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Adaptation of *Staphylococcus xylosus* to Nutrients and Osmotic Stress in a Salted Meat Model

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Staphylococcus xylosus is commonly used as starter culture for meat fermentation. Its technological properties are mainly characterized *in vitro*, but the molecular mechanisms for its adaptation to meat remain unknown. A global transcriptomic approach was used to determine these mechanisms. *S. xylosus* modulated the expression of about 40–50% of the total genes during its growth and survival in the meat model. The expression of many genes involved in DNA machinery and cell division, but also in cell lysis, was up-regulated. Considering that the *S. xylosus* population remained almost stable between 24 and 72 h of incubation, our results suggest a balance between cell division and cell lysis in the meat model. The expression of many genes encoding enzymes involved in glucose and lactate catabolism was up-regulated and revealed that glucose and lactate were used simultaneously. *S. xylosus* seemed to adapt to anaerobic conditions as revealed by the overexpression of two regulatory systems and several genes encoding cofactors required for respiration. In parallel, genes encoding transport of peptides and peptidases that could furnish amino acids were up-regulated and thus concomitantly a lot of genes involved in amino acid synthesis were down-regulated. Several genes involved in glutamate homeostasis were up-regulated. Finally, *S. xylosus* responded to the osmotic stress generated by salt added to the meat model by overexpressing genes involved in transport and synthesis of osmoprotectants, and Na⁺ and H⁺ extrusion.

Keywords: *Staphylococcus xylosus*, meat, transcriptome, nutrient adaptation, osmotic stress

INTRODUCTION

Meat fermentation is an old process that relies on indigenous microbiota and is nowadays carried out under controlled conditions with addition of starter cultures. Meat starter cultures are composed of a combination of lactic acid bacteria and coagulase-negative staphylococci (Ravvyts et al., 2012). *Staphylococcus xylosus* and *Staphylococcus carnosus* are the two main staphylococcal species used as starter cultures. They are associated with different species of *Lactobacillus*, such as *Lactobacillus sakei* or *Lactobacillus curvatus* or *Pediococcus*. Studies over decades of microorganisms used as starters have yielded information about their potential functionalities in meat products. Lactic acid bacteria are well-known acidifiers and bacteriocin producers that affect the technological properties and the microbial stability of products (Leroy et al., 2006). Staphylococci are recognized to play an important role in the color formation through their nitrate

reductase activity (Talon and Leroy, 2006). They also contribute to the flavor development of meat products *via* their catabolism of branched-chain amino acids in odorous volatile methyl compounds (Olesen and Stahnke, 2004; Tjener et al., 2004), their antioxidant properties avoiding rancidity (Barrière et al., 2001; Rosenstein et al., 2009) and their catabolism of pyruvate in diacetyl and acetoin responsible of buttery aroma (Søndergaard and Stahnke, 2002).

The development of -omic techniques has allowed the study of the metabolism of starter cultures *in situ* in the food matrix. The transcriptomic response of *Lactococcus* in milk or cheese has been established (Cretenet et al., 2011; Taïbi et al., 2011), while the proteome of *Streptococcus thermophilus* was studied in milk and the release of bacterial proteins was studied in cheese (Gagnaire et al., 2004; Derzelle et al., 2005).

Very few studies have focused on physiology of meat starter cultures in meat matrices. They concern essentially *L. sakei*. *In vivo* expression technology (IVET) was applied to investigate gene expression of *L. sakei* during meat fermentation (Hüfner et al., 2007). This study revealed the expression of genes encoding proteins that contribute to stress-related functions. A proteomic approach highlighted that *L. sakei* overexpressed dipeptidases in the presence of sarcoplasmic extract and proteins related to energy and pyrimidine metabolism in the presence of myofibrillar extract (Fadda et al., 2010). Concerning staphylococci, the single transcriptomic concerning the response of *S. xylosus* to nitrate and nitrite in meat revealed that *S. xylosus* was subject to nitrosative stress and overcame it by expressing genes involved in iron homeostasis and antioxidant defense (Vermassen et al., 2014).

Functional genomics give us an opportunity to gain insight into the physiological and metabolic capabilities of starters *in situ* in the food matrix. As the molecular mechanisms involved in the adaptation of the bacteria in meat products are little known, we analyzed the transcriptome of *S. xylosus* in a meat model incubated for 72 h at 22°C in conditions that mimic the fermentation step. The *in situ* response of *S. xylosus* was analyzed vs. the *S. xylosus* culture used as inoculum. The results highlighted a total change in gene expression during survival of *S. xylosus* in the meat model, which reflects adaptation to nutrients and mainly to osmotic stress.

MATERIALS AND METHODS

Preparation of the Inoculum and Inoculation of the Meat Model

The *S. xylosus* C2a strain is derived from the type strain DSM20267 cured of its endogenous plasmid pSX267. Its complete genome has been sequenced (LN554884). This strain grows and behaves in meat models and in batter as strains isolated from meat products (our unpublished data). We used this laboratory strain as it is the only one of this species with some genetic background. The strain was cultured overnight at 30°C with shaking (150 rpm) in a minimal medium as already described (Fiegler and Brückner, 1997). The culture was briefly centrifuged and part of the cell pellet called “the inoculum” was

immediately frozen in liquid nitrogen before extraction of RNA. Another part of the cell pellet was resuspended in physiological serum and inoculated in a pork meat model containing 0.5% glucose and 0.47 M NaCl as already described in Vermassen et al. (2014). Petri dishes were completely filled with inoculated pork meat and incubated at 22°C in a wet atmosphere for up to 72 h. These conditions probably created a gradient of oxygen in the samples. At 24, 48, and 72 h of incubation at 22°C, 200 mg meat samples were taken and immediately frozen in liquid nitrogen to stabilize the bacterial RNA. Three independent experiments were done.

To evaluate the inoculum and the growth of *S. xylosus* in meat, bacteria were enumerated after serial dilution on plates of brain-heart infusion agar, which were incubated at 30°C for 24 h.

Physical and Chemical Analysis of the Meat Model

A digital pH meter (MP230, Mettler Toledo, Viroflay, France) with a Inlab[®] 413 electrode was used for measurement of meat pH.

Meat samples (2 g) were homogenized for 30 min with 8 mL of distilled water. The homogenate was then centrifuged for 20 min at 10,000 g at 4°C and the supernatant was filtered. The analyses were carried out as previously described (Nouaille et al., 2009). Briefly, glucose and fermentation products were analyzed by high-performance liquid chromatography (HPLC) using a Bio-Rad HPX87H column under the following conditions: a temperature of 48°C, 5 mM H₂SO₄ as the eluent at a flow rate of 0.5 mL min⁻¹ and dual detection by refractometer and UV analyses. Free amino acids were analyzed after protein precipitation of the supernatant and derivatization. The derivatives were separated by HPLC on a Hypersil AA-ODS column at 40°C by a linear gradient of acetate buffer (pH 7.2) with triethylamine (0.018%), tetrahydrofuran (0.3%) and acetonitrile (60%). Diode array detectors were used at 338 and 262 nm for derivatives.

RNA Extraction, Labeling and Microarray Analyses and Validation

The RNA extraction and labeling of *S. xylosus* either from the inoculum or directly in meat after 24, 48, or 72 h of incubation were carried out as described in Vermassen et al. (2014).

A complete description of the array developed for *S. xylosus* C2a is available at the NCBI Gene Expression Omnibus (GEO) database under platform accession number GPL19201.

Microarrays were analyzed as described in Vermassen et al. (2014). Significant differences in the probe set intensities between the two conditions were identified using a linear model with an empirical Bayes method using all information probes to moderate the standard errors of the estimated log-fold changes (Smyth, 2004). The probabilities were corrected by the Benjamini-Hochberg procedure in order to control the false-discovery rate (FDR) with a *p*-value cut-off of 0.05. All the probes with an FDR = 0.05 are considered to be differentially expressed. Finally, a gene was considered to be differentially expressed if at least 50% of the corresponding probes were differentially

expressed and if the ratio of expression was above 2 or lower than 0.5.

Microarray data were validated as described in Vermassen et al. (2014). The targeted genes for qPCR and primer sequences are listed in Supplementary Table 1. The analyses were performed on the same samples of RNA as used for the microarray experiments. The relative fold change of gene expression, using measured *tuf* housekeeping gene expression, was determined by the $2^{-\Delta\Delta Ct}$ method (Livak and Schmittgen, 2001).

Microarray Data Accession Number

The microarray samples and data have been deposited in the GEO database under accession number GSE69743.

RESULTS AND DISCUSSION

Growth and Survival of *S. xylosus* in the Meat Model and Transcriptome Profile

S. xylosus was collected at the early stationary phase of growth in a minimal medium and was inoculated in the meat model at 7.7 log CFU/g. The growth of *S. xylosus* in the meat model was exponential until 24 h and reached 9.0 log CFU/g and remained almost at this population level until the end of the experiment (72 h).

The *in situ* *S. xylosus* response revealed a global change in gene expression during its survival in the meat model by comparison with the inoculum. There were 1337 (658 down- and 679 up-regulated), 1326 (659 down- and 667 up-regulated), and 1070 (540 down- and 530 up-regulated) genes differentially expressed at 24, 48, and 72 h, respectively (Supplementary Table 2). Notably, 838 genes were differentially expressed at the three times of incubation. This indicated that major transcriptional changes had occurred at 24 h and lasted throughout the incubation (72 h). These genes were classified into different functional categories: the most represented being information storage and processing, cellular processes and metabolism.

To validate the microarray analysis independently, the relative expression of 45 differentially expressed genes representing more than 5% of the common genes with significantly modified expression was measured by qPCR (Supplementary Table 1). The microarray and qPCR results for the tested genes were strongly correlated for the three times of incubation (24 h: $r^2 = 0.931$ and slope = 1.235x, 48 h $r^2 = 0.918$ and slope = 1.273x, 72 h $r^2 = 0.893$ and slope = 1.394x) and the expected trend in the expression pattern was confirmed (Supplementary Figure 1).

The inoculum of *S. xylosus* was grown in a minimal medium with glucose as sole carbon source, NH_4 as nitrogen source, 3 vitamins and trace elements. It then has to adapt its physiology to a pork meat model medium with an approximate composition (expressed as g/per 100 g) of proteins (15–22), lipids (1.5–4.0), minerals and trace amounts of carbohydrate (Toldrá, 2007) with glucose and NaCl added in our conditions. Furthermore, it has to adapt to different environmental conditions: pH variations (pH 5.9 in meat vs. 7.0), solid medium vs. liquid one.

Cell Division and Cell Lysis

Even though *S. xylosus* began a plateau phase at 24 h, it remained metabolically active in the meat model until 72 h, as revealed by the modulation of 60 genes involved in information processing and storage and 36 genes involved in cellular processes (cell division, cell wall/membrane biogenesis) (Table 1, Supplementary Table 2).

The response of the information storage group consisted of 60 genes that were mostly up-regulated, 19 of which were involved in DNA replication and recombination, such as polymerases (*dnaE*, SXYL_01799), topoisomerase IV (*parCE*), helicases (*dnaB*, *ruvB*, *recG*, *recQ1*, *addA*), gyrases (*gyrAB*) and mismatch repair (*mfd*, *mutS1*, *mutS2*) (Table 1). Thirty genes were involved in translation, such as genes encoding ribosomal proteins, amino-acyl tRNA ligases and proteins involved in synthesis of queuosines, which are modified nucleosides present in certain tRNAs (Supplementary Table 2). Furthermore, the gene *fusA* encoding the elongation factor G and the gene *lepA*, a ribosomal elongation factor that recognizes ribosomes after a defective translocation reaction and induces a back-translocation, and two genes *infA* and *infC* encoding translation initiation factors were up-regulated (Table 1).

