



HAL
open science

High-Quality Draft Genome Sequences of Two *Xanthomonas citri* pv. *malvacearum* Strains.

Sébastien Cunnac, Stéphanie S. Bolot, Natalia Forero Serna, Erika Ortiz, Boris Szurek, Laurent Noel, Matthieu Arlat, Marie Agnes Jacques, Lionel Gagnevin, Sebastien Carrere, et al.

► **To cite this version:**

Sébastien Cunnac, Stéphanie S. Bolot, Natalia Forero Serna, Erika Ortiz, Boris Szurek, et al.. High-Quality Draft Genome Sequences of Two *Xanthomonas citri* pv. *malvacearum* Strains.. *Genome Announcements*, 2013, 1 (4), 10.1128/genomeA.00674-13 . hal-02643147

HAL Id: hal-02643147

<https://hal.inrae.fr/hal-02643147>

Submitted on 28 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

High-Quality Draft Genome Sequences of Two *Xanthomonas citri* pv. *malvacearum* Strains

Sébastien Cunnac,^a Stéphanie Bolot,^{b,c} Natalia Forero Serna,^a Erika Ortiz,^a Boris Szurek,^a Laurent D. Noël,^{b,c} Matthieu Arlat,^{b,c,d} Marie-Agnès Jacques,^e Lionel Gagnevin,^f Sébastien Carrere,^{b,c} Michel Nicole,^a Ralf Koebnik^a

UMR 186 IRD-Cirad-Université Montpellier 2 "Résistance des Plantes aux Bioagresseurs," Montpellier, France^a; INRA, Laboratoire des Interactions Plantes Micro-organismes (LIPM), Castanet-Tolosan, France^b; CNRS, Laboratoire des Interactions Plantes Micro-organismes (LIPM), Castanet-Tolosan, France^c; Université de Toulouse, Université Paul Sabatier, Toulouse, France^d; INRA, Institut de Recherche en Horticulture et Semences (IRHS), Beaucauzé, France^e; UMR "Peuplements Végétaux et Bioagresseurs en Milieu Tropical" (PVMT), La Réunion, France^f

Sébastien Cunnac and Stéphanie Bolot contributed equally to this study.

We report high-quality draft genome sequences of two strains (race 18 and 20) of *Xanthomonas citri* pv. *malvacearum*, the causal agent of bacterial blight of cotton. Comparative genomics will help to decipher mechanisms provoking disease and triggering defense responses and to develop new molecular tools for epidemiological surveillance.

Received 30 July 2013 Accepted 31 July 2013 Published 29 August 2013

Citation Cunnac S, Bolot S, Forero Serna N, Ortiz E, Szurek B, Noël LD, Arlat M, Jacques M-A, Gagnevin L, Carrere S, Nicole M, Koebnik R. 2013. High-quality draft genome sequences of two *Xanthomonas citri* pv. *malvacearum* strains. *Genome Announc.* 1(4):e00674-13. doi:10.1128/genomeA.00674-13.

Copyright © 2013 Cunnac et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Ralf Koebnik, koebnik@gmx.de.

Xanthomonas citri pv. *malvacearum* is the causal agent of bacterial blight (BB), one of the most devastating diseases of cotton (*Gossypium* spp.). BB of cotton was first reported in the United States by Atkinson, who described several symptoms, such as angular leaf spots, water-soaked lesions, stem black arm, boll rot, and plantlet burning (1). In the second half of the 20th century, BB became a limiting factor of fiber production in all major cotton-producing regions of Africa, Asia, Australia, and North America (2). The use of resistant cultivars is usually the most efficient way to manage disease. However, resistance was repeatedly overcome by the appearance of new races. More than 20 races have been described, including highly virulent strains that were isolated in Central Africa in the 1980s (2). To better understand the molecular basis of provoking disease and of triggering defense responses and to develop new molecular markers for epidemiological surveillance, we sequenced the genomes of two strains originating from Burkina Faso and belonging to the highly virulent race 20 and the less virulent race 18 (3).

Both strains were sequenced using the Illumina Hi-Seq2000 platform (GATC Biotech, Germany). The shotgun sequencing yielded 40,654,599 read pairs (29,066,551 100-bp paired-end reads with an insert size of 250 bp and 11,588,048 50-bp mate-pair reads with an insert size of 3 kb) and 43,359,649 read pairs (19,340,592 100-bp paired-end reads and 24,019,057 50-bp mate-pair reads) for strains X18 and X20, respectively. A combination of Velvet (4), SOAPdenovo, and SOAPGapCloser (5) yielded 22 contigs larger than 500 bp (N_{50} , 705,301 bp), with the largest contig of 2,062 kb, for a total assembly size of 4,989,917 bp for strain X18 and 17 contigs larger than 500 bp (N_{50} , 1,006,603 bp), with the largest scaffold of 1,770 kb, for a total assembly size of 5,216,199 bp for strain X20.

