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## Total solids content drives high solid anaerobic digestion via mass transfer limitation

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1 **Total solids content drives high solid anaerobic digestion via mass transfer**  
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3  
4 **limitation**  
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24 **Abstract:**

25  
26 The role of the total solids (TS) content on anaerobic digestion was investigated in  
27  
28 batch reactors. A range of TS contents from 10 to 35% was evaluated, four replicates  
29  
30 were performed. The total methane production slightly decreased with TS  
31  
32 concentrations increasing from 10 to 25% TS. Two behaviors were observed at 30%  
33  
34 TS: two replicates had similar performances to that at 25% TS; for the two other  
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36 replicates, the methane production was inhibited as observed at 35% TS. This difference  
37  
38 suggested that 30% TS content corresponded to a threshold of the solids content, above  
39  
40 which methanogenesis was strongly inhibited. The Anaerobic Digestion Model No.1  
41  
42 (ADM1) was used to describe the experimental data. The effects of hydrolysis step and  
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44 liquid/gas mass transfer were particularly investigated. The simulations showed that  
45  
46 mass transfer limitation could explain the low methane production at high TS, and that  
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48 hydrolysis rate constants slightly decreased with increasing TS.  
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1       **Keywords**  
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3       Dry anaerobic digestion; total solids content; mass transfer coefficient; hydrolysis;  
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5       methanogenesis.  
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10       **1. Introduction**

11       The production of waste including municipal solid waste (MSW) has significantly  
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13       increased in recent years. For example, in France, MSW generation has more than  
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15       doubled in less than 50 years, from 175 kg year<sup>-1</sup> capita<sup>-1</sup> in 1960 to 354 kg year<sup>-1</sup>capita<sup>-1</sup>  
16  
17       in 2006 (ADEME, 2009). Different technologies are used for MSW treatment, such as  
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19       incineration or composting, but anaerobic digestion (AD) processes are particularly  
20  
21       interesting in terms of organic matter reduction by converting organic compounds into  
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23       biogas. The methane content in biogas represents a source of renewable energy.  
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26       Based on the total solids (TS) content of solid waste, three main types of technologies  
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28       have been developed: wet ( $\leq 10\%$  TS), semi-dry (10-20% TS) and dry ( $\geq 20\%$  TS)  
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30       processes. Dry technologies, also called “high-solid” anaerobic processes, are attractive  
31  
32       because the quantity of water added to the raw waste is substantially reduced, and  
33  
34       consequently, the digester size is minimized. However, difficulties in operating dry  
35  
36       anaerobic digestion at both laboratory and industrial scales are related to the high  
37  
38       concentration of total solids. Fernandez *et al.* (2008) showed that methane production  
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40       started at day 14 in a reactor with 20% TS and at day 28 in a reactor with 30% TS in  
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42       mesophilic dry anaerobic batch reactors treating the organic fraction of municipal solid  
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44       waste. The total methane production was 17% lower at 30% than at 20% TS. This result  
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46       is consistent with the one obtained by Forster-Carneiro *et al.*(2008), which showed  
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48       better performance of anaerobic reactors operated at 20% TS compared to 25% and  
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1 30% TS. It was suggested that a high TS content could reduce substrate degradation  
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3 and, therefore, biogas production (Fernandez *et al.*, 2008). Le Hyaric *et al.* (2011) used  
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5 a microbial metabolic intermediate, propionate, as substrate to study specifically the  
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7 methanogenic activity in dry reactors. The specific methanogenic activity increased  
8  
9 linearly by a factor of 3.5 when the moisture content increased from 65% to 82%  
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11 (corresponding to 35 to 18% TS, respectively).  
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14  
15 A high TS content also affects the physical properties of the digested solid waste. Water  
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17 distribution was investigated by Garcia-Bernet *et al.* (2011a) in biowastes and  
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19 associated digestates sampled in industrial dry AD plants. Even if the proportion of the  
20  
21 hydration and vicinal water fractions is small ( $0.1 \text{ g}_{\text{water}} \text{ gDM}^{-1}$ ) compared to the total  
22  
23 amount of water, the capillary water fractions of the digestates ranged from 2 to 2.5  
24  
25  $\text{g}_{\text{water}} \text{ gDM}^{-1}$ : this fraction can represent up to 60% of the total water for a digestate  
26  
27 having a TS = 20%. As a consequence, for dry AD systems, the high solids content  
28  
29 strongly affects the rheological behavior of the digestates (Battistoni, 1993; Battistoni *et*  
30  
31 *al.*, 1997; Garcia-Bernet *et al.*, 2011b). Digested media are visco-elastic materials  
32  
33 characterized by high yield stress levels, and yield stress increases with TS content  
34  
35 according to an exponential law (Garcia-Bernet *et al.*, 2011b). Digesters are thus  
36  
37 difficult to mix and homogenize. Indeed, Karim *et al.* (2005) showed that, when the  
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39 concentration of TS increases in reactors, mixing becomes more important for  
40  
41 improving the production of methane. Nevertheless, under unmixed conditions,  
42  
43 transport is likely governed by diffusion processes, which are strongly related to the  
44  
45 porosity of the media and, thus, to the water content. Therefore, the diffusive transport  
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47 resistance of soluble compounds (substrate or by-products) may have a strong influence  
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49 on anaerobic digestion performance in dry systems.  
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1 The aim of this study is to assess the impact of the total solids content on anaerobic  
2 digestion of solid waste. Anaerobic batch reactor experiments were performed with TS  
3 concentrations ranging from 10 to 35%. The methane production performances were  
4 assessed. To better evaluate the impact of the water content on anaerobic digestion, the  
5 Anaerobic Digestion Model No. 1 (ADM1) was then used to describe the experimental  
6 data. The objective of the model application was to test some hypotheses in order to  
7 understand why the TS content affects the global anaerobic digestion performance. The  
8 effects of hydrolysis step and liquid/gas mass transfer were more particularly  
9 investigated.

## 2. Materials and methods

### 2.1. Substrate characterization

#### 2.1.1. Van Soest fractionation

10 The content of cellulose, hemicellulose and lignin-like fractions in the substrate was  
11 analyzed according to the Van Soest procedure (Van Soest, 1963). After shredding and  
12 sieving, 1g of cardboard was placed in a Fiberbag system (Gerhardt Germany). The  
13 sequential fractionation procedure was performed for six samples as follows: (1) The  
14 soluble compounds were obtained by extraction with a neutral detergent ( $30 \text{ g L}^{-1}$   
15  $\text{C}_{12}\text{H}_{25}\text{NaO}_4\text{S}$ ;  $18.61 \text{ g L}^{-1}$   $\text{C}_{10}\text{H}_{14}\text{N}_2\text{Na}_2\text{O}_8, 2\text{H}_2\text{O}$ ;  $6.81 \text{ g L}^{-1}$   $\text{Na}_2\text{B}_4\text{O}_7, 10\text{H}_2\text{O}$ ;  $4.56 \text{ g L}^{-1}$   
16  $\text{Na}_2\text{HPO}_4$ ;  $10 \text{ mL L}^{-1}$   $\text{C}_6\text{H}_{14}\text{O}_4$ ) at  $100^\circ\text{C}$  for 60 min; (2) the hemicellulose-like  
17 compounds were extracted by an acid detergent ( $20 \text{ g L}^{-1}$   $\text{C}_{19}\text{H}_{42}\text{NBr}$ ;  $26.7 \text{ mL L}^{-1}$   
18  $\text{H}_2\text{SO}_4, 95\text{-}97\%$ ) for 60 min at  $100^\circ\text{C}$ ; (3) the lignin-like compounds were obtained by  
19 removing the cellulose-like compounds for 3 h with  $\text{H}_2\text{SO}_4$  (72%). The remaining  
20 fraction corresponds to lignin. At the end of each step, the extracted samples were

1 washed with deionized water and oven-dried at 100°C before the transition to the next  
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3 step.  
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### 8 *2.1.2. The biochemical methane potential*

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10 The biochemical methane potential (BMP) of the cardboard was assessed according to  
11 Angelidaki and Sanders (2004). The assays were conducted in triplicate (and one blank)  
12 in 600 mL serum bottles at 35° C, with a working volume of about 400 mL. The serum  
13 bottles were filled with synthetic growth medium containing nutrients and trace  
14 elements, and inoculated with granular sludge from a mesophilic anaerobic digester of a  
15 sugar factory. The final sludge concentration in the bottles was 20 gVS L<sup>-1</sup>. The bottles  
16 were loaded with 1 g of cardboard (corresponding to 0.77 gVS). Biogas production and  
17 composition were measured daily. The methane production is expressed under standard  
18 condition and accounts for the variation of the gas content in the headspace of the  
19 reactors. The calculated BMP accounts for the global methane production without the  
20 residual (endogenous) methane production measured with the blank assay.  
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### 38 **2.2. Batch preparation and operating conditions**