Fourteen genes involved in nucleotide transport and metabolism were up-regulated (Table 1). This metabolism derived from the salvage of preformed extracellular nucleobases as highlighted by *pbuX* encoding a xanthine permease and *pyrP* a uracil permease. In meat, ATP is rapidly split into ADP and subsequently into AMP and IMP, which is degraded to inosine and hypoxanthine and further oxidized to xanthine (Battle et al., 2000, 2001). Other nucleotide metabolites are present in pork meat such as uracil (Battle et al., 2000). Together with the gene *pbuX*, three other genes (*xpt*, *hpt*, *purA*) encoding functions involved in the conversion of purine bases into nucleotides (IMP, XMP, GMP) were overexpressed. Adenine and guanine compounds are interconvertible through the intermediate formation of IMP. Similarly, in addition to *pyrP*, eight genes (*pdp*, *upp*, *pyrBC*, *carAB*, *pyrE*, *pyrG*) encoding functions involved in the production of UMP and CTP were identified. These genes encode enzymes that belong to either purine or pyrimidine salvage and interconversion pathways as well described for *Bacillus subtilis* (Switzer et al., 2002). The ability of coagulase-negative staphylococci to metabolize adenosine and inosine in a meat simulation medium has been demonstrated and these compounds may serve as potential alternative energy sources (Janssens et al., 2014).

Eight genes involved in cell division were overexpressed (Table 1). Division requires the participation of numerous proteins that can be functionally discriminated in Z-ring organizing proteins, which are involved in peptidoglycan construction and regulatory proteins (den Blaauwen, 2013). The first step is the polymerization of the FtsZ-ring with a number of Z-ring associated proteins such as FtsA and ZapA encoded by *ftsA* and *zapA*. The genes *divIB*, SXYL-00848 encoding proteins associated with septum synthesis and *mreD* involved in cell shape were up-regulated. In *S. xylosus* the genes *ftsA* and *divIB* belonged to a cluster of genes governing cell division and peptidoglycan synthesis as already established for several bacteria including

TABLE 1 | Genes of *Staphylococcus xylosum* discussed in this study differentially expressed over time in meat.

Gene ID	Gene name	Description	Mean ratio of expression		
			24 h	48 h	72 h
CELL DIVISION AND CELL LYSIS					
Replication, recombination, repair, and transcription					
SXYL_01165	<i>dnaE</i>	DNA polymerase III subunit alpha	2.6	3.5	2.6
SXYL_01799		DNA polymerase X family protein	2.4	2.6	2.0
SXYL_01529-30	<i>parCE</i>	DNA topoisomerase 4 subunit A, B	5.0*	3.8*	2.9*
SXYL_00016	<i>dnaB</i>	Replicative DNA helicase	4.9	4.4	3.7
SXYL_01219	<i>ruvB</i>	Holliday junction ATP-dependent DNA helicase RuvB	4.2	3.4	2.5
SXYL_01659	<i>recG</i>	ATP-dependent DNA helicase RecG	3.8	4.0	2.6
SXYL_02129	<i>recQ1</i>	ATP-dependent DNA helicase RecQ	4.8	7.0	5.0
SXYL_01954	<i>addA</i>	ATP-dependent helicase/nuclease subunit A	4.0	3.4	2.4
SXYL_00005-06	<i>gyrAB</i>	DNA gyrase subunits A and B	3.7*	3.9*	2.8*
SXYL_02427	<i>mfd</i>	Transcription-repair-coupling factor	5.8	6.9	4.2
SXYL_01583	<i>mutS1</i>	DNA mismatch repair protein MutS	3.6	4.2	2.9
SXYL_01798	<i>mutS2</i>	Endonuclease MutS2	3.2	4.8	3.5
SXYL_02373	<i>fusA</i>	Elongation factor G	3.5	2.4	
SXYL_01273	<i>lepA</i>	Elongation factor 4	8.4	8.2	5.9
SXYL_00727	<i>infA</i>	Translation initiation factor IF-1	5.8	3.2	2.5
SXYL_01186	<i>infC</i>	Translation initiation factor IF-3	6.2	4.1	3.9
Nucleotides transport and metabolism					
SXYL_02522-21	<i>xpt, pbuX</i>	Xanthine phosphoribosyltransferase, Xanthine permease	7.8*	3.7*	3.6*
SXYL_01690	<i>pyrP</i>	Uracil permease	16.2	6.3	4.1
SXYL_02420	<i>hpt</i>	Hypoxanthine phosphoribosyltransferase	6.7	4.0	2.7
SXYL_00017	<i>purA</i>	Adenylosuccinate synthetase	13.2	8.6	5.0
SXYL_00796	<i>pdp</i>	Pyrimidine-nucleoside phosphorylase	2.0	2.8	2.5
SXYL_00821	<i>upp</i>	Uracil phosphoribosyltransferase	9.6	8.2	6.1
SXYL_01689-85	<i>pyrBC, carAB, pyrF</i>	UMP biosynthesis	7.8*	5.0*	2.9*
SXYL_00807	<i>pyrG</i>	CTP synthase	5.7	5.2	4.3
Division					
SXYL_01703	<i>ftsA</i>	Cell division protein FtsA	2.9	2.8	2.6
SXYL_01801	<i>zapA</i>	Cell division protein ZapA	2.7	2.2	2.2
SXYL_01704	<i>divIB</i>	Cell division protein DivIB	4.8	3.5	2.7
SXYL_00848		Cell division protein, FtsW/RodA/SpoVE family	4.1	2.6	2.0
SXYL_01212	<i>mreD</i>	Cell shape-determining protein MreD	3.2	3.0	2.4
SXYL_01651	<i>smc</i>	Chromosome partition protein Smc	3.5	3.6	2.2
SXYL_01699	<i>sepF</i>	Cell division protein sepF	0.3	0.2	0.3
SXYL_01710	<i>mraZ</i>	Protein MraZ	0.3	0.1	0.2
SXYL_00116	<i>sceD2</i>	Probable transglycosylase SceD 2	34.5	31.9	49.3
Peptidoglycan synthesis					
SXYL_00822	<i>mnaA</i>	UDP-N-acetylglucosamine 2-epimerase	8.5	6.6	4.5
SXYL_01425	<i>murG</i>	UDP-N-acetylglucosamine-transferase	2.4	2.9	2.3
SXYL_02467	<i>sle1</i>	N-acetylmuramoyl-L-alanine amidase Sle1	5.7	5.2	6.1
SXYL_01706-05	<i>mraY, murD</i>	Phospho-N-acetylmuramoyl-pentapeptide-transferase, UDP-N-acetylmuramoylalanine-D-glutamate ligase	8.3*	5.0*	3.1*
SXYL_01914	<i>murE</i>	UDP-N-acetylmuramoyl-L-alanyl-D-glutamate-L-lysine ligase	6.2	5.2	3.6
SXYL_02166	<i>uppP</i>	Undecaprenyl-diphosphatase	2.5	2.3	2.2
SXYL_01624	<i>uppS</i>	Isoprenyl transferase	6.1	3.0	2.5
SXYL_01707	<i>pbp1</i>	Penicillin-binding protein 1	4.3	2.7	2.0
SXYL_01304	<i>pbp3</i>	Penicillin-binding protein 3	2.6	3.3	2.7

(Continued)

TABLE 1 | Continued

Gene ID	Gene name	Description	Mean ratio of expression		
			24 h	48 h	72 h
Teichoic acid synthesis					
SXYL_02208	<i>tagD</i>	Glycerol-3-phosphate cytidyltransferase	0.2	0.2	0.3
SXYL_01022		Teichoic acid translocation permease protein	6.4	5.9	4.6
SXYL_01987-90	<i>dltDCBA</i>	Teichoic acid biosynthesis	7.6*	4.7*	2.6*
Membrane proteins					
SXYL_01650	<i>ftsY</i>	Signal recognition particle receptor FtsY	4.2	4.8	3.0
SXYL_00842	<i>yidC</i>	Membrane protein insertase YidC	2.7	2.0	2.1
SXYL_02419	<i>ftsH</i>	ATP-dependent zinc metalloprotease FtsH	11.7	12.0	8.2
Lipid/Phospholipid synthesis					
SXYL_01167-68	<i>accDA</i>	Acetyl-coenzyme A carboxylase carboxyl transferase subunit beta and alpha	3.6*	3.7*	3.0*
SXYL_01326-27	<i>accBC</i>	Acetyl-CoA carboxylase, biotin carboxyl carrier protein and carboxylase subunit	5.6*	3.2*	2.4*
SXYL_01655-54	<i>fabDG</i>	Malonyl CoA-acyl carrier protein transacylase, 3-oxoacyl-[acyl-carrier protein] reductase	8.9*	8.6*	5.1*
SXYL_01944-43	<i>fabHF</i>	3-oxoacyl-[acyl-carrier-protein] synthase 3 and 2	2.7*	2.6*	2.1
SXYL_01921	<i>fabI</i>	Enoyl-[acyl-carrier-protein] reductase [NADPH]	2.5	2.3	
SXYL_01656	<i>plsX</i>	Phosphate acyltransferase	13.3	9.8	6.6
SXYL_01657	<i>fapR</i>	Transcription factor FapR	8.0	5.1	3.2
Regulator/Sensor					
SXYL_01427-28	<i>arlSR</i>	Signal transduction histidine-protein kinase ArlS, Regulator ArlR	2.6*	3.9*	2.8*
SXYL_00951	<i>vraS</i>	Sensor protein VraS	3.8	4.6	2.7
Lysis					
SXYL_00365-67	<i>cidABC</i>	Holin-like proteins CidA and CidB, Pyruvate oxidase	23.4*	10.4*	9.1*
SXYL_00494	<i>lrgA</i>	Antiholin-like protein LrgA		2.6	
SXYL_01730-82		Phage proteins		13.9*	14.1*
SXYL_01785		Phage repressor-like protein	0.3	0.4	0.5
SXYL_01783		Phage antirepressor protein	2.8	24.6	22.2
CARBOHYDRATE METABOLISM					
SXYL_00699	<i>glcU</i>	Glucose uptake protein	10.2	7.8	5.4
Pentose phosphate pathway					
SXYL_00698	<i>gdh</i>	Glucose 1-dehydrogenase	3.0	3.2	2.8
SXYL_00361		Gluconolactonase	3.9	3.4	2.5
SXYL_00438-40	<i>gntRKP</i>	Gluconate Repressor, Kinase, Permease	19.8*	38.6*	16.1*
SXYL_00568	<i>rpi</i>	Ribose-5-phosphate isomerase		2.5	3.0
EMBDEN-meyerhof-parnas pathway					
SXYL_01308	<i>glkA</i>	Glucokinase	2.8	2.9	
SXYL_02069-67	<i>pgk, tpiA, gpml</i>	Phosphoglycerate kinase, Triosephosphate isomerase, 2,3-bisphosphoglycerate-independent phosphoglycerate mutase	6.5*	4.5*	3.1*
Pyruvate metabolism					
SXYL_00250		L-lactate permease		2.5	2.6
SXYL_00170	<i>lqo</i>	L-lactate-quinone oxidoreductase	2.5		
SXYL_00276	<i>ldhB</i>	L-lactate dehydrogenase	0.2	0.3	0.4
SXYL_01023-24	<i>pflAB</i>	Formate acetyltransferase	2.2*	2.3*	2.1*
SXYL_01839-42	<i>pdhABCD</i>	Pyruvate dehydrogenase complex	3.7*	3.4*	2.9*
SXYL_00194	<i>lplA</i>	Lipoate-protein ligase A	6.4	11.3	6.1
SXYL_02616		Acetate-CoA ligase	3.5	4.3	3.6
TCA CYCLE AND RESPIRATORY CHAIN					
SXYL_01793-95	<i>sdhABC</i>	Succinate dehydrogenase	3.0*	4.4*	3.5*

(Continued)

