



**HAL**  
open science

## Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows

Pierre-Alexis Billa, Yannick Faulconnier, Torben Larsen, Christine Leroux,  
José Pires

### ► To cite this version:

Pierre-Alexis Billa, Yannick Faulconnier, Torben Larsen, Christine Leroux, José Pires. Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. *Journal of Dairy Science*, 2020, 103 (4), pp.3133-3146. 10.3168/jds.2019-17466 . hal-02624734

**HAL Id: hal-02624734**

**<https://hal.inrae.fr/hal-02624734>**

Submitted on 26 May 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NoDerivatives 4.0 International License

1 **Interpretative Summary**, *Billa et al.*, page XX. The evaluation of nutritional deficits in dairy  
2 cows may be performed by measuring the concentrations of plasma metabolites, which  
3 require invasive blood sampling. The use of milk as source of noninvasive indicators would  
4 facilitate monitoring of animals. Concentrations of minor milk constituents were modified  
5 during partial restriction, suggesting their potential use as biomarkers of nutritional status.

6

7

8 **RUNNING HEAD: MILK METABOLITES DURING UNDERNUTRITION**

9

10

11 **Milk metabolites as noninvasive indicators of nutritional status of midlactation Holstein**  
12 **and Montbéliarde cows**

13

14 P.A. Billa\*, Y. Faulconnier\*, T. Larsen†, C. Leroux\*, J.A.A. Pires\*

15

16 \* INRA, Université Clermont Auvergne, VetAgro Sup, UMR Herbivores, F-63122 Saint-  
17 Genès-Champanelle, France

18 † Department of Animal Science, Aarhus University, DK-8830 Tjele, Denmark

19

20

21

22

23

24

25

26

## ABSTRACT

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

The objective was to investigate the effects of feed restriction on concentrations of selected milk metabolites in midlactation Holstein and Montbéliarde cows, and explore their correlations with energy balance and classic plasma and milk indicators of nutritional status. Eight Holstein and 10 Montbéliarde cows ( $165 \pm 21$  DIM) underwent 6 d of feed restriction during which feed allowance was reduced to meet 50% of their net energy for lactation ( $NE_L$ ) requirements. The experiment was divided in four periods: Control (CONT; d -3 to -1), restriction (REST; d 1 to 6), WEEK1 (d 7 to 13) and WEEK2 (d 14 to 18) after refeeding at ad libitum intake. Intake, milk production, energy balance and plasma metabolites were used to validate the feed restriction model. Concentrations of seven milk metabolites, i.e. BHB, glucose, glucose-6-phosphate, isocitrate, glutamate, uric acid and free amino groups were measured in morning milk samples, and fatty acids in pooled PM and AM samples. Feed restriction induced a negative energy balance ( $-42.5 \pm 4.4$  MJ/d), increased plasma non-esterified fatty acids and BHB, and decreased plasma glucose concentrations. Feed restriction increased milk glucose-6-phosphate and isocitrate (+38% and +39%, respectively) and decreased milk BHB, glucose, glutamate, uric acid and free amino group concentrations (-20%, -57%, -65%, -42% and -14%, respectively), compared to pre-restriction. Milk concentrations of medium chain fatty acids (e.g. sum of C10 to C15) decreased and those of long chain (e.g. 18:0, cis-9 18:1) increased during restriction. Breed differences were not detected for the majority of variables. All studied milk metabolites were significantly correlated with energy balance ( $r_s = 0.48, 0.63, -0.31, -0.45, 0.61$  for BHB, glucose, glucose-6-phosphate, isocitrate and glutamate, respectively). Milk glucose and glutamate were the most correlated with plasma metabolites and milk FA associated with lipomobilization. These

49 results suggest that milk metabolites may be used as noninvasive indicators of NEB and  
50 metabolic status of dairy cows.

51

52

## INTRODUCTION

53 Milk composition is modulated by a diversity of factors, including genetics, lactation  
54 stage, nutrition and health status, therefore, milk is an obvious source of biomarkers for the  
55 monitoring of dairy ruminants. Early lactation is characterized by a rapid increase in milk  
56 yield, mobilization of body protein and fat reserves, negative energy balance (**NEB**), and  
57 modifications in milk protein, fat and fatty acid composition. Complex homeorhetic and  
58 homeostatic adaptations are required to the direct limiting nutrients towards the mammary  
59 gland and support milk synthesis during early lactation (Bell and Bauman, 1997). The  
60 occurrence of metabolic disorders related to energy metabolism is typical during this period,  
61 and plasma concentrations of non-esterified fatty acids (**NEFA**) and BHB are classic  
62 indicators for herd troubleshooting (Oetzel, 2004). Experimental feed restriction models are  
63 used to induce nutrient deficits and NEB at different stages of lactation, assess the production  
64 and metabolic responses (Gross et al., 2011a; Bjerre-Harpøth et al., 2012; Friggens et al.,  
65 2016), and the effects of NEB on various biological functions of the dairy cow (Moyes et al.,  
66 2009; Abdelatty et al., 2017; Pires et al., 2019). Metabolic responses to feed restriction models  
67 are of greater amplitude during early lactation compared to later stages (Bjerre-Harpøth et al.,  
68 2012), but midlactation period is more convenient to run experimental protocols and less  
69 prone to confounding due to the dynamic nature of early lactation (Contreras et al., 2016).

70 The reliance on body reserve mobilization for milk synthesis during early lactation is  
71 largely driven by genetics (Friggens et al., 2013). Breed differences between Holstein  
72 (**HOLS**) and Montbéliarde (**MONT**) cows have been described in various production systems  
73 (Dillon et al., 2003; Pomiès et al., 2007; Pires et al., 2015). Holstein cows prioritize milk, fat,

74 protein and lactose secretion compared with MONT (Dillon et al., 2003; Pomiès et al., 2007;  
75 Pires et al., 2015), but experience greater BCS loss and metabolic deviations during early  
76 lactation (Dillon et al., 2003; Pires et al., 2015). Therefore, we hypothesized that breed effects  
77 might be used as a model to induce different production and metabolic responses to a  
78 nutritional challenge.

79 Milk is a source of novel indicators of nutritional status of dairy cows. Milk sampling  
80 is easy to perform in dairy operations, and can be automated for inline analyses. Certain  
81 molecules of intermediary metabolism are present in milk and may be indicators of  
82 physiological and nutritional state of dairy cows (Gross and Bruckmaier, 2019). For instance,  
83 milk is classically used to monitor ketosis by cow-side tests (Oetzel, 2004) and by automated  
84 inline BHB measurements. Milk glucose, glucose-6 phosphate, and acid uric acid  
85 concentrations are modulated by diet digestibility and correlated with DMI in midlactation  
86 cows (Larsen et al., 2016). Furthermore, milk glucose and glucose-6 phosphate concentrations  
87 vary according to DIM (Larsen and Moyes, 2015; Zachut et al., 2016; Ferris et al., 2018).  
88 Milk concentrations of these metabolites may reflect modifications of metabolic pathways in  
89 mammary epithelial cells, including glucose utilization for glycolysis and lactose synthesis  
90 (Chaiyabutr et al., 1981), glucose-6-phosphate and isocitrate to produce reducing potential  
91 (i.e.; NADPH) for de novo fatty acid synthesis (Garnsworthy et al., 2006; Chaiyabutr et al.,  
92 1981), and to counterbalance oxidative stress associated with FA oxidation (Zachut et al,  
93 2016). Milk uric acid originates in part from ruminal digestion of purine bases and has been  
94 suggested as an indicator of microbial protein synthesis (Larsen and Moyes, 2010). Milk  
95 glutamate and free amino acid content may reflect the availability and metabolism of amino  
96 acids.

97 The effects of NEB and breed on concentrations of minor milk metabolites are still  
98 insufficiently documented. We hypothesized that concentrations of selected metabolites in

99 milk are modified during periods of NEB, and may constitute novel indicators of energy  
100 balance and metabolic status of dairy cows. Thus, the objective was to determine the effects  
101 of NEB induced by partial feed restriction on milk concentrations of selected metabolites,  
102 assess potential differences between midlactation HOLS and MONT cows, and evaluate their  
103 relationships with classic indicators of metabolic status and lipomobilization.

104

105

## MATERIALS AND METHODS

106

### *Experimental Design, Animals, Diets and Housing*

108

109 All procedures were approved by the ethic committee on animal experimentation  
110 (APAFIS # 3737-2015043014541577v2). Twenty multiparous midlactation cows ( $165 \pm 21$   
111 DIM), 10 Holstein-Friesian and 10 Montbéliarde ( $1.5 \pm 0.29$  BCS, 0 to 5 scale) were used to  
112 study the effects of 6 d of feed-restriction to meet 50% of  $NE_L$  requirements on milk  
113 production, classic plasma and putative milk biomarkers of metabolic status. Two HOLS cows  
114 were excluded from the study, one due to clinical mastitis and one due to noncompliance with  
115 the restriction protocol. Phenotypic measurements were performed from d -3 to +18 relative  
116 of initiation of restriction, corresponding to the following periods: control (**CONT**; d -3 to -1),  
117 restriction (**REST**; d 1 to 6), week 1 (**WEEK1**; d 7 to 13), and week 2 (**WEEK2**; d 14 to 18;  
118 Figure 1). Five cows of each breed were randomly allocated to a group of 10 animals that  
119 initiated the experimental protocol one day apart. Mammary and liver biopsies were  
120 performed on d 0 and d 6 for complementary studies (Billa et al., 2019). The experiment was  
121 conducted at the INRA Herbipôle experimental farm of 'Marcenat' ( $45^{\circ}18'21''N$ ,  $2^{\circ}50'13''E$ ;  
122 1100 m of altitude; <https://doi.org/10.15454/1.5572318050509348E12>) during April and early  
May.

123 During CONT, WEEK1 and WEEK2 periods, all cows were allowed ad libitum intake  
124 of a TMR (Table 1). During REST period, feed allowance was reduced to meet 50% of  
125 individual  $NE_L$  requirements calculated from BW, feed intake and milk production and  
126 composition recorded before restriction (INRA, 2007). Cows had free access to water and  
127 were housed in a free stall barn equipped with automatic feed bunks that control individual  
128 access and weight feed intake (CRFI, Biocontrol, Rakkestad, Norway). Gates were  
129 programmed to divide individual daily feed allowance in 4 equal portions in 6-h periods.  
130 Cows ate  $3 \pm 0.9$  kg of hay (58.9% NDF, 31.9% ADF, 11.5% CP and 5.4 MJ /kg of DM)  
131 during the two d after refeeding to provide extra fiber and decrease the risk of ruminal  
132 acidosis due to the transition. The ration was analyzed for DM content to calculate individual  
133 DMI. Energy balance was estimated according to the INRA system (INRA, 2007), in which  
134  $NE_L$  is expressed as “unité fourragère lait” (UFL; 1 UFL = 7.12 MJ), as follows:

135  $NE$  intake (UFL) = UFL/kg DM  $\times$  DMI (kg) - E; with E corresponding to the  
136 “digestive interaction”, calculated as a function of the percentage concentrate in the diet (%  
137 Conc; DM-basis) and UFL intake (i.e., UFL/kg DM  $\times$  DMI (kg)), using the formula  $E =$   
138  $(0.00063 \times \%Conc^2) - (0.017 \times UFLintake) + (0.002 \times UFLintake^2)$ .

139  $NE$  production (UFL) = milk yield (kg)  $\times$   $[0.44 + (0.0055 \times (-40 + \text{fat content; g/kg}))$   
140  $+ (0.0033 \times (-31 + \text{protein content; g/kg}))]$ ;

141  $NE$  maintenance (UFL) =  $0.041 \times BW^{0.75} \times 1.1$ ;

142 Energy balance (UFL) =  $NE$  intake -  $NE$  maintenance -  $NE$  production.  
143

#### 144 **Sampling, Measurements and Chemical Analyses**

145 **Milk Sampling and Analysis.** Cows were milked twice daily at approximately 6:30  
146 and 16:00. Milk yield was recorded and milk composition was determined by mid-infrared  
147 spectroscopy (LIAL, Aurillac, France) in morning and evening milk samples. Weighted milk  
148 component means were computed according to PM/AM production and composition.

149 Morning milk samples were collected to determine metabolite concentrations on d -3,  
150 -2, -1, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, and 18 relative to initiation of feed restriction  
151 (Figure 1), before distribution of fresh TMR, and conserved at -20°C until analyses.  
152 Enzymatic-fluorometric methods were used to quantify milk content of BHB (Larsen and  
153 Nielsen, 2005), uric acid (Larsen and Moyes, 2010), isocitrate (Larsen, 2014), glucose and  
154 glucose-6-phosphate (Larsen, 2015), glutamate and free amino groups ( $\text{NH}_2$ ) (Larsen and  
155 Fernández, 2017). Morning and evening milk samples were collected on d -2, -1, 1, 2, 3, 4, 5,  
156 6 and 13 to determine milk fatty acid (FA) composition by gas chromatography, as previously  
157 described (Lerch et al., 2012). Briefly, samples were lyophilized and pooled according PM  
158 and AM production, to provide daily composite samples for each cow. Samples were  
159 methylated and injected into 7890A GC system CN10271102 Series gas chromatograph  
160 equipped with a flame ionization detector (Agilent technologies, Santa Clara, California,  
161 USA). Peaks were routinely identified through comparison of retention times with FA methyl  
162 ester standards. Peak integration was conducted using Chemstation software (Agilent  
163 technologies, Santa Clara, California, USA).

