



**HAL**  
open science

## Sequence Variation and Recognition Specificity of the Avirulence Gene *AvrPiz-T* in *Magnaporthe Oryzae* Field Populations

Chenxi Chen, Meilian Chen, Jinnan Hu, Wenjing Zhang, Zhenhui Zhong, Yulin Jia, Ludovic Allaux, Elisabeth E. Fournier, Didier Tharreau, Guo-Liang Wang, et al.

► **To cite this version:**

Chenxi Chen, Meilian Chen, Jinnan Hu, Wenjing Zhang, Zhenhui Zhong, et al.. Sequence Variation and Recognition Specificity of the Avirulence Gene *AvrPiz-T* in *Magnaporthe Oryzae* Field Populations. *Fungal Genomics & Biology*, 2014, 4 (1), 10.4172/2165-8056.1000113 . hal-02631317

**HAL Id: hal-02631317**

**<https://hal.inrae.fr/hal-02631317>**

Submitted on 27 May 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Sequence Variation and Recognition Specificity of the Avirulence Gene *AvrPiz-T* in *Magnaporthe Oryzae* Field Populations

Chenxi Chen<sup>1,2#</sup>, Meilian Chen<sup>1#</sup>, Jinnan Hu<sup>2</sup>, Wenjing Zhang<sup>1</sup>, Zhenhui Zhong<sup>1</sup>, Yulin Jia<sup>3</sup>, Ludovic Allaux<sup>4</sup>, Elisabeth Fournier<sup>5</sup>, Didier Tharreau<sup>4</sup>, Guo-Liang Wang<sup>2,6</sup>, Zonghua Wang<sup>1</sup>, Wei-Chiang Shen<sup>7</sup>, Guodong Lu<sup>1</sup>, Baohua Wang<sup>1\*</sup>, Thomas K. Mitchell<sup>2\*</sup>

<sup>1</sup>Key Laboratory of Bio-pesticide and Chemistry Biology, Ministry of Education, Fujian Agriculture and Forestry University, Fuzhou, 350002, China

<sup>2</sup>Department of Plant Pathology, The Ohio State University, Columbus, OH, 43210, USA

<sup>3</sup>USDA-ARS Dale Bumpers National Rice Research Center, Stuttgart, AR, 72160, USA

<sup>4</sup>CIRAD, UMR 385 Biologie et Génétique des Interactions Plante-Parasite, F-34398 Montpellier, France

<sup>5</sup>INRA, UMR 385 Biologie et Génétique des Interactions Plante-Parasite, F-34398 Montpellier, France

<sup>6</sup>Hunan Provincial Key Laboratory of Crop Germplasm Innovation and Utilization, College of Agronomy, Hunan Agricultural University, Hunan 410128, China

<sup>7</sup>Department of Plant Pathology and Microbiology, National Taiwan University, Taipei, Taiwan

#These authors contributed equally to this work

## Summary

*Magnaporthe oryzae*, the rice blast pathogen, causes significant annual yield loss of rice worldwide. Currently, the most effective disease control approach is deployment of host resistance through introduction of resistance (*R*) genes into elite cultivars. The function of each *R* gene relies on the specific recognition of an avirulence (*AVR*) gene of the pathogen. However, the introduced resistance can be broken down within a few years, due to mutation of the *AVR* genes in the field populations. It is known from a few case studies that *AVR* mutation patterns are different from one to another. Therefore, knowledge of *AVR* genes sequence diversity serves as fundamental background in introducing new resistance to control rice blast. In this study, we focused on a newly identified *AVR* gene *AvrPiz-t*. We PCR amplified open reading frames (ORFs) of *AvrPiz-t* as well as promoter regions to detect size variation at this locus in 711 isolates of *M. oryzae* collected from 38 countries and regions. Through sequencing and Southern hybridization of the amplified locus, strains with polymorphisms in the ORF were classified into groups based on mutation type and site. Natural selection intensity on this gene was calculated and pathogenicity assays were applied to evaluate the association between *AvrPiz-t* ORF/promoter polymorphism and virulence. In conclusion, sequences at the *AvrPiz-t* locus were revealed to contain variations at both promoter and ORF regions. This locus is undergoing a relatively strong positive selection. The diversity in coding sequence and the insertions of transposable elements in the promoter region enable *M. oryzae* to evade recognition by the cognate *Piz-t R* gene in the host.

**Keywords:** *Magnaporthe oryzae*; *AvrPiz-t*; Transposon insertion; SNP; Gene diversity

## Introduction

Most plants are not affected by the majority of microbes they encounter because of their functional defense system. During the co-evolution of plants and phytopathogens, plants have evolved multiple branches of defenses including one specific reactions triggered by pathogen delivered effectors [1,2]. According to the “gene-for-gene” model proposed by Flor in 1971, plants developed a mechanism to detect avirulence (*Avr*) gene products via corresponding resistance genes (*R* genes), which results in a subsequent hypersensitive reaction (HR) to suppress microbial growth [3].

*Magnaporthe oryzae* is an ascomycete fungus and the causal agent of rice blast disease, which is considered as one of the most devastating diseases of rice with regard to grain loss. This disease has been found in over 85 countries and the amount of rice lost each year could feed upward of 60 million people [4], as severe disease breakouts may lead to as much as 90% yield loss in a field or region [5].

In order to provide a genetic based disease control and understand molecular interactions between plants and fungal pathogens, great effort has been made to identify and clone *R* genes of rice cultivars and *AVR* genes in *M. oryzae*. To date, a large number of *R* genes have been successfully transferred into elite rice cultivars to create resistance rice lines through breeding programs [6]. However, *R* gene mediated resistance can be rapidly broken down after several generations [5], which is hypothesized to be result of the inherent instability and high variation of *M. oryzae* *AVR* genes especially those in subtelomeric regions [7]. For example, while *Piz-t* serves as a proven *R* gene in rice

blast disease control, its effectiveness is challenged by the fast evolution of the *AvrPiz-t* locus, which was cloned in 2009 [8]. In one survey we performed in major rice growing areas in China, although the *Piz-t* gene remains functional in majority areas (Figure 1), the virulence frequency of *AvrPiz-t* containing strains from two disease nurseries in Fujian province has increased from 12.5% to 100% within 14 years (Figure 2). So far, several *AVR* genes have been cloned in *M. oryzae*, including *AVR-Pita* [9], *AVR-CO39* [10], *PWL1* [11], *PWL2* [12], *ACE1* [13], *AVR-Pia*, *AVR-Pii*, and *AVR-Pik/km/kp* [14]. Interestingly, it was found that some *AVR* genes of *M. oryzae* contain higher level of variations compared to other regions of genome. As exemplified by the *AVR-Pita1* gene, its ORF showed high polymorphisms among 30 Thai rice blast isolates resulting in varying pathotypes. It was also found that

