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## To cite this version:

Brigitte Picard, Mohammed Gagaoua, Marwa Al Jammas, Muriel Bonnet. Beef tenderness and intramuscular fat proteomic biomarkers: Effect of gender and rearing practices. Journal of Proteomics, 2019, 200, pp.1-10. 10.1016/j.jprot.2019.03.010 . hal-02627558

**HAL Id: hal-02627558**

**https://hal.inrae.fr/hal-02627558v1**

Submitted on 22 Oct 2021

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Version of Record: https://www.sciencedirect.com/science/article/pii/S1874391919300910
Manuscript_20304f4a9c6363c87472551ccf08d772

# Beef tenderness and intramuscular fat proteomic biomarkers: effect of gender and rearing practices

Brigitte Picard*, Mohammed Gagaoua**, Marwa Al Jammas, Muriel Bonnet

Université Clermont Auvergne, INRA, VetAgro Sup, UMR Herbivores, Saint-Genès-Champanelle, France

**Correspondence:**

* Dr. Brigitte Picard: brigitte.picard@inra.fr
ORCID: 0000-0002-8075-671

## ABSTRACT

This study analyzed the effect of gender on the abundances of 20 protein biomarkers of tenderness and/or intramuscular fat content in five muscles: *Longissimus thoracis*, *Semimembranosus*, *Rectus abdominis*, *Triceps brachii* and *Semitendinosus*, from cows and steers of the Protected Designation Origin Maine Anjou. The protein abundances were quantified using Reverse Phase Protein Array with specific validated antibodies. Among the 20 studied proteins, the abundance of 8 biomarkers involved in energetic metabolism, contraction and cellular stress, was different according to gender. The gender effect was different depending on the muscle type with greater abundances in *Semitendinosus*, *Rectus abdominis* and *Longissimus thoracis* muscles. On the basis of animal characteristics and rearing factors, three rearing practices classes were identified for cows. Among the factors, fattening duration modified the abundance of 12 proteins mainly in *Triceps brachii* muscle. A positive correlation between the abundance of the small HSP20 and slaughter age was observed in the 5 muscles. Two proteins, Four and a half LIM domains 1 (FHL1) and Glycogen phosphorylase (PYGB) appeared to be muscle, gender and rearing practices independent. These results constitute valuable data to understand how to manage beef quality by controlling these different factors.

**Keywords:** Beef proteome, Gender, Skeletal muscles, Rearing factors, Proteomics, Reverse Phase Protein Array

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37 **SIGNIFICANCE**

38 This study is the first to compare the relative abundance of 20 proteins previously
39 identified as biomarkers of tenderness and/or intramuscular fat (IMF) content of beef meat
40 between cows and steers among 5 different muscles. Its originality is in the use of Reverse
41 Phase Protein Array for fast quantification of the proteins and the integration of data from
42 rearing factors, carcass characteristics and biomarkers of meat qualities. The findings provide
43 evidence for modulating biomarker levels by controlling the choice of animal type and rearing
44 factors according to the type of muscle that would produce animals with the desired meat
45 qualities.

46 **1. Introduction**

47 The control of meat qualities is a societal issue that concerns all the meat sectors. Meat
48 qualities are defined by a set of intrinsic and extrinsic properties where the former correspond to
49 safety, health, convenience, nutritional and sensorial qualities; and the later are associated with
50 the product and production system characteristics from the farm-to-fork, including animal
51 welfare, carbon footprint and marketing variables (for review: [1-3]). For beef meat, the most
52 crucial quality traits are tenderness and marbling associated with intramuscular fat (IMF)
53 content. Tenderness defined as the ease with which meat can be sliced or chewed, is a
54 multifactorial quality criterion the most variable and therefore the most difficult to control or
55 predict. The appreciation of beef tenderness is generally positively associated with IMF content,
56 the decrease in IMF content can also reduce tenderness [4]. Indeed, a minimum amount of IMF
57 is needed for the expression of beef flavor as well as better tenderness [5]. IMF also plays an
58 important role in beef juiciness, meat with high IMF content is always less dry than lean meat.
59 Despite industry efforts to control the eating quality of beef, a high level of variability remains
60 in these quality traits, which is one reason for consumer dissatisfaction. Thus, for producers and
61 consumers, the control and management of beef tenderness and IMF content constitute a
62 challenging task for better sustainability of the beef sector.

63 The large literature reported that those beef qualities are the result of complex biological
64 mechanisms involved in muscle biochemistry in the live animals and after slaughtering during
65 the aging period [6, 7]. Over the last decades, numerous studies have analyzed the factors
66 affecting these traits. The effect of factors related to the animal and its production systems such
67 as muscle type, breed, age, sex, physiological stage of animals, nutritional diet, physical activity
68 and fattening duration has been investigated [2, 8]. The earlier results reported that early
69 maturing Anglo-Saxon breeds such as Aberdeen Angus or Japanese black cattle, are
70 characterized with high degree of fatness, on the contrary late maturing breeds such as French

71 beef breeds or double muscled cattle, have high muscle yield and low fatness scores [9]. The
72 development of adipose tissues in some specific muscles appears to disorganize the muscle
73 structure and contributes to tenderization of highly marbled beef during the late fattening period
74 [10]. Increasing age seems to be favorable for juiciness and flavor (due to more intramuscular
75 fat), but unfavorable for tenderness due to connective tissue characteristics despite an
76 attenuation of this effect by high amounts of IMF [11]. Furthermore, gender plays an important
77 role. For example, at the same age, females provide more flavorful, tenderer and intense color
78 beef than steers or bulls[12]. Compared to beef from young bulls, meat from steers contains
79 more IMF [12, 13]. In the large literature, many controversies have been reported regarding the
80 relationships between rearing factors and quality traits, with many conflicting results [8, 12, 14,
81 15] and today there is still no reliable online tools to predict these quality traits and deliver
82 consistent quality beef for consumers. In this context, researches were conducted during the last
83 15 years to better understand the biological mechanisms underpinning tenderness and IMF
84 variability to propose indicators or biomarkers which could be used for their prediction and/or
85 management soon after slaughter of the animals [16, 17]. "Omics" approaches which allow a
86 large number of genes, proteins or metabolites to be simultaneously studied without any *a*
87 *priori*, have been extensively applied (for review [16, 18]). These approaches had revealed that
88 large amount of macromolecules may be potential molecular indicators of muscle mass and
89 growth performance [19], sensory attributes [20-23] or marbling of meat [24, 25]. The question
90 is now how to modulate them in order to control and manage beef quality. The expression or
91 abundance of these biomarkers could be modulated through rearing factors. As the control of
92 the zootechnical performance of animals and the quality of their products is of major economic
93 importance in the context of beef sustainability, the aim of this study was to analyze the gender
94 effect by comparing cows *vs.* steers and link with the rearing factors on the relative abundance
95 of 20 biomarkers of tenderness and/or IMF content in 5 muscles. The proteins were quantified

using the Reverse Phase Protein Array (RPPA) on 101 Protected Designation Origin (PDO) Maine-Anjou cattle [8, 21, 26]. A classification based on rearing factors was applied as described by Gagaoua *et al* [8, 22] to identify rearing practices classes. Then, carcass properties and relative abundances of the biomarkers were analyzed for each class among 5 muscles. The results revealed new insights that could be applied for a better understanding of the biological pathways involved in meat quality according to gender and rearing practices.

