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Response of milk fat concentration and yield to nutrient supply in dairy cows

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Dietary changes alter dairy cow milk fat concentration (MFC) and yield (MFY) through modifications in the supply of nutrients, which act as precursors or inhibitors of mammary fat synthesis. The current models used to formulate dairy cow diets cannot predict changes in milk fat. The knowledge of the effects of the nutrients on milk fat would help to progress toward this prediction. To this end, we quantified and compared the milk fat responses to variations in the supply of seven nutrients derived from digestion: volatile fatty acids, glucose, proteins, long-chain fatty acids (LCFA) and τ 10, τ 12-conjugated linoleic acid (CLA). A database was compiled from studies involving digestive infusions of these nutrients in dairy cows. It included 147 comparisons between a nutrient infusion and a control treatment. The nutrient infusions were limited to the range of physiological variations to mimic nutrient changes after dietary modifications. We established models for the response of MFC, MFY and milk fatty acid (FA) composition to the supply of each nutrient. MFC and MFY responses to the nutrients were significant and linear, except for the MFC response to glucose that was curvilinear. The nutrients differed in their effects on MFC and MFY: acetate, butyrate and LCFA increased MFC and MFY, whereas propionate, glucose and τ 10, τ 12-CLA decreased them. Protein infusions increased MFY and decreased MFC because of an increase in milk yield. The effects of numerous interfering factors related to animals, diets or experimental conditions were tested on the residuals of the response models. The responses of milk FA percentages are also provided. When adjusted to the in vivo variations in the nutrients observed after dietary changes, the effects of the different nutrients were moderate. Finally, this study showed that several of these nutrients could contribute to the changes in milk fat production and composition observed after dietary changes. This is a first step toward predicting milk fat response to changes in nutrient supply.

Keywords: milk fat, dairy cow, meta-analysis, nutrients

Implications

Fluctuations in the milk fat market are driving demands from farmers for ways to control milk fat production and composition in dairy cows. The current models used to estimate dairy cow requirements and formulate diets cannot predict changes in milk fat. These models are based on energy and protein intake and do not consider the pattern of individual nutrients derived from digestion. Using published infusion studies, this meta-analysis quantifies and compares the effects on milk fat secretion of several nutrients derived from digestion. It is a first step toward a model to estimate variations in milk fat based on variations in nutrient supply.

Introduction

The modulation of milk fat production is a concern for dairy farmers wanting to adapt to market demands, and has long been studied by ruminant physiologists. Dietary modifications are a simple way to modulate milk fat concentration (MFC) and yield (MFY), as these effects are rapid (e.g. Roy *et al.*, 2006) and reversible. Milk fat depression (MFD) is an extreme example of nutritional modulation of milk fat, with reductions in MFC of up to 50% (Bauman and Griinari, 2001). In the last 50 years, several theories have been advanced to explain this phenomenon (reviewed in Bauman and Griinari, 2001 and 2003). Currently, the mainstream theory is the biohydrogenation theory: diets inducing MFD cause a change in ruminal lipid metabolism, leading to an increased formation of specific *trans* fatty acids (FA) that inhibit lipid synthesis in the mammary gland (Bauman and

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Griinari, 2001). Using abomasal or duodenal infusions, it has been shown that the isomer τ 10, ϵ 12-conjugated linoleic acid (CLA) and a few other CLA isomers are indeed inhibitors of mammary lipogenesis (Bauman and Griinari, 2003; Shingfield and Griinari, 2007; Harvatine *et al.*, 2009). However, most authors acknowledge that the quantities of these isomers produced *in vivo* in the rumen are insufficient to fully explain the milk fat reductions observed during MFD (Roy *et al.*, 2006; reviews in Harvatine *et al.*, 2009; Shingfield and Griinari, 2007). Two explanations have been suggested: either there are additional FA isomers (still unidentified) that inhibit milk fat secretion and contribute to MFD (Shingfield and Griinari, 2007; Harvatine *et al.*, 2009; Shingfield *et al.*, 2010), or other nutrients or mechanisms are also involved in MFD (Shingfield *et al.*, 2010), as the diets inducing MFD generally disrupt the ruminal fermentation process and thereby alter the supplies of numerous nutrients to the cows.

With this second hypothesis in mind, the aim of this study was to quantify the effects of several nutrients derived from digestion on MFC and MFY, in order to assess whether they could have a significant effect on MFD. A preliminary study had been conducted by Rulquin *et al.* (2007) on the overall effects of energy nutrients on MFC and MFY. This study refines and extends their study, both on the nutrients and variables studied. If the responses of MFC and MFY to these nutrients were determined, they could be used to estimate milk fat changes after dietary modifications, provided that the changes in nutrient supply following dietary modifications can be predicted.

To study the effects of changes in one nutrient without interference from the others, we looked at studies using digestive infusions of nutrients. The response equations were generated by a meta-analysis run from these studies, and the putative interfering factors (stage of lactation, experimental procedures, diet composition, initial milk fat, etc.) on these equations were systematically explored. When enough data were available, we also studied the responses of the milk FA composition to changes in the supply of these nutrients. We established the equations describing the response of milk fat to changes in the supply of seven nutrients derived from digestion for which enough published infusion studies were available: the volatile FA (VFA; acetic (C2), propionic (C3) and butyric (C4) acids), glucose, τ 10, ϵ 12-CLA, long-chain FA (LCFA) and proteins.

Material and methods

Data inclusion

A database was compiled from published studies on dairy cows, reporting the individual effects of seven nutrients derived from digestion: C2, C3, C4, glucose, τ 10, ϵ 12-CLA, LCFA and proteins. Studies were included in the database only when they met two criteria: (i) they used continuous ruminal infusions of individual VFA, duodenal infusions of glucose, proteins, animal or plant lipids (for LCFA) and τ 10, ϵ 12-CLA and (ii) they included a control treatment (unsupplemented). For τ 10, ϵ 12-CLA experiments, the lipid

supplement was pure τ 10, ϵ 12-CLA or a FA mixture, but only studies with at least 30% of τ 10, ϵ 12-CLA in the FA mixture supplement were selected. The sources of infused proteins were casein (80% of the experiments) or plant proteins (soy or cotton, 20% of the experiments). Infusions with only one or a few amino acids (AA) were not included, as they could induce an AA imbalance. The lipids infused in the LCFA studies were high-oleic sunflower oil ($n = 3$), rapeseed oil ($n = 2$), pure FA ($n = 5$), tallow ($n = 2$), olive oil ($n = 2$) and other plant oils ($n = 2$).