164 **Blood Sampling and Analyses.** Jugular blood samples were collected on d -3, -2, 1, 2,  
165 3, 4, 5, 6, 7, 8, 9, 11, 13 and 18 relative to initiation of feed restriction, after morning milking  
166 and before feed distribution. Blood samples were drawn into EDTA (1.95 mg/mL; Terumo  
167 Europe NV, Leuven, Belgium) and Li-heparin (135 USP U; Terumo Europe NV, Leuven,  
168 Belgium) tubes. Plasma was separated by centrifugation at 1,400×g for 15 min at 4°C and  
169 conserved at -20°C until analysis. Plasma (EDTA) glucose, BHB, urea and NEFA  
170 concentrations were quantified spectrophotometrically and insulin measured by RIA (Pires et  
171 al., 2019). Plasma (heparin) glutamine, glutamate and free amino groups ( $\text{NH}_2$ )  
172 concentrations were quantified by enzymatic-fluorometric methods (Larsen and Fernández,  
173 2017).



174

## 175 ***Statistical Analyses***

176 Statistical analyses were performed using SAS enterprise guide (Version 9.4; SAS  
177 Institute INC, Cary, NC). Daily data was analyzed as repeated measures by mixed models that  
178 included day, breed and their interaction as fixed effects, cow as random effect, and Kenward-  
179 Roger adjustment for calculation of degrees of freedom. The Schwarz's Bayesian criterion  
180 was used to compare the fitting of different variance-covariance structures, including spatial  
181 power, AR (1), ARH (1) and CS. The means for each variable within animal and period were  
182 calculated in order to compare periods (CONT, REST, WEEK1 and WEEK2) and explore  
183 breed by period interactions. The models included the fixed effects of period, breed and their  
184 interaction, and the random effect of cow. Significant time effects (i.e., day or period) and  
185 breed by time interactions were explored by the Fisher's protected least significant difference  
186 using the PDIFF and SLICE options of the LSMEANS statement. Residuals were checked for  
187 normality and homoscedasticity. Heterogeneous variance was tested whenever suggested by  
188 residual plots. Least squares means (LSM) and standard error of the mean (SEM) were  
189 estimated from untransformed values, whereas *P*-values may reflect statistical analysis of log-  
190 transformed data when transformation was necessary. Relationships among variables were  
191 explored by Spearman rank correlations. Linear regressions between energy balance and milk  
192 metabolite concentrations were analyzed using PROC REG procedure of SAS. The  
193 significance level was predefined as  $P \leq 0.05$  and trends toward significance at  $0.05 < P \leq$   
194 0.10.

195

196

## **RESULTS AND DISCUSSION**

197

### 198 ***Production Responses***

199 A significant period effect was observed for DMI, energy balance, milk and milk  
200 component yields (Table 2 and Supplemental Figure 1). Per design, energy balance became  
201 negative ( $-42.5 \pm 4.4$  MJ/d) and milk, fat and protein yields decreased during REST. Energy  
202 balance returned to pre-restriction values during WEEK1, and DMI, fat and protein yields  
203 during WEEK2. The effect of restriction on milk, fat and protein yields are in accordance with  
204 previous studies in midlactation Holstein cows (Gross et al., 2011a; Bjerre-Harpøth et al.,  
205 2012; Pires et al., 2016).

206 A significant breed effect was observed for DMI, milk, protein and lactose yields,  
207 which were greater in HOLS than in MONT cows (Table 2). Energy balance and fat yield  
208 tended to be greater in HOLS than in MONT. Nonetheless, milk yield differences between  
209 HOLS and MONT during CONT (+17% for HOLS) were less marked than reported in  
210 previous studies during mid lactation (+21%; Pomiès et al., 2007; Ferlay et al., 2010).  
211 Differences among studies could be explained by feeding systems, milking frequency, and the  
212 lactation stage.

213

#### 214 ***Plasma Metabolite and Insulin Concentrations***

215 Plasma metabolite and insulin concentrations are presented in Table 3 and  
216 Supplemental Figures 2 and 3. Significant time effects were observed for all plasma variables.  
217 Plasma NEFA, BHB and glutamate increased whereas plasma glucose, glutamine, urea,  $\text{NH}_2$   
218 and insulin concentrations decreased during the REST. Plasma NEFA, BHB, glucose and  $\text{NH}_2$   
219 returned to CONT values on WEEK1. Plasma glutamine, urea and  $\text{NH}_2$  returned to CONT  
220 values on WEEK2. Plasma glutamate was lower at the end of WEEK2 than before the  
221 challenge. Plasma glucose concentrations returned to CONT value during WEEK1 and  
222 became greater than CONT concentrations on WEEK2. Plasma insulin was significantly  
223 greater than CONT concentrations on WEEK1, but was lower than CONT on WEEK2.

224 The increase in plasma NEFA concentrations during REST reflects lipomobilisation  
225 (Chilliard et al., 2000a) and is in accordance with previous feed-restriction studies involving  
226 mid- and late lactation cows (Bjerre-Harpøth et al., 2012; Gross and Bruckmaier, 2011a; Pires  
227 et al., 2016). Nonetheless, the increase in plasma NEFA observed during REST in our  
228 experiment was smaller than observed in underfed early lactation cows (Bjerre-Harpøth et al.,  
229 2012; Pires et al., 2019). Plasma BHB concentrations increased during restriction (Table 3 and  
230 Supplemental Figure 2), but remained below the 1.2 mM threshold of subclinical ketosis  
231 (LeBlanc et al., 2005), which is in agreement with previous research in underfed midlactation  
232 cows (Moyes et al., 2009; Gross et al., 2011a; Bjerre-Harpøth et al., 2012). As plasma BHB  
233 originates in part from rumen butyrate (Miettinen and Huhtanen, 1996), its concentrations  
234 observed during REST probably reflect concomitant modifications of DMI and ruminal  
235 butyrate synthesis, and incomplete beta-oxidation of mobilized NEFA. Two cows had  
236 increased concentrations of BHB the morning after refeeding the TMR at ad libitum intake (d  
237 7; Supplemental Figure 3), which may result from increased DMI and a shift in ruminal  
238 butyrate production. Accordingly, insulin was greatest the day after refeeding at ad libitum  
239 intake (d 7, Supplemental Figure 2).

240 The decrease in plasma glutamine and  $\text{NH}_2$  concentrations observed during REST may  
241 reflect reduced DMI and amino acid absorption. Glutamine and glutamate are intermediates in  
242 many anabolic and catabolic pathways, including regulation of metabolic acidosis,  
243 lymphocyte proliferation, and casein synthesis (Lobley et al., 2001). Plasma glutamine,  
244 glutamate and total amino acid concentrations decrease during periods of NEB, such as early  
245 lactation (Meijer et al., 1995) and during experimental feed restriction in midlactation cows  
246 (Girard et al., 2019). Decreased supply of amino acids for intestine absorption may have  
247 contributed more to these changes than amino acid catabolism, because plasma urea  
248 concentration also decreased during REST, as previously suggested (Girard et al., 2019). The

249 increase in plasma glutamate concentration observed during REST (Table 3 and Supplemental  
250 Figure 3) may be due in part to spontaneous glutamine deamination.

251 Holstein cows had greater plasma glucose, glutamate and  $\text{NH}_2$ , concentrations than  
252 MONT (Supplemental Figures 2 and 3). The greater glucose concentrations observed in  
253 HOLS compared to MONT contrast with previous research during the first weeks of lactation  
254 in low input systems (Pires et al., 2015). These discrepancies may be explained by different  
255 lactation stage (early vs. midlactation), diet, and undernutrition model (early lactation  
256 spontaneous NEB vs. midlactation feed restriction). The greater BHB concentrations observed  
257 in HOLS during CONT (Table 3) probably reflect their greater DMI, ruminal butyrate and  
258 BHB synthesis compared with MONT, because all animals were in positive energy balance.

259

#### 260 ***Milk Metabolite Concentrations***

261 Significant time effects were observed for milk concentrations of all metabolites  
262 studied (Table 4, Figures 2 and 3). Milk BHB, glucose, glutamate, uric acid and  $\text{NH}_2$   
263 decreased, whereas glucose-6-phosphate and isocitrate concentrations increased during REST.  
264 Milk glutamate, isocitrate and  $\text{NH}_2$  concentrations returned to pre-restriction values on  
265 WEEK1. Milk BHB, glucose and uric acid returned to pre-restriction concentrations on  
266 WEEK2. Milk glutamate was greater whereas glucose-6-phosphate and isocitrate  
267 concentrations were lower on WEEK2 compared to CONT.

268 The decrease of milk uric acid concentrations observed during REST is in agreement  
269 with results obtained when cows were offered low energy diets (Larsen et al., 2016). Milk  
270 BHB concentrations decreased during REST despite a small but significant increase in plasma  
271 BHB, which suggests that BHB was preferentially metabolized in the mammary gland. In  
272 contrast, BHB concentrations increase both in plasma and milk during early lactation, and  
273 both matrices are used to diagnose subclinical ketosis (Nielsen et al., 2003; Oetzel, 2004).

274 Increased milk glucose-6-phosphate and isocitrate, and decreased milk glucose concentrations  
275 during REST are in agreement with previous observations in starved goats (Chaiyabutr et al.,  
276 1981) and in early lactation cows (Larsen and Moyes, 2015; Zachut et al., 2016). Glucose-6-  
277 phosphate and isocitrate are precursors for NADPH synthesis via the pentose phosphate and  
278 isocitrate dehydrogenase pathways, respectively. Increased milk glucose-6-phosphate and  
279 isocitrate content during REST may reflect a shift in these pathways in mammary epithelial  
280 cells, due to low plasma insulin, decreased de novo FA synthesis and synthesis of other milk  
281 components.

282 Significant breed effects were observed for milk uric acid and isocitrate concentrations  
283 when individual data were analyzed by period (Table 4). Nonetheless, only a trend was  
284 detected for milk isocitrate concentration when daily data were analyzed as repeated  
285 measures, probably because many time points measured after refeeding (WEEK1 and  
286 WEEK2) did not differ between HOLS and MONT (Figures 2 and 3). Breed by time  
287 interactions were not observed for any of the milk metabolites analyzed in this study.

288

### 289 ***Milk Fatty Acid Concentrations***

290 Milk FA concentrations are presented in Table 5 and Supplemental Figure 4.  
291 Significant time effects were observed for all FA and FA classes, except for total odd and  
292 branched chain fatty acids ( $\Sigma$  **OBCFA**). Milk FA concentrations returned to prechallenge  
293 values on d 13, except for OBCFA.

294 Concentrations of FA with 10 to 15 carbons ( $\Sigma$  **10:0 to 15:0**), 16:0 and  $\Sigma$  OBCFA with  
295 carbon chain shorter than 16 ( $\Sigma$  **OBCFA < C16**) decreased during REST, whereas  
296 concentration of 18:0 and *cis*-9 18:1 and of FA with carbon chain greater than 16 ( $\Sigma$  **> C16**)  
297 increased. These modifications in milk FA profile are in agreement with other feed restriction  
298 studies (Chilliard et al., 2000b; Gross et al., 2011b; Abdelatty et al., 2017). The decrease of  $\Sigma$

299 10:0 to 15:0 and 16:0 during REST reflect a diminution of de novo FA synthesis in mammary  
300 gland, due to a decreased availability of precursors (e.g., acetate and propionate) absorbed  
301 from rumen (Chilliard et al., 2000b; Gross et al., 2011b). The increase of 18:0, and *cis*-9 18:1  
302 and  $\Sigma > C16$  FA reflect body fat mobilization (Chilliard et al., 2000b; Gross et al., 2011b ;  
303 Pires et al., 2013).

304 The  $\Sigma$  10:0 to 15:0 and 16:0 decreased gradually until d 3 of restriction (Supplemental  
305 Figure 5), indicating a gradual downregulation of de novo FA synthesis. This downregulation  
306 would reduce NADPH requirements for mammary gland lipogenesis (Bell and Bauman,  
307 1997), and may explain the gradual increase in glucose-6-phosphate and isocitrate  
308 concentrations during REST, which became significant at 48 h of REST (Figure 2). This  
309 pattern may reflect concomitant effects of limiting plasma glucose availability and reduced  
310 glucose uptake by mammary gland during REST, downregulation of lactose synthesis  
311 (Chaiyabutr et al., 1981), and decreased NADPH requirements for de novo fatty acid  
312 synthesis (Chaiyabutr et al., 1981; Garnsworthy et al., 2006). Milk glucose-6-phosphate and  
313 isocitrate concentrations decreased on d 5 and 6 of REST, before refeeding at ad libitum  
314 intake. Epithelial cell homeostatic mechanisms may have reestablished an equilibrium  
315 between cytosolic concentrations of glucose 6-phosphate and isocitrate and the activity of  
316 metabolic pathways for which they are precursors (e.g., NADPH and lactose synthesis).  
317 Zachut et al. (2016) proposed that FA oxidation in mammary cells during periods of  
318 lipomobilization would increase the oxidative stress, requiring the upregulation of the pentose  
319 phosphate pathway to generate reducing potential to neutralize reactive oxygen species.  
320 Mammary gland gene expression shows a shift towards increased reliance on  $\beta$ -oxidation for  
321 energy and sparing of glucose in underfed early lactation cows (Pawłowski et al., 2019).  
322 Oxidative stress may have occurred in our study, inducing a gradual depletion of glucose-6-

323 phosphate and isocitrate on d 5 and 6 of restriction. Indicators of oxidative stress were not  
324 measured in the current study.

