D1, but not D2, dopamine receptor regulates steroid levels during the final stages of pikeperch gametogenesis

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Short title: Dopamine receptors affect pikeperch reproduction

Abstract

In pikeperch, *Sander lucioperca*, aquaculture hormonal treatment is usually applied to synchronize ovulation. However, the effect of dopamine (**DA**) receptor antagonists, in particular those blocking the D1 DA receptors, remains unknown. Thus, the aim of the present study was to investigate and compare the effects of D1 and D2 DA receptor antagonists on the sex-steroid production and reproductive performance of the species. Two experiments were performed during which mature pikeperch

females were injected with different molecules: NaCl 0.9 % (negative control) or hCG 500 IU/kg (positive control) in both experiments, metoclopramide (a D2 receptor antagonist; 4 mg/kg or 20 mg/kg) or SCH23390 (a D1 receptor antagonist; 0.8 mg/kg or 4 mg/kg) alone (experiment one) or in combination with a salmon-GnRH analogue (**sGnRHa** at 25 µg/kg; experiment two). In experiment two, fish were also injected with sGnRHa (25 µg/kg) as positive control. Samplings of oocytes and blood were performed on the day of injection and after 24 h (both experiments), after 48 h (experiment two) and at the time of ovulation (both experiments). In non-ovulating fish, samplings were performed seven days (experiment one) or fourteen days (experiment two) after injection. In experiment two, various zootechnical parameters of fertilized eggs were recorded (survival, hatching and malformation rates). The two antagonists alone were ineffective in inducing the final stages and regulating sex-steroid (testosterone, 11 ketotestosterone, 17β estradiol and 17,20β -dihydroxy-4-pregnen-3-one) production. When administered with sGnRHa, both SCH23390 and metoclopramide induced the final stages. However, only SCH23390 stimulated testosterone (4 mg/kg) and 17β estradiol (0.8 mg/kg) production compared to sGnRHa alone. None of the treatments affected the survival, hatching or malformation rates. This is the first report suggesting that in pikeperch the D1, but not the D2, DA receptor antagonist would be involved in the testosterone and 17β estradiol production as a potentiator of the sGnRHa effect.

Keywords: pikeperch, ovulation, dopamine, antagonist, sex-steroid

Implications

This work provides information about the effects of some hormonal treatments on the induction of pikeperch reproduction in order to produce high quality offspring in a synchronous and predictable way for aquaculture. This work will allow optimizing pikeperch spawning and therefore improving production.

Introduction

Pikeperch, *Sander lucioperca*, is a species of interest for aquaculture diversification. It is a highly valuable economic and commercial species partly thanks to its flesh quality and recreational value (Kestemont *et al.*, 2015). Despite an increase in inland aquaculture production over the past decades, declining captures remain today the main supply to the increasing market demand (Food and Agriculture Organization of the United Nations, 2017). Consequently, great efforts are made to develop pikeperch aquaculture in order to respond to consumer demand. To achieve this objective, controlling the production of high quality offspring in a synchronous and predictable way, year after year, is a crucial step in the culture process (Zarski *et al*., 2015).

Controlled fish spawning is usually supported by the application of hormonal treatment, which allows synchronizing the ovulation process and optimizing egg collection while minimizing the handling and stress to the fish (Zohar and Mylonas, 2001). In pikeperch, various hormonal treatments have already been tested. The most widely used, human chorionic gonadotropin (**hCG**), was found to be highly effective in triggering ovulation (Zarski *et al.,* 2015). However, the spawning effectiveness observed in this species was highly variable, with frequently reported ovulation rates between 75% and 100% and embryonic survival rates between 50%

and 90% (Zarski *et al*., 2015). Besides, some authors demonstrated that application of hCG induces immune (Zohar and Mylonas, 2001) and stress responses (Falahatkar and Poursaeid, 2014), which could alter the subsequent reproductive operations (i.e. need to use higher doses of hCG or ineffectiveness of the treatment) (Zohar and Mylonas, 2001). For these reasons, analogues of the gonadotropin-releasing hormone (**GnRHa**) were applied as an alternative treatment in finfish reproduction (Mylonas *et al*., 2010; Zarski *et al*., 2015). GnRH acts directly at the pituitary level of the hypothalamic-pituitary-gonadal axis stimulating release of gonadotropin (LH), sex-steroid secretion and finally progression of the final stages of gametogenesis (oocyte meiotic maturation and ovulation). However, both basal and GnRH-stimulated LH secretion are under dopaminergic inhibition (Yaron and Levavi-Sivan, 2011). Thus, the combination of dopamine (**DA**) receptor antagonists with GnRHa is usually used (Mylonas *et al*., 2010) partly because antagonists would enhance the reproductive effectiveness of GnRHa therapy (Zarski *et al.*, 2015). Usually, the D2 DA receptor antagonists (metoclopramide **MCP**, domperidone or pimozide) have been used. The application of D2 DA receptor antagonists alone had positive effects on spawning in crucian carp, *Carassius carassius* (Cejko and Kucharczyk, 2015). By contrast, in Senegalese sole, *Solea senegalensis*, these treatments were found to be ineffective (Guzman *et al.,* 2011). In some perciforms, the effects of these antagonists on the reproductive effectiveness are highly contradictory and reduce the significance of the DA effect in this fish order (Dufour *et al.*, 2010; Zarski *et al.,* 2015). Thus, the effectiveness of DA antagonists seems to be species-specific and its use should be verified for each species separately.