From these published studies, we then excluded the experimental treatments for which the amount of nutrients supplied was outside the range of physiological variations, in order to study only those modifications in milk fat secretion that were similar to those induced by dietary changes. The supply of quantities higher than physiological could trigger different biological mechanisms and therefore not be relevant for nutritional applications. The upper limits for the amount supplied were estimated from publications studying changes in nutrients supply following dietary changes. They were set at 1600 g/day for C2, 1000 g/day for C3 and 800 g/day for C4 (values from a database of measured VFA productions, Nozière *et al.*, 2007); 800 g/day for proteins (Ipharraguerre *et al.*, 2005), 1000 g/day for lipids (Christensen *et al.*, 1998) and 5 g/day for τ 10, ϵ 12-CLA. Although the highest reported *in vivo* τ 10, ϵ 12-CLA flows at the duodenum do not exceed 1.5 g/day (reviewed in Shingfield and Griinari, 2007), infusions up to 5 g/day of τ 10, ϵ 12-CLA were selected to have a sufficient number of data (only one publication was available with <1.5 g/day). For the effects of LCFA on milk fat, we used duodenal infusions of animal or plant lipids and excluded treatments with a change in dry matter (DM) intake higher than 1 kg/day following infusions, in order to avoid large changes in the supply of the other nutrients. The maximal change for duodenal glucose was set up at 1500 g/day (estimated from a maximal duodenal starch change of 2.6 kg/day (Overton *et al.*, 1995) and a small intestine starch digestibility of 60%).

There were 9, 11, 3, 14, 7, 8 and 10 publications for C2, C3, C4, glucose, proteins, LCFA and τ 10, ϵ 12-CLA (list in supplementary material).

Databases and statistical analyses

The database was divided into seven sub-databases, one for each nutrient studied. In each sub-database, data were encoded according to studies. Descriptive statistics (the mean, s.d. and ranges of values) were generated for the main characteristics of the studies in each nutrient sub-database, together with the correlations between these characteristics. The differences between sub-databases concerning these characteristics were tested with GLM models.

We had two options for data statistical analysis: either study the absolute values of MFC and MFY before and after the infusions, using mixed models (St-Pierre, 2001); or study the response of MFC and MFY to the infusions. With a view to study the milk fat responses to dietary modifications, we chose to focus on the responses to the nutrient infusions.

The response to the infusions was thus chosen as the dependent variable and was calculated from the MFC and MFY of the infused cows and those of their respective controls. Thus, MFC and MFY responses were given by:

$$\Delta\text{MFC (g/kg milk)} = \text{MFC}_{\text{infused}} - \text{MFC}_{\text{control}}$$

$$\Delta\text{MFY (g/day)} = \text{MFY}_{\text{infused}} - \text{MFY}_{\text{control}}$$

However, for comparison purposes, we also run mixed models on the raw data (with study as a random effect, as recommended by St-Pierre, 2001).

The amount of nutrients supplied was expressed in kilograms per day, except for t10,c12-CLA , which was expressed in grams per day. It was not possible to express the amount of nutrients supplied relative to intake or body weight (BW), as several publications did not report BW or DM intake. No data on milk FA composition were available in the C2 and C4 sub-databases, which prevented us from determining the effects of these two nutrients on the milk FA composition. For lipid infusions, the resulting milk FA composition is directly dependent on the composition of the lipids infused, and thus is meaningless when averaged over different lipid sources. For this reason, milk FA responses are not reported for LCFA supplies. The percentages of 18:3 or odd-chain FA in milk were often unreported and therefore could not be studied for all the nutrients. In addition, because of the heterogeneity among studies in how milk FA were reported, certain data had to be reconciled, especially for C_{18} FA. Thus, for 18:1, 18:2 and 18:3, the values used were either reported overall values or the sums of the corresponding isomers when more detailed FA compositions were provided.

For each study, we thus had a response in MFC and MFY (Y), an amount of nutrient infused (X), and several variables describing the animals, diets, experimental design, etc. used in the study. Statistical analyses (GLM models in Minitab® Statistical Software, version 15 – Minitab Inc., 2007) were performed in two steps. First, the relationships between Y (responses of MFC, MFY or milk FA percentage) and the explanatory variable X (amount of the nutrient supplied) were studied with GLM models without intercept: $Y = b \times X$, where b is the slope of the relationship. Quadratic models were also tested and compared with the linear models. The normality of the residuals was verified using the Shapiro–Will test. Outliers were identified on the basis of residuals, HI leverage and Cook's distance. Graphical examinations were also used at each stage of the meta-analysis process to check that the statistical results were not distorted by some extreme data.

Second, the residuals of these models were regressed on the other variables describing the study, the major potential quantitative interfering factors. The interfering factors are variables that could modify the mean responses to the nutrient supplied. The tested interfering factors were variables describing animals (BW, DM intake (DMI), stage of lactation, milk yield (MY), milk protein and lactose concentrations, etc.), diet composition (proportion of concentrates, dietary

concentrations of protein, fat and NDF, etc.), duration of the experimental period, MFC and MFY of the respective control treatment ($\text{MFC}_{\text{control}}$ and $\text{MFY}_{\text{control}}$) and milk FA percentage of the control treatment ($\% \text{FA}_{\text{control}}$). For qualitative interfering factors, ANOVA were run on the residuals to test for the influence of breed, addition of buffers (only for VFA infusion), main forage, type of infusions (isoenergetic or not), CLA supplement form (pure FA or FA mixture) and type of proteins infused (form of casein or plant protein). When a significant interfering factor was detected on the residuals, it was included as an additional variable in the model, in order to establish whether its inclusion improved or not the residual mean square errors (RMSE) and the adjusted R^2 (R^2_{adj}).

We also run mixed models on the raw data, using the 'Mixed' procedure of SAS (Statistical Analysis Systems Institute, 2000), with study as a random effect. In these models, the nutrient supplies for the control treatments were set up at zero (they were neither reported nor calculable from the publications).

All model parameters and correlations were considered significant at $P < 0.05$, whereas $P < 0.10$ indicated a trend.

Results

Meta-design and description of the sub-databases

All the experiments investigated the effects of the nutrient infusions on MY and composition. Within an experiment, diet and dietary parameters (proportion of concentrates, type of forage, crude protein and NDF concentration) were similar between treatments. Most of the experiments were designed as Latin squares (79% of the designs) and used Holstein cows (77% of the experiments). In all the experiments, cows were milked twice daily. On average, milk composition was determined on 5 days or on 10 consecutive milkings.

Table 1 presents, for each nutrient sub-database, the means and the s.d. of the main characteristics of the control treatments. The proportion of concentrates in the diet was on average 43.9% with no difference between the sub-databases. The stage of lactation differed between sub-databases: the cows used for glucose studies were in early lactation (73 days); for VFA, LCFA and protein studies, they were in mid-lactation and for t10,c12-CLA in late lactation (199 days). The duration of treatments ranged from 4 to 28 days; the experimental periods for t10,c12-CLA studies were significantly shorter (6 days on average) than for the other nutrients (12 to 20 days on average). The DMI, MY and MFY of the control treatments differed between the sub-databases: they were lower in the C2 and C4 sub-databases than for the other nutrients. This is presumably due to the earlier dates of publication for these two nutrients (mostly before 1975). Control MFC was about 40.0 g/kg milk for the VFA, glucose, LCFA and protein studies, but was significantly lower for the t10,c12-CLA sub-database (33.9 g/kg). These differences also appeared in the intercepts of the mixed models (Table A in Supplementary material).