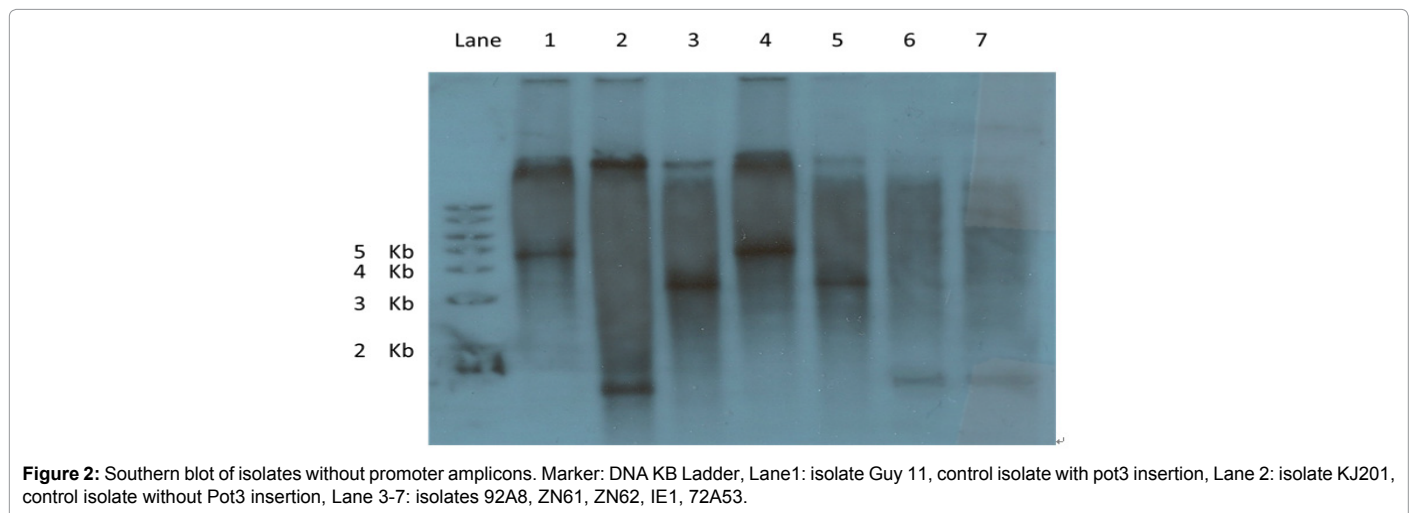
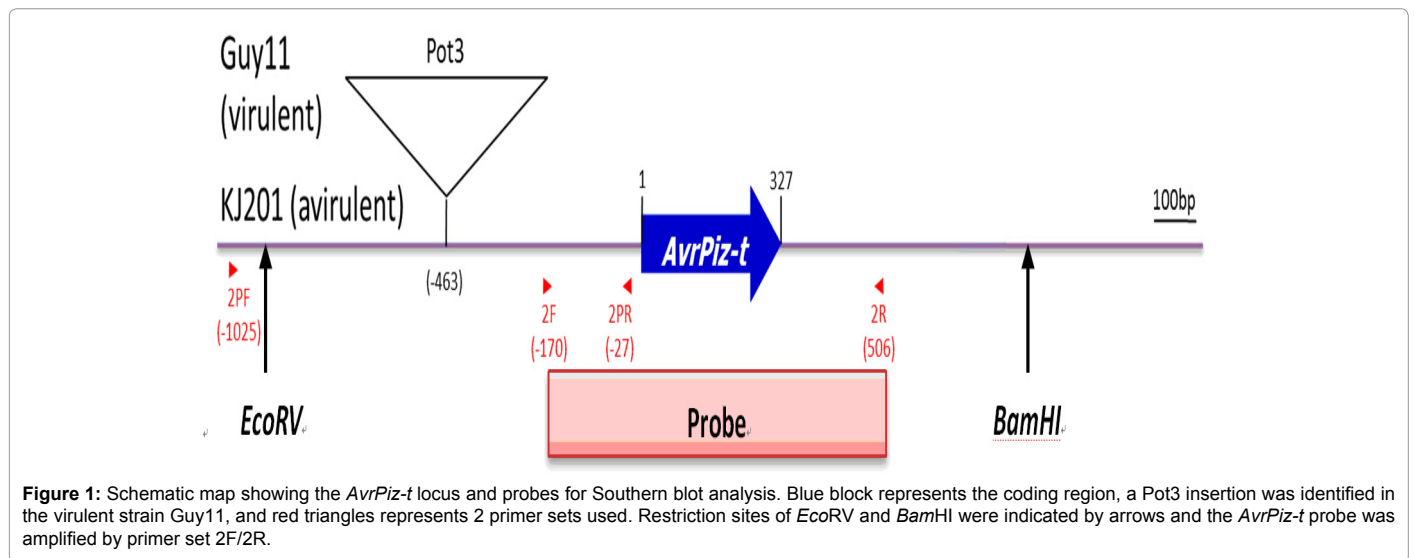
**\*Corresponding authors:** Baohua Wang, Key Laboratory of Bio-pesticide and Chemistry Biology, Ministry of Education, Fujian Agriculture and Forestry University, Fuzhou, 350002, China, Tel: 86-591-86393212; E-mail: [wbaohua@fafu.edu.cn](mailto:wbaohua@fafu.edu.cn)

Thomas Mitchell Department of Plant Pathology, The Ohio State University, Columbus, OH, 43210, USA, Tel: (614) 917-9053; E-mail: [Mitchell.814@osu.edu](mailto:Mitchell.814@osu.edu)

Received May 24, 2014; Accepted July 25, 2014; Published July 28, 2014

**Citation:** Chen C, Chen M, Hu J, Zhang W, Zhong Z, et al. (2014) Sequence Variation and Recognition Specificity of the Avirulence Gene *AvrPiz-T* in *Magnaporthe Oryzae* Field Populations. Fungal Genom Biol 4: 113. doi:10.4172/2165-8056.1000113

**Copyright:** © 2014 Chen C, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.



the *AVR-Pita1* gene locus in the Thai blast isolates were under a positive selection pressure [15]. Similarly, the *AVR-Pita1* alleles in field isolates of the USA were found to be under positive selection. Consequently, mutations at the coding regions resulting amino acid alteration, deletion resulting in frame shift and transposon insertion at a critical motif of the gene have been found to be responsible for defeating the corresponding *Pi-ta R* gene [16-18]. These findings suggest that *M. oryzae* has developed sophisticated mechanisms to survive and retain virulence and that these mechanisms may be different between genes.