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40 A compact cardboard with a density of 1.42 kg m<sup>-3</sup>, branded “Cartonnages Michel”, was  
41 used as a substrate since cardboard represents usually the largest proportion of organic  
42 compounds in municipal solid waste (21.5%) after putrescible waste (32.2%) (ADEME,  
43 2009). The cardboard was shredded using a cutting mill SM-100 and sieved at 2 mm.  
44  
45 Experiments were carried out in 600 mL batch flasks with a working volume of 100  
46 mL. A mixture of cardboard, water, inoculum and oligo-elements was prepared to reach  
47 six TS contents from “wet” to “dry” anaerobic conditions: TS = 10, 15, 20, 25, 30, 35  
48 % . A substrate to biomass ratio S/X of 20 (w/w) was used to limit the influence of the  
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1 inoculum composition on the degradation, S and X representing the initial TS contents  
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3 of the substrate and the biomass, respectively. The inoculum corresponded to a leachate  
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5 of pressed MSW digestate sampled in an industrial plant treating MSW. One mL of an  
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7 oligo-element solution was added to the mixture. This oligo-element solution was  
8  
9 composed of: FeCl<sub>2</sub>, 4 H<sub>2</sub>O (2 g L<sup>-1</sup>), CoCl<sub>2</sub>, 6 H<sub>2</sub>O (0.5g L<sup>-1</sup>), MnCl<sub>2</sub>,4 H<sub>2</sub>O(0.1 g L<sup>-1</sup>),  
10  
11 NiCl<sub>2</sub>, 6 H<sub>2</sub>O (0.1 g L<sup>-1</sup>), ZnCl<sub>2</sub>(0.05 g L<sup>-1</sup>), H<sub>3</sub>BO<sub>3</sub>(0.05 g L<sup>-1</sup>), Na<sub>2</sub>SeO<sub>3</sub>(0.05 g L<sup>-1</sup>),  
12  
13 CuCl<sub>2</sub>, 2 H<sub>2</sub>O (0.04 g L<sup>-1</sup>), Na<sub>2</sub>MoO<sub>4</sub>, 2 H<sub>2</sub>O(0.01 g L<sup>-1</sup>)). The experiments were run  
14  
15 over 298 days under mesophilic conditions (35°C) without mixing. Each TS condition  
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17 was tested in four replicates.  
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## 25 **2.3. Analytical methods**

### 26 *2.3.1 Biogas quantification and composition analysis*

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28 Biogas production and composition were determined daily during the first two months,  
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30 and then once a week after the exponential phase of methane production.  
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33  
34 The biogas production was measured by the water displacement method and then  
35  
36 normalized according to the ambient temperature. The biogas composition was  
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38 determined using a gas chromatograph (Varian  $\mu$ GC-CP4900) by injecting a sample  
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40 volume of 2 mL. This gas chromatograph was equipped with two columns: a Molsieve  
41  
42 5A PLOT column for O<sub>2</sub>, N<sub>2</sub>, CH<sub>4</sub> and CO and a HayeSep A column for CO<sub>2</sub>  
43  
44 quantification. The calibration was carried out with a standard gas composed of 25 %  
45  
46 CO<sub>2</sub>, 2 % O<sub>2</sub>, 10 % N<sub>2</sub> and 63 % CH<sub>4</sub>. The temperatures were 30°C for the oven and  
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48 100°C for the injector and the thermal conductivity detector. The gas carrier of the flow  
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50 was Helium.  
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### 2.3.2 VFA and pH analysis

1 Volatile fatty acids (VFA) were analyzed at the end of the batch test. After  
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6 centrifugation (13000 rpm, 15 min), VFA concentrations were measured by a gas  
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8 chromatograph (Varian  $\mu$ GC-CP3900) equipped with a flame ionization detector (FID).  
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10 The column used was a semi-capillar Econocap FFAP (Alltech). The gas carrier of the  
11  
12 flow was nitrogen. The rest of the assay technique is described in Ganesh *et al.* (2010).  
13  
14 pH was measured directly on the digestate using a pH meter Eutech Instruments pH510  
15  
16 with Mettler Toledo InLab® Expert Pt1000 pH electrodes.  
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### 2.4. Mathematical model

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25 The Anaerobic Digestion Model No. 1 (ADM1) (Batstone *et al.*, 2002) was used to  
26  
27 describe the experimental data. The characterization of the shredded cardboard with  
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29 respect to ADM1 state variables, in particular carbohydrates and particulate inerts, was  
30  
31 based on the results of the Van Soest fractionation and the degradable fraction of fibers  
32  
33 (hemicellulose and cellulose). The degradable fraction of hemicellulose and cellulose  
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35 (major degradable constituents of cardboard) was calibrated with the gas curve at 10%  
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37 TS. The estimated value of 40% is of the same order of magnitude as the degradable  
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39 fraction of fibers reported by Koch *et al.* (2010) for grass silage. In the model, the state  
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41 variable for carbohydrates,  $X_{ch}$ , was considered to be composed of the biodegradable  
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43 fraction of hemicellulose and cellulose, and the soluble organic fraction. The non-  
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45 degradable fraction of hemicellulose and cellulose, as well as the lignin fraction were  
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47 considered as particulate inerts ( $X_I$ ). Overall,  $0.33 \text{ gCOD gTS}^{-1}$  were attributed to  $X_{ch}$   
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49 and  $0.85 \text{ gCOD gTS}^{-1}$  to  $X_I$ . Simulations were run in Matlab/Simulink (Version 7.3.0).  
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1 The model was calibrated to the experimental data of the batch experiments at 10% TS  
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3 using a trial and error approach. Compared to the standard parameters proposed by  
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5 Batstone and Keller (2003) for the anaerobic digestion of sludge, two main parameters  
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7 were modified in order to account for the degradation of the specific substrate (i. e.,  
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9 cardboard) at high total solids contents: the first-order hydrolysis rate constant for  
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11 carbohydrates and the volumetric liquid/gas mass transfer coefficient.  
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14  
15 The default value of the first-order hydrolysis rate constant for carbohydrates ( $k_{hyd,Xch}$ )  
16  
17 is  $10\text{ d}^{-1}$ . For wet anaerobic digestion conditions and under mesophilic conditions,  
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19 hydrolysis rate constants for cellulosic material (cellulose, office paper, cardboard,  
20  
21 newsprint) reported in the literature are about  $0.03$  to  $0.07\text{ d}^{-1}$  (Vavilin et al., 2008;  
22  
23 Liebetrau et al., 2004; Vavilin et al., 2004). However, the present assays were  
24  
25 conducted under semi-dry and dry conditions. At low moisture conditions, Pommier et  
26  
27 al. (2007) observed a strong dependence on the moisture level for both kinetic rates and  
28  
29 ultimate methane production from paper and cardboard waste samples. In a modeling  
30  
31 work, Qu et al. (2009) used a value of  $0.01\text{ d}^{-1}$  for the hydrolysis rate constant of office  
32  
33 paper at TS contents of 40 and 66%. In the present work, the first-order hydrolysis rate  
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35 constant for carbohydrates ( $k_{hyd,Xch}$ ) was, thus, reduced from its default value to  $0.01\text{ d}^{-1}$ .  
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38  
39 The volumetric liquid/gas mass transfer coefficient  $k_T$  (corresponding to the  $k_{La}$   
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41 variable in the model) was reduced from its default value ( $200\text{ d}^{-1}$ ; for wet anaerobic  
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43 digestion of activated sludge) to  $0.5\text{ d}^{-1}$  to account for a reduced gas transfer due the  
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45 pasty texture of the substrate-biomass mixture. To our knowledge, the volumetric  
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47 liquid/gas mass transfer coefficient has never been measured in dry anaerobic digestion  
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49 processes. The volumetric mass transfer coefficient  $k_{La}$  depends on two parameters:  $k_L$   
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51 corresponding to the mass transfer coefficient, and a corresponding to the specific  
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1 surface area. In dry and semi-dry anaerobic digestion processes, the volumetric mass  
2 transfer coefficient  $k_{La}$  is drastically reduced because of two main reasons:  
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6 (1) the solid-liquid/gas interface is low, because of the low biogas bubble  
7 generation, and because of the unmixed conditions.  
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9  
10 (2) The mass transfer coefficient term is also affected by the low moisture  
11 content. The following equation accounts for the difference between the  
12 diffusivity coefficients of soluble compounds in the digestate  $D_i$  (*digestate*)  
13 and in the water  $D_i$  (*water*):  
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$$20 \quad k_L(\text{digestate}) = k_L(\text{water}) \left( \frac{D_i(\text{digestate})}{D_i(\text{water})} \right)^{0.5} \quad (1)$$

