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**OPEN ACCESS – RESEARCH ARTICLE**

**An empirical model that uses light attenuation and plant nitrogen status to predict within-canopy N distribution and upscale photosynthesis from leaf to whole canopy**

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## ABSTRACT

Modelling the spatial and temporal distribution of leaf nitrogen is central to specify photosynthetic parameters and simulate canopy photosynthesis. Leaf photosynthetic parameters depend both on local light availability and whole plant N status. The interaction between these two levels of integration has generally been modelled by assuming an optimal canopy functioning, which is not supported by experiments. During this study we examined how a set of empirical relationships with measurable parameters could be used instead to predict photosynthesis at the leaf and whole canopy levels. The distribution of leaf nitrogen per unit area ( $N_a$ ) within the canopy was related to leaf light irradiance and to the Nitrogen Nutrition Index (NNI), a whole plant variable accounting for plant N status.  $N_a$  was then used to determine the photosynthetic parameters of a leaf gas exchange model. The model was assessed on alfalfa canopies under contrasting N nutrition and with  $N_2$ -fixing and non-fixing plants. Three experiments were carried out to parameterise the relationships between  $N_a$ , leaf irradiance, NNI, and photosynthetic parameters. An additional independent dataset was used for model evaluation. The N distribution model showed it was able to predict leaf nitrogen on the set of leaves tested.  $N_a$  at the top of the canopy appeared to be related linearly to the NNI whereas the coefficient accounting for nitrogen allocation remained constant. Photosynthetic parameters were related linearly to  $N_a$  irrespective of N nutrition and the N acquisition mode. Daily patterns of gas exchange were simulated accurately at the leaf scale. When integrated at the whole canopy scale, the model predicted that raising N availability above an NNI of 1 did not result in increased net photosynthesis. Overall, the model

proposed offered a solution for a dynamic coupling of leaf photosynthesis and canopy N distribution without requiring any optimal functioning hypothesis.

**Keywords:** Light; *Medicago sativa*; nitrogen distribution; nitrogen nutrition index; photosynthesis; transpiration; up-scaling; within-canopy variability.

## INTRODUCTION

A close positive relationship exists between the nitrogen content and photosynthetic capacity of leaves (Field and Mooney 1986; Evans 1989; Hikosaka *et al.* 2004). Such a relationship is the cornerstone of various approaches proposed to up-scale leaf gas-exchange models to the whole canopy level (Leuning 1995; Kull and Jarvis 1995; De Pury and Farquhar 1997; Sinoquet *et al.* 2000; Evers *et al.* 2010). Indeed, it has been shown that the within-canopy variability of photosynthetic parameters can be fully specified at a given time by measurements of the spatial distribution of leaf nitrogen (Harley *et al.* 1992; Le Roux *et al.* 1999; Braune *et al.* 2009). Changes in leaf nitrogen concentration with canopy depth, and the effects of leaf age and leaf light microclimate, have been identified as major sources of spatial variation (Evans 1989). They have been studied extensively in several species and different modelling approaches enable to account for it.

Following the optimization theory, several authors first sought to model nitrogen distribution in order to maximise canopy photosynthesis (Charles-Edwards 1981; Field 1983; Hirose and Werger 1987). The outcome was models predicting a leaf nitrogen concentration which paralleled radiation extinction (Kull and Jarvis 1995; Sands 1995). However, there is no a priori reason for N distribution to follow such a pattern. On the contrary, empirical observations have consistently indicated that the exponential fall in leaf nitrogen with increasing depth into the canopy occurs in most canopies at a slower rate than light extinction (Hirose and Werger 1987; Lemaire *et al.* 1991; Anten 1995; Moreau *et al.* 2012). Alternatively, empirical relationships between light extinction and leaf nitrogen have been used to mimic a local light acclimation and model leaf N distribution by considering potential departures from the light gradient (Sellers *et al.* 1992; Anten 1995; Prieto *et al.* 2012). The approach is

usually based on a coefficient of nitrogen allocation ( $k_N$ ) which shapes the nitrogen profile with respect to relative light extinction ( $I/I_0$ ), and a reference leaf nitrogen concentration at the top of the canopy ( $N_{up}$ ):

$$N_a = N_{up} \cdot (I / I_0)^{k_N} \quad \text{Eq. 1}$$

Leaf nitrogen distribution is not solely a function of light and age, however. It is also dependent on mineral nitrogen availability (Hikosaka *et al.* 1994; Lötscher *et al.* 2003), N demand to support plant growth, and more generally on the nitrogen status of plants (i.e. the relative satisfaction of plant N demand, Lemaire and Gastal 1997). The N demand of a plant at any time in its cycle is generally defined as the amount of N necessary to sustain maximal plant growth. N demand is tightly related to the standing crop mass. On a mass increment basis, it decreases as biomass increases, resulting in an apparent dilution of plant N concentration with plant growth (Greenwood *et al.* 1990; Gastal *et al.* 2015). Canopy N content and leaf N distribution thus respond to changes in the fertilisation rate (Bélanger *et al.* 1992; Dreccer *et al.* 2000) but also to all factors that affect the plant growth rate (e.g. temperature, CO<sub>2</sub> concentration, Pettersson and McDonald 1994). Empirical plant N status indices have been developed to account for both aspects and help to diagnose crop N requirements. For instance, the Nitrogen Nutrition Index (NNI) was assessed on plants as different as C<sub>3</sub> annual crops (e.g. Justes *et al.* 1994; Colnenne *et al.* 1998), C<sub>4</sub> grasses (Plénet and Lemaire 1999) and perennial forage plants (Lemaire and Salette 1984; Lemaire *et al.* 1985). This is based on the concept of critical N dilution which can be applied in dynamic terms and is able to account for temporal changes in the nitrogen nutrition of crops (e.g. STICS crop model, Brisson *et al.* 2009).

Modelling the effect of the interaction between light acclimation and nitrogen limitations on the distribution of leaf N and photosynthetic characteristics has received comparatively less attention (Thornley 1998). One challenge is that light acclimation is a local process driven by the leaf light microclimate (Evans 1989; Hikosaka *et al.* 1994), whereas N demand, plant N status and N allocation are defined at the whole plant scale (Givnish 1988; Lemaire and Gastal 1997; Kull 2002; Gastal *et al.* 2015). Empirical relationships between light extinction and leaf nitrogen generally refer to static canopies at a given developmental stage, and their parameters need to be adjusted between years, sites or nitrogen treatments (Prieto *et al.* 2012). To date, dynamic coupling with plant growth has thus mainly been achieved using approaches based on the optimal distribution theory (Johnson *et al.* 2010). Some studies demonstrated a significant relationship between canopy NNI and the  $k_N$  and  $N_{up}$  parameters (Farrugia *et al.* 2004; Lötscher *et al.* 2003; Gastal *et al.* 2015). These relationships could be tested to make predictions of photosynthetic parameters under contrasting N availabilities without any *a priori* assumptions regarding optimal functioning of the canopy. Such a model would offer a solution to dynamically simulate the interactions between light and nitrogen based on parameters which can be directly measured.

During this study, we developed and assessed a model coupling an empirical canopy N distribution model with a leaf gas exchange model derived from Farquhar *et al.* (1980). The distribution of leaf nitrogen content per unit area ( $N_a$ ) was related to leaf light irradiance and to the canopy Nitrogen Nutrition Index (NNI). The objectives were to determine whether such an empirical approach to leaf N distribution could be used to specify spatial and temporal changes in leaf gas exchange under fluctuating light and N availability. Alfalfa was chosen as a model species because its leaf N

distribution has already been described extensively under non-limiting N and because this species presents limited age dependency of leaf characteristics (Lemaire *et al.* 1991; Evans 1993; Lemaire *et al.* 2005).

## METHODS

### Model description

**Canopy N distribution model** - We assumed that spatial and temporal variations in leaf nitrogen content per unit area ( $N_a$ ) within the canopy and in the course of plant growth can be deduced from leaf light exposure and plant nitrogen status. The effect of relative leaf irradiance on relative  $N_a$  was taken into account using Eq. 1 with the two parameters  $N_{up}$  and  $k_N$ . The effect of N limitation was assumed to affect whole canopy N content in leaves by modulating these two parameters. The Nitrogen Nutrition Index was considered to account for the effect of canopy N status (integrating the effects of soil mineral N and nodule fixation on internal N availability).

At a given time, NNI was defined as:

$$NNI = N_m/N_c \quad \text{Eq. 2}$$

where  $N_m$  represents the actual plant N concentration and  $N_c$  the critical plant N concentration (g N.100 g<sup>-1</sup> plant) corresponding to its mass  $W$  (given by equation  $N_c = 4.8 \cdot W^{0.33}$  in alfalfa, Lemaire *et al.* 1985). When NNI is close to 1, the plant N status is considered as near optimum. Departures from 1 indicate deficiency (NNI <1; the intensity of deficiency is then equal to 1–NNI) or excess nitrogen (NNI >1, the intensity of excess is then equal to NNI–1). Following Farrugia *et al.* (2004), a linear response of  $N_{up}$  to NNI was considered:

$$N_{up} = N_{up}^{opt} + a_2 \cdot (NNI - 1) \quad \text{Eq. 3}$$

where  $N_{up}^{opt}$  represents the nitrogen content of leaves exposed to incoming PAR radiations for a NNI of 1 and  $a_2$  represents the dependency of upper leaf N content on plant N status. Similarly, the coefficient of nitrogen distribution relative to the light gradient was assumed to depend from NNI:

$$k_N = k_N^{opt} + a_3^*(NNI-1) \quad \text{Eq. 4}$$

where  $k_N^{opt}$  represents the allocation coefficient for a NNI of 1 and  $a_3$  represents the dependency of this coefficient on plant N status.