Table 2 presents, for each nutrient, the means, the s.d. and ranges of the amounts of nutrient supplied and the

Table 1 Description of the sub-databases used for the meta-analysis of milk fat responses to variations in the supply of seven nutrients

Sub-database ^b	Number of treatments	Number of control treatments	Stage of lactation (days) ^a	Proportion of concentrates (%) ^a	Data for the control treatments ^a				
					DM intake (kg/day)	MY (kg/day)	Milk fat (g/kg)	Milk fat (g/day)	Milk protein (g/kg)
C2	46	22	102 ± 47	44.8 ± 15.2	12.9 ± 2.4	14.3 ± 3.2	41.8 ± 5.8	590 ± 145	34.2 ± 3.0
C3	33	16	115 ± 48	40.3 ± 12.9	15.6 ± 3.9	21.4 ± 7.8	39.4 ± 3.9	834 ± 284	31.4 ± 2.3
C4	11	5	147 ± 5	49.2 ± 6.6	10.9 ± 2.8	16.1 ± 3.5	40.4 ± 4.0	644 ± 115	33.3 ± 0.9
Glucose	43	18	73 ± 42	45.0 ± 10.1	17.5 ± 3.4	27.4 ± 6.4	40.3 ± 4.2	1080 ± 229	30.2 ± 2.3
†10,c12-CLA	23	10	199 ± 50	48.3 ± 5.8	21.9 ± 2.1	26.7 ± 4.9	33.9 ± 2.8	896 ± 167	29.7 ± 3.6
LCFA	25	9	135 ± 33	49.5 ± 6.7	19.2 ± 2.6	25.9 ± 4.1	38.0 ± 7.0	962 ± 194	32.8 ± 2.7
Proteins	82	36	121 ± 67	39.7 ± 13.9	16.0 ± 4.0	21.3 ± 6.9	41.5 ± 6.8	843 ± 211	32.0 ± 3.0

DM = dry matter; MY = milk yield; C2 = acetic acid; C3 = propionic acid; C4 = butyric acid; CLA = conjugated linoleic acid; LCFA = long-chain fatty acids.

^aData are reported as mean ± s.d.

^bThe full list of the references used in the sub-databases is available as Supplementary material.

Table 2 Overview of the data reported in the publications: mean ± s.d. (and range of values) of the amount of nutrients supplied (Δ nutrient in kg/day, except †10,c12-CLA in g/day) and the responses of DM intake (Δ DM intake, kg/day), MY (Δ MY, kg/day), MPC (Δ MPC, g/kg), MFC (Δ MFC, g/kg) and MFY (Δ MFY, g/day)

Nutrients	<i>n</i> ^a	Δ Nutrient	Δ DM intake	Δ MY	Δ MPC	Δ MFC	Δ MFY
C2	24	+0.72 ± 0.47 (0.16 to 1.60)	+0.35 ± 0.63 (-1.15 to 1.30)	+0.83 ± 0.72 (-0.80 to 2.00)	+0.11 ± 0.72 (-1.50 to 1.10)	+1.86 ± 1.79 (-1.00 to 5.60)	+63 ± 41 (0 to 165)
C3	17	+0.75 ± 0.23 (0.25 to 0.99)	+0.05 ± 0.46 (-1.00 to 0.70)	-0.06 ± 1.15 (-2.40 to 1.40)	+0.43 ± 1.14 (-1.20 to 3.00)	-3.66 ± 1.38 (-6.30 to -1.10)	-80 ± 60 (-195 to 0.0)
C4	6	+0.54 ± 0.07 (0.40 to 0.60)	+0.46 ± 0.05 (0.42 to 0.50)	-0.09 ± 0.82 (-0.90 to 1.40)	+0.45 ± 0.78 (0.00 to 1.9)	+4.23 ± 1.60 (2.10 to 6.80)	+68 ± 47 (30 to 160)
Glucose	25	+0.78 ± 0.44 (0.25 to 1.50)	-0.68 ± 1.12 (-2.80 to 1.10)	+0.61 ± 0.98 (-1.30 to 2.40)	+0.52 ± 0.75 (-0.80 to 2.0)	-3.15 ± 1.7 (-6.10 to -0.40)	-72 ± 76 (-224 to 81)
†10,c12-CLA	13	+3.59 ± 1.27 (1.25 to 5.00)	-0.72 ± 0.94 (-2.80 to 0.70)	+0.57 ± 1.27 (-2.40 to 2.70)	+0.10 ± 0.80 (-1.10 to 2.0)	-8.87 ± 3.34 (-13.70 to -2.10)	-227 ± 83 (-370 to -70)
LCFA	16	+0.30 ± 0.15 (0.07 to 0.50)	-0.09 ± 0.54 (-1.00 to 0.68)	+0.14 ± 1.32 (-1.90 to 2.40)	+0.07 ± 0.60 (-1.00 to 1.00)	+2.81 ± 2.98 (-2.50 to 8.80)	+77 ± 83 (-83 to 299)
Proteins	46	+0.32 ± 0.14 (0.09 to 0.67)	+0.08 ± 0.41 (-1.10 to 0.70)	+1.89 ± 1.10 (0.20 to 5.00)	+1.03 ± 1.09 (-1.80 to 4.4)	-1.49 ± 1.67 (-4.20 to 2.30)	+56 ± 52 (-40 to 181)

CLA = conjugated linoleic acid; DM = dry matter; MY = milk yield; MPC = milk protein concentration; MFC = milk fat concentration; MFY = milk fat yield; LCFA = long-chain fatty acids; C2 = acetic acid; C3 = propionic acid; C4 = butyric acid.

^a*n* = number of difference between a supplemented treatment and its respective control.

responses of DMI, MY, milk protein concentration, MFC and MFY reported in the publications. The amount of nutrients supplied was well distributed over the ranges studied, except for the C3 and †10,c12-CLA sub-databases in which there were few responses with small quantities supplied. Only six data were available for C4.

C2 and protein infusions significantly increased MY:

$$\Delta\text{MY (kg/day)} = 0.96^{***} (\pm 0.18) \times \Delta\text{C2 (kg/day)},$$

$$(n = 24, R^2_{\text{adj}} = 0.54 \text{ and RMSE} = 0.73 \text{ kg/day}),$$

$$\Delta\text{MY (kg/day)} = 5.34^{***} (\pm 0.46) \times \Delta\text{proteins (kg/day)},$$

$$(n = 46, R^2_{\text{adj}} = 0.74 \text{ and RMSE} = 1.10 \text{ kg/day}).$$

The other nutrients did not modify the MY. Milk protein concentration increased with protein infusions (+2.97 g/kg

per kg of added protein, $P < 0.001$, $n = 46$ and RMSE = 1.07 g/kg) and glucose infusions (+0.77 g/kg per kg of added glucose, $P < 0.001$, $n = 23$ and RMSE = 0.57 g/kg).

