5509E (ISO 5509E, 1978) and analysed using an HP 5890 Series II gas chromatograph (Hewlett Packard SA, Barcelona, Spain). Individual fatty acids (FA) were identified by comparison of their retention times with those of pure standards. Quantification was made by using an internal standard calibration with glyceryl tritridecanoate.

2.4.6. Volatile compounds

Furan, acrolein, hexanal, pentylfuran and 2,4-decadienal in sunflower oils and chips were analysed by SPME-GC/MS with a 6850 Network GC system equipped with a 5975C VL MS axis detector (Agilent Technologies, Santa Clara, CA, U.S.A.) and a Combi Pal autosampler (CTC Analytics AG, Zwingen, Switzerland). One gram of sample was added with 1 µL of mixed internal standard solution (acrolein-^{13}C and hexanal-d_{12}, both at 100 mg/L in isopropanol) in a 10 mL glass vial, vortexed for 30 seconds and pre-incubated at 50 ºC for 2 min at a speed of 500 rpm. A SPME DVB/CAR/PDMS fibre assembly (Supelco, Bellefonte - USA) was used with an extraction time of 30 min and constant agitation at 40 ºC. The chromatographic separation was carried out on a DB-5MS column (30 m, 0.250 mm ID, 1.00 µm film thickness; Agilent J&W GC Columns, Santa Clara CA, USA) with helium as carrier gas at a flow of 0.8 mL/min. Initial temperature of the oven was set at 33 ºC, then followed by a 2 ºC/min ramp up to 50 ºC, a 3 ºC/min ramp up to 72 ºC, a 6 ºC/min ramp up to 180 ºC and a 10 ºC/min ramp up to 220 ºC. For quantification purposes, aliquots of samples were spiked with defined amounts of labelled (acrolein-^{13}C and hexanal-d_{12}) and unlabelled compounds in different mass ratios. The ratios of the area counts for the specific ions of the analytes and the labelled standards were plotted against the ratio of the corresponding concentrations, and the response factors were calculated according to Ewert et al. (2011).

195

196 *2.4.7 Acrylamide assessment*

197 Acrylamide was quantified in frying oil and chips by HPLC-MS. One gram of frying oil or
198 potato chips were extracted following the protocol of Al-Taher (2012) based on Quechers.
199 Ten μL of the purified extracts were injected in the Agilent 1200 Series HPLC system,
200 equipped with an Agilent 6100 Series Single Quadrupole MS detector (Agilent Technologies,
201 Inc., CA, USA) and a reverse phase C_{18} column (2.1 i.d. x 100 mm, 3 μm). Elution was
202 carried out isocratically with mobile phase A (water: methanol:formic acid 97.4:2.5:0.1) at a
203 flow rate of 0.2 mL/min. MS detector was operated in positive electrospray ionization mode,
204 and the ion with *m/z* = 72, corresponding to the $[M-H]^+$ of the acrylamide, was monitored.
205 Quantification was made considering the response of the ion with *m/z* = 75, corresponding to
206 the molecular ion of the internal standard (acrylamide 13^C-3).

207

208 *2.4.8 Quantitative Descriptive Analyses*

209 Five Sensory descriptors ("odour roast", "flavour rancid", "flavour roast", "crunchy" and "oil
210 mouth feel") were generated by open discussion in two preliminary sessions by eight trained
211 assessors. A non-structured scoring scale was used, where 0 meant the absence of the
212 descriptor and 10 meant the highest intensity of the descriptor. Sensory evaluation was
213 performed for each session time in two sessions (per sampling time) using chips samples
214 corresponding to a frying experiment. Samples were coded using three random numbers and
215 presented to assessors. The first order and the carry-over effects were balanced according to
216 MacFie et al., (1989). For each frying experiment, the average score of the assessors and
217 sessions have been calculated.

218

2.5. Modelling, Statistics and Aggregation

2.5.1 Multilinear regression and statistic values

Multilinear regression (MLR) coupled to a Step-Wise model (probability for entry: 0.1 and probability for removal: 0.1) was used to develop calibration models on the QTP values from 65 experiments. Two parameters, coefficient of determination of calibration (R^2_{cal}) and probability ($Pr > |t|$) for each explanatory variables (CQAs and CPPs) were reported. Models were determined using the XLSTAT Premium software version 2018.1 (Addinsoft, France).

The different model gives also a predictive equation and a root mean square error of calibration (RMSEC).

$$\mathrm{RMSEC} = \sqrt{\frac{1}{M-1} \times \sum_{i=1}^{M} \left(y_i^{ref} - y_i\right)^2} \qquad \text{EQ.01}$$

Where:

M is the number of samples

y_i^{ref} is the reference value for sample i

y_i is the predicted value for sample i

The different models were tested on a validation set of 33 experiments and the quality of the models on each QTP values was assessed with the root mean square error of prediction (RMSEP), coefficient of determination (R^2_{val}), Bias and range error ratio (RER):

$$\mathrm{RMSEP} = \sqrt{\frac{1}{M} \times \sum_{i=1}^{M} \left(y_i^{ref} - y_i\right)^2} \qquad \text{EQ.02}$$

$$\mathrm{Bias} = \frac{\sum_{i=1}^{M} (y_i^{ref} - y_i)}{M} \qquad \text{EQ.03}$$

$$\mathrm{RER} = \frac{y_{max}^{ref} - y_{min}^{ref}}{\mathrm{RMSEP}} \qquad \text{EQ.04}$$

Where:

240 y_{max}^{ref} and y_{min}^{ref} are respectively the maximum and minimum values of the validation set

241 *2.5.2 Data aggregation*

242 The idea to aggregate QTPs parameters is to have only one data to describe the quality of our
243 potatoes chips product using a mid-level fusion approach (Borràs et al. 2015). To do so a min-
244 max normalisation of selected quality target product profile was done using equation EQ. 05
245 followed by the weighting of normalised data (y_i^{norm}) before calculation of the aggregated
246 data (CDF_i) with EQ. 06.

