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Louis Lefaucheur, Bénédicte Lebret

► **To cite this version:**

Louis Lefaucheur, Bénédicte Lebret. The rearing system modulates biochemical and histological differences in loin and ham muscles between basque and large white pigs. *Animal*, 2020, 14 (9), pp.1976-1986. 10.1017/S175173112000066X . hal-02625150

HAL Id: hal-02625150

<https://hal.inrae.fr/hal-02625150>

Submitted on 26 May 2020

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1 **The rearing system modulates biochemical and histological differences in loin**
2 **and ham muscles between Basque and Large White pigs**

3 L. Lefaucheur ¹ and B. Lebret ¹

4

5 ¹ INRAE, Agrocampus Ouest, PEGASE, 35590 Saint-Gilles, France

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7 Corresponding author: Bénédicte Lebret. E-mail: Benedicte.Lebret@inrae.fr

8

9 Short title: Muscle properties, pig breed and rearing system

10

11 **Abstract**

12 Conventional pork production, based on highly selected breeds for growth efficiency
13 and carcass leanness, is generally considered to decrease pork quality. In contrast,
14 non-selected breeds produced in extensive systems are associated with high pork
15 quality, which is generally attributed to higher intramuscular fat (IMF) content and
16 less glycolytic muscle metabolism. The present study aimed to determine
17 biochemical, histological and quality traits of loin and ham muscles of pigs from
18 selected Large White (LW) and local French, non-selected Basque (B) breeds. Pigs
19 were reared in a conventional indoor (C, slatted floor), alternative (A, indoor bedding
20 and outdoor area) or extensive system (E, free range, B pigs only). A total of 100
21 castrated males were produced in two replicates, each containing 5 groups of 10 pigs
22 based on breed and system: LWC, LWA, BC, BA, and BE. The glycolytic longissimus
23 (LM) and semimembranosus (SM) muscles, and the deep red (RSTM) and superficial
24 white (WSTM) portions of semitendinosus muscle (STM), were studied at 145 kg
25 body weight. Overall, breed induced stronger effects on muscle traits than the rearing

26 system, among which the E system induced greater changes. The lower muscle
27 growth of B pigs was associated with fewer muscle fibers and a smaller cross-
28 sectional area (CSA) of glycolytic fibers ($P < 0.01$). The SM was less glycolytic and
29 more oxidative in B than in LW pigs ($P < 0.001$). The WSTM followed a similar trend,
30 with a larger relative area of type I fibers in B pigs. In contrast, the LM and RSTM
31 were more oxidative in LW pigs. B pigs had higher IMF content and ultimate pH in all
32 muscles, along with lower glycolytic potential, less light and redder meat in the LM
33 and SM ($P < 0.001$). Compared to the C system, the A system induced only a shift
34 towards a more oxidative metabolism in the LM and a smaller fiber CSA in the RSTM
35 of LW pigs ($P < 0.05$), without influencing pork quality traits. Compared to BC pigs,
36 BE pigs had a more oxidative and less glycolytic muscle metabolism, along with
37 higher ultimate pH, lower lightness and redder meat ($P < 0.01$), but similar IMF
38 content. Overall, results indicate that influences of breed and rearing system on
39 muscle properties depend on muscle type, and that IMF content and fiber type
40 composition are unrelated traits that can be modified independently by genetic or
41 rearing factors.

42
43 **Keywords:** muscle fiber type, muscle energy metabolism, intramuscular fat, pork
44 quality, breed

45 46 **Implications**

47 Compared to conventional production of high-performing pig breeds, non-selected
48 local breeds, often produced in outdoor or extensive systems, generally have better
49 pork quality. Muscle biology (fiber type composition, metabolism and lipid content)
50 and pork quality traits differed greatly between selected Large White and local

51 Basque pig breeds. Moreover, compared to conventional systems, the extensive
52 system further improved Basque pork quality, while the alternative system (bedding
53 and outdoor area) had less influence on muscle and pork properties. Independence
54 between fiber type composition and lipid content allows each to be controlled by
55 genetics or pig breeding to optimize pork quality.

56

57 **Introduction**

58 In the context of consumers' increasing expectations for improved sustainability of
59 production systems, animal welfare and eating quality of meat products (Font-i-
60 Furnols and Guerro, 2014), pork products from local pig breeds reared in extensive
61 systems have an excellent image among consumers (Bonneau and Leuret, 2010).
62 Several local pig breeds have been maintained as a genetic-diversity reservoir; they
63 generally have a lower growth rate and lean meat content, as well as higher fatness
64 and eating quality, than selected breeds (Pugliese and Sirtori, 2012). Moreover, their
65 phenotypic traits resulting from their genetic potential can be modulated by rearing
66 factors such as final slaughter weight and age, feeding, climatic conditions, space
67 allowance and physical exercise (Rosenvold and Andersen, 2003; Leuret, 2008).
68 The local Basque breed reared in extensive conditions and slaughtered at ca. 145 kg
69 body weight (BW) in southwestern France is an example of a diversified pork chain
70 that results in high-quality and typical products recognized by the Protected
71 Designation of Origin label (Mercat *et al.*, 2019). Among muscle biological
72 characteristics that confer high quality, intramuscular fat (IMF) content is an important
73 trait that should reach 2-3% of fresh muscle to achieve satisfactory pork tenderness
74 and juiciness and acceptable visual perception by consumers (review by Lustrat *et al.*,
75 2016). However, the association between IMF content and pork quality is not always

76 reported due to the influence of many other factors, such as the post-mortem
77 decrease in pH, aging conditions and cooking methods. Muscle fiber type
78 composition also influences pork quality through its effects on pH, water holding
79 capacity, color and muscle microstructure (review by Lefaucheur, 2010). Increasing
80 the percentage of fast-twitch glycolytic fibers in pig longissimus muscle (LM)
81 increases the rate and extent of the pH decrease and meat lightness, and decreases
82 protein solubility and water holding capacity. In contrast, a higher percentage of slow-
83 twitch oxidative fibers increases pork flavor, tenderness and redness (Kang *et al.*,
84 2011). In addition, increasing the cross-sectional area (CSA) of muscle fiber is widely
85 reported to decrease water holding capacity and pork tenderness (Lefaucheur,
86 2010). Fiber-type composition and IMF content differ greatly among muscles and are
87 influenced by genetic, animal (e.g. age) and environmental factors, but relationships
88 between the two traits remain controversial (Lefaucheur, 2010). It is commonly stated
89 that oxidative muscles contain more IMF than glycolytic muscles (Bereta *et al.*, 2014;
90 Jeong *et al.*, 2017); however, other studies do not support this assessment (Kang *et*
91 *al.*, 2011).