Milk fat responses to the nutrients

Table 3 presents, for each nutrient, the adjusted models of the MFC and MFY responses, R^2_{adj} and RMSE, and indicates the number of data used and the significant interfering factors. Very few outliers were excluded (one each in the C2, C4 and glucose sub-databases). Figure 1 presents the individual responses (within-study differences) and the adjusted models of MFC responses to the supply of each nutrient. The models of absolute values of MFC and MFY following nutrient infusions, adjusted with mixed models (random study effect), are provided in Table A in Supplementary material. The linear coefficients adjusted with these

Table 3 Models of the response of MFC (Δ MFC) and MFY (Δ MFY) to ruminal infusions of C2, C3 and C4, and to duodenal infusions of glucose, proteins, plant or animal lipids (LCFA) and t10,c12-CLA, all expressed in kg/day (except t10,c12-CLA in g/day)

Response	Nutrients	<i>n</i>	Linear coefficient ^a	Quadratic coefficient ^a	RMSE ^b	R^2_{adj} ^b	Interfering factors on the residuals ^c
Δ MFC (g/kg)	C2	24	+2.54 ± 0.33***	ns	1.37	0.71	%C [†]
	C3	17	-4.83 ± 0.32***	ns	1.02	0.93	MFC _{control} ***, period length*
	C4	6	+7.96 ± 0.90***	ns	1.21	0.92	–
	Glucose	25	-6.63 ± 1.04***	2.51 ± 0.84**	1.27	0.88	MFC _{control} [†]
	t10,c12-CLA	13	-2.38 ± 0.21***	ns	2.84	0.91	%F [†] , NDF _f [†]
	LCFA	16	+9.44 ± 1.90***	ns	2.56	0.59	–
	Proteins	46	-3.46 ± 0.78***	ns	1.88	0.28	PI _{control} ***, MY*, DMI*
Δ MFY (g/day)	C2	24	+75.5 ± 9.5***	ns	39.6	0.72	DIM*
	C3	17	-107.2 ± 17.1***	ns	55.0	0.69	–
	C4	6	+127.7 ± 33.4**	ns	44.6	0.68	–
	Glucose	25	-93.5 ± 13.6***	ns	61.3	0.65	–
	t10,c12-CLA	13	-61.1 ± 4.9***	ns	68.1	0.92	%F*, NDF _f *†
	LCFA	16	+246.1 ± 57.0***	ns	76.7	0.53	–
	Proteins	46	+162.9 ± 21.2***	ns	50.6	0.56	MFC _{control} ***

MFC = milk fat concentration; MFY = milk fat yield; C2 = acetic acid; C3 = propionic acid; C4 = butyric acid; CLA = conjugated linoleic acid; LCFA = long-chain fatty acids; %C = proportion of concentrates; MFC_{control} = milk fat concentration of the control treatment; %F = proportion of forage; NDF_f = forage NDF concentration in g/kg DM; PI_{control} = protein intake in the control treatment; MY = milk yield; DMI = dry matter intake; DIM = days in milk.

^aAdjusted coefficient ± s.e.

^bRMSE = residual mean square error; R^2_{adj} : adjusted R^2 .

^cInterfering factors: factors describing animal or diet or experiment, that were significantly correlated with the residuals of the models, expressing an effect on the mean response to the nutrient.

*** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$.

[†]Tendency at $P < 0.1$; ns at $P > 0.1$.

models were very similar to those of the responses to infusions (Table 3), and accordingly only the response of MFC to glucose presented a significant quadratic coefficient. However, the RMSE of the mixed models were lower than those of the within-experiment responses. Table 4 shows the adjusted models of the responses of the milk FA percentages.

Responses to C2 and C4. The ruminal infusions of C2 and C4 significantly increased MFC and MFY. The effects of C4 on MFC and MFY were higher than those of C2 (on a kg/day basis). The MFC response models presented no significant interfering factors on the residuals. However, the proportion of concentrates in the diet tended to increase the response of MFC to C2 ($P = 0.08$, +0.036 g/kg per kg of C2, for each % concentrate). The stage of lactation had a slight effect on the residuals of the MFY response to C2 ($P = 0.05$): the cows in late lactation tended to have a lower response.

Responses to C3 and glucose. The infusions of C3 and glucose decreased MFC and MFY ($P < 0.001$). These reductions were linear except for the MFC response to glucose, which was curvilinear. The MFC response to glucose was linear up to 1.0 kg/day with a mean slope of -4.7 g/kg per kg of added glucose. The MFC_{control} had an effect on the residuals of the MFC models (C3: $P = 0.004$; glucose: $P = 0.09$): the reduction in MFC was higher when MFC_{control} was high (i.e. >40.0 g/kg milk). We thus adjusted a new model with two response coefficients to these nutrients, which improved the

RMSE values and R^2_{adj} .

$$\Delta\text{MFC (g/kg milk)} = -4.28^{**} (\pm 0.43) \times \Delta\text{C3 (kg/day)},$$

if MFC_{control} < 40 g/kg milk,

$$\Delta\text{MFC (g/kg milk)} = -5.58^{**} (\pm 0.43) \times \Delta\text{C3 (kg/day)},$$

if MFC_{control} > 40 g/kg milk,

$$(n = 17, R^2_{adj} = 0.95 \text{ and RMSE} = 0.91 \text{ g/kg}).$$

$$\Delta\text{MFC (g/kg milk)} = -5.50^{**} (\pm 0.94) \times \Delta\text{glucose (kg/day)}$$

$$+ 2.01^{**} (\pm 0.77)$$

$$\times \Delta\text{glucose}^2 \text{ (kg/day)},$$

if MFC_{control} < 40 g/kg milk,

$$\Delta\text{MFC (g/100 g milk)} = -6.92^{**} (\pm 0.94)$$

$$\times \Delta\text{glucose (kg/day)}$$

$$+ 2.01^{**} (\pm 0.77)$$

$$\times \Delta\text{glucose}^2 \text{ (kg/day)},$$

if MFC_{control} > 40 g/kg milk,

$$(n = 25, R^2_{adj} = 0.90 \text{ and RMSE} = 1.13 \text{ g/kg}).$$

The length of the experimental period also had an effect on the residuals of the MFC response to C3 ($P = 0.031$): the reduction in MFC response with C3 was lower for the longer periods.

The effects of the two nutrients on milk FA composition were different. C3 decreased or tended to decrease the