In vertebrates, DA effects are mediated through the binding to two receptor families: D1 and D2 receptor families (Cardinaud *et al.,* 1997; Dufour *et al.,* 2010).

Interestingly, unlike D2 receptors, there is a lack of data concerning the existence and the role of D1 receptors in fish reproduction, although they were suggested to be involved in the control of GnRH release (Yu and Peter, 1992; Kapsimali *et al.,* 2000). In addition, *in vivo* studies showed that the DA/D1 receptor complex regulates the decrease in serum LH levels and aromatase B transcript levels in the hypothalamus of the goldfish, *Carassius auratus* (Popesku *et al.*, 2010 and 2012). These data suggest that species specificity of DA may be associated with different involvement of D1 and D2 receptors in the overall dopamine-related processes. The use of DA antagonists specific to D1 or D2 receptor family alone has never been tested in pikeperch. In addition, the combination of the D1 receptor antagonist with GnRHa has never been studied to date. This, together with the unclear role of DA in perciform reproduction (Dufour *et al.*, 2010; Zarski *et al.*, 2015), creates the need for a detailed investigation of this mechanism which could form the basis for pikeperch-specific hormonal treatment protocols.

Given the lack of data about the effects of dopamine receptor antagonists alone (D1 and D2) or in combination with GnRHa (D1) and the unknown involvement of receptors D1 in pikeperch reproduction, we aimed to investigate the *in vivo* physiological (sex-steroid) and zootechnical responses (gonado-somatic index **(GSI)**, progress of the oocyte meiotic maturation, ovulation rate, latency time, survival, hatching and malformation rates) to D1 and D2 DA receptor antagonists in pikeperch.

Material and methods

This study was split into two independent *in vivo* experiments. The first experiment was performed as a preliminary study to describe the effects of two DA antagonists specific to D1 or D2 receptor, SCH23390 (**SCH**) or MCP, respectively, applied alone

126 on pikeperch reproduction. The second experiment was dedicated to point out the
127 effects of the two antagonists combined with a GnRHa on pikeperch reproduction.
128 Both experiments were performed according to the European and France legislation
129 for fish welfare and approved by the institutional Ethics Committee (APAFIS3073-
130 2016022913149909). Fish were handled after anaesthesia by immersion in a bath
131 containing 150 mg/l of Ethyl 3-aminobenzoate (MS-222; Sigma-Aldrich, Lyon,
132 France).
133
134 *Broodstock management*
135 *Experiment one.* On 1 May 2015, 36 mature females (origin: production pond,
136 Fishery Nove Hrady Ltd, Czech Republic; age: 3 to 4 years old; mean body weight:
137 1.07 ± 0.06 kg) were transported to an outdoor recirculating system (La Bouzule,
138 Laneuvelotte, France). On 4 May 2015, all the fish were individually tagged (ID-100A
139 Microtransponder; Dorset Group BV, Aalten, The Netherlands). Fish were maintained
140 in sub-squared tanks (3000 l, 1 m deep), fed to satiation with forage fish and exposed
141 to natural photoperiod (Nancy, France) and temperature conditions (mean
142 temperature: 17.3 ± 2.7 °C) throughout the two weeks of experiment. Once a week,
143 pH, and ammonia and nitrite concentrations in the water were measured using a
144 WTW 340i pH meter and a CARY I spectrophotometer, respectively. All values
145 remained above 7.5 for pH and below 1 mg/l for nitrites and ammonia.
146
147 *Experiment two.* On 15 January 2016, 47 mature females reared in captivity (origin:
148 Czech Republic; age: 4 to 5 years old; mean body weight: 2.39 ± 0.48 kg) were used
149 in the facilities of the fish farm Asialor (Pierrevillers, France). Tagged fish (FDX-B
150 transponder; Biolog-ID, Bernay, France), were maintained in two 8000 l tanks in a

recirculating aquaculture system under 20 lx of light intensity at the water surface. Prior to the experiment, all fish were subjected to an increase in photoperiod and temperature to reach 14L:10D and 12.5 °C, respectively. Throughout the experiment, fish were kept under automatically controlled photoperiod (from 14L:10D to 15L:9D) and temperature (12.9 ± 0.14 °C) mimicking the environmental conditions prevailing during spawning. Dissolved oxygen (> 6 mg/l) and pH (7.8 ± 0.2) were monitored daily. Ammonia and nitrite concentrations in the water were measured using a colourimetric method once a week and remained below 0.5 mg/l.