247 $$y_i^{norm} = \frac{(y_i - y_{min})}{(y_{max} - y_{min})} \qquad \text{EQ.05}$$

248 $$CDF_i = \sum_{i=1}^{MN} \beta_i \times y_i^{norm} \qquad \text{EQ.06}$$

249 where M N is the number of selected QTPs, β_i is the weight a number between 0 and 1 and
250 have been selected by authors to give more importance to some QTP parameters.

251 Four "negative" quality attributes, colour parameter a*, sensory descriptor "flavour roast",
252 acrylamide content and volatiles content pentylfuran content, have been selected to be
253 aggregated. Four aggregated indexes CDF_{I1}, CDF_{I2}, CDF_{I3} and CDF_{I4} have been calculated
254 using EQ. 06 and different weights β_i. In the first aggregation $CDF_{I1,}$ all quality attributes had
255 the same weight [0.25, 0.25, 0.25, 0.25]. For the second one $CDF_{I2,}$ the weights of volatile
256 quality attribute have been reduced to 0.1 and the others increase to 0.3 in order to take more
257 into accounts safety attribute and attributes related with consumer perception. For the third
258 CDF_{I3} [0.2, 0.3, 0.4, 0.1] and fourth CDF_{I4} [0.2, 0.2, 0.5, 0.1] aggregation more emphasis was
259 given to safety issues realty with acrylamide content. In the first aggregation index, $CDF_{I1,}$ all
260 quality attributes [a*, roast, acrylamide, pentylfuran] had the same weight [0.25, 0.25, 0.25,
261 0.25]. For the second index, $CDF_{I2,}$ the weight of pentylfuran content has been reduced to 0.1
262 and the others increased to 0.3 in order to highlight safety (acrylamide content) and consumer

perception. For the third CDF_{I3} [0.2, 0.3, 0.4, 0.1] and fourth CDF_{I4} [0.2, 0.2, 0.5, 0.1] indexes more emphasis was given to safety issues related with acrylamide content. Weights for a*, flavour roast, acrylamide and pentylfuran are [0.25, 0.25, 0.25, 0.25] for CDF_{I1}, [0.3, 0.3, 0.3, 0.1] for CDF_{I1}, [0.2, 0.3, 0.4, 0.1], for CDF_{I3} and [0.2, 0.2, 0.5, 0.1] for CDF_{I4}. A principal component analysis (PCA) has been carried out on the four quality parameters and the first PCA factor was retained as an additional aggregated index (PCA factor 1).

3. RESULTS

Table 1 shows the average, standard deviation, maximum and minimum values for the selected CQAs as well as for CPPs for the calibration and validation sets. Most of the CQAs display important standard deviations indicating substantial variations in the composition of the raw materials and deep frying conditions and, therefore, including in the predictive models sources of variations usually found in the real processes.

3.1. Multilinear analysis on single QTPs parameters

The coefficient of determination from calibration set (R^2_{cal}), the root mean square error of calibration (RMSEC), the standardized regression coefficients and the p-values from the multilinear regressions calculation are presented in table 2. R^2_{cal} gives the strength of a relationship between exploratory variables and QTPs and it is generally admitted (Moore et al. 2013) that a coefficient above 0.7 indicates that the proposed model explains correctly the variation of the QTPs. Colour parameters a* and b*, sensory descriptors “Odour roast” and “Flavour roast” and volatile parameters hexanal and pentylfuran presented coefficients of determination above 0.7. Others QTPs such as sensory descriptor “Flavour rancid”, acrylamide content and 2.4 decadienal content, showed R^2_{cal} between 0.5 and 0.7, indicating

that the predictive models do not explain completely their variations. L*, sensory descriptors "crunchy" and "oil mouth feel" had R^2_{cal} below 0.5, indicating that our models do not explain their variation. Table 2 shows that, out of 29 explanatory variables, 2 to 8 have been retained to explain the variation of each QTPs. On the opposite, 7 explanatory variables (Fructose content, reducing sugars content, TPM, p-anisidine value, fatty acid (FA) 18:2 cis-9 trans-12, ∑FA ω6, ∑FA trans and monosaturated fatty acids or MUFA) have not been retained by none of the models to explain variation of the QTPs and were discarded.

MLR models describing QTPs a* and b*, retained respectively 4 and 8 exploratory variables related with raw materials, oil quality, volatile, fatty acids, variables related with oil temperature and process time. For sensory descriptors "odour roast" and "flavour roast", 5 and 4 explanatory variable were respectively retained, related with Sucrose content, L*, hexanal content, saturated FA, oil temperature $TC°_E$ and time. For acrylamide content, the MLR model retained 4 explanatory variables related with red colour, volatile, ratio ω6/ω3 and $TC°_E$ oil temperature. For QTPs volatiles pentylfuran and 2.4 decadienal, MLR model did not retain any explanatory variable of raw materials, but it retained oil quality parameters, volatile parameter, Saturated FA and $TC°_E$ oil temperature for the first. For QTP 2.4 decadienal only 4 explanatory variables related with oil quality, volatiles and fatty acids. For QTP hexanal, 3 explanatory variables are related with raw materials and 4 with oil characteristics (volatile and fatty acids).

In 5 of the 6 QTPs with a R^2_{cal} above superior to 0.7, exploratory variables related with CPPs have a positive standardized regression coefficients indicating that an increase of temperature or time will increase the different QTPs. Only sensory descriptor "flavour rancid" presents a negative standardized regression coefficient for the exploratory variables $TC°_{av}$. Considering raw materials and oil exploratory variables, positive and negative standardized regression coefficients have been calculated by the model for QTPs a*, b*, "odour Roast", "flavour

312 rancid" and "flavour roast". For volatiles, all QTPs present positive standardized regression
313 coefficients indicating that an increase of all exploratory variables will lead to an increase of
314 the volatiles in the chips. For acrylamide content, an increase of exploratory variable a* will
315 lead to an increase of acrylamide content while an increase of hexanal and ratio ω6/ω3 will
316 have the opposite effect.