*AvrPiz-t* was cloned in 2009 using a map-based strategy and encodes a 108-amino-acid predicted secreted protein [19]. The structure of the *AvrPiz-t* protein has recently identified based on Nuclear Magnetic Resonance (NMR) measurements, which forms a six-strand  $\beta$ -sandwich structure, while both N- and C-termini are disordered [20]. The mechanism of its function has also been explored in a recent study: during the infection process, *AvrPiz-t* accumulates in a specific structure called biotrophic interfacial complex (BIC) and is then transported into rice cells [21]. In the susceptible rice host, *AvrPiz-t* suppresses the reactive oxygen species (ROS) generation and thus strengthens pathogenicity in cultivars without *Piz-t R* gene [22].

It was proposed by Jones and Dangl [2] that *AVR* genes are usually

flanked by transposable elements (TEs) or telomeres, which may serve as a simple strategy for pathogen to evade host detection. In *M. oryzae*, it has been reported that *AVR* genes are sometimes tightly associated with diverse TEs [13,23,24] in either the promoter or coding regions that can alter strain virulence, as been reported in *Avr-Pita* [16,25]. For instance, functional *Avr-Pita* homologs were found located in different chromosomes in a *M. oryzae* population survey, and were accompanied by a variety of TEs such as Inago1, Inago2, Pyret, Pot2, and Pot3. Based on the strong association between the *Avr-Pita* translocation events and the flanking TEs, the authors suggested that the inserted TEs may be essential for the mobility of *Avr-Pita* [26]. In *M. oryzae* strain GUY11, a virulence strain to *Piz-t* containing cultivar, *AvrPiz-t* was found present but having a transposable element Pot3 inserted in its promoter region [8]. In the 146 kb *AvrPiz-t* locus of the strain 70-15 which is derived from GUY11, TEs sequences were also found for as much as 43.2% of the genomic content at this locus, suggesting the dynamic evolution in this region [27].

To evaluate the diversity of the *AvrPiz-t* gene, and determine its effect to virulence, we characterized 711 *M. oryzae* field strains in total: 313 isolates from different geographic origins (38 countries), which are referred to as the “Global strains” group, and an additional 398

isolates from different regions in China, which are referred to as the “Chinese strains” group. Most of the strains were isolated from rice but that some “non rice” strains were also included. In this study, we PCR amplified *AvrPiz-t* ORFs as well as promoter regions of each strain, and then compared the sequence and structure variation using Sanger sequencing and Southern hybridization. Strains with polymorphisms in the ORF were classified into groups based on mutation type and site. Pathogenicity assays were applied to evaluate the association between *AvrPiz-t* gene/promoter polymorphism and virulence.

## Materials and Methods

### Fungal isolates

Fungal isolates used in this study were collected through collaborating labs from over 38 countries and regions worldwide: China, Egypt, India, The Philippines, Burundi, Brazil, Ivory Coast, Colombia, Cameroon, South Korea, France, Gabon, French, Guyana, Hungary, Japan, Kenya, Morocco, Madagascar, Mali, Portugal, Russia, Rwanda, Spain, Thailand, USA and Vietnam. Figure 4 shows the world map where these isolates were collected. Another batch of field isolates collected from different regions of China was added into analysis. Single spores were isolated from each of these strains and plated on complete media agar plates (0.75% yeast extract, 0.75% casamino acid, 0.1% sucrose and 1% agar) covered with desiccated filter paper, which were then incubated at room temperature for 7 days. Fungal mycelia were stored on filter papers at -20°C.

### DNA preparation

Genomic DNA was extracted using an in-house method optimized for fungal DNA. Isolates were transferred and grown in liquid complete media at room temperature in dark condition with shaking for 7 days. Mycelia were collected using funnel, squeezed to get rid of water and frozen dried overnight. Dried mycelia were grounded into powder with a pestle and petrol. One quarter of 2 ml tube was filled with mycelia powder and mixed with DNA extraction buffer (100 mM Tris-HCl pH=8.0, 100 mM EDTA pH=8.0 and 250 mM NaCl) and proteinase K, followed by incubating at 50 °C for 1 hr. 100 ul of 10% N-laurylsarcosyl was then added and incubated at 55°C for 1hr. Supernatant containing genomic DNA was obtained by centrifuge for 15 mins at 5,000 rpm at room temperature. Genomic DNA was extracted and purified using phenol-chloroform extraction method [28]. Final concentration of genomic DNA was adjusted to 50 ng/ul.

### PCR amplification and sequencing

The ORFs and promoter regions of *AvrPiz-t* were amplified from genomic DNA of each Global strains with 2F/2R and 2PF/2PR primer pairs respectively, while for Chinese strains 9F/10R and 13F/14R were used (Table 1). *Taq* PCR kit (New England Biolabs, Inc., MA, USA)

was used to perform 20 µl PCR reactions: 2 ul genomic DNA (25 ng), 2 ul thermo-buffer, 1 ul dNTP (400 mM of each dNTP), 1 ul of 10 uM forward primer, 1 ul of 10 uM reverse primer, 0.5 ul *Taq* and 12.5 ul distilled H<sub>2</sub>O. Reactions were performed using the following cycle: 95°C for 3min, 25 cycles of 95°C for 1 min, 55-60°C for 30 seconds (varies with different primer pairs) and 72°C for 1 min, followed by a final extension step at 72°C for 7 min. The size of amplified fragment was estimated by electrophoresis gel with 1kb DNA ladder. All amplicons were purified using QIAquick PCR Purification Kit (Qiagen Inc., CA, USA) and sent for Sanger sequencing with 2F/2R and 2PF/2PR for coding region and promoter region respectively from both forward and reverse direction.

### Southern hybridization

Genomic DNA of selected isolates was digested with *EcoRV* and *BamHI* restriction enzymes (New England Biolabs Inc., USA). The digestion scheme diagram was shown in Figure 2. Digested genomic DNA was purified using phenol-chloroform method and electrophoresed on a 1.5% agarose gel at 40 voltages overnight to separate fragments by size. Fragments were transferred onto Hybond N+ membrane by capillary blotting. Probe was amplified with primer pair 2F and 2R, labeled and hybridized onto DNA membrane. Signals were detected on CXS high speed blue film. All performances were followed by the instruction of Amersham ECL Direct Labeling and Detection System kit (GE Healthcare Life Sciences, PA, USA).