Evaluation of oocyte maturation stages

For both experiments, each female was catheterized for evaluation of the oocyte maturation stages at different sampling times (as described in the sampling strategy) according to the classification by Zarski *et al.* (2012). Briefly, oocytes were sampled using a catheter (CH06; 1.2 mm internal and 2 mm external diameter) and placed in Serra's solution (ethanol/formalin/glacial acetic acid, 6:3:1 v/v/v). After mixing slowly oocytes in Serra's solution and waiting (about 5 min) until the cytoplasm of the oocyte will become clarified, the oocyte maturation stage was evaluated under binocular microscope, magnification × 4 (Motic® SFC-11 Series). In pikeperch, the final stages of maturation were divided into seven morphological stages, from stage I to stage VII (ovulation; Zarski *et al.*, 2012). This allowed following the progression of the oocyte meiotic maturation [hereinafter termed final oocyte maturation (**FOM)**] until ovulation.

Hormonal treatments

Experiment one. On 4 May 2015, oocyte maturation stages were evaluated for each female. Females between stages II and IV were randomized, sampled for blood (0 h)

and injected intraperitoneally with one of the following treatments: (1) negative control with saline solution, the vehicle of all molecules (NaCl 0.9%, n = 5); (2) positive control with hCG (500 IU/kg, n = 5; Sigma-Aldrich); a D2 DA receptor antagonist, (3) **MCP 4** (4 mg/kg, n = 6; Sigma-Aldrich) or (4) **MCP 20** (20 mg/kg, n = 7); a D1 DA receptor antagonist, (5) **SCH 0.8** (0.8 mg/kg, n = 6; Abcam, Paris, France) or (6) **SCH 4** (4 mg/kg, n = 7). The doses applied for hCG and MCP 20 were the most commonly used in controlled reproduction of percids (Zarski *et al*., 2015). For SCH, the doses applied were chosen to be consistent with the literature and to get a common dose between MCP and SCH.

Experiment two. On 15 January 2016, after determination of the oocyte maturation stage, all the females at stage I were randomized and injected intraperitoneally with one of the following treatments: (1) negative control with NaCl (0.9%, n = 9); (2) a first positive control with hCG (500 IU/kg, n = 7); (3) a second positive control with a salmon-GnRHa [25 µg/kg **(sGnRHa)**, n = 7; Pyr-His-Trp-Ser-Tyr-D-Arg-Trp-Leu-Pro-NHEt acetate salt; Syndel Laboratories Ltd, Nanaimo, Canada]; (4) MCP 4 or (5) MCP 20 in combination with sGnRHa (25 µg/kg, n = 6 and n = 5, respectively); (6) SCH 0.8 or (7) SCH 4 in combination with sGnRHa (25 µg/kg, n = 7 and n = 6, respectively). Before injection, 10 females were randomly sampled for blood (0 h). The doses applied in both positive control groups were the most commonly used in controlled reproduction of percids (Zarski *et al.*, 2015).

Sampling strategy

Experiment one. Blood and oocytes were sampled 0 and 24 h after injection and at the time of ovulation or 7 days after injection (Final sampling time, if the female did

not ovulate). Blood was sampled from the caudal vein and placed in tubes containing heparin (28 mg/ml; 100 kU Sigma-Aldrich). Plasma was obtained by centrifugation (15 min at 10 000 rpm) and stored at -80 °C until further steroid hormone analysis. At the final sampling time, the fish were killed by overexposure to anaesthetics MS-222 (240 mg/l) and the whole gonads were cut out and weighed.

The following parameters were recorded: the progression of FOM (= number of oocyte meiotic maturation stages between injection and the time of ovulation (or the end of the experiment); the GSI (= 100 × gonad weight / total fish weight); the ovulation rate for each treatment group (= 100 × number of ovulating females / total number of females); and the latency time (= time interval between injection and ovulation).

Experiment two. Blood and oocytes were sampled 0, 24 and 48 h after injection and at the time of ovulation or 14 days after injection (Final sampling time). Then, plasma was recovered and stored at -80 °C until further steroid hormone analysis. From 48 h, if the females did not reach stage VI, oocyte maturation stages were determined every 2 days. At stage VI, the genital papilla was sewn (as described by Zarski *et al.*, 2015) in order to prevent spontaneous releasing of eggs into the tank. From stage VI, ovulation control was performed every 6 h by gentle massage of the abdomen. At the time of ovulation, eggs were collected in dry plastic containers, weighed and then kept tightly covered at 11 °C for no longer than 30 min until fertilization.