Multilocus sequence analyses of six housekeeping genes described earlier for xanthomonads confirmed that strains X18 and

X20 belong to *X. citri* pv. *malvacearum* (2,745 bp with 100% identity) (6), corresponding to DNA-DNA homology group 9.5, which also includes *X. citri* pv. *citri* (2,730 identical residues) and *X. citri* pv. *glycines* (2,729 identical residues) (7). The genome encodes a canonical type III protein secretion system (8) and several type III effectors, including transcriptional activator-like (TAL) effectors, which are responsible for symptom formation and avirulence reactions on cotton (9). The availability of two genome sequences of *X. citri* pv. *malvacearum* will critically aid in developing new molecular typing tools for epidemiological surveillance and guiding breeding programs based on rapid and accurate identification of predominant lineages.

Nucleotide sequence accession numbers. These whole-genome shotgun projects have been deposited in GenBank under the accession numbers [ATMA000000000](https://www.ncbi.nlm.nih.gov/nuccore/ATMA000000000) for strain X18 and [ATMB000000000](https://www.ncbi.nlm.nih.gov/nuccore/ATMB000000000) for strain X20.

ACKNOWLEDGMENT

This work was supported by grant ANR-2010-GENM-013 from the French Agence Nationale de la Recherche.

REFERENCES

- Atkinson GF. 1891. The black rust of cotton. *Ala. Agric. Exp. Stn. Bull.* 27:1–16.
- Delannoy E, Lyon BR, Marmey P, Jalloul A, Daniel JF, Montillet JL, Essenberg M, Nicole M. 2005. Resistance of cotton towards *Xanthomonas campestris* pv. *malvacearum*. *Annu. Rev. Phytopathol.* 43:63–82. doi: [10.1146/annurev.phyto.43.040204.140251](https://doi.org/10.1146/annurev.phyto.43.040204.140251).
- Martinez C, Baccou JC, Bresson E, Baissac Y, Daniel JF, Jalloul A, Montillet JL, Geiger JP, Assigbetsé K, Nicole M. 2000. Salicylic acid mediated by the oxidative burst is a key molecule in local and systemic responses of cotton challenged by an avirulent race of *Xanthomonas campestris* pv. *malvacearum*. *Plant Physiol.* 122:757–766. doi: [10.1104/pp.122.3.757](https://doi.org/10.1104/pp.122.3.757).
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read

- assembly using de Bruijn graphs. *Genome Res.* 18:821–829. doi: [10.1101/gr.074492.107](https://doi.org/10.1101/gr.074492.107).
5. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. *Gigascience* 1:18. doi: [10.1186/2047-217X-1-18](https://doi.org/10.1186/2047-217X-1-18).
 6. Almeida NF, Yan S, Cai R, Clarke CR, Morris CE, Schaad NW, Schuenzel EL, Lacy GH, Sun X, Jones JB, Castillo JA, Bull CT, Leman S, Guttman DS, Setubal JC, Vinatzer BA. 2010. PAMDB, a multilocus sequence typing and analysis database and website for plant-associated microbes. *Phytopathology* 100:208–215. doi: [10.1094/PHYTO-100-3-0208](https://doi.org/10.1094/PHYTO-100-3-0208).
 7. Rademaker JL, Louws FJ, Schultz MH, Rossbach U, Vauterin L, Swings J, de Bruijn FJ. 2005. A comprehensive species to strain taxonomic framework for *Xanthomonas*. *Phytopathology* 95:1098–1111. doi: [10.1094/PHYTO-95-1098](https://doi.org/10.1094/PHYTO-95-1098).
 8. Büttner D. 2012. Protein export according to schedule: architecture, assembly, and regulation of type III secretion systems from plant- and animal-pathogenic bacteria. *Microbiol. Mol. Biol. Rev.* 76:262–310. doi: [10.1128/MMBR.05017-11](https://doi.org/10.1128/MMBR.05017-11).
 9. Yang Y, De Feyter R, Gabriel DW. 1994. Host-specific symptoms and increased release of *Xanthomonas citri* and *X. campestris* pv. *malvacearum* from leaves are determined by the 102-bp tandem repeats of *pthA* and *avrB6*, respectively. *Mol. Plant Microbe Interact.* 7:345–355. doi: [10.1094/MPMI-7-0345](https://doi.org/10.1094/MPMI-7-0345).