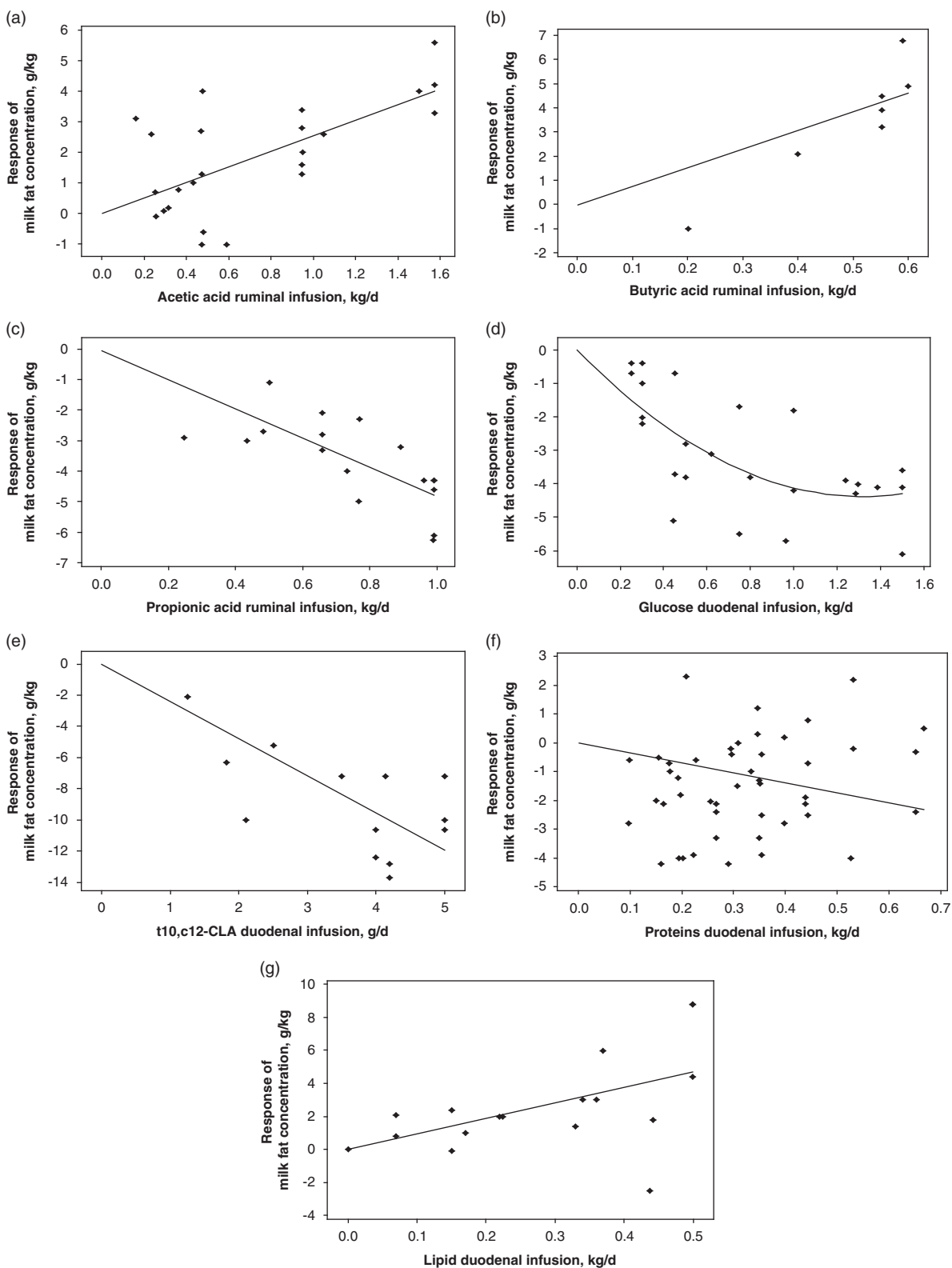


Figure 1 Responses of milk fat concentration (MFC; g/kg milk) to ruminal infusions of acetic (a), butyric (b) and propionic (c) acids, and to duodenal infusions of glucose (d), t10,c12-CLA (e), proteins (f) and plant or animal lipids (LCFA; g). The amount of nutrients infused is expressed in kilogram per day (except t10,c12-CLA in g/day). Each dot represents a difference between a supplemented treatment and the respective control treatment (amount of nutrient infused in X, MFC response in Y); the lines represent the adjusted models reported in Table 3.

Table 4 Responses of the milk FA percentages to ruminal infusions of C3 and duodenal infusions of glucose, τ 10,c12-CLA and proteins

Nutrients	Response	% FA _{control} ^a	n	Linear coefficient ^b	RMSE ^c	R ² _{adj} ^c
C3 (kg/day)	4:0	2.59 ± 0.12	8	-0.476 ± 0.051***	0.124	0.91
	6:0	1.97 ± 0.16	8	-0.315 ± 0.045***	0.109	0.86
	8:0	1.31 ± 0.12	8	-0.144 ± 0.047**	0.085	0.66
	10:0	3.48 ± 0.33	8	+0.087 ± 0.089 ^{ns}	0.217	ns
	12:0	4.27 ± 0.35	8	+0.182 ± 0.083 [†]	0.202	0.30
	14:0	13.72 ± 0.79	8	-0.378 ± 0.190 [†]	0.463	0.25
	16:0	36.22 ± 2.65	8	+0.179 ± 0.565 ^{ns}	1.375	ns
	18:0	9.05 ± 1.33	8	-0.948 ± 0.248**	0.603	0.62
	18:1	17.69 ± 1.87	8	+0.522 ± 0.351 ^{ns}	0.855	ns
	18:2	2.08 ± 0.56	8	+0.028 ± 0.064 ^{ns}	0.156	ns
	15 + 17	2.83 ± 0.44	8	+1.072 ± 0.085***	0.208	0.95
Glucose (kg/day)	4:0	2.87 ± 0.31	17	-0.341 ± 0.030***	0.131	0.88
	6:0	2.16 ± 0.24	17	-0.089 ± 0.017***	0.073	0.62
	8:0	1.39 ± 0.16	17	+0.044 ± 0.017*	0.072	0.26
	10:0	3.36 ± 0.35	17	+0.449 ± 0.063***	0.274	0.74
	12:0	4.02 ± 0.45	17	+0.893 ± 0.076***	0.331	0.89
	14:0	13.00 ± 0.55	17	+0.963 ± 0.156***	0.675	0.68
	16:0	36.23 ± 1.89	17	+2.578 ± 0.350***	1.520	0.76
	18:0	9.45 ± 0.89	17	-2.182 ± 0.191***	0.824	0.88
	18:1	17.59 ± 1.93	17	-2.606 ± 0.276***	1.195	0.84
	18:2	2.20 ± 0.57	17	-0.107 ± 0.032***	0.137	0.38
	15 + 17	3.10 ± 0.26	17	+0.035 ± 0.032 ^{ns}	0.192	ns
τ 10,c12-CLA (g/day)	4:0	4.10 ± 1.15	10	+0.043 ± 0.058 ^{ns}	0.728	ns
	6:0	2.03 ± 0.27	10	-0.088 ± 0.015***	0.187	0.76
	8:0	1.10 ± 0.25	10	-0.057 ± 0.007***	0.082	0.88
	10:0	2.45 ± 0.83	10	-0.120 ± 0.015***	0.188	0.86
	12:0	2.88 ± 1.11	10	-0.082 ± 0.022**	0.275	0.56
	14:0	10.31 ± 2.22	10	-0.068 ± 0.043 ^{ns}	0.533	ns
	16:0	28.72 ± 3.57	10	-0.544 ± 0.071***	0.885	0.85
	18:0	11.07 ± 2.88	10	+0.399 ± 0.065***	0.811	0.78
	18:1	26.00 ± 5.71	10	+0.414 ± 0.122**	1.520	0.51
		τ 10,c12-CLA	0.002 ± 0.004	10	+0.040 ± 0.002***	0.019
	18:3	0.476 ± 0.302	9	+0.020 ± 0.015 ^{ns}	0.178	ns
	15 + 17	1.62 ± 0.48	10	-0.005 ± 0.025 ^{ns}	0.316	ns
Proteins (kg/day)	4:0	3.99 ± 1.19	17	+0.294 ± 0.325 ^{ns}	0.536	ns
	6:0	2.35 ± 0.42	17	+0.163 ± 0.085 [†]	0.139	0.13
	8:0	1.39 ± 0.20	17	+0.200 ± 0.081*	0.134	0.23
	10:0	2.26 ± 0.46	17	+0.791 ± 0.183***	0.302	0.51
	12:0	3.90 ± 0.62	17	+0.899 ± 0.184***	0.303	0.57
	14:0	11.92 ± 1.34	17	+1.391 ± 0.467**	0.769	0.31
	16:0	36.32 ± 4.94	17	-2.862 ± 1.302*	2.150	0.18
	18:0	9.04 ± 2.76	17	-0.624 ± 0.316 [†]	0.520	0.14
	18:1	18.33 ± 4.01	17	-0.485 ± 0.700 ^{ns}	1.155	ns
	18:2	1.90 ± 0.55	17	-0.230 ± 0.109 [†]	0.180	0.16