### Rice cultivar and pathogenicity assay

To evaluate virulence of *AvrPiz-t* polymorphic isolates on different hosts, 7-8 week-old rice cultivar Toride (for Global strains) or IRBL11 (for Chinese strains) containing *Piz-t*, and Nipponbare lacking *Piz-t* were used in this study. Conidia of selected strains were harvested from 7-day-old V8 juice agar plates and suspended in 250 ppm Tween 20 with a concentration of 5×10<sup>5</sup> conidia per ml. Ten ul conidial suspension of each strains were dropped onto punctuated rice leaves and wrapped with transparent tape, followed by incubation at 25°C in dark and moisture incubator. Disease severity was evaluated 7 days after inoculation using the rating system described by Valent [29].

### Data mining and sequence analysis

ORFs of the *AvrPiz-t* gene in each isolates were assembled and aligned by CLC sequence viewer 6.0 (CLC bio Inc.). Haplotypes, polymorphic sites, sliding window analysis and natural selection test were performed by DnaSP 5.0 [30].

## Results

Four primer sets were used in this study (Table 1). Two primer sets were used for global strains ORF amplification and promoter

Haplotypes	Positions									
	27	56	68	80	120	156-159	178	192	238	325
I										T/C
II			G/T							
III									G/C	
IV							G/A			
V	G/T	G/A			G/C	AGGG/CAAC			G/C	
VI		G/A			G/C	AGGG/CAAC			G/C	
VII								A ins		
VIII				T del						

**Table 1:** The SNP, insertion, and deletion identified in *AvrPiz-t* coding region and their positions. Isolates were categorized into 8 different haplotypes based on the comparison to sequence from strain KJ201.

amplification (Figure 1), and another two primer sets were designed and used specifically for Chinese strains. Out of the total 711 isolates, 606 had their *AvrPiz-t* promoters successfully amplified and 637 had their full ORFs amplified. The possible reason for the failure of PCR amplification may be due to either the high rate of polymorphisms occurring in the primer regions, or deletions/insertions, which greatly alter the length of amplified region. The sizes of promoter amplicons were compared between isolates via agarose gel electrophoresis to identify the existence of transposons. To analyze for polymorphisms in the coding region, selected ORF amplicons were subjected to Sanger sequencing.

### Sequence diversity in the promoter region

The promoter regions of Chinese strains showed a higher PCR amplification rate (394 successes out of 398) compared to the amplification rate of Global strains (212 successes out of 313). Based on the analysis of the amplified promoter regions in, 287 isolates out of 394 Chinese strains and 197 out of 212 Global strains showed the same size as reference avirulent strain KJ201. In the 15 Global strains with different size, 4 isolates including GUY11 contain a 4 kb promoter region, and 11 isolates contain a 5.8 kb promoter region.

As shown in the work of Li et al. [19], a Pot3 transposon inserted in the promoter region of this gene was detected for GUY11. Sequencing of random picked samples from Chinese strains validated at least two types of insertions: a “retrotransposon Inago2” insertion at 41 bp upstream ORF, and a 1870 bp insertion at 462 bp upstream the ORF.

### Southern hybridization validation of transposon insertions

There were 101 Global strains that failed in amplification of their promoter regions. The failure may be due to transposon insertion in the promoter region that interrupts in the primer binding sequences, or to large transposon element insertion that makes the region length exceed PCR limit. In order to verify the transposon insertion in the promoter regions for these isolates, Southern hybridization was designed using a probe covering the *AvrPiz-t* coding region and 170 bp upstream promoter sequences (Figure 1). This analysis was applied to Global strains that have ORF amplicon but failed in promoter amplification. Three strains, 92A8, ZN61, ZN62, were randomly picked for validation, while KJ201, GUY11 and two other strains IE1 and 72A53 were chosen as controls. As shown in Figure 2, there is a 3 kb size difference between GUY11 and KJ201 controls in their promoter region. The two control strains IE1 and 72A53 which were known to have no TE insertion showed the same product size as KJ201. ZN61 showed the same size hybridizing band as GUY11, while 92A8 and ZN62 showed a 2 kb insertion compared to KJ201.

### Polymorphisms in ORF region

The low diversity in the *AvrPiz-t* ORF region has been confirmed in this study. In Chinese strains, more than half of the isolates collected from Taiwan showed a 1858 bp insertion in coding region at 211 bp downstream of the start codon (Figure 3). This long insertion altered the structure of the *AvrPiz-t* protein as well as its function, which has been demonstrated to gain virulence to *Piz-t* containing rice in pathogenicity assay.

Of all the 243 Global strains with ORF sequenced, only 14 strains showed polymorphisms in the ORF region compared to the KJ201 gene sequence. These 14 *AvrPiz-t* sequences represented 8 different haplotypes. Table 1 and Figure 3 showed the 8 haplotypes of SNPs and protein differences, respectively. All DNA polymorphisms led to

altered protein sequences except the SNP at 27 bp and 120 bp. Frame shift occurs in haplotypes VII and VIII, thus the sequences of the entire protein changed after the inserted/deleted position and the proteins are expected to lose function. Table 2 shows hosts and country of origin the 14 strains. Haplotypes V and VI are very similar to each other except that there is an additional polymorphism site 27 in haplotype V. Another common polymorphism site among haplotypes III, V and VI is at position 238 with a G to C substitution and the absence of nucleotide insertions or deletions. In the survey, three fungal strains belong to haplotype III, five fungal strains belong to type VI and one strain belongs to type V, which were collected from Asia, Africa and South America, respectively. Each has hosts that belong to weed species except one mainly infects *Zea mays* in haplotype VI. The remaining four isolates collected from Africa, Asia, North and South America on rice are classified into four different polymorphism types. Isolates with a grass host were separated from isolates that can infect rice based on the polymorphism similarity in *AvrPiz-t* coding region.

Notably, three interesting findings can be summarized from the table: 1) Three out of eight haplotypes are detected in non rice strains only; 2) Rice strains do not share haplotypes with non-rice strains; 3) Haplotype VI is shared by distant strains in the phylogeny.