325 The decrease in  $\sum$  OBCFA < C16 observed during restriction (Table 5 and  
326 Supplemental Figure 4) may be explained by reduced ruminal synthesis and incorporation of  
327 absorbed FA into milk fat. The increase in milk  $\sum$  OBCFA C>16 content (data not shown)  
328 during REST suggests that these FA were mobilized from adipose tissue. The majority of milk  
329 OBCFA originates from digested rumen bacteria. Branched chain FA are synthesized by  
330 elongation of carbon chains originating from branched chain amino acids (e.g. valine, leucine  
331 and isoleucine), and part of the odd chain FA are synthesized by elongation of propionate and  
332 valerate. A smaller proportion is synthesized de novo from elongation of propionate  
333 (Vlaeminck et al., 2005, 2015).

334 A breed effect was observed for milk 18:0 content, which was greater in HOLS than  
335 MONT cows. No breed by time interaction was observed for milk FA concentrations.

336

### 337 ***Correlation and regression analyses***

338 Correlations are presented in Tables 6 and 7 and regressions between energy balance  
339 and milk metabolites are presented in Figure 4 and Supplemental Figure 5. All variables  
340 presented were significantly correlated with energy balance, except plasma urea concentration  
341 and milk total OBCFA content. Among plasma metabolites, NEFA and glucose had the  
342 greatest absolute correlation with energy balance ( $r_s = -0.72$  and  $r_s = 0.64$ , respectively; Table  
343 6). Among milk metabolites, milk glucose and glutamate had the greatest absolute  
344 correlations with energy balance ( $r_s > 0.60$ ) and plasma NEFA ( $r_s = -0.67$ , Table 6), which is a  
345 classic indicator of lipomobilization. Moreover, milk glucose was correlated with plasma  
346 glucose ( $r_s = 0.61$ ) which is also modulated by NEB (Meijer et al., 1995; Bjerre-Harpøth et  
347 al., 2012; Girard et al., 2019). Milk isocitrate and glucose-6-phosphate concentrations were

348 negatively correlated with energy balance ( $r_s = -0.45$  and  $-0.31$ , respectively; Table 6). Milk  
349 glucose and milk glutamate concentrations present the best regressions with energy balance  
350 ( $r^2 = 0.49$  and  $0.46$ , respectively;  $P < 0.001$ ; Figure 4). Whereas milk glucose-6-phosphate and  
351 milk isocitrate present a weak regression with energy balance ( $r^2 = 0.10$  and  $0.26$ ,  
352 respectively;  $P < 0.001$ ). These results suggest that milk glucose and glutamate concentrations  
353 may be good indicators of NEB and metabolic status of dairy cows.

354 Table 7 presents the milk FA correlations with selected variables. Milk FA composition  
355 is used as a biomarker of lipomobilization in this study, and was measured in pooled samples  
356 of PM/AM milkings, therefore it integrates modifications occurring during 24 h periods, as  
357 for the calculations of energy balance. Milk  $\Sigma > C16$ ,  $18:0$  and *cis*- $9\ 18:1$  were negatively  
358 correlated with energy balance ( $r_s = -0.54$  to  $-0.56$ ), whereas milk  $\Sigma\ 10:0$  to  $15:0$ ,  $16:0$  and  $\Sigma$   
359 OBCFA  $< C16$  were positively correlated with energy balance ( $r_s = 0.39$  to  $0.52$ ). These  
360 results were expected due to the known associations among NEB, lipomobilization and milk  
361 FA composition (Gross et al., 2011b; Pires et al., 2013). Accordingly, FA potentially  
362 synthesized de novo in mammary gland, such as  $\Sigma\ 10:0$  to  $15:0$  and OBCFA  $< C16$ , were  
363 positively correlated with plasma glucose, and negatively correlated with plasma NEFA and  
364 BHB. Opposite correlations were observed among these plasma metabolites and  $\Sigma > C16$ ,  
365 which are FA potentially mobilized from adipose. Moreover, milk glucose, glutamate and  
366 isocitrate concentrations were significantly correlated with all reported milk FA and FA  
367 classes ( $r_s > 0.45$  in absolute value), except for total OBCFA, further supporting potential of  
368 milk glucose, glutamate and isocitrate as biomarkers of NEB and metabolic status of dairy  
369 cows.

370

371

## CONCLUSIONS



372 The feed restriction model induced NEB and modified plasma metabolites and milk  
373 FA concentrations in midlactation cows as expected. The absence of marked breed differences  
374 for most variables may be explained by the short duration of the nutritional challenge (6 d)  
375 and lactation stage, as phenotypical differences exist between HOLS and MONT during early  
376 lactation. Milk concentrations of FA and selected metabolites were modulated by feed  
377 restriction in midlactation cows. Milk glucose and glutamate concentrations presented the  
378 strongest correlations with classic indicators of metabolic status and the best regressions with  
379 energy balance under this experimental model. Milk glucose and glutamate concentrations  
380 may constitute good noninvasive indicators of energy balance, lipomobilization and metabolic  
381 status. Further research is warranted in early lactation cows, because feed restriction models  
382 during established lactation lead to relatively smaller deviations of classic indicators of NEB  
383 and metabolic status compared to early lactation.

384

385

#### ACKNOWLEDGEMENTS

386 The authors thank the staff at Herbipôle INRA (UE1414, Marcenat, France) for animal  
387 care and sampling; A. Delavaud, S. Bes, D. Chadeyron, E. Tixier and M. Tourret (INRA,  
388 UMR1213, Saint-Genès-Champanelle, France) for sample collection and laboratory analyses;  
389 J. Clausen and C. Berthelsen (Aarhus University, Tjele, Denmark) for milk metabolite  
390 analyses. This research was partially funded by FEDER and Région Auvergne S3 project nr.  
391 23000794, and Compte d'affection Spécial au Développement Agricole et Rural (CASDAR;  
392 Paris, France) project nr. 00001908 (Biomarq'lait).

393

394

#### REFERENCES

395

- 396 Abdelatty, A.M., M.E. Iwaniuk, M. Garcia, K.M. Moyes, B.B. Teter, P. Delmonte, A.K.G.  
397 Kadegowda, M.A. Tony, F.F. Mohamad, and R.A. Erdman. 2017. Effect of short-term  
398 feed restriction on temporal changes in milk components and mammary lipogenic  
399 gene expression in mid-lactation Holstein dairy cows. *J. Dairy Sci.* 100:4000–4013.  
400 doi:10.3168/jds.2016-11130.
- 401 Bell, A.W., and D.E. Bauman. 1997. Adaptations of glucose metabolism during pregnancy  
402 and lactation. *J. Mammary Gland Biol. Neoplasia* 2:265–278.  
403 doi:10.1023/A:1026336505343.
- 404 Billa, P.A., Y. Faulconnier, T. Ye, M. Chervet, F. Le Provost, J.A.A. Pires, and C. Leroux.  
405 2019. Deep RNA-Seq reveals miRNome differences in mammary tissue of lactating  
406 Holstein and Montbéliarde cows. *BMC Genomics* 20:621. doi:10.1186/s12864-019-  
407 5987-4.
- 408 Bjerre-Harpøth, V., N.C. Friggens, V.M. Thorup, T. Larsen, B.M. Damgaard, K.L. Ingvarsen,  
409 and K.M. Moyes. 2012. Metabolic and production profiles of dairy cows in response  
410 to decreased nutrient density to increase physiological imbalance at different stages of  
411 lactation. *J. Dairy Sci.* 95:2362–2380. doi:10.3168/jds.2011-4419.
- 412 Chaiyabutr, N., A. Faulkner, and M. Peaker. 1981. Changes in the concentrations of the minor  
413 constituents of goat's milk during starvation and on refeeding of the lactating animal  
414 and their relationship to mammary gland metabolism. *Br. J. Nutr.* 45:149–157.  
415 doi:10.1079/BJN19810087.
- 416 Chilliard, Y., A. Ferlay, Y. Faulconnier, M. Bonnet, J. Rouel, and F. Bocquier. 2000a. Adipose  
417 tissue metabolism and its role in adaptations to undernutrition in ruminants. *Proc.*  
418 *Nutr. Soc.* 59:127–134. doi:10.1017/S002966510000015X.

- 419 Chilliard, Y., A. Ferlay, R.M. Mansbridge, and M. Doreau. 2000b. Ruminant milk fat  
420 plasticity: nutritional control of saturated, polyunsaturated, trans and conjugated fatty  
421 acids. *Ann. Zootech.* 49:181–205. doi:10.1051/animres:2000117.
- 422 Contreras, G.A., K. Thelen, S.E. Schmidt, C. Strieder-Barboza, C.L. Preseault, W. Raphael,  
423 M. Kiupel, J. Caron, and A.L. Lock. 2016. Adipose tissue remodeling in late-lactation  
424 dairy cows during feed-restriction-induced negative energy balance. *J. Dairy Sci.*  
425 99:10009–10021. doi:10.3168/jds.2016-11552.
- 426 Dillon, P., F. Buckley, P. O'Connor, D. Hegarty, and M. Rath. 2003. A comparison of different  
427 dairy cow breeds on a seasonal grass-based system of milk production: 1. Milk  
428 production, live weight, body condition score and DM intake. *Livest. Prod. Sci.*  
429 83:21–33. doi:10.1016/S0301-6226(03)00041-1.
- 430 Ferlay, A., B. Martin, S. Lerch, M. Gobert, P. Pradel, and Y. Chilliard. 2010. Effects of  
431 supplementation of maize silage diets with extruded linseed, vitamin E and plant  
432 extracts rich in polyphenols, and morning v. evening milking on milk fatty acid  
433 profiles in Holstein and Montbéliarde cows. *Animal.* 4:627–640.  
434 doi:10.1017/S1751731109991224.
- 435 Ferris, C.P., P.J. Purcell, A.W. Gordon, T. Larsen, and M. Vestergaard. 2018. Performance of  
436 Holstein and Swedish-Red × Jersey/Holstein crossbred dairy cows within low- and  
437 medium-concentrate grassland-based systems. *J. Dairy Sci.* 101:7258–7273.  
438 doi:10.3168/jds.2017-14107.
- 439 Friggens, N.C., L. Brun-Lafleur, P. Faverdin, D. Sauvant, and O. Martin. 2013. Advances in  
440 predicting nutrient partitioning in the dairy cow: recognizing the central role of

441 genotype and its expression through time. *Anim. Int. J. Anim. Biosci.* 7:89–101.  
442 doi:10.1017/S1751731111001820.

443 Friggens, N.C., C. Duvaux-Ponter, M.P. Etienne, T. Mary-Huard, and P. Schmidely. 2016.  
444 Characterizing individual differences in animal responses to a nutritional challenge:  
445 Toward improved robustness measures. *J. Dairy Sci.* 99:2704–2718.  
446 doi:http://dx.doi.org/10.3168/jds.2015-10162.

447 Garnsworthy, P.C., L.L. Masson, A.L. Lock, and T.T. Mottram. 2006. Variation of Milk  
448 Citrate with Stage of Lactation and De Novo Fatty Acid Synthesis in Dairy Cows. *J.*  
449 *Dairy Sci.* 89:1604–1612. doi:10.3168/jds.S0022-0302(06)72227-5.

450 Girard, C.L., N. Vanacker, V. Beaudet, M. Duplessis, and P. Lacasse. 2019. Glucose and  
451 insulin responses to an intravenous glucose tolerance test administered to feed-  
452 restricted dairy cows receiving folic acid and vitamin B12 supplements. *J. Dairy Sci.*  
453 102:6226–6234. doi:10.3168/jds.2019-16298.

454 Gross, J., H.A. van Dorland, R.M. Bruckmaier, and F.J. Schwarz. 2011a. Performance and  
455 metabolic profile of dairy cows during a lactational and deliberately induced negative  
456 energy balance with subsequent realimentation. *J. Dairy Sci.* 94:1820–1830.  
457 doi:10.3168/jds.2010-3707.

458 Gross, J., H.A. van Dorland, R.M. Bruckmaier, and F.J. Schwarz. 2011b. Milk fatty acid  
459 profile related to energy balance in dairy cows. *J. Dairy Res.* 78:479–488.  
460 doi:10.1017/S0022029911000550.

461 Gross, J.J., and R.M. Bruckmaier. 2019. Review: Metabolic challenges in lactating dairy cows  
462 and their assessment via established and novel indicators in milk. *animal* 13:s75–s81.  
463 doi:10.1017/S175173111800349X.

- 464 INRA. 2007. Alimentation des bovins, ovins et caprins: besoins des animaux, valeurs des  
465 aliments. Editions Quae Versailles, France.
- 466 Larsen, T. 2014. Fluorometric determination of free and total isocitrate in bovine milk. *J.*  
467 *Dairy Sci.* 97:7498–7504. doi:10.3168/jds.2014-8018.
- 468 Larsen, T. 2015. Fluorometric determination of free glucose and glucose 6-phosphate in cows'  
469 milk and other opaque matrices. *Food Chem.* 166:283–286.  
470 doi:10.1016/j.foodchem.2014.06.017.
- 471 Larsen, T., L. Alstrup, and M.R. Weisbjerg. 2016. Minor milk constituents are affected by  
472 protein concentration and forage digestibility in the feed ration. *J. Dairy Res.* 83:12–  
473 19. doi:10.1017/S0022029915000692.
- 474 Larsen, T., and C. Fernández. 2017. Enzymatic-fluorometric analyses for glutamine,  
475 glutamate and free amino groups in protein-free plasma and milk. *J. Dairy Res.* 84:32–  
476 35. doi:10.1017/S0022029916000789.
- 477 Larsen, T., and K.M. Moyes. 2010. Fluorometric determination of uric acid in bovine milk. *J.*  
478 *Dairy Res.* 77:438–444. doi:10.1017/S0022029910000580.
- 479 Larsen, T., and K.M. Moyes. 2015. Are free glucose and glucose-6-phosphate in milk  
480 indicators of specific physiological states in the cow?. *Anim. Int. J. Anim. Biosci.*  
481 9:86–93. doi:10.1017/S1751731114002043.
- 482 Larsen, T., and N.I. Nielsen. 2005. Fluorometric Determination of  $\beta$ -Hydroxybutyrate in Milk  
483 and Blood Plasma. *J. Dairy Sci.* 88:2004–2009. doi:10.3168/jds.S0022-  
484 0302(05)72876-9.