FA = fatty acid; C3 = propionic acid; CLA = conjugated linoleic acid.

^a%FA_{control} = means ± s.d. for control (unsupplemented) treatments (expressed in g/100 g total FA).

^bAdjusted coefficient ± s.e.

^cRMSE = residual mean square error; R²_{adj} = adjusted R².

***P < 0.001; **P < 0.01; *P < 0.05.

[†]Tendency at P < 0.1; ns at P > 0.1.

percentage of several even-chain FA (4:0, 6:0, 8:0, 12:0, 14:0 and 18:0, P < 0.1, n = 8), whereas glucose decreased the percentage of short-chain FA (SCFA; 4:0 and 6:0, P < 0.001, n = 17) and LCFA (all C₁₈, P < 0.001, n = 17) and increased the percentage of medium-chain FA (MCFA; 10:0 to 16:0,

P < 0.05, n = 17). C3 increased the percentage of the odd-chain FA (+1.07% of C₁₅ + C₁₇ per kg/day of C3, P < 0.001, n = 8), but glucose did not alter them. No interfering factors had an effect on the milk FA responses to C3. The responses of 4:0, 14:0, 16:0 and C₁₈ FA to glucose were

influenced by their percentage with the control treatment ($P < 0.05$). The increases were higher when the initial percentage was low, and conversely, the reductions were higher when the initial percentage was high.

Response to LCFA. The infusions of animal or plant lipids linearly increased MFC and MFY. These models had higher RMSE and lower adjusted R^2 compared with the VFA, glucose and CLA models. No significant interfering factor was identified on the MFC and MFY response models.

Response to t10,c12-CLA . The MFC and MFY decreased linearly with t10,c12-CLA infusions (up to 5 g/day). No interfering factors had a significant effect on the residuals of the MFC response. However, a high proportion of forage ($P = 0.084$, -0.25 g/kg per %forage) and a high forage NDF concentration ($P = 0.082$, -0.04 g/kg per g/kg DM) tended to promote the decrease in MFC caused by t10,c12-CLA . Similarly, the proportion of forage ($P = 0.012$, $n = 12$) and the forage NDF concentration ($P = 0.021$, $n = 12$) had a significant effect on the residuals of the MFY response. However, these two variables were not significant when introduced as covariables in the initial model. The infusions of t10,c12-CLA significantly decreased the percentage of all *de novo* synthesized FA ($P < 0.01$, $n = 10$) except for 4:0 and 14:0, and increased the percentage of the preformed FA (18:0 and 18:1, $P < 0.001$, $n = 10$). The percentage of milk t10,c12-CLA increased linearly with t10,c12-CLA infusions, by 0.04% per gram of t10,c12-CLA infused. No interfering factors influenced the milk FA responses to t10,c12-CLA .

Response to proteins. The duodenal infusions of proteins increased MFY and decreased MFC. Although these models were significant, they had low R^2_{adj} , which were 56% and 28%, respectively. Several interfering factors had a significant effect on the residuals of MFC response: the amount of protein intake of control treatment ($\text{PI}_{\text{control}}$, $P < 0.001$), DM intake ($P = 0.016$) and MY ($P = 0.014$). The reduction in MFC was lower for high $\text{PI}_{\text{control}}$, MY or DM intake. All these factors were correlated ($P < 0.05$). Inclusion of $\text{PI}_{\text{control}}$ as a covariate in the model (centered on the overall mean 1.52 kg/day) was the model that best improved RMSE and R^2_{adj} :

$$\begin{aligned} \Delta\text{MFC (g/kg milk)} &= -3.49^{***} (\pm 0.70) \\ &\quad \times \Delta\text{proteins (kg/day)} \\ &\quad - 1.68^{***} (\pm 0.47) \\ &\quad \times (1.52 - \text{PI}_{\text{control}} \text{ (kg/day)}), \\ (n = 46, R^2_{\text{adj}} &= 0.45 \text{ and RMSE} = 1.68 \text{ g/kg}). \end{aligned}$$

The MFY response to proteins was affected by $\text{MFC}_{\text{control}}$ ($P < 0.001$): the increase in MFY was lower for the low $\text{MFC}_{\text{control}}$. Inclusion of the $\text{MFC}_{\text{control}}$ as a covariable in the model (centered on the overall mean of 42.0 g/kg milk)

improved the RMSE and R^2 :

$$\begin{aligned} \Delta\text{MFY (g/day)} &= 164.4^{***} (\pm 18.7) \\ &\quad \times \Delta\text{proteins (kg/day)} \\ &\quad - 3.40^{***} (\pm 0.93) \\ &\quad \times (42.0 - \text{MFC}_{\text{control}} \text{ (g/kg)}), \\ (n = 46, R^2_{\text{adj}} &= 0.66 \text{ and RMSE} = 44.8 \text{ g/day}). \end{aligned}$$

The percentage of SCFA and MCFA increased or tended to increase (6:0 to 14:0, $P < 0.1$) with protein infusions, whereas the percentage of 16:0, 18:0 and 18:2 tended to decrease. However, these models explained $< 50\%$ of the observed variability in the milk FA composition.

Discussion

Individual effects of the nutrients on milk fat

This study provides equations to estimate the responses of milk fat production and composition to changes in the supply of seven nutrients derived from digestion: C2, C4, C3, glucose, t10,c12-CLA , protein and LCFA. Very few interfering factors had a significant effect on the residuals. The RMSE of the models were similar to the inherent variability of milk fat measurements: they were lower than the day-to-day variations in milk fat measured *in vivo* (Syrstad, 1977; Forsback *et al.*, 2010) and were similar to the residual variations (r.s.d.) calculated from the publications included in the database (the r.s.d. were calculated from the s.e.m. reported in the publications and the number of animals in the experimental groups): between 1.21 and 7.8 g/kg for MFC, and between 35 and 360 g/day for MFY. The nutrients differed in their effects on MFC and MFY.