TABLE 1 | Continued

Gene ID	Gene name	Description	Mean ratio of expression		
			24 h	48 h	72 h
SXYL_01636-37	<i>sucCD</i>	Succinyl-CoA synthase		2.5*	2.2*
SXYL_00579	<i>mgo</i>	Malate:quinone oxidoreductase	5.0	6.3	4.7
SXYL_00824-31	<i>atpBEFHAGDC</i>	F0F1-type ATP synthase	8.7*	5.5*	4.0*
SXYL_00823	<i>atpl</i>	Putative ATP synthase protein I	7.3	3.7	2.6
SXYL_00539-42	<i>narGHJ</i>	Respiratory nitrate reductase	4.2*	4.4*	3.1*
SXYL_00534-36	<i>nirBD, sirB</i>	Nitrite reductase	6.3*	3.6*	3.3*
SXYL_00547	<i>narT</i>	Nitrate transporter NarT	2.3		
SXYL_00530		Formate/nitrite transporter	2.9	2.2	2.4
Regulators					
SXYL_00544	<i>nreB</i>	Oxygen sensor histidine kinase NreB	2.1		
SXYL_01366	<i>srrB</i>	Sensor histidine kinase SrrB	2.7	2.1	
SXYL_00883	<i>rex</i>	Redox-sensing transcriptional repressor Rex	0.4	0.4	0.4
COFACTOR, VITAMIN SYNTHESIS					
Molybdenum					
SXYL_00675-77	<i>modABC</i>	Molybdate ABC transporter	5.5*	5.0*	3.7*
SXYL_00682-87	<i>moeA, moeB, moaAE</i>	Molybdenum biosynthesis	3.0*	3.1*	2.9*
SXYL_00679	<i>moeB</i>	Molybdopterin biosynthesis protein MoeB	2.1	1.8	1.7
Heme					
SXYL_01196-99	<i>hemCDBL1</i>	Heme biosynthesis	3.3*	4.0*	3.2*
SXYL_01274	<i>hemN</i>	Oxygen-independent coproporphyrinogen oxidase III	6.6	5.0	3.2
SXYL_00972	<i>hemL2</i>	Glutamate-1-semialdehyde 2,1-aminomutase 2	3.1	3.3	2.5
SXYL_01040-41	<i>hemHG</i>	Heme biosynthesis	4.2*	4.9*	5.6*
SXYL_01819	<i>ctaB</i>	Protoheme IX farnesyltransferase	2.8		2.5
SXYL_00751-53	<i>htsABC</i>	heme transport system	2.9*	2.8*	2.4*
SXYL_00755	<i>isdG</i>	Heme-degrading monooxygenase	0.2	0.2	0.3
SXYL_00749	<i>sfaB</i>	Siderophore biosynthesis protein, lucA/lucC family	0.1	0.2	0.2
SXYL_00944	<i>ftnA</i>	Ferritin	0.2	0.3	0.4
Menaquinone					
SXYL_01079-80	<i>menEC</i>	Menaquinone biosynthesis	2.3	2.8*	2.1
SXYL_01891-92	<i>menHD</i>	Menaquinone biosynthesis	3.2*	5.2*	4.2*
Riboflavin					
SXYL_01097-100	<i>ribDEBAH</i>	Riboflavin biosynthesis protein	0.2*	0.2*	0.3*
Folate					
SXYL_02414-16	<i>folKBP</i>	Folate biosynthesis	5.1*	5.1*	3.4*
SXYL_01203	<i>folC</i>	Tetrahydrofolate synthase	3.6	2.2	
SXYL_01872	<i>folD</i>	Bifunctional protein FolD	7.3	4.6	3.3
SXYL_01261	<i>aroE</i>	Shikimate dehydrogenase	4.4	4.3	2.7
PEPTIDE AND AMINO ACID METABOLISM					
Transport					
SXYL_00300-02		Oligopeptide ABC transporter	2.9*	2.9*	2.3*
SXYL_01184	<i>lysP</i>	Lysine-specific permease	7.1	6.9	4.5
SXYL_01528		Sodium:alanine symporter	5.11		
SXYL_01919		Sodium:alanine symporter	0.45		2.78
SXYL_02518	<i>gltT</i>	Proton/sodium-glutamate symporter	2.3		
Peptidase					
SXYL_01247-48		U32 family peptidases	5.8*	4.5*	3.2*
SXYL_00957	<i>ampS</i>	Leucyl aminopeptidase	3.4	2.7	2.2

(Continued)

TABLE 1 | Continued

Gene ID	Gene name	Description	Mean ratio of expression		
			24 h	48 h	72 h
SXYL_00948	<i>map</i>	Methionine aminopeptidase	3.0	3.0	2.0
Valine, leucine, isoleucine					
SXYL_00867-75	<i>ilvA</i> , <i>leuDCBA</i> , <i>ilvCNBD1</i>	Valine, leucine, isoleucine biosynthesis	0.3*	0.1*	0.1*
SXYL_02469	<i>ilvD2</i>	Dihydroxy-acid dehydratase	0.2	0.1	0.1
SXYL_01337-40	<i>lpdA</i> , <i>bkdA1A2</i>	alpha-keto acid dehydrogenase complex	4.9*	3.7*	2.8*
Tryptophan, phenylalanine, tyrosine					
SXYL_01383-85	<i>aroABC</i>	Aromatic acid biosynthesis	0.3*	0.3*	0.2*
SXYL_02022	<i>aroD</i>	3-dehydroquinate dehydratase	0.2	0.3	0.3
SXYL_01497	<i>trpA</i>	Tryptophan synthase alpha chain	0.4	0.4	0.5
SXYL_01128		DAHPh synthetase-chorismate mutase	0.1	0.0	0.1
Histidine					
SXYL_00460-68	<i>hisZGDCBHAFI</i> Cysteine, methionine	Histidine biosynthesis	0.1*	0.1*	0.1*
SXYL_02644	<i>metE</i>	5-methyltetrahydropteroyltriglutamate-homocysteine methyltransferase	0.1	0.1	0.2
SXYL_00372		D-cysteine desulfhydrase	0.3	0.4	0.4
Arginine					
SXYL_00238-41	<i>rocD1, argCJB</i>	Arginine biosynthesis	0.3*	0.2*	0.3*
SXYL_00252	<i>arcB</i>	Ornithine carbamoyltransferase	0.3	0.3	0.4
SXYL_01961-62	<i>argGH</i>	Arginine biosynthesis	0.3*	0.2*	0.2*
SXYL_01355	<i>proC</i>	Pyrroline-5-carboxylate reductase	0.3	0.3	0.3
Aspartate					
SXYL_01002	<i>panD</i>	Aspartate 1-decarboxylase	0.4	0.2	0.2
SXYL_01558		Aspartokinase	0.4	0.3	0.4
SXYL_01373	<i>asnA</i>	L-asparaginase	2.6	2.8	2.4
Glutamate, glutamine					
SXYL_01964	<i>gluD1</i>	Glutamate dehydrogenase		2.3	
SXYL_02326	<i>gluD2</i>	Glutamate dehydrogenase	3.5		
SXYL_02459-61	<i>gltBCD</i>	Glutamate synthase	2.6*		
SXYL_00105		Membrane protein, EutH superfamily	20.3	29.3	12.9
SXYL_00106		Short-chain dehydrogenase	41.3	29.4	13.9
SXYL_00107	<i>glnA2</i>	Glutamine synthetase	41.7	29.2	15.7
SXYL_00108		Aldehyde dehydrogenase	41.3	29.5	15.4
RESPONSE TO OSMOTIC STRESS					
SXYL_01536	<i>mscL</i>	Large-conductance mechanosensitive channel	0.3	0.2	0.3
Synthesis and accumulation of osmoprotectant					
SXYL_00488-91	<i>opuCABCD</i>	Glycine betaine/carnitine/choline ABC transporter	24.8*	6.1*	4.3*
SXYL_00486	<i>lcdH</i>	L-carnitine dehydrogenase	22.8	5.0	3.3
SXYL_00223-26	<i>cutTCA</i> , <i>betA</i>	Glycine betaine/carnitine/choline ABC transporter	7.9*	3.7*	3.4*
SXYL_01171	<i>aapA</i>	D-serine/D-alanine/glycine transporter	2.6	2.8	2.3
SXYL_00317		D-serine/D-alanine/glycine transporter	4.9	6.1	4.4
SXYL_02528-29	<i>sdaAA1AB1</i>	L-serine dehydratase, alpha subunit and beta subunit	3.8*	5.0*	4.0*
SXYL_00820	<i>glyA</i>	Serine hydroxymethyltransferase	10.7	7.4	5.4
SXYL_01084	<i>metK</i>	S-adenosylmethionine synthetase	6.8	8.3	4.8
SXYL_02526		Sodium:dicarboxylate symporter	11.2	3.7	
Na⁺/H⁺ antiporter					
SXYL_01970-76	<i>mnhA1B1C1D1E1F1G1</i>	Na(+)/H(+) antiporter	6.1*	6.3*	3.4*

(Continued)

TABLE 1 | Continued

Gene ID	Gene name	Description	Mean ratio of expression		
			24 h	48 h	72 h
SXYL_02220-26	<i>mnhG2F2E2D2C2B2A2Na(+)/H(+)</i>	antiporter	4.8*	4.6*	2.8*
Regulator					
SXYL_00859	<i>rsbU</i>	Serine phosphatase RsbU, regulator of sigma subunit	4.2	2.9	2.8
SXYL_00860	<i>rsbV</i>	Anti-sigma-B factor antagonist	0.3	0.2	0.2
SXYL_00861	<i>rsbW</i>	Serine-protein kinase RsbW	0.2	0.2	0.2
STRESS RESPONSE AND PIGMENTATION					
SXYL_01551	<i>katB</i>	Catalase B	0.2	0.3	0.4
SXYL_02533	<i>katC</i>	Catalase C	0.2	0.2	0.2
SXYL_01572	<i>bsaA</i>	Glutathione peroxidase	0.2	0.4	0.4
SXYL_00374		Thioredoxin	0.1	0.2	0.2
SXYL_00051-54	<i>crtPQMN</i>	pigment biosynthesis	7.8*	6.3*	3.9*
SXYL_00358	<i>mvaS</i>	3-hydroxy-3-methylglutaryl CoA synthase	2.2		
SXYL_00359-60	<i>mvaCA</i>	Acetyl-CoA acetyltransferase, Hydroxymethylglutaryl-CoA reductase	2.4*	2.2*	
SXYL_02258-56	<i>mvaK1DK2</i>	Mevalonate kinase, Diphosphomevalonate decarboxylase, Phosphomevalonate kinase	5.0*	4.4*	2.9*
SXYL_00599	<i>fni</i>	Isopentenyl-diphosphate delta-isomerase	2.2	3.2	2.5
SXYL_01332	<i>ispA</i>	Geranyltranstransferase	4.6	3.8	2.5
SXYL_00309		Universal stress protein	0.1	0.2	0.2
SXYL_01162		Universal stress protein	0.2	0.2	0.2
SXYL_00196		General stress protein	0.0	0.0	0.0

*Means of the expression of the clustered genes differentially expressed.

S. aureus (Margolin, 2000). The gene *smc*, which plays a role in chromosome partitioning was up-regulated and two genes (*sepF*, *mraZ*) encoding proteins involved in septation and separation of one cell into two daughter cells were down-regulated. Finally, the gene *sceD2* encoding a transglycosylase potentially able to cleave peptidoglycan and affect separation of bacterial cells was highly up-regulated.

Concomitantly, 8 genes (*mnaA*, *murG*, *sle1*, *mraY*, *murD*, *murE*, *uppP*, *uppS*) involved in peptidoglycan synthesis and two genes (*pbp1*, *pbp3*) encoding penicillin-binding proteins involved in its translocation across the membrane were up-regulated (Table 1). The *arlSR* genes, which encode the two-component regulatory system ArlSR, were up-regulated. An *arlS* mutant in *S. aureus* exhibited altered activity of peptidoglycan hydrolases, which participate in important processes occurring during cell growth and division, such as cell wall synthesis, peptidoglycan turnover and recycling (Fournier and Hooper, 2000). Also, *vraS* encoding the sensor protein of the two-component system VraSR was overexpressed in *S. xylosum*. In *S. aureus*, VraSR positively modulates the regulation of the cell wall biosynthesis pathway (Kuroda et al., 2003). The *tagD* gene encoding the synthesis of teichoic acids was down-regulated in *S. xylosum*, while the gene SXYL_01022 and the *dlt* operon encoding, respectively, a permease protein involved in their translocation across the membrane and proteins involved in their alanylation were up-regulated.

Sixteen genes encoding membranous proteins were modulated, with 12 of them up-regulated (Table 1,

Supplementary Table 2). Among these, we identified *ftsY*, which encodes FtsY involved in targeting and insertion of nascent membrane proteins into the cytoplasmic membrane, and *yidC*, which encodes YidC, which functions as a membrane protein insertase independent of the Sec protein-conducting channel. YidC can also assist in the lateral integration and folding of membrane proteins that insert into the membrane via the Sec pathway (Samuelson et al., 2000). The *ftsH* gene was involved in the quality control of integral membrane proteins.