## 2. Materials and Methods

### *2.1. Animals, handling and slaughtering*

A total of 101 cattle including 86 cows and 15 steers from the French PDO (Protected Designation of Origin) Maine-Anjou, using "Rouge des Prés" breed [21], were collected [26]. The PDO Maine-Anjou animals originated in the northwestern part of France from a cooperative of livestock farmers located in the department of Maine-et-Loire. This breed was the second (since 2004) among the four breeds allowed to be used in France for PDO meat production. It is composed of around 80% of cows (justifying the high number of animals in this study), younger than 10 years of age, having calved at least once and a minimal carcass weight of 380 kg. Steers over 30 months of age with a carcass weight of 400 kg minimum can also be found (20%). PDOs are of special importance for the valorization of local breeds, and the specifications of animal products under PDO are paid increasing attention [21]. The rearing practices of each animal were surveyed by a questionnaire as detailed in Gagaoua *et al.* [8] based on the study by Couvreur *et al*. Briefly, the questionnaire included variables about (i) the finishing period [part of hay, haylage, and/or grass in the finishing diet (% w/w); total amount of concentrate (kg); fattening duration (days); physical activity of the animals (% days out)] and (ii) the animal characteristics by the age at slaughter in months. Those variables were used to identify rearing practices as detailed in the statistical section of this manuscript.

120 Before slaughter, all animals were food deprived for 24 h and had free access to water. The
121 slaughtering was performed in the same industrial abattoir (Charal, Sablé sur Sarthes, France).
122 The animals were stunned using captive-bolt pistol prior to exsanguination and dressed
123 according to standard commercial practices. The slaughtering was also performed in
124 compliance with the French welfare regulations and respecting EU regulations (Council
125 Regulation (EC) No. 1099/2009).

126 After slaughter, the carcasses were characterized and graded according to the European beef
127 grading system (CE 1249/2008). Thus, information for each carcass were measured, namely hot
128 carcass weight (HCW, kg), EUROP conformation score (EUROP grid), carcass fat weight and
129 fat to muscle ratio (% w/w) as described by Gagaoua *et al.* [11, 27].

130 ***2.2. Muscle sampling***

131 The carcasses were not electrically stimulated and they were chilled at 3 to 4°C until 24 h
132 *post-mortem*. The right half carcass was used for muscles measurements. Then, aliquots of five
133 muscles: *Longissimus thoracis* (LT), *Semimembranosus* (SM), *Rectus abdominis* (RA), *Triceps*
134 *brachii* (TB) and *Semitendinosus* (ST), from each carcass of the 101 PDO Maine Anjou cattle
135 were sampled. These heterogeneous muscles were chosen according to their differences in
136 contractile and metabolic type [26]. The LT muscle was excised from the 6$^{th}$ rib as detailed by
137 Gagaoua *et al.* [27]. As the samples were for omics biomarkers analysis, the muscles (an
138 approximate of 2 g) trimmed of connective and superficial fat tissue were immediately and
139 carefully frozen in liquid nitrogen and stored at -80°C until analysis following the protocol
140 previously described by Picard *et al.* [26].

141 ***2.3. Protein biomarkers quantification by Reverse Phase Protein Array***

142 The relative abundance of 20 protein biomarkers of tenderness and/or IMF content was
143 measured in the 5 muscles by the Reverse Phase Protein Array (RPPA) recently described by

144 our group [20, 22, 26]. The specificity of the 20 antibodies on bovine muscle and their
145 conditions of use have been previously defined by western blotting which uses the same
146 technical principle as the RPPA method [26]. Briefly, the samples were firstly disrupted in a
147 Laemmli buffer containing 50 mM Tris pH =6.8, 2% SDS, 5% glycerol, 2 mM DTT, 2.5 mM
148 EDTA, 2.5 mM EGTA, 1x HALT Phosphatase inhibitor (Perbio 78420), Protease inhibitor
149 cocktail complete MINI EDTA-free (Roche 1836170, 1 tablet/10 mL), 2 mM $Na_3VO_4$ and 10
150 mM NaF, using a Precellys (Bertin). Extracts were then boiled for 10 min at 100°C, sonicated
151 to reduce viscosity and centrifuged 10 min at 15000 rpm. The supernatant was harvested and
152 stored at -80°C. Protein concentration was determined using the Pierce BCA reducing agent
153 compatible kit (ref 23252).

154 The sample extracts were then deposited onto nitrocellulose covered slides (Supernova,
155 Grace Biolabs) using a dedicated arrayer (2470 arrayer, Aushon Biosystems). Four serial
156 dilutions, ranging from 2000 to 250 µg/ml, and two technical replicates per dilution were
157 printed for each sample. Arrays were labeled with each of the 20 specific antibodies or without
158 primary antibody (negative control), using an Autostainer Plus (Dako) as detailed in our
159 previous papers [20, 22, 26]. After protein quantification by RPPA, the raw data were
160 normalized using Normacurve following the procedure described by [28], which normalizes for
161 fluorescent background per spot, a total protein stain and potential spatial bias on the slide.
162 Next, each RPPA slide was median centered and scaled (divided by median absolute deviation).
163 We then corrected for remaining sample loadings effects individually for each array by
164 correcting the dependency of the data for individual arrays on the median value of each sample
165 over all 20 arrays using a linear regression.

166 ***2.5. Statistical analysis***

167 The statistical analyses were performed using SAS statistical software (SAS 9.1, SAS
168 Institute INC, Cary, NC, USA) and XLSTAT 2017.19.4 (AddinSoft, Paris, France). Before

169 analysis, raw data means were scrutinized for data entry errors and outliers. Normal distribution
170 and homogeneity of the dataset was first tested by the Shapiro-Wilk test ($P > 0.05$). The PROC
171 GLM procedure of SAS was then used to study the muscle type (5 muscles), gender (cows *vs.*
172 steers) and interactions effects on the relative abundances of the proteins. Significant
173 differences among muscles were performed using Tukey's test at a significance level of $P <$
174 $0.05$. Similarly, the protein abundances were further compared between the two genders within
175 each muscle separately and the effect of rearing practices on the abundances of the 20 proteins
176 was analyzed for both cows and steers.

177 For the 86 cows only, rearing practices classes were created using the statistical approach
178 described by Gagaoua *et al.* [8, 20] based on principal component analysis (PCA) combined to
179 *k*-means clustering. For that, the fattening period data (part of hay, haylage and/or grass in the
180 finishing diet (% w/w)); total amount of concentrate (kg); duration (days) and physical activity
181 (% days out) of the animals at the farm were used [8]. Two factors with eigenvalues >1.0 were
182 extracted on the basis of the scree plot and evaluation of the factor loading matrix after
183 orthogonal rotation. These allowed us to identify using Z-scores on the two axis 3 rearing
184 practices that were named to simplify the discussion as follow: Class 1= "Hay class"; Class 2 =
185 "Grass class", and Class 3 = "Haylage class", respectively (Table 3). Z-scores represent the
186 deviation of each observation relative to the mean of the corresponding individual in each
187 rearing practice and were calculated using PROC STANDARD of SAS that standardizes data to
188 a mean of 0 and standard deviation of 1. These normalized data were used to build PCAs to
189 depict the relationships between the rearing practices of the 86 PDO Maine-Anjou cows with i)
190 animal, rearing factors and carcass characteristics, and with ii) the 20 protein biomarkers from
191 the 5 muscles quantified by RPPA technique within the rearing factors. The Kaiser-Meyer-
192 Olkin (KMO) measure, known also as Kaiser's Measure of Sampling Adequacy (MSA) was
193 applied to test the validity of the sampling [29]. Subsequently, unsupervised hierarchical

194 clustering heatmap was generated using the same data to assess the differences among the 5
195 muscles based on the normalized data for each rearing practice. For the 15 steers, only two
196 rearing practices were identified (grass (n = 5) and haylage (n = 10)) and were considered in the
197 analyses in same manner than cows.

198 Finally, the PROC CORR of SAS after Z-scores calculation was used to compute the
199 Pearson's correlations of coefficients between the 20 proteins and the animal, rearing factors
200 and carcass characteristics of the whole data of the 86 cows. Correlation coefficients were
201 considered significant at $P < 0.05$.