92 Lebret *et al.* (2015) observed that pigs of the local, non-selected Basque (B) breed
93 had higher quality pork than pigs of the conventional Large White (LW) breed. The
94 objective of the present study was to determine the biological properties of muscle
95 that explained this difference. We hypothesized that B pigs have higher muscle
96 oxidative metabolism and IMF content and lower fiber CSA than LW pigs. Since
97 outdoor or free-range rearing of pigs can increase muscles' oxidative metabolism and
98 influence their energy stores (i.e. IMF content and glycolytic potential (GP), which
99 can influence pork quality (Bee *et al.*, 2004; Gentry *et al.*, 2004)), we also
100 hypothesized that biological properties of B and LW pig muscles would differ

101 depending on their rearing conditions. Therefore, we aimed to determine the
102 biochemical, metabolic and histological traits of loin and ham muscles that influence
103 the pork quality of B and LW pigs reared in a conventional indoor, alternative (indoor
104 bedding and outdoor area) or extensive free-range system (B pigs only), with the
105 ultimate aim to identify breeding and rearing practices that improve pork quality.

106

107 **Materials and methods**

108

109 *Animals and experimental design*

110 Lebret *et al.* (2014, 2015) described the experimental design in detail. Briefly, 100
111 castrated male pigs from the pure local B (n = 60) or LW breed (n = 40) were used.
112 The B pigs came from two breeding farms of the Basque pork chain (Mercat *et al.*,
113 2019), while the LW pigs came from the INRA experimental herd. All animals were
114 free of the RYR1 and PRKAG3 mutated alleles. Pigs were produced in two
115 experimental replicates (R1 and R2), each including 30 B and 20 LW pigs. In each
116 replicate, at a mean BW of 35 kg, 20 B littermates (from 10 B litters each in R1 and
117 R2) and 20 LW littermates (from 10 and 7 LW litters in R1 and R2, respectively) were
118 placed in a conventional (C, one pen per breed, indoor slatted floor, 1.0 m²/pig) or
119 alternative housing system (A, one pen per breed, indoor bedding and free access to
120 an outdoor area, 2.4 m²/pig), until slaughter at ca. 145 kg BW. LWC, LWA, BC and
121 BA pigs (n = 10 (one pen) per treatment and replicate) were fed the same standard
122 growing (2.29 Mcal/kg net energy, 18.0% CP, 3.40% crude fat, 0.83% digestible
123 lysine; from 35 up to 75 kg BW) and finishing (2.14 Mcal/kg net energy, 14.7% CP,
124 1.60% crude fat, 0.75% digestible lysine ; from 75 kg BW to slaughter) diets. Pigs
125 were fed *ad libitum* until the average feed intake of 2.5 kg/d and per pig was reached,

126 corresponding approximately to 75 kg BW in each treatment. Then, the daily feed
127 allowance was progressively increased up to 3.0 kg/d and per pig up to 110 kg BW
128 and maintained at 3.0 kg/d and per pig until 145 kg BW. Pigs were weighed at the
129 start, during and at the end of the experiment, and average daily gain (ADG) was
130 calculated individually. Feed consumption per pen was recorded weekly, and
131 average daily feed intake (ADFI) and feed efficiency were calculated per pen for each
132 treatment and replicate. LWC, LWA, BC and BA pigs were slaughtered at the INRA
133 slaughterhouse in four sessions, each including pigs from the four treatments.
134 In addition, in each replicate, 10 B pigs, half-littermates of BC and BA pigs, were
135 placed at 35 kg BW in an extensive production system (E, outdoor free-range
136 system, 2.5 ha/pig) on a Basque chain farm, until slaughter at ca. 145 kg BW. BE
137 pigs had free access to natural food resources (acorns and chestnuts) and were fed
138 a standard supplementary growing-finishing diet (2.15 Mcal/kg net energy, 15.5%
139 CP, 1.95% crude fat, 0.70% digestible lysine) according to local farming practices
140 (allowance of 1.4 kg/d up to 2.2 kg/d and per pig from 35 kg up to 75 kg BW; 2.2 kg/d
141 up to 2.6 kg/d and per pig from 75 kg up to 110 kg BW; 2.6 kg/d to 2.3 kg/d and per
142 pig from 110 kg up to 130 kg BW; and 2.0 kg/d and per pig from 130 kg until
143 slaughter at around 145 kg BW; Lebret *et al.*, 2014; Mercat *et al.*, 2019). BE pigs
144 were weighed individually to calculate ADG. The ADFI and feed efficiency of BE pigs
145 could not be calculated because their feed consumption, including both the
146 supplementary diet and the natural food resources, could not be determined. BE pigs
147 were included in the experiment 5 months, and BC and BA pigs 3 months, before
148 LWC and LWA pigs in order to slaughter all pigs at 145 kg BW during winter in
149 January (R1) or February (R2). BE pigs were slaughtered in a commercial

150 slaughterhouse (Saint-Jean-Pied-de-Port, France). Slaughtering conditions of the two
151 slaughterhouses used in the experiment were standardized as much as possible.

152

153 *Carcass and pork quality traits*

154 According to the handling practices of the Basque pork chain, all pigs were
155 slaughtered after 36 h of fasting by exsanguination after electrical stunning (350 V, 4
156 A), in compliance with national regulations for slaughterhouses. Weights of the hot
157 carcass and cold half-side carcass, as well as the loin and back fat primal cuts, were
158 recorded. Pork quality traits were measured for all animals in the loin (LM, first
159 lumbar vertebra level), as presented by Lebret *et al.* (2015), and in the ham
160 semimembranosus muscle (SM). Measurements included IMF content, GP, ultimate
161 pH (pHu, 24 h post mortem), lightness and hue angle. The semitendinosus muscle
162 (STM) was completely excised from the LWC, LWA, BC and BA pigs of R1 (n = 40)
163 and weighed. Since the STM is heterogeneous, with a deep red (RSTM) and a
164 superficial white (WSTM) portion (Figure 1; Listrat *et al.*, 2016), the IMF content and
165 pHu of both portions were measured. See Supplementary Material S1 for details of
166 the methods.

167

168 *Metabolic enzyme activities*

169 Enzyme activities were measured in the LM and SM of all R1 and R2 pigs, and in the
170 RSTM and WSTM of LWC, LWA, BC and BA pigs of R1. Within 40 min post mortem,
171 muscle samples were taken from the LM (last rib level), SM (external part, 2 cm
172 deep) and central portions of the RSTM and WSTM; cut into small pieces; promptly
173 frozen in liquid nitrogen and stored at -80°C. Activities of lactate dehydrogenase
174 (LDH), citrate synthase (CS) and β -hydroxy-acyl-CoA dehydrogenase (HAD) were

175 determined as markers of glycolytic metabolism, oxidative capacity (tricarboxylic acid
176 cycle) and lipid β -oxidation potential, respectively, as described by Lebret *et al.*
177 (2002) and detailed in Supplementary Material S1.