In parallel, 13 genes involved in membrane lipid synthesis were up-regulated (Table 1). The mechanism of fatty acid synthesis is conserved and proceeds in two stages: initiation and elongation (Schujman and de Mendoza, 2008). The *accDA* genes encode the two subunits of carboxyltransferase, while the *accBC* genes encode the biotin acetyl CoA carboxylase, all of which are involved in the initiation step. In *B. subtilis*, the transcription of the operon *accBC* is under growth rate control, the rate of transcription decreasing with decreased growth (Marini et al., 2001). The genes *fabDG*, *fabHF*, *fabI* are involved in the elongation step, while *plsX* is involved in phospholipid synthesis. PlsX in *B. subtilis* plays an important role in the coordination of production of fatty acids and phospholipids (Schujman et al., 2003). FapR is a global transcriptional repressor that controls the expression of the *fap* regulon including all the *fab* genes and the *plsX* one (Schujman et al., 2003). The binding of FapR to its DNA targets is specifically inhibited by malonyl-CoA, a cellular pool that provides a mechanism for sensing the status of fatty acid synthesis. In our conditions, up-regulation of

acc genes could lead to malonyl-CoA that can control the activity of FapR encoded by *fabR* up-regulated.

Fifty-one genes involved in cell lysis were modulated (Table 1, Supplementary Table 2). The *cidABC* cluster was overexpressed. In *S. aureus*, Cid and Lrg proteins are involved in the control of cell lysis (Ranjit et al., 2011). The *S. aureus cid* operon expresses two overlapping transcripts: a full-length *cidABC* transcript that is expressed at low levels during exponential growth and a *cidBC* transcript also expressed during exponential growth but at high levels (Rice et al., 2004). In *S. xylosus*, the *cidA* gene was highly overexpressed by comparison with the *cidB* and *cidC* genes (Supplementary Table 2). In *S. aureus*, *cidA* encodes a holin-like protein with a positive effect on murein hydrolase activity and *lrgA* encodes an antiholin-like protein with an inhibitory effect on these enzymes (Ranjit et al., 2011). In *S. aureus*, the *cidA* mutant displayed decreased lysis during biofilm formation while the *lrgAB* mutant was shown to increase lysis (Mann et al., 2009). In our conditions, the very high expression of *cidA* compared with expression of *lrgA* led us to suppose that cell lysis occurred during the survival of *S. xylosus* in the meat model.

Furthermore, a cluster of 46 genes (SXYL_01730-88) encoding phage proteins was modulated with 44 highly up-regulated at 48 and 72 h of incubation (Table 1, Supplementary Table 2). The gene SXYL_01785 encoding a phage repressor-like protein was down-regulated, while the gene SXYL_01783 coding a phage antirepressor protein was highly up-regulated. In *Salmonella* phage P22, the antirepressor overcomes the repressor protein involved in the maintenance of lysogeny (Levine et al., 1975). Consequently, in our meat model, particularly after 48 and 72 h of incubation, 44 genes involved in phage multiplication reflecting a lytic phase were highly up-regulated. Moreover, we have shown that a lytic phage can be induced by mitomycin C in *S. xylosus* C2a (data not shown).

Taken together these results (many up-regulated genes involved in DNA machinery, cellular process, cell lysis and the *S. xylosus* population remaining stable) suggested that a balance between cell division and cell lysis could be observed in our meat model up to 72 h.

Glucose and Lactate Catabolized Simultaneously

The concentrations of glucose added to the meat model and of lactate originating from the meat were close at T0 and decreased similarly during incubation to undetectable levels at 72 h (Table 2). Concomitantly, an increase of acetate was noted in the meat model throughout incubation associated with a small

pH decrease of 0.27 after 48 h of incubation. *S. xylosus* produced mainly acetate as end-product and simultaneously metabolized glucose and lactate in the meat model. It has been shown that *S. aureus* is able to grow using glucose and lactate simultaneously to form acetate under aerobiosis (Ferreira et al., 2013).

Our transcriptomic data revealed that several genes involved in carbohydrate metabolism were overexpressed (Table 1, Figure 1). Glucose could be transported by the glucose uptake protein (GlcU). Then it could be metabolized via the glucose 1-dehydrogenase (Gdh). In *S. xylosus*, the genes *glcU* and *gdh* are co-localized and co-expressed, suggesting that GlcU recruited glucose for glucose dehydrogenase (Fiegler et al., 1999). GlcU serves in addition to PTS in taking up glucose during *S. xylosus* growth in laboratory medium (Fiegler et al., 1999). The Gdh and gluconolactonase activities resulted in formation of gluconate and consequently there was overexpression of the cluster *gntRKP*, encoding the repressor protein, gluconate kinase and gluconate permease, respectively, and involved in gluconate utilization via the pentose phosphate (PP) pathway (Table 1). For *Bacillus subtilis*, expression of *gnt* is derepressed in the presence of gluconate (Fujita and Fujita, 1987; Yoshida et al., 1995). In our conditions, gluconate 6-P could be further metabolized via the PP pathway and entered the Embden-Meyerhof-Parnas (EMP) pathway via D-glyceraldehyde 3-P (Table 1, Figure 1). In parallel, the gene *glkA* encoding glucokinase could fuel the EMP pathway with four genes up-regulated (Table 1). Deviation of part of the catabolism of glucose to the PP pathway could be an advantage as it will generate two NADH contributing to the redox status.

Finally, lactate used as carbohydrate substrate could be imported via the L-lactate permease encoded by the gene SXYL_00250, which is overexpressed in the meat model, and catabolized to pyruvate by the lactate-quinone oxidoreductase encoded by *lqo*, which was also overexpressed (Table 1, Figure 1). *S. aureus* consumes lactate by a lactate-quinone oxidoreductase that is critical to respiratory growth on L-lactate (Fuller et al., 2011). Interestingly, *ldhB* encoding lactate dehydrogenase was down-regulated (Table 1, Figure 1). Pyruvate could be catabolized to acetyl-CoA by both formate acetyltransferase and the pyruvate dehydrogenase complex (PDH) (Table 1). Moreover the *lplA* gene encoding a lipote-protein ligase A responsible for lipoylation in the presence of exogenous lipoic acid, a factor required for PDH, was also up-regulated (Table 1). Acetyl CoA could therefore be catabolized to acetate by acetate-CoA ligase encoded by SXYL_02616 and excreted in the meat model as early as the first 24 h of incubation (Tables 1, 2).

TABLE 2 | Glucose and lactate consumption and acetate production in a meat model over time.

Sampling (h)	Glucose/100 g		Lactate/100 g		Acetate/100 g		pH
	mg	mmole	mg	mmole	mg	mmole	
0	633	35	763	85	0	0	5.82
24	521	29	510	57	72	12	5.65
48	248	14	256	28	182	30	5.55
72	0	0	0	0	262	44	5.68

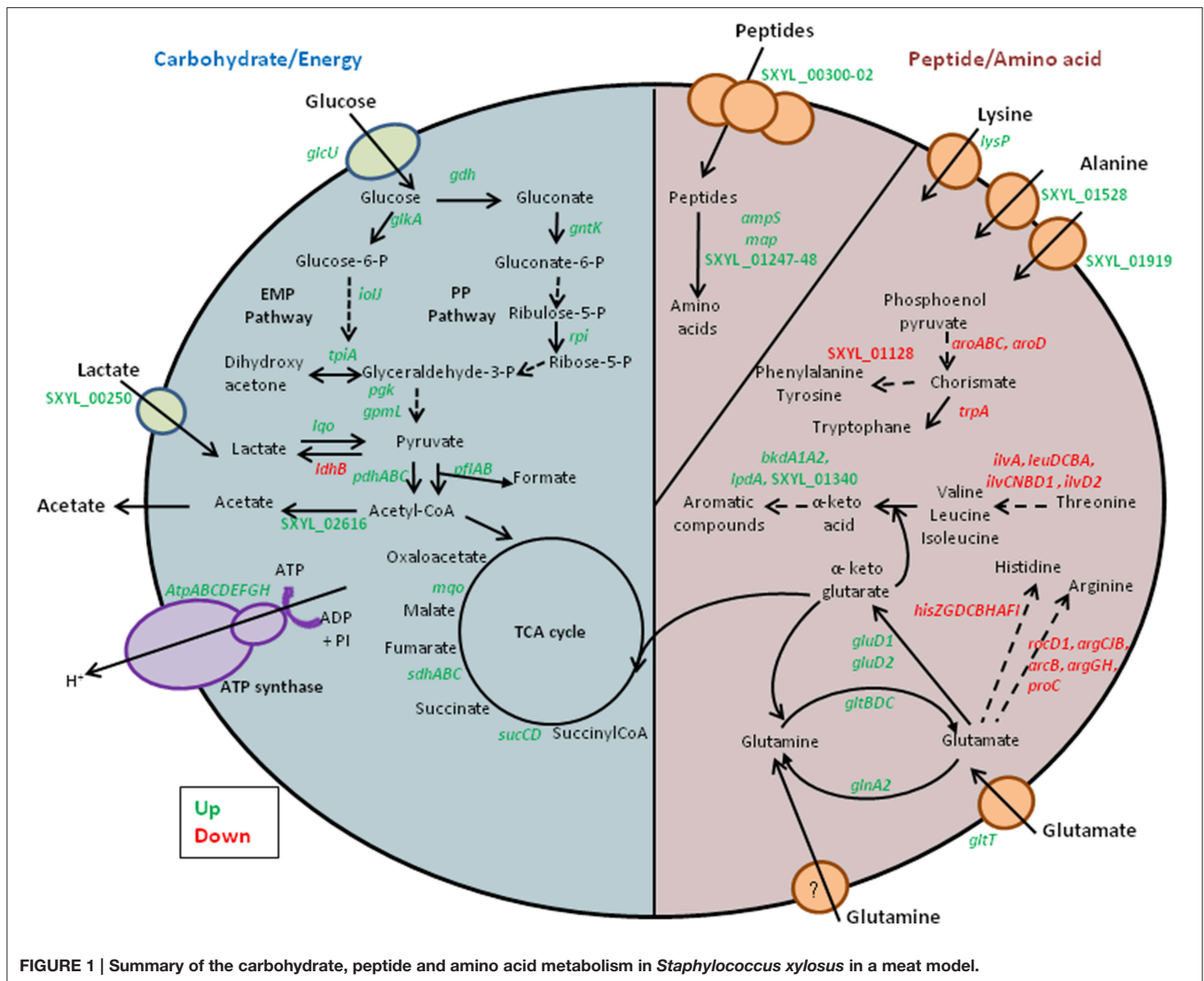


FIGURE 1 | Summary of the carbohydrate, peptide and amino acid metabolism in *Staphylococcus xylosus* in a meat model.

In our conditions, 100% of the two carbon sources (glucose and lactate) were consumed toward EMP and PP (Table 2, Figure 1). Their catabolism generated acetyl-CoA with 55% (up to 170 mmoles) arising from lactate and 45% (up to 140 mmoles) from glucose. Acetyl-CoA can be then catabolized in acetate or feed the TCA cycle. Only small amounts of acetate were formed (Table 2). So we can hypothesize that the acetyl-CoA to acetate conversion rate did not exceed 14%.

TCA Cycle and Respiratory Chain

Acetyl-CoA is catabolized through the tricarboxylic acid (TCA) cycle, which is a central pathway for the catabolism of carbohydrate, fatty and amino acids, which enter the cycle at several points. Some TCA cycle genes were up-regulated in the meat model, such as the succinate dehydrogenase (*sdhABC*), succinyl-CoA synthase (*sucCD*), and malate:quinone oxidoreductase (*mgo*) genes (Table 1). For *S. aureus* in biofilm, it was found that the up-regulation of succinate dehydrogenase genes is an advantage under oxygen-limited conditions (Gaupp

et al., 2010). The succinate dehydrogenase is involved in the electron transfer of the respiratory chain driving ATP synthesis. We observed that 8 genes encoding the F₀F₁-type ATP synthase (*atpBEFHAGDC*) and the gene *atpI*, upstream of the *atp* genes and encoding a putative ATP synthase protein I, were highly overexpressed and could furnish energy for the survival of *S. xylosus* in meat (Table 1, Figure 1).