For in vitro fertilization, sperm was collected from 30 males of pikeperch (origin: Czech Republic; age: 4 to 5 years old; mean body weight: 2.39 ± 0.36 kg). For each spawn, sperm from three males, injected with hCG (250 IU/kg) at 0 h, was collected in a dry syringe. For each fertilization procedure, only freshly collected sperm (15 min

maximum prior to fertilization), with a motility rate above 80% evaluated under a light microscope (magnification × 400; Motic® B3 Series) (Cejko *et al*., 2010) was used. For each female, three egg samples (approximately 50-100 eggs each) were placed in three glass Petri dishes containing 5 ml of hatchery water. Simultaneously, 50 µl of pooled sperm was added in each dish. After vigorous agitation for 15 s*,* each dish was incubated in plastic cups containing 500 ml of water at 12 °C.

The progression of FOM, the ovulation rate and the latency time were recorded. The GSI was not calculated due to the necessity to keep the fish alive. Additional zootechnical parameters were recorded: the embryo survival rate at 72 h post fertilization (= 100 × number of viable eggs / total number of eggs), the hatching rate (= 100 × number of larvae / total number of viable eggs) and the malformation rate [= 100 × number of larvae showing malformations (lordosis, cardiac oedema, kyphosis, fragmentation of oil droplet, yolk sac oedema, spinal curvature, scoliosis, C-shaped larvae) / total number of larvae]*.*

Levels of sex steroid hormones in blood plasma

For both experiments, sex-steroid hormones [17β estradiol (**E_2**), testosterone (**T**), 11 ketotestosterone (**11KT**) and 17,20β-dihydroxy-4-pregnen-3-one (**DHP**)] were measured in plasma using commercially available competitive ELISA kits. The kits for E_2 (KAP0621) and T (KAPD1559) were obtained from Diasource (Louvain-La-Neuve, Belgium), the kit for 11KT (582751) from Cayman Chemical (Ann Arbor, USA) and the kit for DHP (MBS2602842) from MyBiosource (San Diego, USA). The sensitivity limit, and the intra- and interassay CV were respectively 0.005 ng/ml (range: 0 - 0.935 ng/ml), < 4% and < 5% for E_2; 0.083 ng/ml (range: 0 - 16 ng/ml), < 10% and <

9% for T; 1.3 pg/ml (range: 0.78 - 100 pg/ml), < 9% and < 13% for 11KT; and 0.06 ng/ml (range: 0.312 - 20 ng/ml), < 9% and < 13% for DHP.

Statistical analysis

Statistical analyses were performed using the free software R version 3.3.1. For all dependent variables, homogeneity of variances was tested using Levene test (leveneTest, package 'car', Fox and Weisberg, 2011). For sex-steroids, GSI, latency time, survival rate, hatching rate and malformation rate, data were analysed by a linear mixed model (lmer, package 'lme4', Bates *et al.*, 2015) with hormonal treatment and sampling time as fixed effects, and either the fish and the maturation stage at P0 (experiment one) or only the fish (experiment two) as random effects: model=lmer(*Y*~treatment*sampling_time+(1|fish)+(1|maturation_stage) with *Y*: dependent variable. For model validation, residuals were tested for homogeneity and normality using residual *vs* fitted value and sample *vs* theoretical quantile (Q-Q) plots, respectively (plotresid, package 'RVAideMemoire', Hervé, 2016). If necessary, data were log transformed, root square transformed or arcsin root square transformed (only for data expressed in percentage). When the model was validated, an anova table was performed to calculate F-tests (Anova, package 'car', Fox and Weisberg, 2011) followed by a Least-squares means (predicted marginal means) multiple comparison between treatments, sampling times and/or their interaction as post-hoc test (lsmeans, package 'lsmeans', Lenth, 2016). When data, even transformed, did not meet the assumptions for the linear mixed model, we used the aligned rank transformation for nonparametric factorial analysis (aligned.rank.transform, package 'ART', Villacorta, 2015) followed by a pairwise comparison using Dunn test (posthoc.kruskal.dunn.test, package 'PMCMR', Pohlert,

2016). For the ovulation rate, data were analysed with a chi-square test (chisq.test, package 'MASS', Venables and Ripley, 2002). Data are expressed as mean ± SEM. The level of significance used in all tests *was* $P < 0.05$.