202 **3. Results and discussion**

203 ***3.1. Gender effect***

204 The gender (cows *vs.* steers) had a highly significant effect on the relative abundance of 8
205 proteins among the 20 analyzed: HSP20, PGK1 ($P<0.001$), PRDX6, ALDOA ($P<0.01$), MDH1,
206 TPI1, MyHC-IIX, TNNT1 ($P<0.05$) (Table 2). All muscle combined, the cows comparatively to
207 steers had significantly ($P< 0.01$) higher abundance of HSP20, ALDOA, MDH1, MyHC-IIX,
208 and lower abundance of PGK1, PRDX6, TPI1 and TNNT1 (Table 2).

209 Of the 20 proteins analyzed, only HSP20 had an abundance that differed between steers and
210 cows irrespective of the considered muscle. An interaction of muscle x gender was observed for
211 this protein (Table 2) which was more abundant in cows for LT, SM, ST muscles, and was not
212 different in RA and TB. Figure 1 illustrates higher differences between muscles in steers than in
213 cows. In the two genders, the abundance of HSP20 was the highest in RA muscle. On another
214 hand, the abundances of CRYAB, HSP27, HSP40, HSP70-1, FHL1, TRIM72, PYGB,
215 ALDH1A1, ENO3, TTN, MLC1F and α-tubulin were not different between steers and cows.

216 Our results showed that the muscles of cows comparatively to steers differed by the
217 abundance of 8 proteins among the 20 analyzed. Thus, gender affects less proteins than muscle

218 type reported to modify the abundance of 16 of 20 proteins, only 4 proteins namely HSP40
219 (Heat shock protein), FHL1 (Four and a half LIM domains protein 1), PYGB (Glycogen
220 phosphorylase B) and MDH1 (Malate dehydrogenase), were found to do not differ among the 5
221 muscles [26]. Thus, according to these two studies, HSP40, FHL1 and PYGB were not
222 modified either by gender or muscle type while HSP20, PRDX6, PGK1, ALDOA, MyHC-IIX,
223 TNNT1 and TPI1 showed both muscle and gender effects.

224 The analysis of gender effect in each of the 5 muscles showed that it was most important for
225 ST, RA, and LT muscles. It is particularly significant for HSP20 which abundance between
226 cows and steers was not modified in TB and RA muscles and was significantly different
227 between the two genders for the three other muscles. In TB muscle, the abundances of 19 of 20
228 proteins were not different between cows and steers. This indicates that this muscle is
229 insensitive to the sex or gender effect (Table 2). This result is coherent with previous data of our
230 group showing no effect of castration on contractile and metabolic properties of TB muscle
231 while the effect of castration was the greatest in ST and LT muscles [30], in accordance with
232 the results of the present study. Indeed, in the present study, the most important differences
233 between the two genders were observed in ST muscle as the abundances of 8 proteins were
234 different between cows and steers, whereas 6 were different in LT and RA and 5 in SM muscle.
235 To our knowledge, very few studies in the literature have compared the muscle proteome
236 properties of cows comparatively to steers in different muscle types. Previous results of our
237 group showed that RA muscle of heifers comparatively to steers, was more oxidative with
238 greatest ICDH and COX activities and less glycolytic with a lowest LDH activity [21, 31].
239 These data are coherent with the present results showing modifications of contractile [MyHC-
240 IIX (fast glycolytic isoform), TNNT1 (slow isoform)] and metabolic [ALDOA (glycolytic
241 enzyme involved in glycogen storage), MDH1 (involved in tricarboxylic acid cycle), PGK1
242 (glycolytic enzyme) and TPI1 (involved in gluconeogenesis and carbohydrate biosynthesis)]

243 properties of the muscles between cows and steers. This effect could be explained mainly by
244 differences in sex hormones between the two genders. The effect of estrogens on skeletal
245 muscle properties has been largely studied in different species [32, 33]. Indeed, estrogens and
246 their receptors play key roles in the regulation of energy metabolism pathways, including
247 glucose transport, glycolysis, tricarboxylic acid cycle, mitochondrial respiratory chain,
248 adenosine nucleotide translocator and fatty acid β-oxidation and synthesis [34]. A higher insulin
249 sensitivity was also reported in female, and the ratio of glycolytic/oxidative enzyme activities
250 within skeletal muscle correlated negatively. These modifications in muscle physiology induced
251 by estrogens are in accordance with the modifications in protein abundances observed in this
252 study.

253 Among the differential proteins, HSP20 and PGK1 showed and all muscles confounded the
254 highest differences between the two genders (Table 2). To the best of our knowledge, only one
255 publication reported a higher abundance of HSP20 (*HSPB6* gene) and a lower abundance of
256 PGK1 in muscle from women than men as observed for cattle in this study [35]. Few data are
257 available in the literature about the effect of castration or estrogens on *HSPB6* gene expression
258 (HSP20). In line to this scarcity of studies in the large literature, a recent review by Gianazza *et*
259 *al.* [36] reported that the first proteomic survey on the proteome of male *vs* female serum in
260 humans is also as recent as 2010 [37]. Therefore, it is difficult to compare the findings of this
261 study to the literature.

262 The findings of HSP20 protein may be partly linked to its binding to structural proteins such
263 as TNNT1 [38]. These data are coherent with the differences observed between cows and steers
264 for both HSP20 and TNNT1. Moreover, earlier studies demonstrated that HSP20 is
265 phosphorylated in response to insulin in skeletal muscle [39] and the authors proposed HSP20
266 as a potential modulator of insulin's functions. The differences in TNNT1 abundance between
267 cows and steers could be the consequence of insulin sensitivity induced by estrogens. The

268 action of estrogen is also through circulating adipokines as adiponectin and leptin which levels
269 are higher in females [40]. These adipokines are involved in muscle metabolism and fat
270 deposition.

271 The main effect of gender in the present study was observed for PGK1 as it is the only
272 protein among the 20 analyzed which was more abundant in steers comparatively to cows in
273 each of the 5 muscles. This protein is involved in glycolysis as it is the first ATP-generating
274 enzyme in the glycolytic pathway, catalyzing the conversion of 1,3-diphosphoglycerate to 3-
275 phosphoglycerate. It has been recently shown that PGK1 translocates to the mitochondria where
276 it specifically phosphorylates pyruvate dehydrogenase kinase [41]. These data are in accordance
277 with a high effect of sex hormone on glucose metabolism [42] that would also be linked to IMF
278 deposition within steers [43]. Several data of the literature indicated that castrated cattle have
279 higher fast-twitch glycolytic fiber proportion and lower slow-twitch oxidative fiber than intact
280 males.

281 ***3.2. Effect of rearing practices***

282 The variance analysis showed that the abundance of very few proteins was modified by
283 rearing practices (Table 3). In cow muscles, only 3 proteins were significantly different
284 ($P$<0.05): PRDX6, PGK1, ALDOA, and 3 others showed tendencies ($P$< 0.1): HSP20, ENO3,
285 MDH1. In steer muscles, we observed no significant differences between the two rearing
286 practices for 18 proteins and only 2 tended to be different: ALDOA and ALDH1A1. Only the
287 abundance of ALDOA was affected by rearing practices in both cows and steers. It is
288 worthwhile to note that the abundance of this protein was also different among the 5 muscles in
289 cows and in steers. An effect of gender was observed only in LT muscle with a lower
290 abundance in LT of steers comparatively to cows. The results demonstrated that the effect of
291 rearing practices on the abundance of the 20 biomarkers is weak, and lower than the effect of
292 gender which is weaker than muscle type effect.