**Leaf gas exchange model** - The leaf gas exchange model is described in details in Prieto *et al.* (2012) and has originally been assessed on grapevine. It combines the biochemical photosynthetic model developed by Farquhar *et al.* (1980) with a semi-empirical stomatal conductance model that was originally proposed by Ball *et al.* (1987) and then modified by Leuning (1995). All the equations, variables and parameters are presented in Appendix 1. The coupling of this leaf gas exchange model with the previously presented canopy N distribution model was performed through the dependency of the principal photosynthesis parameters (Value of  $V_{cmax}$ ,  $J_{max}$ ,  $TPU$  and  $R_d$  at 25 °C) to  $N_a$ . A linear relationship was assumed [Eq. (A9)] (Harley *et al.* 1992; Le Roux *et al.* 1999; Braune *et al.* 2009).

### Model calibration

Three experiments were carried out at the INRA Lusignan research station, France (46.43N, 0.18E), to calibrate this model and assess the impacts of light, N nutrition and leaf age on the distribution of leaf photosynthetic parameters in alfalfa (*Medicago sativa*). The three experiments were based on the same cultivars (cv. "Orca" as a regular nitrogen fixing material, and cv. "Agate NF" as a non-N<sub>2</sub>-fixing material, Barnes *et al.* 1990).

**Experiment 1** - The first experiment was performed in a growth chamber between March and June 2010. All plants were grown in 1.5L pots (10 x 20 cm cylindrical pots) filled with an N-free substrate (fine quartz sand, 0.8-1.4 mm mesh). The pots were arranged in a quincunx and two plants were transplanted into each pot, resulting in a planting density of about 230 plants.m<sup>-2</sup>. Three canopies comprising 81 pots each (i.e. 162 plants each) were grown under contrasting N availabilities at 22°C/17°C (day/night) under a 14h photoperiod. The incident photosynthetic photon flux density (PPFD) was about 400 μmol.m<sup>-2</sup>.s<sup>-1</sup>. Each canopy was surrounded by a row of border plants grown under the same conditions. Two of these canopies were sown using the “Orca” cultivar and were ferti-irrigated every four hours (daily amount of 200 mL.pot<sup>-1</sup>) with either a complete nutrient solution (N+, 8 mMol N) or a low N nutrient solution (N-, 0.5 mMol N). The nitrogen concentration of the N+ solution was non-limiting for growth and prevented the nodulation of alfalfa roots. With the N- solution however, nodulation and nitrogen fixation did occur in the Orca cultivar (with natural strains of rhizobium, since the plants were not inoculated). The third canopy was sown with the “Agate NF” cultivar grown with the N- nutrient solution, so that nitrogen fixation could not compensate for low mineral N availability. In order to induce a size hierarchy into the canopy, and to decorrelate the vertical position of leaves from their age, alternate rows were sown with a 17-day delay in each canopy. The study focused on the initial growth period (no defoliation). Two samplings were performed in order to characterise the leaf N distribution. The plants were at the 12<sup>th</sup> visible leaves stage (40 days after the first sowing) and beginning of bloom stage (58 days after the first sowing), based on development of the Orca-N+ canopy. At each date, eight pots (16 plants) were collected from the centre of each canopy.

**Experiment 2** - The second experiment was performed outdoors between April and August 2009 using the “Orca” cultivar. The average incident photosynthetic photon flux density (PPFD) was about  $725 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ . All plants were grown in individual 1 L pots (5x52 cm cylindrical pots), resulting in a plant density of 460 plants. $\text{m}^{-2}$ . The canopy was made up of 100 study pots surrounded by three rows of border plants grown under the same conditions. All pots were filled with a growing medium that comprised sterile potting mix sand and clay-sandy-loam soil from a field in Lusignan (1:1:1, v/v). They were ferti-irrigated three times a day with the N+ nutrient solution. At the end of the second regrowth (beginning of bloom stage), 20 plants were sampled from the centre of the canopy for the characterisation of leaf N distribution. The plants in this canopy had previously been shown to be highly size-structured (Baldissera *et al.* 2014).

**Experiment 3** - The third experiment was carried out in a greenhouse between February and June 2012 using the “Orca” cultivar. The average incident photosynthetic photon flux density (PPFD) was about  $540 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ . All plants were grown in 1.1L pots (10 x 10 x 11 cm) filled with an N-free substrate (fine quartz sand, 0.8-1.4 mm mesh). A single plant was transplanted into each pot, resulting in a density of 100 plants. $\text{m}^{-2}$ . Just after transplantation, the seedlings were inoculated with a solid commercial preparation for the coating of alfalfa seeds (*Sinorhizobium meliloti*, strain 2011, Becker Underwood). The pots were automatically ferti-irrigated five times a day with a complete nutrient solution devoid of mineral nitrogen (N0, 0 mMol N). The nutrient solution was sampled weekly to determine the absence of NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> and ensure that nitrogen fixation was the only source of nitrogen supplied to the alfalfa plants. The experimental design consisted of four contiguous blocks of 49 pots each. At the end of the initial growth period (mid-bloom stage), four

plants were sampled from the centre of each block in order to characterise the leaf N distribution.

**Measurement of canopy N distribution and NNI** - At each sampling date specified in the three experiments, each plant was separated into stems, flowers (when present) and leaves. The leaves were sub-divided into 10 cm strata from the bottom to the top of the plant. The leaf area into each strata was determined using an LI-3100 planimeter (Li-Cor, Lincoln, NE, USA). Plant samples were dried at 60°C for 2 days, weighed to determine the dry mass and finally ground in a vibrating ball mill (MM400, Retsch GmbH and Co, Haan, Germany). Each sample was analysed with an elemental analyser to determine the nitrogen content (model EA 1108, Carlo Erba Instruments, Milan, Italy). For each canopy, the leaf area index (LAI) was calculated as the average leaf area of the collected plant multiplied by the plant density. The nitrogen nutrition index (NNI) was calculated as the ratio between the canopy N concentration ( $N_m$ ) and the critical N concentration ( $N_c$ , Eq. 2).

**Leaf photosynthetic capacity** - In Exp. 1 to 3, gas exchanges were measured using the same procedure with a portable Licor 6400 photosynthesis system (LI-6400, Li-Cor, Lincoln, NE, USA). The photosynthetic parameters were determined through the response of  $A$  to the internal  $\text{CO}_2$  concentration ( $C_i$ ) at the sub-stomatal level ( $A-C_i$  curves). Different levels of  $C_i$  were obtained by modifying the ambient  $\text{CO}_2$  concentration ( $C_a$ ) in the leaf measurement chamber. The  $A-C_i$  curves were determined as proposed by Long and Bernacchi (2003). Each  $C_a$  step was maintained for 5 minutes in order to record stable values. The three parameters ( $V_{cmax}$ ,  $J_{max}$ , and  $TPU$ ) were estimated simultaneously by fitting the Farquhar model to the whole  $A-C_i$  curve according to the procedure proposed by Sharkey *et al.* (2007). All curves were determined at  $1500 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  of photosynthetic photon flux

density (PPFD), while the leaf temperature was controlled at 25°C and the vapour pressure deficit (VPD) between the leaf and the air was kept at  $1 \pm 0.5$  kPa. Night respiration was estimated at the end of the night on a sub-sample of leaves which had previously been used for photosynthesis measurements ( $A-C_i$  curves). A different leaflet from the same leaf was used. Night respiration was considered to be equal to day respiration ( $R_d$ ).

In each of the canopies studied, leaves were sampled at three to four levels from the bottom to the top of the canopy, just before the plants were collected to assess canopy N content and leaf N distribution. Measurements were made on the central leaflet of primary leaves. A total of 102  $A-C_i$  curves were analysed over the three experiments (70, 20 and 12 for Exp. 1, 2 and 3, respectively).

**Stomatal conductance** - In Exp. 2, the daily evolution of leaf transpiration was recorded on leaves with contrasting positions within the canopy. Measurements were carried out during a series of sunny and cloudy days in summer. The parameters of the stomatal conductance model [Eq. (A10)] were estimated using the data obtained during a period which cumulated about five days of measurements.

**Leaf traits** - For each of the leaves on which  $A-C_i$  curves were determined, the leaf age was calculated by the thermal time difference (in °Cd) between the date of measurement and the date of leaf appearance. Thermal time was calculated from the daily integration of air temperatures minus the base temperature (5°C). Immediately after the gas exchange measurements, the three leaflets were collected and scanned (Konica Minolta C352/C300, Konica Minolta Sensing, Osaka, Japan). The leaf area was determined using image analysis (ImageJ software, <http://rsbweb.nih.gov/ij/>). The leaves were then dried at 60°C for 2 days, weighed to determine the dry mass and then ground in a vibrating ball mill (MM400, Retsch GmbH and Co, Haan,

Germany). Leaf samples were analysed with an elemental analyser (model EA 1108, Carlo Erba Instruments, Milan, Italy) to determine their N concentration. The Specific Leaf Area ( $SLA$ ,  $m^2.g^{-1}$ ), leaf nitrogen content per unit dry mass (%) and leaf nitrogen content per unit of area ( $N_a$ ,  $g\ N.m^{-2}$ ) were then calculated.

**Determination of local light conditions** - In Exp. 1, the PPFD values at the top of the canopy and at the leaf level (for each leaf used for the  $A-C_i$  characterisations) was measured using a portable LI-189 quantum meter (Li-Cor, Lincoln, NE, USA). In addition, in all the canopies studied, the vertical distributions of leaf area measured at each sampling date were used to compute light extinction and average PPFD levels corresponding to each leaf stratum, using the RATP model (Sinoquet *et al.* 2000). A leaf angle distribution was derived from measurements of alfalfa architecture in Exp. 3 (see Barillot *et al.* 2011, for details).

