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RESEARCH ARTICLE

Assessment of Sublethal and Transgenerational Effects of Pirimicarb on the Wheat Aphids *Rhopalosiphum padi* and *Sitobion avenae*

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Abstract

The wheat aphids, *Rhopalosiphum padi* (Linnaeus) and *Sitobion avenae* (Fabricius), are key pests on wheat crops worldwide. Management practices rely primarily on insecticides. The pirimicarb (carbamate) is used extensively as an effective insecticide to control these two aphids. In addition to the mortality caused by pirimicarb, various sublethal effects may occur in aphids when exposed to low lethal or sublethal doses. Understanding the general effect of pirimicarb on aphids could help increasing rational use of this insecticide. Under laboratory conditions, we assessed the sublethal effects of a low lethal concentration of pirimicarb (LC₂₅) on biological traits and acetylcholinesterase (AChE) activity of *R. padi* and *S. avenae*. Both direct and transgenerational effects, i.e. on parent and the F1 generations were assessed, respectively. We found that *R. padi* and *S. avenae* responded differentially to the LC₂₅ of pirimicarb. The parent generation of *R. padi* showed a 39% decrease in fecundity and multiple transgenerational effects were observed in the F1 generation; overall juvenile development, reproductive period, adult longevity and lifespan were longer than those of the control group. By contrast, LC₂₅ of pirimicarb showed almost no effects on *S. avenae* biological traits in both the parent and F1 generations; only the pre-reproductive duration was reduced in F1 generations. Demographic parameter estimates (e.g. r_m) showed similar trend, i.e. significant negative effect on *R. padi* population growth and no effect on *S. avenae*. However, AChE activity decreased in both *R. padi* and *S. avenae* treated by the LC₂₅ of pirimicarb. We demonstrated sublethal and transgenerational effects of pirimicarb in the two wheat aphid species; it hinted at the importance of considering sublethal effects (including hormesis) of pirimicarb for optimizing Integrated Pest Management (IPM) of wheat aphids.

Introduction

The bird cherry-oat aphid, *Rhopalosiphum padi* (Linnaeus) and grain aphid, *Sitobion avenae* (Fabricius) are destructive pests of wheat, sorghum and other small grain crops worldwide [1–3]. These two species usually coexist at the late stage of wheat growth in China [4], causing direct damage through feeding. Moreover, they also serve as vectors of barley yellow dwarf virus [5]. Outbreaks of these two aphids lead to severe yield losses [6].

In China, insecticides application is primary strategy to control aphids. Organophosphate and carbamate insecticides are extensively used for wheat aphid control [7]. Carbamate insecticides are esters of carbamic acid, and they inhibit hydrolysis of neurotransmitter acetylcholine (ACh) by acetylcholinesterase (AChE) resulting in the disruption of normal nervous system functions. Pirimicarb is a selective carbamate aphicide and mainly used for management of wheat aphid [8, 9]; it has been used for the control of wheat aphids since the 1990s in China [10].

In addition to their lethal effects, insecticides may also impair various key biological traits of aphids in the exposed insects through sublethal effects [11]. Sublethal effects are defined as physiological and/or behavioral effects on individuals that survived from exposure to a pesticide at low or sublethal concentrations/doses [11]. Sublethal effects of insecticides could affect population dynamics through impaired behavior and physiological traits, such as life span, development rates, fertility and fecundity [12–16]. Studies have demonstrated that exposure to lethal or sublethal doses/concentrations of pesticides could reduce insect longevity and fecundity [17–21]. Therefore, assessment of sublethal effects is crucial to acquiring knowledge on global insecticide efficacy on insect pests, as well as on possible selectivity towards non-target organisms [22, 23]. In addition, sublethal insecticide exposure could be assessed using specific biomarkers e.g. AChE and glutathione S-transferase (GST) activity [24–27]. Numerous studies have focused on insect AChE because it is target of organophosphates and carbamates insecticides, which are two major pesticide classes used worldwide for pest management [28].

In China, *R. padi* and *S. avenae* are susceptible to pirimicarb according to established susceptible base line; thus pirimicarb is used efficiently to control these aphids [29, 30]. However, the extensive use of this insecticide and possible inappropriate and/or sub-optimal applications may promote the development of resistances as reported for many other pest-pesticide couples [31–34]. In addition, abiotic disturbances are known to affect ecological interactions among species [35]; an insecticide-mediated shift in ecological dominance between the wheat aphids might occur through the persistence of low pesticide concentrations in fields. Understanding the general effect of pirimicarb on the aphids would help increase rational use of this insecticide in wheat and also increase its sustainable use. In this context, we investigated the direct effect of pirimicarb on wheat aphids under a low lethal concentration (LC_{25}) on exposed individuals i.e. the parent generation, as well as the transgenerational effect on the subsequent generation (F1) [33]. Assessment of such effects could be highly relevant for *R. padi* and *S. avenae* that have high reproductive rates and short life cycles. In addition, the activity of AChE was characterized to complement the assessment of effects of pirimicarb on wheat aphids. The LC_{25} was used because such low concentration may occur in fields when insecticides degrade following initial application ([e.g. see [36]).

Materials and Methods

Insects

Colonies of *R. padi* and *S. avenae* were initiated from apterous aphid clones collected from a wheat field of the Agricultural Experiment Station, China Agricultural University (N40°03', E116°28'). Colonies of the two aphid species were maintained in the laboratory without

insecticides exposure since May 2005. They were reared on wheat seedlings under laboratory conditions (18–25°C, relative humidity $60 \pm 10\%$ and a photoperiod of 17:7 L:D) according to Lu and Gao (2007)[4].

Insecticides and chemicals

Pirimicarb (95% technical grade) was obtained from Wuxi Ruize Chemical Co. Ltd, China. For assessment of the AChE activity, S-Acetylthiocholine iodide (ATChI) and 5, 5'-Dithiobis (2-nitrobenzoic acid) (DTNB) were purchased from Sigma (St. Louis, MO, USA). Bovine serum albumin (BSA) was purchased from Tongzheng Biological Company (Beijing, China) and was used as standard for assessment of AChE activity.