The overexpression of several genes involved in the EMP and PP pathways, of the clusters *nar* and *nir* related to nitrate respiration, although nitrate was not present, of the nitrate and formate/nitrite transporter genes and of *pfl* encoding formate acetyl transferase, suggested anaerobic conditions (Table 1), as already mentioned for *S. aureus* grown *in vitro* in the absence of oxygen (Fuchs et al., 2007). The two-component regulatory system NreBC, which is considered as an oxygen-sensing system, stimulates the expression of genes of nitrate respiration under anaerobic conditions in *S. carnosus* and *S. aureus* (Schlag et al., 2008; Reinhart et al., 2010). The targets are the genes encoding the nitrate and nitrite reductases and the nitrate transporter.

In our conditions, the regulatory gene *nreB* was up-regulated at 24 h of growth, thus explaining the up-regulation of *nar*, *nir*, and *narT* (Table 1). Moreover, *ssrB* encoding a sensor that autophosphorylates in the absence of O₂ was up-regulated. This two-component system SrrAB is induced under oxygen-limiting conditions in staphylococci (Green et al., 2014). In *S. aureus*, a redox-sensing transcriptional repressor Rex is involved in the regulation of anaerobic gene expression (Pagels et al., 2010). The binding activity of Rex is enhanced by NAD⁺ while NADH decreases it. Rex regulates the expression of pathways that lead to anaerobic NAD⁺ regeneration, nitrate respiration and ATP synthesis. In our study, the transcription of *rex* was down-regulated and could explain the overexpression of genes involved in EMP and PP pathways, nitrate respiration and ATP synthesis.

Cofactor, Vitamin Synthesis

In the meat model, *S. xylosum* overexpressed the cluster *modABC* encoding the molybdate ABC-type transporter and six genes, including *moeB*, of the cluster involved in the synthesis of the molybdenum cofactor (Table 1). In *S. carnosus*, anaerobic growth conditions enhanced transcription of *moeB* (Neubauer et al., 1998). Moreover, *S. carnosus* mutants defective either in *modABC* or in *moeB* were defective in nitrate reductase activity, the nitrate reductase being a molybdoenzyme (Neubauer et al., 1999). As we mentioned above, *S. xylosum* in the meat model also up-regulated the cluster *nar* encoding the nitrate reductase.

The cluster *hemCDBL1* and five other genes (*hemN*, *hemL2*, *hemHG*, *ctaB*) encoding proteins involved in the synthesis of heme were up-regulated (Table 1). Heme is vital to many biological systems and a multitude of redox enzymes engage heme as a catalyst of electron transfer. *S. aureus* can employ heme as a cofactor required for respiration, both by aerobic electron transport and by the anaerobic nitrate reductase complex (von Eiff et al., 2006). Moreover, the cluster *htsABC* encoding a heme transport system was up-regulated in *S. xylosum* (Table 1). In *S. aureus*, inactivation of HtsABC reduced its ability to import heme iron (Skaar et al., 2004). Heme can be an iron source, but in the meat model *S. xylosum* down-regulated the gene *isdG* encoding a heme-degrading monooxygenase that promotes catalytic degradation of heme with subsequent release of iron. Furthermore, there was down-regulation of *sfaB* belonging to the cluster *sfa* involved in synthesis of the siderophore staphyloferrin A and of *ftnA* involved in iron storage (Table 1). Thus, in the meat model, i.e., in iron-replete conditions, the synthesis of heme is more probably intended for proteins involved in respiration, such as succinate dehydrogenase encoded by *sdh* (Table 1). The genes *menEC* and *menHD* involved in synthesis of menaquinone from chorismate were overexpressed (Table 1). Menaquinone is an important cofactor that is exploited in electron transport pathways. Menaquinone biosynthesis in facultative anaerobes is increased by anaerobiosis and in *S. aureus* menaquinone deficiency is accompanied by impaired nitrate respiration (Bentley and Meganathan, 1982). During respiration, menaquinone donate electrons to the heme molecules located within cytochromes (Wakeman et al., 2012).

The cluster *ribDEBAH* involved in riboflavin synthesis was down-regulated in our conditions (Table 1). The active forms

of riboflavin in the cell are flavin mononucleotide (FMN) and flavin adenine dinucleotide (FAD), but riboflavin can be found as free compounds (Perkins and Pero, 2002). In contrast, the cluster *folKBP* and 3 other genes involved in the synthesis of folate were overexpressed (Table 1). Cells require folate cofactors as acceptor/donor of one carbon unit in numerous processes such as methionine, purine, and thymine synthesis and in some degradative reactions (Rossi et al., 2011).

Peptide and Amino Acid Metabolism

Proteins are the main components of meat. They are classified as water-soluble (sarcolemmal, 30%), soluble in high salt concentration (myofibrillar, 55%) and insoluble (stromal proteins, proteins from connective tissue) (Lafarga and Hayes, 2014). The sarcolemmal and myofibrillar proteins are hydrolyzed during fermentation and ripening of fermented sausages (Hughes et al., 2002). The initial degradation is essentially due to the activity of endogenous proteinases that release peptides that can be further hydrolyzed by bacterial enzymes (Hughes et al., 2002). These peptides can be transported into *S. xylosum* by the oligopeptide transport system encoded by the three genes, SXYL_00300-302 (Table 1, Figure 1). Peptides played a key role in bacterial nutrition; their transport is generally regulated at the transcriptional level by intracellular pools of amino acids and by environmental changes such as anaerobic conditions (Hiron et al., 2007). Two genes (SXYL_01247-48) encoding peptidases from U32 family and the genes *ampS* and *map* encoding leucyl- and methionine- aminopeptidases, respectively, were overexpressed. These peptidases will furnish amino acids for *S. xylosum* (Table 1). The starter cultures (*Pediococcus pentosaceus* plus *S. xylosum* or *Lactobacillus sakei* plus *S. carnosus*) influenced the proteolytic activity and the release of amino acids in sausages (Hughes et al., 2002; Aro Aro et al., 2010). Moreover, *L. sakei* overexpressed two dipeptidases in the presence of sarcolemmal proteins (Fadda et al., 2010).

The meat model contained variable levels of free amino acids with a broad pattern of 19 amino acids (Table 3). The total initial concentration (157 mg/100 g) increased up to 24 h and could be mainly attributed to endogenous meat peptidases. After 24 h there was a decrease of the total free amino acid content. Notably, the level of lysine decreased after 24 h and *lysP* encoding a lysine-specific permease was overexpressed in *S. xylosum* (Tables 1, 3). Likewise, the concentration of alanine decreased sharply in the meat model after 48 h of incubation and two genes encoding sodium:alanine symporters were modulated during the incubation (Tables 1, 3). In parallel, in the meat model, *S. xylosum* down-regulated 26 genes involved in the synthesis of branched-chain amino acids (nine genes belonging to the cluster *ilv-leu* and the gene *ilvD2*), aromatic amino acids (*aroABC*, *aroD*, *trpA*, SXYL_01128), histidine (*hisZGDCBHAFI*) and cysteine-methionine (*metE*, SXYL_00372) (Table 1, Figure 1). In *S. aureus*, CodY contributes to the regulation of more than 200 genes including most of the ones listed above (Majerczyk et al., 2010). CodY repression occurs in *Bacillus* cells growing in a medium containing glucose and amino acids (Shivers and Sonenshein, 2004). Furthermore, *S. xylosum* down-regulated 8 genes involved in synthesis of

TABLE 3 | Concentration of free amino acids in a meat model over time.

Amino Acid	0 h mg/100 g	24 h mg/100 g	48 h mg/100 g	72 h mg/100 g
VAL	7.4	8.8	8.2	5.0
LEU	8.1	10.5	9.7	5.0
ILE	3.8	3.3	2.9	1.0
THR	8.0	7.9	5.7	1.4
TYR	3.8	6.1	6.2	7.3
PHE	5.0	7.6	7.9	12.4
HIS	0.0	0.0	6.9	7.6
ASP	3.6	3.7	3.1	2.9
ASN	2.9	4.3	4.3	2.1
GLU	16.0	15.7	14.8	17.5
GLN	20.5	16.9	12.0	2.1
SER	5.4	5.4	3.5	0.8
GLY	12.8	8.7	3.6	0.0
ALA	27.6	34.5	27.3	4.0
ARG	6.7	11.3	11.4	6.1
CYS	1.4	1.1	0.8	0.6
MET	3.3	4.9	5.3	6.4
LYS	16.8	19.1	11.8	7.6
PRO	3.9	5.3	6.8	5.2
Total	157.0	175.1	152.2	95.0

arginine (*rocD1-argCJB*, *arcB*, *argGH*, *proC*) (Table 1). In *S. aureus*, arginine biosynthesis is regulated through the global regulator CcpA and it was shown that a mutant *ccpA* facilitates the synthesis of arginine via the urea cycle (Nuxoll et al., 2012).

The genes encoding the synthesis of branched-chain amino acids were down-regulated, whereas four genes *lpdA*, *bkdA1A2*, *SXYL_01340* organized in a cluster and involved in the catabolism of branched-chain amino acids were up-regulated (Table 1, Figure 1). The two genes (*bkdA1A2*) encode the subunits of branched-chain alpha-keto acid dehydrogenase E1 involved in the production of 3-methyl butanoyl-CoA, the precursor of 3-methyl butanoic acid (Table 1). The concentrations of leucine, isoleucine and valine decreased in the meat model after 48 h of incubation (Table 3). The pathway leading to the synthesis of 3-methyl butanoic acid, 3-methyl butanal and 3-methyl butanol from the catabolism of leucine has been characterized in *S. xylosus* (Beck et al., 2004). These aroma compounds and more generally methyl aldehydes, methyl acids and methyl alcohols contribute to the flavor of fermented sausages (Berdagué et al., 1993; Stahnke, 1995; Søndergaard and Stahnke, 2002).

Three genes involved in the aspartate metabolism were modulated in *S. xylosus* grown in the meat model, with two genes, *panD* and *SXYL_01558*, down-regulated, while *asnA* encoding L-asparaginase was up-regulated (Table 1). The gene *asnA* coding L-asparaginase in *L. sakei* was induced during the growth of this bacterium in a meat model and the corresponding mutant Δ *asnA2* showed reduced growth in this model, suggesting that asparagine could be a source of nitrogen (Hüfner et al., 2007).

Glutamate and glutamine were found in high concentrations in the meat model (Table 3). In contrast to the concentration of glutamate, which remained stable throughout incubation, the concentration of glutamine decreased sharply in the meat model (Table 3). Glutamate could be imported by the glutamate symporter protein encoded by *gltT* and catabolized by the glutamate dehydrogenases encoded by *gluD1* and *gluD2* (Table 1, Figure 1). Glutamate dehydrogenase activity provides α -ketoglutarate, which is required for amino acid transamination, which initiates the conversion of amino acids to aromatic compounds. Glutamate could be synthesized by the glutamate synthase from α -ketoglutarate, an intermediate of TCA cycle, and glutamine. This enzyme is encoded by the *gltBCD* cluster, which was overexpressed at 24 h of incubation (Table 1, Figure 1). Glutamate serves as the major amino group donor for all nitrogen-containing compounds, as a link between nitrogen and carbon metabolism. The glutamate synthesizing and degrading reactions must be tightly controlled to maintain its homeostasis (Gunka and Commichau, 2012). A cluster of four genes (*SXYL_00105-108*) was highly overexpressed at the three times of incubation (Table 1). This cluster is uncharacterized, but one of these genes potentially encodes a glutamine synthetase (*glnA2*). In *S. xylosus* C2a, another gene encodes a glutamine synthetase, *glnA1* located in a cluster with *glnR*. In *B. subtilis*, the *glnRA* operon is expressed in the absence of glutamine (Schreier et al., 1989). In the meat model, *glnR* and *glnA1* were not differentially expressed (Supplementary Table 2). Similarly, two genes encoding glutamine synthetases were identified in *Halobacillus halophilus*. In this bacterium, the expression of *glnA2*, but not *glnA1*, was increased in the presence of NaCl (Saum et al., 2006). The up-regulation of *glnA2* and the other genes of the cluster in *S. xylosus* could be linked to the osmotic stress generated by the presence of salt in the meat model.

