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Lateral roots: random diversity in adversity

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Abstract

Lateral roots are essential for soil foraging and uptake of minerals and water. They feature a large morphological diversity that results from divergent primordia or root growth and development patterns. Besides a structured diversity, resulting from the hierarchical and developmental organisation of root systems, there exists a random diversity, occurring between roots of similar age, of the same hierarchical order and exposed to uniform conditions. The physiological bases and functional consequences of this random diversity are largely ignored. Here we review the evidence for such random diversity throughout the plant kingdom, present innovative approaches based on statistical modelling to account for such diversity and set the list of its potential benefits in front of a variable and unpredictable soil environment.

Structured and random diversity of lateral roots

Lateral roots are essential for the uptake of minerals and water as well as for the many interactions with the surrounding soil. They are the first place where architectural responses of root systems to external abiotic and biotic stimuli take place ([1], [2]) suggesting that these responses contribute to the plant acclimation to the various soil conditions encountered. Within a plant, all lateral roots are not equal but rather display contrasting morphological (length, orientation) and anatomical (size of vessels, proportion of stele...) features with consequences on their function [3]. Such diversity is visible in the absence of soil heterogeneity and can thus be considered as an intrinsic property of the plant (Fig 1) by contrast with an environment-induced plasticity component. Intrinsic diversity can be partly accounted for by structural and developmental patterns within the root system. For instance, lateral roots branched on primary roots are often thicker, longer and display different anatomy than those branched on secondary roots and the same holds for tertiary as compared to secondary roots [eg [4] in trees, [5] in grasses, Fig 1A]. Similarly, lateral roots close to the parent root apex are shorter because they are younger as compared to more distal ones, due to the acropetal sequence of lateral root formation [6]. These sources (and others) lead to what we suggest to call *structured diversity* of lateral roots, arising from known sources of variation. Aside from this structured diversity, however, a *random diversity* term (Fig 1A) can be detected between lateral roots branched into the same region of a parent root *ie.* that are of the same order and of presumably the same age. The random nature of this diversity was

recently tested with appropriate statistical tools [7]. Figure 1 conceptualizes the distinction between intrinsic structured and random diversity of lateral roots and illustrates (Fig 1B-E) random diversity in various annual monocots and dicots as well as in trees. Additional illustrations can be found for sunflower (*Helianthus annuus*) [8], rubber tree (*Hevea brasiliensis*) [9], but also for the model plant *Arabidopsis thaliana* [10], [11] suggesting random diversity is a common feature within the plant kingdom. It has been proposed that it is prompted by developmental instability, a concept often used in to account for variation between individuals of the same genotype within the same (but unpredictable) environment [12] but that was applied to the case of diversity of lateral roots within a single individual plant [13]. Random diversity is visible through differences in length, diameter and anatomy of neighbouring lateral roots ([14] - [15] and Fig 1F) but also of neighbour primordia ([16], [17]). Within species, there exists genetic variability for structured diversity of lateral roots (eg. maize (*Zea mays* L.) genotypes with on average longer (and fewer) or shorter laterals, [18]). By contrast, very little information (but see [19]) is available on the occurrence of genetic variability of random diversity (*ie.* within a species, are there genotypes with more or less diverse lateral roots).

There is evidence that both structured and random diversity contribute to the responses of the root system to spatial and temporal soil heterogeneities. ([20], [21], Fig 1G), although in a different way. Within the structured component of diversity, similar roots (same order, same age) tend to respond similarly. A clear illustration is the ‘xerobranching’ pattern induced by transient lack of water ([22], Fig 1H) where all primordia passing by a given developmental window during the stress are permanently arrested while emerged roots continue to grow or recover. Another example is the development of cluster roots to low P in some species such as white lupin (*Lupinus albus*) [23]. In the random component, different roots may respond differently. An example is the response to N spatial heterogeneity with longer roots responding more steeply to high N (illustrated in Fig 1G).

Paradoxically, while the development of lateral roots and of their primordia has been the matter of intense research during the last two decades, in particular in *Arabidopsis* (Banda et al 2019 in this issue of TIPS gives a timely update on the molecular networks operating on these processes), the random diversity of lateral roots has been largely overlooked. While root system architecture is increasingly targeted as a lever to improve plant resource capture efficiency under poor and/or fluctuating (minerals, water) soil ([24], [25]) and more broadly

low-input agriculture ([26], [18]), it becomes essential to better figure out (i) when, where and how lateral root random diversity takes place, (ii) how it can be accounted for by models and (iii) what can be the benefits of this diversity in front of an adverse, spatially and temporally variable and unpredictable soil environment. These points are the matter of the present review.

Lateral root random diversity: when? where?

In order to understand the outset of lateral root random diversity, we propose to recall the various stages of lateral root development and localize where random diversity is taking place. This different stages and the stochastic representation of the different trajectories of primordia and roots is shown in Figure 2. The current paradigm emanating from the model plant *Arabidopsis* [27] states that regular oscillations of molecular responsiveness to auxin at the basal meristem / elongation zone region of the primary root prime those pericycle cells which flow through that zone during peaks of responsiveness. These early oscillations can be affected by environmental clues, such as an ABA-mediated repression by a lack of water [22]. These primed regions, named pre-branch sites, become competent for later asymmetrical division and primordia initiation [28]. The activation of cell division of primed cells is, however, not systematic and depends on local and global clues [29]. For example, primordia initiation is promoted under conditions of high auxin [30]. Priming and initiation can thus be formalised by an endogenous, process-dependent probability (occurrence or not of the auxin oscillation, activation or not of pre-branch sites, Fig 2). These probabilities are under the control of local environment such as patches with high nitrate [31], high water availability [32] or low P in the case of cluster roots [23] and auxin is again a good candidate to integrate these local environmental fluctuations.

Once initiated, the fate of primordia is not unique. This can be seen in (rare) studies where primordia size (length, width, cell numbers) is recorded as a function of distance to the parent root apex ([17], [16]). If primordia initiation is considered acropetal (although there are debates on this depending on species, see [29] and [6]), a high variance of primordium size at a given distance from the parent root apex means that primordia develop at various rates, possibly equipped with various anatomical attributes (Fig 2). An extreme example of this diversity is the S and L primordia in rice (*Oryza sativa*) that give rise to S (short, determinate growth) and L (long) lateral roots [33] which differ strongly in their anatomy [34]. Beyond auxin ([35], [36]), the list of hormonal and molecular signals able to influence probabilities

associated with primordia development is long, starting with cytokinins ([37], [38]) and regularly updated: sugars [39], brassinosteroids [40], ethylene [41], jasmonic acid [42] but also signalling peptides [43], small RNA [44] or more recently very long chain fatty acids [45] all play their part in this process, often with some crosstalks [38]. How these signals contribute to maintaining or exacerbating lateral root primordia diversity both under non-stressful conditions or as mediator of environmental clues is essentially not known and should attract the attention of the lateral root community in the coming years.

Ultimately, primordia may or may not emerge. A lack of emergence of a proportion of primordia has been repeatedly observed in response to high [46] but also very low [47], [35], nitrate, high phosphorus [10], [48] or water stress [49] but it is less acknowledged that this occurs also in the absence of stresses ([6], [50]). Lucas et al. [51] identified auxin again as a major actor governing the succession of arrested primordia/emerged lateral roots in *Arabidopsis*. According to our framework, this means that auxin likely alters the different probabilities associated with primordia development (Fig 2).

Once emerged, lateral roots display a range of growth patterns, from a rapid deceleration leading to growth arrest shortly after emergence, up to a steep acceleration and sustained growth for several days (Fig 2). This diversity is consistently revealed in studies in which time series of lateral root growth rate were measured (see examples in *Arabidopsis* [11], in oak tree (*Quercus robur*) [52], in peach tree (*Prunus persica*) [53], in rubber tree [9], sunflower [8], maize and pearl millet (*Pennisetum glaucum*) [7]). Diversity can take the form of a continuum [11], [52], or of distinct types (classically 2-4), that have been either strictly defined (as in the case of rice, [54] or millet [55]) or defined with some overlap as in the case of maize [7]. Short lateral roots are usually thin, with limited transport capacity [55] and their meristem show signs of exhaustion or at least shrinkage [56], [57] as in the case of determinate primary roots as found in some species such as Cactaceae [58]. Lateral root length or growth rate are often correlated – though loosely – to apical or basal diameter, displaying an ‘envelope curve’, suggesting that root diameter (and thus likely the surface area of xylem or phloem vessels and their conductivity) defines a potential growth rate that cannot be exceeded due to conductivity limits ([59], [14]). The diversity of lateral root fate within that envelope curve occurs in the absence of environmental heterogeneity and contributes to what was defined as random diversity above. In addition, evidence can be found that random diversity also contributes to environment-induced plasticity of lateral root fate. For example,

in *Arabidopsis*, root segments in contact with high nitrate patches show increased proportion of long, likely fast growing roots [60] while phosphorus deficiency leads to a reduced number of fast and long roots ([10], [20]). Similarly, the response to soil moisture in rice involves a change of the proportion of L-types (and there exists a large genetic variability for this trait [19]). This proportion also depends on internal clues. Once again, auxin is a well-known modifier of the proportion of lateral roots but only within certain temporal limits, out of which it has little influence [61]. Sugars also stimulate lateral root elongation rate [62] and change the proportion of fast and slow growing roots [11] suggesting sugars are also important component of the signalling landscape influencing probabilities associated with lateral root development (Fig 2).