178

179 *Histological analyses*

180 Muscle fibers were typed in the LM of all R1 and R2 pigs and in the RSTM and
181 WSTM of LWC, LWA, BC and BA pigs of R1. From the samples taken to measure
182 enzyme activities, sub-samples parallel to the muscle fiber axis were promptly frozen
183 in isopentane cooled by liquid nitrogen and stored at -80°C until analysis, performed
184 as described by Larzul *et al.* (1997) and detailed in Supplementary Material S1.
185 Briefly, 10 μm thick transverse serial sections were cut in a cryostat (-20°C), mounted
186 on glass slides and stained for acto-myosin ATPase after preincubation at pH 4.35 to
187 distinguish contractile types I, IIA and IIB, or stained for succino-dehydrogenase to
188 identify metabolic red oxidative (R) and white glycolytic (W) fibers. Combining both
189 stains, fibers were classified as type I (slow-twitch oxidative), IIA (fast-twitch oxido-
190 glycolytic), IIBR (fast-twitch oxido-glycolytic) or IIBW (fast-twitch glycolytic). From
191 each sample, four fields containing ca. 250 fibers each were randomly chosen to
192 determine the percentage, CSA and relative area of each fiber type using digital
193 image analysis. The total number of fibers (TNF) in the RSTM and WSTM was
194 determined by multiplying the mean number of fibers per unit area (calculated from
195 three fields per sample using a projection microscope) by the transverse area of each
196 portion (determined as described in Supplementary material S1).

197

198 *Statistical analyses*

199 Statistical Analysis System (SAS) software (version 9.4, 2013, SAS Institute, Cary,
200 NC, USA) was used to analyze the data (details in Supplementary Material S1). Data
201 for the ADFI and feed efficiency of LWA, LWC, BA and BC pigs, for which the pen
202 was considered the statistical unit, were analyzed using ANOVA (GLM procedure),
203 considering the treatment (4 levels) and replicate (2 levels) as fixed effects. Contrasts
204 between breeds and between the A and C systems for a given breed were
205 determined (Lebret *et al.*, 2014). For all other traits, the animal was considered the
206 statistical unit. First, data were analyzed using ANOVA (GLM procedure), considering
207 the treatment (LWC, LWA, BC, BA and BE) and replicate (R1 and R2) as fixed
208 effects to calculate residues. The normality of residues was checked (Shapiro-Wilk
209 test, $P \geq 0.05$). When necessary, data were log- or square-root transformed to obtain
210 a normal distribution of residues, which were checked using the same test.
211 Subsequently, contrasts between breeds were determined from the ANOVA using
212 only the LWC, LWA, BC and BA pigs, to balance the production systems between
213 breeds. Within each breed, contrasts between rearing systems were determined to
214 evaluate effects of the A vs. C system for the LW breed, and the A vs. C and E vs. C
215 systems for the B breed. Means were calculated by treatment. When residues could
216 not be normalized, a non-parametric method (NPAR1WAY procedure, Kruskal-Wallis
217 test) was used to determine effects of the breed and of the rearing system within a
218 given breed; their medians were calculated by treatment.

219

220 **Results**

221

222 *Growth performance and carcass traits*

223 As previously reported in more detail (Lebret *et al.*, 2014), B pigs had lower ADG
224 than LW pigs (Table 1). ADG did not differ between LWC and LWA pigs, but was
225 higher for BA pigs and lower for BE pigs than for BC pigs. Consequently, B pigs,
226 especially BE, were older than LW pigs at slaughter. LW pigs had higher ADFI and
227 feed efficiency than B pigs. LWA pigs had similar ADFI as, but lower feed efficiency
228 than, LWC pigs, while BA pigs had higher ADFI than, but similar feed efficiency as,
229 BC pigs. B pigs had fatter carcasses, with higher back fat and lower loin proportions,
230 than LW pigs. Compared to the C system, the A system increased the loin proportion
231 in LW pigs, but did not influence proportions of carcass cuts of B pigs. In contrast, the
232 E system decreased the carcass fatness of B pigs.

233

234 *Biochemical and quality traits of longissimus and semimembranosus muscles*

235 B pigs had higher IMF content in both the LM and SM than LW pigs (Table 2). No
236 significant differences in IMF content between A and C systems were observed
237 within each breed, or between E and C systems within the B breed. B pigs had
238 higher pHu and lower GP, lightness and hue angle (indicating redder meat) in both
239 muscles than LW pigs. Within each breed, the A vs. C system did not influence pHu,
240 GP, lightness or hue angle. In contrast, BE pigs had higher pHu and lower lightness
241 in both muscles than BC pigs. BE pigs had lower GP in the SM than BC pigs, but
242 they did not differ in the LM.

243 Breed effects on metabolic enzyme activities differed between muscles (Table 3). B
244 pigs had lower CS and HAD activities in the LM than LW pigs, while the opposite was
245 observed in the SM. LDH activity in the LM did not differ between breeds but was
246 lower in the SM of B than LW pigs. Within each breed, compared to the C system,
247 the A system increased CS activity in both muscles and HAD activity in the SM of LW

248 pigs, but did not influence LDH activity in either muscle. BE pigs had higher CS and
249 HAD activities than, but similar LDH activity as, BC pigs in the LM. In contrast, BE
250 pigs had lower LDH activity than, but similar CS and HAD activities as, BC pigs in the
251 SM.

252

253 *Histological properties of the longissimus muscle*

254 B pigs had a lower percentage of IIA fibers and a higher percentage of IIBW fibers in
255 the LM than LW pigs, but they did not differ in the percentages of I and IIBR fibers
256 (Table 4). B pigs had lower mean fiber CSA than LW pigs, but differences varied
257 among fiber types. B pigs had smaller IIBW fibers and larger I fibers than, and similar
258 CSA of IIA and IIBR fibers as, LW pigs. Only the relative area of I fibers was higher in
259 B pigs than in LW pigs. The influence of the A vs. C system on fiber types varied by
260 breed. LWA pigs had higher IIBR and lower IIBW percentages than LWC pigs, while
261 BA pigs had a higher IIA percentage than BC pigs. The A vs. C system did not
262 influence fiber CSA in either breed, but resulted in a lower relative area of IIBW fibers
263 in LW pigs and a higher relative area of IIA fibers in B pigs. Compared to BC pigs, BE
264 pigs had a larger CSA of I fibers and a higher percentage and larger relative area of
265 IIA fibers at the expense of IIBW fibers.

266

267 *Biochemical, quality traits and histological characteristics of the semitendinosus* 268 *muscle*

269 B pigs had a lower proportion of STM in the half-carcass than LW pigs, but the
270 proportion of STM did not differ between A and C systems within each breed (Table
271 5). B pigs had higher IMF content (Table 5) and visual marbling (Figure 1) in both the
272 RSTM and WSTM than LW pigs, especially in the WSTM. B pigs had higher pHu in

273 both STM portions than LW pigs, with generally higher pHu in the RSTM than in the
274 WSTM. Breed did not influence CS or HAD activities in the RSTM or WSTM, but B
275 pigs had higher LDH activity in the RSTM than LW pigs. Regardless of the breed, the
276 A and C systems did not differ in enzyme activities, IMF content or pHu in the RSTM
277 or WSTM.