Results

Experiment one

Effect of dopamine receptor antagonists alone on reproductive performance. In fish treated with hCG, 100% of ovulation was recorded associated with higher GSI and progression of FOM (3 - 5 stages) than in the other groups. In the latter, similar GSI and progression of FOM (0 - 3) were noted. Consequently, MCP and SCH failed to induce ovulation and did not trigger a significant progression of FOM (Table 1).

Effect of dopamine receptor antagonists alone on plasma steroid concentrations.

Plasma T and 11KT levels varied significantly as a function of the interaction between hormonal treatments and sampling times ($P < 0.05$; Figure 1). Only females injected with hCG showed a significant decrease in T and 11KT concentrations between 24 h and the final sampling time ($P < 0.001$). However, no significant difference in plasma androgen levels was observed between the groups at each sampling time.

Only a time effect was monitored for E_2 with a drop observed at the final sampling time compared to 0 h and 24 h ($P < 0.01$; Figure 2a).

No significant difference related to hormonal treatments and sampling times was obtained for plasma DHP levels (Figure 2b).

For all steroid levels, high variability was observed.

Experiment two

Effect of dopamine receptor antagonists in combination with salmon-GnRH analogue on reproductive performance. Only the negative control group did not promote the progression of FOM and ovulation. All the other treatments triggered 100% of ovulation. The latency time and zootechnical parameters (survival rate, hatching rate, malformation rate) were similar in all the groups (Table 2).

Effect of dopamine receptor antagonists in combination with salmon-GnRH analogue on plasma steroid concentrations. Plasma T concentrations showed significant differences related to the interaction between hormonal treatments and sampling times ($P < 0.001$; Figure 3a). All hormonal treatments induced a marked increase in T compared to the negative control at 24 h ($P < 0.05$). At 48 h, the SCH 0.8, SCH 4, MCP 4 and hCG treatments were still effective in maintaining higher T levels compared to the negative control ($P < 0.05$). At the final sampling time, all hormonal treatments reduced the T concentrations ($P < 0.05$). The comparison between sGnRHa alone and the combination of DA receptor antagonists with sGnRHa showed that T levels increased with SCH 4 at 48 h ($P < 0.01$).

Concerning 11KT, significant differences were observed related to the interaction between hormonal treatments and sampling times ($P < 0.001$; Figure 3b). All hormonal treatments induced an increase in 11KT levels at 24 h and 48 h ($P < 0.001$), while at the final sampling time they were significantly abolished but only after SCH and MCP 20 treatments ($P < 0.001$). However, for each sampling time, the DA

receptor antagonist failed to change the 11KT level compared to the sGnRHa treatment alone.

For E_2, concentrations varied significantly as a function of the interaction between hormonal treatments and sampling times ($P < 0.001$; Figure 4a). All hormonal treatments induced an increase in E_2 levels at 24 h and 48 h ($P < 0.05$) except for MCP 4 and SCH 4 at 24 h, and sGnRHa and MCP 4 at 48 h. At the final sampling time, all treatments decreased E_2 levels ($P < 0.01$). Here also, there was not any significant difference between sGnRHa alone and the combination of DA receptor antagonists with sGnRHa except with SCH 0.8 at 48 h which increased E_2 levels ($P < 0.01$).

No significant difference related to hormonal treatments and sampling times was obtained for plasma DHP levels (Figure 4b).

Mild variability was observed for all steroid levels.

Discussion

This study investigated, for the first time, the potential regulatory effects of dopamine receptor antagonists alone on reproductive mechanisms in pikeperch. Our results first confirmed the high effectiveness of hCG in inducing the progression of FOM and ovulation in this species (Zarski *et al.,* 2015). During FOM, oocytes undergo a phenomenon of hydration inducing, in turn, an increase in follicle weight (Mañanós *et al.,* 2008). After hCG treatment, the rise in GSI may thus stem from the hydration process even if the latter remains to be demonstrated in pikeperch. Contrary to this positive control, no antagonist treatments were found to induce either GSI increase or ovulation. The latter finding is in accordance with previous reports on the Senegalese sole (Guzman *et al.*, 2011) and the common tench, *Tinca tinca*

(Podhorec *et al.,* 2016). In contrast, Cejko and Kucharczyk (2015) showed that the injection of metoclopramide induces ovulation in the crucian carp. This inter-species difference may be due to variable potency of the DA inhibition as already suggested (Mañanós *et al.*, 2008; Zarski *et al.*, 2015). Consequently, we hypothesize that DA receptor antagonists applied alone would be ineffective in inducing ovulation in pikeperch because of a weakness in the DA inhibition during the spontaneous progression of FOM. Alternatively, we might speculate that the doses chosen were not sufficient to generate a GnRH and/or LH endogenous surge. However, when combined with sGnRHa, these DA receptor antagonists do not prevent sGnRHa from triggering ovulation. They do not thus appear as inhibitors of the final stages of gametogenesis in pikeperch. Conversely, DA receptor antagonists would not by themselves allow the reproductive performance to be improved or impaired.