317

318 3.2 Prediction with multilinear models

319 Multilinear model have been used to predict the evolution of selected QTPs with a validation
320 set of 33 experiments. Quality parameters of the prediction are reported in table 3. Taking into
321 account colour parameters of the potatoes chips, only a* presents a coefficient of
322 determination of validation (R^2_{val}) superior to 0.7. For colour parameter b*, results are
323 disappointing with R^2_{val} below 0.5. Models for the sensory descriptors "odour roast" and
324 "flavour rancid" have a R^2_{val} between 0.6 and 0.7, and "flavour roast" has a R^2_{val} above 0.7.
325 For the acrylamide content, when 2 outliers are removed from the analysis, R^2_{val} are between
326 0.5 and 0.7. Concerning the volatile parameter hexanal, the step-wise model give a R^2_{val}
327 below 0.5, while for volatile parameters pentylfuran and 2-4 decadienal, R^2_{val} are between 0.5
328 and 0.7.

329 To summarise, only 2 QTPs (a* and "flavour roast") have a R^2_{val} above 0.7, while others 5
330 ("odour roast", acrylamide content; hexanal, pentylfuran and 2.4-decadienal) have a R^2_{val}
331 between 0.5 and 0.7. The quality of the models could also be provided by the RER
332 parameters. The QTP acrylamide gives a value of RER of 5.0, while our best predictive
333 models were obtained for sensory descriptors "flavour rancid" and "odour roast" with a
334 respective RER of 6.9 and 6.6. The best RER values ranged between 4.0 and 10.0 indicating
335 that our models have a performance corresponding to screening target (AACC Method 39-
336 00.01).

337

338 3.3 Aggregation of QTPs parameters

339 The contribution of each QTPs to the first PCA factor was 37.2% for a*, 27.8% for "flavour
340 roast", 27.4% for acrylamide content and 7.6% for pentylfuran. Multilinear regression
341 analyses were conducted on different aggregated indexes and results on the calibration set are
342 shown in Table 4. R^2_{cal} is above superior to 0.7 for 3 of the 4 indexes, CDF_{I4} being the
343 exception with a value of 0.692, and for the first PCA factor, thus indicating that our models
344 can explain the variation of aggregated chips quality parameters. It can be noted that, an
345 increase of the weight of acrylamide content in aggregated indexes, had the effect to reduce
346 R^2_{cal}. Number of explanatory variables retained by the MLR model have been reduced to 7: a*
347 in CDF_{I2}, CDF_{I3} and CDF_{I4}; b* in only one case (CDF_{I1}), when all selected QTPs have the
348 same weight; glucose content in only one case (CDF_{I4}), when the weight of acrylamide
349 content has been set up at 0.5; hexanal volatile content of the oil in CDF_{I2}, CDF_{I3} and CDF_{I4};
350 ω6 content of the oil in only one case (CDF_{I1}); MUFA in CDF_{I2} and CDF_{I3}; Oil temperature
351 $TC°_E$ in all aggregated index. It is significant that all oil quality parameters (TPM, acidity, p-
352 anisidine and peroxide value) have been discarded by the model as well as Time. All
353 standardized regression coefficients of oil temperature $TC°_E$ are positive as well as MUFA
354 and a* and glucose when they are retained by the model. On the contrary, b*, hexanal and ω6
355 present a negative standardized regression coefficients when they are retained.

356 Models have been applied to the validation data set to explain the variation of our aggregated
357 indexes (table 5). Predictive results of the variation of CDF_{I1}, CDF_{I2} and CDF_{I3} are
358 encouraging with R^2_{val} between 0.668 and 0.728. RER values are between 6.2 and 7.8,
359 indicating a performance target corresponding to screening target. Although first PCA factor
360 shows the best coefficient of determination of validation R^2_{val} with one outlier, the aggregated
361 index CDF_{I2} explained by the Step-Wise model seems to be a good option (Figure 2). Model

362 for the aggregated index CDF_{I2} used only 4 explanatory variables (colour a*, hexanal content,
363 MUFA and oil temperature $TC°_{E}$), had a R^2_{val} of 0.718 and no outliers in the validation set.

364

365 ## 4. DISCUSSION

366 In order to define the final chips product a total of 12 QTPs, including 3 colour parameters, 5
367 sensory attributes, 3 volatiles parameters and acrylamide content, have been used. Usually,
368 research works evaluate the impact of some processing parameters on single compounds, like
369 the acrylamide content (Zhang et al. 2015) or texture and oil intake in the potatoes (Pedeschi
370 et al. 2005) but few had a more global approach (Yang et al 2016; Santos et al. 2018).

371 In the present study only results from MLR algorithm are presented even if non-linear
372 algorithms (Random forest regression and log-linear regression models) have been tested on
373 our dataset. Results of non-linear algorithms have proven to be disappointing. The limited
374 number of independent experiments seems to be a limiting factor to use such non-linear
375 approaches.

376 Our results show that colour parameters L* and a* had a significant variation that can be
377 explained by CPPs parameters such as the average oil temperature. Yang et al. (2016) had
378 have compared the evolution of colour of potatoes strips retrieved issue from Agria,
379 Kennebec and Red Pontiac cultivars regarding oil temperatures and frying time 190°C / 160 s,
380 170°C / 240 s, 150°C / 330 s. In contrast with our results, few colour variations of the final
381 products have been measured for Agria cultivar, much more have been detected for the other
382 two cultivars. Pedreschi et al. (2005) proved that the oil temperature and time of frying is
383 related to the colour a* parameter of the potato and the acrylamide formation. Our predictive
384 results for acrylamide are lower than expected but some positive points could be extracted.
385 Yang et al. (2016) established that the correlations between selected studied factors of raw

materials (such as asparagine, fructose, glucose, sucrose, reducing sugar, oil uptake, colour L*, colour b* and shear force) were significant to explain the acrylamide content in the final product. Some of the parameters have been measured in our study and the explanatory variables colour a*, hexanal content, ratio ω6/ω3 and average frying temperature have been used by the MLR model to explain and predict the variation of acrylamide content. Our study, as a new approach, took into account sensory attributes, because chip taste is related with Maillard reactions, which is the main responsible for the formation of acrylamide (Lee & Shibamoto, 2002). However, no clear relationship (R^2<0.5) could be found between measured acrylamide content and sensory descriptors or other compositional parameters of potatoes chips. Even if such results are in discrepancy with finding of Pedreschi et al. (2005), it should be pointed out that a different cultivar was used (Agria versus Panda) and that our experiment was carried out with a continuous semi-industrial fryer and using oil at different degree of oxidation to mimic the industrial condition. On the other hand, formation of acrylamide involves complex mechanism reactions that probably the CQAs and CPPs included in the model cannot describe completely (Purlis, 2010).