Recently, using appropriate statistical tools, we found no evidence of any structuration of the occurrence of long, short or intermediate laterals along primary roots in maize and pearl millet suggesting this typology results from a random process [7]. The molecular and genetic bases of this random process, largely overlooked, now need careful attention. While genetic regions controlling lateral root density are being revealed and validated at the molecular level [63], an identification of genetic regions controlling lateral root diversity in crops will become the next challenge.

Lateral root random diversity: a journey through a progressive decrease of entropy

Root biologists conventionally report lateral root descriptors (e.g. diameter, growth rate, angle) that are averaged (see inset in Fig 1) at the level of the plant or group of plants (e.g. genotypes, conditions). This group-centered approach maintains the vision of differentiation as a stereotyped program that all lateral roots execute identically, starting at the specification of the pre-branch site. Contrastingly, in the animal domain, there is growing acknowledgement that stochasticity is an essential aspect of differentiation, as illustrated by the great variability of gene expression at the single cell level in undifferentiated tissues [64]. In line with this, some studies refer to the theoretical framework of non-equilibrium thermodynamics, which offers innovative paths to analyse biological processes related to differentiation [65]. In that context, cell differentiation is viewed as a succession of irreversible transition(s) from one steady state to another, entailing a continuous decrease of **entropy** [66]. Between successive transitions along the differentiation journey, cells remain in a (meta)-stable state with a characteristic gene expression profile that is self-maintained by the cell gene regulatory network. Transitions occur through a transient increase of **entropy**

triggered by some tissue signals and are followed by a decrease of **entropy** as the cell stabilises in a new state [67]. This theoretical framework also copes with multi-stability *i.e.* multiple possible fates. Importantly, it provides observables, such as Shannon **entropy**, that can aid identifying key time-points or actors which break the stability of the system required for differentiation [64].

Consistent with this view, we propose that the succession of events leading to the random diversity of primordia and lateral root fates can be seen as a journey through a progressive decrease of Shannon **entropy**, *i.e.* an increase of information and a decrease of the deformability of the system (Fig 3). If we unroll the different phases of primordia and lateral root development, the lack of priming of pre-branch sites depicted in the previous section is analogous to a steep decrease of **entropy** since non-primed pericycle cells definitely lose their ability to initiate primordia. Primed sites experience a lower decrease of **entropy** since they may or may not lead to primordium initiation (Fig 3). Pre-branch sites that do not give rise to primordia remain stuck in a steady state with no further possible initiation (**entropy** sink). Once initiated, primordia developmental trajectories differ. If these trajectories are mirrored by different anatomies as suspected ([7], [55]) with more or less, smaller or larger xylem and phloem vessels, changes of trajectories may become less likely. Finally, the emergence of lateral roots leads to a decrease of **entropy** with different amplitudes. Those lateral roots quickly decelerating also fall in an **entropy** sink since they most often lose their meristem [56]. Similarly, accelerating roots have their meristem enlarging with time [56] which suggests they are well equipped to grow for a long time. As in the case of primordia, roots with intermediate behaviour could exhibit different fates awaiting for signals depending on intrinsic or environmental conditions [7]. Illustrations of the flexibility of these trajectories come from classical studies showing that modifications of sugar supply or local IAA or tissue ABA induces massive changes in lateral root fate at the different transitions (Fig 3, [22], [32], [11], [61], [68], [69], [70]).

Lateral root primordia development therefore sounds like an appropriate material to explore the benefits of using the concept of **entropy** in plant developmental biology as recently reviewed [64]. This would require defining a metric for **entropy** and testing signals (Fig 3) potentially increasing or reducing **entropy**. It could also stimulate studies on gene expression network at sub-tissular or even single cell levels levels, as it is recently currently developing [71], for instance in incipient primordia.

Stochastic models as a keystone to characterize lateral root diversity

In order to consider lateral root diversity as a trait, and characterize genetic and/or environmental impacts, some effort is needed to define variables and ways to measure them. Because diversity in growth patterns has been often related to diversity in diameters ([9], [55]), the coefficient of variation of basal or apical diameter of lateral roots was recently used to characterize the diversity of lateral roots across 140 mono and dicot species [14]. Based on the central role of root apical diameter in determining growth potential, the ArchiSimple model [72] was developed that satisfactorily reproduce the root architectural features of a large number of species with a low number of parameters (Fig 4A). In that case, the diversity of lateral root growth patterns is modelled by incorporating a **stochastic** function for the assignment of lateral root diameter at emergence, with lateral root types emerging as a consequence of this stochasticity. Several other root models incorporate stochasticity which is implemented either directly (each lateral root growth rate is sampled from a normal distribution with mean and standard deviation set by the user) or indirectly through diameters as in the case of ArchiSimple. A subset of root architectural models accounting for stochasticity and thus lateral root types is listed in Table 1. It notably includes CRootBox [73], DigR [74], OpenSimRoot [75], or RootTyp [76]. These models also have the capacity to define explicitly several lateral root types with additional parameters including type-specific probabilities of generation by the parent root. In that case, diversity is not an emergent property of the model and requires a more extensive parameterization. As far as we know, neither the effect of environmental clues, nor the effect of genetic variation has been implemented to modulate proportions of lateral root types in these models. This step certainly constitutes a future challenge in order to propose a new generation of root architecture models able to respond to a fluctuating environment through modification of lateral root growth patterns. Another challenge is to couple these root architectural models to soil models in order to evaluate the impact of root architecture, including lateral root random diversity, on resource capture performance [77].

Inasmuch as functional-structural models are perfect tools to closely reproduce real root architectures based on growth and development processes, statistical models are perfect tools to identify patterns of lateral root diversity and thus be used to perform genetic analyses or evaluate the impact of environmental stimuli. In line with this, a new phenotyping pipeline was recently developed and evaluated on two cereals: maize and pearl millet [7]. This pipeline

starts with the SmartRoot image-analysis system [78], which is used to reconstruct consistent spatio-temporal data on the basis of successive snapshots of root system architecture, with temporal and spatial statistical models. Daily growth rates of individual lateral roots (Fig 4B) are then analyzed using semi-Markov switching linear models (SMS-LM) that synthesize the information into a probability to enter a growth stage, a duration before growth cessation and a linear trend describing the change in growth rate with time. For both maize and pearl millet, three types were identified, with similar characteristics across species [79]. In pearl millet, these types matched anatomical differences in whole root or stele diameter as well as in number and size of xylem vessels. Interestingly, introducing genetic (auxin signalling mutant) or environmental (severe shading) perturbations in maize only changed the proportion of roots in each type, but not the definition of types. This suggests that environmental or genetic effects could be captured directly through effects on subsets of statistical parameters [72], [73]. For instance, environmental effects could be formalized through a parameterized response curve between a transition probability as depicted in Fig 2 and an environmental variable. These parameters can then be considered as genotype-dependent, as in the case of models accounting for genetic and environmental effect on shoot growth ([80], [81]).

Lateral root random diversity for efficiency in adversity

Addressing the functional significance of lateral root random diversity means dealing with several levels of complexity: (i) environmental clues are perceived locally (*e.g.* local patches of nitrate associated with intense mineralization) but heritable benefits are seen at the whole plant level and on the long term (fitness); (ii) developmental decisions leading to lateral root diversity are taken during short time windows (Fig 2) with only a partial knowledge of the future whole plant context and soil conditions; (iii) benefits are (soil) context-dependent: few long lateral roots are better to capture the mobile nitrate while many short ones are preferable to capture the immobile phosphate [82], [18]; (iv) benefits need to be evaluated in reference to carbon costs; (v) lateral roots and their environment are part of the same system and they influence each other; (vi) finally, root developmental patterns or responses are most often, if not always, described through the filter of averaging while distributions would be certainly more insightful. Nevertheless, in the following, we attempt to connect the available results (either from rare studies or most often from simulations) around five different, yet overlapping, aspects of the functional significance of lateral roots random diversity that are sketched in Fig 5.

Functional complementarity

Several examples indicate that the functions of the root system are, at least in part, distributed between the different types of roots. A first example is the ratio of radial-to-axial hydraulic conductance which is predicted to affect the distribution of root water uptake along a lateral root [83]. Specifically, in thin lateral roots with a large ratio [84], the water influx is expected to be biased towards the root base, while thick lateral roots with a small ratio (mainly because axial conductance is much larger in thick than in thin roots), would distribute a larger influx on their entire length (Fig 5A). In this context, a random diversity of lateral roots allows the plant to optimise the route of water from the bulk soil to the shoot and to avoid localized hotspots of uptake that are prone to cause localized soil drying and drop of rhizosphere conductivity [85]. The co-existence of lateral roots with different hydraulic properties and which do not take up water equally even when they are exposed to the same rhizosphere moisture is also expected to smooth out the distribution of water uptake through the soil profile.