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25 Diffusivity coefficients decrease drastically with the decrease of the porosity  
26 (i. e., the water content) and the increase of viscosity, which is also strongly  
27 related to the TS content (Battistoni, 1993; Battistoni *et al.*, 1997; Garcia-  
28 Bernet *et al.*, 2011b).  
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35 Based on these theoretical considerations, the order of magnitude of the volumetric  
36 mass transfer coefficient is strongly affected by the TS content. A  $k_T=0.5 \text{ d}^{-1}$  was  
37 considered for the assay conducted at TS = 10%.  
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### 45 **3. Results and Discussion**

#### 46 **3.1. Impact of TS content on anaerobic digestion performance**

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48 Figure 1 shows the cumulated methane production for the reactors operated at six  
49 different TS contents during the whole experiment. Each curve represents the average of  
50 four replicates, except for TS = 30% where two distinguished behaviors were observed.  
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57 On the first 7 days (shown in the insert in Figure 1), all the cumulated curves are  
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1 similar. The initial rate of anaerobic degradation was, thus, identical for all TS contents,  
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3 indicating that the intrinsic activity of the microbial consortium was the same for the  
4  
5 different conditions, with an average methane rate of about  $0.48 \pm 0.05 \text{ mL gVS}^{-1} \text{ d}^{-1}$ .  
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8 After the first 7 days, the cumulated methane production was inversely proportional to  
9  
10 the TS content (the first 40 days in Figure 1).  
11  
12

13 Then, from 30-40 days of operation, methane production rates increased for the  
14  
15 replicates having a TS content ranging from 10 to 25% (maximum methane rates are  
16  
17 shown in Figure 2a), indicating the beginning of the exponential growth phase. During  
18  
19 this phase, two different behaviors were observed at 30% TS. Two replicates followed a  
20  
21 similar trend to that of the experiments conducted at 35% TS (named 30%-a in Figure  
22  
23 1), whereas the two remaining replicates showed a behavior close to that at 25% TS  
24  
25 (named 30%-b in Figure 1). For 30%-b and 35% TS, the acceleration phase,  
26  
27 corresponding to the exponential growth phase, never started. The highest  $\text{CH}_4$   
28  
29 production was about  $197 \text{ mL gVS}^{-1}$  at 10% TS. Since the biochemical methane  
30  
31 potential (BMP) was about  $214 \text{ mL CH}_4 \text{ gVS}^{-1}$ , the biodegradation reaction was almost  
32  
33 completed at 10% TS.  
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40 Considering that the COD of the substrate was  $1.44 \text{ gCOD gVS}^{-1}$ , the BMP was, thus,  
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42  $149 \text{ mL CH}_4 \text{ gCOD}^{-1}$  and corresponded to about 40% of the theoretical BMP value of  
43  
44  $350 \text{ mL CH}_4 \text{ gCOD}^{-1}$  (Buffière *et al.*, 2008). The low BMP value was probably due to  
45  
46 the composition of the substrate: indeed only 73% of the TS content of this cardboard  
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48 corresponded to volatile solids, and the remaining 27% to a mineral non-biodegradable  
49  
50 fraction. Among the volatile solids (VS), the organic matter was composed of only 4%  
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52 of soluble fraction and 14% of hemicellulosic fraction: these two minor fractions  
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54 represent the most degradable part of the organic matter. The two additional and most  
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1 abundant fractions corresponding to cellulose and lignin represented 66% and 16% of  
2  
3 the organic matter, respectively. These two fractions are resistant to biodegradation,  
4  
5 especially under anaerobic conditions (Buffière *et al.*, 2008).  
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### 10 11 3.1.1. Methane production rate

12 Methane production rates were estimated from the cumulative methane production  
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14 curves. The maximum methane production rate is shown in Figure 2a for the different  
15  
16 operating conditions. The maximum methane rate decreased with increasing TS  
17  
18 contents. This result is consistent with previous work of Fernandez *et al.* (2010) who  
19  
20 showed that the maximum methane rate was higher at 20% than at 30% TS. In the  
21  
22 present work, the maximum methane rates were almost the same at 30%-b and 35% TS.  
23  
24 However, the maximum methane rates were measured during the first seven days after  
25  
26 the start-up of the experiment, corresponding therefore to the initial rate of anaerobic  
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28 digestion. This initial rate was identical for all TS concentrations, because it  
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30 corresponded to the intrinsic activity of the microbial consortium. For other TS  
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32 concentrations, the maximum methane rates were observed between 40 to 80 days of  
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34 experiment.  
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### 45 3.1.2. Maximum methane yield

46 The cumulative methane yields measured after 298 days are shown in Figure 2b. They  
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48 were clearly dependent on the TS content. Analyses of variance (ANOVA) were  
49  
50 performed, distinguishing three groups: the first group (G1), which included all  
51  
52 replicates of 10, 15, 20, 25% TS, was significantly different from the second group  
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54 (G2), which included 30%-a replicates (\*p = 0.0011); the third group (G3), which  
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1 included replicates of 30%-b and 35% TS, was different from G1 and G2, with a very  
2 significant\*\*\*p of  $2.2 \cdot 10^{-16}$  and  $2.02 \cdot 10^{-6}$ , respectively (\*p < 0.05; \*\*p < 0.005; \*\*\*p <  
3  $5 \cdot 10^{-5}$ ; n.s., not significant (p >0.05)). The methane production was inversely correlated  
4 to the TS concentration. The methane production was  $176 \pm 7$ ,  $142 \pm 1$  and  $30 \pm 10$  mL  
5 gVS<sup>-1</sup> for the groups G1, G2 and G3, respectively. Concerning the replicates named  
6 30%-a, the reduction of the cumulated methane production is consistent with previous  
7 works. Indeed, Fernandez *et al.* (2008) observed a higher methane yield at 20% TS than  
8 at 30% TS for mesophilic conditions in batch tests. Likewise, Forster-Carneiro *et al.*  
9 (2008) reported a higher methane production in thermophilic batch tests at 20% TS  
10 compared to 25% TS, and even higher than at 30% TS.

11 At 30%-b and 35% TS, lower methane productions of about 37 and 24 mL gVS<sup>-1</sup>,  
12 respectively, were observed. The difference in the behavior between the four replicates  
13 at 30% TS suggested that 30% TS could be considered as a threshold concentration for  
14 an inhibitory effect in high solids anaerobic digestion. This threshold could correspond  
15 to an inhibition of anaerobic digestion at high solids content due to a higher  
16 accumulation of metabolic by-products, such as volatile fatty acids.

### 3.1.3. pH and VFA concentration

17 For a better understanding of the failure of dry anaerobic digestion at TS >30%, volatile  
18 fatty acids (VFA) and pH were analyzed at the end of the experiments (Figure 2c). For  
19 batch experiments with a good anaerobic digestion performance (between 10% TS and  
20 30%-a), there was no accumulation of VFA and the average pH was  $7.43 \pm 0.31$ .