Insecticide bioassays

Insecticide toxicity was assayed following the method of exposure to pesticide residues in glass tubes (diameter: 2 cm, length: 5.2 cm) [29]. Pirimicarb was diluted into six different concentrations in analytical grade acetone. An aliquot of 200 μ l insecticide-acetone solution was applied to each tube. These tubes were immediately rotated using a micro-rotator (American Wheaton Company) until solutions were dried. Twenty aphids were treated for each concentration with three replications. Controls were treated with acetone only. The aphids were then reared under laboratory conditions and mortality was checked after three hours. After this 3-h period the mortality was assessed; aphids that did not move legs when touched with a fine brush (no reflex movement) were considered dead [37]. The results of testing the six pirimicarb concentrations were used to estimate the LC_{25} for the following experiments on sublethal effects of pirimicarb.

Impact of LC_{25} pirimicarb on biological traits in parent generation of the aphids

The assays were conducted on wheat seedling stage using life table construction. Wheat seedlings were grown in plastic dishes (diameter: 4 cm) with a slice of wadding paper-filter put at the bottom of plastic dish to keep humidity under laboratory conditions. Pirimicarb was prepared in acetone and diluted to the LC_{25} with distilled water containing 0.05% (v/v) Triton X-100. The adult aphids fed on wheat seedlings that were treated with LC_{25} of pirimicarb for 10 s following the method of leaf dipping [37]. Mortality was calculated at 24 h post treatment and the survivals were gently moved into the new wheat seedlings without any insecticide. Survival aphids were kept in plastic dish individually and each plastic dish was sealed with fitted ventilated plastic tubes in order to prevent aphid escape. The number of progeny nymphs was recorded and then removed daily until the adult aphid was dead. Wheat seedlings were replaced every week during the experiment period. Three replicates were carried out with twenty-five aphids for each replicate. The aphids in control group fed on the seedlings treated with distilled water containing 0.05% (v/v) Triton X-100.

Transgenerational sublethal effects of pirimicarb in F1 generation of the aphids

Using the same experimental protocol as described above, we exposed adult aphids to LC_{25} pirimicarb. Five survival adult aphids were transferred into the new wheat seedling after treatment with LC_{25} of pirimicarb at 24 h post treatment, and then maintained in the plastic dishes as described above. At 24 h after the transfer, all of the aphids were removed leaving only one neonate nymph in the wheat seedling. Thereafter, we recorded (i) the duration of development, (ii) the numbers of survivors at different development stages, and (iii) the numbers of nymphs

produced per aphid every day. These recorded data were used to estimate (i) duration of nymphs or generation (in days), (ii) pre-reproductive or reproductive period (in days), (iii) longevity of adults (in days), and (iv) total number of nymphs laid per aphid during its life (cited as fecundity hereafter). During the reproductive period, the newborn nymphs were counted and then removed daily. Data from alate aphids were not included in the analysis. Three replicates were carried out with twenty-five aphids for each replicate. The aphids in control group fed on the seedlings treated with distilled water containing 0.05% (v/v) Triton X-100.

The life-table parameters, which include the (i) net reproductive rate ($R_0 = \sum l_x m_x$): the population growth rate per generation with regard to the number of female offspring produced per female, (ii) mean generation time ($T = \sum x l_x m_x / R_0$): the average interval separating births from one generation to the next, (iii) the intrinsic rate of increase ($r_m = \ln(R_0)/T$): the maximum exponential increase rate in a population growing within defined physical conditions and (iv) population doubling time ($Dt = \ln(2)/r_m$): the time required by a population, when growing exponentially, to double when it increase as a given r_m were calculated as follows for control and treatment. The age-specific survival rate (l_x) is the proportion of individuals in the initial cohort alive at age x time (days), and the age-specific fecundity (m_x) is the mean number of female progeny produced per female alive at the age interval, x days.

Determination of AChE activity

AChE activity was assessed with ATChI as substrates according to the method of Ellman et al. (1961) [38] and Gao (1987) [39]. The enzyme preparation was replicated three times and each replicate was prepared by homogenizing 60 wingless adult aphids. They were placed in 1 ml ice-cold phosphate buffer (0.04 M, pH 7.5, containing 0.1% (v/v) Triton X-100). The homogenates were centrifuged at 4°C, 10, 800 g (Eppendorf centrifuge 5417R, Germany) for 30 min, and the supernatant was used for subsequent AChE activity assay. The assay mixture contain 100 µl of enzyme preparation, 100 µl of substrate solution (5 mM). The reaction was stopped by the addition of 3.6 ml DNTB (0.125 mM) with 40% ethanol after incubating at 30°C for 15 min. The optical density (OD) at 412 nm was measured using spectrophotometer (Lambda Bio 40, PE, USA). The control samples contained no enzyme during the incubation. AChE activity was based on protein content which was determined by the method of Bradford (1976) [40], using BSA as the standard. AChE activities were expressed as nmol ATCh hydrolyzed per min per mg protein using the extinction coefficient of $1.36 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$.

Data analysis

Datasets were first tested for normality and homogeneity of variance using the Kolmogorov-Smirnov and the Cochran tests respectively and were transformed if necessary. The LC_{25} of pirimicarb for *R. padi* and *S. avenae* was determined using a log-probit model [41]. The life table and population growth datasets were subjected to t-tests to compare pirimicarb-exposed groups to respective control groups (GraphPad Software, San Diego, CA, USA).

Results

LC_{25} of pirimicarb

The LC_{25} of pirimicarb were $0.29 \mu\text{g mL}^{-1}$ and $0.60 \mu\text{g mL}^{-1}$ for *R. padi* and *S. avenae*, respectively (Table 1). When these concentrations were used for subsequent assessment of sublethal effects, the corrected mortality were 25.8% and 23.2% for *R. padi* and *S. avenae*, respectively.

Table 1. LC₂₅ values and mortality for pirimicarb toxicity on *R. padi* and *S. avenae* after 3 h exposure to treated glass tubes.

Insect	Slope ± SE ^a	LC ₅₀ ^b	LC ₂₅ ^b	χ ^{2c}	P values	Mortality ^d (%)
<i>R. padi</i>	1.43 ± 0.27	0.53(0.40–0.64)	0.29 (0.18–0.39)	1.29	0.206	25.80
<i>S. avenae</i>	1.57 ± 0.38	1.36(0.78–1.93)	0.60 (0.25–9.49)	2.44	0.584	23.22

^a SE = standard error.