- 485 LeBlanc, S.J., K.E. Leslie, and T.F. Duffield. 2005. Metabolic Predictors of Displaced  
486 Abomasum in Dairy Cattle. *J. Dairy Sci.* 88:159–170. doi:10.3168/jds.S0022-  
487 0302(05)72674-6.
- 488 Lerch, S., A. Ferlay, K.J. Shingfield, B. Martin, D. Pomiès, and Y. Chilliard. 2012. Rapeseed  
489 or linseed supplements in grass-based diets: Effects on milk fatty acid composition of  
490 Holstein cows over two consecutive lactations. *J. Dairy Sci.* 95:5221–5241.  
491 doi:10.3168/jds.2012-5337.
- 492 Lobley, G.E., S.O. Hoskin, and C.J. McNeil. 2001. Glutamine in Animal Science and  
493 Production. *J. Nutr.* 131:2525S-2531S. doi:10.1093/jn/131.9.2525S.
- 494 Meijer, G. a. L., J.V.D. Meulen, J.G.M. Bakker, C.J.V.D. Koelen, and A.M.V. Vuuren. 1995.  
495 Free Amino Acids in Plasma and Muscle of High Yielding Dairy Cows in Early  
496 Lactation. *J. Dairy Sci.* 78:1131–1141. doi:10.3168/jds.S0022-0302(95)76730-3.
- 497 Miettinen, H., and P. Huhtanen. 1996. Effects of the Ratio of Ruminal Propionate to Butyrate  
498 on Milk Yield and Blood Metabolites in Dairy Cows. *J. Dairy Sci.* 79:851–861.  
499 doi:10.3168/jds.S0022-0302(96)76434-2.
- 500 Moyes, K.M., J.K. Drackley, J.L. Salak-Johnson, D.E. Morin, J.C. Hope, and J.J. Loor. 2009.  
501 Dietary-induced negative energy balance has minimal effects on innate immunity  
502 during a *Streptococcus uberis* mastitis challenge in dairy cows during midlactation. *J.*  
503 *Dairy Sci.* 92:4301–4316. doi:10.3168/jds.2009-2170.
- 504 Nielsen, N.I., K.L. Ingvarstsen, and T. Larsen. 2003. Diurnal Variation and the Effect of Feed  
505 Restriction on Plasma and Milk Metabolites in TMR-fed Dairy Cows. *J. Vet. Med. A*  
506 *Physiol. Pathol. Clin. Med.* 50:88–97. doi:10.1046/j.1439-0442.2003.00496.x.

- 507 Oetzel, G.R. 2004. Monitoring and testing dairy herds for metabolic disease. *Vet. Clin. North*  
508 *Am. Food Anim. Pract.* 20:651–674. doi:10.1016/j.cvfa.2004.06.006.
- 509 Pawłowski, K., J.A.A. Pires, Y. Faulconnier, C. Chambon, P. Germon, C. Boby, and C.  
510 Leroux. 2019. Mammary Gland Transcriptome and Proteome Modifications by  
511 Nutrient Restriction in Early Lactation Holstein Cows Challenged with Intra-  
512 Mammary Lipopolysaccharide. *Int. J. Mol. Sci.* 20:1156. doi:10.3390/ijms20051156.
- 513 Pires, J.A.A., Y. Chilliard, C. Delavaud, J. Rouel, D. Pomiès, and F. Blanc. 2015.  
514 Physiological adaptations and ovarian cyclicity of Holstein and Montbéliarde cows  
515 under two low-input production systems. *Anim. Int. J. Anim. Biosci.* 9:1986–1995.  
516 doi:10.1017/S1751731115001317.
- 517 Pires, J.A.A., C. Delavaud, Y. Faulconnier, D. Pomiès, and Y. Chilliard. 2013. Effects of body  
518 condition score at calving on indicators of fat and protein mobilization of  
519 periparturient Holstein-Friesian cows. *J. Dairy Sci.* 96:6423–6439.  
520 doi:10.3168/jds.2013-6801.
- 521 Pires, J.A.A., K. Pawłowski, J. Rouel, C. Delavaud, G. Foucras, P. Germon, and C. Leroux.  
522 2019. Undernutrition modified metabolic responses to intramammary  
523 lipopolysaccharide but had limited effects on selected inflammation indicators in  
524 early-lactation cows. *J. Dairy Sci.* doi:10.3168/jds.2018-15446.
- 525 Pires, J.A.A., L.F. Stumpf, I.D. Soutullo, J.B. Pescara, S.E. Stocks, and R.R. Grummer. 2016.  
526 Effects of abomasal infusion of nicotinic acid on responses to glucose and  $\beta$ -agonist  
527 challenges in underfed lactating cows. *J. Dairy Sci.* 99:2297–2307.  
528 doi:10.3168/jds.2015-10308.

- 529 Pomiès, D., B. Martin, Y. Chilliard, P. Pradel, and B. Rémond. 2007. Once-a-day milking of  
530 Holstein and Montbéliarde cows for 7 weeks in mid-lactation. *Anim. Int. J. Anim.*  
531 *Biosci.* 1. doi:10.1017/S1751731107000778.
- 532 Vlaeminck, B., C. Dufour, A.M. van Vuuren, A.R.J. Cabrita, R.J. Dewhurst, D. Demeyer, and  
533 V. Fievez. 2005. Use of Odd and Branched-Chain Fatty Acids in Rumen Contents and  
534 Milk as a Potential Microbial Marker. *J. Dairy Sci.* 88:1031–1042.  
535 doi:10.3168/jds.S0022-0302(05)72771-5.
- 536 Vlaeminck, B., R. Gervais, M.M. Rahman, F. Gadeyne, M. Gorniak, M. Doreau, and V.  
537 Fievez. 2015. Postruminal synthesis modifies the odd- and branched-chain fatty acid  
538 profile from the duodenum to milk. *J. Dairy Sci.* 98:4829–4840. doi:10.3168/jds.2014-  
539 9207.
- 540 Zachut, M., G. Kra, Y. Portnik, F. Shapiro, and N. Silanikove. 2016. Milk glucose-6-  
541 phosphate dehydrogenase activity and glucose-6-phosphate are associated with  
542 oxidative stress and serve as indicators of energy balance in dairy cows. *RSC Adv.*  
543 6:65412–65417. doi:10.1039/C6RA11924G.
- 544  
545  
546



547  
548  
549  
550  
551

**Table 1:** Diet ingredient and nutrient composition

Ingredient (% DM)	Corn silage	66.3
	Barley straw	8.0
	Corn grain	7.6
	Soybean meal	17.4
	Mineral and vitamin mix <sup>1</sup>	0.7
Nutrient composition (DM basis)	Net Energy (MJ/kg DM)	6.9
	PDI (g/kg DM)	106
	CP (% DM)	13.8
	Fat (% DM)	2.4
	Starch (% DM)	23.3
	NDF (% DM)	34.6
	ADF (% DM)	18.7

552 <sup>1</sup>Mineral and vitamin contained 4.5% P, 23% Ca, 4.5% Mg, 1% S, 400,000 IU/kg of vitamin  
553 A, 100,000 IU/kg of vitamin D3, 1,600 IU/kg of vitamin E, 400 IU/kg of vitamin B1, 1 g/kg  
554 of Cu, 5 g/kg of Zn, 4 g/kg of Mn, 0.1 g/kg of I, 40 mg/kg of Co and 24 mg/kg of Se;  
555 Galaphos Midi Repro granule, CCPA, Aurillac, France.

556

557 **Table 2.** Effects of feed restriction on energy balance, DMI, milk yield, and milk component  
 558 yield in midlactation Holstein (HOLS) and Montbéliarde (MONT) cows<sup>1</sup>.  
 559

	Period	SEM		P-value					
		CONT	REST	WEEK 1	WEEK 2	Breed	Period	Breed × Period	
Energy balance (MJ/d)	HOLS	37 <sup>a</sup>	-42 <sup>b</sup>	34 <sup>a</sup>	34 <sup>a</sup>	5	0.09	0.001	0.09
	MONT	23	-43	23	32	4			
DMI (kg/d)	HOLS	25 <sup>a</sup>	9 <sup>c</sup>	22 <sup>b</sup>	24 <sup>a</sup>	0.8	0.01	0.001	0.08
	MONT	21	8	19	23	0.8			
Milk yield (kg/d)	HOLS	29 <sup>a</sup>	19 <sup>d</sup>	24 <sup>c</sup>	27 <sup>b</sup>	1.2	0.01	0.001	0.25
	MONT	24	16	21	24	1.1			
Fat yield (g/d)	HOLS	1030 <sup>a</sup>	770 <sup>b</sup>	836 <sup>b</sup>	964 <sup>a</sup>	43	0.07	0.001	0.48
	MONT	896	695	739	919	38			
Protein yield (g/d)	HOLS	883 <sup>a</sup>	565 <sup>c</sup>	722 <sup>b</sup>	844 <sup>a</sup>	38	0.03	0.001	0.75
	MONT	785	494	628	782	36			
Lactose yield (g/d)	HOLS	1466 <sup>a</sup>	966 <sup>d</sup>	1197 <sup>c</sup>	1316 <sup>b</sup>	68	0.01	0.001	0.28
	MONT	1217	797	1007	1180	63			
Body weight (kg)	HOLS	680 <sup>a</sup>	628 <sup>c</sup>	-	661 <sup>b</sup>	21	0.35	0.001	0.64
	MONT	661	605	-	626	19			

560  
 561 <sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to  
 562 6). Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after  
 563 restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

564 <sup>a, b, c, d</sup> Concern main period effects pooled across both breeds. Period LSMEANS not sharing a  
 565 common superscript differ ( $P \leq 0.05$ ).

566 <sup>y, z</sup> Breed LSMEANS not sharing a common superscript differ within the period ( $P \leq 0.05$ ),  
 567 presented when Breed × Period effect was significant.

568

569 **Table 3.** Effects of feed restriction on plasma metabolite and insulin concentrations in  
 570 midlactation Holstein (HOLS) and Montbéliarde (MONT) cows<sup>1</sup>.  
 571

		Period	SEM	P -value					
		CONT	RES T	WEEK 1	WEEK 2	Breed	Period	Breed × Perio d	
NEFA (mM)	HOLS	0.11 <sup>b</sup>	0.69 <sup>a</sup>	0.09 <sup>b</sup>	0.08 <sup>b</sup>	0.03	0.93	0.001	0.98
	MON T	0.11	0.68	0.09	0.10	0.03			
BHB (mM)	HOLS	0.61 <sup>y,bc</sup>	0.64 <sup>a</sup>	0.56 <sup>ab</sup>	0.51 <sup>c</sup>	0.04	0.53	0.003	0.05
	MON T	0.48 <sup>z</sup>	0.62	0.59	0.52	0.03			
Glucose (mM)	HOLS	3.84 <sup>b</sup>	3.50 <sup>c</sup>	4.04 <sup>ab</sup>	3.95 <sup>a</sup>	0.07	0.08	0.001	0.77
	MON T	3.77	3.32	3.93	3.87	0.06			
Glutamat e (mM)	HOLS	0.10 <sup>b</sup>	0.11 <sup>a</sup>	0.09 <sup>c</sup>	0.09 <sup>c</sup>	0.003	0.07	0.001	0.17
	MON T	0.09	0.10	0.09	0.09	0.003			
Glutamin e (mM)	HOLS	0.16 <sup>a</sup>	0.11 <sup>c</sup>	0.16 <sup>b</sup>	0.15 <sup>ab</sup>	0.01	0.61	0.001	0.13
	MON T	0.16	0.10	0.13	0.15	0.01			
Urea (mM)	HOLS	4.61 <sup>a</sup>	3.50 <sup>b</sup>	3.71 <sup>b</sup>	4.63 <sup>a</sup>	0.41	0.07	0.001	0.3
	MON T	5.36	4.61	4.10	4.99	0.38			
NH <sub>2</sub> (mM) <sup>2</sup>	HOLS	1.43 <sup>a</sup>	1.21 <sup>b</sup>	1.47 <sup>a</sup>	1.48 <sup>a</sup>	0.04	0.01	0.001	0.19
	MON T	1.4	1.09	1.28	1.38	0.04			
Insulin (μU/ml)	HOLS	20.3 <sup>b</sup>	8.5 <sup>d</sup>	25.2 <sup>a</sup>	16.1 <sup>c</sup>	1.8	0.13	0.001	0.6
	MON T	15.7	7.2	24.5	13.7	1.6			

572  
 573 <sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to  
 574 6). Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after  
 575 restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

576 <sup>a, b, c, d</sup> **Concern main period effects pooled across both breeds.** Period LSMEANS not sharing a  
 577 common superscript differ ( $P \leq 0.05$ ).