### Natural selection force assay

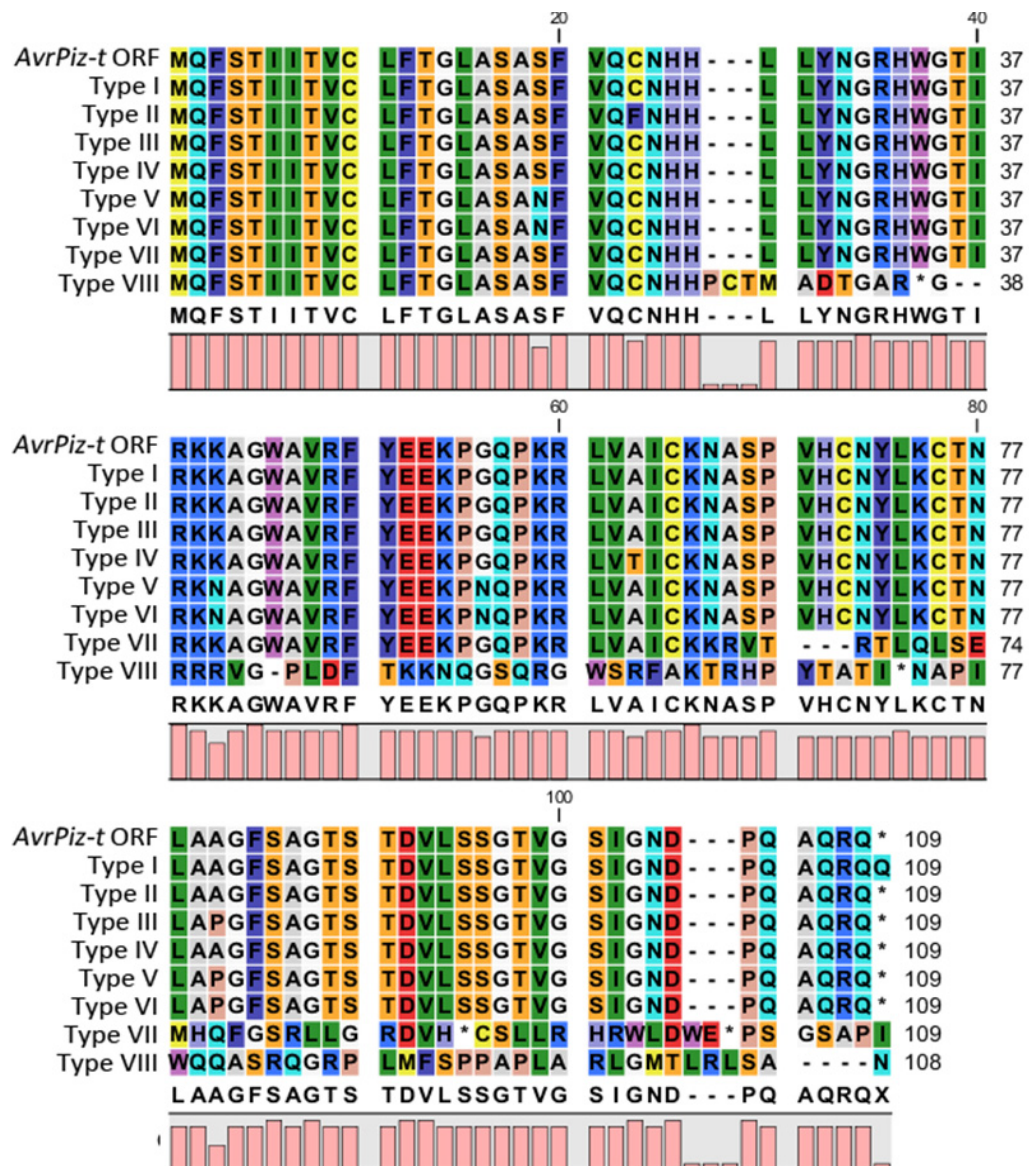
The nucleotide diversity level was calculated based on the ORF sequences from Global strains. To estimate the number of nucleotide diversity per site [31,32],  $\pi$  ( $\pi=0.0023$ ) and  $\theta$  ( $\theta=0.014$ ) were measured on the entire *AvrPiz-t* ORF. The  $\pi$  value is lower than that of the reported *Avr-Pita1* sites within Thai isolates population ( $\pi=0.00891$ ) [17]. A sliding window analysis of  $\pi$  value across the entire coding region was performed, as shown in Figure 4. A major peak can be found from 150-160bp in the variation distribution of *AvrPiz-t* alleles.

To investigate whether the *AvrPiz-t* gene is under directional selection force, its ORF sequence was examined with three statistical parameters: Tajima's D [33], Fu and Li' D and Fu and Li's F [34]. As shown in Table 3, the value of these three indicators is -2.307 ( $p<0.01$ ), -6.010 ( $p<0.01$ ) and -5.449 ( $p<0.01$ ). The negative value indicates that this locus is undergoing a directional natural selection. To further determine the direction of selection power, the ratio of non-synonymous and synonymous rate was calculated, with the assumption that ratio equals 1 indicating neutral selection. The ratio calculated for *AvrPiz-t*  $\pi_{non}/\pi_{syn} = 1.286$ , which is greater than 1, indicates that *AvrPiz-t* is under a positive selection.

### Pathogenicity assays

Identified polymorphisms in either the *AvrPiz-t* promoter or coding regions may alter its function and thus change pathogenicity. To evaluate the effect of insertion on protein function, all the Chinese strains were used for pathogenicity assays (see inoculation sheet for detailed results). It can be summarized from the results that while all the avirulent strains contain wild type *AvrPiz-t* promoter and coding regions, 72.8% (107 of 147) of the virulent strains contain size variation in promoter regions, 21.8% (32 of 147) contain insertions in coding regions, 2.7% (4 of 147) failed in promoter or coding region amplifications, and 2.7% (4 of 147) showed no obvious size variations. The fact that majority of virulent mutant strains comes from the promoter size variation group suggests that TE insertion in promoter region plays an important role in the ability for *M. oryzae* to break *Piz-t* gene resistance in host.

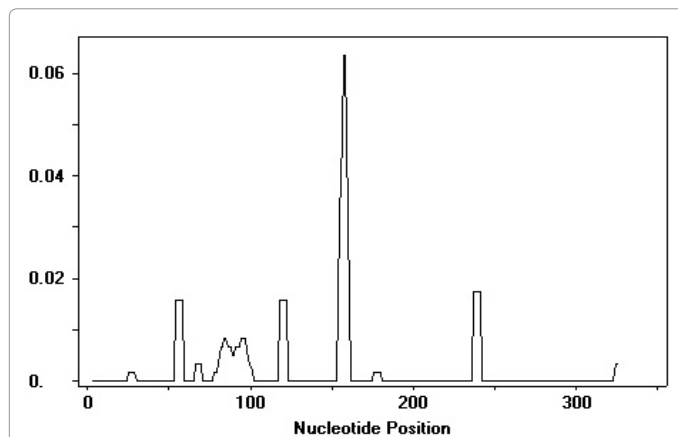
In addition to insertions, SNPs observed in coding regions



**Figure 3:** Alignment of the amino acid sequence of the *AvrPiz-t* protein in KJ201 (the first row) and in 8 haplotypes. Height of bars in the last row represent the conserve level at this position.