^b Expressed in µg mL⁻¹; 95% CI of LC₂₅ are given in bracket.

^c Chi-square testing linearity of dose-mortality responses.

^d LC₂₅ induced mortality.

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Longevity and fecundity of parent generation of *R. padi* and *S. avenae*

The longevity of *R. padi* and *S. avenae* from parent generation was not affected by the LC₂₅ of pirimicarb when compared to their respective control (Table 2, $P = 0.847$ and $P = 0.972$, respectively). The fecundity of *R. padi* exposed to pirimicarb was significantly lower than that in the control group ($P < 0.001$) (Table 2). However, no significant difference was found in the fecundity of *S. avenae* exposed to pirimicarb ($P = 0.694$) (Table 2).

Fecundity of *R. padi* and *S. avenae* in F1 generation

There was no significant impact of pirimicarb on total fecundity in F1 generation of *R. padi* ($P = 0.757$), whereas daily fecundity significantly decreased after exposed to pirimicarb ($P < 0.001$) (Table 3). There was no significant impact of pirimicarb on total fecundity ($P = 0.690$) and daily fecundity ($P = 0.745$) in F1 generation of *S. avenae* (Table 3).

Development of *R. padi* and *S. avenae* in F1 generation

The effects of pirimicarb exposure on development in F1 generation of *R. padi* were reported in Fig 1A. The development time of the first and second instar of *R. padi* were not affected, while the development time of the third instar was significantly longer (1.38 ± 0.11 days) after pirimicarb exposure than in the control group (1.04 ± 0.07 days) ($t = 2.599$, $P = 0.013$). The development time of the fourth instar of *R. padi* was significantly longer (1.83 ± 0.08) after exposed to pirimicarb than that of control (1.38 ± 0.09 days) ($t = 3.817$, $P < 0.001$). The overall juvenile development of *R. padi* was significantly longer (6.42 ± 0.18 days) in the treatment than that of the control (5.50 ± 0.14 days) ($t = 3.943$, $P < 0.001$). In general, the duration of generation of *R. padi* significantly increased (7.21 ± 0.23 days) after exposure to pirimicarb compared to the control (6.04 ± 0.11 days) ($t = 4.639$, $P < 0.001$). Exposure to LC₂₅ of pirimicarb significantly increased the reproductive period of *R. padi* ($t = 3.621$, $P < 0.001$). The adult longevity of *R. padi* was also significantly longer (23.54 ± 2.10 days) after exposed to pirimicarb than that of the control (17.38 ± 1.54 days) ($t = 2.370$, $P = 0.022$). The lifespan of *R. padi* was

Table 2. Effect of LC₂₅ of pirimicarb on longevity and fecundity of *R. padi* and *S. avenae* from the parent generation.

Insect	Longevity of adults (days ± SD)					Number of nymphs laid per aphid ± SD				
	Control	Pirimicarb	P	t	df	Control	Pirimicarb	P	t	df
<i>R. padi</i>	12.84 ± 1.03	13.29 ± 2.12	0.847	0.194	47	49.44 ± 3.50	29.79 ± 5.51	<0.001	4.162	47
<i>S. avenae</i>	14.41 ± 0.98	13.70 ± 1.88	0.972	0.035	67	14.74 ± 1.61	14.48 ± 1.77	0.694	0.395	67

The values (mean ± SD) in bold text are significantly different when compared to their respective control (t-test) at $P < 0.05$.

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Table 3. Effect of LC₂₅ of pirimicarb on fecundity of *R. padi* and *S. avenae* from the F1 generation.

Insect	Number of nymphs laid per aphid ± SD					Number of nymphs laid per aphid daily ± SD				
	Control	Pirimicarb	P	t	df	Control	Pirimicarb	P	t	df
<i>R. padi</i>	60.50 ± 3.74	62.38 ± 4.60	0.758	0.314	46	5.26 ± 0.28	3.63 ± 0.26	<0.001	4.250	46
<i>S. avenae</i>	23.44 ± 1.99	22.12 ± 2.73	0.690	0.400	86	1.67 ± 0.09	1.61 ± 0.13	0.745	0.327	86

The values (mean ± SD) in bold text are significantly different when compared to their respective control (t-test) at $P < 0.05$.

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significantly longer (29.96 ± 2.16 days) after pirimicarb exposure than that of the control (22.88 ± 1.58 days) ($t = 2.644, P = 0.011$). When *S. avenae* was exposed to LC₂₅ of pirimicarb, no biological traits were significantly affected compared to their respective control, except for the pre-productive period. The pre-productive period of *S. avenae* was significantly shorter (0.88 ± 0.12 days) after exposure to pirimicarb than that of the control (1.46 ± 0.12 days) ($t = 3.243, P = 0.002$) (Fig 1B).

Demographic parameters of *R. padi* and *S. avenae* when exposed to LC₂₅ pirimicarb

The intrinsic rate of increase (r_m) of *R. padi* were significantly lower when exposed to LC₂₅ of pirimicarb when compared to the control (Table 4). In addition, the mean generation time (T) and population doubling time (Dt) of *R. padi* were significantly higher than control when exposed to LC₂₅ of pirimicarb. By contrast, exposure to LC₂₅ of pirimicarb showed no significant effect on *S. avenae* demographic parameters (Table 5).

AChE activity of parent generation in *R. padi* and *S. avenae*

We evaluated the effect of LC₂₅ of pirimicarb on the total AChE enzyme activity. Our study showed that exposure by LC₂₅ of pirimicarb significantly reduced the AChE activity of *R. padi* and *S. avenae* when compared to those of their respective control. The AChE activity in pirimicarb-exposed *R. padi* decreased up to 52.5% (when compared to those of the control) after 48 h post-treatment (Fig 2A). The AChE activity in pirimicarb-exposed *S. avenae* decreased by up to 69.6% (when compared to those of the control) after 12 h post-treatment (Fig 2B).