The specificity of lateral root responses to phosphorus, nitrogen and water shortage, is also an indication of functional complementarity. This functional complementarity is a cornerstone of multifunctional root system ideotypes (e.g. drought- and low nitrogen-tolerant) that are proposed to rely on a diversity of lateral root growth types optimised to the distribution of water and nitrogen in the soil profile ([86], [87]). In addition to the different lengths of lateral roots, this functional diversity can be due to the position of the roots relative to their neighbour, and to the extent of secondary branching that is often exacerbated in long laterals [61]. Finally, it can be due to the anatomy and molecular arsenal of each lateral root (cluster roots being an extreme case of functional specificity [88]).

Foraging efficiency

The diversity of lateral roots discussed in this paper essentially focuses on length and thickness. As these are tightly correlated with construction and maintenance costs, one can expect lateral root diversity to be an important lever to adjust the foraging efficiency *i.e.* soil volume explored per unit carbohydrates invested into roots (Fig 5B-C). This foraging efficiency has been predicted to increase with the diversity of lateral root growth patterns [89].

Beyond its effect on the volume of soil explored, the way carbon is invested among different root types (including lateral root types) also affects how resources flow in the soil-plant

system, in a resource-dependent manner. For example, S-type lateral roots in rice root system may act as long-lived super root hairs (Fig 5B-C), increasing the root surface area and nutrient uptake while at the same time being carbon-cost efficient compared to the large L-type lateral roots [90]. Mimicking root hairs at high density, these fine S-type roots will further improve soil exploration at minimal cost [91]. The overlap between the rhizosphere of neighbour lateral roots (and thus an inefficient carbon investment) is expected to decrease with alternating diverse roots types [89]. Consistent with this, rice exhibits a decrease of lateral root density with crown root rank that also shows some genotypic differences [92].

More generally, it has been suggested that root system foraging efficiency could be cast in a more general context of behavioural ecology, where behaviour classically refers to the action or reaction of a foraging unit (e.g. an individual root) to an event or stimulus [93]. For instance, as pointed out by Forde [13], diversity of individual root trajectories resembles random walk strategy used by animals to find resource-rich patches when they have no *a priori* knowledge of the location of their target. In that perspective, selection should favour a diversity of lateral root behaviours that are recruited to enhance whole plant fitness, rather than the performance of each and every foraging unit. In agreement with this view, using the auxin insensitive mutant *axr-4*, it was shown that diversity and responsiveness to nutrient patches (responsiveness being lost in the mutant) are particularly important for plant fitness to better capture the immobile phosphorus and much less to capture the highly mobile nitrate [94].

Risk management

The optimal carbon investment in lateral roots is largely context dependent. For example, when the spatial and temporal distribution of resources is known, plants can afford a low cost strategy and develop lateral roots preferentially into resource-rich patches (Fig 5F) or layers [1]. However, in **stochastic** environments where local conditions change over time, such strategy is inherently risk prone, while plants that invest in many foraging units with lower resource gains might be considered risk averse [93]. It was recently shown that plants are able of risk-sensitive behaviours by which they adjust their relative allocation to stable *vs.* variable soil compartments, depending on the average nutrient availability [95]. The role of lateral roots diversity in that context remains to be addressed. However, the observation that the distribution of lateral root growth rate is modified according to assimilate availability [11] suggests that lateral root diversity contributes to some form of risk management.

The context in which decisions concerning lateral root development and growth are taken extends beyond resource availability and carbon allocation. In particular, cavity expansion pressure and bending stiffness, which influence root penetration and likelihood of buckling, are largely dependent on the root shape, on its rate of penetration and on the soil mechanical properties ahead of and around the tip [96]. Their importance may be revealed by the evolution of developmental responses such as **hydropatterning** [32] and **xerobranching** [22], which repress the initiation of lateral roots that would otherwise emerge in air (dry soil macropore), with limited lateral mechanical support in the elongation zone which may prevent them re-entering the bulk soil [97]. Since the shape and growth rate of emerging lateral roots are determined very early (Fig 2), potentially before the lateral roots can be informed of the soil structure (pore size, deformability), an intrinsic lateral root diversity should increase the chances for a lateral root to better seek out cracks and pores (Fig 5D) and to minimise the likelihood of buckling when it enters the soil [98].

Unleashed phenotypic plasticity

The abundant literature on lateral roots plasticity, certainly biased towards *Arabidopsis* seedlings, implicitly assumes the existence of a unique lateral root type which responds to environmental cues, leading to a range of lateral roots phenotypes (reaction norms). However, careful observations have shown that different lateral root types do not respond similarly to the same variation of their substrate [99], [100] and that their responses are affected by genotype-by-environment interactions [19]. From an evolutionary perspective, a root system comprising different types of lateral roots whose proportions and behaviour can be modulated in type-specific manners benefits from many more degrees of freedom than a root system made of lateral roots that respond all in the same way (Fig 5E-F).

Information system

Since root tips provide the plant with a variety of signals [101], some being transported throughout the plant [102], [103], a root system can be seen as a distributed network of sensors. In such distributed systems, decisions at the local level (for a lateral root, to grow or not to grow) need to be taken considering both the local environment as well as some integrated control by the whole plant. In this context, lateral roots diversity, leading to a random spatial distribution of lateral root tips (Fig 5G), avoiding clusters of root tips in the same area, may better contribute to providing the plant with a balanced level of signals and, in turn, set-up the right decision after integrating both the local and whole plant information.

Concluding remarks and future perspectives

Most of the characterizations of lateral roots in plants rely on cumulative or averaged variables (e.g. total length, average diameter, specific root length). Moreover, a vast majority of studies on lateral root primordia have focused on the identification of standard patterns of development. Here, we defend the idea that considering the diversity of lateral roots and primordia is essential because it is probably at the basis of the huge phenotypic plasticity of the root system exposed to a variable and unpredictable soil environment. We therefore suggest that diversity of lateral roots (and primordia) should be more often (if not systematically whenever possible) quantified through appropriate traits such as probabilities, or intra-plant distributions of length / diameter (see simple methods for measuring intra-plant diameter distribution in [14] and [15]). We therefore urge the root community to adopt standards for capturing and sharing this information. Recently, the RSML [104] format was proposed. It has the virtue (among others) to store detailed variables at the lateral root scale. Given the large number of groups evaluating root plasticity in response to the environment in a variety of phenotyping displays ([105], [106], [107], [108]) the recourse to a common data format should multiply the possibilities of meta-analyses.

How the many molecular signals involved at some point in lateral root development interfere with diversity is essentially not known although auxin certainly plays one of the first roles. In this regard, the lateral root primordium sounds like an ideal organ to explore the role of **stochastic** processes in development. Incorporating the concept of **entropy** in Plant Biology is likely to be an important step in line with the advances it allowed in animal embryogenesis. A lot must now be done to explore the genetic variability of intrinsic and environmentally-driven lateral root diversity in crops. In parallel, models are essential to predict the consequences of more or less diversity or more or less plastic diversity to the environment. With this information in hands, one could envisage to provide breeding schemes with new traits (diversity) allowing the selection of efficient cultivars able to capture resources at low (carbon) cost in conditions where resource fluctuations in time and space are particularly high, as in poor soils and/or under low input agriculture conditions (see also outstanding questions).

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References

- 1 Drew, M.C. *et al.* (1973) Nutrient Supply and the Growth of the Seminal Root System in Barley. THE EFFECT OF NITRATE CONCENTRATION ON THE GROWTH OF AXES AND LATERALS. *J Exp Bot* 24, 1189–1202
- 2 Malamy, J.E. Intrinsic and environmental response pathways that regulate root system architecture. *Plant, Cell & Environment* 28, 67–77
- 3 Comas, L. *et al.* (2013) Root traits contributing to plant productivity under drought. *Frontiers in plant science* 4, 442
- 4 Guo, D. *et al.* (2008) Anatomical traits associated with absorption and mycorrhizal colonization are linked to root branch order in twenty-three Chinese temperate tree species. *New Phytologist* 180, 673–683
- 5 Picon-Cochard, C. *et al.* (2012) Effect of species, root branching order and season on the root traits of 13 perennial grass species. *Plant Soil* 353, 47–57
- 6 Dubrovsky, J.G. *et al.* (2006) Lateral root initiation in Arabidopsis: Developmental window, spatial patterning, density and predictability. *Annals of Botany* 97, 903–915
- 7 Passot, S. *et al.* (2018) A new phenotyping pipeline reveals three types of lateral roots and a random branching pattern in two cereals. *Plant Physiology* DOI: 10.1104/pp.17.01648
- 8 Aguirrezabal, L.A.N. and Tardieu, F. (1996) An architectural analysis of the elongation of field-grown sunflower root systems. Elements for modelling the effects of temperature and intercepted radiation. *Journal of Experimental Botany* 47, 411–420
- 9 Thaler, P. and Pagès, L. (1996) Root apical diameter and root elongation rate of rubber seedlings (*Hevea brasiliensis*) show parallel responses to photoassimilate availability. *Physiologia Plantarum* 97, 365–371
- 10 AL-GHAZI, Y. *et al.* (2003) Temporal responses of Arabidopsis root architecture to phosphate starvation: evidence for the involvement of auxin signalling. *Plant, Cell & Environment* 26, 1053–1066
- 11 Freixes, S. *et al.* (2002) Root elongation and branching is related to local hexose concentration in *Arabidopsis thaliana* seedlings. *Plant, Cell & Environment* 25, 1357–1366
- 12 Valladares, F. *et al.* (2002) Plasticity, instability and canalization: is the phenotypic variation in seedlings of sclerophyll oaks consistent with the environmental unpredictability of Mediterranean ecosystems? *New Phytologist* 156, 457–467