293 The analysis of animal and rearing factors on cows allowed to distinguish 3 rearing practices
294 classes that differed by 9 factors (Table 4). The most discriminating factors were animal
295 activity, percentage of grass, haylage or hay in the diet during the fattening period ($P$<0.001)
296 (Table 4). Accordingly, these 3 classes were called "grass", "hay" and "haylage" [8]. For steers,
297 we have identified "grass" and "haylage" rearing practices only (data not shown) and they were
298 not different for any of the studied biomarkers, therefore the results are not discussed in the
299 following sections (Table 3).

300 Comparatively to the "hay" and "haylage" classes, the "grass" class was characterized by
301 higher animal activity, longer fattening period duration and the carcasses of the animals had a
302 lower conformation score (Table 4 and Figure 2a). The haylage class was characterized by a
303 higher carcass weight than the two other classes.

304 For the effect of rearing practices on the studied protein biomarkers, the "grass" class had an
305 impact mainly on the properties of the SM and ST muscles known as fast glycolytic muscles
306 (Figure 2b). This class was characterized by high relative abundance of MLC1F (fast isoform),
307 PRDX6 (an antioxidant enzyme) and of three glycolytic enzymes (PGK1, TPI1 and ENO3).
308 Hay finishing practices affected the properties of RA muscle known as slow oxidative muscle.
309 This class was characterized by high abundance of small Heat Shock Proteins (HSP20, 27 and
310 CRYAB) as well as HSP70-1A, TNNT1 (slow structural protein isoforms) and ALDH1A, and
311 by a low abundance of MyHC-IIX (fast glycolytic). Furthermore, the results revealed that LT
312 and TB muscles, known as mixed oxido-glycolytic muscles, were less impacted by rearing
313 practices than the 3 other muscles. Interestingly, the abundance of 3 proteins FHL1, MDH1 and
314 PYGB was not different among the 3 rearing practices classes whatever the muscle (Figure 3).
315 Abundance of HSP40 and α-tubulin was modified in the Hay class only.

316 One of the main results of the present study is to show that rearing practices classes are
317 different according to the studied muscle. Grass class is composed mainly of SM and ST

318 muscles (fast glycolytic muscles); haylage class groups LT and TB muscles (mixed oxydo-
319 glycolytic muscles) and hay class contains only RA muscle. These data indicate that the impact
320 of rearing practices is muscle type dependent. In this study, the fast glycolytic muscles were the
321 most impacted by grass finishing diet. These modifications are interesting in term of beef
322 tenderness as well as other sensory qualities [44]. Indeed, we have recently showed that ST
323 muscle is more tender when it is more fast glycolytic [45]. A recent study of our group showed
324 that the LT muscle of Rouge de Prés cows with grass diet had lower proportions of IIX fibres
325 (fast glycolytic and higher proportion of IIA fibres fast oxydo-glycolytic) [8]. An opposite
326 effect of rearing practices on LT and ST muscles has already been observed. However, despite
327 an opposite response, the effect of a grass finishing diet has a positive impact on tenderness in
328 both muscles, since for LT, unlike ST, the less glycolytic are the most tender [45].

329 ***3.3. Correlations between biomarkers and the carcass and rearing factors***

330 The correlation analyses, although they are weak but coherent, showed that among the 9
331 factors discriminating the 3 rearing practices classes of cows, fattening duration and age at
332 slaughter had an influence on the protein abundances in the 5 muscles (Figure 4). Fattening
333 duration modified the abundance of 12 among the 20 studied proteins (Figure 4). This effect
334 was the most important in TB muscle as the abundance of 6 proteins was modified. For TB
335 muscle, the abundances of MLC1F, PYGB, PRDX6 and FHL1 decreased when fattening
336 duration increased whereas abundance of HSP70-1A and TTN increased. The abundance of
337 PYGB was also modified in LT and ST muscles (with a negative correlation between fattening
338 duration and PYGB abundance) but not in RA and SM muscles. HSP70-1A was modified also
339 in RA but inversely in comparison with TB muscle. We observed also that the abundance of
340 ENO3 was inversely correlated with fattening duration in LT (positively) and SM (negatively).
341 The present abundance variations seem to be related to the composition of the fibrous part of
342 the diet and/or animal activity that was independent of the slaughter weight and age. These are

343 consistent with previous observations by our group highlighting that fattening duration is the
344 most influencing rearing factors for meat quality, particularly tenderness [2, 8, 11].

345 For slaughter age, the main effect observed was a positive correlation with the abundance of
346 HSP20 in the 5 muscles (Figure 4). It is the only protein which abundance was modified in the
347 same way in the 5 muscles with an increase with age at slaughter of the animals. Interestingly,
348 HSP20 discussed above to be affected by gender was the only protein which abundance was
349 modified in the same way in the 5 muscles. HSP20 belongs to a family of at least 10 different
350 small HSPs [17]. HSP20 is expressed in multiple tissues but it is more abundant in muscle [46].
351 In human and rat, an increase of its expression with age has been reported in accordance with
352 the present results [47, 48]. This increase is considered in the literature as an essential cellular
353 response to fiber aging; according to our results this response seems to be muscle type
354 independent. The modifications of HSP20 abundances with slaughter age are in accordance
355 with the modification of contractile and metabolic properties observed in aged muscles in cows
356 and steers toward a shift from fast glycolytic to slow oxidative [8, 16, 45]. The main effect of
357 slaughter age was observed for RA muscle with a correlation with the abundance of 5 proteins:
358 positively with HSP20, FHL1, ALDH1A1, TNNT1 and negatively with MyHC-IIX. EUROP
359 conformation and carcass weight were linked to the studied proteins in 4 muscles unless TB
360 muscle which was not influenced as any correlation with proteins abundances were observed
361 (Figure 4). The EUROP conformation had an impact mainly in SM muscle in which it was
362 correlated with 4 proteins: positively with TTN, MDH1, TRIM72 and negatively with PGK1.

363 Factors associated with diet composition had weak effects on protein abundances. Grass %
364 was correlated with 4 proteins in LT: positively with TPI1, negatively with HSP70-1A, MDH1,
365 PYGB. Total concentrate (in kg) was correlated with proteins abundances in 4 muscles and no
366 correlations were observed in LT muscle. It was negatively correlated with MDH1 abundance
367 in RA, SM and ST muscles, but not for LT and TB. The abundance of this protein in LT and

368 RA was negatively correlated with animal activity, no correlations were observed for the 3
369 other muscles. It was also negatively correlated with animal slaughter age in LT and SM. We
370 observed that in LT muscle, the abundance of this protein was correlated negatively with 4
371 rearing factors: animal activity, % grass in the diet, carcass weight and fattening duration. On
372 another hand, animal activity showed no correlations with the protein abundances of ST and TB
373 muscles. In each of the three other muscles, animal activity was correlated with the abundances
374 of 3 proteins.

375 Of the 5 muscles, the proteins in TB muscle were the least sensitive to variations in rearing
376 practices. No correlations were observed with any proteins irrespective of rearing practices with
377 EUROP conformation and carcass weight. Only one protein was correlated with the activity of
378 the animals at the farm, mainly MLC-1F as well as with total concentrate for FHL1. However,
379 TB muscle was the most modified muscle by fattening duration. On the contrary, RA and SM
380 muscles were the most sensible to rearing practices as correlations with all rearing factors
381 except grass% for RA and haylage % for SM, were observed.

382 ***3.4. Proteins that did not discriminate the rearing practices classes with no difference among***
383 ***muscles and genders***

384 The abundances of FHL1 (Four and a half LIM domains protein 1) and PYGB (Glycogen
385 phosphorylase B) were not different between the three rearing practices classes. Interestingly,
386 the abundances of these proteins were not significantly different among the two genders and
387 among the5 muscles in cows and in steers. This indicates that the abundances of these proteins
388 are muscle, gender and rearing practices independent.