Response to Osmotic Stress Generated by Added NaCl

S. xylosus has to adapt to the high concentration of sodium chloride (0.47 M) in the meat model compared with 0.017 M in the medium used for the preparation of the inoculum. A primary response of *S. xylosus* to the presence of NaCl was the down-regulation of *mscL*, encoding a large conductance mechanosensitive channel, to prevent water efflux and maintain the physical integrity of the bacterial cells (Table 1, Figure 2).

In addition, genes encoding enzymes involved in three different pathways for the synthesis of glycine betaine, a powerful osmoprotectant, were up-regulated (Table 1, Figure 2). Among them, the cluster *opuC* (*opuCABCD*) and *lcdH* encoding L-carnitine dehydrogenase were identified. The cluster *opuC* encodes a glycine betaine/carnitine/choline ABC transporter identified as the only uptake route for L-carnitine in *B. subtilis* and *Listeria monocytogenes* (Kappes and Bremer, 1998; Angelidis and Smith, 2003). L-carnitine is found in raw meats with levels varying from 6.5 to 87.7 mg/100 g (Demarquoy et al., 2004). In the meat model, L-carnitine could be imported via the OpuC and catabolized to glycine betaine by L-carnitine dehydrogenase. The cluster of four genes (*cudTCA*, *betA*) encoding a choline transporter (*cudT*) and

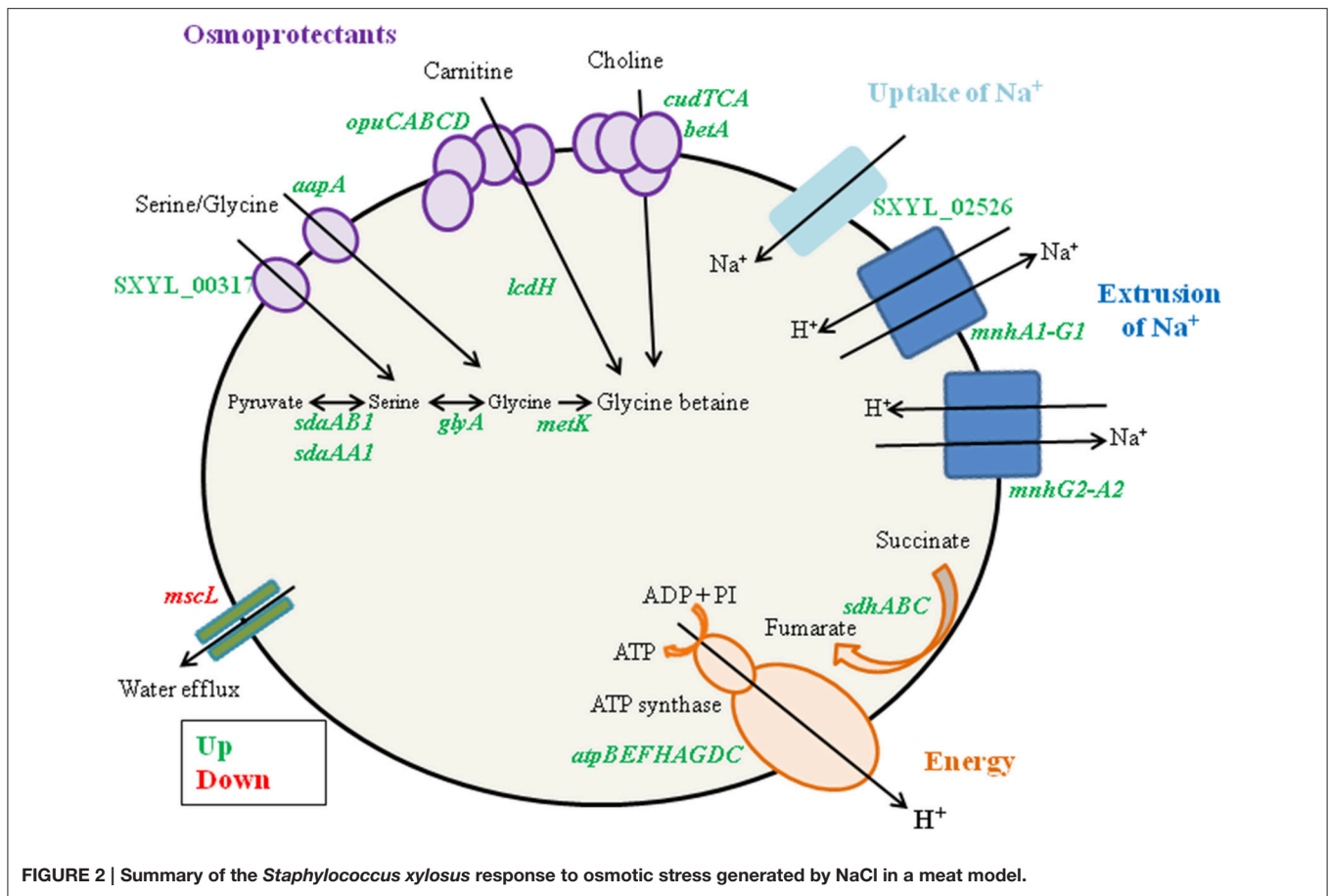


FIGURE 2 | Summary of the *Staphylococcus xylosus* response to osmotic stress generated by NaCl in a meat model.

enzymes for dehydrogenation (*cudA*, *betA*) to form glycine betaine from choline were also up-regulated. These genes were transcriptionally regulated when *S. xylosus* C2a was cultured *in vitro* at high NaCl concentration (Rosenstein et al., 1999). Moreover, the genes *aapA* and SXYL_00317, which encode D-serine D-alanine glycine transporters, three genes (*sdaAA1*, *sdaAB1*, *glyA*) encoding enzymes involved in the synthesis of glycine, and *metK*, which encodes S-adenosyl-methionine, which serves as methyl donor for synthesis of glycine betaine, were up-regulated (Table 1, Figure 2). Concomitantly, we observed a sharp decrease of the concentrations of glycine and serine during the incubation in the meat model (Table 3).

The presence of NaCl induced the gene SXYL_02526 encoding a symporter of Na⁺ and the two clusters *mnh* (*mnhA1-G1*, *mnhG2-A2*) encoding Na⁺/H⁺ antiporter systems (Table 1, Figure 2). The modulation of *mnh2* expression was previously noted in *S. xylosus* grown in a meat model supplemented with nitroso compounds, the presence of nitrate and nitrite leading to the down-regulation of *mnh2* (Vermassen et al., 2014). In *S. aureus*, an active antiporter Mnh resulted in increased transcription of genes encoding respiratory chain components, such as the succinate dehydrogenase locus (*sdh*) (Swartz et al., 2007). The same observation was made in our meat model (Table 1). Moreover, all the genes involved in the synthesis of the multisubunit F₀F₁-ATPase were overexpressed by *S. xylosus*,

as already mentioned (Table 1, Figure 2). This ATPase links the production of ATP to the transmembrane proton motive force (PMF) and can either generate ATP at the expense of the PMF or generate a PMF using ATP produced by fermentative substrate level phosphorylation (Cotter and Hill, 2003). The PMF can expel the protons, resulting from the activity of the two *S. xylosus* Mnh antiporters, from the cytoplasm to maintain pH homeostasis. In *Bacillus pseudofirmus*, proton pumping by the F₀F₁-ATPase generated a proton motive force across the membrane that powered Mnh antiporter activity after addition of Na⁺ (Morino et al., 2014).

Sigma factors play a role in the response of bacteria to environmental stress conditions. The expression of *rsbU* was up-regulated while *rsbV* and *rsbW* of the sigma B cluster were down-regulated (Table 1). In *S. epidermidis*, osmolarity increased the synthesis of polysaccharide intercellular adhesion (PIA) and biofilm formation. The induction of PIA synthesis in the presence of NaCl depends on a functional *rsbU* gene (Knobloch et al., 2001). Our results with the up-regulation of only *rbsU* in *S. xylosus* in the presence of salt could suggest a regulation independent of sigma B.

Response to Other Stresses

S. xylosus overexpressed the cluster *dlt* involved in D-alanylation of lipoteichoic and wall teichoic acids (Table 1). The degree

of D-alanylation varies depending on environmental conditions such as modification of pH or salt concentration (Neuhaus and Baddiley, 2003). Esterification in *S. aureus* was shown to increase with a decrease of pH (Neuhaus and Baddiley, 2003). Inactivation of *dltC* in *Streptococcus mutans* resulted in the generation of an acid-sensitive strain that could not grow below pH 6.5 (Cotter and Hill, 2003). In the meat model, *S. xylosus* has to adapt to acidification as the inoculum was grown at pH 7.0 before its inoculation in the meat model at pH 5.9 with a slight decrease during the incubation (Table 2). Thus, the up-regulation of *dlt* could be the result of adaptation to the acidic environment of meat. It is noteworthy that in the conditions of the meat model, the high concentration of NaCl (0.47 M) did not repress the expression of *dlt*, as reported for *S. aureus in vitro* conditions in the presence of 0.325 M NaCl (Koprivnjak et al., 2006).

Growth in the meat model did not seem to generate oxidative stress, as the genes *katC*, *katB*, *bsaA* and *SXYL_00374* encoding, respectively, catalases, glutathione peroxidase and thioredoxin involved in the response to this stress, were down-regulated (Table 1). Whereas in our former study, nitrate and nitrite added to the meat model generated nitrosative stress that induced the up-regulation of genes involved in antioxidant defenses (Vermassen et al., 2014).

S. xylosus overexpressed the *crtPQMN* genes involved in the carotenoid biosynthesis pathway (Table 1). Moreover, eight genes (*mvaS*, *mvaCA*, *mvaK1DK2*, *fni*, *ispA*) encoding proteins of mevalonate pathway involved in the synthesis of farnesyl diphosphate, a precursor of the carotenoid synthesis, were up-regulated (Table 1). The C2a strain produces a yellow pigment after cultivation on agar medium, as do about 50% of *S. xylosus* strains (our unpublished data). *S. aureus* produced the intermediary yellow pigment 4,4' diaponeurosporene, which is then converted to the yellow-orange end-product, staphyloxanthin, after prolonged cultivation (Wieland et al., 1994). In *S. aureus*, the CrtM and CrtN enzymes are responsible for the synthesis of the yellow pigment from farnesyl diphosphate (Wieland et al., 1994). Then, CrtP, CrtQ, CrtO, and AldH catalyze the oxidation, glycosylation and esterification reactions to convert this pigment into staphyloxanthin (Pelz et al., 2005; Kim and Lee, 2012). Staphyloxanthin protects *S. aureus* against oxidative stress by scavenging free radicals (Clauditz et al., 2006). But more generally carotenoids play a role in overall fitness (Clauditz et al., 2006; Johler et al., 2010) and could contribute to the growth and survival of *S. xylosus* in meat.

Finally, three genes encoding proteins that are enhanced when the cell is exposed to stress agents were down-regulated (Table 1).

All these data suggest that the conditions of growth in meat did not seem to generate stress other than osmotic and acid for *S. xylosus*.

CONCLUSION

The global gene expression of *S. xylosus in situ* in salted meat has allowed us to unravel its adaptation to this complex matrix in conditions that mimic the fermentation period of sausage manufacture. In these conditions, *S. xylosus* reached a plateau phase after 24 h and a balance between cell division and cell lysis was highlighted during this phase. *S. xylosus* adapted its metabolism to the meat nutrients and anaerobic conditions. It simultaneously used glucose and lactate as carbon sources. It used peptides and amino acids furnished by the meat. It has to cope essentially with the osmotic stress generated by addition of NaCl. It counteracted this stress by multiple strategies and particularly by the synthesis of glycine betaine, a powerful osmoprotectant.