- 13 Forde, B.G. (2009) Is it good noise? The role of developmental instability in the shaping of a root system. *J Exp Bot* 60, 3989–4002
- 14 Pagès, L. (2016) Branching patterns of root systems: comparison of monocotyledonous and dicotyledonous species. *Annals of botany* 118, 1337–1346
- 15 Wu, Q. *et al.* (2016) Relationships between root diameter, root length and root branching along lateral roots in adult, field-grown maize. *Ann Bot* 117, 379–390
- 16 MacLEOD, R.D. (1976) GROWTH OF LATERAL ROOT PRIMORDIA IN VICIA FABA L. *New Phytologist* 76, 143–151
- 17 MacLeod, R.D. (1990) Lateral root primordium inception in *Zea mays* L. *Environmental and Experimental Botany* 30, 225–234
- 18 Zhan, A. and Lynch, J.P. (2015) Reduced frequency of lateral root branching improves N capture from low-N soils in maize. *Journal of Experimental Botany* 66, 2055–2065
- 19 Niones, J.M. *et al.* (2015) QTL associated with lateral root plasticity in response to soil moisture fluctuation stress in rice. *Plant and Soil* 391, 63–75
- 20 Williamson, L.C. *et al.* (2001) Phosphate Availability Regulates Root System Architecture in *Arabidopsis*. *Plant Physiol.* 126, 875–882
- 21 Remans, T. *et al.* (2006) A Central Role for the Nitrate Transporter NRT2.1 in the Integrated Morphological and Physiological Responses of the Root System to Nitrogen Limitation in *Arabidopsis*. *Plant Physiol.* 140, 909–921
- 22 Orman-Ligeza, B. *et al.* (2018) The xerobranching response represses lateral root formation when roots are not in contact with water. *Current Biology* 28, 3165–3173
- 23 Gallardo, C. *et al.* (2018) Anatomical and hormonal description of rootlet primordium development along white lupin cluster root. *Physiologia plantarum*
- 24 Julkowska, M. *et al.* (2017) Genetic components of root architecture remodeling in response to salt stress. *The Plant Cell*
- 25 Dowd, T.G. *et al.* (2018) Maize lateral root developmental plasticity induced by mild water stress. I: Genotypic variation across a high-resolution series of water potentials. *Plant, cell & environment*
- 26 Lynch, J.P. (2018) Rightsizing root phenotypes for drought resistance. *J Exp Bot* 69, 3279–3292
- 27 Moreno-Risueno, M.A. *et al.* (2010) Oscillating gene expression determines competence for periodic *Arabidopsis* root branching. *Science* 329, 1306–1311
- 28 Laskowski, M. and Kirsten, H. (2017) Periodic lateral root priming: What makes it tick? *The Plant Cell* 29, 432–444
- 29 McCleery, W.T. *et al.* (2017) Root branching plasticity: collective decision-making results from local and global signalling. *Current opinion in cell biology* 44, 51–58
- 30 Laskowski, M. (2013) Lateral root initiation is a probabilistic event whose frequency is set by fluctuating levels of auxin response. *Journal of experimental botany* 64, 2609–2617
- 31 Mounier, E. *et al.* (2014) Auxin-mediated nitrate signalling by NRT 1.1 participates in the adaptive response of *Arabidopsis* root architecture to the spatial heterogeneity of nitrate availability. *Plant, cell & environment* 37, 162–174
- 32 Bao, Y. *et al.* (2014) Plant roots use a patterning mechanism to position lateral root branches toward available water. *PNAS* 111, 9319–9324
- 33 Kawata, S.-I. and Shibayama, H. (1965) On the lateral root primordia formation in the crown roots of rice plants. *Japanese Journal of Crop Science* 33, 423–431
- 34 Coudert, Y. *et al.* (2010) Genetic control of root development in rice, the model cereal. *Trends in Plant Science* 15, 219–226
- 35 Krouk, G. *et al.* (2010) Nitrate-regulated auxin transport by NRT1. 1 defines a mechanism for nutrient sensing in plants. *Developmental cell* 18, 927–937
- 36 Celenza, J.L. *et al.* (1995) A pathway for lateral root formation in *Arabidopsis thaliana*. *Genes Dev.* 9, 2131–2142
- 37 Marhavý, P. *et al.* (2014) Cytokinin Controls Polarity of PIN1-Dependent Auxin Transport during Lateral Root Organogenesis. *Current Biology* 24, 1031–1037

- 38 Liu, J. *et al.* (2017) Crosstalk complexities between auxin, cytokinin and ethylene in Arabidopsis root development: from experiments to systems modelling, and back again. *Molecular plant*
- 39 Bingham, I.J. *et al.* (1997) Site, scale and time-course for adjustments in lateral root initiation in wheat following changes in C and N supply. *Annals of Botany* 80, 97–106
- 40 Wei, Z. and Li, J. (2016) Brassinosteroids regulate root growth, development, and symbiosis. *Molecular plant* 9, 86–100
- 41 Ivanchenko, M.G. *et al.* (2008) Ethylene–auxin interactions regulate lateral root initiation and emergence in Arabidopsis thaliana. *The Plant Journal* 55, 335–347
- 42 Huang, H. *et al.* (2017) Jasmonate action in plant growth and development. *Journal of experimental botany* 68, 1349–1359
- 43 Oh, E. *et al.* (2018) Signaling peptides and receptors coordinating plant root development. *Trends in plant science* 23, 337–351
- 44 Yu, N. *et al.* (2015) The role of miR156/SPL s modules in Arabidopsis lateral root development. *The Plant Journal* 83, 673–685
- 45 De Bigault Du Granrut, A. and Cacas, J.-L. (2016) How Very-Long-Chain Fatty Acids Could Signal Stressful Conditions in Plants? *Front. Plant Sci.* 7,
- 46 De Smet, I. *et al.* (2003) An abscisic acid-sensitive checkpoint in lateral root development of Arabidopsis. *The Plant Journal* 33, 543–555
- 47 Mounier, E. *et al.* (2014) Auxin-mediated nitrate signalling by NRT1. 1 participates in the adaptive response of Arabidopsis root architecture to the spatial heterogeneity of nitrate availability. *Plant, cell & environment* 37, 162–174
- 48 Nacry, P. *et al.* (2005) A role for auxin redistribution in the responses of the root system architecture to phosphate starvation in Arabidopsis. *Plant Physiol* 138, 2061–74
- 49 Dubrovsky, J.G. *et al.* (1998) Root growth, developmental changes in the apex, and hydraulic conductivity for Opuntia ficus-indica during drought. *The New Phytologist* 138, 75–82
- 50 Barlow, P.W. and Adam, J.S. (1988) The position and growth of lateral roots on cultured root axes of tomato, Lycopersicon esculentum (Solanaceae). *Plant Systematics and Evolution* 158, 141–154
- 51 Lucas, M. *et al.* (2008) An Auxin Transport-Based Model of Root Branching in Arabidopsis thaliana. *PLoS ONE* 3, e3673
- 52 Pagès, L. (1995) Growth patterns of the lateral roots of young oak (Quercus robur) tree seedlings Relationship with apical diameter. *New Phytologist* 130, 503–509
- 53 Pagès, L. *et al.* (1993) Development of the Root System of Young Peach Trees (Prunus persia L. Batsch): A Morphometrical Analysis. *Annals of Botany* 71, 369–375
- 54 Rebouillat, J. *et al.* (2009) Molecular genetics of rice root development. *Rice* 2, 15–34
- 55 Passot, S. *et al.* (2016) Characterization of Pearl Millet Root Architecture and Anatomy Reveals Three Types of Lateral Roots. *Front Plant Sci* 7,
- 56 Moreno-Ortega, B. *et al.* (2017) Identifying Developmental Zones in Maize Lateral Root Cell Length Profiles using Multiple Change-Point Models. *Front. Plant Sci.* 8,
- 57 Varney, G.T. and McCully, M.E. (1991) The branch roots of Zea. II. Developmental loss of the apical meristem in field-grown roots. *New Phytologist* 118, 535–546
- 58 Shishkova, S. *et al.* (2008) Determinate root growth and meristem maintenance in angiosperms. *Annals of botany* 101, 319
- 59 Pagès, L. (2014) Branching patterns of root systems: quantitative analysis of the diversity among dicotyledonous species. *Ann Bot* 114, 591–598
- 60 Remans, T. *et al.* (2006) The Arabidopsis NRT1. 1 transporter participates in the signaling pathway triggering root colonization of nitrate-rich patches. *Proceedings of the National Academy of Sciences* 103, 19206–19211
- 61 Kawai, T. *et al.* (2017) Compensatory growth of lateral roots responding to excision of seminal root tip in rice. *Plant Root* 11, 48–57