21 When anaerobic digestion was inhibited (30%-b and 35% TS), high VFA  
22 concentrations were measured, with values ranging from 29 to 36 gVFA kgVS<sup>-1</sup>. Under  
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1 semi-dry anaerobic digestion conditions, Li *et al.* (2010) already observed an increase of  
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3 the VFA concentration with increasing VS content (for TS contents of 16.0%, 13.5%  
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5 and 11.0%, the maximum VFA value was 22.4 g L<sup>-1</sup>, 6.8 g L<sup>-1</sup> and 4.2 g L<sup>-1</sup>,  
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7  
8 respectively), and high concentrations of VFA inhibit the methane production (Ahring  
9  
10 and Westermann, 1988). Mainly, total VFAs affect more specifically methanogens  
11  
12 (Vedrenne, 2007). In addition, the average pH value was about 5.95 and 6.10 at 30%-b  
13  
14 and 35% TS, respectively. These low pH values associated with VFA concentrations of  
15  
16 29 and 36 g L<sup>-1</sup> might explain the inhibition of the anaerobic digestion process for these  
17  
18 two TS contents, since VFA accumulation contributed to the decrease of pH. Moreover,  
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20 at low pH, between 5 and 7, VFA are in their undissociated form, which is toxic for  
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22 microorganisms (Zehnder, 1978).  
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### 30 **3.2. Model results**

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32 For better understanding of the origin of the limitation of anaerobic degradation at high  
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34 solids contents, the Anaerobic Digestion Model No. 1 (ADM1) was used to describe  
35  
36 this experimental data. Figure 3 presents exemplarily simulation results for 10% TS and  
37  
38 the critical concentration of 30% TS. Model predictions closely followed the  
39  
40 experimental data of cumulative methane production for a TS concentration of 10%,  
41  
42 except for the first 50 days of the experiment. However, the model clearly over-  
43  
44 predicted experimental data obtained for a TS concentration of 30%, using the same set  
45  
46 of calibrated parameter values ( $k_T=0.5\text{ d}^{-1}$  and  $k_{\text{hyd},X_{\text{ch}}}=0.01\text{ d}^{-1}$ ). As the methane  
47  
48 production rate production rate decreased with increasing total solids content (see  
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50 section ‘3.1.1 Methane production rate’) and hydrolysis is the limiting step in methane  
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52 production (Vavilin *et al.*, 2004), it was hypothesized that the first-order hydrolysis rate  
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1 decreases with increasing TS concentration. Consequently, the first-order hydrolysis  
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3 rate for carbohydrates,  $k_{\text{hyd},X_{\text{ch}}}$ , was decreased step by step with increasing TS  
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5 concentration to fit simulation results, in particular the total cumulative methane  
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7 production, to experimental data. The first-order hydrolysis rate needed to be reduced to  
8  
9 0.0045  $\text{d}^{-1}$  in order to properly describe the final cumulative  $\text{CH}_4$  production for the two  
10  
11 experiments that were not inhibited by high VFA concentrations and low pH at 30% TS  
12  
13 (Figure 3.b, solid line). The experimental results of the two experiments, in which  
14  
15 methane production stalled due to very high VFA concentrations and reactor  
16  
17 acidification, could not be described by further decreasing the hydrolysis rate (see  
18  
19 section '3.2.1 Influence of the hydrolysis rate constant'). Thus, another mechanism must  
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21 have been responsible for the inhibition of methane production.  
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27 Due to the pasty texture of the substrate-biomass mixture, the initial value of the  
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29 liquid/gas transfer coefficient  $k_T$  was already reduced to 0.5  $\text{d}^{-1}$  for a TS concentration  
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31 of 10% to account for reduced mass transfer. As the viscosity of the substrate-biomass  
32  
33 mixture increased and the porosity decreased with increasing TS content, it was  
34  
35 hypothesized that the overall mass transfer decreases with increasing TS concentration.  
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37 The overall mass transfer coefficient ( $k_T$ ) was, therefore, decreased to match measured  
38  
39 and simulated total cumulative methane production for the two inhibited batch  
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41 experiments at 30% TS. Reducing the value of  $k_T$  from 0.5  $\text{d}^{-1}$  to 0.02  $\text{d}^{-1}$  resulted in an  
42  
43 adequate representation of the final cumulative methane production for the two  
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45 inhibited experiments (Figure 3.b, dashed line).  
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51 Although the calibration of  $k_T$  and  $k_{\text{hyd},X_{\text{ch}}}$  allowed for describing the final cumulative  
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53 methane production, the ADM1 did not describe very well the initial phase of  
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55 cumulative methane production. The ADM1 describes hydrolysis as a simple first-order  
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1 process. According to Vavilin *et al.* (2008), hydrolysis of certain complex substrates  
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3 can, however, be better represented by the Contois model, which considers that  
4  
5 hydrolysis kinetics depends on both substrate and biomass concentration (concentration  
6  
7 of the biomass that produces the enzymes for hydrolysis). The Contois model is a more  
8  
9 general model than the first-order hydrolysis kinetics and allows for better representing  
10  
11 the sigmoid-type curve of cumulative methane production (Vavilin *et al.*, 2008). Using  
12  
13 the Contois model, however, has no impact on the prediction of the final cumulative  
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15 methane production (data not shown). As the aim of the modeling study was to find an  
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17 explanation for the general trend of the experimental data (inhibition of anaerobic  
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19 digestion of cardboard at high total solids contents), the first-order hydrolysis model is  
20  
21 the simplest approximation of the hydrolysis step. In addition, the more complex  
22  
23 structure of the Contois model makes it more difficult to identify and distinguish  
24  
25 mechanisms influencing anaerobic digestion at high TS concentrations. Thus, first-order  
26  
27 hydrolysis kinetics was used for deeper analysis of mechanisms affecting anaerobic  
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29 digestion at high TS concentrations. Based on the simulation results presented above,  
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31 two different mechanisms were identified that have a considerable impact on anaerobic  
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33 digestion at high TS concentrations: 1) the rate of hydrolysis of particulate substrates  
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35 (here only carbohydrates), and 2) the transfer of produced gas from the liquid phase to  
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37 the gas phase. The influence of both phenomena is now presented in more detail.  
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### 50 *3.2.1 Influence of the hydrolysis rate constant*

51 For each TS concentration, the first-order hydrolysis rate constant for carbohydrates was  
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53 calibrated for batch experiments that were not inhibited by high VFA concentrations  
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55 and low pH to meet the total cumulative methane production at the end of each batch  
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1 experiment. Figure 4 shows the calibrated hydrolysis rate constants for each TS  
2  
3 concentration. The hydrolysis rate constants obtained for TS concentrations of 10% and  
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5 15% were similar. But the calibrated hydrolysis rate constants linearly decreased with  
6  
7 increasing TS concentration for  $15\% \leq \text{TS} \leq 30\%$ . This finding is in agreement with  
8  
9 results presented by Pommier *et al.* (2007), who observed a strong impact of the  
10  
11 moisture content on kinetic rates and maximum methane production in solid waste  
12  
13 anaerobic digestion. Extrapolation of the hydrolysis rate constant for a TS concentration  
14  
15 of 35% using the linear relationship shown in Figure 4 gave a value of  $0.00275 \text{ d}^{-1}$ .  
16  
17  
18 With this hydrolysis rate constant and a  $k_T$  value of  $0.5 \text{ d}^{-1}$  (as used for all non-inhibited  
19  
20 batch experiments), the ADM1 predicted a total cumulative methane production of 107  
21  
22  $\text{mL CH}_4 \text{ gVS}^{-1}$  and no VFA accumulation for  $\text{TS} = 35\%$ . The total cumulative methane  
23  
24 production predicted by the model was 4.5 times higher than that observed in the  
25  
26 experiment ( $23.5 \text{ mL CH}_4 \text{ gVS}^{-1}$ ). Thus, a further decrease in the hydrolysis rate  
27  
28 constant could not explain why methane production stalled at a total solids  
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30 concentration of 35%.

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33 In summary, a decreasing first-order hydrolysis rate constant with increasing TS content  
34  
35 could well describe experimental methane production at different TS in non-inhibited  
36  
37 batch experiments. But a decreasing hydrolysis rate constant could not explain  
38  
39 considerable VFA accumulation, reactor acidification, and limited methane production  
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41 observed in two batch experiments at 30% TS and all batch experiments at 35% TS.  
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### 52 3.2.2. Influence of the overall mass transfer coefficient