578 <sup>y, z</sup> Breed LSMEANS not sharing a common superscript differ within the period ( $P \leq 0.05$ ),  
 579 presented when Breed × Period effect was significant.

580 <sup>2</sup> Free amino groups (NH<sub>2</sub>): Estimation of free amino acid concentration

581



583 **Table 4.** Effects of feed restriction on milk metabolite concentrations in midlactation Holstein  
 584 (HOLS) and Montbéliarde (MONT) cows<sup>1</sup>.

	Period	SEM		P-value					
		CON T	RES T	WEEK 1	WEEK 2	Breed	Period	Breed × Period	
BHB (μM)	HOLS	54.7 <sup>b</sup>	40.3 <sup>c</sup>	64.5 <sup>a</sup>	46.7 <sup>b</sup>	5.2	0.93	0.001	0.55
	MONT	49.1	41.6	62.0	51.8	4.6			
Glucose (mM)	HOLS	0.54 <sup>a</sup>	0.22 <sup>c</sup>	0.47 <sup>b</sup>	0.51 <sup>ab</sup>	0.03	0.15	0.001	0.17
	MONT	0.47	0.21	0.45	0.45	0.02			
Glucose-6-phosphate (μM)	HOLS	20.5 <sup>b</sup>	32.2 <sup>a</sup>	14.0 <sup>c</sup>	27.2 <sup>a</sup>	8.9	0.24	0.001	0.12
	MONT	35.5	45.9	18.8	48.7	7.9			
Glutamate (mM)	HOLS	0.39 <sup>a</sup>	0.14 <sup>c</sup>	0.39 <sup>a</sup>	0.33 <sup>b</sup>	0.06	0.77	0.001	0.99
	MONT	0.38	0.13	0.38	0.31	0.06			
Isocitrate (mM)	HOLS	0.12 <sup>c</sup>	0.17 <sup>a</sup>	0.13 <sup>c</sup>	0.13 <sup>b</sup>	0.01	0.05	0.001	0.29
	MONT	0.14	0.19	0.13	0.16	0.01			
Uric acid (mM)	HOLS	0.15 <sup>a</sup>	0.09 <sup>c</sup>	0.13 <sup>b</sup>	0.16 <sup>a</sup>	0.01	0.003	0.001	0.54
	MONT	0.12	0.07	0.09	0.12	0.01			
NH <sub>2</sub> (mM) <sup>1</sup>	HOLS	0.81 <sup>a</sup>	0.71 <sup>b</sup>	0.84 <sup>a</sup>	0.83 <sup>a</sup>	0.04	0.31	0.001	0.72
	MONT	0.79	0.66	0.79	0.79	0.03			

585  
 586 <sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to 6).  
 587 Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after  
 588 restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

589 <sup>a, b, c, d</sup> **Concern main period effects pooled across both breeds.** Period LSMEANS not sharing a  
 590 common superscript differ ( $P \leq 0.05$ ).

591 <sup>2</sup> Free amino groups (NH<sub>2</sub>): Estimation of free amino acid concentration (Larsen and Fernández,  
 592 2017).

593  
 594  
 595

596 **Table 5.** Effects of feed restriction on milk fatty acid concentrations (g/100g of fatty acids) in  
 597 midlactation Holstein (HOLS) and Montbéliarde (MONT) cows<sup>1</sup>. Milk fatty acid composition  
 598 was analyzed in pooled PM/AM samples.

	Period	SEM		P-value				
		CON T	RES T	WEEK 1	Breed	Period	Breed × Period	
Σ 10:0 to 15:0 <sup>2</sup>	HOLS	23.3 <sup>a</sup>	14.9 <sup>b</sup>	22.8 <sup>a</sup>	1.3	0.38	0.001	0.49
	MONT	23.9	14.5	25.3	1.1			
16:0	HOLS	35.0 <sup>b</sup>	26.6 <sup>c</sup>	35.2 <sup>a</sup>	1.1	0.89	0.001	0.18
	MONT	33.6	25.8	37.8	0.9			
18:0	HOLS	7.2 <sup>b</sup>	11.1 <sup>a</sup>	8.1 <sup>b</sup>	0.6	0.02	0.001	0.25
	MONT	6.9	10.4	5.7	0.5			
<i>cis</i> -9 18:1	HOLS	18.0 <sup>b</sup>	29.6 <sup>a</sup>	17.5 <sup>b</sup>	1.6	0.97	0.001	0.4
	MONT	18.7	31.2	15.1	1.4			
Σ >C16 <sup>3</sup>	HOLS	33.0 <sup>b</sup>	52.2 <sup>a</sup>	33.8 <sup>b</sup>	2.2	0.43	0.001	0.3
	MONT	33.2	53.2	28.3	1.9			
Σ OBCFA	HOLS	3.9	3.7	4.0	0.2	0.37	0.68	0.11
	MONT	3.7	3.8	3.6	0.2			
Σ OBCFA <C16 <sup>4</sup>	HOLS	2.2 <sup>b</sup>	1.8 <sup>c</sup>	2.5 <sup>a</sup>	0.2	0.64	0.001	0.81
	MONT	2.1	1.8	2.4	0.1			

599  
 600 <sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to 6).  
 601 Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after  
 602 restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

603 <sup>a, b, c, d</sup> **Concern main period effects pooled across both breeds.** Period LSMEANS not sharing a  
 604 common superscript differ ( $P \leq 0.05$ ).

605 <sup>2</sup>Sum of FA with between 10 and 15 carbons.

606 <sup>3</sup>Sum of FA with more than 16 carbons.

607 <sup>4</sup>Sum of OBCFA with less than 16 carbons.

508  
509  
510  
511

**Table 6.** Spearman rank correlations among milk metabolites and classic indicators of metabolic status in midlactation Holstein and Montbéliarde cows<sup>1</sup>. Metabolite concentrations were measured in morning milk and plasma samples. Number in brackets correspond to the number of observations (\*\*\*  $P < 0.001$ ; \*\*  $P < 0.01$ ; \*  $P < 0.05$ ; †  $P < 0.1$ ).

	Energy balance	Plasma NEFA	Plasma BHB	Plasma Glucose	Plasma NH <sub>2</sub> <sup>1</sup>	Milk BHB	Milk Glucose	Milk Glucose-6-phosphate	Milk Glutamate	Milk Isocitrate	Milk Uric acid	Milk NH <sub>2</sub> <sup>1</sup>													
Energy balance	1.00	-0.72	**	-0.17	*	0.64	**	0.34	***	0.51	***	0.48	**	0.63	**	-0.31	**	0.61	**	-0.45	**	0.30	**	0.41	***
	(306)	(231)	(231)	(231)	(231)	(231)	(231)	(230)	(231)	(231)	(231)	(286)	(285)	(286)	(285)	(286)	(286)	(286)	(286)	(286)	(286)	(284)	(284)	(284)	(284)
Plasma NEFA		1.00	0.17	*	-0.53	**	0.45	***	-0.44	**	-0.67	**	0.36	**	-0.67	**	0.48	**	-0.44	**	-0.46	***	***	***	***
		(249)	(249)	(249)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(247)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(246)	(246)	(246)	(246)	(246)
Plasma BHB			1.00	-0.33	**	0.28	***	-0.17	**	-0.27	**	-0.09	ns	-0.11	†	0.31	**	-0.31	**	-0.06	ns	ns	ns	ns	ns
			(249)	(249)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(247)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(246)	(246)	(246)	(246)	(246)
Plasma glucose				1.00	-	***	0.46	***	0.35	**	0.61	**	-0.25	**	0.56	**	-0.30	**	0.31	**	0.46	***	***	***	***
				(249)	(249)	(248)	(248)	(248)	(248)	(248)	(248)	(247)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(246)	(246)	(246)	(246)
Plasma glutamate					1.00	-	***	0.17	**	-0.14	*	-0.36	**	0.19	**	-0.39	**	0.33	**	-0.22	**	-0.25	***	***	***
					(249)	(249)	(248)	(248)	(248)	(248)	(248)	(247)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(248)	(245)	(245)	(245)	(245)
Plasma NH <sub>2</sub> <sup>1</sup>						1.00	0.26	**	0.50	**	-0.20	**	0.59	**	-0.34	**	0.50	**	0.44	***	***	***	***	***	***
						(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)	(300)

Version preprint

	(248)	(247)	(246)	(247)	(246)	(247)	(247)	(247)	(245)	
<b>Milk BHB</b>	1.00	0.30** *	-0.18**	**	0.45** *	-0.08	ns	-0.03	ns	0.42***
<b>Milk glucose</b>	(304)	(303)	(304)	(303)	(303)	(304)	(304)	(304)	(302)	
		1.00	-0.18**	**	0.70** *	-0.50** *	**	0.51** *	**	0.51***
<b>Milk glucose-6-phosphate</b>		(303)	(303)	(303)	(303)	(303)	(303)	(303)	(302)	
			1.00	-0.28** *	0.34** *	**	0.05	ns	-0.29***	
<b>Milk glutamate</b>			(304)	(303)	(304)	(304)	(304)	(304)	(302)	
				1.00	-0.36** *	**	0.38** *	**	0.72***	
<b>Milk isocitrate</b>				(303)	(303)	(303)	(303)	(303)	(302)	
					1.00	-0.17	**	-0.26	***	
<b>Milk uric acid</b>						(304)	(304)	(302)	(302)	
							1.00	0.28	***	
<b>Milk NH<sub>2</sub><sup>2</sup></b>								(304)	(302)	
									(302)	

612  
613  
614  
615  
616  
617

<sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to 6). Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

<sup>2</sup> Free amino groups (NH<sub>2</sub>): Estimation of free amino acid concentrations (Larsen and Fernández, 2017).

Comment citer ce document :

Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. Journal of Dairy Science, 103 (4), 3133–3146.



**Table 7.** Spearman rank correlations among milk fatty acids and milk metabolites and classic indicators of metabolic status in midlactation Holstein and Montbéliarde cows<sup>1</sup>. Milk fatty acid composition was analyzed in pooled PM/AM milk samples, and metabolite concentrations were measured in morning milk and plasma samples. Number in brackets corresponds to the number of observations. (\*\*\*  $P < 0.001$ ; \*\*  $P < 0.01$ ; \*  $P < 0.05$ ; †  $P < 0.1$ ).

	$\Sigma$ 10:0 to 15:0 <sup>2</sup>	16:0	18:0	<i>Cis</i> -9 18:1	$\Sigma$ >C16 <sup>3</sup>	$\Sigma$ OBCFA <C16 <sup>4</sup>	$\Sigma$ OBCFA								
Energy balance				***	***	-0.56 (162)	***	-0.54 (162)	***	-0.55 (162)	***	0.39 (162)	***	-0.09 (162)	ns
Plasma NEFA				***	***	0.61 (143)	***	0.68 (143)	***	0.69 (143)	***	-0.58 (143)	***	-0.09 (143)	ns
Plasma BHB				***	***	0.37 (143)	***	0.43 (143)	***	0.44 (143)	***	-0.48 (143)	***	-0.27 (143)	**
Plasma glucose				***	***	-0.36 (143)	***	-0.45 (143)	***	-0.44 (143)	***	0.28 (143)	***	-0.07 (143)	ns
Plasma glutamate				***	***	0.52 (143)	***	0.52 (143)	***	0.55 (143)	***	-0.55 (143)	***	-0.13 (143)	ns
Plasma NH <sub>2</sub> <sup>5</sup>				***	***	-0.41 (143)	***	-0.55 (143)	***	-0.52 (143)	***	0.51 (143)	***	0.18 (143)	*
Milk BHB				***	***	-0.23 (161)	**	-0.38 (161)	***	-0.35 (161)	***	0.15 (161)	ns	-0.23 (161)	**
Milk glucose				***	***	-0.57 (160)	***	-0.66 (160)	***	-0.66 (160)	***	0.45 (160)	***	-0.06 (160)	ns
Milk glucose-6-phosphate				ns	ns	0.25 (161)	**	0.09 (161)	ns	0.15 (161)	†	-0.05 (161)	ns	0.13 (161)	ns
Milk glutamate				***	***	-0.68 (160)	***	-0.70 (160)	***	-0.71 (160)	***	0.57 (160)	***	0.03 (160)	ns
Milk isocitrate				***	***	0.61 (161)	***	0.56 (161)	***	0.61 (161)	***	-0.47 (161)	***	0.04 (161)	ns
Milk uric acid				***	***	-0.28 (161)	***	-0.48 (161)	***	-0.43 (161)	***	0.53 (161)	***	0.40 (161)	***
Milk NH <sub>2</sub> <sup>5</sup>				***	***	-0.43 (160)	***	-0.53 (160)	***	-0.52 (160)	***	0.31 (160)	***	-0.16 (160)	*
$\Sigma$ 10:0 to 15:0 <sup>2</sup>					***	-0.85 (162)	***	-0.95 (162)	***	-0.96 (162)	***	0.89 (162)	***	0.28 (162)	***
16:0						-0.78 (162)	***	-0.91 (162)	***	-0.92 (162)	***	0.70 (162)	***	0.04 (162)	ns
18:0						1.00		0.78	***	0.86	***	-0.74	***	-0.10	ns

Comment citer ce document :

Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. *Journal of Dairy Science*, 103 (4), 3133–3146.