### **Assessment of the leaf gas exchange model at the leaf level**

A separate experiment was carried out between March and June 2011 according to the same design as Exp. 1. A dataset of ten leaves was used to assess the ability of the model to predict leaf N from canopy NNI and local leaf irradiance. These leaves were collected from the Orca-N+ and Orca-N- treatments (NNI values of 1.1 and 0.9, respectively) at the beginning of the bloom stage. To evaluate the ability of the model to simulate responses to rapid changes in environmental conditions, the sampled plants were placed outdoors and daily evolutions of the leaf gas exchange were recorded. Measurements were taken during sunny and cloudy days on leaves at different heights within the canopies. On very cloudy days, the plants were placed under a shelter to protect the material from the rain. A total of 14 days was analysed. The incident photosynthetic photon flux density (PPFD), leaf temperature, VPD and  $C_a$  were measured.

## Assessment of the leaf gas exchange model at the whole canopy level

The behaviour of the leaf gas exchange model when up-scaled to the whole canopy level was also assessed. The leaf N distribution was simulated for contrasting canopies (i.e. LAI values of 1.5, 3 and 5 m<sup>2</sup>.m<sup>-2</sup>) at NNI values ranging from 0.3 to 1.4. Leaf area was assumed to be distributed homogeneously into eight vertical strata, and the leaf N concentration in each stratum was assumed to be acclimated to the relative light irradiance integrated over the day. A leaf angle distribution was derived from measurements of alfalfa architecture in Exp. 3 (see Barillot *et al.* 2011, for details). The light distribution within the canopy was calculated hourly using the RATP model (Sinoquet *et al.* 2000). Simulations were performed for contrasting days in the series used for validation at the leaf level. Net photosynthesis was calculated within each stratum, and then summed to determine aboveground whole canopy net gas exchanges.

### Statistical analyses

Statistical analyses were performed using R software (R Development Core Team, 2012). Curve fittings were realised with the *nls* procedure for Eq. 2 and with the *lm* procedure for linear regressions (Eq. 3 and 4). Analyses of covariance (ANCOVA, *lm* procedure) were used to test for the effects of continuous and categorical variables simultaneously and to compare the slopes and intercepts of linear relationships between nitrogen concentration and photosynthetic parameters.

Predicted and measured values of leaf nitrogen concentration and net photosynthesis were compared using the root mean square error (*RMSE*) and bias (*Bias*) of the model, calculated as follows:

$$RMSE = \sqrt{\frac{\sum_{i=1}^n (s_i - m_i)^2}{n}} \quad \text{Eq. 5.}$$

$$\text{Bias} = \frac{\sum_{i=1}^n (s_i - m_i)}{n} \quad \text{Eq. 6}$$

where  $s_i$  and  $m_i$  are the  $i^{\text{th}}$  simulated and measured values respectively, and  $n$  is the number of observations.

## RESULTS

### Impact of NNI on the relationship between leaf irradiance and leaf nitrogen

The relationship between  $N_a$  and relative leaf irradiance was markedly affected by the nitrogen nutrition of the plants. Table 1 summarises the parameters obtained by fitting Eq. 1 to the different nitrogen nutrition situations studied. Parameter  $N_{up}$  was the most affected, ranging from about 2.4 g.m<sup>-2</sup> under N+ treatments to 0.8 g.m<sup>-2</sup> in N- plants reliant on mineral nitrogen assimilation alone. It related linearly to the NNI of the plant stand (Fig. 1a; Eq. 3). Variations in  $N_{up}$  thus reflected variations in N nutrition and internal N availability. By comparison, parameter  $k_N$ , which accounted for N allocation with respect to relative leaf irradiance, displayed little variation. For  $k_N = 1$ , the nitrogen gradient parallels the light gradient within the canopy. All observed values were clearly inferior to unity (<0.5), indicating a more-than-proportional N allocation to leaves with high irradiance and making the N concentration decrease more slowly than relative irradiance. Most  $k_N$  values were within the narrow range between 0.2 and 0.3 (except for one 0.09) with no clear relation to NNI. A single  $k_N$  parameter (0.25) enabled us to fit the normalised  $N_a$  distributions (Fig. 1b).

The effect of leaf age on  $N_a$  distributions was also assessed (Figure Sup. A and B). Due to an upward age gradient in alfalfa canopies, leaf nitrogen per unit area was related to both leaf age and local irradiance when considered separately. A multiple regression analysis confirmed the dominant effect of relative irradiance (t-value=

5.11;  $P < 10^{-6}$ ), but showed a non significant impact of leaf age *per se* and no interaction with irradiance (t-value= 1.54;  $P > 0.12$  for the age term).

The canopy N distribution model was parameterised on the basis of these relationships ( $a_2=2.15$ ,  $N_{up}^{opt}=2.17$ ,  $a_3=0$ . and  $k_N^{opt} = 0.25$ ). Figure Sup. C shows the change in leaf nitrogen concentration as a function of relative leaf irradiance and NNI as predicted by this model.

### Parameters of the photosynthetic and stomatal conductance model

Parameters  $V_{cmax}^{25}$ ,  $J_{max}^{25}$ ,  $TPU^{25}$  and  $R_d^{25}$  were related linearly to  $N_a$  (Fig. 2). The range of values observed for the different leaf parameters varied significantly between experiments, in relation to the minimum and maximum values taken by  $N_a$  (i.e. up to 2.8, 2, 1.8 and 1.1 g N.m<sup>-2</sup> in canopies relying on the N+ solution, N- solution and fixation, fixation only and N- solution only, respectively). The range of variations in  $N_a$  values resulted from both the nitrogen nutrition of plants and leaf-to-leaf variations in the light microclimate. However, a single relationship was found for each parameter between  $P^{25}$  and  $N_a$ , independently of the N nutrition. No significant difference in the slopes (ANCOVA, t-value < 0.91;  $p > 0.38$  for interactions terms between  $P^{25}$  and  $N_a$ ) and intercepts (ANCOVA, t-value < 0.31,  $p > 0.75$ ) were found between the nitrogen treatments. A larger dispersion of the points within the 'N+-assimilation' dataset was observed. This was due to a difference between indoor and outdoor  $N_a$  values as shown by slightly higher intercepts in Exp.2. (ANCOVA, t-value < -3.84,  $p < 0.001$  for the intercept term).

An unique set of measurements from Exp. 2 was used to determine the stomatal conductance parameters ( $a_1$ ,  $D_o$ ). It contained leaves from different positions within the canopy and days with contrasting meteorological conditions. Parameter values are presented in Appendix A2.

### Quantitative assessment of the nitrogen distribution model

Figure 3 compares the simulated  $N_a$  values (Eq. 1 and 3) with values measured on leaves at various heights within canopies grown under low and high N availability (NNI ranging from 0.45 to 1.1). Most inter-leaf variance in  $N_a$  values was explained by the nitrogen distribution model ( $r^2=0.85$ ). The model error remained low (RMSE=0.28 g N.m<sup>-2</sup>), but a significant positive bias was observed. Predicted values of  $N_a$  appeared to be slightly higher on average (Bias=+0.20 g N.m<sup>-2</sup>), particularly in leaves at an intermediate height within the canopy.

### Quantitative assessment of the leaf gas exchange model

The photosynthesis and transpiration sub-models were further assessed using directly measured leaf  $N_a$ . In a first step, the photosynthetic parameters ( $V_{cmax}^{25}$ ,  $J_{max}^{25}$ ,  $TPU^{25}$  and  $R_d^{25}$ ) were calculated using measured  $N_a$  as an input. The gas exchange model was then run to simulate the daily patterns of  $A$  and  $E$  in a range of contrasting leaves (taken from various heights within canopies grown under low and high N availability). The model correctly predicted the diurnal patterns of  $A$  and  $E$  in various leaves under contrasting environmental conditions (Fig. 4). Cumulated over a day, the relationship between the observed and simulated values of  $A$  and  $E$  did not differ significantly from the 1:1 line (Fig. 5;  $P<0.05$ ). The model accurately estimated the diurnal patterns of  $A$  and its variation associated with climatic scenarios and leaves under high or low N status (RMSE=0.04, no significant bias). The predictions also agreed satisfactorily for  $E$ , but the model errors were greater. Significant discrepancies were observed on  $E$  predictions for leaves with a high N content on sunny days (e.g. DOY 177 for a leaf at 1.96 g N.m<sup>-2</sup>, Fig. 4h). An underestimation of transpiration of up to 20% was observed under such conditions. This bias did not

result from unpaired temporal predictions at a particular time of the day, but from a general underestimation throughout the day.

### **Model predictions of the whole canopy response to nitrogen availability**

The behaviour of the model integrated at the whole canopy level was assessed for canopies growing under a range of nitrogen availabilities. Examples of daily integrated canopy assimilation are presented in Figure 6 for three contrasting days (DOY 176, 177 and 157 with an average PPFD decreasing from 709 to 610 and to 263  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  and average air temperatures of 20.1°C, 26.3°C and 17.9°C, respectively). All canopies presented a saturating response curve to nitrogen availability. As expected, canopy assimilation was lower during cloudy days (Fig. 6a-c). Canopies with a leaf area index lower than that required for canopy closure (LAI below 3  $\text{m}^2\cdot\text{m}^{-2}$ ) always displayed a lower assimilation rate per unit of soil area. Further increasing the LAI after canopy closure (LAI above 3  $\text{m}^2\cdot\text{m}^{-2}$ ) did not improve canopy assimilation. The threshold at which canopy assimilation ceased to respond to N availability was very close to an NNI value of 1 for closed canopies during sunny and moderately cloudy days (Fig. 6d-e), and for open canopies during very cloudy days (Fig. 6f). Slight shifts of threshold were predicted, depending on the canopy LAI and light availability. Open canopies appeared to be more able to valorise high nitrogen availability and displayed delayed thresholds (e.g. at an NNI of about 1.2 on sunny days). By contrast, dense canopies presented anticipated thresholds that were particularly apparent on cloudy days.