389 FHL1 also named SLIM1 or KyoT1, belongs to the FHL protein family composed of four
390 and a half Lin-11, Isl-1, and Mec-3 (LIM) domains. FHL LIM domains mediate protein –
391 protein interactions, scaffolding signaling proteins in the cytoplasm, and transcription factors in
392 the nucleus. FHL1 as mentioned above is considered as a regulator of skeletal muscle mass, and

393 strength enhancement by binding with the calcineurin-regulated transcription factor NFATc1
394 [49]. This protein is confined to the *Z*-line of skeletal muscle and its proteolysis is linked to the
395 release of intact α-actinin from bovine myofibrils and contributes to the weakening of the Z-line
396 during meat tenderizing [50]. FHL1 may also interact with other biological pathways, namely
397 metabolic enzymes [26, 51] in response to both hypoxia, apoptosis and oxidative stress [52].
398 This protein seems to play a fundamental role in muscle mass and muscular strength which
399 could explain why its expression is relatively stable according to muscle, gender or rearing
400 practices. For example, FHL1 increased the myostatin activity on a SMAD reporter and
401 increased myostatin dependent myotube wasting [53]. According to these authors, FHL1 is
402 expressed at higher levels in type II than in type I fibers raising the possibility that it contributes
403 to the greater sensitivity of type II fibers to myostatin. However, these differences in fiber types
404 expression were not observed among our 5 muscles as previously reported by our group [26].
405 On another hand, PYGB is a Glycogen Phosphorylase which catalyzes the glycogen
406 degradation. Its activity is positively regulated by AMP and negatively regulated by ATP, ADP,
407 and glucose-6-phosphate [6].The non-variation on this protein abundance would be due to a
408 lack of an enhanced glycogen degradation by the factors considered in this publication.

409 **4. Conclusion**

410 This study is the first to consider the effect of gender and rearing practices on the
411 abundances of biomarkers of tenderness and IMF content in five different muscles in cattle. The
412 main results showed a higher effect of muscle type than gender or rearing practices. Moreover,
413 factors associated with diet composition had few effects on proteins abundances. This
414 knowledge constitutes important information to understand how to manage the expression of
415 biomarkers of tenderness and IMF content according to gender and rearing practices.

416

417

## Author contributions

BP and MB defined the experiment design, managed the experiment, co-wrote the paper, and approved the final draft of the manuscript. MG managed the database, analyzed the data, prepared figures and/or tables, co-wrote the paper and approved the final draft of the manuscript. MEJ participated in the database preparation. All authors collaborated with interpretation and discussion of the results. All authors have given approval to the final versions of the manuscript.

## Conflict of interest

The authors declare no competing financial interest


## Funding Sources

This experiment was conducted with funding from the regional council of Pays de Loire and SICA Rouge des Prés (France).

## Acknowledgements

The authors thank Albéric Valais, Ghislain Aminot and Marlène Pécot from SICA Rouge des Prés for muscle sampling and data on animal, rearing factors and carcass properties. They thank Leanne De Koning, Aurélie Cartier and Bérengere Ouine from Institut Curie, RPPA Plateform, Paris France, for the quantification of the biomarkers.


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## Tables and Figures

### Figure captions

**Figure 1.** Interaction between muscle x gender for HSP20 protein.

**Figure 2.** Principal component analysis (PCA) depicting the relationships between the rearing practices of the 86 PDO Maine-Anjou cows identified following the procedure by Gagaoua *et al.* [8] with **A)** animal, rearing factors and carcass characteristics, and with **B)** the 20 protein biomarkers from the 5 muscles quantified by RPPA technique within the rearing factors. The projection of the individuals of haylage class (red), hay class (bleu) and grass class (green) are encircled in ellipses (x,y-means ± x,y-standard deviation (SD)) using the corresponding schematic colors. Furthermore, the barycenter of each muscle with the corresponding color are given.

**Figure 3.** Unsupervised hierarchical classification heatmap highlighting the differences in the quantified proteins in the five muscles and among the three rearing practices for cows. The proteins that were not affected by rearing practices or muscle type are shown by "*". Colors correspond to the z-scores of the standardized values of protein fold-change between the muscles according to the 3 rearing factors.

**Figure 4.** Significant correlations ($P < 0.05$) between the 20 protein biomarkers and animal, rearing factors and carcass characteristics by muscle type. The negative correlations are given in red and the positive in green. The summary of the number of the correlations by muscle with the animal, rearing factors and carcass characteristics are given in a gradient-blue dependent color legend at the right down of the graph from 1 to 6 correlations in each muscle and with the same factor. For example, for TB muscle 6 significant correlations (intense bleu color) were found with fattening duration compared to animal activity where only one correlation was found (light bleu color).

**Table 1.** List of the 20 protein biomarkers quantified using the Reverse Phase Protein Array (RPPA) technique. The suppliers and conditions for each primary antibody used in this study after western blotting validation are given as in Picard *et al.* [26] and Gagaoua *et al.* [11, 27].

| Protein biomarkers name (*gene*) | Uniprot ID | Monoclonal (Mo) or Polyclonal (Po) antibodies references | Antibody dilutions |
|---|---|---|---|
| ***Metabolic enzymes*** | | | |
| Malate dehydrogenase (*MDH1*) | **P40925** | Mo. anti-pig Rockland 100-601-145 | 1/1000 |
| β-enolase 3 (*ENO3*) | **P13929** | Mo. anti-human Abnova Eno3 (M01), clone 5D1 | 1/30 000 |
| Retinal dehydrogenase 1 (*ALDH1A1*) | **P48644** | Po. anti-bovine Abcam ab23375 | 1/500 |
| Triosephosphate isomerase (*TPI1*) | **Q5E956** | Po. anti-human Novus NBP1-31470 | 1/50 000 |
| Phosphoglycerate kinase 1 (*PGK1*) | **Q3T0P6** | Po. anti-human Abcam ab90787 | 1/5000 |
| Fructose-bisphosphate aldolase (*ALDOA*) | **A6QLL8** | Po. anti-human Sigma AV48130 | 1/4000 |
| Glycogen phosphorylase (*PYGB*) | **Q3B7M9** | Po. anti-human Santa Cruz SC-46347 | 1/250 |
| ***Heat shock proteins*** | | | |
| αB-crystallin (*CRYAB*) | **P02511** | Mo. anti-bovine Assay Designs SPA-222 | 1/1000 |
| Hsp20 (*HSPB6*) | **O14558** | Mo. anti-human Santa Cruz HSP20-11:SC51955 | 1/500 |
| Hsp27 (*HSPB1*) | **P04792** | Mo. anti-human Santa Cruz HSP27 (F-4):SC13132 | 1/3000 |
| Hsp40 (*DNAJA1*) | **P31689** | Mo. anti-human Santa Cruz HSP40-4 (SPM251):SC-56400 | 1/250 |
| Hsp70-1A (*HSPA1A*) | **Q27975** | Mo. anti-human RD Systems MAB1663 | 1/1000 |
| ***Oxidative proteins*** | | | |
| Peroxiredoxin6 (*PRDX6*) | **P30041** | Mo. anti-human Abnova PRDX6 (M01), clone 3A10-2A11 | 1/500 |
| ***Structural proteins*** | | | |
| MLC-1F (*MYL1*) | **P05976** | Po. anti-human Abnova MYL1 (A01) | 1/1000 |
| Myosin heavy chain-IIx (*MYH1*) | **P12882** | Mo anti-bovine Biocytex 8F4 | 1/500 |
| Troponin T, slow skeletal muscle (*TNNT1*) | **Q8MKH6** | Po. anti-human Sigma SAB2102501 | 1/4000 |
| Titin (*TTN*) | **Q8WZ42** | Mo. anti-human Novocastra NCL-TITIN | 1/100 |
| Tubulin alpha-4A chain (*TUBA4A*) | **P81948** | Mo anti-human Sigma T6074 | 1/1000 |
| ***Cell death, protein binding and proteolysis*** | | | |
| Tripartite motif protein 72 (*Trim72*) | **E1BE77** | Po. anti-human Sigma SAB2102571 | 1/2000 |
| Four and a half LIM domains 1 (*FHL1*) | **Q3T173** | Po. anti-human Sigma AV34378 | 1/5000 |