Aggregated indexes with different QTPs parameters describing potatoes chips characteristics have also been analysed, in order to predict a global potatoes chips quality. In food science, low and mid-level data fusion have been undertaken for a wide range of applications such as quality parameters correlation, sensory properties assessment, cultivar selection or origin authentication (Borras et al., 2015). In our case, four parameters describing potatoes chips have been used, and different weight has been given to acrylamide content. Using aggregated data indexes a compromise have been found between the need to obtain safe products with lower acrylamide contents, but taking into accounts the sensory profile. Whatever the aggregated index selected to obtain the "best product", within the experimental domain here studied and with our frying equipment, we should use fresh potatoes with highest intensity of

yellow/green colour (highest b* and lowest a* values) and the lowest frying oil temperature (150 ºC). As time did not appear as an explanatory variable in aggregated indexes, we could use the shortest time (150 seconds) to achieve the maximum production efficiency. If we consider CDF_{I4}, which gives more importance to acrylamide content, fresh potatoes with the lowest glucose content should be selected. MUFA, hexanal and ω6 oil contents are indicators of the oil quality. The variation of these parameters with respect to those of the fresh oil could be used to establish the oil turnover, which will depend on the aggregated index selected.

In the present work, online measurements were possible for some of the attributes, such as colour parameters (L*, a* and b*) in raw materials, oil quality (TPM) and process parameters (time and temperature), but others key parameters (sugar content of raw materials, volatiles, fatty acids) were analysed off-line at laboratory scale. So, future improvements of Quality by Design approach are also strictly linked to the implementation of suitable online analytical methods for a comprehensive monitoring of the process.

5. CONCLUSION

The Quality by Design approach has been used to identify the main quality and process parameters that can be modified for the production of deep-fried potatoes "chips". To conduct processing, specific target parameters related with sensory descriptors could be predicted with MLR models with some accuracy by measurement of few explanatory variables related with potatoes brightness, oil volatile, saturated fatty acid and oil temperature, but for safety issues such as acrylamide content the predictive models are far from satisfactory. A general aggregated index incorporating 4 different quality parameters of the chips can be predicted with a reasonable accuracy, and can be used to establish the optimal process conditions. They are still a number of complex mechanisms and factors to be identified that can influence the quality parameters of potatoes chips. The work had shown the need of further studies to

explore the data fusion strategies for quality parameters of the final products to define single parameter that can be easily predicted and still full fit the goal to optimise sustainable processing.

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447 REFERENCES

448 Al-Taher F. (2012). Analysis of acrylamide in French fries using Agilent bond Elut Quechers
449 AOAC kit and LC/MS/MS. *Agilent Technologies application note nº5990-5940EN.*
450 https://www.agilent.com/cs/library/applications/5990-5940EN.pdf

451 Bakeev K.A. and Menezes J.C. (2010). *Chapter 16, Future Trends for PAT for Increased*
452 *Process Understanding and Growing Applications in Biomanufacturing*. In Process
453 Analytical Technology: Spectroscopic Tools and Implementation Strategies for the Chemical
454 and Pharmaceutical Industries. Edited by K. Bakeev, a Wiley & Sons Ltd., Publication,

455 Borràs E., Ferré J., Boqué R., Mestres M., Acena L. and Busto O. (2015). Data fusion
456 methodologies for food and beverage authentication and quality assessment – a review.
457 *Analytica Chimica Acta*, 891, 1-14.

458 Chatzilazarou A., Gortzi O., Lalas S., Zoidis E. and Tsaknis J. (2006). Physicochemical
459 changes of olive oil and selected vegetable oils during frying. *Journal of Food Lipids*, 13(1),
460 27 – 35. ·

461 Chen W.-A., Chiu C. P., Cheng W.-C., Hsu C.-K. and Kuo M.-I. (2013). Total polar
462 Compounds and acid values of repeatedly used in frying oils measured by standard and rapid
463 methods. *Journal of Food and Drug Analysis*, 21(1), 58-65

464 Cullen P.J., O'Donnell C.P. and Fagan C.C. (2014). *Benefits and Challenges of Adopting PAT*
465 *for the food industry*. In Process Analytical Technology for the Food Industry, C.P. O'Donnel,
466 C. Fagan and P.J Cullen editors).

467 Ewert A., Granvolg M. and Schierberle (2011). Development of Two Stable Isotope Dilution
468 Assays for the Quantitation of Acrolein in Heat-Processed Fats. *Journal of Agriculture Food*
469 *Chemistry*, 59 (8), pp 3582–3589

FDA, U.S. Department of Health and Human Services, Food and Drug Administration (2004). Guidance for industry: PAT—a framework for innovative pharmaceutical development, manufacturing, and quality assurance. https://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/ucm070305.p. Consulted 18th June 2018.

Folgado R., Sergeant K., Renaut J., Swennen R., Hausman J.F. and Panis B. (2014). Changes in sugar content and proteome of potato in response to cold and dehydration stress and their implications for cryopreservation. *Journal of Proteomics*, 98:99-111.

González-Martinez G., Ahrné L., Gekas V. and Sjöholm I. (2004). Analysis of temperature distribution in potato tissue during blanching and its effect on the absolute residual pectin methylesterase activity. *Journal of Food Engineering*, 65, 433-441.