	(162)	(162)	(162)	(162)	(162)	(162)
<b>Cis-9 18:1</b>		1.00	0.97 ***	-0.85 ***	-0.23 **	
		(162)	(162)	(162)	(162)	
<b>Σ &gt;C16<sup>3</sup></b>			1.00	-0.83 ***	-0.16 *	
			(162)	(162)	(162)	
<b>Σ OBCFA &lt;C16<sup>4</sup></b>				1.00	0.61 ***	
				(162)	(162)	
<b>Σ OBCFA</b>					1.00	
					(162)	

---

<sup>1</sup> Intake was limited to meet 50% of NE<sub>L</sub> requirements during restriction period (REST, d 1 to 6). Cows were allowed ad libitum intake during control period (CONT, d -3 to -1) and after restriction (WEEK1, d 7 to 13; WEEK2, d 14 to 18).

<sup>2</sup>Sum of FA with between 10 and 15 carbons.

<sup>3</sup>Sum of FA with more than 16 carbons.

<sup>4</sup>Sum of OBCFA with less than 16 carbons.

<sup>5</sup> Free amino groups (NH<sub>2</sub>): Estimation of amino acid concentration (Larsen and Fernández, 2017).

Comment citer ce document :

Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. Journal of Dairy Science, 103 (4), 3133–3146.

630 **Figure 1:** Sampling timeline. Control period (CONT; d -3 to -1); restriction period (REST; d 1  
 631 to 6, thick line), when feed intake was restricted to meet 50% of  $NE_L$  requirements calculated  
 632 during the CONT period; week 1 after refeeding (WEEK1; d 7 to 13); week 2 after refeeding  
 633 (WEEK2: d 14 to 18).  $\diamond$  Milk yield and composition each milking.  $\infty$  Milk sampling for milk  
 634 metabolite analyses (d 1 corresponds to samples collected at 24 h of feed restriction).  $\circ$  Blood  
 635 sampling for plasma metabolite and insulin analyses (d 1 corresponds to samples collected at  
 636 24 h of feed restriction).  $\Delta$  Milk sampling for milk fatty acid composition analyses (pooled  
 637 PM and AM samples).

638 **Figure 2.** Effects of feed restriction on milk concentrations of (A) glucose, (B) glucose-6-  
 639 phosphate, (C) isocitrate and (D)  $\beta$ -hydroxybutyrate (BHB) in midlactation Holstein (HOLS;  
 640  $\bullet$ , solid lines) and Montbéliarde (MONT;  $\blacktriangle$ , dashed lines) cows. Intake was limited to meet  
 641 50% of  $NE_L$  requirements during restriction period (d 1 to 6). Cows were allowed ad libitum  
 642 intake during control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM  $\pm$   
 643 SEM.

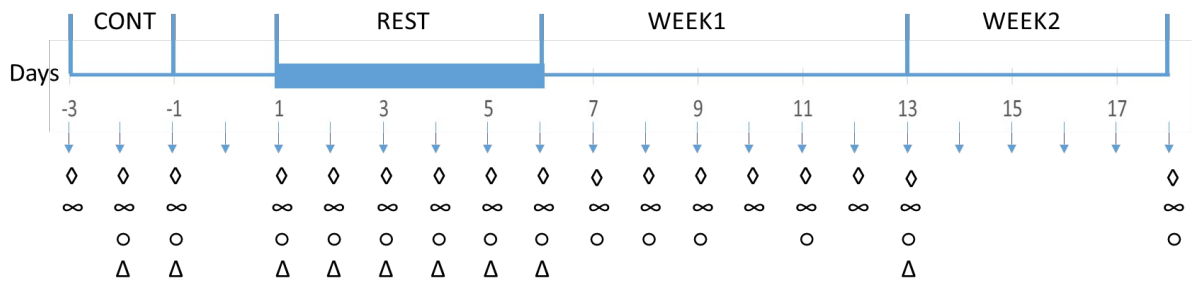
644  
 645 **Figure 3.** Effects of feed restriction on milk concentrations of (A) free amino group ( $NH_2$ ),  
 646 (B) glutamate and (C) uric acid in midlactation Holstein (HOLS;  $\bullet$ , solid lines) and  
 647 Montbéliarde (MONT;  $\blacktriangle$ , dashed lines) cows. Intake was limited to meet 50% of  $NE_L$   
 648 requirements during restriction period (d 1 to 6). Cows were allowed ad libitum intake during  
 649 control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM  $\pm$  SEM.

650  
 651 **Figure 4.** Relationship between energy balance and (A) milk glucose ( $y = 171.0x - 62.1, r^2 =$   
 652  $0.49$ ), (B) natural log of milk glutamate ( $y = 42.7x + 62.7, r^2 = 0.46$ ), (C) natural log of milk  
 653 glucose-6-phosphate ( $y = -9.5x - 37.8, r^2 = 0.10$ ) and (D) milk isocitrate ( $y = -458.4 + 70.7, r^2$   
 654  $= 0.26$ ) in midlactation Holstein (HOLS;  $\bullet$ ) and Montbéliarde (MONT;  $\blacktriangle$ ) cows ( $P < 0.001$ ).  
 655 Intake was limited to meet 50% of  $NE_L$  requirements during restriction period (d 1 to 6). Cows  
 656 were allowed ad libitum intake during control period (d -3 to -1) and after restriction (d 7 to  
 657 18).

658  
 659

660 Billa et al. Figure 1

661



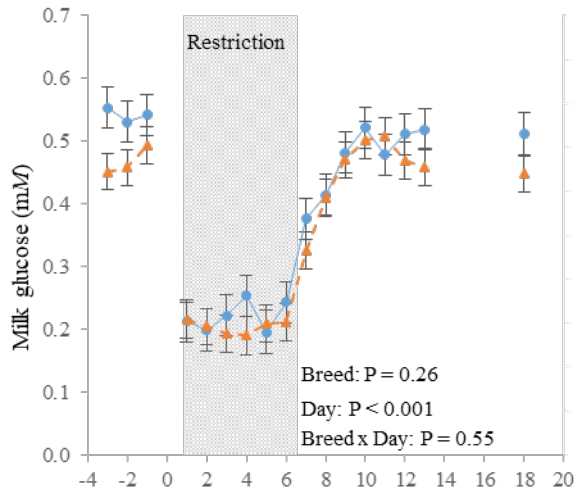
662

663 Billa et al. Figure 2

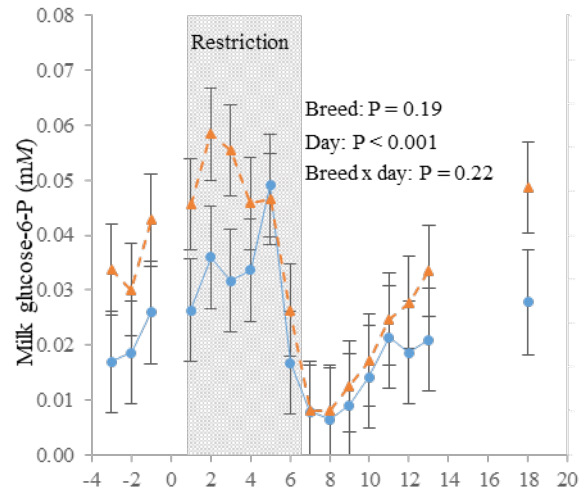
664

665

**A**



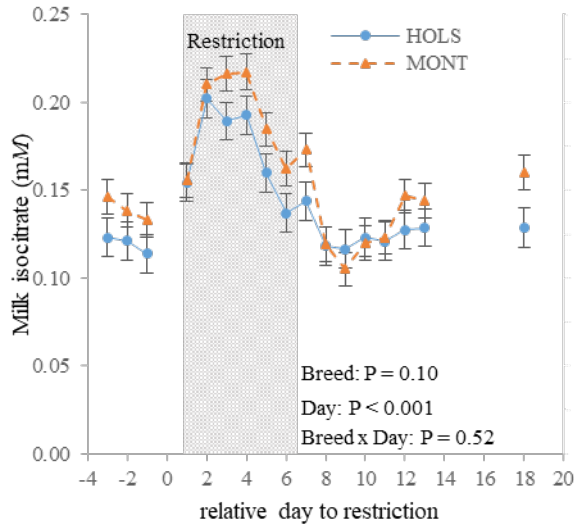
**B**



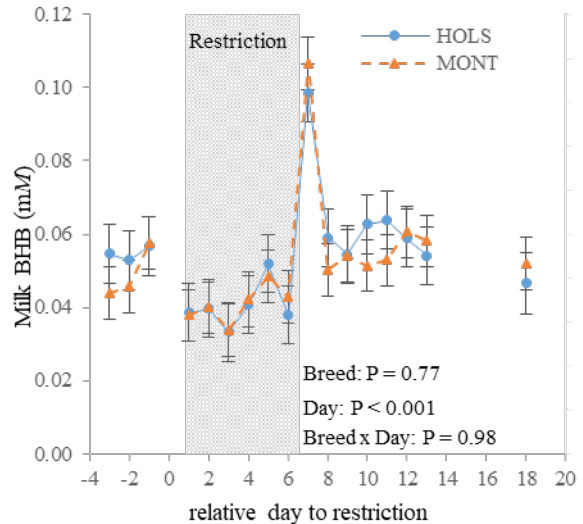
666

667

**C**



**D**



668

669

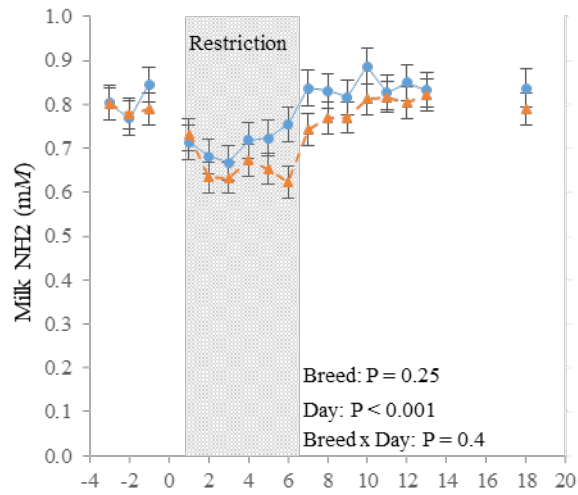
670

Comment citer ce document :

Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. *Journal of Dairy Science*, 103 (4), 3133–3146.