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54 The influence of the overall mass transfer coefficient  $k_T$  was evaluated by varying its  
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56 value between  $0.001 \text{ d}^{-1}$  and  $100 \text{ d}^{-1}$  for each studied TS concentration. The default  
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1 value (named  $k_{La}$  in the original ADM1) is  $200 \text{ d}^{-1}$  for modeling anaerobic digestion of  
2  
3 activated sludge (wet digestion) (Rosen and Jeppsson, 2006). The first-order hydrolysis  
4  
5 rate constant for carbohydrates was kept constant at  $0.01 \text{ d}^{-1}$  for all simulations to focus  
6  
7 on the effect of the overall mass transfer coefficient. Figure 5 shows the final  
8  
9 cumulative methane production obtained for different values of  $k_T$  and different TS  
10  
11 contents. A sudden and steep decrease in the cumulative methane production can be  
12  
13 observed for  $k_T$  values between  $0.04 \text{ d}^{-1}$  and  $0.18 \text{ d}^{-1}$  depending on the TS content.  
14  
15 Above a critical  $k_T$  value, methane production was not influenced by overall mass  
16  
17 transfer and the cumulative methane production remained almost constant  
18  
19 independently of the  $k_T$  value. In this case, no VFA accumulation was observed. Below  
20  
21 the critical  $k_T$  value, limited overall mass transfer clearly affected methane production  
22  
23 and resulted in considerably lower cumulative methane production. Methane production  
24  
25 was more sensitive to overall mass transfer limitations at high TS concentrations as the  
26  
27 methane production dropped already at higher  $k_T$  values (at  $0.18 \text{ d}^{-1}$  for 35% TS  
28  
29 compared to  $0.04 \text{ d}^{-1}$  for 10% TS). Low methane production always went along with  
30  
31 accumulation of VFA and reactor acidification (data not shown).  
32  
33 The different behaviors in the batch experiments at 30% TS content might be related to  
34  
35 slightly different actual total solids contents in the serum bottles as a result of minor  
36  
37 inaccuracies in the preparation of the bottles, including measurement errors. In  
38  
39 simulations, methane production at 30% TS content dropped from  $174.5 \text{ mL CH}_4 \text{ gVS}^{-1}$   
40  
41 to  $51.9 \text{ mL CH}_4 \text{ gVS}^{-1}$  when the mass transfer coefficient was reduced from  $0.16 \text{ d}^{-1}$  to  
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43  $0.15 \text{ d}^{-1}$ . At 35% TS, the methane production dropped already between  $k_T = 0.18 \text{ d}^{-1}$  and  
44  
45  $k_T = 0.17 \text{ d}^{-1}$  from  $174.5 \text{ mL CH}_4 \text{ gVS}^{-1}$  to  $32.8 \text{ mL CH}_4 \text{ gVS}^{-1}$ . In addition, the lines for  
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47 TS 30% and 35% in Figure 5 are very close to each other. Thus, a small increase in the  
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1 actual TS concentration at a desired TS content of 30% can have already a significant  
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3 impact on the cumulative methane production.  
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6 According to model results, overall mass transfer limitation resulted in an accumulation  
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8 of inorganic carbon (CO<sub>2</sub>), dissolved methane, and dissolved hydrogen, and can have an  
9  
10 inhibitory effect on methanogenesis. Indeed, the CO<sub>2</sub> produced during methanogenesis  
11  
12 remains trapped in the matrix since the release of dissolved compounds is low in the  
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14 digestate: therefore, the local CO<sub>2</sub> concentration can increase, which may cause a local  
15  
16 acidification, and hence inhibition of methanogenesis from the beginning of the  
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18 experiment implicating VFA accumulation. The accumulation of dissolved hydrogen  
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20 can also cause inhibition of valerate, butyrate, and propionate degradation and generate  
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22 an accumulation of these VFA. “Obligated Hydrogen-Producing Acetogens” bacteria  
23  
24 convert VFA and high molecular weight long-chain fatty acids to acetate, carbon  
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26 dioxide and hydrogen. These  $\beta$ -oxidation reactions are thermodynamically not possible  
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28 under standard conditions ( $\Delta G^\circ > 0$ ). In fact,  $\beta$ -oxidation occurs only at very low  
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30 hydrogen partial pressure. Thus, produced hydrogen needs to be locally consumed (e.g.,  
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32 syntrophic association between hydrogen producing species and methanogenic species)  
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34 (Ahring, 2003), or to be removed from the liquid phase through liquid/gas mass  
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36 transfer.  
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45 An effect of a limitation of the overall mass transfer on hydrolysis cannot be reported as  
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47 the ADM1 does not consider inhibition of hydrolysis. Overall, the simulation results  
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49 clearly show that overall mass transfer limitation can explain limited methane  
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51 production at 30% and 35% TS. But the decreasing cumulative methane production for  
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53 TS concentrations between 15% and 30% cannot be attributed to mass transfer  
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55 limitation.  
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#### 4. Conclusion

Based on experimental observations and ADM1 simulations, it was shown that the TS content affects the global anaerobic digestion performance of solids through (i) reduction of the microbial hydrolysis rate for  $10\% \leq \text{TS} \leq 25\%$  and (ii) physical limitation related to liquid/gas mass transfer for  $\text{TS} \geq 30\%$ . Further investigations need to be done to understand the role of the TS content on the behavior of the microbial community structure involved in the anaerobic digestion degradation of solids from wet to dry technologies.

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1           **References**  
2  
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- 4           1. ADEME, 2009. <http://www2.ademe.fr>  
5  
6           2. Ahring, B.K., 2003. Biomethanisation, Springer-Verlag, Berlin Heidelberg New  
7           York.  
8  
9  
10          3. Ahring, B.K., Westermann, P., 1988. Product Inhibition of butyrate metabolism by  
11           acetate and hydrogen in a thermophilic coculture. *Appl. Environ. Microbiol.* 54  
12           (10), 2393-2397.  
13  
14  
15  
16  
17  
18          4. Angelidaki, I., Sanders, W., 2004. Assessment of the anaerobic biodegradability of  
19           macropollutants. *Rev. Environ. Sci. Biotechnol.* 3 (2), 117-129.  
20  
21  
22  
23          5. Batstone, D.J., Keller, J., 2003. Industrial applications of the IWA anaerobic  
24           digestion model No. 1 (ADM1). *Water Sci. Technol.* 47 (12), 199-206.  
25  
26  
27  
28          6. Batstone, D.J., Keller, J., Steyer, J.P., 2006. A review of ADM1 extensions,  
29           applications, and analysis: 2002-2005. *Water Sci. Technol.* 54 (4), 1-10.  
30  
31  
32  
33          7. Batstone, D.J., Keller, J., Angelidaki, I., Kalyuzhnyi, S., Pavlostathis, S.G., Rozzi, A.,  
34           Sanders, W.T.M., Siegrist, H., Vavilin, V.A., 2002. *Anaerobic Digestion Model*  
35           No.1 (ADM1). IWA Publishing, London, UK.  
36  
37  
38  
39  
40          8. Battistoni, P., 1997. Pre-treatment, measurement execution procedure and waste  
41           characteristics in the rheology of sewage sludges and the digested organic fraction  
42           of municipal solid wastes. *Water Sci. Technol.* 36 (11), 33-41.  
43  
44  
45  
46  
47          9. Battistoni, P., Fava, G., Stanzini, C., Cecchi, F., Bassetti, A., 1993. Feed  
48           Characteristics and Digester Operative Conditions as Parameters Affecting the  
49           Rheology of Digested Municipal Solid-Wastes. *Water Sci. Technol.* 27 (2), 37-45.  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
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65