Discussion

Sublethal effects of pesticides on arthropods may be negative or positive [11], e.g. positive effects may occur through hormesis [42]. Due to the variable distribution and continuous degradation of active ingredients on plants after initial pesticide applications [36], arthropod populations may be exposed to sublethal concentrations of pesticides. Therefore, research on sublethal effects of pesticides on target pests is of great importance for increasing their rational use, e.g. help design rational pesticide-based IPM (Integrated Pest Management) methods that would delay the development of insecticide resistances in pests. Such studies may also help shedding light on potential for insecticide-mediated shift in ecological dominance between species [43, 31]. In China, the carbamate pesticide pirimicarb is widely used in wheat fields against the aphid pest *R. padi* and *S. avenae*. Therefore, it may also cause a range of sublethal effects on *R. padi* and *S. avenae* besides its initial lethal effect; thorough knowledge of these possible effects would help optimize the IPM in wheat crops in China.

The lethal and sublethal effects of pirimicarb have been investigated on many arthropods, e.g. on *Aphidius ervi* [44], *Myzus persicae* [45] and *Coccinella undecimpunctata* [46]. Sublethal

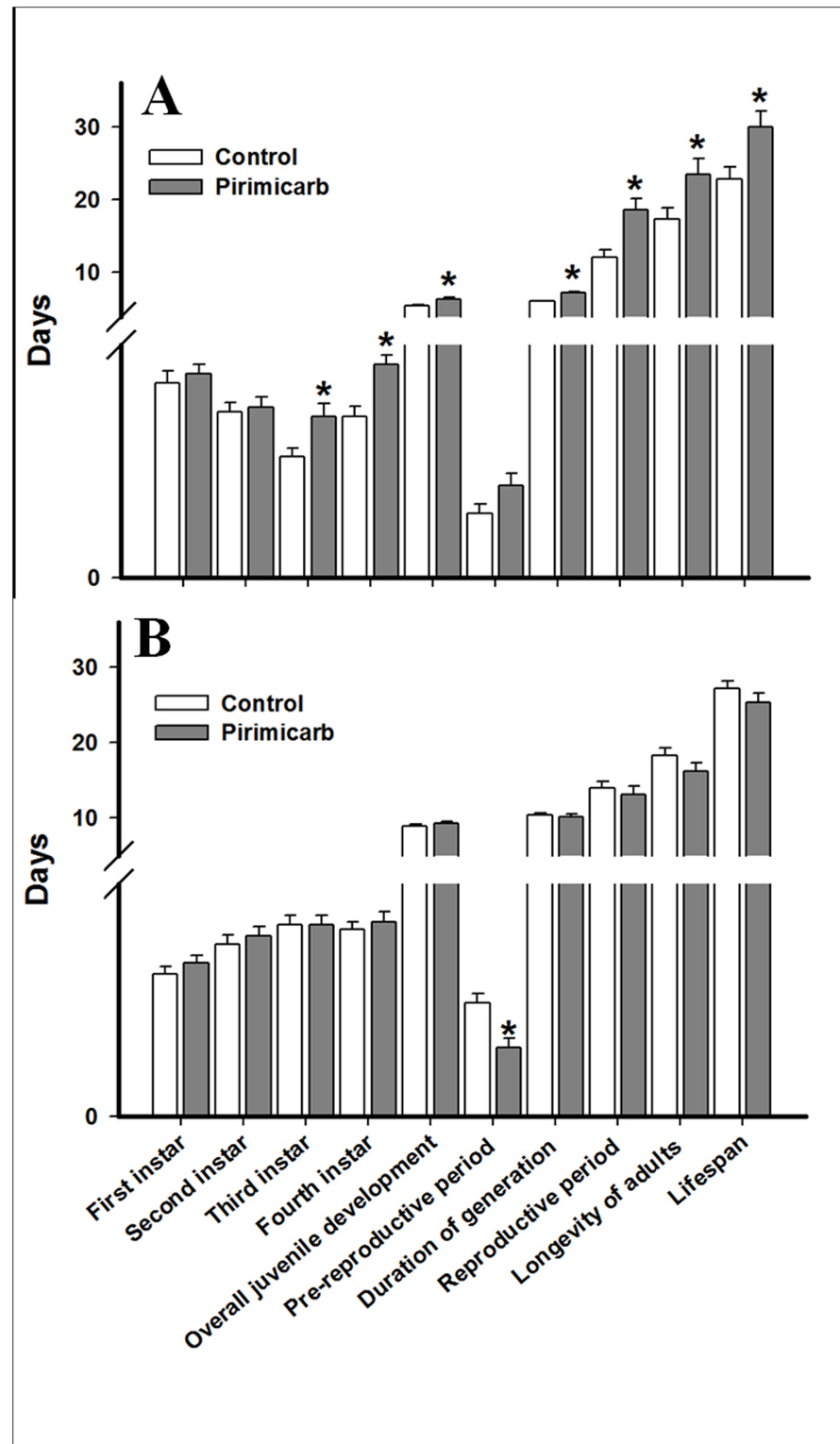


Fig 1. Effects of LC₂₅ of pirimicarb on biological traits of *R. padi* (A) and *S. avenae* (B) in the F1 generation. Asterisk on the bars of the histogram indicate significant differences between treatment and respective control (t-test) at $P < 0.05$.

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Table 4. Effect of LC₂₅ of pirimicarb on demographic parameters of *R. padi*.

Demographic Parameter	<i>R. padi</i>		Statistics		
	Control	Treatment	<i>P</i>	<i>t</i>	df
Net reproductive rate: R_0	56.20 ± 6.34	55.50 ± 5.74	0.077	3.226	46
Intrinsic rate of increase: r_m	0.36 ± 0.01	0.27 ± 0.02	< 0.001	6.835	46
Mean generation time: T (day)	11.33 ± 0.67	14.75 ± 0.54	< 0.001	7.290	46
Population doubling time: Dt (day)	1.95 ± 0.11	2.55 ± 0.21	< 0.001	6.447	46

The values (mean ± SD) in bold text are significantly different when compared to their respective control (t-test) at $P < 0.05$.