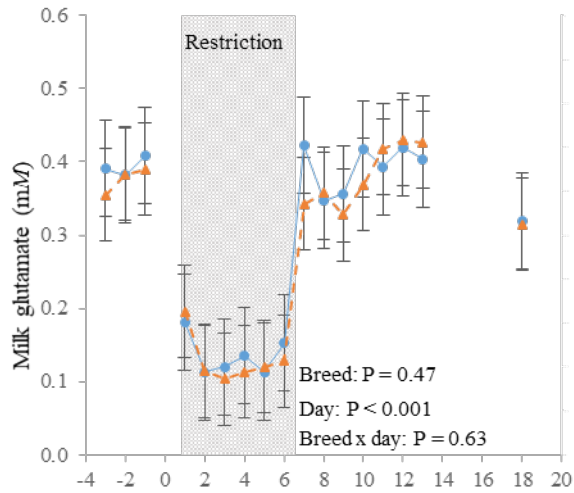
671 Billa et al. Figure 3

672 **A**



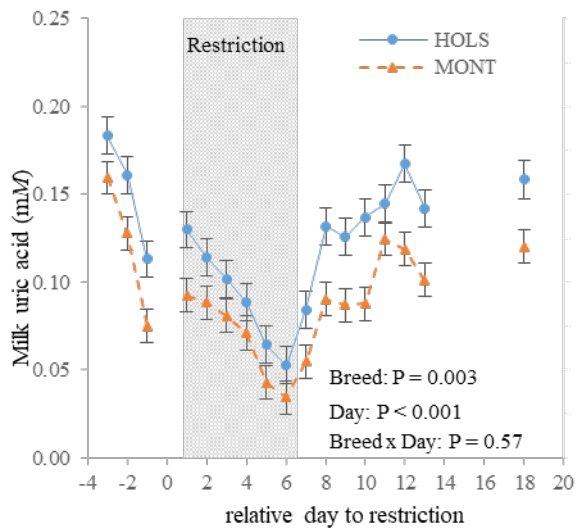
673

674 **B**



675

676 **C**

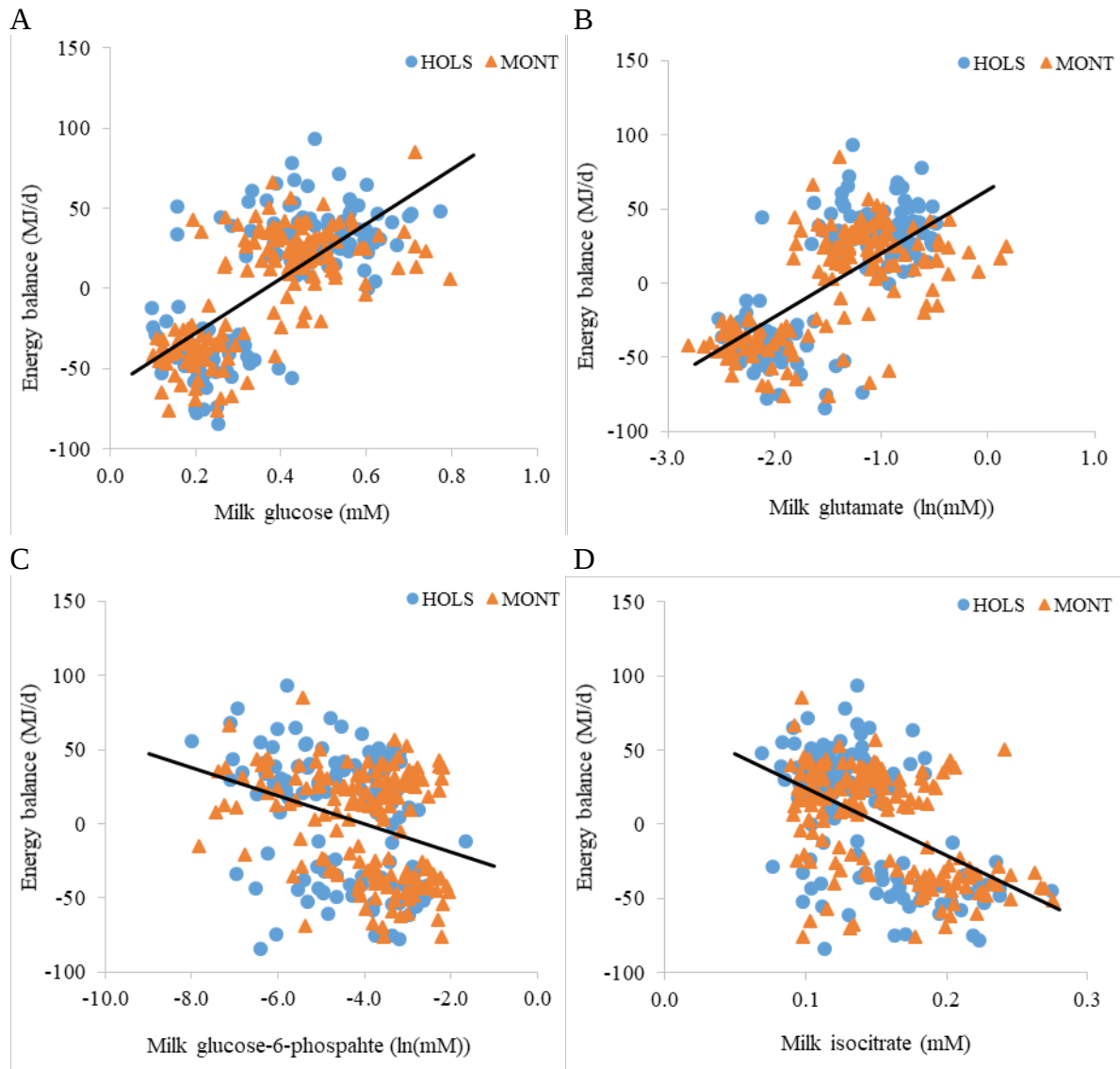


677

678 Billa et al. Figure 4

679

680



681

682

683

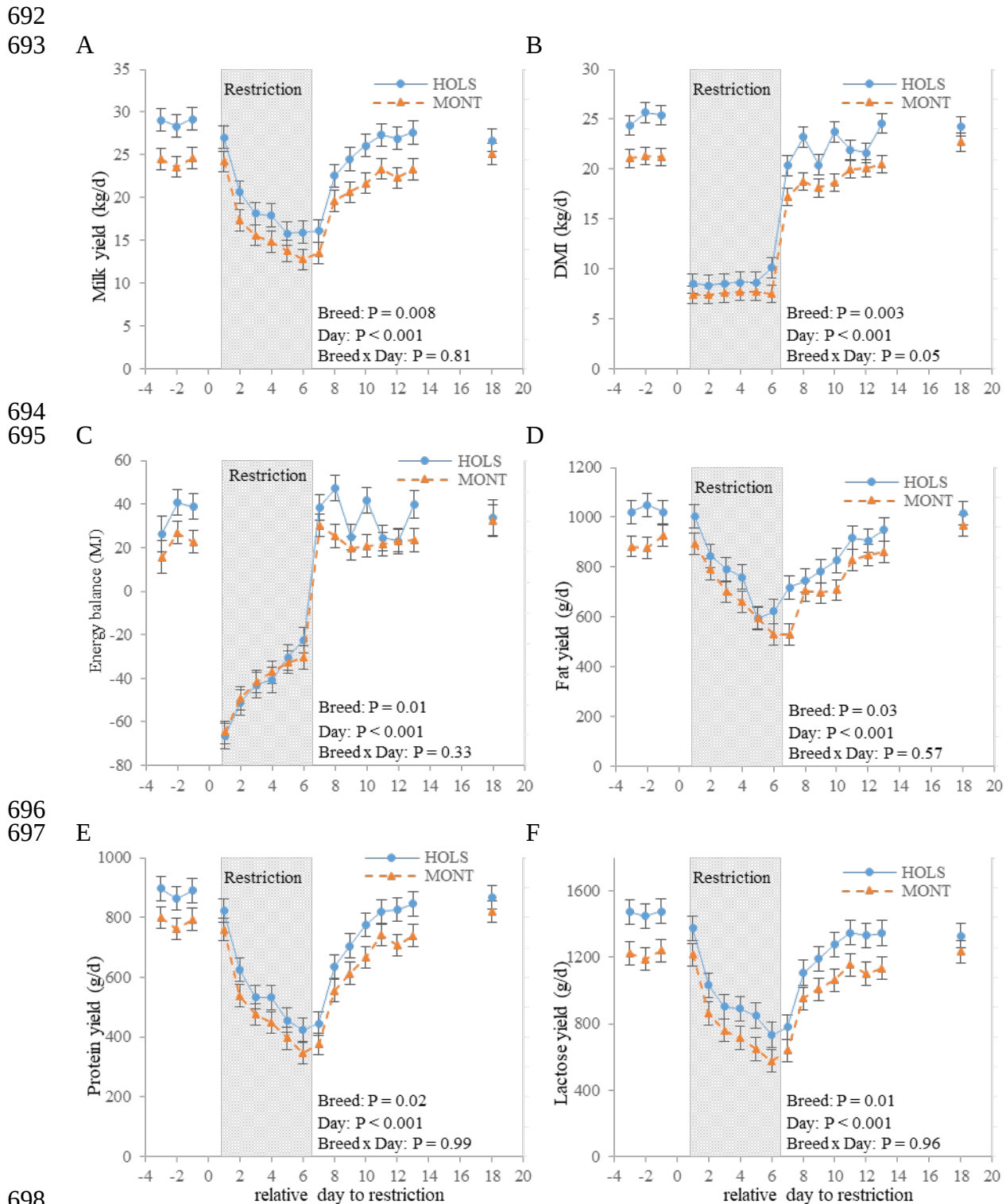
684

685

Comment citer ce document :

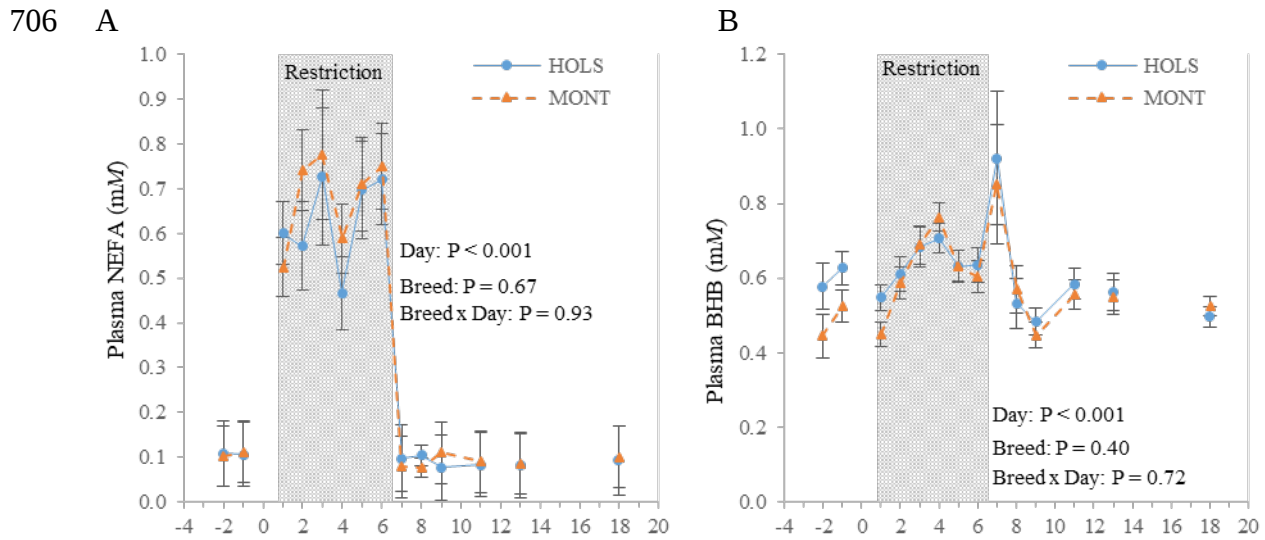
Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. *Journal of Dairy Science*, 103 (4), 3133–3146.

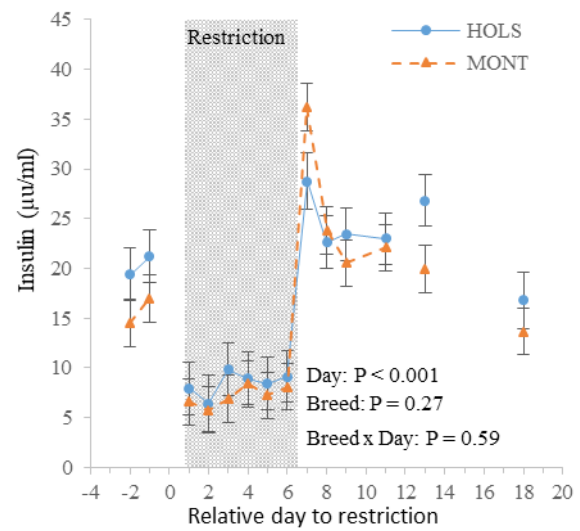
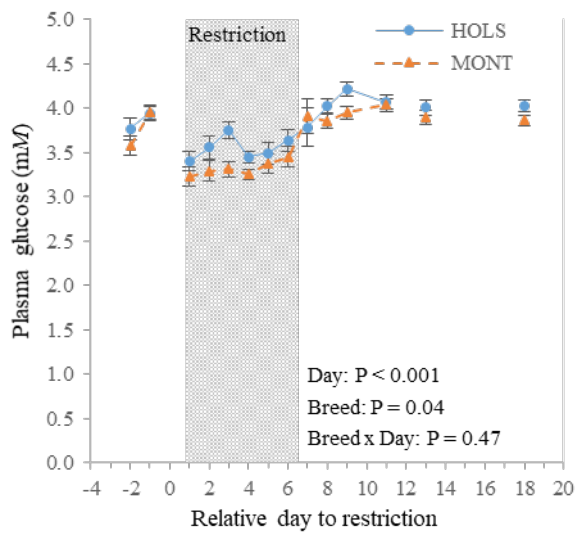
686 **Supplemental Figure 1.** Effects of feed restriction on milk yield (A), DMI (B), **energy**  
 687 **balance (C)**, fat (D), protein (E) and lactose (F) yield in midlactation Holstein (HOLS; ●,  
 688 solid lines) and Montbéliarde (MONT; ▲, dashed lines) cows. Intake was limited to meet  
 689 50% of  $NE_L$  requirements during restriction period (d 1 to 6). Cows were allowed *ad libitum*  
 690 intake during control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM  $\pm$   
 691 SEM.





699 **Supplemental Figure 2.** Effects of feed restriction on plasma concentrations of non-esterified  
 700 fatty acid (NEFA) (A), BHB (B), glucose (C) and insulin (D) in midlactation Holstein (HOLS;  
 701 ●, solid lines) and Montbéliarde (MONT; ▲, dashed lines) cows. Intake was limited to meet  
 702 50% of  $NE_L$  requirements during restriction period (d 1 to 6). Cows were allowed *ad libitum*  
 703 intake during control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM  $\pm$   
 704 SEM.  
 705