Haplotypes	Strains	Countries	Host
I	BD0024	Burundi	<i>Oryza sativa</i>
	FC23	Columbia	<i>Oryza sativa</i>
III	PH0059	Philippines	<i>Leersia hexandra</i>
	PH0075	Philippines	<i>Brachiaria nutica</i>
	VT0032	Vietnam	<i>Leersia hexandra</i>
IV	TH0012	Thailand	<i>Hordeum vulgare</i>
V	KN0006	Kenya	<i>Hordeum vulgare</i>
VI	BR0032	Brazil	<i>Triticum</i>
	BR0062	Brazil	<i>Eleusine indica</i>
	GN0001	Gabon	<i>Zea mays</i>
	JP0028	India	<i>Eleusine sp.</i>
	KN0001	Japan	<i>Eragrostis curvala</i>
	KN0001	Kenya	<i>Hordeum vulgare</i>
VII	49D	US	<i>Oryza sativa</i>
VIII	EG85	Egypt	<i>Oryza sativa</i>

**Table 2:** Isolates in different haplotypes with their hosts and collected countries.



**Figure 4:** Distribution of nucleotide variation along *AvrPiz-t* coding region. X axis is the nucleotide position and Y axis is the nucleotide diversity  $\pi$  value.

Coding sites	H <sup>a</sup>	S <sup>b</sup>	$\pi$	$\theta$	Tajima's D	Fu and Li's D	Fu and Li's T
Coding	8	28	0.0023	0.014	-2.307*	-6.101*	-5.449*

<sup>a</sup>: Number of haplotypes

<sup>b</sup>: Number of polymorphic (segregating) site, number of polymorphic (segregating) site's estimator,

\*: significant statistics < 0.01

**Table 3:** Polymorphism test of *AvrPiz-t* coding region among population.

may also affect protein function. To estimate the effect of *AvrPiz-t* ORF mutation to the fungal virulence, several strains from different haplotypes were chosen for pathogenicity assays, including strain BD0024 (Type I), FC23 (Type II), 49D (Type VII), and EG85 (Type VIII). The four strains, as well as negative control strain KJ201, were inoculated on rice cultivar Nipponbare (NPB) and the *Piz-t* harboring cultivar Toride. Figure 5 shows that NPB is sensitive to KJ201, which induces a hypersensitive reaction on rice cultivar Toride. EG85 and FC23 can overcome resistance mediated by *Piz-t* and established growth on Toride, while 49D cannot infect either of these two cultivars. According to the DNA sequences at the *AvrPiz-t* locus of these strains, a 'T' insertion caused frame shift in the EG85 ORF and an early stop codon resulting in a truncated 36 amino acid protein. This suggests that this truncated protein is not a functional factor and cannot trigger host defense. A single base mutation in the FC23 ORF led to an amino acid change close to the N terminal suggesting that this region may contain a core recognition signal or is critical for maintaining the protein structure. In addition, an 'A' insertion also occurred in the 49D ORF but closer to the C terminal ending up with a truncated 88 amino acid protein suggesting that the functional recognition signal is close to the N terminus.

Strain BD0024 was also tested for pathogenicity and showed avirulence result (Data not show). These results suggest that some modifications in the protein may not alter avirulence (ie recognition) or that other avirulence genes in this strain triggered Toride 1 resistance (which may have several R genes).

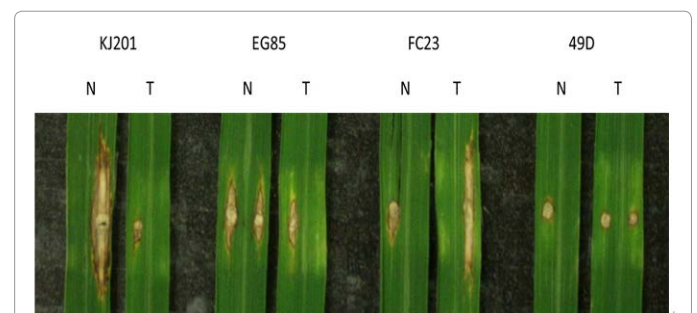
## Discussion

Successful plant defense relies on recognition and interaction with pathogen effectors by R genes. The detailed molecular mechanisms describing how pathogens overcome specific resistance gene defense is not well understood, but it is certain that the instability of AVR genes leads to the gain of virulence. To better understand this association,

genome organizations at AVR gene loci have been investigated. For instance, the *AvrPi-ta1* gene in *M. oryzae* has been well studied for years. It has been demonstrated to interact with its cognate *Pi-ta* gene in rice directly [35,36], and one study shows that coding region polymorphisms exist in 5 out of 11 *M. oryzae* field isolates collected in China [37]. In another study on *Avr-Pita1* alleles of 151 US isolates, 26 haplotypes were identified in the coding region based on DNA sequencing [17]. The high genetic diversity of the *Avr-Pita1* locus is well supported by the fact that it is located in the subtelomeric region, a highly unstable region [26,38]. In addition, several other AVR genes have been mapped near the telomeric region including *Avr-Pii*, *Avr-Pia*, *Avr-Pit*, *Avr1-Ku86*, *Avr1-MedNoi* and *PWL1* [11,39-41]. Another example on the promoter of *Avr-CO39* showed that repetitive sequences including REP1, RETRO5 and MGR691/MGR508 were identified at the 5' terminus of the *Avr-CO39* locus in *M. oryzae* [24]. In this study, we conducted similar analysis, and 8 ORF haplotypes were identified among 243 sequenced isolates, in which haplotypes II and VIII showed gain of virulence in the pathogenicity assays compared to avirulent isolate KJ201, suggesting the possible association of virulence with DNA sequence diversity in the coding region. The coding region diversity is lower compared to the previous report of *Avr-Pita1* locus. However, the result is consistent with the study conducted by Yoshida et al., who found that the majority (78%) of 1032 analyzed the AVR loci are identical in field isolates [42].

The *AvrPiz-t* gene resides in a region that comprises high transposable element density on chromosome 7 [43]. It was further identified that a Pot3 transposable element inserted in the 500 bp upstream region in the virulent isolate GUY11 compared to the avirulent isolate KJ201 [27]. In this study, the fact that more diversity has been identified in the promoter region of the *AvrPiz-t* locus compared to its coding region suggests the function of this gene in a population may be affected from the present/absent polymorphism of transposable element more often than from variation in its coding sequence. In addition to transposon elements inserted in *AvrPiz-t* promoter region in this study, a Pot3 element was shown to be present in the promoter region and coding region of the *Avr-Pita1* gene in two different virulent isolates [25]. Also, a 1.9 kb MINE element was identified in the exon region of AVR gene *ACE1* in a virulence isolates [13]. Combined with the similar findings in other fungi isolates and the AVR loci, transposon elements insertion close or inside coding region of AVR loci are crucial mechanisms the fungi use to protect themselves from being recognized by R genes.