- 1 10. Buffiere, P., Frederic, S., Marty, B., Delgenes, J.P., 2008. A comprehensive method  
2  
3 for organic matter characterization in solid wastes in view of assessing their  
4  
5 anaerobic biodegradability. *Water Sci. Technol.* 58 (9), 1783-1788.  
6  
7
- 8 11. Fernández, J., Pérez, M., Romero, L.I., 2008. Effect of substrate concentration on  
9  
10 dry mesophilic anaerobic digestion of organic fraction of municipal solid waste  
11  
12 (OFMSW). *Bioresour. Technol.* 99 (14), 6075-6080.  
13  
14
- 15 12. Fernández, J., Pérez, M., Romero, L.I., 2010. Kinetics of mesophilic anaerobic  
16  
17 digestion of the organic fraction of municipal solid waste: influence of initial total  
18  
19 solid concentration. *Bioresour. Technol.* 101 (16), 6322-6328.  
20  
21
- 22 13. Forester, J., Pérez, M., Romero, L.I., 2008. Influence of total solid and inoculum  
23  
24 contents on performance of anaerobic reactors treating food waste. *Bioresour.*  
25  
26 *Technol.* 99 (15), 6974-7002.  
27  
28
- 29 14. Ganesh, R., Rajinikanth, R., Thanikal, J.V., Ramanujam, R.A., Torrijos, M., 2010.  
30  
31 Anaerobic treatment of winery wastewater in fixed bed reactors. *Bioprocess*  
32  
33 *Biosyst. Eng.* 33 (5), 619-628.  
34  
35
- 36 15. Garcia-Bernet, D., Buffière, P., Latrille, E., Steyer, J.P., Escudíé, R., 2011a. Water  
37  
38 distribution in biowastes and digestates of dry anaerobic digestion technology.  
39  
40 *Chem. Eng. J.* 172 (2-3), 924-928.  
41  
42
- 43 16. Garcia-Bernet, D., Loisel, D., Guizard, G., Buffière, P., Steyer, J.P., Escudíé, R.,  
44  
45 2011b. Rapid measurement of the rheological properties of anaerobically digested  
46  
47 solid waste using slump test. *Waste Manag.* 31 (4), 631-635.  
48  
49
- 50 17. Karim, K., Hoffmann, R., Klasson, K.T., Al-Dahhan, M.H., 2005. Anaerobic  
51  
52 digestion of animal waste: effect of mode of mixing. *Water Res.* 9 (15), 3597-  
53  
54 3606.  
55  
56  
57  
58  
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55  
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58  
59  
60  
61  
62  
63  
64  
65
18. Koch., K., Lübken., M., Gehring. T., Wichern., M., Horn. H., 2010. Biogas from grass silage - Measurements and modeling with ADM1. *Bioresour. Technol.* 101 (21), 8158-8165.
  19. Le Hyaric, R., Chardin, C., Benbelkacem, H., Bollon, J., Bayard, R., Escudié, R., Buffière, P., 2011. Influence of substrate concentration and moisture content on the specific methanogenic activity of dry mesophilic municipal solid waste digestate spiked with propionate. *Bioresour. Technol.* 102 (2), 822-827.
  20. Li, D., Yuan, Zh.H., Sun, Y.M., 2010. Semi-dry mesophilic anaerobic digestion of water sorted organic fraction of municipal solid waste (WS-OFMSW). *Bioresour. Technol.* 101 (8), 2722-2728.
  21. Liebetrau, J., Kraft, E., Bidlingmaier, W., 2004. The influence of the hydrolysis rate of co-substrates on process behaviour. In: Guiot, S.G. (Ed.), *Proceedings of the Tenth World Congress on Anaerobic. Canadian Association on Water Quality, Montreal*, pp. 1296-1300.
  22. Pommier, S., Chenu, D., Quintard, M., Lefebvre, X., 2007. A logistic model for the prediction of the influence of water on the solid waste methanization in landfills. *Biotechnol. Bioeng.* 97 (3), 473-482.
  23. Qu, X., Vavilin, V.A., Mazéas, L., Lemunier, M., Duquennoi, C., He, P.-J., Bouchez, T., 2009. Anaerobic biodegradation of cellulosic material: Batch experiments and modelling based on isotopic data and focusing on aceticlastic and non-aceticlastic methanogenesis. *Waste Manag.* 29 (6), 1828-1837.
  24. Rosen, C., Jeppsson, U., 2006. Aspects on ADM1 Implementation within the BSM2 Framework. Technical report. Dept. of Industrial Electrical Engineering and Automation, Lund University, Lund, Sweden.

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54  
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60  
61  
62  
63  
64  
65
25. Van Soest, P.J., 1963. Use of detergents in the analysis of fibrous feeds. II. A rapid method for the determination of fiber and lignin. *J. Assoc. Off. Agric.Chem.* 46, 825-835.
  26. Vavilin, V.A., Fernandez, B., Palatsi, J., Flotats., X., 2008. Hydrolysis kinetics in anaerobic degradation of particulate organic material: an overview. *Waste Manag.* 28 (6), 941-953.
  27. Vavilin, V.A., Lokshina, L.Y., Jokela, J., Rintala, J., 2004. Modeling solid waste decomposition. *Bioresource Technol.* 94 (1), 69-81.
  28. Vedrenne, F., Beline, F., Dabert, P., Bernet, N., 2008. The effect of incubation conditions on the laboratory measurement of the methane producing capacity of livestock measurement wastes. *Bioresour. Technol.* 99 (1), 146-155.
  29. Zehnder, A.J.B., 1978. Ecology of methane formation. *Water pollution microbiology*, Vol. 2, Edited by Ralph Mitchell. John Wiley and Sons, New York, NY, pp. 349-376



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2  
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## Figure captions

**Figure 1:** Cumulated methane production from cardboard according to the TS content.

The insert corresponds to the first 40 days of the experiment.

**Figure 2:** Influence of TS content on anaerobic digestion performance.

a) Maximum methane production rate.

b) Final methane yield (at the end of the experiment).

c) pH and VFA concentration at the end of the experiment.

**Figure 3:** Experimental and simulated cumulative CH<sub>4</sub> production.

a) At 10% TS, with calibrated values for  $k_T$  and  $k_{hyd,Xch}$ .

b) At 30% TS, with calibrated values for  $k_T$  and  $k_{hyd,Xch}$ .

**Figure 4:** Influence of the first-order hydrolysis rate constant for carbohydrates on anaerobic digestion of cardboard for unlimited mass transfer as predicted by the ADM1.

**Figure 5:** Influence of the overall mass transfer coefficient  $k_T$  on anaerobic digestion of cardboard as predicted by the ADM1.

Figure1

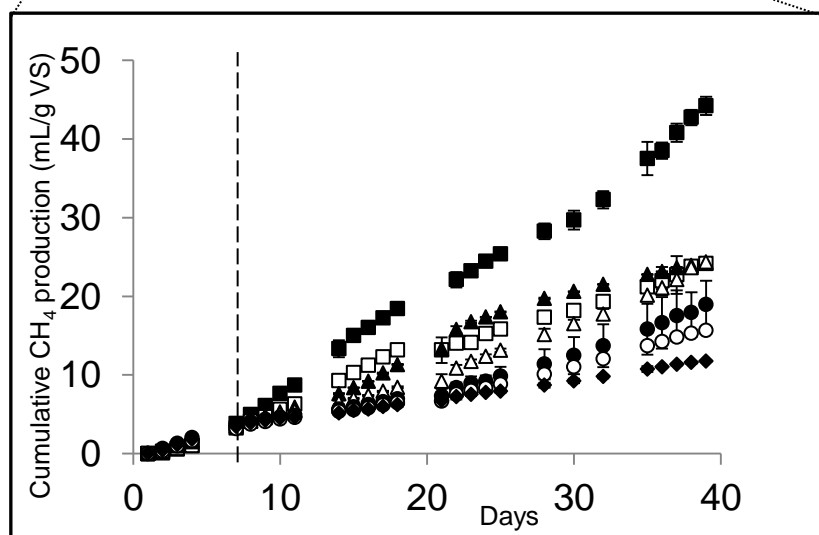
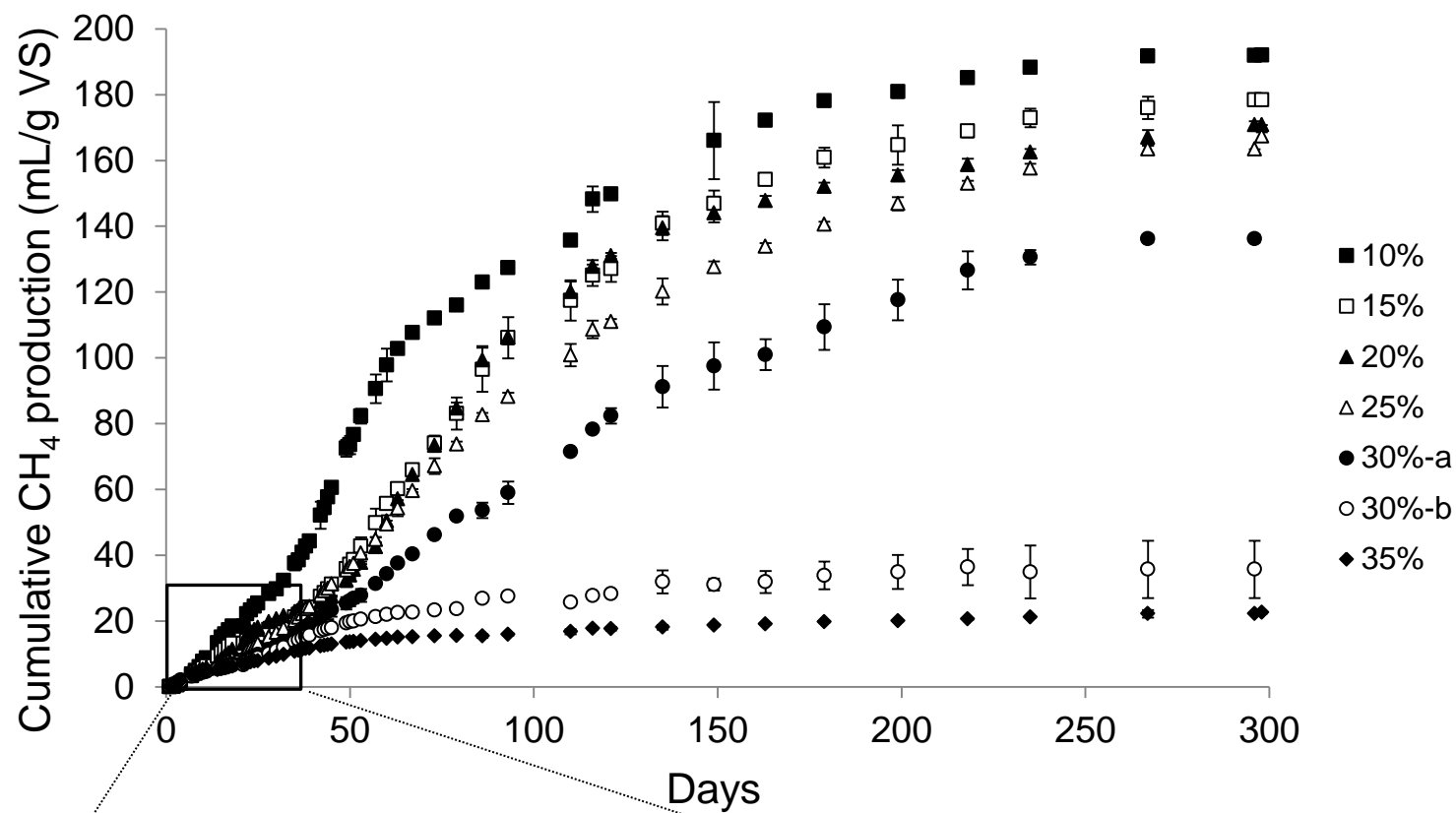