278

279 *Fiber type composition of the semitendinosus muscle*

280 B pigs had 16% smaller TNF in the STM than LW pigs, exclusively due to a smaller
281 TNF in the WSTM (Table 6). TNF did not differ between the A and C systems in
282 either breed. Breed differences in fiber types varied by STM portion. B pigs had a
283 lower percentage of I fibers in the RSTM than LW pigs but higher percentages of
284 IIBR and IIBW fibers. In contrast, B pigs had a higher percentage of I fibers in the
285 WSTM than LW pigs but similar percentages of IIA, IIBR and IIBW fibers. B pigs had
286 smaller mean fiber CSA in the RSTM and WSTM than LW pigs, due to a smaller CSA
287 of all fiber types in the RSTM, but only of IIB fibers in the WSTM. Compared to LW
288 pigs, B pigs had a larger relative area of IIBR at the expense of IIA fibers in the
289 RSTM, and of I fibers at the expense of IIB fibers in the WSTM. No effect of the A vs.
290 C system was observed on fiber type percentages in either the STM portion or breed.
291 The A system resulted in smaller CSA of all fiber types in the RSTM of LW pigs, with
292 no effect in B pigs. Fiber CSA in the WSTM did not differ between A and C systems
293 regardless of fiber type or breed. Overall, relative areas of fiber types did not differ
294 between A and C systems in the RSTM or WSTM.

295

296 **Discussion**

297 Unlike the local B breed, which has not experienced genetic selection, the LW breed
298 has been selected for a high growth rate, feed efficiency and carcass leanness,
299 which likely indirectly resulted in selecting animals with a larger adult body size.
300 Consequently, at a given BW, B pigs are older and fatter (Lebret *et al.*, 2014), and
301 closer to their adult size, meaning that they are physiologically more mature than LW
302 pigs. Muscle mass depends on the TNF, CSA and length of fibers. Since the TNF in
303 pigs is established by the end of the fetal period (review by Lefaucheur, 2010), post-
304 natal muscles grow exclusively by increasing both the CSA and length of fibers. In
305 our study, the lower muscle mass of B pigs than of LW pigs was associated with both
306 smaller TNF and mean fiber CSA, indicating that the smaller TNF in B pigs was not
307 compensated by larger fiber CSA at the same slaughter weight. Similarly, smaller
308 TNF and fiber CSA were reported in Meishan, a Chinese breed with low muscle
309 growth potential, than in the LW breed (Lefaucheur *et al.*, 2004). Gil *et al.* (2008)
310 made the same observation when comparing other local pig breeds to selected pig
311 breeds. Smaller CSA of IIB fibers and larger relative area of I fibers have been
312 associated with higher pork quality, especially water holding capacity and tenderness
313 (Lefaucheur, 2010). These histological properties observed in the LM of B pigs but
314 not LW pigs may contribute to the higher eating quality of B pork (Lebret *et al.*, 2015).
315 Within each breed, the lack of difference in the mean fiber CSA in the LM and WSTM
316 between the A and C systems agrees with the systems' relatively weak effects on
317 body composition. Similarly, the mean CSA in the LM and loin proportion in B pigs
318 did not differ between the E and C systems. However, the smaller CSA of all fiber
319 types in the RSTM of LWA pigs than of LWC pigs was not expected and requires
320 further study. Unfortunately, the STM characteristics of BE pigs could not be
321 assessed in the present study.

322 Pork quality traits are influenced by genetic factors, environmental factors and
323 slaughter conditions (Rosenvold and Andersen, 2003), and higher quality is generally
324 reported in local rather than “conventional” selected pig breeds (review by Bonneau
325 and Lebret, 2010). The LM and SM of B pigs had lower GP, higher pHu and IMF
326 content, lower lightness and a redder color than those of LW pigs, which explains
327 their higher eating and technological quality (Lebret *et al.*, 2015). Based on
328 descriptions of differences in muscle metabolism between wild or non-selected pigs
329 and selected pigs (Wimmers *et al.*, 2008; Lefaucheur, 2010), B pigs were expected to
330 have lower glycolytic and/or higher oxidative metabolism than LW pigs. These
331 differences were observed in the SM, in which B pigs had higher CS and HAD
332 activities and lower LDH activity than LW pigs, which is consistent with their lower
333 GP, higher pHu and redder meat. In contrast, the LM of B pigs had less oxidative
334 metabolism, probably due to their older age and greater maturity at a given BW,
335 since fast-oxido-glycolytic fibers in the LM are converted into fast-glycolytic fibers as
336 pigs age (Lefaucheur and Vigneron, 1986). Because type I fibers have the lowest
337 glycogen content (Lefaucheur, 2010), their larger relative area in the LM of B pigs
338 than in LW pigs can partly explain their lower GP. The redder meat in the LM and SM
339 of B pigs could be due to their higher muscle myoglobin content with age (Mayoral *et al.*
340 *et al.*, 1999). The percentage of I fibers was similar in the LM of B and LW pigs, in
341 agreement with lack of difference for this trait between Meishan and LW pigs
342 (Lefaucheur *et al.*, 2004). However, other studies reported a higher percentage of I
343 fibers in the LM of wild pigs or other local breeds (Ruusunen and Puolanne, 2004;
344 Wimmers *et al.*, 2008). These discrepancies could be explained by differences in
345 physical exercise, climatic conditions, BW, age and physiological maturity between
346 the animals in specific studies. As in the LM and SM, B pigs had higher pHu in the

347 RSTM and WSTM than LW pigs. This difference was associated with a higher
348 percentage of I fibers in the WSTM, and, surprisingly, with a higher glycolytic
349 metabolism in the RSTM of B pigs than LW pigs, suggesting that different portions of
350 the same muscle can be influenced differently by breed. The relative areas of the two
351 STM portions may also have differed between breeds, but they could not be studied
352 due to the difficulty in delineating the portions (Figure 1).

353 Overall, breed influenced the biological properties of muscle and pork quality traits
354 more than the rearing system. Within each breed, the A and C systems did not differ
355 in the IMF content, GP, pHu, lightness or hue angle in the LM and SM, in agreement
356 with studies that showed no consistent effect of indoor bedding or free access to a
357 small outdoor area on these traits (Lebret *et al.*, 2002; Millet *et al.*, 2005). Although
358 the rearing system yielded similar quality traits, it influenced the muscle metabolism
359 of LW pigs, with higher oxidative metabolism in the LM and SM of LWA pigs than of
360 LWC pigs. These results agree with the shift from fast glycolytic to fast oxido-
361 glycolytic fibers in the LM of pigs offered access to an outdoor area (Lebret *et al.*,
362 2002) and in the SM of free-range pigs during winter, unlike those of pigs housed
363 indoors (Gentry *et al.*, 2004). No such effects were observed in the LM and SM of B
364 pigs, or in the RSTM and WSTM of either breed, suggesting greater robustness of
365 muscle properties of B pigs to environmental factors. Comparing winter free-range to
366 indoor housing of Swiss LW pigs, Bee *et al.* (2004) also observed higher oxidative
367 metabolism in the LM but no differences in the RSTM or WSTM. Petersen *et al.*
368 (1998) observed higher oxidative metabolism in the STM of pigs physically trained or
369 reared in large pens, but they did not distinguish the RSTM and WSTM portions.