Statistical analysis of the selection force on *AvrPiz-t* suggests that this gene is under positive selection and has favored amino acid



**Figure 5:** Pathogenicity assay of the selected isolates with ORF polymorphisms. Tested strains were marked at top, leaves marked as N represent the rice cultivar Nipponbare lacking the *Piz-t* gene, and leaves marked as T represent the rice cultivar Toride containing the *Piz-t* gene.

substitution in its coding region. This finding is consistent with other studies that *AVR* genes are prone to change in positive selection [17,42,43]. Given the dynamic nature of telomeric regions, repetitive elements and transposon elements, the fact that many *AVR* genes are associated with these regions may provide fungi advantages to modify *AVR* genes adapting to *R* genes. For example, multiple *AVR* haplotypes of *AVR-Pita1* were found in natural populations where only one resistant haplotype of the cognate *R* gene *Pi-ta* was found suggesting that *AVR-Pita1* engages trench warfare with *Pi-ta*.

Now we confirmed the diversity of *AvrPiz-t* and the connection between sequence variance and pathogenicity, a potential method to apply this finding is design biomarkers for *AvrPiz-t* gene that can be used to determine the virulence of the tested strain to corresponding *R* gene. This can serve as an approach to estimate field population containing functional *AVR* or nonfunctional alleles and deploy *Piz-t* resistance effectively.

In conclusion, we suggest that the *AvrPiz-t* alleles in world field pathogen populations have been undergoing a relatively strong positive selection that prefers amino acid substitution. The coding sequence is dynamic and transposon element insertion in the promoter region or the coding region enable *M. oryzae* to prevent them from being recognized by cognate *R* genes in their hosts.

#### Acknowledgements

We thank all investigators who collected fungal isolates and supported this study around the world, and Mr. Jian Chen and Ms. Qinying Qu for analyzing samples. This work was supported the Ohio Agricultural Research and Development Center and by 973 (Project No. 2012CB114001), NSFC (No. 91231121 and No. U1305211). This work was also supported by the IRMA grant (ANR-07-GPLA-007) from the Génoplante program and the GEMO project (ANR-09-GENM-029) from the ANR-Génomique program. The U.S. Department of Agriculture is an equal opportunity provider and employer.

#### References

- Chisholm ST, Coaker G, Day B, Staskawicz BJ (2006) Host-microbe interactions: shaping the evolution of the plant immune response. *Cell* 124: 803-814.
- Jones JD, Dangl JL (2006) The plant immune system. *Nature* 444: 323-329.
- Flor HH (1971) Current status of the gene-for-gene concept. *Annu Rev Phytopathol* 9: 275-296.
- Zeigler RS, Leong SA, Teng PS (1994) Rice Blast Disease. CAB International, Wallingford.
- Ou SH (1985) Blast, Rice Diseases. (2nd edn). Commonwealth Agricultural Bureaux., Slough, UK, pp. 109-201. Pennisi, E., 2010. Armed and dangerous. *Science* 327: 804-805.
- Ballini E, Morel JB, Droc G, Price A, Courtois B, et al. (2008) A genome-wide meta-analysis of rice blast resistance genes and quantitative trait loci provides new insights into partial and complete resistance. *Mol Plant Microbe Interact* 21: 859-868.
- Kiyosawa S (1982) Genetics and Epidemiological Modeling of Breakdown of Plant-Disease Resistance. *Annu Rev Phytopathol* 20: 93-117.
- Li W, Wang B, Wu J, Lu G, Hu Y, et al. (2009) The *Magnaporthe oryzae* avirulence gene *AvrPiz-t* encodes a predicted secreted protein that triggers the immunity in rice mediated by the blast resistance gene *Piz-t*. *Mol Plant Microbe Interact* 22: 411-420.
- Orbach MJ, Farrall L, Sweigard JA, Chumley FG, Valent B (2000) A telomeric avirulence gene determines efficacy for the rice blast resistance gene *Pi-ta*. *Plant Cell* 12: 2019-2032.
- Farman ML, Leong SA (1998) Chromosome walking to the AVR1-CO39 avirulence gene of *Magnaporthe grisea*: discrepancy between the physical and genetic maps. *Genetics* 150: 1049-1058.
- Kang S, Sweigard JA, Valent B (1995) The PWL host specificity gene family in the blast fungus *Magnaporthe grisea*. *Mol Plant Microbe Interact* 8: 939-948.