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effects such as reductions in reproductive capacity and longevity could result in negative impacts on insect population growth [11]. In our study, we found no significant difference in longevity of *R. padi* parent generation between the treated and control group. However, the fecundity of parent generation of *R. padi* exposed to pirimicarb (LC₂₅) was significantly lower than those in the control (Table 2). These results were consistent with previous studies on other carbamate insecticides, such as *Plutella xylostella* exposed to sublethal concentrations of carbaryl [47], *Choristoneura fumiferana* treated with sublethal concentration of aminocarb [48] and *Hippodamia undecimnotata* treated with sublethal concentration of carbofuran [49]. Such effect on adults may result from disturbances in the neurosecretory system caused by the LC₂₅ of pirimicarb. The reproduction process is largely regulated by neurohormones in arthropods and neurohormonal imbalance resulting from insecticide poisoning may affect normal reproductive functions [50]. By contrast, the longevity and fecundity of parent generation of *S. avenae* were not affected by exposure to the LC₂₅ of pirimicarb (Table 2). Similar lack of sublethal effect of pirimicarb has been reported by Cabral et al. (2008) [46] studying *C. undecim-punctata*, and by He et al. (2013) [51] assessing sublethal effects of carbosulfan on the fecundity of *Bemisia tabaci*. These contrasting effects stress the need to assess sublethal effects of pesticides on the specific biological models considered for further IPM optimization. In the present study, we observed contrasting results despite studying sublethal effects in two species that were (i) closely related in phylogeny [52] and (ii) that shared the same host plant (wheat). The contrasting effects of pirimicarb may not prompt an insecticide-mediated shift in ecological dominance between the two species: main demographic parameters of *R. padi* remained higher than these of *S. avenae* despite the negative effects of LC₂₅ of pirimicarb on the former (though further studies on this issue are needed because effects of pirimicarb might add up to other biotic and/or abiotic effects in field conditions).

Transgenerational effects were observed in the F1 generation of *R. padi* when parents were exposed to LC₂₅ of pirimicarb. We found that the overall juvenile development of *R. padi* was

Table 5. Effect of LC₂₅ of pirimicarb on demographic parameters of *S. avenae*.

Demographic Parameter	<i>S. avenae</i>		Statistics		
	Control	Treatment	<i>P</i>	<i>t</i>	df
Net reproductive rate: R_0	20.19 ± 2.85	19.10 ± 1.39	0.648	0.504	86
Intrinsic rate of increase: r_m	0.18 ± 0.01	0.18 ± 0.01	0.373	0.924	86
Mean generation time: T (day)	16.44 ± 0.74	16.01 ± 0.69	0.831	0.204	86
Population doubling time: Dt (day)	3.79 ± 0.13	3.76 ± 0.09	0.596	0.617	86

The values (mean ± SD) in bold text are significantly different when compared to their respective control (t-test) at $P < 0.05$.

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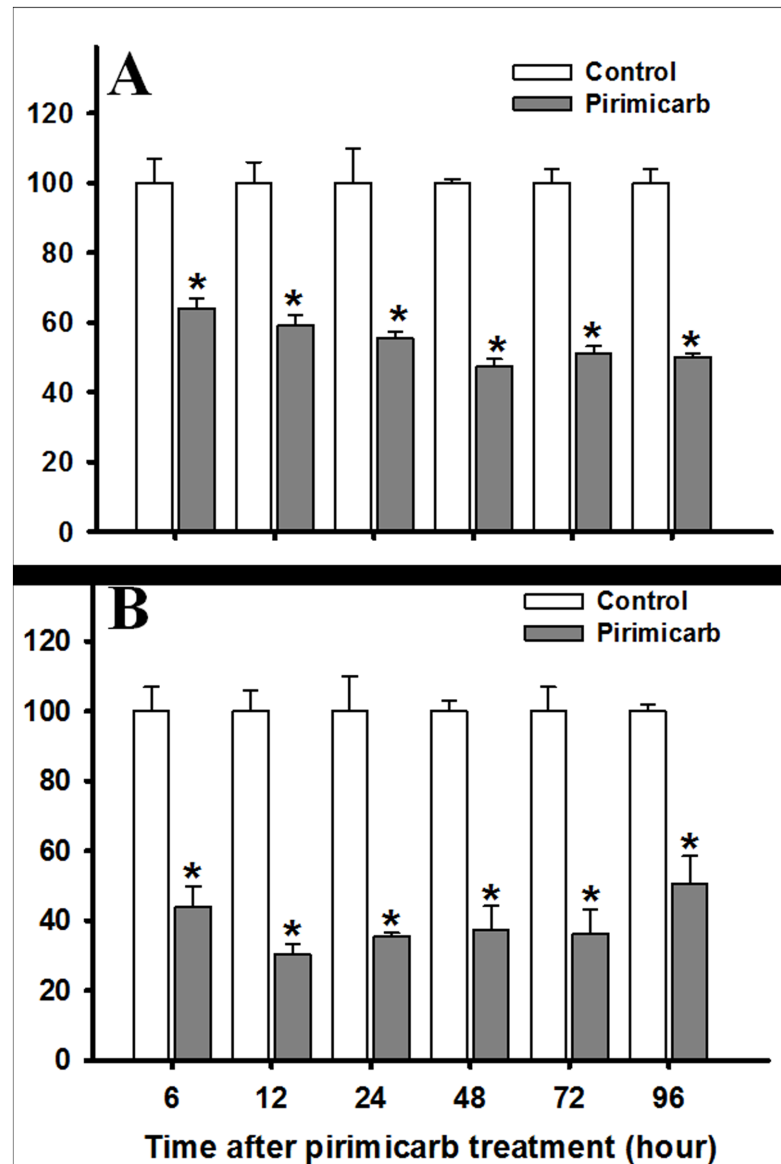


Fig 2. Effects of LC₂₅ of pirimicarb on AChE activities of *R. padi* (A) and *S. avenae* (B) from the parent generation. AChE activities were expressed as nmol of ATCh hydrolyzed/min/mg protein. The results are presented as the mean and standard errors of three replicates. Asterisk on the bars of the histogram indicate significant differences between treatment and respective control (t-test) at $P < 0.05$.

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longer than those in the control. This result was consistent with an earlier study demonstrating that larvae development of *C. fumiferana* was significantly extended after exposure to sublethal concentration of carbaryl [48]. Moreover, traits like the duration of generation, reproductive period and adult longevity were also longer than those of the control (Fig 1A). Similar effect, i.e. increased adults longevity in offspring from parents previously exposed to a pesticide, was also observed in *P. xylostella* exposed to sublethal concentrations of carbaryl [47] and *Anopheles stephensi* exposed to sublethal concentrations of propoxur [53]. Moreover, the longer reproductive period in treated group led to an average fecundity (per reproductive day) significantly lower than in the control group; there was no significant difference was observed on total

fecundity (Table 3). As previously observed in case of individuals in the parent generation, the sublethal effects of the pirimicarb on *R. padi* were not found in *S. avenae* (Fig 1B).