AUTHOR CONTRIBUTIONS

AV: Contribution to acquisition, analysis and interpretation of the data, contribution in drafting the article. ED: Contribution to experimental and data analysis, critical revising of the manuscript. AD: Statistical data analysis. PM: Contribution to data analysis. VL: Chemical analysis. SL, RT: equally contribute to this paper, design of the work, analysis and interpretation of data, writing of the manuscript

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2016.00087>

REFERENCES

- Angelidis, A. S., and Smith, G. M. (2003). Role of the glycine betaine and carnitine transporters in adaptation of *Listeria monocytogenes* to chill stress in defined medium. *Appl. Environ. Microbiol.* 69, 7492–7498. doi: 10.1128/AEM.69.12.7492-7498.2003
- Aro Aro, J. M., Nyam-Osor, P., Tsuji, K., Shimada, K.-I., Fukushima, M., and Sekikawa, M. (2010). The effect of starter cultures on proteolytic changes and amino acid content in fermented sausages. *Food Chem.* 119, 279–285. doi: 10.1016/j.foodchem.2009.06.025
- Barrière, C., Centeno, D., Lebert, A., Leroy-Setrin, S., Berdague, J. L., and Talon, R. (2001). Roles of superoxide dismutase and catalase of *Staphylococcus xylosus* in the inhibition of linoleic acid oxidation. *FEMS Microbiol. Lett.* 201, 181–185. doi: 10.1016/S0378-1097(01)00271-3
- Battle, N., Aristoy, M.-C., and Toldrà, F. (2000). Early postmortem detection of exudative pork meat based on nucleotide content. *J. Food Sci.* 65, 413–416. doi: 10.1111/j.1365-2621.2000.tb16018.x

- Battle, N., Aristoy, M.-C., and Toldrà, F. (2001). ATP metabolites during aging of exudative and nonexudative pork meats. *J. Food Sci.* 66, 68–71. doi: 10.1111/j.1365-2621.2001.tb15583.x
- Beck, H. C., Hansen, A. M., and Lauritsen, F. R. (2004). Catabolism of leucine to branched-chain fatty acids in *Staphylococcus xylosum*. *J. Appl. Microbiol.* 96, 1185–1193. doi: 10.1111/j.1365-2672.2004.02253.x
- Bentley, R., and Meganathan, R. (1982). Biosynthesis of vitamin K (menaquinone) in bacteria. *Microbiol. Rev.* 46, 241–280.
- Berdagué, J. L., Monteil, P., Montel, M. C., and Talon, R. (1993). Effects of starter cultures on the formation of flavour compounds in dry sausage. *Meat Sci.* 35, 275–287. doi: 10.1016/0309-1740(93)90033-E
- Clauditz, A., Resch, A., Wieland, K.-P., Peschel, A., and Götz, F. (2006). Staphyloxanthin plays a role in the fitness of *Staphylococcus aureus* and its ability to cope with oxidative stress. *Infect. Immun.* 74, 4950–4953. doi: 10.1128/IAI.00204-06
- Cotter, P. D., and Hill, C. (2003). Surviving the acid test: responses of Gram-positive bacteria to low pH. *Microbiol. Mol. Biol. Rev.* 67, 429–453. doi: 10.1128/MMBR.67.3.429-453.2003
- Cretenet, M., Laroute, V., Ulve, V., Jeanson, S., Nouaille, S., Even, S., et al. (2011). Dynamic analysis of the *Lactococcus lactis* transcriptome in cheeses made from milk concentrated by ultrafiltration reveals multiple strategies of adaptation to stresses. *Appl. Environ. Microbiol.* 77, 247–257. doi: 10.1128/AEM.01174-10
- Demarquoy, J., Georges, B., Rigault, C., Royer, M.-C., Clairet, A., Soty, M., et al. (2004). Radioisotopic determination of l-carnitine content in foods commonly eaten in Western countries. *Food Chem.* 86, 137–142. doi: 10.1016/j.foodchem.2003.09.023
- den Blaauwen, T. (2013). Prokaryotic cell division: flexible and diverse. *Curr. Opin. Microbiol.* 16, 738–744. doi: 10.1016/j.mib.2013.09.002
- Derzelle, S., Bolotin, A., Mistou, M.-Y., and Rul, F. (2005). Proteome analysis of *Streptococcus thermophilus* grown in milk reveals pyruvate formate-lyase as the major upregulated protein. *Appl. Environ. Microbiol.* 71, 8597–8605. doi: 10.1128/AEM.71.12.8597-8605.2005
- Fadda, S., Anglade, P., Baraige, F., Zagorec, M., Talon, R., Vignolo, G., et al. (2010). Adaptive response of *Lactobacillus sakei* 23K during growth in the presence of meat extracts: a proteomic approach. *Int. J. Food Microbiol.* 142, 36–43. doi: 10.1016/j.ijfoodmicro.2010.05.014
- Ferreira, M. T., Manso, A. S., Gaspar, P., Pinho, M. G., and Neves, A. R. (2013). Effect of oxygen on glucose metabolism: utilization of lactate in *Staphylococcus aureus* as revealed by *in vivo* NMR Studies. *PLoS ONE* 8:e58277. doi: 10.1371/journal.pone.0058277
- Fiegler, H., Bassias, J., Jankovic, I., and Brückner, R. (1999). Identification of a gene in *Staphylococcus xylosum* encoding a novel glucose uptake protein. *J. Bacteriol.* 181, 4929–4936.
- Fiegler, H., and Brückner, R. (1997). Identification of the serine acetyltransferase gene of *Staphylococcus xylosum*. *FEMS Microbiol. Lett.* 148, 181–187. doi: 10.1111/j.1574-6968.1997.tb10286.x
- Fournier, B., and Hooper, D. C. (2000). A new two-component regulatory system involved in adhesion, autolysis, and extracellular proteolytic activity of *Staphylococcus aureus*. *J. Bacteriol.* 182, 3955–3964. doi: 10.1128/JB.182.14.3955-3964.2000
- Fuchs, S., Pané-Farré, J., Kohler, C., Hecker, M., and Engelmann, S. (2007). Anaerobic gene expression in *Staphylococcus aureus*. *J. Bacteriol.* 189, 4275–4289. doi: 10.1128/JB.00081-07
- Fujita, Y., and Fujita, T. (1987). The gluconate operon *gnt* of *Bacillus subtilis* encodes its own transcriptional negative regulator. *Proc. Natl. Acad. Sci. U.S.A.* 84, 4524–4528. doi: 10.1073/pnas.84.13.4524
- Fuller, J. R., Vitko, N. P., Perkowski, E. F., Scott, E., Khatri, D., Spontak, J. S., et al. (2011). Identification of a lactate-quinone oxidoreductase in *Staphylococcus aureus* that is essential for virulence. *Front. Cell. Infect. Microbiol.* 1:19. doi: 10.3389/fcimb.2011.00019
- Gagnaire, V., Piot, M., Camier, B., Vissers, J.-P., Jan, G., and Léonil, J. (2004). Survey of bacterial proteins released in cheese: a proteomic approach. *Int. J. Food Microbiol.* 94, 185–201. doi: 10.1016/j.ijfoodmicro.2004.01.010
- Gaupp, R., Schlag, S., Liebeke, M., Lalk, M., and Götz, F. (2010). Advantage of upregulation of succinate dehydrogenase in *Staphylococcus aureus* biofilms. *J. Bacteriol.* 192, 2385–2394. doi: 10.1128/JB.01472-09
- Green, J., Rolfe, M. D., and Smith, L. J. (2014). Transcriptional regulation of bacterial virulence gene expression by molecular oxygen and nitric oxide. *Virulence* 5, 794–809. doi: 10.4161/viru.27794
- Gunka, K., and Commichau, F. M. (2012). Control of glutamate homeostasis in *Bacillus subtilis*: a complex interplay between ammonium assimilation, glutamate biosynthesis and degradation. *Mol. Microbiol.* 85, 213–224. doi: 10.1111/j.1365-2958.2012.08105.x
- Hiron, A., Borezée-Durant, E., Piard, J.-C., and Juillard, V. (2007). Only one of four oligopeptide transport systems mediates nitrogen nutrition in *Staphylococcus aureus*. *J. Bacteriol.* 189, 5119–5129. doi: 10.1128/JB.00274-07
- Hüfner, E., Markieton, T., Chaillou, S., Crutz-Le Coq, A. M., Zagorec, M., and Hertel, C. (2007). Identification of *Lactobacillus sakei* genes induced during meat fermentation and their role in survival and growth. *Appl. Environ. Microbiol.* 73, 2522–2531. doi: 10.1128/AEM.02396-06
- Hughes, M. C., Kerry, J. P., Arendt, E. K., Kenneally, P. M., McSweeney, P. L., and O'Neill, E. E. (2002). Characterization of proteolysis during the ripening of semi-dry fermented sausages. *Meat Sci.* 62, 205–216. doi: 10.1016/S0309-1740(01)00248-0
- Janssens, M., Van der Mijnsbrugge, A., Sánchez Mainar, M., Balzarini, T., De Vuyst, L., and Leroy, F. (2014). The use of nucleosides and arginine as alternative energy sources by coagulase-negative staphylococci in view of meat fermentation. *Food Microbiol.* 39, 53–60. doi: 10.1016/j.fm.2013.11.004
- Johler, S., Stephan, R., Hartmann, I., Kuehner, K. A., and Lehner, A. (2010). Genes involved in yellow pigmentation of *Cronobacter sakazakii* ES5 and influence of pigmentation on persistence and growth under environmental stress. *Appl. Environ. Microbiol.* 76, 1053–1061. doi: 10.1128/AEM.01420-09
- Kappes, R. M., and Bremer, E. (1998). Response of *Bacillus subtilis* to high osmolarity: uptake of carnitine, crotonobetaine and gamma-butyrobetaine via the ABC transport system OpuC. *Microbiology* 144, 83–90. doi: 10.1099/00221287-144-1-83
- Kim, S. H., and Lee, P. C. (2012). Functional expression and extension of staphylococcal staphyloxanthin biosynthetic pathway in *Escherichia coli*. *J. Biol. Chem.* 287, 21575–21583. doi: 10.1074/jbc.M112.343020
- Knobloch, J. K., Bartscht, K., Sabottke, A., Rohde, H., Feucht, H. H., and Mack, D. (2001). Biofilm formation by *Staphylococcus epidermidis* depends on functional RsbU, an activator of the *sigB* operon: differential activation mechanisms due to ethanol and salt stress. *J. Bacteriol.* 183, 2624–2633. doi: 10.1128/JB.183.8.2624-2633.2001
- Koprivnjak, T., Mlakar, V., Swanson, L., Fournier, B., Peschel, A., and Weiss, J. P. (2006). Cation-induced transcriptional regulation of the *dlt* operon of *Staphylococcus aureus*. *J. Bacteriol.* 188, 3622–3630. doi: 10.1128/JB.188.10.3622-3630.2006
- Kuroda, M., Kuroda, H., Oshima, T., Takeuchi, F., Mori, H., and Hiramatsu, K. (2003). Two-component system *VraSR* positively modulates the regulation of cell-wall biosynthesis pathway in *Staphylococcus aureus*. *Mol. Microbiol.* 49, 807–821. doi: 10.1046/j.1365-2958.2003.03599.x
- Lafarga, T., and Hayes, M. (2014). Bioactive peptides from meat muscle and by-products: generation, functionality and application as functional ingredients. *Meat Sci.* 98, 227–239. doi: 10.1016/j.meatsci.2014.05.036
- Leroy, F., Verluyten, J., and De Vuyst, L. (2006). Functional meat starter cultures for improved sausage fermentation. *Int. J. Food Microbiol.* 106, 270–285. doi: 10.1016/j.ijfoodmicro.2005.06.027
- Levine, M., Truesdell, S., Ramakrishnan, T., and Bronson, M. J. (1975). Dual control of lysogeny by bacteriophage P22: an antirepressor locus and its controlling elements. *J. Mol. Biol.* 91, 421–438. doi: 10.1016/0022-2836(75)90270-3
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta Ct}$ Method. *Methods* 25, 402–408. doi: 10.1006/meth.2001.1262
- Majerczyk, C. D., Dunman, P. M., Luong, T. T., Lee, C. Y., Sadykov, M. R., Somerville, G. A., et al. (2010). Direct targets of CodY in *Staphylococcus aureus*. *J. Bacteriol.* 192, 2861–2877. doi: 10.1128/JB.00220-10
- Mann, E. E., Rice, K. C., Boles, B. R., Endres, J. L., Ranjit, D., Chandramohan, L., et al. (2009). Modulation of eDNA release and degradation affects *Staphylococcus aureus* biofilm maturation. *PLoS ONE* 4:e5822. doi: 10.1371/journal.pone.0005822