The DA receptor antagonists alone did not induce significant changes in the sex-steroid levels. These results are in accordance with prior studies in striped bass, *Morone saxatilis* (King *et al.*, 1994), and in Senegalese sole (Guzman *et al.*, 2011) in which application of the D2 DA receptor antagonist did not modify the T and E_2 levels in plasma. The attempt to block the D1 receptor family did not succeed in altering those hormonal levels as well. The ineffectiveness of DA receptor antagonists alone in changing the sex-steroid secretion is consistent with the absence of effect on the ovulation and progression in oocyte maturation. Additionally, this would indicate absence of or weakness in the dopaminergic inhibition by the application of D1 or D2 receptor antagonist at the tested doses during the non-hormonally manipulated oocyte maturation process in pikeperch.

Interestingly, we observed that the hCG treatment was unequally effective in inducing changes in the sex-steroid levels. These results are quite surprising considering that this molecule is a common substitute for natural (endogenous) fish LH, which induces fish spawning by direct action on gonads and sex-steroid levels (Mylonas *et al.*, 2010). Considering our results from experiment one, we noticed that at the same sampling time (e.g. 24 h), fish exhibited different stages of oocyte maturation, contrary to the synchronous stages in experiment two. This high inter-individual variability may have induced a larger range in steroid concentrations among the fish, which could potentially explain statistically irrelevant endocrine response following hCG treatment. However, other differences between the two populations (e.g. environmental conditions, stress status, final number of individuals) may also explain this difference in sensitivity to hCG. In contrast, our experimental setup, that is the presence of two experiments, the number of modalities, the common number of fish per group (King *et al.*, 1994; Barry *et al.*, 1995; Guzman *et al*., 2011), the number of sampling times and the *P*-values observed lead us to think that we have satisfactory conditions to have a sufficient statistical power for observing potential effects.

The sGnRHa treatment stimulated the production of sex-steroids confirming the widely observed activation of the gonadotropic axis in finfishes after such a treatment (Yaron and Levavi-Sivan, 2011). Interestingly, T and E_2 production was stimulated when sGnRHa was complemented with SCH depending on the dose. These results lead us to hypothesize that SCH would boost the sGnRHa effect on T and E_2 secretion. To our knowledge, this is the first report of plasma sex-steroid change after exposure to D1 receptor family antagonist in fish. Either this antagonist would directly block the D1 receptors at the gonad level as shown in rats (Venegas-Meneses *et al.*, 2015), which in turn would stimulate the ovarian steroidogenesis. Or, the blockage of

those receptors in the brain would disrupt the aromatase activity and the further metabolism of T and E_2 (Marsh *et al*., 2006; Popesku *et al*., 2012). Also, the blockage of D1 receptors could induce a surge in blood LH (Popesku *et al.*, 2010) which would stimulate the sex-steroid production. Finally, we cannot rule out some indirect effects of SCH through other metabolic factors. Further studies would be needed to check presence of those receptors in the ovary and to test these regulations by investigating the aromatase expression and activity in brain and oocytes as well as LH in the blood plasma.

Plasma DHP concentrations remained basal and stable over time in all the treatments. In many teleosts, the steroid DHP is the maturation-inducing steroid (**MIS**) (Nagahama and Yamashita, 2008). Progression of FOM was found to be linked to a significant increase in DHP levels in walleye, *Sander vitreus* (Barry *et al.,* 1995), and in striped bass (Mylonas *et al.*, 1997). However, in our study, even after application of hCG or sGnRHa with which all the fish ovulated, no peak of DHP concentration was observed. Surprisingly, these results are not consistent with GnRH commonly known effects on the gonadotrope axis (Yaron and Levavi-Sivan, 2011) and with a prior study in which hCG induced an increase in plasma DHP levels during the progression of FOM in white perch, *Morone americana*, and white bass, *Morone chrysops* (King *et al.*, 1995). Several hypotheses may explain these findings. First, as reported in walleye, the DHP could be rapidly removed from the plasma after its conjugation to a non-immunodetectable molecule such as 17,20-P-sulphate or 17,20-P-glucuronate (Scott and Canario, 1992). Second, in a closely related species, the Eurasian perch, *Perca fluviatilis*, Migaud *et al.* (2003) found very low DHP levels in plasma and suggested that these levels may be higher in the early morning than at