Ruminal infusions of C2 and C4 linearly increased MFC and MFY. This positive effect of C2 and C4 stems from their role of substrate for the mammary FA synthesis (Bauman and Davis, 1974; Barber *et al.*, 1997). Our models showed that the effects of C2 on milk fat secretion were lower than those of C4. This finding could be explained by the specific role of β -hydroxybutyrate (synthesized from C4) as a precursor to initiate *de novo* FA elongation (Palmquist *et al.*, 1969; McCarthy and Smith, 1972). However, the effect of C2 on MFC could be slightly underestimated because C2 significantly increased MY (dilution effect), unlike C4. In the database, most C2 infusions represented an energy supplement compared with the controls, which could explain this increase in MY. Owing to a lack of published data, the milk FA composition responses to C2 and C4 could not be quantified. However, Storry and Rook (1965) observed an increase in the percentage of all FA from 4:0 to 16:0 and a decrease in C_{18} FA after ruminal infusions of C2 and C4.

C3 and glucose infusions decreased MFC and MFY. The observed milk fat reduction used to be explained by an increase in insulin secretion, causing FA to be preferentially used by adipose tissues rather than the mammary gland (review in Bauman and Griinari, 2001). However, experiments

using hyperinsulinemic–euglycemic clamps (Griinari *et al.*, 1997; Corl *et al.*, 2006) showed that insulin could not be fully responsible for the observed milk fat reduction. Thus, the mechanisms involved in milk fat reduction with C3 and glucose remain unclear, but seem to differ, as the resulting milk FA compositions are different. The reduction in milk fat with glucose is mainly due to a decrease in the yield of LCFA, whereas with C3 the reduction in milk fat is associated with a decrease in the yield of all even-chain FA. Both nutrients decreased the plasma concentrations of milk fat precursors: C2, β -hydroxybutyrate, non-esterified FA and total glycerides (Rigout *et al.*, 2003; Lemosquet *et al.*, 2009a). Hurtaud *et al.* (1998) and Rigout *et al.* (2002) proposed that the reduction in LCFA with glucose could also be due to a specific inhibiting effect of glucose on adipose tissues mobilization, leading to a lower contribution of LCFA to milk fat, or a decrease in lipoprotein lipase activity and mammary FA esterification. The responses of MFC to C3 and glucose were dependent on MFC_{control}: the reductions were lower when MFC_{control} was low. In our database, the low MFC_{control} values were associated with starch-rich diets, which induced high digestive flows of C3 or glucose, and possibly FA biohydrogenation products. The effects of C3 and glucose may thus be saturable: less effect would be observed with diets inducing high digestive flows of these nutrients.

To estimate the positive effects of LCFA on milk fat (LCFA are a substrate for milk fat synthesis), we used duodenal infusions of animal and plant lipids, in order to avoid the effects of dietary lipids on rumen fermentations. The responses of MFC and MFY were the highest of the nutrients studied: +9.4 g/kg and +246.1 g/day per kg, respectively, of added lipids. In a preliminary study of dietary protected tallow supplementation (as a proxy of LCFA supply, data not shown), the responses were significantly lower than those of duodenal infusions (+4.35 g/kg and +159.2 g/day per kg/day, respectively, of added tallow for MFC and MFY). The figures for duodenal infusions are also much higher than what is reported for dietary plant lipid supplementation (e.g. Glasser *et al.*, 2008). The higher responses found in this study probably result from the fact that duodenal infusions avoid the detrimental effects of dietary lipids on rumen fermentations, diet digestibility and milk fat synthesis (through the production of *trans* FA in the rumen) and also from the selection of infusion studies in which lipid infusions did not depress DMI to a large extent. The intestinal absorption of the lipids infused (mainly as triglycerides) was probably not limiting in this data set, given the magnitude of the response. Moreover, most lipid sources used in the sub-database were rich in saturated and *cis* monounsaturated FA, which are highly incorporated in milk lipids (unlike some *trans* or polyunsaturated FA that can inhibit mammary lipogenesis). These responses obtained with duodenal infusions thus correspond to a maximum positive effect of LCFA supply on milk fat synthesis (which was our objective), without the side effects observed with diet supplementation (decrease in DMI and sometimes diet digestibility, rumen production of

FA biohydrogenation isomers that inhibit mammary lipogenesis, etc.). The data set used was relatively limited, and did not enable a detailed study of interfering factors. The stage of lactation, for example, is known to have an effect on the response to lipid infusion, cows in early lactation being less responsive to lipid supplementation (Gagliostro and Chilliard, 1991). The effects of lipid supplements on milk FA composition have been extensively reviewed (e. g. Chilliard *et al.*, 2001 and 2007). They generally increase the percentage of C₁₈ FA in milk and decrease the SCFA and MCFA. The reduction in *de novo* synthesized FA could be linked to an inhibition of *de novo* synthesis by LCFA (Barber *et al.*, 1997) or a substitution of the SCFA and MCFA by LCFA on milk triglycerides (Hansen and Knudsen, 1987). Within this general frame, their precise effects depend on the source and presentation of the supplement, and will not be detailed here.

Duodenal infusions of τ 10, τ 12-CLA linearly decreased MFC and MFY. This isomer is naturally present in duodenal contents, but in low amounts (*in vivo* data do not exceed 1.5 g/day, Shingfield and Griinari, 2007). It is considered as one of the isomers responsible for diet-induced MFD, and the only one for which extensive data are available (Shingfield *et al.*, 2010). De Veth *et al.* (2004) and Shingfield and Griinari (2007) found an exponential decay model to describe the relationship between the changes in MFY and the duodenal infusions of τ 10, τ 12-CLA. However, these authors included very high doses of τ 10, τ 12-CLA in their models (i.e. >5 g/day). In this study, the highest doses of τ 10, τ 12-CLA were excluded and the relationships were linear, and thus are likely applicable to the range of values of duodenal flows measured *in vivo*. The τ 10, τ 12-CLA infusions reduced the secretion of all FA, the reduction being generally greater for the *de novo* synthesized FA. Several recent publications (Harvatine and Bauman, 2006; Gervais *et al.*, 2009) have shown that τ 10, τ 12-CLA decreases milk fat through a transcriptional downregulation of enzymes and proteins involved in mammary lipid synthesis (see also Shingfield *et al.*, 2010).

Duodenal infusion of proteins increased MFY and decreased MFC. We are not aware of any meta-analysis on the effects of dietary protein intake on milk fat, which could be used for comparison. The supply of proteins increased mammary lipid synthesis, but this increase was proportionally smaller than the increase in MY, so that MFY increased, whereas MFC decreased. The mechanism explaining the increase in milk fat synthesis after protein supplementation is unclear. Proteins seem to stimulate the *de novo* FA synthesis, as suggested by the increase in the percentages of SCFA and MCFA (and the consecutive decrease in LCFA percentages). This increase in SCFA and MCFA could be linked to an increase in arterial concentrations or extraction rates of some mammary FA synthesis precursors (C2 and non-esterified FA: Guinard *et al.*, 1994; β -hydroxybutyrate: Vanhatalo *et al.*, 2003; Lemosquet *et al.*, 2009b). The hypothesis of dietary protein stimulating lipomobilization is not consistent with the observed decrease in LCFA percentages. The models of milk fat responses to proteins showed the lowest R^2_{adj} and were influenced by

numerous interfering factors in comparison with the other nutrients. This underlines the fact that proteins have only an indirect effect on milk fat secretion.