Kalogianni E.P., Karastogiannidou C. and Karapantsios T.D. (2010). Effect of potato presence on the degradation of extra virgin olive oil during frying. *International of Food Science and Technology*, 45, 765 – 775.

Kalogianni E.P., Georgiu D, Romaidi M., Exarhopoulos S., Petridis D., Karastogiannidou C., Dimitreli G. and Karakosta P. (2017). Rapid Methods for Frying Oil quality Determination: Evaluation with Respect to Legislation Criteria. *Journal of the American Oil Chemist' Society*, 94, 19-36.

Kersey A.D., Davis M.A., Patrick H. J., LeBlanc M., Koo K. P., Askins C. G., Putnam M.A. and Friebele E.J. (1997). *Fiber grating sensors*. *Journal of Lightwave Technology*, v15(8), 1442-1463.

Kourti T. 2006. Process Analytical Technology Beyond Real-Time Analyzers: The Role of Multivariate Analysis. *Critical Reviews in Analytical Chemistry*, 36: 257–278.

493 Mach, N., Devant, M., Díaz, I., Font-Furnols, M., Oliver, M.A., García Regueiro, J.A. and
494 Bach, A. (2006). Increasing the amount of omega-3 fatty acid of meat from young Holstein
495 bulls through nutrition. *Journal Animal Science,* 84, 3039–3048.

496 MacFie, H. J., Bratchell, N., Greenhoff, H., and Vallis, L. V. (1989). Designs to balance the
497 effect of order of presentation and first-order carry-over effects in hall test. *Journal of Sensory*
498 *Studies,* 4, 129–149.

499 Moore, D. S., Notz, W. I, and Flinger, M. A. (2013). *The basic practice of statistics* (6th ed.).
500 New York, NY: W. H. Freeman and Company. Page (138)

501 Lee, K.-G. and Shibamoto, T. (2002). Toxicology and antioxidant activities of non-enzymatic
502 browning reaction products: Review. *Food Review International*,18, 151-175

503 Panikuttira B. and O'Donnell C.P. (2018). *Process Analytical Technology for the Fruit Juice*
504 *Industry*. In "Fruit Juices. Extraction, Composition, Quality and Analysis". Rajauria G. &
505 Tiwari B.K. Editors., Academic Press Publisher (Cambridge, US). pp. 835–847 .

506 Pedreschi F., Moyano P., Kaack K. and Granby K. (2005).Color changes and acrylamide
507 formation in fried potato slices. *Food Research International,* 38, 1-9..

508 Purlis E. (2010). Browning development in bakery products – A review. *Journal of Food*
509 *Engineering*, 99, 239-249.

510 Rathore A. and Kapoor G. (2016). Im^lmentation of Quality by Design for processing of
511 food products and biotherapeutics. *Food and Bioproducts Processing*, 99, 231-243.

512 Rojo J.A. and Perkins E.G. (1987). Cyclic fatty acid monomer formation in frying fats. I.
513 Determination and structural study. *Journal of the American Oil Chemist' Society*, 64(2), 414-
514 421.

515 Romani S., Bacchiocca M., Rocculi P. and Dalla Rosa M. (2009). Influence of frying
516 conditions on acrylamide content and other quality characteristics of French fries. *J. Food*
517 *Compos. Analysis*, 22: 582–588.

518 Santos C.S.P., Molina-garcia l., Cunha S.C. and Casal S. (2018). Fried potatoes: impact of
519 prolonged frying in monounsatureated oils. *Food Chemistry*, 243, 192-201.

520 Tajmmal Munir M., Yu W., Young B.R. and Wilson D.I. (2015). The current status of process
521 analytical technologies in the dairy industry. *Trend in Food science & technology*, 43, 205-
522 218

523 van den Berg F., Lyndgaard C.B., Sorensen K.M. and Engelsen S.B. (2013). Process
524 analytical technology in the Food Industry- a review. *Food Science &Technology,* 31, 27-35.

525 Vitrac O., Trystram G. and Raoult-Wack A.L. (2003). Continuous measurement of convective
526 heat flux during deep-frying: validation and application to inverse modelling. *Journal of Food*
527 *Engineering*. 60: 111–124.

528 Yang Y., Achaerandio I. and Pujolà M. (2016). Classification of potato cultivars to establish
529 their processing aptitude. *Journal of Food Science and Agriculture*, 96, 413-421.

530 Zhang Q., Saleh A.S.M., Chen J., and Shen Q. (2012). Chemical alterations taken place
531 during deep-fat frying based on certain reaction products: A review. *Chemistry and Physics of*
532 *Lipids.* 165: 662–681