other times. Due to experimental conditions, our sampling times were performed at the beginning of the afternoon, so we may have missed the peak of detectable DHP in the blood plasma. Third, a close hormone, the 17α,20β, 21-trihydroxy-4-pregnen-3-one (**20β-S** or 17,20,21-P), identified as MIS in other perciforms (Nagahama and Yamashita, 2008) could also play this role in pikeperch. That would explain the mild level of DHP in this species, even after injection with hCG or sGnRHa. Indeed, although Barry *et al.* (1995) demonstrated that 20β-S was not detectable in walleye, others studied in an other perciform, the European sea bass, *Dicentrarchus labrax,* suggested that DHP and 20β-S could be both considered as MIS (Sorbera *et al.,* 1999; Asturiano *et al*., 2000). The DHP would be involved in the initiation of the maturation, just before our first sampling time, explaining the absence of detection in our study, and the 20β-S just before the ovulation. This indicates that both methodical and physiological studies on MIS in pikeperch should be reconsidered in the future. In any case, by considering DHP as the primary MIS in pikeperch, the lack of DA receptor antagonist effects (positive or negative) on FOM progression would support the absence of DHP regulation by these treatments.

In conclusion, this study, in the current experimental conditions, showed the ineffectiveness of DA receptor antagonist treatments alone, whatever the receptor family (D1 or D2), in inducing sex-steroid changes, FOM and ovulation in pikeperch. Combined with sGnRHa, these DA receptor antagonists did not prevent sGnRHa from triggering ovulation. However, in this combination with sGnRHa, SCH but not MCP proved efficiency to increase sGnRHa-stimulated steroid levels. Thus, only SCH would be involved in the regulation of sex-steroids indicating a putative potentiator effect of sGnRHa through D1 DA receptor blockage. The use of *in vitro*

448 biological tests of organ culture (e.g. brain, pituitary and ovary) could be useful in the
449 future to pinpoint these endocrine mechanisms.
450
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Table 1 *Effect of hormonal treatments on reproductive performance[1] in pikeperch females.*

	Treatments												
	Controls[2]				Antagonist groups[3]								
Parameters	NaCl	SEM	hCG	SEM	MCP 4	SEM	MCP 20	SEM	SCH 0.8	SEM	SCH 4	SEM	*P*-value
GSI (%)	13.5^{A}	0.3	19.5^{B}	1.3	11.8^{A}	0.8	11.6^{A}	1.4	11.5^{A}	0.8	10.4^{A}	0.4	< 0.001
FOM prog	0 - 1		3 - 5		0 - 1		0 – 3		0 - 1		0 - 1		
OR (%)	0^{A}		100^{B}		0^{A}		17^{A}		0^{A}		0^{A}		< 0.001
LT (h)			53.4	6.2			47.1						

GSI = gonado-somatic index; FOM prog = final oocyte maturation progression; OR = ovulation rate; LT = latency time (time between injection and ovulation); NaCl = saline solution; hCG = human chorionic gonadotropin; MCP 4 or MCP 20 = metoclopramide at 4 mg/kg or 20 mg/kg; SCH 0.8 or SCH 4 = SCH23390 at 0.8 mg/kg or 4 mg/kg

[1] Reproductive performance parameters are GSI, FOM prog, OR and LT

[2] Controls include negative control (NaCl) and positive control (hCG)

[3] Antagonist groups are treatments with one dopamine receptor antagonist (SCH or MCP)

[A,B] Means within the same row with different superscripts differ significantly at $P < 0.001$.

585 **Table 2** *Effect of hormonal treatments on reproductive performance[1] in pikeperch females.*

	Treatments													
	Controls[2]					Antagonist groups with GnRH[3]								
Parameters	NaCl	hCG	SEM	sGnRHa	SEM	MCP 4	SEM	MCP 20	SEM	SCH 0.8	SEM	SCH 4	SEM	*P*-value
FOM prog	0 - 1	6		6		6		6		6		6		
OR (%)	0^{A}	100^{B}		100^{B}		100^{B}		100^{B}		100^{B}		100^{B}		< 0.001
LT (h)		129.7	5.4	136.9	20.0	168.5	32.6	208.3	17.0	187.8	27.1	159.7	37.6	> 0.05
SR (%)		46.8	8.1	15.0	10.7	28.3	7.7	44.0	15.2	41.5	10.2	24.7	18.0	> 0.05
HR (%)		32.0	8.3	11.8	10.5	21.0	7.0	37.0	13.4	36.5	12.8	9.0	5.5	> 0.05
MR (%)		22.6	4.1	40.7	26.9	34.5	15.5	14.3	3.2	35.5	3.9	38.5	28.5	> 0.05