The demographic parameters in pirimicarb-exposed and control aphid populations provided insight on possible effects of pirimicarb on *R. padi* and *S. avenae* at the generational scale. After exposure of parents to pirimicarb (LC₂₅), the r_m values of *R. padi* from the F1 generation was lower than those of the control group, whereas both the T and Dt values were higher than those of the control. In addition, no significant effects on these parameters were observed when *S. avenae* exposed to LC₂₅ of pirimicarb. This suggests that the LC₂₅ of pirimicarb may slow down population growth of *R. padi* but *S. avenae*.

Hormesis refers to possible enhanced performances of individuals occurring at low levels of exposure to toxic agents that are harmful at high levels of exposure [42]. Pesticide-induced hormesis may be an alternate mechanism explaining pest resurgences; this is a serious problem in agriculture [54]. Hormesis in insects exposed to sublethal concentrations of insecticides has been documented for several taxa and compounds [55]. In the present study, we assessed whether hormesis effects, especially on fecundity and aphid population growth-related traits, may occur in *R. padi* and *S. avenae* during and/or after exposure (for parent and F1 generation, respectively) to a low dose of pirimicarb. The parent generation of *R. padi* exposed to LC₂₅ pirimicarb had significantly lower numbers of nymphs produced, and most of aphid development-related traits recorded in F1 generation showed a slow-down development (e.g. increased duration of pre-reproductive period). LC₂₅ pirimicarb had no significant effect on the fecundity and longevity in parent generation and various biological traits in F1 generation of *S. avenae*. Therefore, no stimulatory effect on fecundity was induced by LC₂₅ pirimicarb in *R. padi* and *S. avenae*, and no resurgence may occur in *R. padi* and *S. avenae* when exposed to low concentrations of pirimicarb.

AChE is an important biochemical marker in ecotoxicology. In our study, AChE activities in survival aphids provided a measure of sublethal effects of pirimicarb. The results demonstrated that the AChE activity of *R. padi* was reduced after exposure to pirimicarb, and the similar trend of AChE activity was observed in *S. avenae*. These results were in concordance with the previous studies on organophosphate insecticides which have same mode action with carbamate insecticides, e.g. for *Micromus tasmaniae* when exposed to azinphos-methyl and methyl parathion [24] and when exposed to diazinon and chlorpyrifos [56]. AChE activity of *Helicoverpa armigera* decreased after exposure to LD₁₀ of phoxim [57] and exposure to dimethoate induced an inhibition of AChE activity in both *M. tasmaniae* and *R. padi* [25]. Moreover, the specific activity of AChE in *M. persicae* was reduced significantly when treated with different sublethal doses of imidacloprid [58].

In summary, the present study indicated that a low concentration of pirimicarb would affect the fecundity of *R. padi* of parent generation, and would also trigger biological traits variation at the F1 generation of *R. padi*. In addition, no stimulatory effects on fecundity were observed in parent and F1 generation of *S. avenae* and *R. padi*. In practice, these findings under laboratory conditions highlighted the importance of sublethal effects on aphids and how they may translate to population dynamic in the field. Therefore, our study provided evidence for promoting the continued use of pirimicarb for wheat aphid control.

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Author Contributions

Conceived and designed the experiments: XWG DX. Performed the experiments: DX TY PH. Analyzed the data: DX TY ND XWG. Contributed reagents/materials/analysis tools: DX TY XWG. Wrote the paper: DX XWG ND PH.