## DISCUSSION

### A simple empirical model to link leaf nitrogen distribution with plant N status and light distribution

To date, modelling the interaction between nitrogen limitations and light acclimation has been tackled using 'goal seeking' or optimal distribution theory (Chen *et al.* 1993; Thornley 1998; Johnson *et al.* 2010). Our study demonstrated how a combination of empirical relationships might be a promising option for this purpose too. The strategy proposed is based on modulation of the  $N_{up}$  and  $k_N$  parameters used in the empirical distribution model as a function of plant N status (NNI). A linear relationship was found between  $N_{up}$  and NNI over the range of alfalfa canopies studied. Similar results had previously been reported in different grass species, where the relationship was shown to be stable under contrasting growth conditions and canopy structures (Farrugia *et al.* 2004; Gastal *et al.* 2015). In these species,  $N_{up}$  has even been used as a routine proxy to facilitate the determination of NNI in the field (Louarn *et al.* 2015; Maamouri *et al.* 2015). The second parameter in the empirical relationship,  $k_N$ , was shown to be independent from NNI during the present study. Depending on the species however, contradictory results have been reported concerning the effect of N limitation on  $k_N$ . In some cases, limited effects have been observed (Sinclair and Shiraiwa 1993; Sadras *et al.* 2012), whereas in others a steeper N gradient has been found in N stressed plants (Milroy *et al.* 2001; Dreecer *et al.* 2000). Moreau *et al.* (2012) suggested that the size of the canopy (indirectly reduced by N stress), rather than a direct NNI effect, might explain the steeper gradient in N limited wheat canopies. In line with our results, Lemaire *et al.* (1991) did not show any variation of  $k_N$  in alfalfa canopies at contrasting developmental stages. Different types of plant architecture may affect N reallocation strategies and

contribute to explaining these differences in the  $k_N$  response. Some species, such as alfalfa or sunflower (Archontoulis et al. 2011), are made up of leaves distributed in different strata along the vertical light gradient, and may adjust more efficiently than long-leaf species (such as grasses or cereals) in which each leaf may simultaneously experience light conditions from the bottom to the top of the canopy. In those cases, the parameter  $a_3$  representing the dependency of  $k_N$  on plant N status (Eq. 3) is likely to take values different from zero.

### **Assumptions and potential limitations of the leaf N distribution model**

Species differ in the plasticity of their leaf traits and in the within canopy variation of photosynthetic characteristics (Niinemets et al. 2015). The present model assumes that the distribution of leaf N is mainly driven by two factors: the light gradient within the canopy and the plant N status. No significant age effects were recorded in alfalfa, as previously shown in several other species (Evans 1993; Hikosaka *et al.* 1994). This is not a general feature however, and many plant species display age-dependent leaf traits, such as decreasing SLA in ageing leaves for instance. This can alter the light-  $N_a$  relationship and limit the validity of our model (Prieto et al. 2012). In their recent review, Niinemets et al. (2015) distinguished two main groups of species: a first group with high rates of canopy development and leaf turnover, exhibiting highly dynamic light environments, active change photosynthetic characteristics by nitrogen reallocation among leaves, and a second group made up of species with slow leaf turnover exhibiting a passive  $N_a$  acclimation response, primarily determined by the acclimation of leaf structure. The proposed model appears clearly best suited to the first group of species because they are less susceptible to leaf ageing effects.

Another limitation of empirical models is their validity out of their domain of calibration. Other environmental factors, such as water stress (Errecart et al. 2014), extreme temperatures (Zaka et al. 2015) or extreme light environments (as shown by the indoor/outdoor effect in our dataset), can affect leaf growth and leaf traits. In some case this is likely to imply a reassessment model parameter values. Finally, the two-parameter model used (Eq. 1, Sup. Table A) could present a lack of flexibility in some species. This formalism was previously used on other crops (Moreau et al. 2012; Sadras et al. 2012), but studies comparing a large number of species have generally relied on three-parameter models because they presented an overall better fit (Lötsher et al. 2003; Niinemets et al. 2015).

### **The N acquisition mode did not affect the photosynthetic parameters**

Our study confirmed in alfalfa a linear relationship between Farquhar photosynthetic parameters ( $P_{25}$ ) and leaf nitrogen per unit leaf area (Field 1983; Evans 1989) and showed that it holds true for leaves in different canopies grown under contrasting mineral N availabilities. Acclimation to light and plant N status both affected the nitrogen concentration of leaves, but the  $N_a - P_{25}$  relationships remained unchanged, as shown previously (Braune *et al.* 2009). In addition, our study examined the effects of the N acquisition mode in legumes, comparing fixing and non fixing genotypes of alfalfa under different N nutrition statuses. There was no significant impact of the N acquisition mode on the  $N_a - P_{25}$  relationships. We thus showed that, contrary to the whole plant level (Gosse *et al.* 1986), no-extra-cost to carbon acquisition was associated with nitrogen fixation at the leaf level (Boller and Heichel 1984). No gain, associated to an extra carbon-sink, was observed either. Overall, the  $N_a - P_{25}$  relationships established during this study on a perennial forage legume displayed slopes (e.g.  $sNa$  of the  $N_a - V_{cmax}^{25}$  relationship at  $53 \mu\text{mol.g}$

$\text{N}^{-1}.\text{s}^{-1}$ ) that were intermediate between those of cereals (60 and  $63 \mu\text{mol.g N}^{-1}.\text{s}^{-1}$  for wheat and barley, respectively; Müller *et al.* 2005; Braune *et al.* 2009) and those of C3 trees and vines (e.g. about 30 and  $38 \mu\text{mol.g N}^{-1}.\text{s}^{-1}$  in walnut tree and grapevine, respectively; Le Roux *et al.* 1999; Prieto *et al.* 2012). This positioning was consistent with other productive grassland species (e.g. 36 to  $50 \mu\text{mol.g N}^{-1}.\text{s}^{-1}$  reported for cocksfoot and red clover; Wohlfahrt *et al.* 1998).

### **Performance of the leaf gas exchange model at the leaf and whole canopy levels**

The gas exchange model correctly estimated daily cumulated values of net assimilation ( $A$ ) and transpiration ( $E$ ) at the leaf level and their diurnal patterns. Model errors, however, were greater with respect to transpiration. This might partly be related to the absence of a direct relationship between the leaf nitrogen and stomatal conductance parameters considered in the model. Indeed, the scaling parameter  $a_1$  has been shown to increase for leaves with a very low N concentration (Braune *et al.* 2009). In our case however, discrepancies in  $E$  mainly concerned the top leaves under high N availability. An alternative explanation might be a less robust parameterization of the stomatal conductance model. Leaves from a single experiment were used, covering a more limited range of environmental conditions than that encountered in the validation dataset (in terms of VPD in particular). Future work will incorporate the response to water deficit in the model, which should enable the refinement of this parameterisation.

When up-scaled at the whole canopy level, the gas exchange model coupled with the N distribution model displayed interesting properties regarding the response to N availability. The relationship between nitrogen and the assimilation rate switched from a linear function at the leaf level to a saturating function at the whole canopy

scale. Above a certain threshold, the model predicted that an increase in N did not result in increased canopy assimilation. Such a point had previously been reported during numerous experiments comparing a broad range of N fertilisation rates (Justes *et al.* 1994; Lemaire and Gastal, 1997), and it defines the critical N concentration on which NNI calculations are based. Remarkably, such behaviour emerged from our canopy-integrated model. Furthermore, the NNI value corresponding to this transition happened to be very close to 1 during sunny and moderately cloudy days. This complied with the theoretical definition of NNI, which states that a value of 1 corresponds to the critical N concentration. The threshold was predicted to be lower during cloudy days. However, as in practice the critical N concentrations are determined from cumulated values of biomass production, critical N is likely to be primarily driven by sunny days (accounting for most biomass accumulation) rather than cloudy days.

No direct measurements were carried out to quantitatively assess the gas exchange model on alfalfa canopies. However, the simulated plateau values were consistent with previous studies measuring the daily net carbon exchange in closed canopies of alfalfa under non limiting N. For instance, Heichel *et al.* (1988) reported net rates of 1.17, 0.81 and 0.45 mol CO<sub>2</sub>.m<sup>-2</sup>.day<sup>-1</sup> in a 2.7 LAI canopy during days with average PPFD values of 1100, 700 and 400 μmol.m<sup>-2</sup>.s<sup>-1</sup>, respectively (corresponding roughly to the light conditions prevailing during the three days shown in Fig 6). Similarly, Woodward and Sheehy (1979) reported rates ranging from 0.27 to 1.39 mol CO<sub>2</sub>.m<sup>-2</sup>.day<sup>-1</sup> after canopy closure during a spring regrowth. As in these two studies, the model outputs concerned the net aboveground carbon exchange of N fertilized alfalfa (with presumably limited N fixation). Allocation to the root system and to the respiration of roots and nodules needs to be implemented in future

versions of the model in order to account for a potential cost of N fixation in terms of the carbon balance at the canopy level (Gosse *et al.* 1986). Root growth and respiration indeed represent a substantial share of carbohydrate use in fixing alfalfa (Thomas and Hill 1937; Layzell *et al.* 1981).