- Sweigard JA, Carroll AM, Kang S, Farrall L, Chumley FG, et al. (1995) Identification, cloning, and characterization of PWL2, a gene for host species specificity in the rice blast fungus. *Plant Cell* 7: 1221-1233.
- Fudal I, Böhnert HU, Tharreau D, Lebrun MH (2005) Transposition of MINE, a composite retrotransposon, in the avirulence gene ACE1 of the rice blast fungus *Magnaporthe grisea*. *Fungal Genet Biol* 42: 761-772.
- Yoshida K, Saitoh H, Fujisawa S, Kanzaki H, Matsumura H, et al. (2009) Association genetics reveals three novel avirulence genes from the rice blast fungal pathogen *Magnaporthe oryzae*. *Plant Cell* 21: 1573-1591.
- Kasetsomboon T, KNS, Sriwongchai T, Zhou B, Jantaturiyarat C (2013) Sequence variation of avirulence gene AVR-Pita1 in rice blast fungus, *Magnaporthe oryzae*. *Mycological Progress* 12: 617-628.
- Zhou E, Jia Y, Singh P, Correll JC, Lee FN (2007) Instability of the *Magnaporthe oryzae* avirulence gene AVR-Pita alters virulence. *Fungal Genet Biol* 44: 1024-1034.
- Dai Y, Jia Y, Correll J, Wang X, Wang Y (2010) Diversification and evolution of the avirulence gene AVR-Pita1 in field isolates of *Magnaporthe oryzae*. *Fungal Genet Biol* 47: 973-980.
- Xing J, Jia Y, Correll JC, Lee FN, Cartwright R, et al. (2013) Analysis of genetic and molecular identity among field isolates of the rice blast fungus with an International Differential System, Rep-PCR, and DNA Sequencing. *Plant Dis* 97: 491-495.
- Zhou B, Qu S, Liu G, Dolan M, Sakai H, et al. (2006) The eight amino-acid differences within three leucine-rich repeats between Pi2 and Piz-t resistance proteins determine the resistance specificity to *Magnaporthe grisea*. *Mol Plant Microbe Interact* 19: 1216-1228.
- Zhang ZM, Zhang X, Zhou ZR, Hu HY, Liu M, et al. (2013) Solution structure of the *Magnaporthe oryzae* avirulence protein AvrPiz-t. *J Biomol NMR* 55: 219-223.
- Khang CH, Berruyer R, Giraldo MC, Kankanala P, Park SY, et al. (2010) Translocation of *Magnaporthe oryzae* effectors into rice cells and their subsequent cell-to-cell movement. *Plant Cell* 22: 1388-1403.
- Park CH (2012) The *Magnaporthe oryzae* effector AvrPiz-t targets the RING E3 ubiquitin ligase APIP6 to suppress pathogen-associated molecular pattern-triggered immunity in rice. *Plant Cell* 24: 4748-4762.
- Khang CH, Park SY, Lee YH, Valent B, Kang S (2008) Genome organization and evolution of the AVR-Pita avirulence gene family in the *Magnaporthe grisea* species complex. *Mol Plant Microbe Interact* 21: 658-670.
- Farman ML, Eto Y, Nakao T, Tosa Y, Nakayashiki H, et al. (2002) Analysis of the structure of the AVR1-CO39 avirulence locus in virulent rice-infecting isolates of *Magnaporthe grisea*. *Mol Plant Microbe Interact* 15: 6-16.
- Kang S, Lebrun MH, Farrall L, Valent B (2001) Gain of virulence caused by insertion of a Pot3 transposon in a *Magnaporthe grisea* avirulence gene. *Mol Plant Microbe Interact* 14: 671-674.
- Chuma I, Isobe C, Hotta Y, Ibaragi K, Futamata N, et al. (2011) Multiple translocation of the AVR-Pita effector gene among chromosomes of the rice blast fungus *Magnaporthe oryzae* and related species. *PLoS Pathog* 7: e1002147.
- Li P, Bai B, Zhang HY, Zhou H, Zhou B (2012) Genomic organization and sequence dynamics of the AvrPiz-t locus in *Magnaporthe oryzae*. *J Zhejiang Univ Sci B* 13: 452-464.
- Chomczynski P, Sacchi N (2006) The single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction: twenty-something years on. *Nat Protoc* 1: 581-585.
- Valent B, Farrall L, Chumley FG (1991) *Magnaporthe grisea* genes for pathogenicity and virulence identified through a series of backcrosses. *Genetics* 127: 87-101.
- Rozas J, Sánchez-DelBarrio JC, Messeguer X, Rozas R (2003) DnaSP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* 19: 2496-2497.
- Saitou N, Nei M (1987) On the Maximum-Likelihood Method for Molecular Phylogeny. *Japanese Journal of Genetics* 62: 547-548.
- Watterson GA (1975) On the number of segregating sites in genetical models without recombination. *Theor Popul Biol* 7: 256-276.



33. Tajima F (1989) Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* 123: 585-595.
34. Fu YX, Li WH (1993) Statistical tests of neutrality of mutations. *Genetics* 133: 693-709.
35. Bryan GT, Wu KS, Farrall L, Jia Y, Hershey HP, et al. (2000) tA single amino acid difference distinguishes resistant and susceptible alleles of the rice blast resistance gene Pi-ta. *Plant Cell* 12: 2033-2046.
36. Jia Y, McAdams SA, Bryan GT, Hershey HP, Valent B (2000) Direct interaction of resistance gene and avirulence gene products confers rice blast resistance. *EMBO J* 19: 4004-4014.
37. Ma BT, Qu GL, Shi J, Chen de X, Lin YF, et al. (2008) [Genetic diversity of AVR-pita alleles of rice blast fungus *Magnaporthe grisea*]. *Fen Zi Xi Bao Sheng Wu Xue Bao* 41: 495-499.
38. Starnes JH, Thornbury DW, Novikova OS, Rehmeyer CJ, Farman ML (2012) Telomere-targeted retrotransposons in the rice blast fungus *Magnaporthe oryzae*: agents of telomere instability. *Genetics* 191: 389-406.
39. Yasuda N, Noguchi MT, Fujita Y (2006) Partial mapping of avirulence genes AVR-Pii and AVR-Pia in the rice blast fungus *Magnaporthe oryzae*. *Canadian Journal of Plant Pathology-Revue Canadienne De Phytopathologie* 28: 494-498.
40. Diah W, Tharreau D, Notteghem JL, Orbach M, Lebrun MH (2000) Mapping of avirulence genes in the rice blast fungus, *Magnaporthe grisea*, with RFLP and RAPD markers. *Mol Plant Microbe Interact* 13: 217-227.
41. Chen QH, Wang YC, Li AN, Zhang ZG, Zheng XB (2007) Molecular mapping of two cultivar-specific avirulence genes in the rice blast fungus *Magnaporthe grisea*. *Mol Genet Genomics* 277: 139-148.
42. Miki S, Matsui K, Kito H, Otsuka K, Ashizawa T, et al. (2009) Molecular cloning and characterization of the AVR-Pia locus from a Japanese field isolate of *Magnaporthe oryzae*. *Mol Plant Pathol*. 10: 361-374.
43. Thon MR, Martin SL, Goff S, Wing RA, Dean RA (2004) BAC end sequences and a physical map reveal transposable element content and clustering patterns in the genome of *Magnaporthe grisea*. *Fungal Genet Biol* 41: 657-666.

**Citation:** Chen C, Chen M, Hu J, Zhang W, Zhong Z, et al. (2014) Sequence Variation and Recognition Specificity of the Avirulence Gene *AvrPiz-T* in *Magnaporthe Oryzae* Field Populations. *Fungal Genom Biol* 4: 113. doi:[10.4172/2165-8056.1000113](https://doi.org/10.4172/2165-8056.1000113)

### Submit your next manuscript and get advantages of OMICS Group submissions

#### Unique features:

- User friendly/feasible website-translation of your paper to 50 world's leading languages
- Audio Version of published paper
- Digital articles to share and explore

#### Special features:

- 350 Open Access Journals
- 30,000 editorial team
- 21 days rapid review process
- Quality and quick editorial, review and publication processing
- Indexing at PubMed (partial), Scopus, EBSCO, Index Copernicus and Google Scholar etc
- Sharing Option: Social Networking Enabled
- Authors, Reviewers and Editors rewarded with online Scientific Credits
- Better discount for your subsequent articles

Submit your manuscript at: <http://www.omicsonline.org/submission>