Figure2a

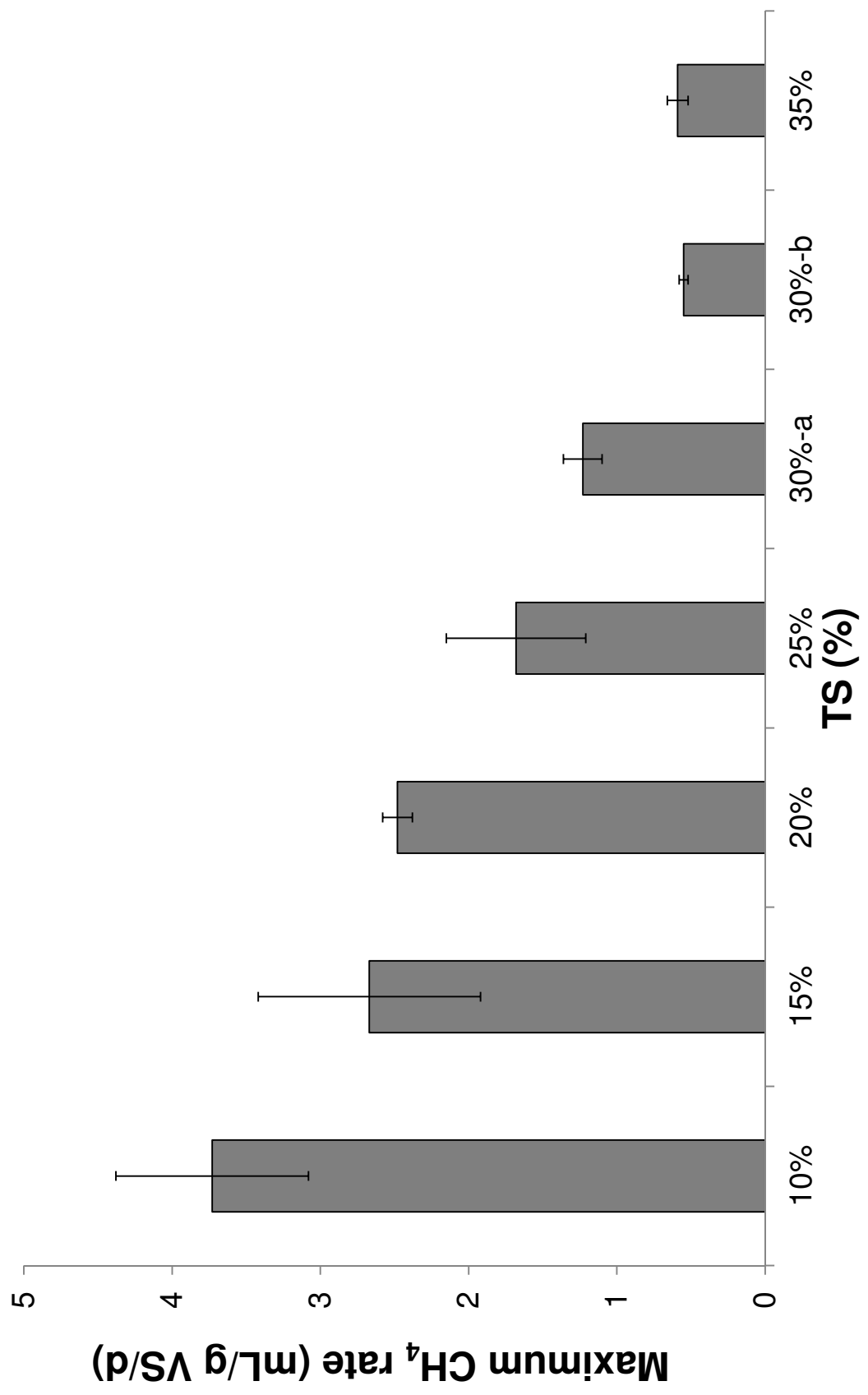


Figure2b

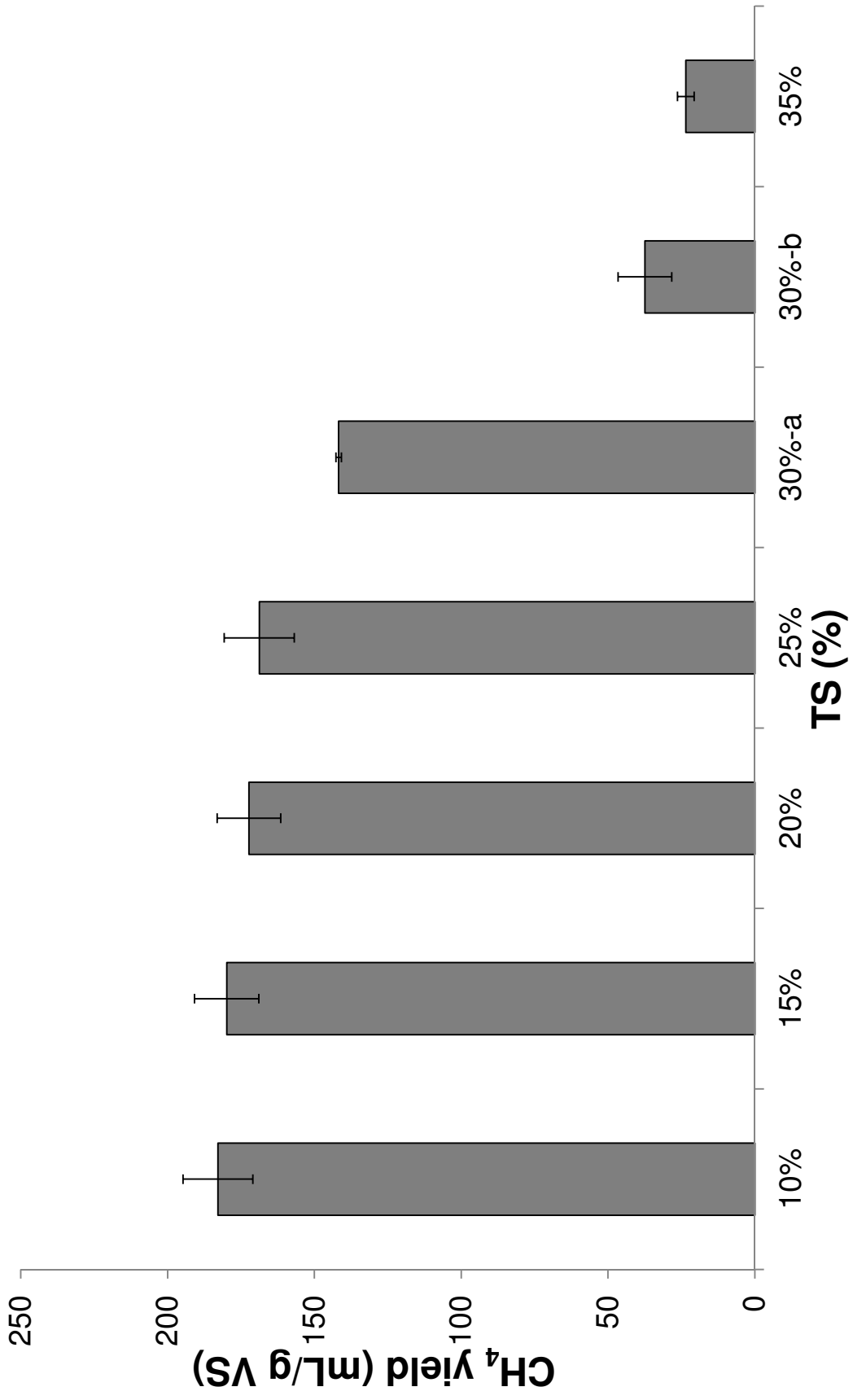


Figure 2c