Fewer references were available to compare the response induced by nitrogen limitation. We thus compared the simulation results with those relative to regular non fixing C3 plants and found a relative reduction in the net assimilation rate measured by Belanger *et al.* (1992) in a range of tall fescue canopies (Fig. 6). The simulated response curve of dense canopies on sunny and moderately cloudy days appeared to be particularly close to the measured curve, suggesting a good ability of the up-scaled leaf model to capture the N stress response of a whole canopy.

## CONCLUSIONS

Overall, the set of empirical relationships introduced in this paper to distribute leaf N was simple and effective at predicting leaf nitrogen concentration in response to light and plant N status. The leaf gas exchange model proved accurate and produced consistent predictions in terms of whole canopy assimilation under contrasting soil N availability scenarios. Even if the genericity of the coupled model still needs to be challenged in a broader range of species, this work constitutes a further step toward models which can bridge local acclimation to light with N acquisition and global N nutrition status, without presuming an optimal carbon gain or N distribution. Such a model rely on parameters which can all be directly measured and may help us to infer and better understand the differences in nitrogen use efficiency observed between species or genotypes.

## SUPPORTING INFORMATION

The following [SUPPORTING INFORMATION] is available in the online version of this article:

**Figure Sup. A:** Impact of leaf ageing and relative leaf irradiance on the specific leaf area and leaf nitrogen concentration per unit area in alfalfa.

**Figure Sup. B:** Relationship between leaf age and the residuals of the fit of Eq.1 to leaf nitrogen concentration.

**Figure Sup. C:** Variations in leaf nitrogen concentrations predicted for alfalfa as a function of the nitrogen nutrition index of the canopy and the relative leaf irradiance.

**Table Sup. A:** Comparison of two and three parameter models to account for distribution leaf nitrogen concentrations with respect to relative irradiance in alfalfa.

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## CONTRIBUTIONS BY THE AUTHORS

G.L., E.L. and J.A.P contributed to model development. G.L. and E.F. designed the experiments and conducted measurements. They performed data analyses with the help of S.Z. All of the authors contributed to writing the manuscript.

## CONFLICT OF INTEREST STATEMENT

None declared.

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**Table 1:** Canopy characteristics and nitrogen distribution parameters determined during the different experiments and nitrogen treatments studied. Parameters were obtained by fitting Eq. 1 to the nitrogen content measured in leaves separated in 10-cm-strata. Standard errors are indicated into brackets.

Experiment	Nutrient solution	N acquisition mode	LAI	$N_{up}$	$k_n$	$r^2$
1	N+	Assimilation	5.1	2.12 (0.075)	0.23 (0.013)	0.95
1	N+	Assimilation	2.6	2.31 (0.106)	0.24 (0.020)	0.96
2	N+	Assimilation	8.1	2.59 (0.123)	0.21 (0.021)	0.96
1	N-	Assimilation	1.	1.09 (0.259)	0.20 (0.088)	0.64
1	N-	Assimilation	0.7	0.76 (0.130)	0.09 (0.312)	0.20
1	N-	Assimilation+Fixation	3.3	1.77 (0.134)	0.15 (0.033)	0.87
1	N-	Assimilation+Fixation	2.1	1.69 (0.072)	0.29 (0.031)	0.94
3	N0	Fixation	2.0	1.71 (0.181)	0.24 (0.054)	0.75

## Appendix A1: Equations for the photosynthesis and stomatal conductance models

Equation	Description	N°
PHOTOSYNTHESIS MODEL		
$A = V_c - 0.5 \cdot V_o - R_d = V_c \cdot \left(1 - \frac{\Gamma^*}{C_i}\right) - R_d$	Net photosynthetic rate ( $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ )	(A1)
$V_c = \text{Min} \{A_c, A_j, A_p\}$	Carboxylation rate ( $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ )	(A2)
$A_c = \frac{V_{\text{max}} \cdot C_i}{C_i + K_c \cdot \left(1 + \frac{O}{K_o}\right)}$	RUBISCO-limited photosynthetic rate	(A3)
$A_j = \frac{J \cdot C_i}{4 \cdot C_i + 8 \cdot \Gamma^*}$	RuBP regeneration-limited photosynthetic rate	(A4)
$A_p = \frac{3 \cdot \text{TPU}}{\left(1 - \frac{\Gamma^*}{C_i}\right)}$	TPU-limited photosynthetic rate	(A5)
$J = \frac{\alpha \cdot \text{PPFD}}{\sqrt{1 + \frac{\alpha^2 \cdot \text{PPFD}^2}{J_{\text{max}}^2}}}$	Electron transport rate, dependence on the radiance level	(A6)
$P = e^{\left(\frac{c - \frac{\Delta H_a}{R \cdot T_k}}{R \cdot T_k}\right)}$	Arrhenius function, temperature dependence for $K_c$ , $K_o$ , $\Gamma^*$ and $R_d$	(A7)
$P = \frac{e^{\left(\frac{c - \frac{\Delta H_a}{R \cdot T_k}}{R \cdot T_k}\right)}}{1 + e^{\left(\frac{\Delta S \cdot T_k - \frac{\Delta H_d}{R \cdot T_k}}{R \cdot T_k}\right)}}$	Arrhenius function, temperature dependence for $V_{\text{max}}$ , $J_{\text{max}}$ and TPU	(A8)
$P^{25} = S_{N_a} \cdot N_a - b$	Nitrogen dependence function for $V_{\text{max}}$ , $J_{\text{max}}$ , TPU and $R_d$ at 25 C°	(A9)
STOMATAL CONDUCTANCE MODEL		
$g_s = g_o + \frac{a_1 \cdot A}{\left(1 + \frac{\text{VPD}}{D_o}\right) \cdot (C_s - \Gamma)}$	Stomatal conductance	(A10)
$C_s = C_a - A \cdot \frac{1.37}{g_b}$	CO <sub>2</sub> partial pressure at the leaf surface	(A11)
$C_i = C_a - A \cdot \left(\frac{1.6}{g_s} + \frac{1.37}{g_b}\right)$	C <sub>i</sub> value by coupling A and g <sub>s</sub>	(A12)

## Appendix A2: Symbols, values and units of different parameters, variables and constants used in the photosynthetic and stomatal conductance models

Symbol	Value	Unit	Description
PHOTOSYNTHESIS MODEL			
$\alpha$	0.20 <sup>a</sup>	$\mu\text{mol CO}_2 \mu\text{mol photon}^{-1}$	Photochemical efficiency or initial quantum yield
$\Gamma^*$	-	Pa	Compensation point for $\text{CO}_2$ in the absence of mitochondrial respiration
A	-	$\mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$	Net photosynthetic rate
$A_c$	-	$\mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$	RUBISCO-limited photosynthetic rate
$A_j$	-	$\mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$	Electron transport rate-limited photosynthetic rate
$A_p$	-	$\mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$	Triose phosphate utilisation-limited photosynthetic rate
c	-	-	Scaling constant
$C_a$	-	Pa	Ambient $\text{CO}_2$ partial pressure
$C_i$	-	Pa	Intercellular $\text{CO}_2$ partial pressure
$\Delta H_a$	-	$\text{KJ mol}^{-1}$	Enthalpy of activation
$\Delta H_d$	200 <sup>b</sup>	$\text{KJ mol}^{-1}$	Enthalpy of deactivation
$K_c$	-	Pa	Michaelis-Menten constant of RUBISCO for $\text{CO}_2$
$K_o$	-	KPa	Michaelis-Menten constant of RUBISCO for $\text{O}_2$
J	-	$\mu\text{mol electron m}^{-2} \text{s}^{-1}$	Electron transport rate
$J_{\text{max}}$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Maximum electron transport rate
$N_a$	-	$\text{g m}^{-2}$	Area based nitrogen content
$N_{\text{amin}}$	-	$\text{g m}^{-2}$	Minimum value of $N_a$ at which $P_{25} \rightarrow 0$
<b>O</b>	21	KPa	Oxygen partial pressure
$P^{25}$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Value of $V_{\text{cmax}}$ , $J_{\text{max}}$ , TPU or $R_d$ at 25 C°
PPFD	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Photosynthetic photon flux density
R	0.00831	$\text{KJ mol}^{-1} \text{K}^{-1}$	Universal gas constant for perfect gases
$R_d$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Mitochondrial respiration in light
$\Delta S$	0.635 <sup>b</sup>	$\text{KJ mol}^{-1}$	Entropy term
$S_{N_a}$	-	$\mu\text{mol g}^{-1} \text{s}^{-1}$	Slope of the relationship between $N_a$ and $V_{\text{cmax}}$ , $J_{\text{max}}$ , TPU or $R_d$
$T_{\text{leaf}}$	-	°C	Leaf temperature in degrees Celsius
$T_k$	-	Kelvin degrees	Leaf temperature in Kelvins
TPU	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Triose phosphate utilisation rate
$V_c$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Carboxylation rate
$V_o$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Oxygenation rate
$V_{\text{cmax}}$	-	$\mu\text{mol m}^{-2} \text{s}^{-1}$	Maximum rate of RUBISCO carboxylation

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**STOMATAL CONDUCTANCE MODEL**