- 62 Bingham, I.J. and Stevenson, E.A. (1993) Control of root growth: effects of carbohydrates on the extension, branching and rate of respiration of different fractions of wheat roots. *Physiol Plant* 88, 149–158
- 63 Wang, F. *et al.* (2018) Genome-wide association and gene validation studies for early root vigour to improve direct seeding of rice. *Plant, Cell & Environment* 41, 2731–2743
- 64 Richard, A. *et al.* (2016) Single-Cell-Based Analysis Highlights a Surge in Cell-to-Cell Molecular Variability Preceding Irreversible Commitment in a Differentiation Process. *PLOS Biology* 14, e1002585
- 65 Goldbeter Albert (2018) Dissipative structures in biological systems: bistability, oscillations, spatial patterns and waves. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences* 376, 20170376
- 66 Barbacci, A. *et al.* (2015) Thermodynamical journey in plant biology. *Front. Plant Sci.* 6,
- 67 Mojtahedi, M. *et al.* (2016) Cell Fate Decision as High-Dimensional Critical State Transition. *PLOS Biology* 14, e2000640
- 68 Brown, N.A. and Gardner, F.E. (1936) Galls produced by plant hormones, including a hormone extracted from *Bacterium tumefaciens*. *Phytopath* 26, 708–713
- 69 Eysholdt-Derzso, E. and Sauter, M. (2017) Root bending is antagonistically affected by hypoxia and ERF-mediated transcription via auxin signaling. *Plant physiology*
- 70 Bingham, I.J. *et al.* (1998) Relationship between tissue sugar content, phloem import and lateral root initiation in wheat. *Physiologia Plantarum* 103, 107–113
- 71 Shulze, C.N. *et al.* (2019) High-Throughput Single-Cell Transcriptome Profiling of Plant Cell Types. *Cell Reports* 27, 2241–2247.e4
- 72 Pagès, L. *et al.* (2014) Calibration and evaluation of ArchiSimple, a simple model of root system architecture. *Ecological Modelling* 290, 76–84
- 73 Schnepf, A. *et al.* (2018) CRootBox: a structural–functional modelling framework for root systems. *Annals of botany* 121, 1033–1053
- 74 Barczi, J.-F. *et al.* (2018) DigR: a generic model and its open source simulation software to mimic three-dimensional root-system architecture diversity. *Annals of botany* 121, 1089–1104
- 75 Postma, J.A. *et al.* (2017) OpenSimRoot: widening the scope and application of root architectural models. *New Phytol.*
- 76 Pagès, L. *et al.* (2004) RootTyp: a generic model to depict and analyse the root system architecture. *Plant Soil* 258, 103–119
- 77 Javaux, M. *et al.* (2008) Use of a three-dimensional detailed modeling approach for predicting root water uptake. *Vadose Zone Journal* 7, 1079–1088
- 78 Lobet, G. *et al.* (2011) A novel image analysis toolbox enabling quantitative analysis of root system architecture. *Plant physiology*
- 79 Lièvre, M. *et al.* (2016) Identifying developmental phases in the *Arabidopsis thaliana* rosette using integrative segmentation models. *New Phytologist* at <<http://onlinelibrary.wiley.com/doi/10.1111/nph.13861/pdf>>
- 80 Reymond, M. *et al.* (2003) Combining quantitative trait Loci analysis and an ecophysiological model to analyze the genetic variability of the responses of maize leaf growth to temperature and water deficit. *Plant Physiol* 131, 664–75
- 81 Sadok, W. *et al.* (2007) Leaf growth rate per unit thermal time follows QTL-dependent daily patterns in hundreds of maize lines under naturally fluctuating conditions. *Plant, Cell & Environment* 30, 135–146
- 82 Jia, X. *et al.* (2018) Greater lateral root branching density in maize improves phosphorus acquisition from low phosphorus soil. *J Exp Bot* 69, 4961–4970
- 83 Draye, X. *et al.* (2010) Model-assisted integration of physiological and environmental constraints affecting the dynamic and spatial patterns of root water uptake from soils. *J Exp Bot* 61, 2145–2155

- 84 Doussan, C. *et al.* (1998) Modelling of the Hydraulic Architecture of Root Systems: An Integrated Approach to Water Absorption—Distribution of Axial and Radial Conductances in Maize. *Ann Bot* 81, 225–232
- 85 Lobet, G. *et al.* (2014) Plant water uptake in drying soils. *Plant Physiology*
- 86 Lynch, J.P. *et al.* (2014) Root anatomical phenes associated with water acquisition from drying soil: targets for crop improvement. *J Exp Bot* 65, 6155–6166
- 87 Zhan, A. *et al.* (2015) Reduced lateral root branching density improves drought tolerance in maize. *Plant Physiology* 168, 1603–1615
- 88 Lambers, H. *et al.* (2006) Root Structure and Functioning for Efficient Acquisition of Phosphorus: Matching Morphological and Physiological Traits. *Ann Bot* 98, 693–713
- 89 Pagès, L. (2011) Links between root developmental traits and foraging performance. *Plant, Cell & Environment* 34, 1749–1760
- 90 Nestler, J. *et al.* (2016) Root hair formation in rice (*Oryza sativa* L.) differs between root types and is altered in artificial growth conditions. *J Exp Bot* 67, 3699–3708
- 91 Nestler, J. and Wissuwa, M. (2016) Superior root hair formation confers root efficiency in some, but not all, rice genotypes upon P deficiency. *Frontiers in plant science* 7, 1935
- 92 Mori, A. *et al.* (2016) The role of root size versus root efficiency in phosphorus acquisition in rice. *J Exp Bot* 67, 1179–1189
- 93 McNickle, G.G. *et al.* (2009) Focusing the metaphor: plant root foraging behaviour. *Trends in Ecology & Evolution* 24, 419–426
- 94 Linkohr, B.I. *et al.* Nitrate and phosphate availability and distribution have different effects on root system architecture of *Arabidopsis*. *The Plant Journal* 29, 751–760
- 95 Dener, E. *et al.* (2016) Pea plants show risk sensitivity. *Current Biology* 26, 1763–1767
- 96 Bengough, A.G. *et al.* (2011) Root elongation, water stress, and mechanical impedance: a review of limiting stresses and beneficial root tip traits. *J Exp Bot* 62, 59–68
- 97 Hirth, J.R. *et al.* (2005) Ability of seedling roots of *Lolium perenne* L. to penetrate soil from artificial biopores is modified by soil bulk density, biopore angle and biopore relief. *Plant and Soil* 272, 327–336
- 98 Whiteley, G.M. *et al.* (1982) The buckling of plant roots. *Physiologia Plantarum* 54, 333–342
- 99 Tanaka, S. *et al.* (1993) Cultivar difference in the response of root system to nitrogen application in rice plant. *Jpn J Crop Sci* 62, 447–455
- 100 Tanaka, S. *et al.* (1995) Root system morphology of four rice cultivars: response of different component roots to nitrogen application. *Jpn. J. Crop Sci.* 64, 148–155
- 101 Motte, H. *et al.* (2019) Molecular and Environmental Regulation of Root Development. *Annual review of plant biology* 70,
- 102 Okamoto, S. *et al.* (2016) Long-distance peptide signaling essential for nutrient homeostasis in plants. *Current opinion in plant biology* 34, 35–40
- 103 Kehr, J. and Kragler, F. (2018) Long distance RNA movement. *New Phytologist* 218, 29–40
- 104 Lobet, G. *et al.* (2015) Root System Markup Language: toward a unified root architecture description language. *Plant Physiology*
- 105 Atkinson, J.A. *et al.* (2015) Phenotyping pipeline reveals major seedling root growth QTL in hexaploid wheat. *Journal of Experimental Botany* 66, 2283–2292
- 106 Le Marié, C. *et al.* (2016) RADIX: rhizoslide platform allowing high throughput digital image analysis of root system expansion. *Plant methods* 12, 40
- 107 Clark, R.T. *et al.* (2013) High-throughput two-dimensional root system phenotyping platform facilitates genetic analysis of root growth and development. *Plant, cell & environment* 36, 454–466
- 108 Trachsel, S. *et al.* (2011) Shovelomics: high throughput phenotyping of maize (*Zea mays* L.) root architecture in the field. *Plant and Soil* 341, 75–87
- 109 Barczy, J.-F. *et al.* (2008) AmapSim: a structural whole-plant simulator based on botanical knowledge and designed to host external functional models. *Ann. Bot.* 101, 1125–1138

- 110 Diggle, A.J. (1988) ROOTMAP-a model in three-dimensional coordinates of the growth and structure of fibrous root systems. *Plant Soil* 105, 169178
- 111 Gu, D. *et al.* (2017) Quantitative classification of rice (*Oryza sativa* L.) root length and diameter using image analysis. *PloS one* 12, e0169968

Table 1. Structural functional plant models able to generate lateral roots random diversity

Model name	Growth stochasticity ^a	Various lateral root types ^b	Lateral root types / proportions modified by environment ^c	Refs
AMAPsim	Yes	No	No	[109]
ArchiSimple	Yes	Yes (continuum)	No	[72]
CRootBox	Yes	Yes	No	[73]
DigR	Yes	Yes	No	[74]
OpenSimRoot	Yes	Yes	No	[75]
RootMap	No	No	No	[110]
RootTyp	Yes	Yes	No	[76]

^a *Growth stochasticity* means that growth parameters are not fixed, but chosen randomly within a given distribution. ^b *Various lateral root types* refers to the possibility, for the user, to explicitly input several lateral root types for each mother root. ^c *Lateral root types / proportions modified by environment* refers to the ability of the model to modulate the types and proportions of lateral roots based on environmental clues.