370 Both physical exercise and cold exposure, like for pigs in the A system (Lebret *et al.*,
371 2014), have been shown to increase muscle oxidative metabolism (Lefaucheur,

2010). In the present study, however, they were not sufficient to modify the metabolic or histological traits of the RSTM or WSTM in either breed.

Among the rearing systems considered, the E system of the Basque pork chain induced the strongest effects on muscle and pork traits. The higher pHu and redness and lower lightness of the LM and SM of BE pigs than of BC pigs agree with the higher oxidative metabolism in the LM and lower glycolytic metabolism in the SM. These results are consistent with the increased IIA:IIB fiber ratio found in the LM of free-range pigs (Petersen *et al.*, 1998; Bee *et al.*, 2004; Gentry *et al.*, 2004). Because BE pigs were older, have more physical activity and encounter colder temperatures, higher oxidative capacity was also expected in their SM, which is involved in locomotion, but this increase was not observed. The redder meat of BE pigs would thus be more likely explained by their greater age at slaughter, which induces higher myoglobin content (Mayoral *et al.*, 1999). The higher pHu in BE pigs than in BC pigs was logically accompanied by a lower GP in the SM, but not in the LM, confirming that muscle properties other than GP determine the pHu (Scheffler *et al.*, 2013).

Unlike our results, extensive pig rearing during winter has been shown to increase GP, especially in muscles involved in movement, such as the SM (Lebret, 2008). This difference may be due to higher pre-slaughter physical activity of BE pigs than BC pigs, despite the high standardization of pre-slaughter handling in the experiment (Lebret *et al.*, 2015).

B pigs had a higher IMF content in all muscles, regardless of their contractile and metabolic profiles, than LW pigs, in agreement with the positive correlation between the IMF content in different muscles in pigs (Quintanilla *et al.*, 2011). Interestingly, the IMF content in both breeds was higher in the glycolytic WSTM than in the oxidative RSTM, with stronger breed differences in the WSTM. Differences in IMF

397 content between Duroc, a breed with high IMF content, and the LW breed were also
398 stronger in the LM than in the more oxidative psoas major muscle (Wood *et al.*,
399 2004). Assessing relationships between IMF content and metabolic enzyme
400 activities, the percentage of type I fibers or mean CSA in the LM, WSTM and RSTM,
401 revealed no significant correlation between IMF content and these traits, either
402 between or within muscles (Figure 2). Similarly, IMF content has been shown to be
403 genetically unrelated to fiber type composition in the LM (Larzul *et al.*, 1997),
404 suggesting that IMF content and muscular contractile and metabolic characteristics
405 are unrelated traits that can be manipulated separately by breeding or rearing
406 factors.

408 **Conclusion**

409 Overall, effects of breed on pork quality and muscle biochemical and histological
410 characteristics were greater than those of the rearing system, among which the E
411 system induced the greatest changes. Both smaller TNF and CSA of glycolytic fibers
412 were associated with the lower muscle mass of B pigs than of LW pigs. In all
413 muscles, B pigs had higher IMF content and pHu and smaller mean fiber CSA than
414 LW pigs, which likely explains the former's higher pork quality. The expected shift
415 towards a less glycolytic and more oxidative muscle metabolism in B pigs than in LW
416 pigs was observed in the SM and, to a lesser extent, the WSTM, but not in the LM or
417 RSTM, which were more oxidative in LW pigs. Interestingly, the glycolytic WSTM
418 contained more IMF than the oxidative RSTM, especially in B pigs, supporting the
419 independence between IMF content and contractile and metabolic muscle properties.

420

421 **Acknowledgements**

422 The authors thank the staff of the INRA experimental farm, slaughterhouse and
423 laboratories, especially Patrick Ecolan and Nathalie Bonhomme, and the breeders
424 and slaughterhouse staff of the Basque pork chain.

425

426 **Declaration of interest**

427 None.

428

429 **Ethics statement**

430 All experimental procedures complied with European Union (Directive 86/609/CEE)
431 and French legislation (Décret no. 2001-464 29/05/01). INRA UEPR held the pig
432 experimentation agreement C-35–275–32. All technical and scientific staff involved in
433 the experiment had an individual agreement for experimenting on living animals,
434 provided by the Veterinary Services of the French Ministry of Agriculture.

435

436 **Software and data repository resources**

437 None of the datasets were deposited in an official repository.

438

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521

522 **Table 1.** Growth performance and carcass traits by pig breed (Large White, LW; Basque, B) and rearing system
 523 (Conventional, C; Alternative, A; Extensive, E) (Lebret *et al.*, 2014)

	Treatment: breed x rearing system ¹					Significance ²					
						Breed		Rearing system within breed			
	LWC	LWA	BC	BA	BE	RMSE	B vs. LW	LW A vs. C	B A vs. C	B C	B E vs. C
N of pigs	20	19	20	20	20						
N of pens	2	2	2	2	2						
Initial live weight (kg) ³	38.5	37.9	34.8	35.3	35.7	5.55	0.017	0.71	0.81	0.64	
Initial age (d) ⁴	85	85	106	106	106		<0.001	0.79	0.87	0.91	
Final live weight (kg) ³	148.0	144.8	139.9	146.3	141.8	8.81	0.10	0.24	0.024	0.50	
Final age (d) ⁴	228	230	320	312	423		<0.001	0.84	0.24	<0.001	
ADG (g/d) ^{4,5}	772	755	498	544	335		<0.001	0.61	0.040	<0.001	
ADFI (kg/d) ^{3,5}	2.88	2.88	2.39	2.67	-	0.047	0.002	0.99	0.009	-	
Feed efficiency (kg/kg) ^{3,5}	0.27	0.25	0.21	0.20	-	0.003	<0.001	0.033	0.14	-	
Hot carcass weight (kg) ³	118.2	113.8	114.2	118.7	113.5	7.29	0.75	0.065	0.052	0.84	

Loin (%) ^{3,6}	22.8	23.7	17.7	18.0	17.7	0.89	<0.001	0.002	0.34	0.84
Back fat (%) ^{3,6}	7.9	8.0	15.2	15.7	11.3	1.37	<0.001	0.68	0.45	<0.001

524 ¹Mean, or median when a non-parametric test was performed.

525 ²*P*-values of contrasts between breeds (determined using A and C pigs of both breeds, i.e. n=39 LW and n=40 B) or rearing systems within each
526 breed and RMSE obtained from ANOVA or *P*-values of a non-parametric test when data could not be normalized.