$C_s$	-	Pa	CO <sub>2</sub> partial pressure at the leaf surface
$g_b$	2.357 <sup>c</sup>	mol m <sup>-2</sup> s <sup>-1</sup>	Boundary layer conductance
$g_s$	-	mmol m <sup>-2</sup> s <sup>-1</sup>	Stomatal conductance
$g_o$	0.020	mmol m <sup>-2</sup> s <sup>-1</sup>	Residual stomatal conductance when A→ 0
VPD	-	KPa	Water vapour pressure deficit
$D_o$	2.86	KPa	Empirical factor assessing stomata sensitivity to VPD
$a_1$	12.5	-	Empirical stomatal conductance factor

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<sup>a</sup> Values taken from Schultz 2003

<sup>b</sup> Values taken from Harley et al. 1992

<sup>c</sup> Constant used for measurement in the leaf-chamber of the LcPro (ADC Lcpro, BioScientific Ltd, Hoddesdon, Herts, UK)

**Appendix A3.** Values of  $c$  (scaling constant), enthalpies of activation ( $\Delta H_a$ ) describing the temperature response for parameters of the photosynthesis model

Parameter	Value at 25°C	$c$	$\Delta H_a$ (kJ mol <sup>-1</sup> )
$V_{cmax}$	-	26.35 <sup>a</sup>	65.33 <sup>a</sup>
$J_{max}$	-	17.7 <sup>b</sup>	43.9 <sup>b</sup>
TPU	-	21.46 <sup>c</sup>	53.1 <sup>c</sup>
$R_d$	-	18.72 <sup>a</sup>	46.39 <sup>a</sup>
$\Gamma^*$	42.75 <sup>a</sup>	19.02 <sup>a</sup>	37.83 <sup>a</sup>
$K_c$	404.9 <sup>a</sup>	38.05 <sup>a</sup>	79.43 <sup>a</sup>
$K_o$	278.4 <sup>a</sup>	20.30 <sup>a</sup>	36.38 <sup>a</sup>

<sup>a</sup> Values taken from Bernacchi et al. 2001

<sup>b</sup> Values taken from Bernacchi et al. 2003

<sup>c</sup> Values taken from Harley et al. 1992

## FIGURE LEGENDS

**Figure 1:** Relationships between (a) canopy nitrogen nutrition index and leaf nitrogen concentration at the top of the canopy ( $N_{up} = 2.15 \cdot \text{NNI} + 0.02$ ,  $r^2=0.91$ ), and (b) relative leaf irradiance and leaf nitrogen concentration relative to the leaf nitrogen concentration at the top of the canopy ( $k_n=0.247$ ;  $r^2=0.73$ ).

**Figure 2:** Relationship between the values of photosynthetic parameters at a leaf temperature of 25°C and leaf nitrogen concentration ( $N_a$ ) across the different experiments and nitrogen treatments studied. Linear relationships were found for  $V_{cmax}^{25}$  (a,  $V_{cmax}^{25}=53.03 \cdot N_a - 14.74$ ,  $r^2=0.86$ ),  $J_{max}^{25}$  (b,  $J_{max}^{25}=90.91 \cdot N_a - 13.83$ ,  $r^2=0.83$ ),  $TPU^{25}$  (c,  $TPU^{25}=6.72 \cdot N_a - 0.72$ ,  $r^2=0.78$ ) and  $R_d^{25}$  (d,  $R_d^{25}=0.69 \cdot N_a - 0.005$ ,  $r^2=0.77$ ), respectively.

**Figure 3:** Relationship between leaf nitrogen concentrations ( $N_a$ ) observed at various positions within the canopy and the corresponding values simulated. Open and filled symbols indicate canopies grown with N- and N+ nutrient solutions, respectively.

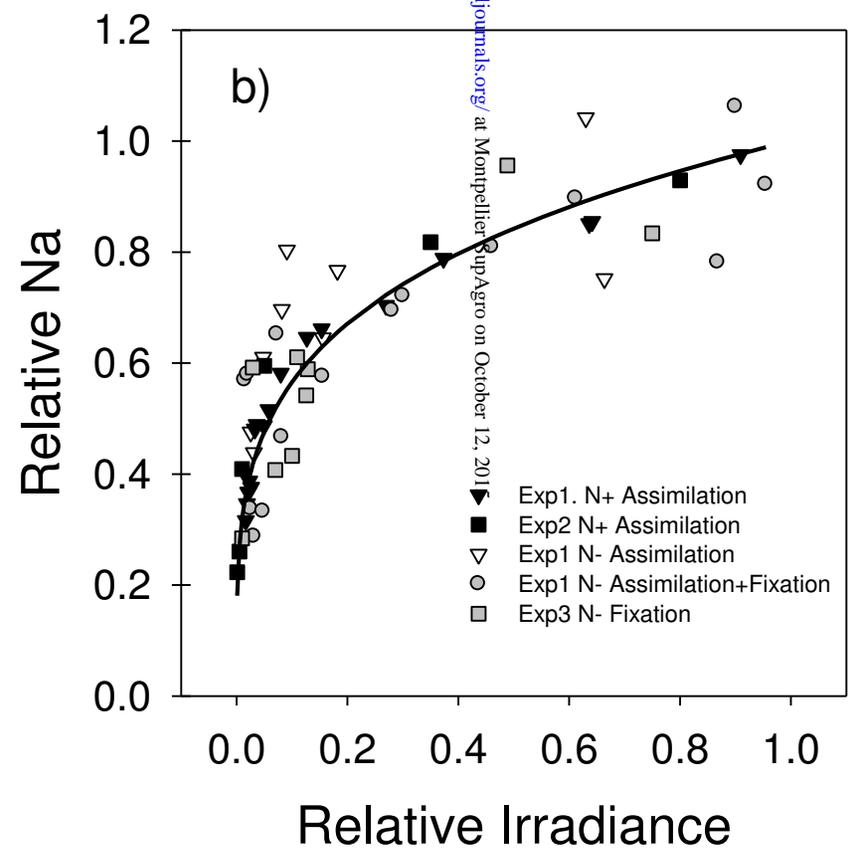
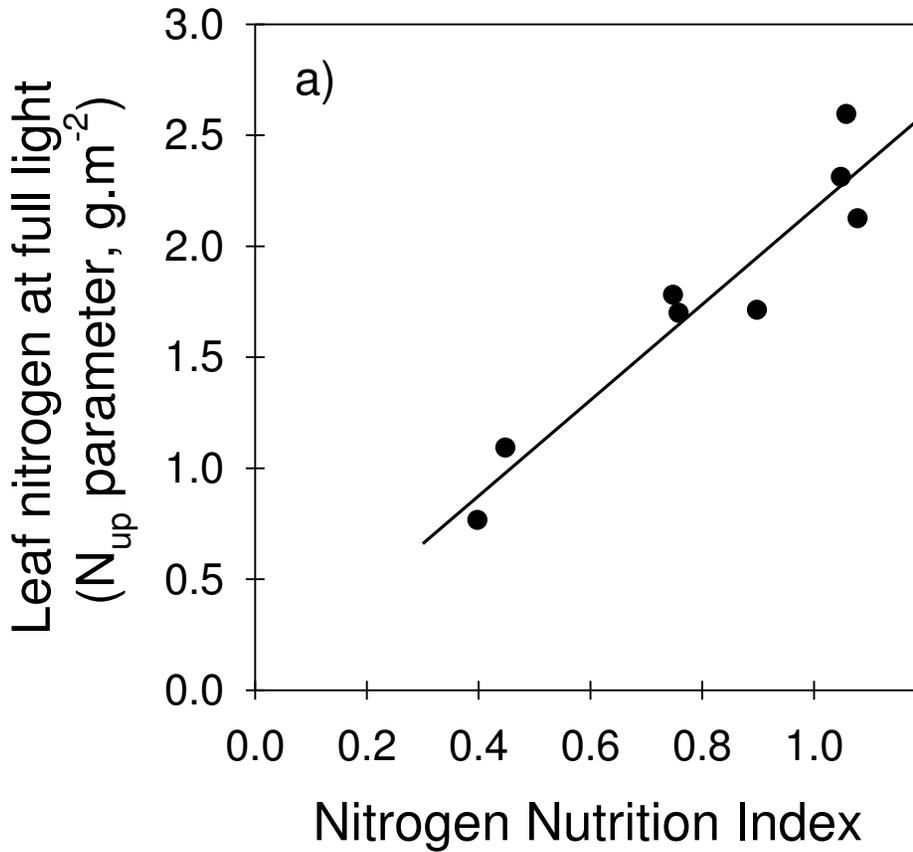
**Figure 4:** Measured instantaneous photosynthetic photon flux density at the leaf level (a–c), leaf temperature (black) and vapour pressure deficit (grey, d–f), and the measured (open circles) and predicted (solid line) net photosynthesis (g–i), and transpiration rates (j–l), for three leaves in Lusignan in 2011. DOY, day of the year.

**Figure 5:** Comparison of measured and predicted values of daily net photosynthesis (a), and transpiration rates (b). Dashed lines: regressions between measured and

predicted values; solid lines: 1:1 relationships. Open and filled symbols indicate leaves from canopies grown with N- and N+ nutrient solutions, respectively.

**Figure 6:** Simulations for three contrasting days of whole canopy net assimilation in response to changes in the canopy nitrogen nutrition index (NNI) and leaf area index (LAI) (a-c) and their corresponding responses normalised by the assimilation rate at an NNI value of 1 (d-f). Grey circles in the lower panel represent the relative reduction in radiation use efficiency measured by Bélanger *et al.* (1992) in response to NNI.