Figure 1. Structured and random diversity of lateral roots in plants. (A): Structured (right) and random (left) morphological and anatomical diversity of lateral roots as intrinsic feature of root system architecture in plants. In the structured diversity component, 1st order laterals are longer and thicker than 2nd order etc. Within a root order, root lengths and growth patterns are essentially determined by the age of the lateral root and thus the distance from the apex of the parent root causing a highly regular pattern. In the random diversity component, presumably caused by developmental instability, length and anatomy differ among roots of the same order, branched on the same parent root and of the same age. Both structured and random diversity take place in the absence of environmental heterogeneity and is referred to as intrinsic diversity. Random diversity is detectable in most if not all plant species. Illustrations are given for monocots ((B): rice, and (C): maize), annual dicot ((D): pea and tree dicot (E: oak tree,) with arrows pointing towards long (red), short (green) or intermediate (blue) lateral roots (scale bar = 5 mm). Diversity in length can be associated with diversity in anatomy as illustrated with transverse sections in three types of lateral roots in pearl millet (F, Scale bar: 20 μ m, note that for the large root on the left, only the stele is shown). Within species, there exists large genetic variability (upper panel, plain grey arrows) for structured

diversity (*eg.* genotypes with on average longer or shorter laterals, longer or shorter apical unbranched zone). By contrast, little is known (dotted grey arrow) about the genetic variability of random diversity (*ie.* within a species, are there genotypes with more or less diverse laterals). In response to environmental heterogeneity, structured and random diversity respond in a different way. On the structured component of diversity, all roots essentially respond in the same way. An example of this is the ‘xerobranching’ pattern leading to a complete absence of primordia due to transient lack of water ([20], (H) shows an image of a barley root). Another example is the the response of cluster roots to low P in white lupin [23]. By contrast, in the random component, different roots respond differently. This is illustrated by the branching patterns (G) along primary roots of maize exposed to high or low N in split-roots system, where longer lateral roots respond much more steeply to high N. Whereas structured diversity is essentially studied through the filter of an average behaviour, random diversity must be seen through the lens of distributions (of lengths, growth rates etc..) as illustrated by the small inset sowing bars or distributions. Both structured and random diversity certainly contribute to efficiency in response to environmental heterogeneity and unpredictability. All pictures from the authors except (B) from [111], (D) courtesy C Salon and (F) from [55].

Figure 2. Longitudinal sequence of stochastic events contributing to the diversity of lateral roots branched along a primary root and formalized using a Markov chain. 0. establishment of pre-branch sites following oscillations of the response to auxin in a zone located between the basal meristem and the elongation zone with a probability $p_{(pbs)}$. 1. Once initiated, primordia develop with a diversity of rates, here arbitrarily discretized into fast (1a), intermediate (1b) or slow (1c) associated with corresponding probabilities ($p_{(a)}$, $p_{(b)}$, $p_{(c)}$). 2. Primordia may or may not emerge. If emerged, lateral roots grow with a diversity of rates for more or less long durations. According to [7], this diversity is not homogenous but can be discretized into types (here three, set as fast and accelerating (2A), slow and rapidly stopping (2C) and slowly decelerating (2B). Again, each state is reached with a probability that also depends on the previous status (for instance, $p_{(A|b)}$ is the probability to enter the stage 2A after being a primordia under the 1b status). An ultimate stage (3) represents arrested roots. Diversity of growth patterns translate into a divergence of lateral root length after a few days (red, blue and green dotted lines showing the shape of the envelope of root tips of the three types). A

Markov chain is represented by straight arrows when an object (primordium, root) moves to a new status and by a loop (called self-transitions) when the object remain in the same status until the next time step. Dashed arrows correspond to the less probable transitions. The coloured loops and arrows correspond to non-homogeneous transition distributions (i.e. distributions are time-dependent, consistent with the idea of developmental window when 'decisions' are made). The self-transitions whose probabilities increase with time and converge to 1 are in orange. This is the case of the prebranch sites which have a delineated window to initiate primordia. Similarly, slowly developing primordia have probability to arrest which increases with time. As a consequence, probabilities to exit these states converge to zero (orange arrows). For states 2A and 2B, the self-transition probability decreases then stabilizes to a value > 0 (pink loop), while the probability of moving to the growth arrest state (pink arrow) increases then stabilizes to a value < 1 , the asymptotic probability of staying in state A being far higher than that to remain in state B.

Figure 3. A journey through a progressive decrease of entropy during primordia development. The entropy characterizes the degree of totipotency of a group of cells (pre-branch site, primordium or root tip). The journey starts with the targeting (0B) or not (0A) of pre-branch sites (PBS) with a slight (0B) or steep (0A) decrease of entropy. PBS (grey beads) that do not give rise to primordia (black beads) remain stuck in a stable state in an entropy sink. Initiated primordia develop with different trajectories from large/rapid primordia to small and/or slow growing primordia with possible intermediates (a-red, c-green and b-blue respectively). Entropy sufficiently decreases so that changes in primordia fate becomes more difficult (symbolized as beads being trapped in an entropy sink). Then, lateral roots emerge (2) but some primordia do not (additional black beads). At that stage, the fate of lateral roots from extreme types is no longer flexible while intermediate ones can still behave differently based on external stimuli. The journey can be altered upon environmental and/or hormonal treatments. For instance, the supply of IAA to the root, the excision of the primary root tip or the local bending of the primary root induce a massive flush of IAA and a drop of entropy (1'). This drives all developing primordia to behaving in the same way (large / fast growing). Sugars also stimulate fast development of primordia. By contrast, IAA has a much lesser impact on emerged laterals since it occurs at a stage where entropy is too low to allow redirection of fate (2'). Impact of signals is also seen at earlier stages such as during hydropatterning or xerobranhing with ABA being an intermediate inhibiting primordia initiation. In that case, all PBS fall in an entropy sink.

Figure 4. Modelling approaches to capture lateral root random diversity. (A). Representation of two extreme branching patterns along a dicot taproot, either highly diverse (left) or not (right) generated using the ArchiSimple model [70]. The only difference between the two root systems is the variance of lateral root diameter at emergence which is 2,5 higher on the left than on the right root system. (B). Model-based characterization of the diversity of lateral root growth patterns. The diversity of growth patterns in pearl millet is represented by the upper right panel which shows growth trajectories of all laterals from one single maize primary root (grey lines). Four contrasted patterns are shown in black. This diversity is analysed using Semi-Markov switching linear model (SMS-LM). In that case, a four-state SMS-LM best accounts for the diversity of patterns [11]. The three growth states correspond to lateral root types called A, (fast and accelerating pattern), B (intermediate, slowly declining) and C (rapidly arrested). All roots may ultimately stop growing (consistent with the formalization shown in Fig 2). The left part of the panel shows (top to bottom) (i) the distribution of growth phase duration, that is much longer for A types than for B or C type roots, (ii) the graphs of transitions with probabilities associated with transitions states and (iii) the linear trend models estimated for each state. With this model, each root can be tagged and assigned to a given type (lower right panel).

Figure 5: Proposed contributions of lateral roots random diversity to efficiency. (A). LR with different sizes and hydraulic properties (left) complement each other to smooth the spatial distribution of root water uptake. (B) and (C). Diverse sets of lateral roots (left) explore a given soil volume and acquire a limiting resource at a reduced cost. (D). An intrinsic diversity (left) increases the likelihood for lateral roots to better seek out cracks and pores and to minimise the likelihood of buckling. (E). Low cost - low gain foraging units (left) are risk averse in stochastic environments. (F). In rich environments, plants can afford high cost - high gain foraging units (right). (G). A set of lateral roots with diverse growth rates (left) sense soil properties with a better coverage of the soil volume then if they had the same growth pattern (right).

Glossary

Developmental instability. It refers to the ability of an individual to develop non identical phenotype under given environmental and genetic conditions.