- Margolin, W. (2000). Themes and variations in prokaryotic cell division. *FEMS Microbiol. Rev.* 24, 531–548. doi: 10.1111/j.1574-6976.2000.tb00554.x
- Marini, P. E., Perez, C. A., and de Mendoza, D. (2001). Growth-rate regulation of the *Bacillus subtilis* accBC operon encoding subunits of acetyl-CoA carboxylase, the first enzyme of fatty acid synthesis. *Arch. Microbiol.* 175, 234–237. doi: 10.1007/s002030100256
- Morino, M., Suzuki, T., Ito, M., and Krulwich, T. A. (2014). Purification and functional reconstitution of a seven-subunit Mrp-type Na⁺/H⁺ antiporter. *J. Bacteriol.* 196, 28–35. doi: 10.1128/JB.01029-13
- Neubauer, H., Pantel, I., and Götz, F. (1998). Characterization of moeB- part of the molybdenum cofactor biosynthesis gene cluster in *Staphylococcus carnosus*. *FEMS Microbiol. Lett.* 164, 55–62. doi: 10.1016/S0378-1097(98)00196-7
- Neubauer, H., Pantel, I., Lindgren, P. E., and Götz, F. (1999). Characterization of the molybdate transport system ModABC of *Staphylococcus carnosus*. *Arch. Microbiol.* 172, 109–115. doi: 10.1007/s002030050747
- Neuhaus, F. C., and Baddiley, J. (2003). A continuum of anionic charge: structures and functions of d-alanyl-teichoic acids in gram-positive bacteria. *Microbiol. Mol. Biol. Rev.* 67, 686–723. doi: 10.1128/MMBR.67.4.686-723.2003
- Nouaille, S., Even, S., Charlier, C., Le Loir, Y., Coccagn-Bousquet, M., and Loubière, P. (2009). Transcriptomic response of *Lactococcus lactis* in mixed culture with *Staphylococcus aureus*. *Appl. Environ. Microbiol.* 75, 4473–4482. doi: 10.1128/AEM.02653-08
- Nuxoll, A. S., Halouska, S. M., Sadykov, M. R., Hanke, M. L., Bayles, K. W., Kielian, T., et al. (2012). CcpA regulates arginine biosynthesis in *Staphylococcus aureus* through repression of proline catabolism. *PLoS Pathog.* 8:e1003033. doi: 10.1371/journal.ppat.1003033
- Olesen, P. T., and Stahnke, L. H. (2004). The influence of environmental parameters on the catabolism of branched-chain amino acids by *Staphylococcus xylosum* and *Staphylococcus carnosus*. *Food Microbiol.* 21, 43–50. doi: 10.1016/S0740-0020(03)00048-0
- Pagels, M., Fuchs, S., Pané-Farré, J., Kohler, C., Menschner, L., Hecker, M., et al. (2010). Redox sensing by a Rex-family repressor is involved in the regulation of anaerobic gene expression in *Staphylococcus aureus*. *Mol. Microbiol.* 76, 1142–1161. doi: 10.1111/j.1365-2958.2010.07105.x
- Pelz, A., Wieland, K.-P., Putzbach, K., Hentschel, P., Albert, K., and Götz, F. (2005). Structure and biosynthesis of staphyloxanthin from *Staphylococcus aureus*. *J. Biol. Chem.* 280, 32493–32498. doi: 10.1074/jbc.M505070200
- Perkins, J. B., and Pero, J. (2002). “Vitamin biosynthesis,” in *Bacillus subtilis and Its Closest Relatives from Genes to Cells*, eds A. L. Sonenshein, J. A. Hoch, and R. Losick (New York, NY: ASM Press), 271–286.
- Ranjit, D. K., Endres, J. L., and Bayles, K. W. (2011). *Staphylococcus aureus* CidA and LrgA proteins exhibit holin-like properties. *J. Bacteriol.* 193, 2468–2476. doi: 10.1128/JB.01545-10
- Ravys, F., Vuyst, L. D., and Leroy, F. (2012). Bacterial diversity and functionalities in food fermentations. *Eng. Life Sci.* 12, 356–367. doi: 10.1002/elsc.201100119
- Reinhart, F., Huber, A., Thiele, R., and Uden, G. (2010). Response of the oxygen sensor NreB to air *in vivo*: Fe-S-containing NreB and Apo-NreB in aerobically and anaerobically growing *Staphylococcus carnosus*. *J. Bacteriol.* 192, 86–93. doi: 10.1128/JB.01248-09
- Rice, K. C., Patton, T., Yang, S.-J., Dumoulin, A., Bischoff, M., and Bayles, K. W. (2004). Transcription of the *Staphylococcus aureus* cid and lrg murein hydrolase regulators is affected by sigma factor B. *J. Bacteriol.* 186, 3029–3037. doi: 10.1128/JB.186.10.3029-3037.2004
- Rosenstein, R., Futter-Bryniok, D., and Götz, F. (1999). The choline-converting pathway in *Staphylococcus xylosum* C2A: genetic and physiological characterization. *J. Bacteriol.* 181, 2273–2278.
- Rosenstein, R., Nerz, C., Biswas, L., Resch, A., Raddatz, G., Schuster, S., et al. (2009). Genome analysis of the meat starter culture bacterium *Staphylococcus carnosus* TM300. *Appl. Environ. Microbiol.* 75, 811–822. doi: 10.1128/AEM.01982-08
- Rossi, M., Amaretti, A., and Raimondi, S. (2011). Folate production by probiotic bacteria. *Nutrients* 3, 118–134. doi: 10.3390/nu3010118
- Samuelson, J. C., Chen, M., Jiang, F., Möller, I., Wiedmann, M., Kuhn, A., et al. (2000). YidC mediates membrane protein insertion in bacteria. *Nature* 406, 637–641. doi: 10.1038/35020586
- Saum, S. H., Sydow, J. F., Palm, P., Pfeiffer, F., Oesterheld, D., and Müller, V. (2006). Biochemical and molecular characterization of the biosynthesis of glutamine and glutamate, two major compatible solutes in the moderately halophilic bacterium *Halobacillus halophilus*. *J. Bacteriol.* 188, 6808–6815. doi: 10.1128/JB.00781-06
- Schlag, S., Fuchs, S., Nerz, C., Gaupp, R., Engelmann, S., and Liebecke, M. (2008). Characterization of the oxygen responsive NreABC regulon of *Staphylococcus aureus*. *J. Bacteriol.* 190, 7847–7858. doi: 10.1128/JB.00905-08
- Schreier, H. J., Brown, S. W., Hirschi, K. D., Nomellini, J. F., and Sonenshein, A. L. (1989). Regulation of *Bacillus subtilis* glutamine synthetase gene expression by the product of the glnR gene. *J. Mol. Biol.* 210, 51–63. doi: 10.1016/0022-2836(89)90290-8
- Schujman, G. E., and de Mendoza, D. (2008). Regulation of type II fatty acid synthase in Gram-positive bacteria. *Curr. Opin. Microbiol.* 11, 148–152. doi: 10.1016/j.mib.2008.02.002
- Schujman, G. E., Paoletti, L., Grossman, A. D., and de Mendoza, D. (2003). FapR, a bacterial transcription factor involved in global regulation of membrane lipid biosynthesis. *Dev. Cell.* 4, 663–672. doi: 10.1016/S1534-5807(03)00123-0
- Shivers, R. P., and Sonenshein, A. L. (2004). Activation of the *Bacillus subtilis* global regulator CodY by direct interaction with branched-chain amino acids. *Mol. Microbiol.* 53, 599–611. doi: 10.1111/j.1365-2958.2004.04135.x
- Skaar, E. P., Humayun, M., Bae, T., DeBord, K. L., and Schneewind, O. (2004). Iron-source preference of *Staphylococcus aureus* infection. *Science* 305, 1626–1628. doi: 10.1126/science.1099930
- Smyth, G. K. (2004). Linear models and empirical bayes methods for assessing differential expression in microarray experiments. *Stat. Appl. Genet. Mol. Biol.* 3, 1–25. doi: 10.2202/1544-6115.1027
- Søndergaard, A. K., and Stahnke, L. H. (2002). Growth and aroma production by *Staphylococcus xylosum*, *S. carnosus* and *S. equorum*, a comparative study in model systems. *Int. J. Food Microbiol.* 75, 99–109. doi: 10.1016/S0168-1605(01)00729-2
- Stahnke, L. H. (1995). Dried sausages fermented with *Staphylococcus xylosum* at different temperatures and with different ingredient levels - Part II. Volatile components. *Meat Sci.* 41, 193–209. doi: 10.1016/0309-1740(94)00069-J
- Swartz, T. H., Ito, M., Ohira, T., Natsui, S., Hicks, D. B., and Krulwich, T. A. (2007). Catalytic properties of *Staphylococcus aureus* and *Bacillus* members of the secondary cation/proton antiporter-3 (Mrp) family are revealed by an optimized assay in an *Escherichia coli* host. *J. Bacteriol.* 189, 3081–3090. doi: 10.1128/JB.00021-07
- Switzer, R. L., Zalkin, H., and Saxild, H. H. (2002). “Purine, pyrimidine, and pyridine nucleotide metabolism,” in *Bacillus subtilis and Its Closest Relatives from Genes to Cells*, eds A. L. Sonenshein, J. A. Hoch and R. Losick (New York, NY: ASM Press), 255–269.
- Taïbi, A., Dabour, N., Lamoureux, M., Roy, D., and LaPointe, G. (2011). Comparative transcriptome analysis of *Lactococcus lactis* subsp. *cremoris* strains under conditions simulating Cheddar cheese manufacture. *Int. J. Food Microbiol.* 146, 263–275. doi: 10.1016/j.ijfoodmicro.2011.02.034
- Talon, R., and Leroy, S. (2006). “Latest developments in meat bacterial starters,” in *Advanced Technologies for Meat Processing*, eds L. M. L. Nollet and F. Toldrà (New York, NY: Taylor and Francis Group), 401–418.
- Tjener, K., Stahnke, L. H., Andersen, L., and Martinussen, J. (2004). The pH-unrelated influence of salt, temperature and manganese on aroma formation by *Staphylococcus xylosum* and *Staphylococcus carnosus* in a fermented meat model system. *Int. J. Food Microbiol.* 97, 31–42. doi: 10.1016/j.ijfoodmicro.2004.04.007
- Toldrà, F. (2007). “Biochemistry of meat and fat,” in *Handbook of Fermented Meat and Poultry*, ed F. Toldrà (Oxford: Blackwell Publishing), 51–58.
- Vermassen, A., de la Foye, A., Loux, V., Talon, R., and Leroy, S. (2014). Transcriptomic analysis of *Staphylococcus xylosum* in the presence of nitrate and nitrite in meat reveals its response to nitrosative stress. *Front. Microbio.* 5:691. doi: 10.3389/fmicb.2014.00691
- von Eiff, C., McNamara, P., Becker, K., Bates, D., Lei, X.-H., Ziman, M., et al. (2006). Phenotype microarray profiling of *Staphylococcus aureus* menD and hemB mutants with the small-colony-variant phenotype. *J. Bacteriol.* 188, 687–693. doi: 10.1128/JB.188.2.687-693.2006

- Wakeman, C. A., Hammer, N. D., Stauff, D. L., Attia, A. S., Anzaldi, L. L., Dikalov, S. I., et al. (2012). Menaquinone biosynthesis potentiates haem toxicity in *Staphylococcus aureus*. *Mol. Microbiol.* 86, 1376–1392. doi: 10.1111/mmi.12063
- Wieland, B., Feil, C., Gloria-Maercker, E., Thumm, G., Lechner, M., Bravo, J.-M., et al. (1994). Genetic and biochemical analyses of the biosynthesis of the yellow carotenoid 4,4'-diaponeurosporene of *Staphylococcus aureus*. *J. Bacteriol.* 176, 7719–7726.
- Yoshida, K., Ohmori, H., Miwa, Y., and Fujita, Y. (1995). *Bacillus subtilis* gnt repressor mutants that diminish gluconate-binding ability. *J. Bacteriol.* 177, 4813–4816.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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