586 FOM prog = final oocyte maturation progression; OR = ovulation rate; LT = latency time (time between injection and ovulation); SR = survival rate; HR =
587 hatching rate; MR = malformation rate; NaCl = saline solution; hCG = human chorionic gonadotropin; sGnRHa = analogue of gonadotropin releasing hormone;
588 MCP 4 or MCP 20 = metoclopramide at 4 mg/kg or 20 mg/kg; SCH 0.8 or SCH 4 = SCH23390 at 0.8 mg/kg or 4 mg/kg
589 [1] Reproductive performance parameters are FOM prog, OR, LT, SR, HR and MR
590 [2] Controls include negative control (NaCl) and positive controls (hCG and sGnRHa)
591 [3] Antagonist groups with sGnRHa are treatments combining GnRH with one dopamine receptor antagonist (SCH or MCP)
592 [A,B] Means within the same row with different superscripts differ significantly at $P < 0.001$.
593
594

595 **Figure captions**

596

597 **Figure 1 Effect of hormonal treatments on plasmatic concentrations of (a)**
598 **testosterone (T) and (b) 11 ketotestosterone (11KT) in pikeperch**. Fish were
599 injected with NaCl (0.9%, n = 5), hCG (500 IU/kg, n = 5), MCP 4 or MCP 20
600 [metoclopramide at 4 mg/kg (n = 6) or 20 mg/kg (n = 7), respectively] and SCH 0.8 or
601 SCH 4 [(SCH23390 at 0.8 mg/kg (n = 6) or 4 mg/kg (n = 7), respectively]. Blood was
602 sampled at three sampling times: 0 and 24 h after injection and at the time of
603 ovulation or 7 days after injection (if ovulation did not occur; Final). Values are means
604 ± SEM. Different lowercase letters indicate significant differences for the interaction
605 between hormonal treatments and sampling times ($P < 0.05$).

606

607 **Figure 2 Effect of hormonal treatments on plasmatic concentrations of (a) 17β**
608 **estradiol (E_2) and (b) 17,20β-dihydroxy-4-pregnen-3-one (DHP) in pikeperch**.
609 Fish were injected with NaCl (0.9%, n = 5), hCG (500 IU/kg, n= 5), MCP 4 or MCP 20
610 [metoclopramide at 4 mg/kg (n = 6) or 20 mg/kg (n = 7), respectively] and SCH 0.8 or
611 SCH 4 [(SCH23390 at 0.8 mg/kg (n = 6) or 4 mg/kg (n = 7), respectively]. Blood was
612 sampled at three sampling times: 0 h and 24 h after injection and at the time of
613 ovulation or 7 days after injection (if ovulation did not occur; Final). Values are means
614 ± SEM. Different lowercase letters indicate significant differences between sampling
615 times ($P < 0.05$).

616

617 **Figure 3 Effect of hormonal treatments on plasmatic concentrations of (a)**
618 **testosterone (T) and (b) 11 ketotestosterone (11KT) in pikeperch**. Fish were
619 injected with NaCl (0.9%, n = 9), hCG (500 IU/kg, n = 7), sGnRHa (25 µg/kg, n = 7),

sGnRHa (25 µg/kg) in combination with MCP 4 or MCP 20 [metoclopramide at 4 mg/kg (n = 6) or 20 mg/kg (n = 5), respectively] or with SCH 0.8 or SCH 4 [(SCH23390 at 0.8 mg/kg (n = 7) or 4 mg/kg (n = 6), respectively]. Blood was sampled at four sampling times: 0, 24 and 48 h after injection and at the time of ovulation or 14 days after injection (if ovulation did not occur; Final). Values are means ± SEM. Different capital letters indicate significant differences between hormonal treatments for the same sampling time ($P < 0.05$). Different lowercase letters indicate significant differences between sampling times for the same hormonal treatment ($P < 0.05$).

Figure 4 Effect of hormonal treatments on plasmatic concentrations of (a) 17β estradiol (E_2) and (b) 17,20β-dihydroxy-4-pregnen-3-one (DHP) in pikeperch. Fish were injected with NaCl (0.9%, n = 9), hCG (500 IU/kg, n = 7), sGnRHa (25 µg/kg, n = 7), sGnRHa (25 µg/kg) in combination with MCP 4 or MCP 20 [metoclopramide at 4 mg/kg (n = 6) or 20 mg/kg (n = 5), respectively] or with SCH 0.8 or SCH 4 [(SCH23390 at 0.8 mg/kg (n = 7) or 4 mg/kg (n = 6), respectively]. Blood was sampled at four sampling times: 0, 24 and 48 h after injection and at the time of ovulation or 14 days after injection (if ovulation did not occur; Final). Values are means ± SEM. Different capital letters indicate significant differences between hormonal treatments for the same sampling time ($P < 0.05$). Different lowercase letters indicate significant differences between sampling times for the same hormonal treatment ($P < 0.05$)