Entropy. *Sensu* Shanon, entropy is a measure of unpredictability of the state, or equivalently, of its average information content.

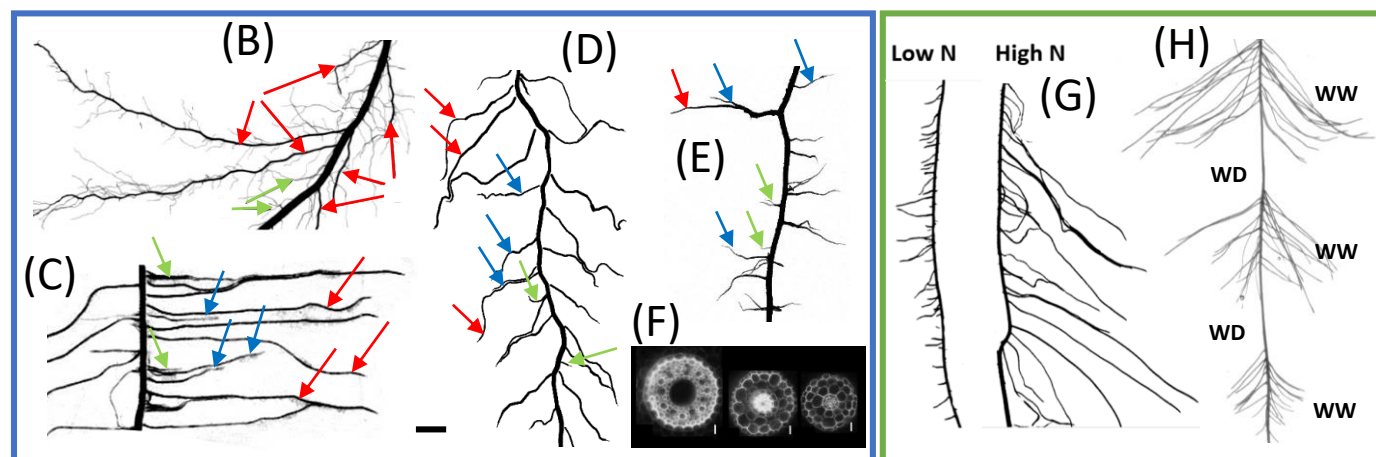
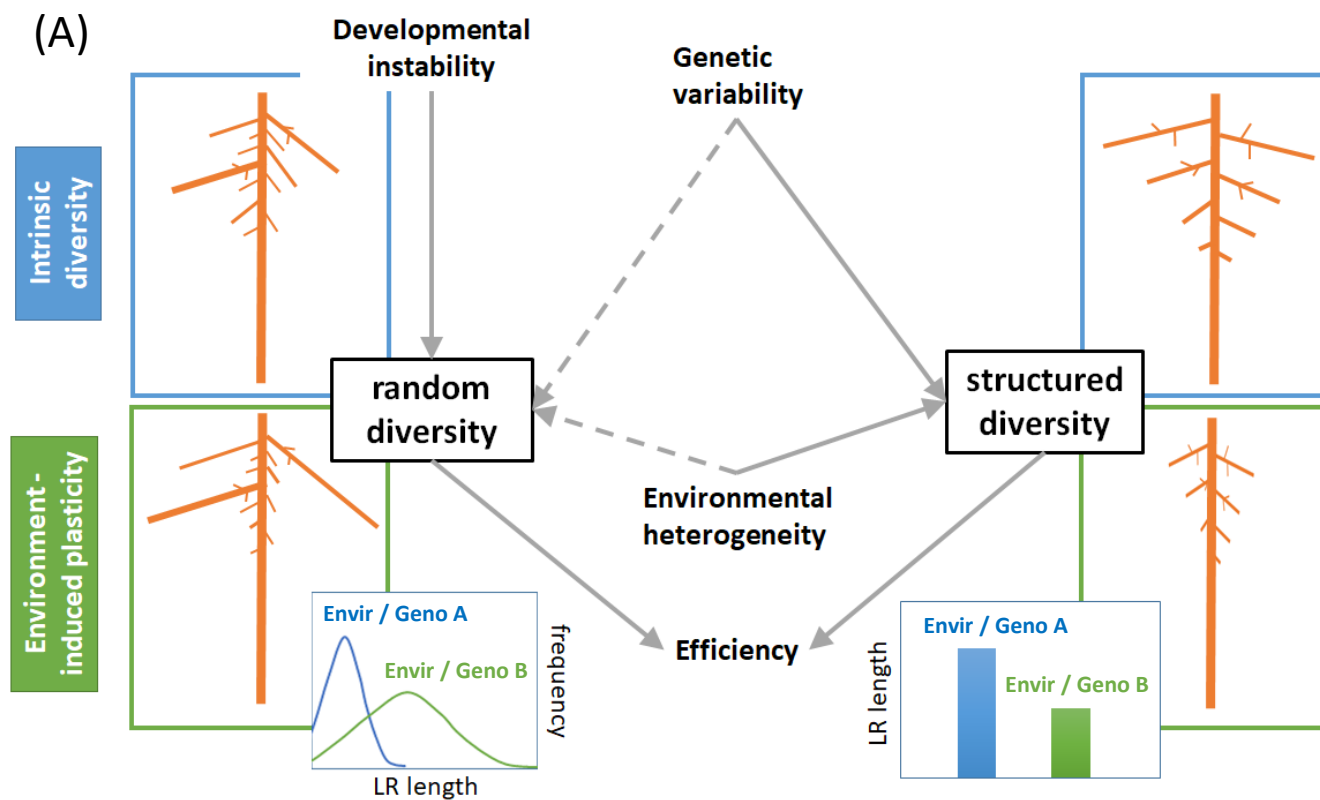
Hydropatterning. Plants can sense microscale heterogeneity in water availability across their circumference, which causes dramatic differences in the patterning of tissues along this axis, in particular branching. Auxin biosynthesis and transport is essential for regulating this process.

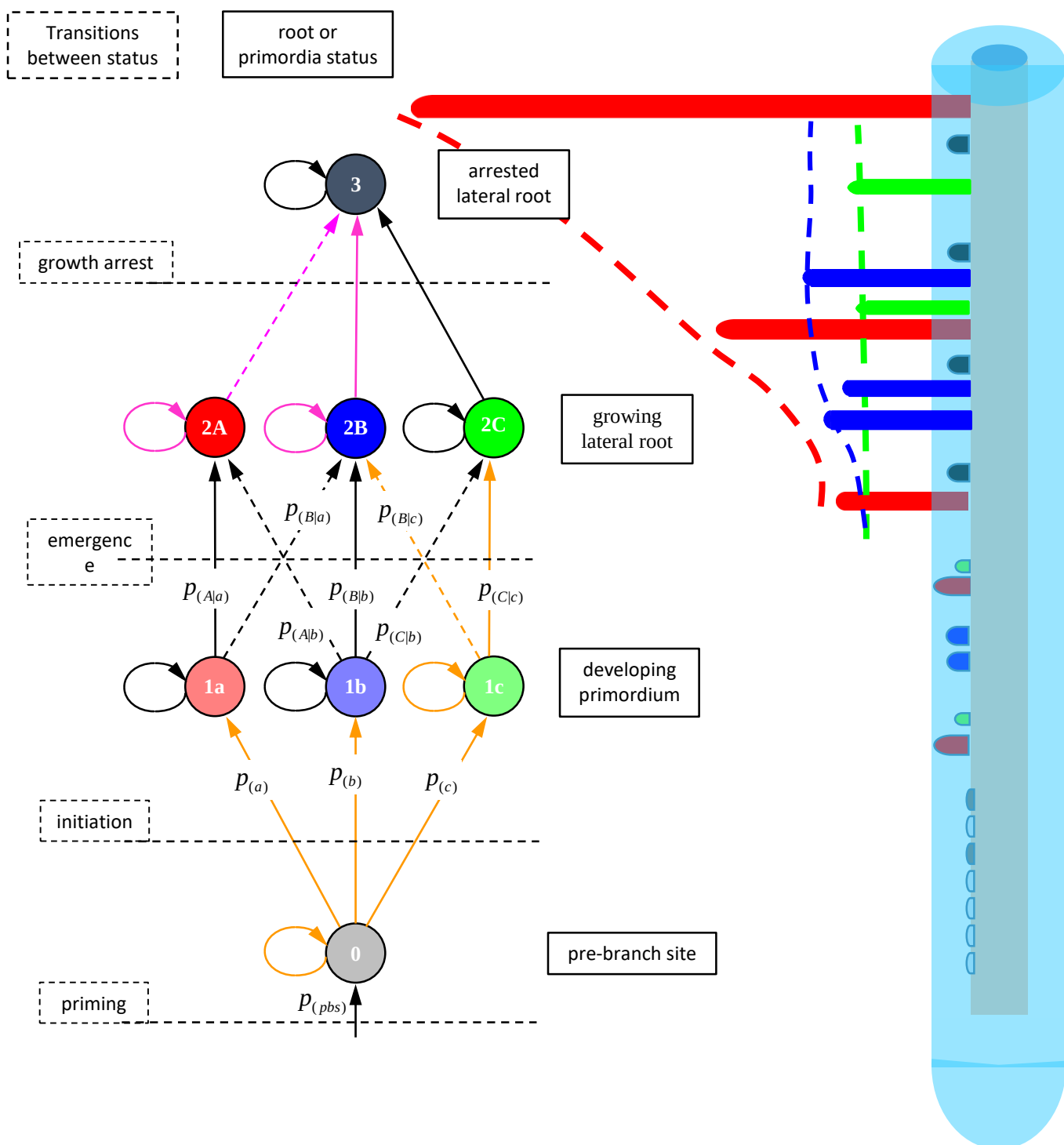
Markov chain. A Markov chain is a stochastic model describing a sequence of possible events in which each event depends only on the previous event.