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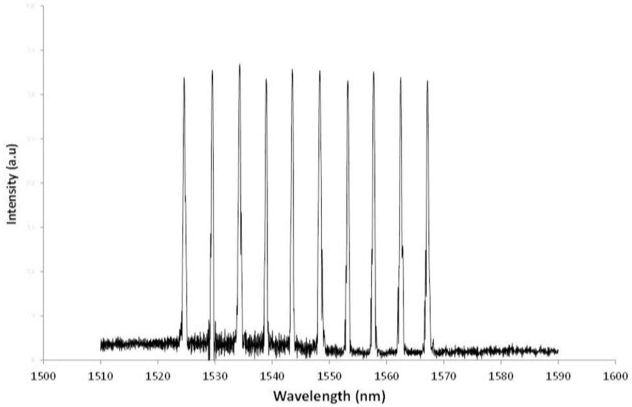
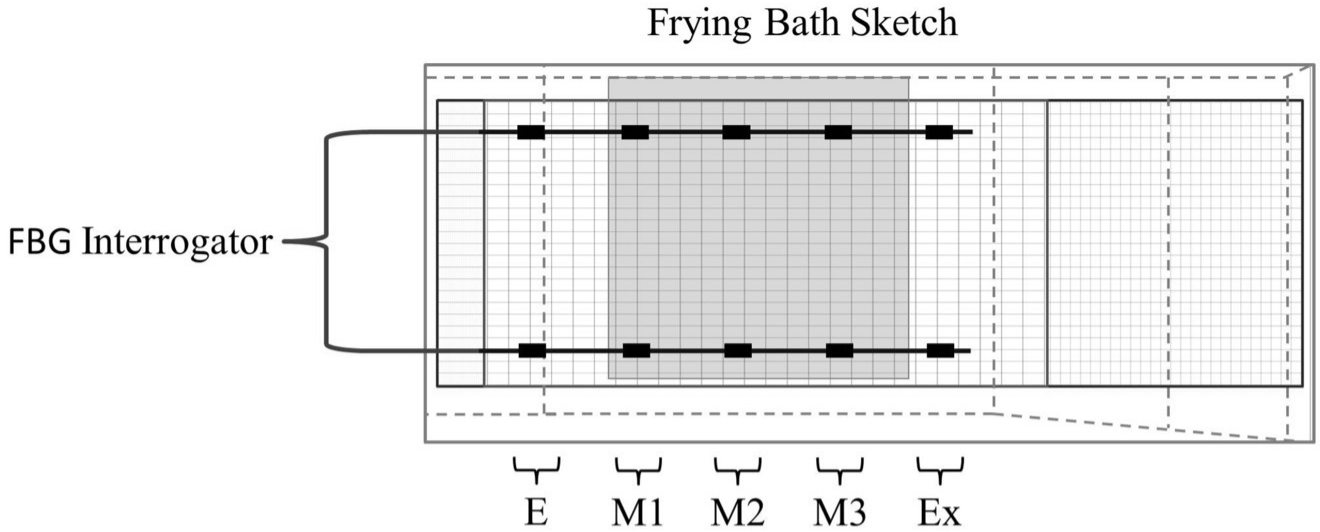


FBG Interrogator signal



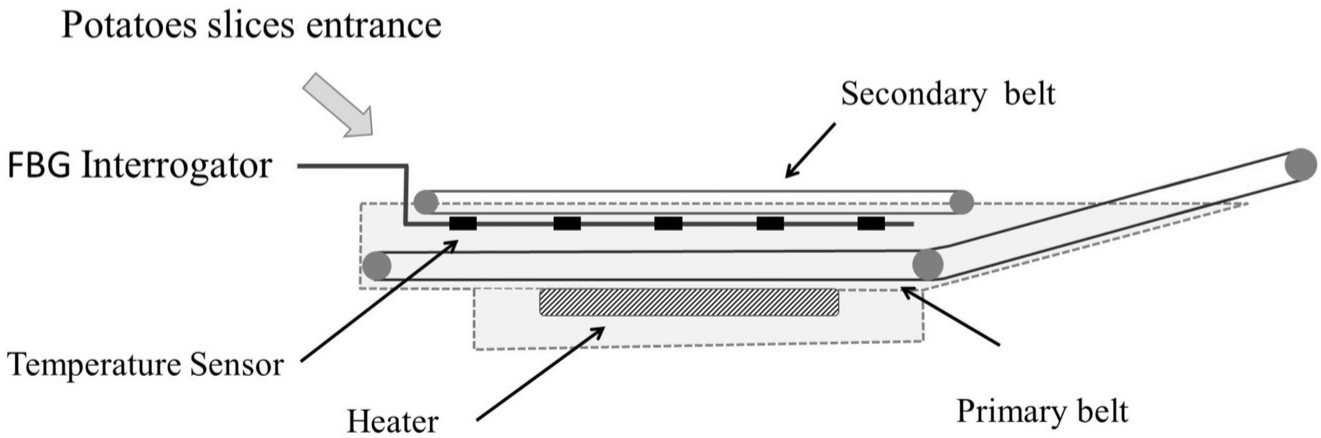
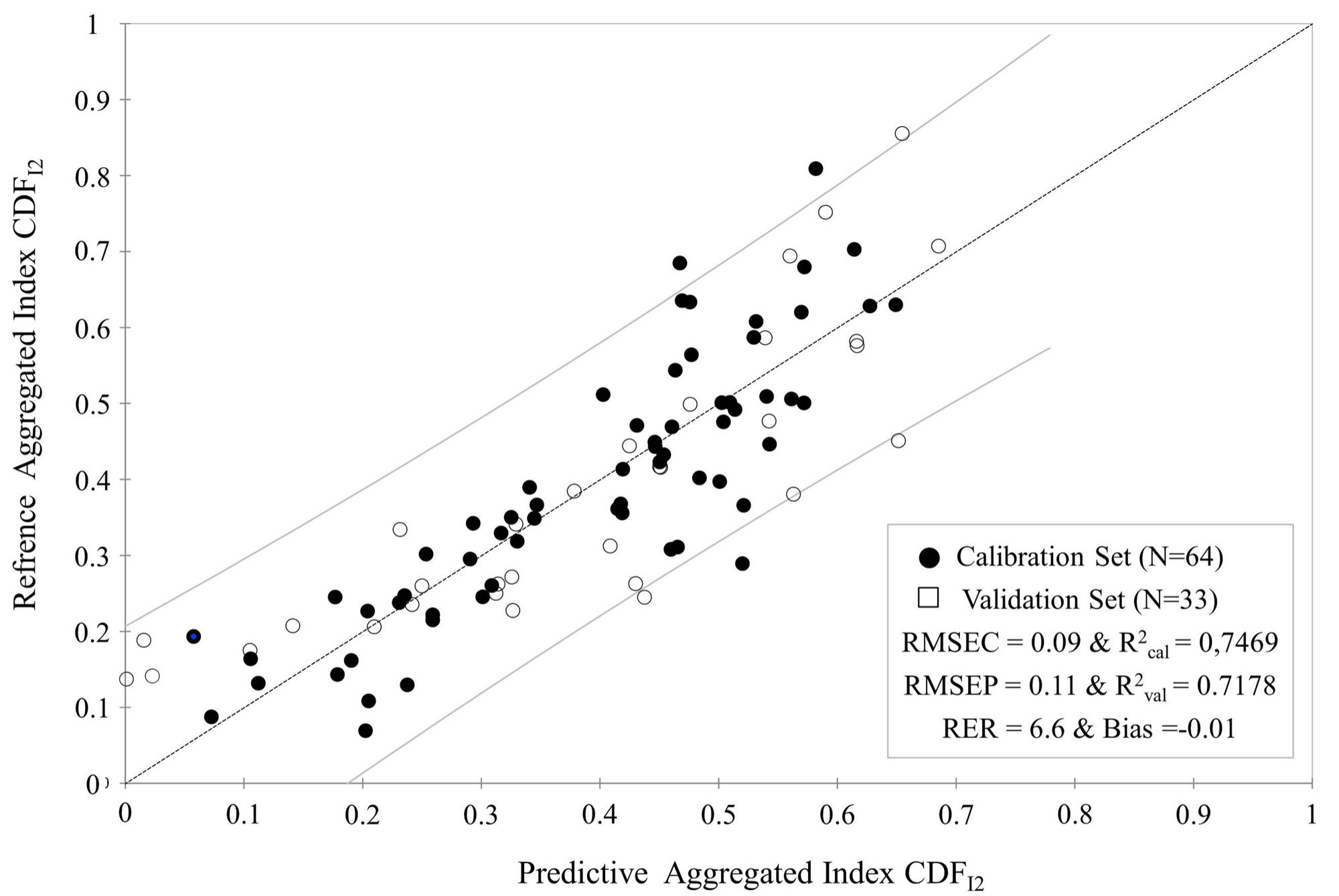