References

1. Aqueel MA, Leather SR (2011) Effect of nitrogen fertilizer on the growth and survival of *Rhopalosiphum padi* (L.) and *Sitobion avenae* (F.) (Homoptera: Aphididae) on different wheat cultivars. *Crop Prot* 30: 216–221.
2. Caballero-López B, Blanco-Moreno JM, Pérez-Hidalgo N, Michelena-Saval JM, Pujade-Villar J, Guerrieri E, et al. (2012) Weeds, aphids, and specialist parasitoids and predators benefit differently from organic and conventional cropping of winter cereals. *J Pest Sci* 85: 81–88.
3. Savaris M, Lampert S, Salvadori JR, Lau D, Pereir PRVS, Smaniotto MA (2013) Population growth and damage caused by *Rhopalosiphum padi* (L.) (Homoptera, Aphididae) on different cultivars and phenological stages of wheat. *Neotrop Entomol* 42: 539–543.
4. Lu YH, Gao XW (2007) A method for mass culture of wheat aphid. *Chin Bull Entomol* 44: 289–290.
5. Brault V, Herrbach É, Reinbold C (2007) Electron microscopy studies on luteovirus transmission by aphids. *Micron* 38: 302–312. PMID: [16750376](#)
6. Larsson H (2005) A crop loss model and economic thresholds for the grain aphid, *Sitobion avenae* (F.), in winter wheat in southern Sweden. *Crop Prot* 24: 397–405.
7. Chen MH, Han ZJ, Qiao XF, Qu MJ (2007) Resistance mechanisms and associated mutations in acetylcholinesterase genes in *Sitobion avenae* (Fabricius). *Pestic Biochem Physiol* 87: 189–195.
8. Lu YH, Wu KM, Jiang YY, Guo YY, Desneux N (2012) Widespread adoption of Bt cotton and insecticide decrease promotes biocontrol services. *Nature* 487: 362–365. doi: [10.1038/nature11153](#) PMID: [22722864](#)
9. Vanaclocha P, Vidal-Quist C, Oheix S, Monton H, Planes L, Catalan J, et al., (2013) Acute toxicity in laboratory tests of fresh and aged residues of pesticides used in citrus on the parasitoid *Aphytis melinus*. *J Pest Sci* 86: 329–336.
10. Lu YH, He YP, Gao XW (2013) Comparative studies on acetylcholinesterase characteristics between the aphids, *Sitobion avenae* and *Rhopalosiphum padi*. *J Insect Sci* 13: 9. doi: [10.1673/031.013.0901](#) PMID: [23879406](#)
11. Desneux N, Decourtye A, Delpuech JM (2007) The sublethal effects of pesticides on beneficial arthropods. *Annu Rev Entomol* 52: 81–106. PMID: [16842032](#)
12. Kerns DL, Stewart SD (2000) Sublethal effects of insecticides on the intrinsic rate of increase of cotton aphid. *Entomol Exp Appl* 94: 41–49.
13. Stark JD, Banks JE (2003) Population-level effects of pesticides and other toxicants on arthropods. *Annu Rev Entomol* 48: 505–519. PMID: [12221038](#)
14. Biondi A, Zappalà L, Stark JD, Desneux N (2013) Do biopesticides affect the demographic traits of a parasitoid wasp and its biocontrol services through sublethal effects? *PLoS ONE* 8: e76548. doi: [10.1371/journal.pone.0076548](#) PMID: [24098793](#)
15. Fogel MN, Schneider MI, Desneux N, González MB, Ronco AE (2013) Impact of the neonicotinoid acetamiprid on immature stages of the predator *Eriopsis connexa* (Coleoptera: Coccinellidae). *Ecotoxicology* 22: 1063–1071. doi: [10.1007/s10646-013-1094-5](#) PMID: [23793295](#)
16. Saber M, Abedi Z (2013) Effects of methoxyfenozide and pyridalyl on the larval ectoparasitoid *Habrobracon hebetor*. *J Pest Sci* 86: 685–693.
17. Pan HS, Liu YQ, Liu B, Lu YH, Xu XY et al. (2014) Lethal and sublethal effects of cycloxyaprid, a novel cis-nitromethylene neonicotinoid insecticide, on the mirid bug *Apolygus lucorum*. *J Pest Sci* 87: 731–738
18. Desneux N, Ramirez-Romero R, Kaiser L (2006) Multistep bioassay to predict recolonization potential of emerging parasitoids after a pesticide treatment. *Environ Toxicol Chem* 25: 2675–2682.
19. Bao HB, Liu SH, Gu JH, Wang XZ, Liang XL, Liu ZW (2009) Sublethal effects of four insecticides on the reproduction and wing formation of brown planthopper, *Nilaparvata lugens*. *Pest Manag Sci* 65: 170–174. doi: [10.1002/ps.1664](#) PMID: [18937216](#)
20. Tan Y, Biondi A, Desneux N, Gao XW (2012) Assessment of physiological sublethal effects of imidacloprid on the mirid bug *Apolygus lucorum* (Meyer-Dur). *Ecotoxicology* 21: 1989–1997. doi: [10.1007/s10646-012-0933-0](#) PMID: [22740097](#)

21. Planes L, Catalán J, Tena A, Porcuna JL, Jacas J A, Izquierdo J. et al., (2013) Lethal and sublethal effects of spirotetramat on the mealybug destroyer, *Cryptolaemus montrouzieri*. *J Pest Sci* 86: 321–327.
22. Croft BA (1990) *Arthropod Biological Control Agents and Pesticides*. Wiley and Sons, New York.
23. Biondi A, Mommaerts V, Smagghe G, Viñuela E, Zappalà L, Desneux N (2012) The non-target impact of spinosyns on beneficial arthropods. *Pest Manag Sci* 68: 1523–1536. doi: [10.1002/ps.3396](https://doi.org/10.1002/ps.3396) PMID: [23109262](https://pubmed.ncbi.nlm.nih.gov/23109262/)
24. Rumpf S, Hetzel F, Frampton C (1997) Lacewings (Neuroptera: Hemeroibiidae and Chrysopidae) and integrated pest management: enzyme activity as biomaker of sublethal insecticide exposure. *J Econ Entomol* 90: 102–108.
25. Booth LH, Wratten SD, Kehrl P (2007) Effects of reduced rates of two insecticides on enzyme activity and mortality of an aphid and its lacewing predator. *J Econ Entomol* 100: 11–19. PMID: [17370803](https://pubmed.ncbi.nlm.nih.gov/17370803/)
26. Kumrungsee N, Pluempanupat W, Koul O, Bullangpoti V (2014) Toxicity of essential oil compounds against diamondback moth, *Plutella xylostella*, and their impact on detoxification enzyme activities. *J Pest Sci* 87:721–729
27. Peng Y, Luo Y, Nie XP, Liao W, Yang YF, Ying GG (2013) Toxic effects of Triclosan on the detoxification system and breeding of *Daphnia magna*. *Ecotoxicology* 22: 1384–1394. PMID: [24022518](https://pubmed.ncbi.nlm.nih.gov/24022518/)
28. Weill M, Fort P, Berthomieu A, Dubois MP, Pasteur N, Raymond M (2002) A novel acetylcholinesterase gene in mosquitoes codes for the insecticide target and is non-homologous to the *ace* gene *Drosophila*. *Proc R Soc Lond B* 269: 2007–2016.
29. Lu YH, Gao XW (2009) Multiple mechanisms responsible for differential susceptibilities of *Sitobion avenae* (Fabricius) and *Rhopalosiphum padi* (Linnaeus) to pirimicarb. *Bull Entomol Res* 99: 611–617. doi: [10.1017/S0007485309006725](https://doi.org/10.1017/S0007485309006725) PMID: [19413913](https://pubmed.ncbi.nlm.nih.gov/19413913/)
30. Lu YH, Yang T, Gao XW (2009) Establishment of baseline susceptibility data to various insecticides for aphids *Rhopalosiphum padi* (Linnaeus) and *Sitobion avenae* (Fabricius) (Homoptera: Aphididae) by the method of residual film in glass tube. *Acta Entomol Sinica* 52: 52–58.
31. Liang P, Tian YA, Biondi A, Desneux N, Gao XW (2012) Short-term and transgenerational effects of the neonicotinoid nitenpyram on susceptibility to insecticides in two whitefly species. *Ecotoxicology* 21: 1889–1898. doi: [10.1007/s10646-012-0922-3](https://doi.org/10.1007/s10646-012-0922-3) PMID: [22661314](https://pubmed.ncbi.nlm.nih.gov/22661314/)
32. Shad SA, Sayyed AH, Fazal S, Saleem MA, Zaka SM, Ali M (2012) Field evolved resistance to carbamates, organophosphates, pyrethroids, and new chemistry insecticides in *Spodoptera litura* Fab. (Lepidoptera: Noctuidae). *J Pest Sci* 85: 153–162.
33. Guo L, Desneux N, Sonoda S, Liang P, Han P, Gao XW (2013) Sublethal and transgenerational effects of chlorantraniliprole on biological traits of the diamondback moth, *Plutella xylostella* L. *Crop Prot* 48: 29–34.
34. Tong H, Su Q, Zhou XM, Bai LY (2013) Field resistance of *Spodoptera litura* (Lepidoptera: Noctuidae) to organophosphates, pyrethroids, carbamates and four newer chemistry insecticides in Hunan, China. *J Pest Sci* 86: 599–609.
35. Shea K, Roxburgh SH, Rauscher ESJ (2004) Moving from pattern to process: coexistence mechanisms under intermediate disturbance regimes. *Ecol Lett* 7: 491–508.
36. Desneux N, Fauvergue X, Dechaume-Moncharmont FX, Kerhoas L, Ballanger Y, Kaiser L (2005) *Diaeretiella rapae* limits *Myzus persicae* populations following applications of deltamethrin in oilseed rape. *J Econ Entomol* 98: 9–17. PMID: [15765661](https://pubmed.ncbi.nlm.nih.gov/15765661/)
37. Moores GD, Gao XW, Denholm I, Devonshire AL (1996) Characterisation of insensitive acetylcholinesterase in insecticide-resistant cotton aphids, *Aphis gossypii* Glover (Homoptera: Aphididae). *Pestic Biochem Physiol* 56: 102–110.
38. Ellman GL, Courtney KD, Andres VJR, Featherstone RM (1961) A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol* 7: 88–95. PMID: [13726518](https://pubmed.ncbi.nlm.nih.gov/13726518/)
39. Gao XW (1987) Introduction of Ellman procedure for assay of cholinesterases in crude enzymatic preparations modified by Gorun. *Chin Bull Entomol* 33: 245–246.
40. Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248–254. PMID: [942051](https://pubmed.ncbi.nlm.nih.gov/942051/)
41. Finney DJ (1971) *Probit analysis*. Cambridge University Press, Cambridge
42. Calabrese EJ, Baldwin LA (2003) Toxicology rethinks its central belief-Hormesis demands a reappraisal of the way risks are assessed. *Nature* 421: 691–692. PMID: [12610596](https://pubmed.ncbi.nlm.nih.gov/12610596/)
43. Gianoli E (2000) Competition in cereal aphids (Homoptera: Aphididae) on wheat plants. *Env Entomol* 29: 213–219.