527 ³ANOVA of raw data.

528 ⁴Non-parametric test.

529 ⁵ADG: Average daily gain; ADFI: Average daily feed intake; Feed efficiency: weight gain:feed intake

530 ⁶Proportion of carcass right side weight.

531 **Table 2.** Biochemical characteristics and meat quality traits of the longissimus (LM) and semimembranosus (SM) muscles
 532 of pigs by breed (Large White, LW; Basque, B) and rearing system (Conventional, C; Alternative, A; Extensive, E)

	Treatment: breed x rearing system ¹					Significance ²				
						Breed		Rearing system within breed		
	LWC	LWA	BC	BA	BE	RMSE	B vs. LW	LW A vs. C	B A vs. C	B E vs. C
N	20	19	20	19	20					
Intramuscular fat (%) ³										
LM	2.32	2.14	3.79	4.07	3.28	0.117	<0.001	0.34	0.60	0.11
SM	2.21	1.98	3.91	4.00	3.80	0.126	<0.001	0.24	0.78	0.76
Glycolytic potential (μmol eq. lactate/g)										
LM ⁴	164	173	136	138	136	18.9	<0.001	0.16	0.69	0.99
SM ⁵	164	166	137	145	128		<0.001	0.86	0.57	0.015
pH 24 h ⁵										
LM	5.47	5.48	5.58	5.54	5.67		<0.001	0.94	0.40	0.005
SM	5.50	5.48	5.55	5.59	5.75		<0.001	0.41	0.51	<0.001
Lightness ⁴										
LM	53.6	53.8	51.2	51.6	48.1	2.74	<0.001	0.86	0.68	<0.001

SM	52.2	51.9	47.3	47.0	44.7	2.90	<0.001	0.80	0.76	0.006
Hue angle ⁴										
LM	37.7	38.3	34.5	35.3	27.8	2.80	<0.001	0.53	0.36	<0.001
SM	36.1	35.7	29.6	29.2	24.8	2.50	<0.001	0.66	0.58	<0.001

533 ¹Mean, or median when a non-parametric test was performed.

534 ²*P*-values of contrasts between breeds (determined using A and C pigs of both breeds, i.e. n=39 LW and n=39 B) or rearing systems within each
535 breed and RMSE obtained from ANOVA or *P*-values of a non-parametric test when data could not be normalized.

536 ³ANOVA of log values to fit a normal distribution.

537 ⁴ANOVA of raw data.

538 ⁵Non-parametric test.

539 **Table 3.** Metabolic enzyme activities¹ of lactate dehydrogenase (LDH), citrate synthase (CS) and β -hydroxy-acyl-Co-A
 540 dehydrogenase (HAD) in longissimus (LM) and semimembranosus (SM) muscles of pigs by breed (Large White, LW;
 541 Basque, B) and rearing system (Conventional, C; Alternative, A; Extensive, E)

	Treatment: breed x rearing system ²					RMSE	Significance ³			
							Breed		Rearing system within breed	
	LWC	LWA	BC	BA	BE		B vs. LW	LW A vs. C	B A vs. C	B E vs. C
N	20	19	20	20	20					
LDH										
LM ⁴	2663	2645	2603	2605	2595	0.1	0.24	0.77	0.97	0.90
SM ⁴	2608	2556	2321	2269	2191	0.1	<0.001	0.49	0.43	0.047
CS										
LM ⁴	5.63	6.34	4.94	5.33	6.22	0.067	<0.001	0.025	0.14	<0.001
SM ⁴	8.72	9.95	10.67	11.36	10.66	0.067	<0.001	0.010	0.21	0.98
HAD										
LM ⁴	3.38	3.62	2.97	3.12	3.55	0.074	<0.001	0.23	0.37	0.002
SM ⁴	4.77	5.35	6.04	6.42	6.26	0.071	<0.001	0.030	0.24	0.49

542 ¹Expressed as micromol of substrate per min per g of fresh muscle.

543 ²Mean of treatment groups.

544 ³P-values of contrasts between breeds (determined using A and C pigs of both breeds, i.e. n=39 LW and n=40 B) or rearing systems within each

545 breed and RMSE obtained from ANOVA.

546 ⁴ANOVA of log values to fit a normal distribution.

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547 **Table 4.** Histochemical characteristics of longissimus muscle by pig breed (Large White, LW; Basque, B) and rearing
 548 system (Conventional, C; Alternative, A; Extensive, E)

	Treatment: breed × rearing system ¹					Significance ²				
						Breed		Rearing system within breed		
	LWC	LWA	BC	BA	BE	RMSE	B vs. LW	LW	B	B
							A vs. C	A vs. C	E vs. C	
N	20	19	20	20	20					
Fiber percentage (%) ^{3,4}										
I	8.7	10.1	9.0	8.0	10.1	3.04	0.20	0.15	0.30	0.28
IIA	7.8	8.8	4.6	6.7	6.6	2.18	<0.001	0.19	0.005	0.006
IIBR	5.7	7.4	6.1	6.3	6.6	2.66	0.54	0.045	0.86	0.56
IIBW	77.7	73.6	80.2	79.0	76.6	3.43	<0.001	<0.001	0.27	0.001
Cross-sectional area (μm ²) ^{3,5}										
I	3026	2767	3161	3529	3586	0.1	0.002	0.17	0.085	0.049
IIA	2257	2122	2270	2583	2513	0.1	0.066	0.43	0.093	0.18
IIBR	3115	2863	2859	3168	2947	0.1	0.91	0.39	0.29	0.76
IIBW	5514	5300	4343	4565	4823	0.1	<0.001	0.50	0.40	0.076

Mean	4841	4500	4029	4238	4389	0.1	0.003	0.20	0.37	0.13
Relative area (%) ^{3, 6}										
I	5.1	6.0	6.8	6.5	8.0	0.39	0.017	0.17	0.61	0.087
IIA	3.6	3.9	2.5	4.0	3.6	0.32	0.055	0.43	<0.001	0.002
IIBR	3.5	4.6	4.2	4.5	4.4	0.53	0.52	0.12	0.63	0.81
IIBW	87.3	85.0	85.9	84.5	83.3	0.14	0.12	0.007	0.12	0.003

549 ¹Mean of treatment groups.

550 ²P-values of contrasts between breeds (determined using A and C pigs of both breeds, i.e. n=39 LW and n=40 B) or rearing systems within each
551 breed and RMSE obtained from ANOVA.

552 ³ Fibers I : slow-twitch oxidative, IIA : fast-twitch oxido-glycolytic, IIBR : fast-twitch oxido-glycolytic, IIBW : fast-twitch glycolytic.

553 ⁴ANOVA of raw data.

554 ⁵ANOVA of log values to fit a normal distribution.

555 ⁶ANOVA of square root values to fit a normal distribution.