Figure 1


Refrence Aggregated Index CDF_{I2}
Predictive Aggregated Index CDF_{I2}
0
0.1
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1
Calibration Set (N=64)
Validation Set (N=33)
RMSEC = 0.09 & R^2_{cal} = 0,7469
RMSEP = 0.11 & R^2_{val} = 0.7178
RER = 6.6 & Bias =-0.01

Figure 2

Table 1: Mean ±standard deviation (SD), maximum and minimum of the different critical quality attributes (CQAs) and Quality Process Parameters (CPPs) measured for the calibration set (N=65) and Validation Set (N=33). TPM stands for total polar materials; FA stands for fatty acid; MUFA stands for monosaturated fatty acids; PUFA stands for polysaturated fatty acids.

		Calibration Set (N = 65)			Validation Set (N=33)		
		Mean	Max	Min	Mean	Max	Min
Potatoes CQAs	$L^*_{(CIELAB)}$	66.4 ±1.1	68.5	62.7	66.4 ±1.6	68.5	62.7
	$a^*_{(CIELAB)}$	-3.6 ±0.8	-2.7	-5.6	-3.6 ±0.9	-2.7	-5.6
	$b^*_{(CIELAB)}$	14.4 ±5.0	25.1	10.2	14.4 ±5.4	25.1	10.2
	TSS (°Brix)	1.8 ±0.3	2.5	1.2	1.8 ±0.4	2.5	1.2
	Sucrose (mg/100L)	575 ±157	759	217	633 ±124	759	217
	Glucose (mg/100 L)	215 ±128	500	26	236 ±146	500	26
	Fructose (mg/100 L)	299 ±60	447	198	319 ±64	447	198
Oil CQAs	TPM (%)	8.6 ±3.6	15.1	1.1	8.3 ±3.6	14.3	1.7
	Acidity index (%)	0.30 ±0.22	0.81	0.03	0.26 ±0.19	0.73	0..04
	p-anisidine value	14.2 ±15.3	46.9	0.5	13.3 ±15.7	48.6	0.5
	Peroxide index (meqO2/kg)	4.8 ±2.8	14.5	1.0	4.2 ±2.2	10.4	1.2
	Acrolein (ppb)	499 ±245	1205	150	548 ±237	1017	155
	Furan (ppb)	38 ±28	139	1	35 ±25	133	4
	Hexanal (ppm)	2.15 ±0.77	5.21	0.59	2.26 ±0.78	4.40	1.24
	Pentylfuran (ppm)	1.71 ±0.68	3.69	0.12	1.76 ±0.81	5.10	0.51
	2,4-decadienal (ppm)	137 ±96	445	0	158 ±115	553	23
	FA 18:1 trans ω9 (%)	0.13 ±0.08	0.28	0	0.11±0.07	0.27	0.00
	FA 18:2 cis-9 trans-12 (%)	0.07 ±0.02	0.16	0.04	0.07±0.01	0.09	0.02
	FA 18:2 trans-9 cis-12 (%)	0.07 ±0.01	0.11	0.05	0.07±0.01	0.10	0.04
	∑FA ω6 (%)	8.4 ±1.1	10.0	6.5	8.5 ±1.0	9.8	6.5
	∑FA trans (%)	0.27 ±0.07	0.41	0.13	0.25 ±0.07	0.38	0.14
	Ratio ω6/ ω3	152 ±64	414	40	147 ±56	229	26
	∑FA ω3 (%)	0.06 ±0.03	0.24	0.02	0.07 ±0.05	0.32	0.04
	Saturated FA (%)	9.3 ±0.2	9.8	8.9	9.3 ±0.3	9.8	8.8
	MUFA (%)	82 ±1	84	80	82 ±1	84	80
	PUFA (%)	8.4±1.1	10.0	6.5	8.5 ±1.0	9.9	6.6
CPPs	Time (s)	164 ±10	180	150	164 ±10	180	150
	$TC°_{av}$ (°C)	159 ±7	172	147	158 ±8	172	147
	$TC°_{E}$ (°C)	157 ±7	170	142	156 ±8	169	144

Table 2: Standardized regression coefficients and p-value (Pr > |t|) in parenthesis of the F statistic from an analysis of variance (ANOVA) and coefficient of determination R^2_{cal} , Root Mean Square Error of calibration (RMSEC) of the multi linear regression (MLR) using the model Step-wise (probability for entry: 0.1 and probability for removal: 0.1) for the different QTPs of potatoes chips. FA 18:2 trans(2) stands for FA 18:2 trans-9 cis-12; FA stands for fatty acid; PUFA stands or polysaturated fatty acids.