**Table 2.** Muscle, gender and muscle x gender interaction effects on the 20 beef tenderness and intramuscular fat proteomic biomarkers.

| Proteins[1] | G | Muscle (M)[2] | | | | | Gender (G) | | P-values[3] | | |
|---|---|---|---|---|---|---|---|---|---|---|---|
| | | TB | ST | RA | SM | LT | Cows (C) | Steers (S) | M | G | M*G |
| *CRYAB* | C | -0.15$^{bc}$ | -0.62$^{d}$ | 1.03$^{a}$ | -0.21$^{c}$ | -0.02$^{b}$ | 0.03 | -0.09 | *** | ns | ns |
| | S | -0.18$^{b}$ | -0.67$^{c}$ | 0.57$^{a}$ | -0.35$^{b}$ | -0.06$^{b}$ | | | *** | | |
| *Sign.*[4] | | ns | ns | ** | ns | ns | | | | | |
| *HSP20* | C | -0.23$^{c}$ | -0.25$^{c}$ | 0.29$^{a}$ | 0.01$^{b}$ | 0.17$^{a}$ | 0.05$^{a}$ | -0.33$^{b}$ | *** | *** | ** |
| | S | -0.34$^{b}$ | -0.78$^{b}$ | 0.16$^{a}$ | -0.46$^{b}$ | -0.42$^{b}$ | | | ** | | |
| *Sign.* | | ns | *** | ns | *** | *** | | | | | |
| *HSP27* | C | -0.06$^{b}$ | -0.08$^{b}$ | 0.61$^{a}$ | -0.44$^{c}$ | -0.04$^{b}$ | 0.03 | -0.14 | *** | ns | ns |
| | S | -0.19$^{b}$ | -0.27$^{b}$ | 0.27$^{a}$ | -0.65$^{c}$ | -0.05$^{b}$ | | | *** | | |
| *Sign.* | | ns | * | * | * | ns | | | | | |
| *HSP70-1A* | C | -0.20$^{c}$ | -0.36$^{c}$ | 0.28$^{a}$ | 0.17$^{ab}$ | 0.08$^{b}$ | -0.02 | 0.05 | *** | ns | ns |
| | S | -0.02 | -0.17 | 0.17 | 0.01 | 0.15 | | | ns | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *HSP40* | C | -0.11 | 0.02 | -0.05 | 0.06 | -0.11 | -0.05 | 0.03 | ns | ns | ns |
| | S | -0.03 | 0.30 | 0.09 | 0.12 | -0.01 | | | ns | | |
| *Sign.* | | ns | * | ns | ns | ns | | | | | |
| *FHL1* | C | 0.12 | -0.16 | 0.04 | -0.03 | 0.01 | -0.01 | 0.07 | ns | ns | ns |
| | S | 0.13 | 0.03 | 0.14 | -0.05 | 0.13 | | | ns | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *TRIM72* | C | 0.41$^{a}$ | -0.08$^{b}$ | 0.01$^{b}$ | -0.11$^{b}$ | 0.32$^{a}$ | 0.13 | 0.04 | *** | ns | ns |
| | S | 0.34$^{a}$ | -0.20$^{c}$ | -0.04$^{bc}$ | -0.07$^{bc}$ | 0.11$^{b}$ | | | *** | | |
| *Sign.* | | ns | * | ns | ns | ** | | | | | |
| *PRDX6* | C | 0.16$^{a}$ | 0.12$^{ab}$ | -0.03$^{b}$ | 0.26$^{a}$ | -0.33$^{c}$ | 0.00$^{b}$ | 0.23$^{a}$ | *** | ** | ns |
| | S | 0.37$^{a}$ | 0.35$^{a}$ | 0.33$^{a}$ | 0.38$^{a}$ | -0.07$^{b}$ | | | * | | |
| *Sign.* | | ns | ns | ** | ns | * | | | | | |
| *MDH1* | C | 0.09 | 0.04 | 0.01 | -0.11 | 0.07 | 0.04$^{a}$ | -0.09$^{b}$ | ns | * | ns |
| | S | -0.01 | -0.08 | -0.15 | -0.14 | -0.09 | | | ns | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *PYGB* | C | 0.08 | 0.11 | -0.02 | 0.05 | 0.01 | 0.04 | 0.07 | ns | ns | ns |
| | S | 0.06 | 0.08 | 0.07 | 0.18 | -0.08 | | | ns | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *PGK1* | C | 0.11$^{b}$ | 0.39$^{a}$ | -0.95$^{c}$ | 0.35$^{a}$ | 0.11$^{b}$ | -0.06$^{b}$ | 0.30$^{a}$ | *** | *** | ns |
| | S | 0.33$^{b}$ | 0.83$^{a}$ | -0.52$^{c}$ | 0.59$^{ab}$ | 0.39$^{b}$ | | | *** | | |
| *Sign.* | | * | *** | * | * | * | | | | | |
| *ALDOA* | C | -0.04$^{b}$ | 0.26$^{a}$ | -0.24$^{c}$ | 0.16$^{a}$ | -0.02$^{b}$ | 0.04$^{a}$ | -0.11$^{b}$ | *** | ** | ns |
| | S | -0.03$^{ab}$ | 0.25$^{a}$ | -0.25$^{b}$ | 0.02$^{ab}$ | -0.27$^{b}$ | | | ** | | |
| *Sign.* | | ns | ns | ns | ns | * | | | | | |
| *ALDH1A1* | C | -0.16$^{bc}$ | -0.07$^{b}$ | 0.73$^{a}$ | -0.28$^{c}$ | -0.15$^{bc}$ | 0.00 | 0.14 | *** | ns | ns |
| | S | -0.06$^{b}$ | 0.11$^{b}$ | 0.67$^{a}$ | -0.03$^{b}$ | 0.05$^{b}$ | | | *** | | |
| *Sign.* | | ns | ns | ns | * | ns | | | | | |
| *ENO3* | C | 0.22$^{bc}$ | 0.58$^{a}$ | -1.22$^{d}$ | 0.33$^{b}$ | 0.10$^{c}$ | -0.03 | 0.14 | *** | ns | ns |
| | S | 0.17$^{b}$ | 0.76$^{a}$ | -0.70$^{c}$ | 0.31$^{b}$ | 0.27$^{b}$ | | | *** | | |
| *Sign.* | | ns | * | ** | ns | ns | | | | | |
| *TPI1* | C | 0.04$^{c}$ | 0.55$^{a}$ | -1.02$^{d}$ | 0.31$^{b}$ | -0.03$^{c}$ | -0.08$^{b}$ | 0.18$^{a}$ | *** | * | ns |
| | S | 0.11$^{b}$ | 0.86$^{a}$ | -0.56$^{c}$ | 0.45$^{b}$ | 0.24$^{b}$ | | | *** | | |
| *Sign.* | | ns | ** | * | ns | ns | | | | | |

[1] Least-square means in the same row with different superscript letters are significantly different ($P$<0.05).
[2] Muscle abbreviation:
TB: *Triceps brachii*; ST: *Semitendinosus* ; RA: *Rectus abdominis* ; SM: *Semimembranosus* ; LT: *Longissimus thoracis*
[3] Significances: ns: not significant; * $P$<0.05; ** $P$<0.01; *** $P$<0.001
[4] Gender effect significance on the proteins by muscle.