556 **Table 5.** Physicochemical characteristics and metabolic enzyme activities of lactate dehydrogenase (LDH), citrate
 557 synthase (CS) and β -hydroxy-acyl-CoA dehydrogenase (HAD) in the red (RSTM) and white (WSTM) portions of
 558 semitendinosus muscle (STM) by pig breed (Large White, LW; Basque, B) and rearing system (Conventional, C;
 559 Alternative, A; Extensive, E)

	Treatment: breed \times rearing system ¹				RMSE	Significance ²		
						Breed B vs. LW	Rearing system within breed	
	LWC	LWA	BC	BA			LW A vs. C	B A vs. C
N	10	10	10	10				
STM (%) ^{3,4}	1.14	1.13	0.68	0.69	0.040	<0.001	0.71	0.93
RSTM								
Intramuscular fat (%) ⁴	3.71	4.14	5.58	5.37	0.087	<0.001	0.23	0.67
pH 24 h ⁵	5.85	5.70	5.91	6.08	0.198	0.001	0.089	0.069
LDH ^{4,6}	815	791	927	899	0.1	0.004	0.61	0.60
CS ^{4,6}	19.6	19.7	21.1	19.7	0.06	0.37	0.92	0.27
HAD ^{4,6}	14.6	15.2	15.7	14.4	0.07	0.86	0.57	0.24
WSTM								

Intramuscular fat (%) ⁴	6.28	5.20	16.0	15.2	0.11	<0.001	0.10	0.66
pH 24 h ⁵	5.55	5.49	5.75	5.72	0.083	<0.001	0.13	0.41
LDH ^{4,6}	1463	1365	1534	1421	0.1	0.18	0.13	0.097
CS ^{4,6}	5.45	5.55	6.16	5.37	0.093	0.51	0.86	0.16
HAD ^{4,6}	3.07	3.01	3.49	3.09	0.091	0.25	0.82	0.20

560 ¹Mean of treatment groups.

561 ²P-values of contrasts between breeds or rearing systems within each breed and RMSE obtained from ANOVA.

562 ³Proportion of right carcass side weight.

563 ⁴ANOVA of log values to fit a normal distribution.

564 ⁵ANOVA of raw data.

565 ⁶Expressed as micromol of substrate per min per g of fresh muscle.

566 **Table 6.** Histochemical characteristics of the red (RSTM) and white (WSTM) portions
 567 of semitendinosus muscle by pig breed (Large White, LW; Basque, B) and rearing
 568 system (Conventional, C; Alternative, A)

	Treatment: breed × rearing system ¹				Significance ²			
					RMSE	Breed	Rearing system within breed	
	LWC	LWA	BC	BA		B vs. LW	LW A vs. C	B A vs. C
N	10	10	10	10				
TNF (10 ⁻³) ^{3,4}	610	642	520	526	0.1	0.004	0.54	0.90
RSTM								
TNF (10 ⁻³) ^{3,4}	231	248	229	225	0.1	0.53	0.54	0.88
Fiber percentage (%) ^{5, 6}								
I	45.4	46.1	39.0	40.8	6.51	0.007	0.80	0.54
IIA	23.1	22.1	19.3	20.4	4.82	0.080	0.63	0.60
IIBR	10.7	9.8	17.5	14.2	6.87	0.013	0.77	0.33
IIBW	20.8	22.0	24.2	24.3	4.37	0.042	0.54	0.95
Cross-sectional area (μm ²) ^{4, 5}								
I	5719	4427	4391	4261	0.1	0.005	<0.001	0.67
IIA	4314	3848	3251	3186	0.1	<0.001	0.071	0.74
IIBR	5467	4516	3583	3651	0.1	<0.001	0.020	0.81
IIBW	7565	6284	4745	4746	0.1	<0.001	0.026	0.99
Mean	5457	4484	3940	3917	0.1	<0.001	0.003	0.93
Relative area (%) ^{5, 7}								
I	45.0	43.3	41.4	42.5	0.41	0.19	0.49	0.64
IIA	17.4	17.9	15.1	15.7	0.51	0.10	0.78	0.74
IIBR	10.0	7.8	14.2	12.3	0.91	0.028	0.38	0.51

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Lefaucheur, L., Lebret, B. (2020). The rearing system modulates biochemical and histological differences in loin and ham muscles between basque and large white pigs. *Animal*, sous presse (sous presse), sous presse. , DOI : 10.1017/S175173112000066X

IIBW	26.7	29.2	28.0	28.2	0.45	0.93	0.24	0.93
WSTM								
TNF (10 ⁻³) ^{3,4}	374	390	289	297	0.08	<0.001	0.61	0.74
Fiber percentage (%) ^{5,6}								
I	1.5	1.3	4.1	4.7	2.03	<0.001	0.83	0.54
IIA	8.4	7.9	8.2	8.5	4.28	0.88	0.82	0.86
IIBR	12.1	14.5	12.0	12.2	4.75	0.42	0.27	0.92
IIBW	78.0	76.3	75.7	74.6	5.19	0.23	0.46	0.63
Cross-sectional area (μm ²) ^{4,5}								
I	3526	3773	3412	3421	0.10	0.42	0.57	0.98
IIA	3532	3563	3617	3455	0.10	0.96	0.93	0.65
IIB ⁸	4282	4254	3673	3592	0.07	0.003	0.93	0.76
Mean	4194	4195	3668	3569	0.07	0.004	0.99	0.69
Relative area (%) ^{5,7}								
I	0.8	1.0	3.6	4.1	0.55	<0.001	0.76	0.63
IIA	6.4	5.9	7.9	8.1	0.74	0.14	0.77	0.92
IIB ⁸	91.8	92.0	87.8	87.3	0.23	0.004	0.94	0.80

569 ¹Mean of treatment groups.

570 ²P-values of contrasts differences between breeds or rearing systems within breed and RMSE
571 obtained from ANOVA.

572 ³Total number of muscle fibers.

573 ⁴ANOVA of log values to fit a normal distribution.

574 ⁵Fibers I : slow-twitch oxidative, IIA : fast-twitch oxido-glycolytic, IIBR : fast-twitch oxido-glycolytic,
575 IIBW : fast-twitch glycolytic.

576 ⁶ANOVA of raw data.

577 ⁷ANOVA of square root values to fit a normal distribution.

578 ⁸Cross-sectional areas of type IIBR and IIBW fibers could not be determined for technical reasons.

Comment citer ce document :

Lefaucheur, L., Lebret, B. (2020). The rearing system modulates biochemical and histological differences in loin and ham muscles between basque and large white pigs. *Animal*, sous presse (sous presse), sous presse. , DOI : 10.1017/S175173112000066X

579

580 **Figure 1.** Mid-cross-trans sectional section of semitendinosus muscle (STM) from
581 representative Large White (A) and Basque (B) pigs (adapted from Listrat *et al.*, 2016). R:
582 red portion of STM; W: white portion of STM; IMF: intramuscular fat content (%). Scale bar: 2
583 cm.