Figure 1



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Figure 2

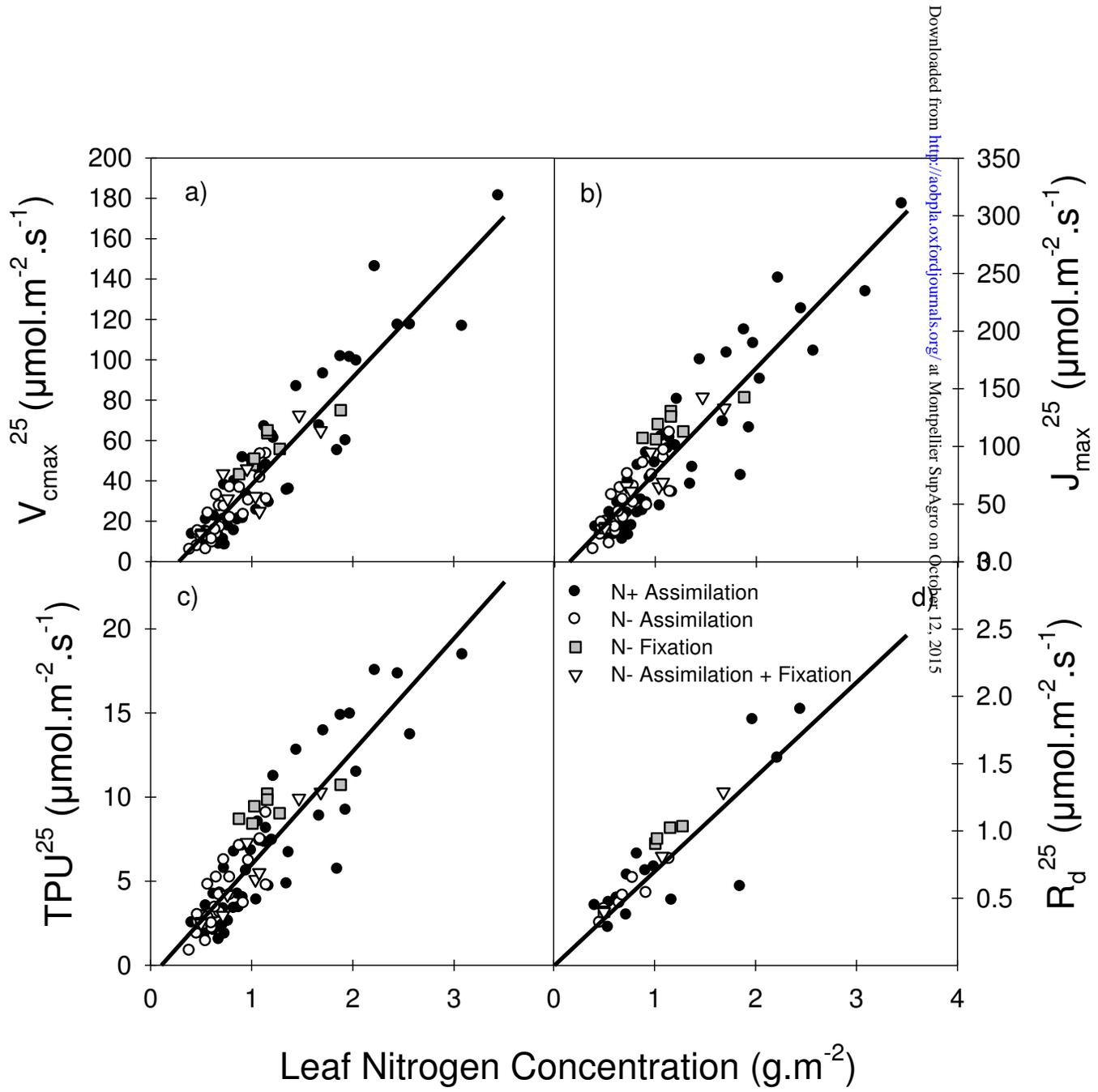


Figure 3

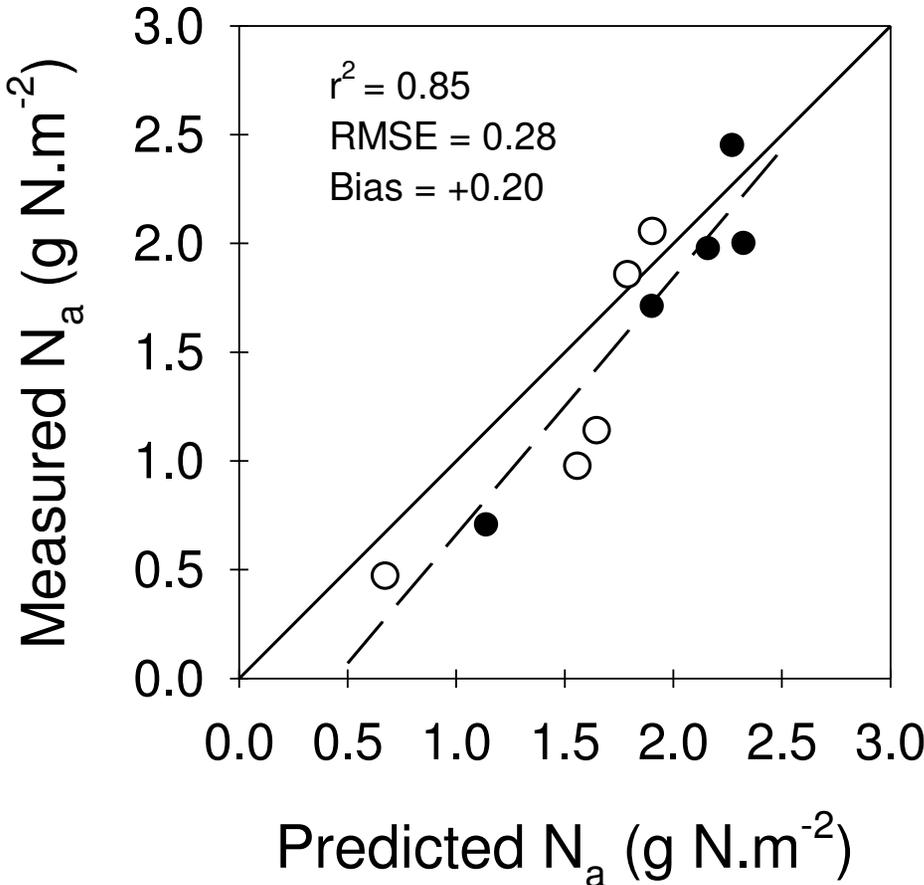


Figure 4

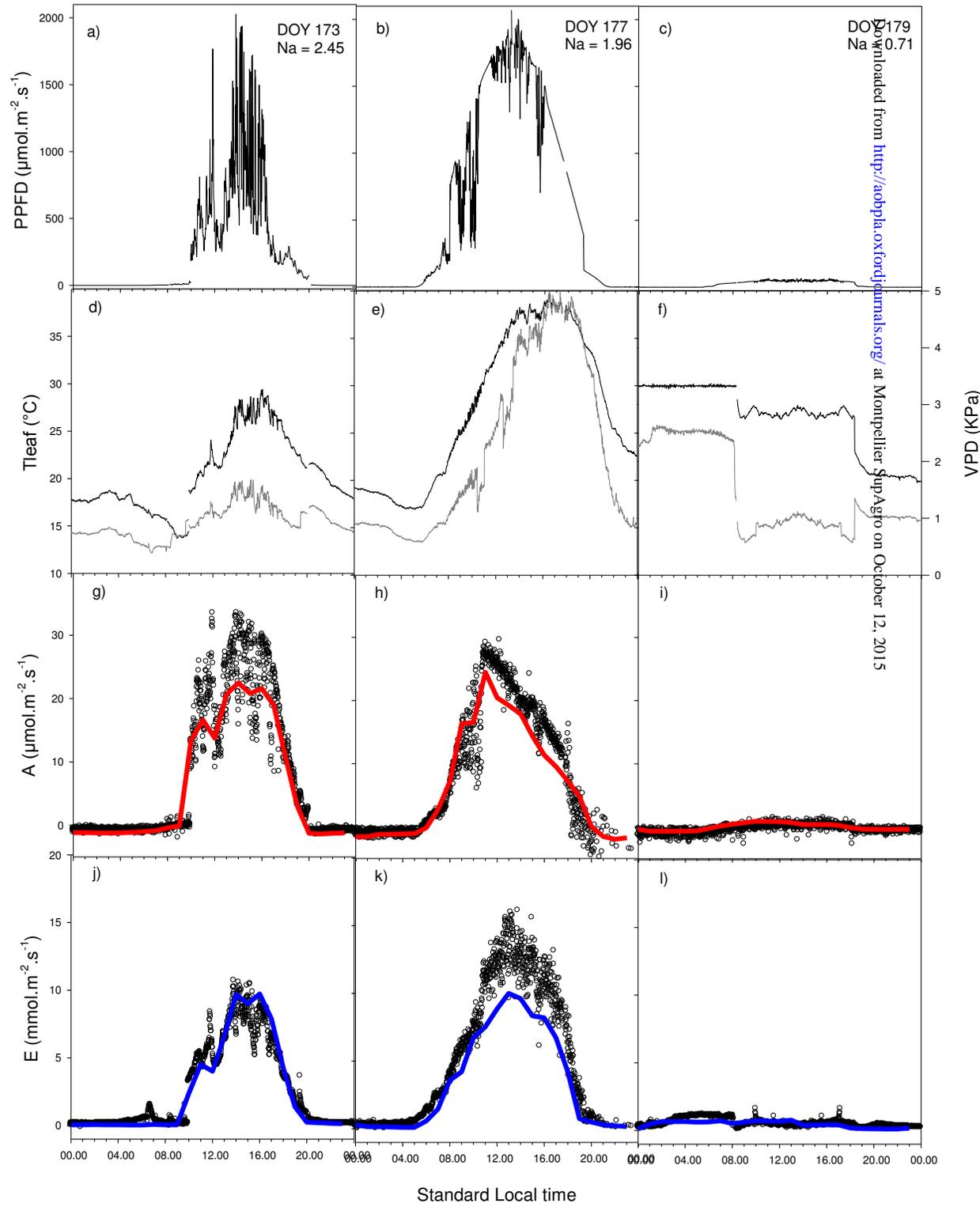