	Quality Target Parameters (QTPs) of potatoes chips											
	Colour			**Sensory**					**Safety**	**Volatiles**		
	$L^*_{(CIELAB)}$	$a^*_{(CIELAB)}$	$b^*_{(CIELAB)}$	Odour Roast	Flavour rancid	Flavour Roast	Crunchy	Oil Mouth feel	Acrylamide	Hexanal	Pentylfuran	2.4decadienal
R^2_{Cal}	0.375	0.711	0.739	0.777	0.633	0.764	0.439	0.480	0.539	0.729	0.755	0.642
RMSEC	3.6	1.4	1.7	0.7	0.7	0.8	0.5	0.6	0.68 ppm	99 ppb	82 ppb	10 ppm
$L^*_{(CIELAB)}$						0.12 (0.066)		0.33 (0.004)		0.26 (0.004)		
$a^*_{(CIELAB)}$									0.46 (<0.001)			
$b^*_{(CIELAB)}$		-0.20 (0.037)	0.51 (< 0.001)									
TSS			-0.17 (0.020)					-0,23 (0.031)		-0.22 (0.010)		
Sucrose				-0.17 (0.056)	-0.16 (0.070)					-0.35 (<0.001)		
Glucose		0.39 (< 0.001)			0.39 (<0.001)							
Acidity			-0.49 (<0.001)				-0.45 (< 0.001)	-0.28 (0.095)			0.36 (0.006)	
peroxide											0.16 (0.089)	0.21 (0.018)
Acrolein			-0.17 (0.042)					-0.22 (0.034)				
Furan				-0.32 (0.002)	-0.30 (0.024)	-0,14 (0.039)	0.18 (0.093)					
Hexanal	0.41 (<0.001)	-0.19 (0.017)					0.26 (0.009)		-0.35 (<0.001)	0.20 (0.028)		0.37 (< 0.001)
Pentylfuran					0.36 (0.007)					0.43 (< 0.001)	0.47 (< 0.001)	
2.4-decadienal										-0.21 (0.013)		0.30 (0.001)
FA 18:1 trans ω9				0.21 (0.043)				-0.36 (0.320)				
FA 18:2 trans(2)										0.33 (<0.001)		
∑FA trans												
Ratio ω6/ ω3									-0.21 (0.063)			0.30 (0.001)
∑FA ω3			0.17 (0.025)		-0.20 (0.037)							
Saturated FA						-0,16 (0.020)					0.33 (< 0.001)	
PUFA			-0.34 (0.015)									
Time (s)			0.14 (0.063)	0.13 (0.083)	0.20 (0.027)							
$TC°_{av}$ (°C)			0.40 (< 0.001)		-0.44 (< 0.0001)		0.42 (< 0.0001)					
$TC°_{E}$ (°C)	-0.48 (< 0.001)	0.79 (< 0.001)		0.77 (< 0.001)		0.83 (< 0.001)			0.54 (< 0.001)		0.23 (0.002)	

Table 3: Validation of the different models used to explain the variability of selected QTPs. N_v: number of experiments from the validation set; R^2_{Val}: coefficient of determination of the validation set; RMSEP: root mean square error of prediction; Bias: model bias; RER: range error ratio.

QTPs	N_v	R^2_{Val}	RMSEP	Bias	RER
$a^*_{(CIELAB)}$	33	**0.789**	1.6	0.0	5.1
$b^*_{(CIELAB)}$	31	0.316	2.5	-0.4	4.8
Odour Roast	32	0.656	0.8	0.0	6.4
Flavour Rancid	33	0.614	0.7	0.0	6.9
Flavour Roast	33	**0.736**	0.9	0.0	6.6
Acrylamide (ppm)	31	0.520	0.9	0.0	5.0
Hexanal (ppb)	32	0.319	137	13	4.1
Pentylfuran (ppb)	32	0.613	91	25	5.7
2.4decadienal (ppm)	32	0.514	10	1.4	5.5

Table 4: Standardized regression coefficients and p-value (Pr > |t|) in parenthesis of the F statistic from an analysis of variance (ANOVA) and coefficient of determination R^2_{cal} , Root Mean Square Error of calibration (RMSEC) of the multi linear regression (MLR) using the model Step-wise (probability for entry: 0.1 and probability for removal: 0.1) for PCA factor 1 and aggregated indexes CDF_{I1}, CDF_{I2}, CDF_{I3} and CDF_{I4}. MUFA stands for monosaturated fatty acids

	CDF_{I1}	CDF_{I2}	CDF_{I3}	CDF_{I4}
R^2_{Cal}	0.778	0.747	0.719	0.692
RMSEC	0.08	0.09	0.09	0.10
$a^*_{(CIELAB)}$		0.29 (<0.001)	0.33 (<0.001)	0.39 (< 0.001)
$b^*_{(CIELAB)}$	-0.27 (0.001)			
Glucose				0.23 (0.010)
Hexanal		-0.16 (0.028)	-0.19 (0.015)	-0.25 (0.002)
∑FA ɯ6	-0.37 (< 0.001)			
MUFA		0.24 (0.003)	0.21 (0.010)	
$TC°_E$	0.81 (< 0.001)	0.84 (< 0.001)	0.82 (< 0.001)	0.81 (< 0.001)

Table 5: Validation of the different models used to explain the variability of PCA factor 1 and aggregated indexes (CDF_{I1}, CDF_{I2}, CDF_{I3} and CDF_{I4}). N_v: number of experiments from the validation set; R^2_{Val}: coefficient of determination of the validation set; RMSEP: root mean square error of prediction; Bias: model bias; RER: range error ratio.

	N_v	R^2_{Val}	RMSEP	Bias	RER
PCA factor 1	32	**0.747**	0.84	-0.07	7.1
CDF_{I1}	32	**0.728**	0.09	0.00	6.9
CDF_{I2}	33	**0.718**	0.11	-0.01	6.6
CDF_{I3}	33	0.668	0.12	0.00	6.2
CDF_{I4}	32	0.650	0.14	0.00	5.5