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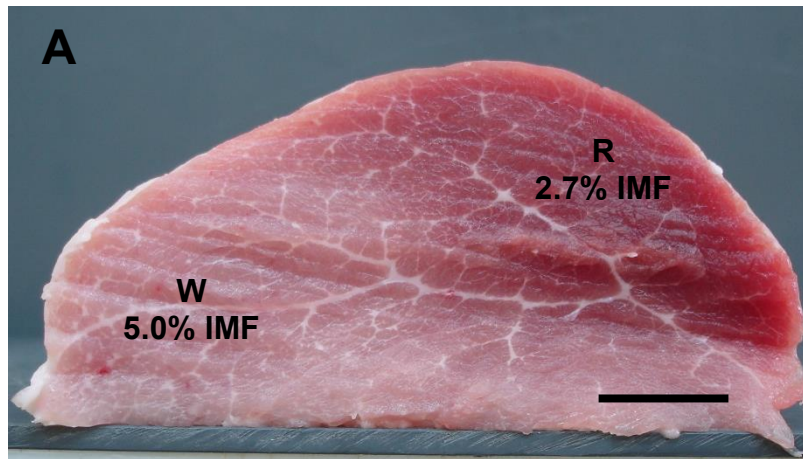
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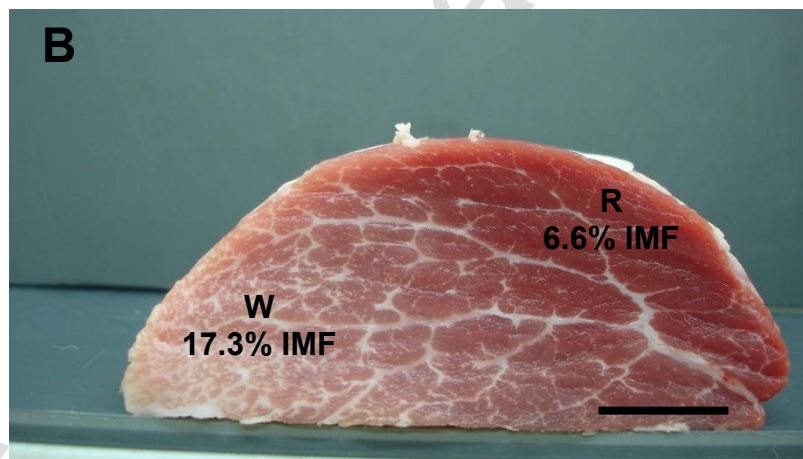
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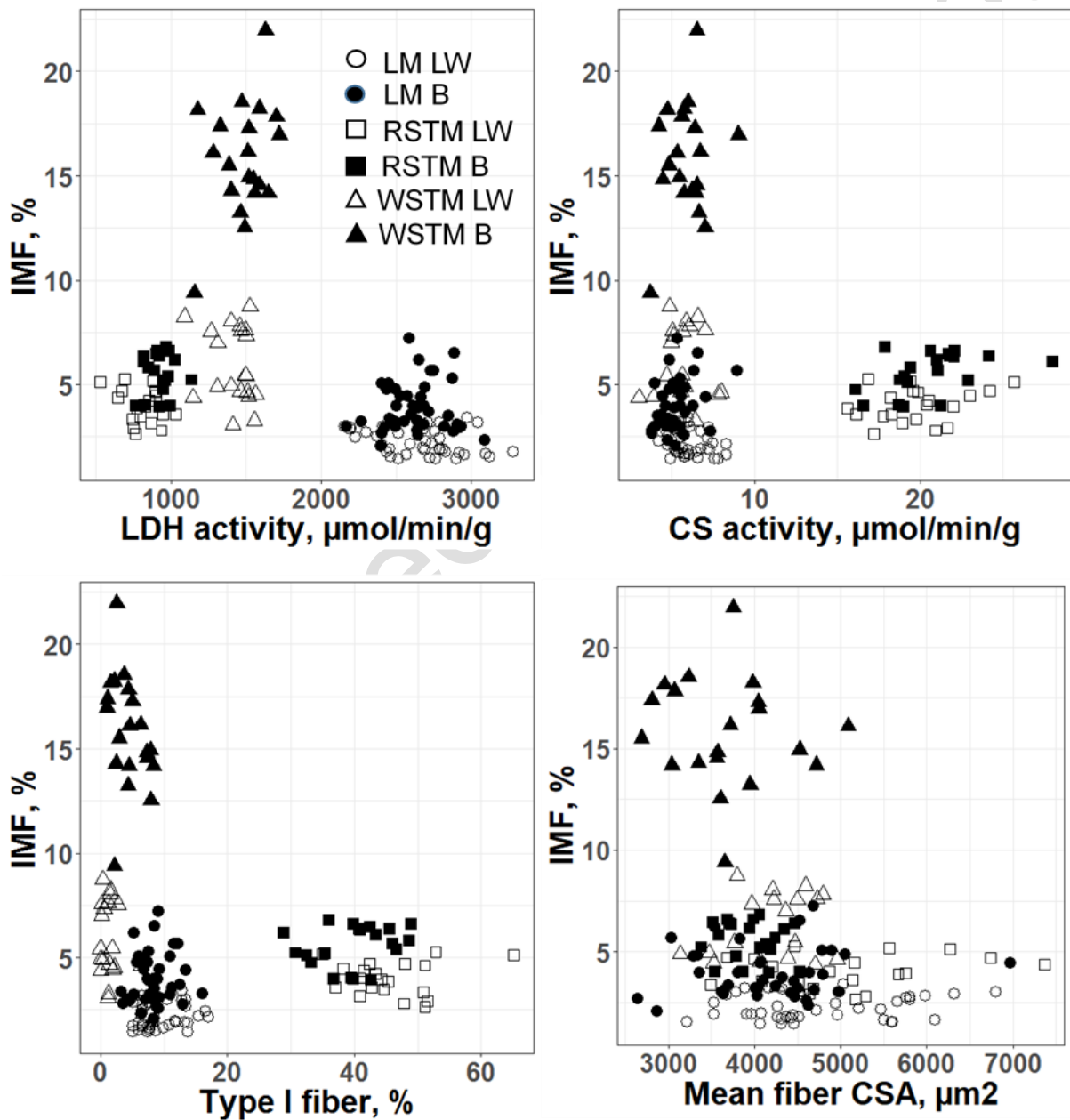
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598 **Figure 2.** Co-variation of intramuscular fat (IMF) content and lactate dehydrogenase
 599 (LDH), citrate synthase (CS) activities, percentage of type I (slow-twitch oxidative)
 600 fibers and mean fiber cross sectional area (CSA) in longissimus (LM) and red
 601 (RSTM) and white (WSTM) portions of semitendinosus muscles (STM) of Large
 602 White (LW) and Basque (B) pigs. Only pigs from the conventional (C) and alternative
 603 (A) rearing systems, represented by both breeds, are shown.



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