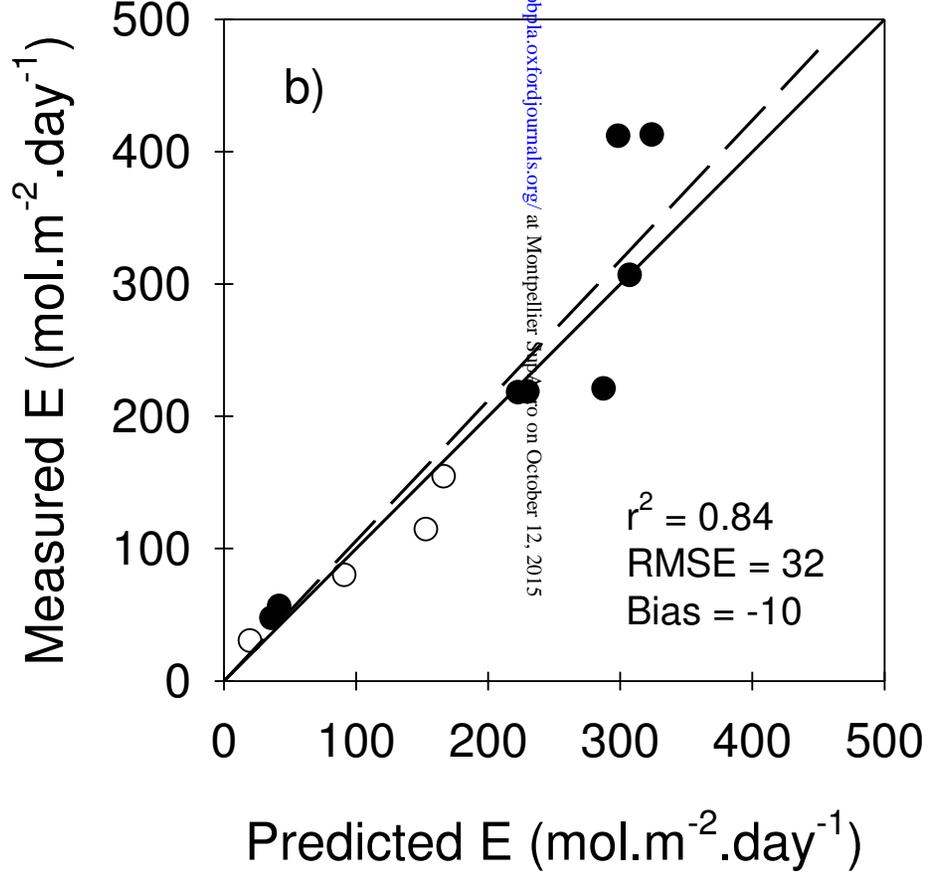
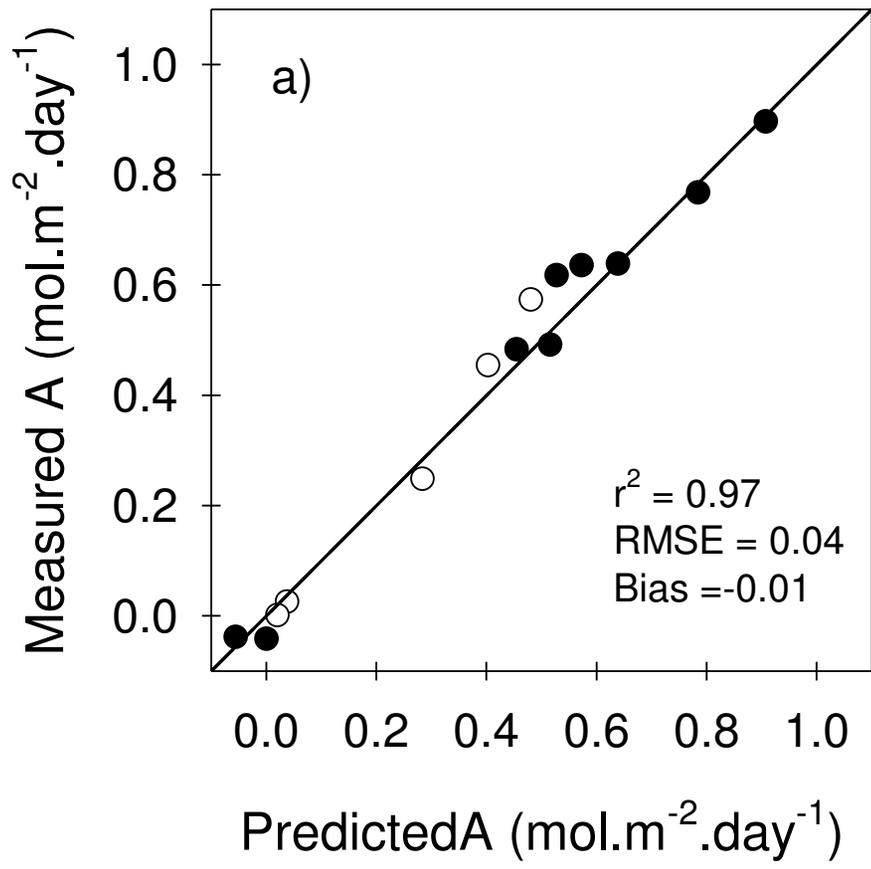
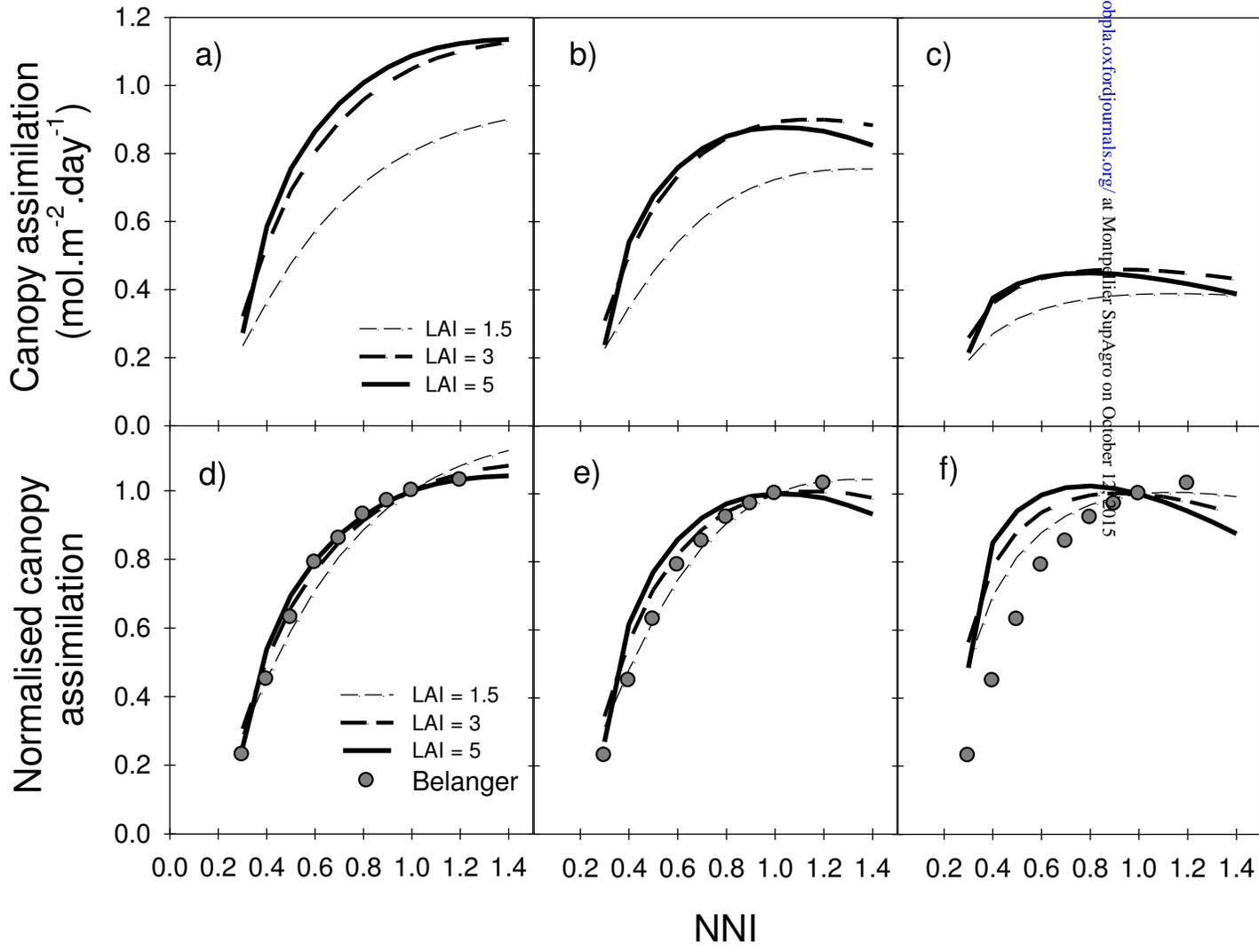


Figure 5



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Figure 6



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