44. Desneux N, Rafalimanana H, Kaiser L (2004) Dose-response relationship in lethal and behavioural effects of different insecticides on the parasitic wasp *Aphidius ervi*. *Chemosphere* 54: 619–627. PMID: [14599507](#)
45. Wang XY, Yang ZQ, Shen ZR, Lu J, Xu WB (2008) Sublethal effects of selected insecticides on fecundity and wing dimorphism of green peach aphid (Hom., Aphididae). *J Appl Entomol* 132: 135–142.
46. Cabral S, Garcia P, Soares AO (2008) Effects of pirimicarb, buprofezin and pymetrozine on survival, development and reproduction of *Coccinella undecimpunctata* (Coleoptera: Coccinellidae). *Biocontrol Sci Techn* 18: 307–318.
47. Kumar K, Chapman RB (1984) Sublethal effects of insecticides on the Diamondback Moth *Plutella xylostella* (L.). *Pestic Sci* 15: 344–352.
48. Alford AR, Holmes JA (1986) Sublethal effects of carbaryl, aminocarb, fenitrothion and *Bacillus thuringiensis* on the development and fecundity of the spruce budworm (Lepidoptera: Tortricidae). *J Econ Entomol* 79: 31–34.
49. Papachristos DP, Milonas PG (2008) Adverse effects of soil applied insecticides on the predatory coccinellid *Hippodamia undecimnotata* (Coleoptera: Coccinellidae). *Biol Control* 47: 77–81.
50. Lee CY (2000) Sublethal effects of insecticides on longevity, fecundity and behaviour of insect pests: a review. *J Bioscience* 11: 107–112.
51. He YX, Zhao JW, Zheng Y, Weng QY, Biondi A, Desneux N. et al., (2013) Assessment of potential sublethal effects of various insecticides on key biological traits of the tobacco whitefly, *Bemisia tabaci*. *Int J Biol Sci* 9: 246–255. doi: [10.7150/ijbs.5762](#) PMID: [23494876](#)
52. Desneux N, Blahnik R, Delebecque CJ, Heimpel GE (2012) Host phylogeny and host specialization in parasitoids. *Ecology Letters* 15: 453–460. doi: [10.1111/j.1461-0248.2012.01754.x](#) PMID: [22404869](#)
53. Sanil D, Shetty NJ (2012) The effect of sublethal exposure to temephos and propoxur on reproductive fitness and its influence on circadian rhythms of pupation and adult emergence in *Anopheles stephensi* Liston—a malaria vector. *Parasitol Res* 111: 423–432. doi: [10.1007/s00436-012-2857-2](#) PMID: [22392132](#)
54. Guedes RNC, Cutler GC (2014) Insecticide-induced hormesis and arthropod pest management. *Pest Manag Sci* doi: [10.1002/ps.3669](#)
55. Cutler GC (2013) Insects, insecticides and hormesis: evidence and considerations for study. *Dose-Response* 11: 154–177. doi: [10.2203/dose-response.12-008.Cutler](#) PMID: [23930099](#)
56. Hodge S, Longley M, Booth L, Heppelthwaite V, O'Halloran K (2000) Cholinesterase activity in the tasmanian lacewing as a biomarker for organophosphorus pesticides. *Aust J Ecotoxicol* 6: 109–116.
57. Liu B, Gao XW, Zheng BZ (2003) Effects of sublethal doses of anticholinesterase agents on toxicity of insecticides and their induction to acetylcholinesterase (AChE) activity in *Helicoverpa armigera*. *Acta Entomol Sinica* 46: 691–696.
58. Zeng CX, Wang JJ (2007) Time and dose effects of sublethal imidacloprid concentrations on acetylcholinesterase in *Myzus persicae*. *Plant Prot* 33: 50–54.