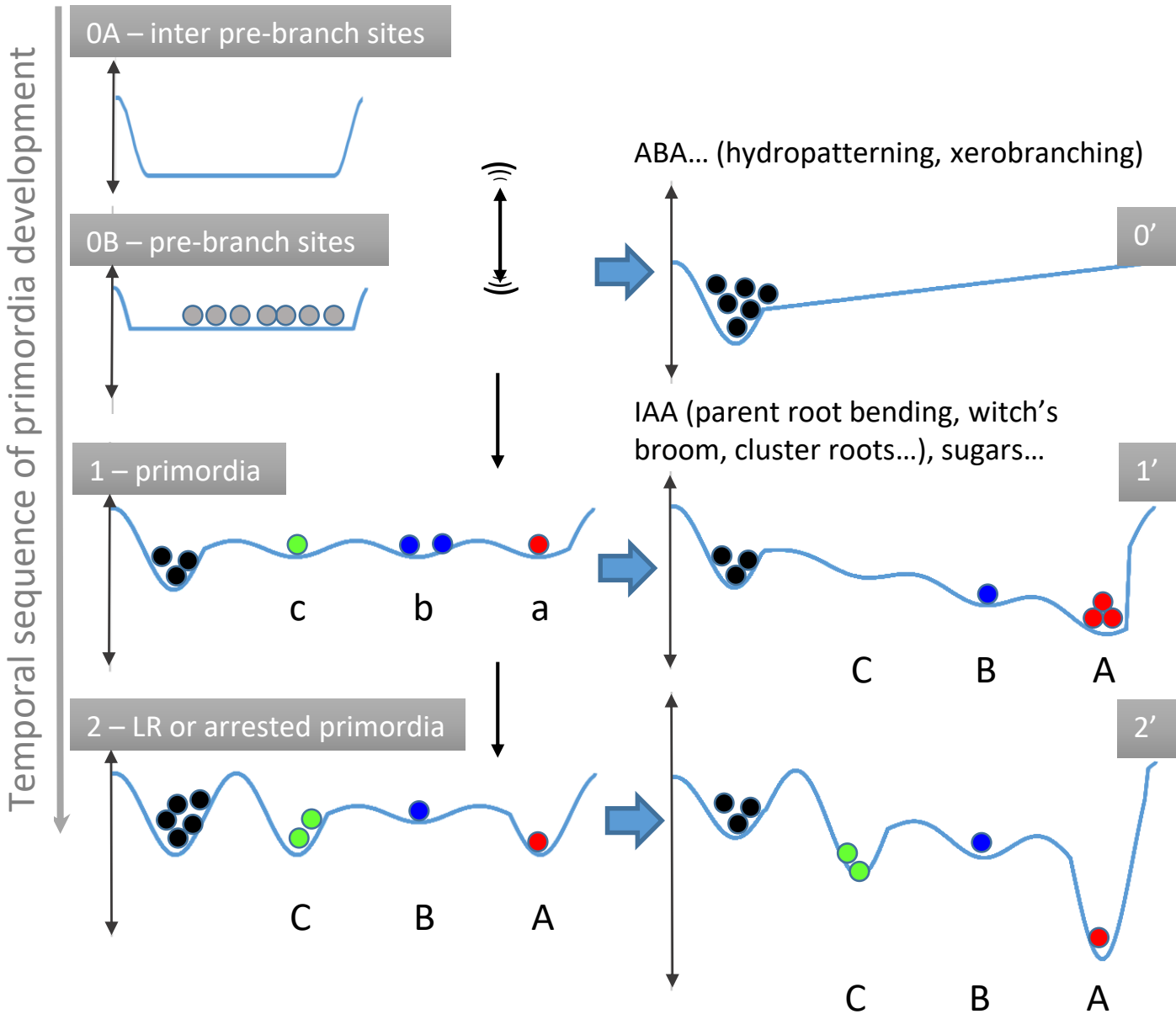
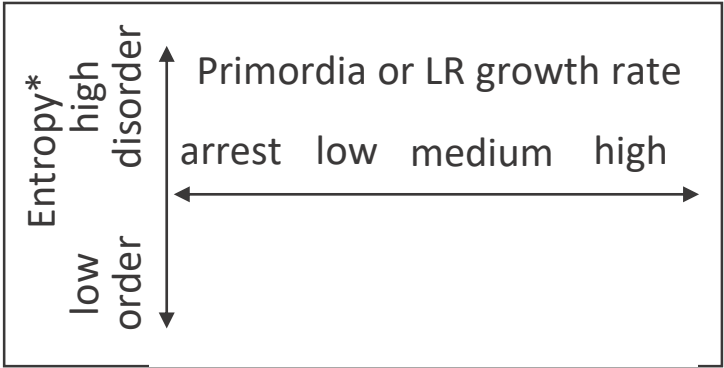
Stochastic. Refers to the randomness of a process.

Xerobbranching. An ABA-dependent adaptive mechanism allowing roots to rapidly respond to changes in water availability in their local micro-environment

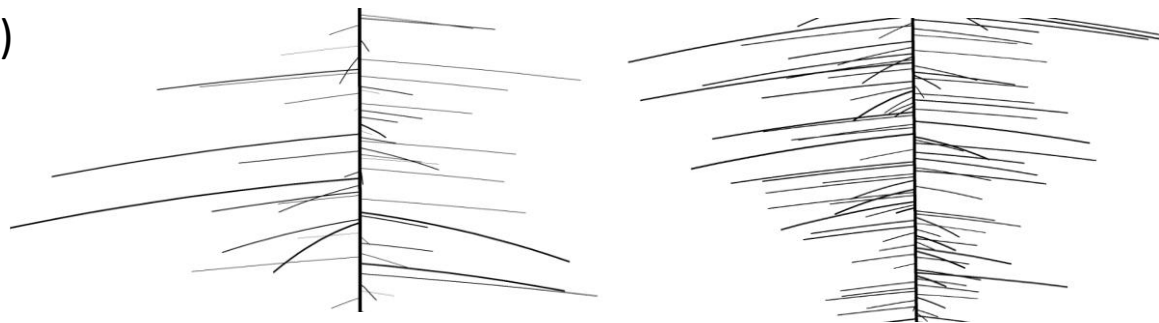
(A)



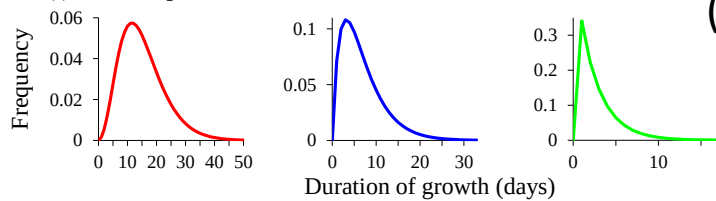




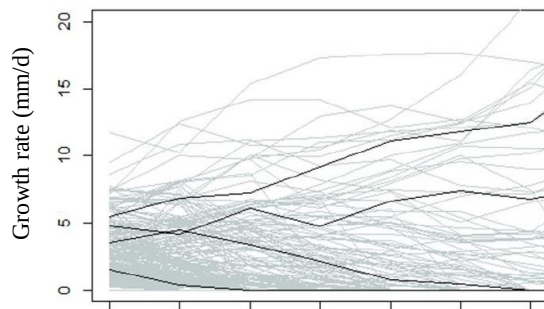
(A)



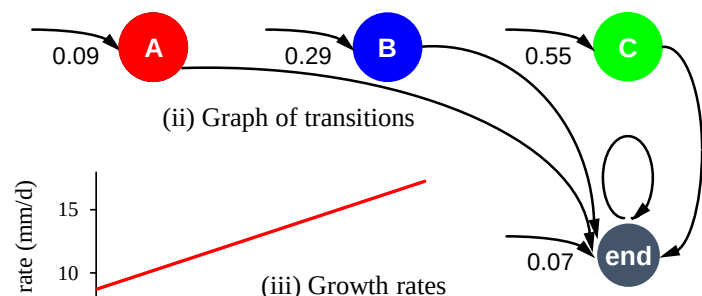
(i) Growth phase succession and durations



(B)



(ii) Graph of transitions



(iii) Growth rates within phases