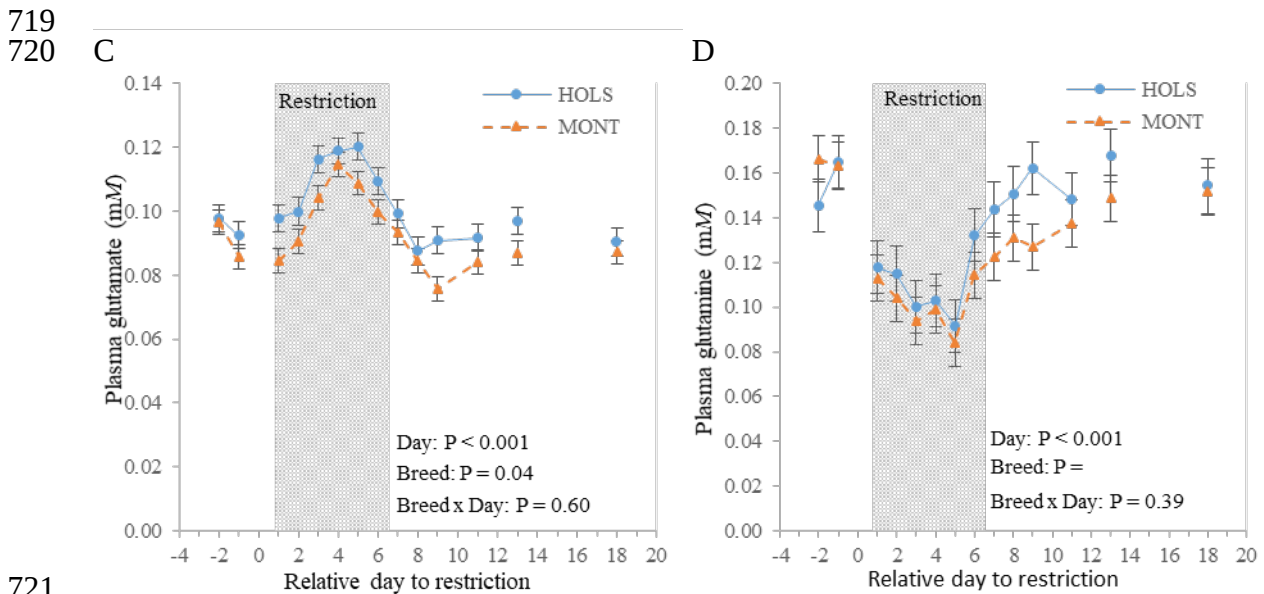
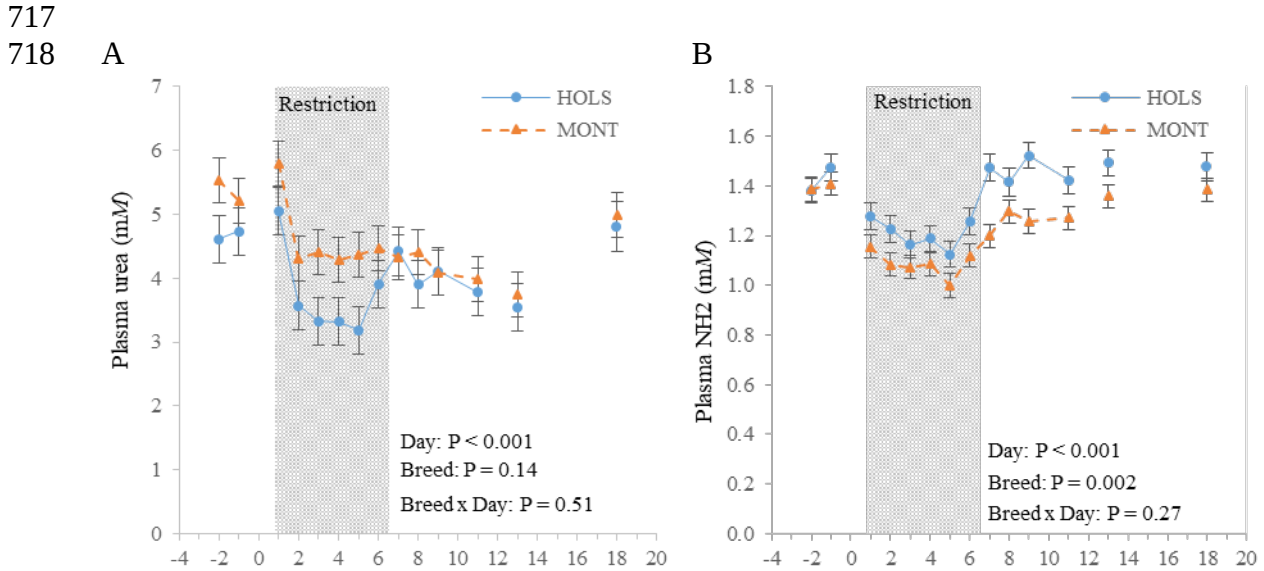




709

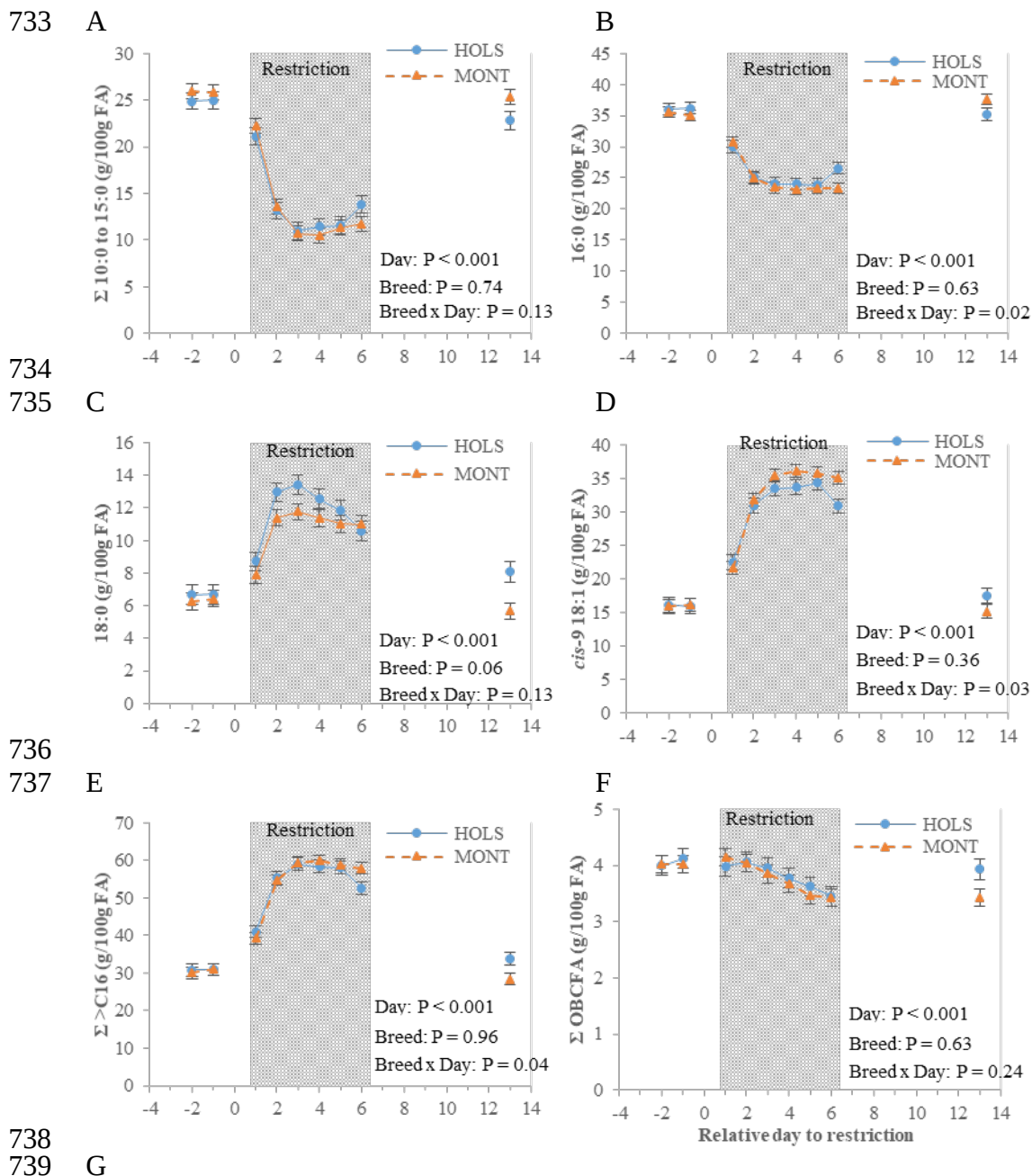
710

711 **Supplemental Figure 3.** Effects of feed restriction on plasma concentrations of urea (A), free  
 712 amino groups (NH<sub>2</sub>) (B), glutamate (C) and glutamine (D) in midlactation Holstein (HOLS; ●,  
 713 solid lines) and Montbéliarde (MONT; ▲, dashed lines) cows. Intake was limited to meet  
 714 50% of NE<sub>L</sub> requirements during restriction period (d 1 to 6). Cows were allowed ad libitum  
 715 intake during control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM ±  
 716 SEM.



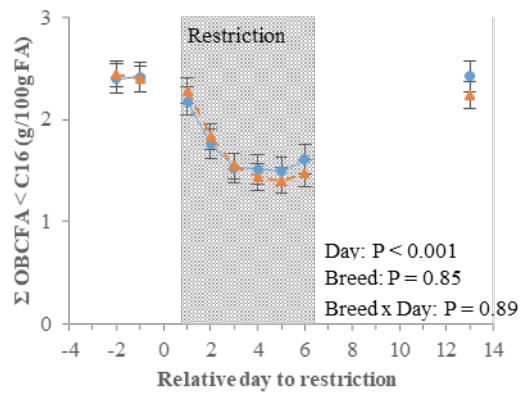
721  
722  
723

724 **Supplemental Figure 4.** Effects of feed restriction on fatty acid concentrations of sum of FA  
 725 with 10 to 15 carbons ( $\Sigma$  10:0 to 15:0) (A), 16:0 (B), 18:0 (C), *cis*-9 18:1 (D), sum greater  
 726 than C16 carbon ( $\Sigma$ >C16) (E), sum odd and branched fatty acid ( $\Sigma$  OBCFA) (D), sum  
 727 OBCFA with less than 16 carbon ( $\Sigma$  OBCFA<C16) (G) in midlactation Holstein (HOLS; ●,  
 728 solid lines) and Montbéliarde (MONT; ▲, dashed lines) cows. Fatty acid composition was  
 729 analyzed in pooled PM/AM milk samples. Intake was limited to meet 50% of  $NE_L$   
 730 requirements during restriction period (d 1 to 6). Cows were allowed *ad libitum* intake during  
 731 control period (d -3 to -1) and after restriction (d 7 to 18). Values are LSM  $\pm$  SEM.  
 732  
 733



Comment citer ce document :

Billa, P.-A., Faulconnier, Y., Larsen, T., Leroux, C., Pires, J. (2020). Milk metabolites as noninvasive indicators of nutritional status of mid-lactation Holstein and Montbéliarde cows. Journal of Dairy Science, 103 (4), 3133–3146.

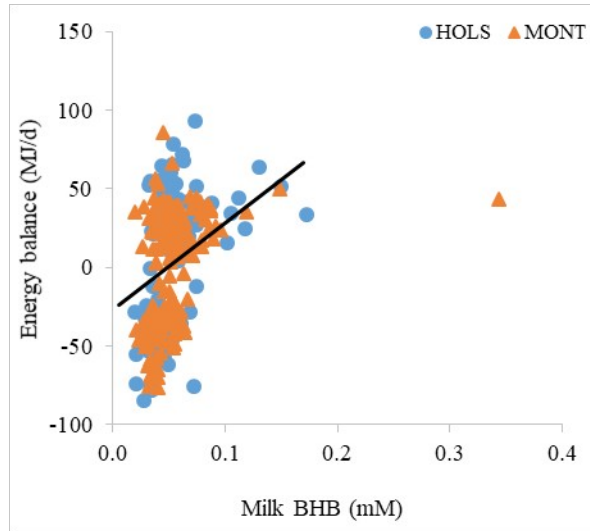


740  
741

742

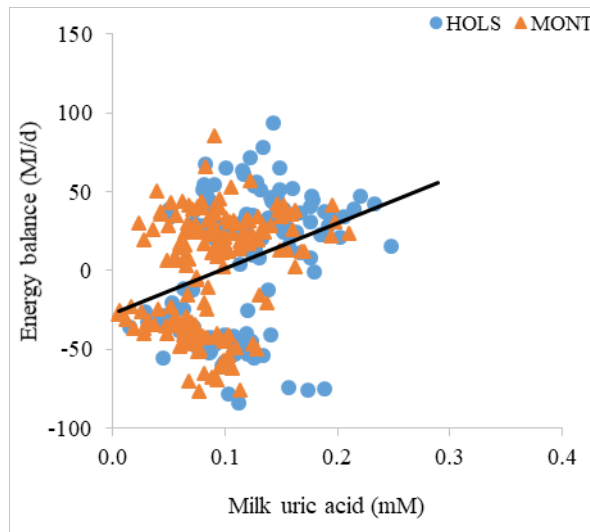
743 **Supplemental Figure 5.** Relationship between energy balance and (A) milk BHB ( $y = 549.5x$   
744  $- 26.8$ ,  $r^2 = 0.14$ ), (B) milk uric acid ( $y = 288.4x - 27.5$ ,  $r^2 = 0.11$ ) and (C) milk total free  
745 amino groups ( $\text{NH}_2$ ;  $y = 129.3x - 96.5$ ,  $r^2 = 0.17$ ) in midlactation Holstein (HOLS; ●) and  
746 Montbéliarde (MONT; ▲) cows ( $P < 0.001$ ). Intake was limited to meet 50% of  $\text{NE}_L$   
747 requirements during restriction period (d 1 to 6). Cows were allowed ad libitum intake during  
748 control period (d -3 to -1) and after restriction (d 7 to 18).  
749  
750

A



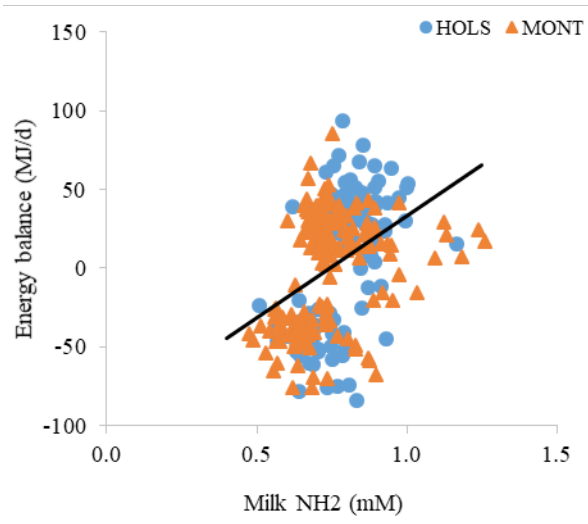
751  
752

B



753  
754

C



755  
756  
757