Compared effects of the nutrients on milk fat

The data used to establish the response equations were different between nutrient sub-databases. However, we checked many putative interfering factors (including animal, diet and other experimental characteristics), and included them in the models when they were significant. This, plus the fact that the models are based on within-experiment responses, ensures that our models are generic. Thus, it is coherent to compare the responses between nutrients, or even use them together if necessary. However, it is not relevant to compare directly the coefficients of the response models, because the *in vivo* flows of the nutrients differ greatly. To compare their potential *in vivo* effects following dietary changes, the equations had to be adjusted to the digestive flows of each nutrient. To this end, the maximal changes of these flows following dietary changes were estimated from published experiments studying the changes in nutrient production: 1.5 kg/day for C2, 1.0 kg/day for C3, 0.5 kg/day for C4, 1.5 kg/day for glucose, 0.8 kg/day for proteins, 1.5 g/day for CLA and 1.0 kg/day for LCFA. By applying our response equations to these maximal changes in digestive flows, the maximal responses of MFC (in g/kg milk) would be +3.8 for C2, -4.8 for C3, +4.0 for C4, -4.3 for glucose, -2.7 for proteins, -3.6 for τ 10, τ 12-CLA and +9.4 for LCFA, respectively. These maximal effects are moderate (all lower than 5 g/kg except for LCFA, when reductions up to 15 g/kg are observed following dietary changes). Proteins have the lowest effect, consistent with an indirect effect of proteins on milk fat. These results imply that none of these nutrients alone can fully explain MFD. Hence, several of these nutrients must be contributing simultaneously to the changes in milk fat observed following dietary changes, or additional nutrients must also be acting, or there are some synergistic interactions between nutrients (or some combination of these mechanisms).

Potential effect of additional nutrients

Some other nutrients, not studied here, could be involved in milk fat changes after dietary modification. Two other CLA isomers have been identified as inhibitors of milk fat synthesis in dairy cows: τ 9, τ 11-CLA, with less potent effects than τ 10, τ 12-CLA (Perfield *et al.*, 2007); possibly τ 10, τ 12-CLA (Saebø *et al.*, 2005), but with indirect proofs and despite any increase of this isomer with MFD diets (Shingfield *et al.*, 2010). These two CLA isomers have been examined in only one study at one dose and therefore their effects remain to be confirmed.

The inhibiting role of some *trans*-18:1 isomers is also suspected, but it has not been thoroughly studied owing to the technical difficulty of obtaining pure preparations of these isomers. Among these *trans* FA, some authors have suggested that τ 10-18:1 was the most probable candidate responsible for the inhibitory effects on milk fat synthesis

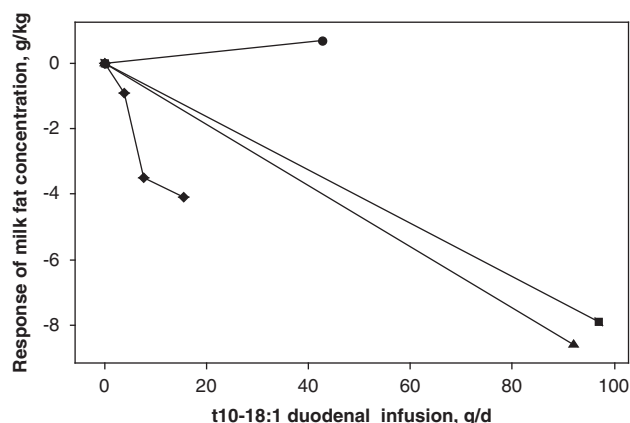


Figure 2 Milk fat concentration response (g/kg milk) to duodenal infusions of τ 10-18:1 (g/day) in lactating dairy cows using data from Shingfield *et al.* (2009; ▲); Lock *et al.* (2007; ●); Piperova *et al.* (2004; ◆); Romo *et al.* (2000; ■).

(Griinari *et al.*, 1998; Piperova *et al.*, 2004, Roy *et al.*, 2006). Only four infusion studies are available for τ 10-18:1, and they report inconsistent effects (Figure 2 shows the relationship between the changes in MFC and the amount of τ 10-18:1 infused): the postprandial infusion of a relatively pure (95%) τ 10-18:1 preparation (Lock *et al.*, 2007) had no effect on MFC and MFY; the other experiments (Romo *et al.*, 2000; Piperova *et al.*, 2004; Shingfield *et al.*, 2009) observed a reduction in milk fat secretion. However, these three experiments used FA mixtures in which τ 10-18:1 represented only between 9% to 37% of the total FA supplied. The effect of τ 6+ τ 7+ τ 8-18:1 has been suspected in cows (Kadegowda *et al.*, 2008) and an inhibition of mammary lipogenesis by τ 7-18:1 has been recently shown in mice (Kadegowda *et al.*, 2010). The isomers τ 6+ τ 7+ τ 8-18:1 are present in the FA mixtures used by Piperova *et al.* (2004) and Romo *et al.* (2000) and could contribute to the observed inhibition. Further studies are required to confirm the putative effects of τ 10-18:1 and τ 6+ τ 7+ τ 8-18:1 on milk fat synthesis.

Correlations between individual milk FA and MFC suggest that several other *trans* or CLA isomers may be involved in MFD (Loor *et al.*, 2005; Roy *et al.*, 2006; Kadegowda *et al.*, 2008). However, their direct effects on milk fat secretion remain to be established.

Interactions between the nutrients

This study quantified the individual effects of nutrients on milk fat secretion. However, these nutrients vary simultaneously after dietary changes. They have a combined action on milk fat synthesis and it is not known whether they have additive or interactive effects. There are very few reports of simultaneous infusions of several of these nutrients into dairy cows. Rook *et al.* (1965) studied, in the same experimentation, the effects of VFA infused separately or simultaneously into dairy cows. Their results show that the effects of C2, C3 and C4 on MFC and MFY are additive. Maxin *et al.* (2010) also observed additive effects of C3 and τ 10, τ 12-CLA

on milk fat and FA concentration and yield. The linearity of the responses to each nutrient (except for glucose) also reflects a form of 'additivity' of the amounts supplied within each nutrient. If these findings were validated for other nutrients, they could be used to estimate the total milk fat changes following dietary modification, based on the additivity of the effects of the nutrients.

Conclusions and perspectives

This study gives equations to estimate milk fat production and composition responses to changes in the supply of seven nutrients derived from digestion. Very few interfering factors had a significant effect on residuals and RMSE were similar to the inherent variability in the *in vivo* measurement of milk fat. In addition, data are very scarce or old for C2 and C4: more recent data, if they were available, would help to improve the models.

This study is a first step toward predicting overall milk fat responses to dietary changes, provided that the changes in nutrient supply following dietary modifications can be predicted.

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