**Table 2.** *Continued*

| Proteins[1] | G | Muscle (M)[2] | | | | | Gender (G) | | P-values[3] | | |
|---|---|---|---|---|---|---|---|---|---|---|---|
| | | TB | ST | RA | SM | LT | Cows (C) | Steers (S) | M | G | M*G |
| *TTN* | C | 0.30[a] | -0.33[c] | -0.05[b] | -0.31[c] | 0.34[a] | -0.01 | 0.04 | *** | ns | ns |
| | S | 0.34[a] | -0.32[b] | -0.20[b] | -0.05[b] | 0.22[a] | | | ** | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *MHC-IIX* | C | 0.27[b] | 0.75[a] | -0.91[d] | 0.06[b] | -0.21[c] | 0.03[a] | -0.24[b] | *** | * | ns |
| | S | 0.08[a] | 0.30[a] | -0.83[b] | -0.07[a] | -0.54[b] | | | *** | | |
| *Sign.* | | ns | ** | ns | ns | ns | | | | | |
| *MLC1F* | C | 0.26[ab] | 0.39[a] | -0.56[c] | 0.08[b] | 0.09[b] | 0.06 | -0.02 | *** | ns | ns |
| | S | 0.20[a] | 0.24[a] | -0.54[b] | 0.09[a] | 0.09[a] | | | *** | | |
| *Sign.* | | ns | ns | ns | ns | ns | | | | | |
| *TNNT1* | C | 0.09[b] | -0.97[d] | 0.88[a] | -0.13[c] | 0.08[b] | -0.02[b] | 0.19[a] | *** | * | ns |
| | S | 0.27[b] | -0.76[c] | 0.87[a] | 0.05[b] | 0.28[b] | | | *** | | |
| *Sign.* | | ns | ns | ns | ns | * | | | | | |
| *α-Tubulin* | C | 0.05[a] | -0.03[ab] | 0.10[a] | -0.02[ab] | -0.13[b] | 0.01 | -0.08 | ** | ns | ns |
| | S | 0.05 | 0.03 | -0.01 | -0.16 | -0.27 | | | ns | | |
| *Sign.* | | ns | ns | ns | * | ns | | | | | |

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[2] Muscle abbreviation:
TB: *Triceps brachii*; ST: *Semitendinosus* ; RA: *Rectus abdominis* ; SM: *Semimembranosus* ; LT: *Longissimus thoracis*
[3] Significances: ns: not significant; * $P<0.05$; ** $P<0.01$; *** $P<0.001$
[4] Gender effect significance on the proteins by muscle.

**Table 3.** Variance analyses of the rearing practices and muscle x rearing practices interaction effects on the 20 beef tenderness and intramuscular fat proteomic biomarkers for cows and steers.

| Proteins[1] | Gender | Effects[3] | |
|---|---|---|---|
| | | Rearing practices | Rearing practices x muscle |
| *CRYAB* | Cows (C) | ns | ns |
| | Steers (S)[2] | ns | ns |
| *HSP20* | C | 0.073 | ns |
| | S | ns | ns |
| *HSP27* | C | ns | ns |
| | S | ns | ns |
| *HSP70-1A* | C | ns | ns |
| | S | ns | 0.093 |
| *HSP40* | C | ns | ns |
| | S | ns | ns |
| *FHL1* | C | ns | ns |
| | S | ns | ns |
| *TRIM72* | C | ns | ns |
| | S | ns | ns |
| *PRDX6* | C | 0.019 | ns |
| | S | ns | ns |
| *MDH1* | C | 0.088 | ns |
| | S | ns | ns |
| *PYGB* | C | ns | 0.087 |
| | S | ns | ns |
| *PGK1* | C | 0.038 | ns |
| | S | ns | ns |
| *ALDOA* | C | 0.035 | ns |
| | S | 0.098 | ns |
| *ALDH1A1* | C | ns | ns |
| | S | 0.056 | ns |
| *ENO3* | C | 0.056 | ns |
| | S | ns | ns |
| *TPI1* | C | ns | ns |
| | S | ns | ns |
| *TTN* | C | ns | ns |
| | S | ns | ns |
| *MHC-IIX* | C | ns | ns |
| | S | ns | ns |
| *MLC1F* | C | ns | ns |
| | S | ns | ns |
| *TNNT1* | C | ns | ns |
| | S | ns | ns |
| *α-Tubulin* | C | ns | ns |
| | S | ns | ns |

[1] Least-square means in the same row with different superscript letters are significantly different ($P<0.05$).
[2] Only two rearing factors were identified for steers (Grass class (n = 5) and Haylage class (n= 10)).
[3] ns: not significant ($P > 0.1$).

**Table 4.** Differences in animal, rearing factors and carcass characteristics among the three identified rearing practices.

| Variables | Grass class (n = 24) | Haylage class (n = 21) | Hay class (n = 41) | P-values[1] |
|---|---|---|---|---|
| Animal activity, % | 78.79[a] | 2.81[b] | 5.29[b] | *** |
| Grass, % | 19.10[a] | 0.80[b] | 0.53[b] | *** |
| Haylage, % | 59.71[b] | 81.99[a] | 4.70[c] | *** |
| Hay, % | 21.15[b] | 17.21[b] | 94.77[a] | *** |
| Total concentrate, kg | 857 | 741 | 788 | ns |
| Fattening duration, days | 120.3[a] | 100.3[b] | 99.5[b] | * |
| Age, months | 64.50[b] | 65.19[b] | 71.22[a] | t |
| Carcass weight, kg | 461.33[a] | 434.10[b] | 462.24[a] | * |
| Conformation score | 3.54[b] | 4.10[a] | 3.85[ab] | t |

[1] Significances: ns: not significant; t $P < 0.1$; * $P<0.05$; *** $P<0.001$

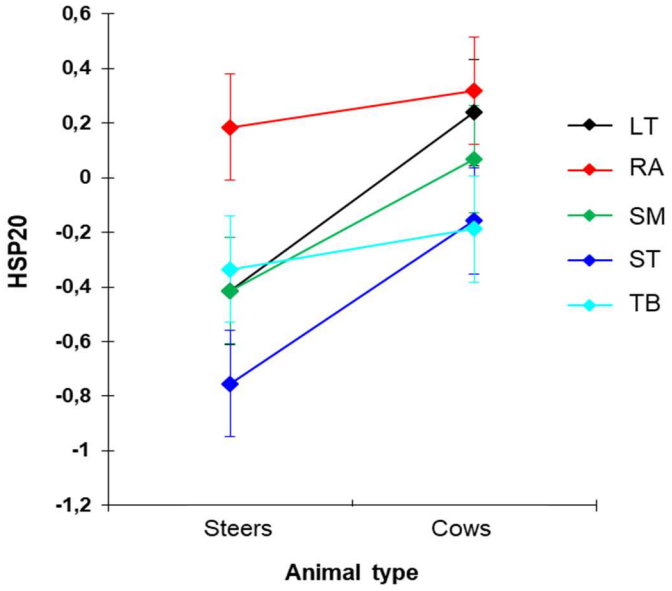

0,6
0,4
0,2
0
-0,2
-0,4
-0,6
-0,8
-1
-1,2
HSP20
Steers
Cows
Animal type
LT
RA
SM
ST
TB

**Figure 1.**

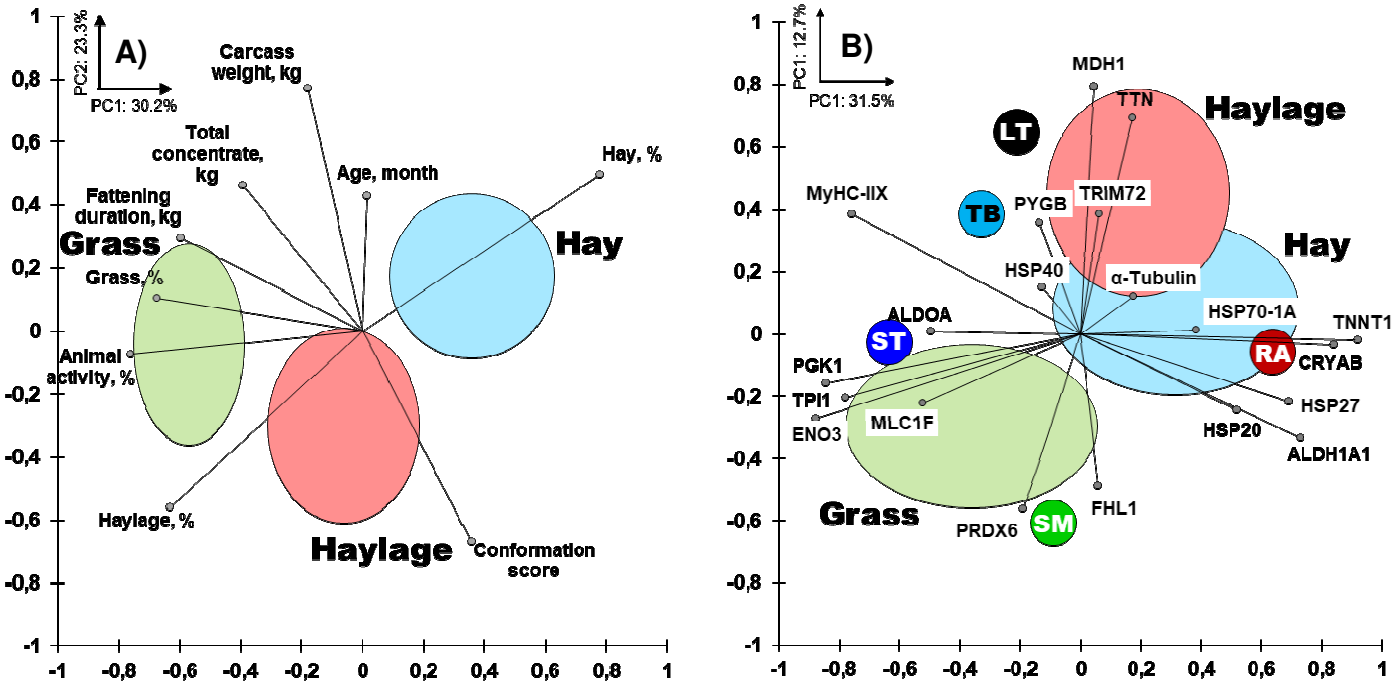

A)
PC2: 23.3%
PC1: 30.2%
Carcass weight, kg
Total concentrate, kg
Age, month
Hay, %
Fattening duration, kg
Grass
Hay
Grass, %
Animal activity, %
Haylage, %
Haylage
Conformation score
B)
PC1: 12.7%
PC1: 31.5%
MDH1
TTN
Haylage
LT
MyHC-IIX
TB
PYGB
TRIM72
Hay
HSP40
α-Tubulin
ALDOA
HSP70-1A
TNNT1
ST
RA
CRYAB
PGK1
TPI1
HSP27
ENO3
MLC1F
HSP20
ALDH1A1
Grass
FHL1
PRDX6
SM

**Figure 2.**

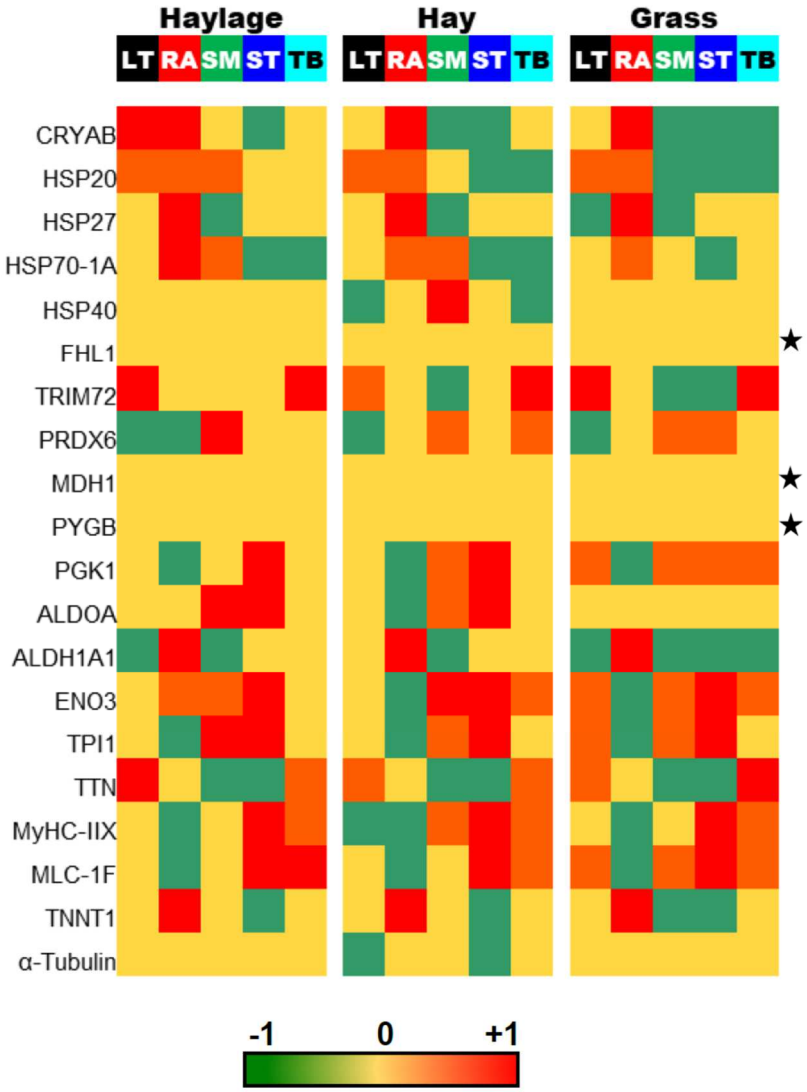

Haylage
Hay
Grass
LT RA SM ST TB
LT RA SM ST TB
LT RA SM ST TB
CRYAB
HSP20
HSP27
HSP70-1A
HSP40
FHL1
TRIM72
PRDX6
MDH1
PYGB
PGK1
ALDOA
ALDH1A1
ENO3
TPI1
TTN
MyHC-IIX
MLC-1F
TNNT1
α-Tubulin
-1
0
+1

**Figure 3.**

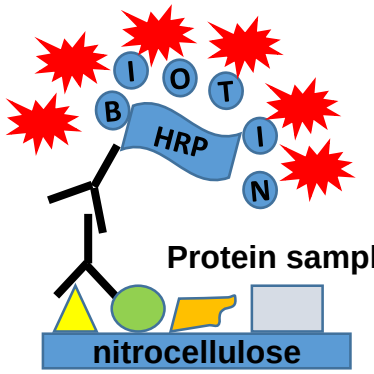
Animal activity, %
Grass, %
Haylage, %
Hay, %
Total concentrate, kg
EUROP conformation
Age at slaugher, months
Carcass weight, kg
Fattening duration, days
LT RA SM ST TB
CRYAB
HSP20
HSP27
HSP70-1A
HSP40
FHL1
TRIM72
PRDX6
MDH1
PYGB
PGK1
ALDOA
ALDH1A1
ENO3
TPI1
TTN
MyHC-IIX
MLC-1F
TNNT1
α-Tubulin
Animal activity, %
Grass, %
Haylage, %
Hay, %
Total concentrate, kg
EUROP conformation
Age at slaugher, months
Carcass weight, kg
Fattening duration, days
#Color legend
1
2
3
4
5
6

**Figure 4.**

Effect of

- Gender: cows *vs* steers
- Muscle type
- Rearing factors

Quantification of beef tenderness / intramuscular fat content protein biomarkers




*Reverse Phase Protein Array*

Management of beef meat qualities by rearing practices