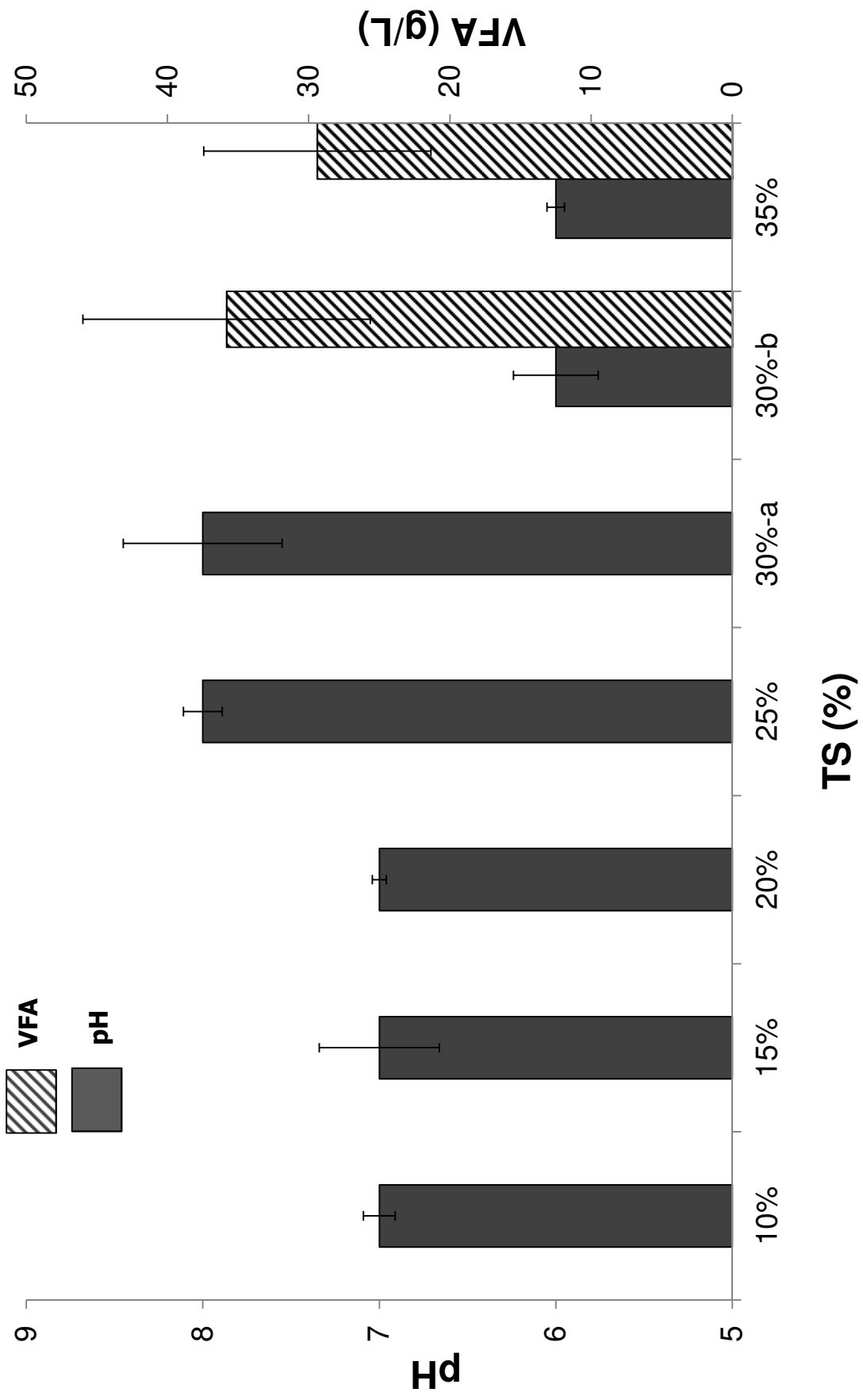


Figure3a

### TS 10%

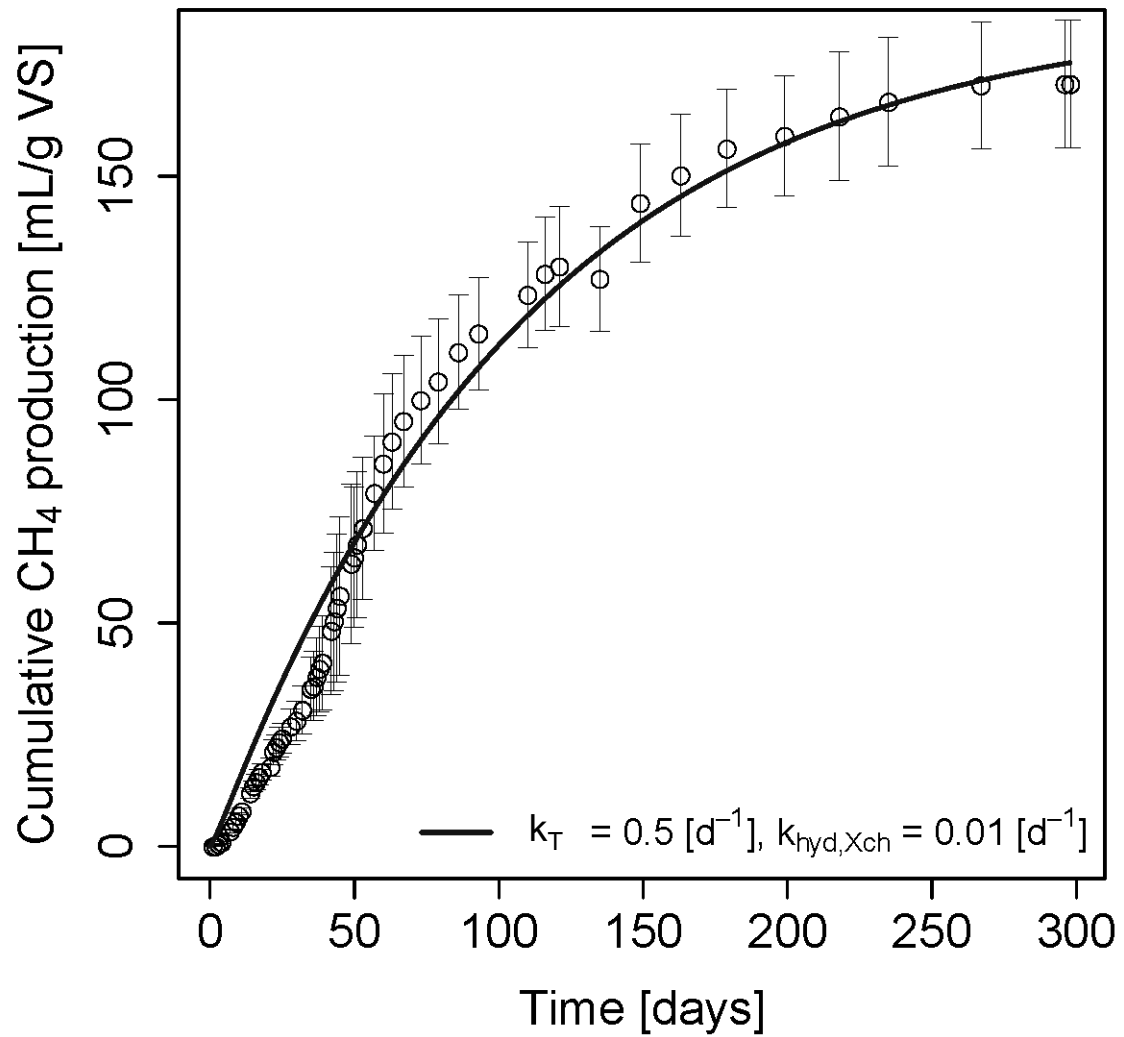


Figure 3b

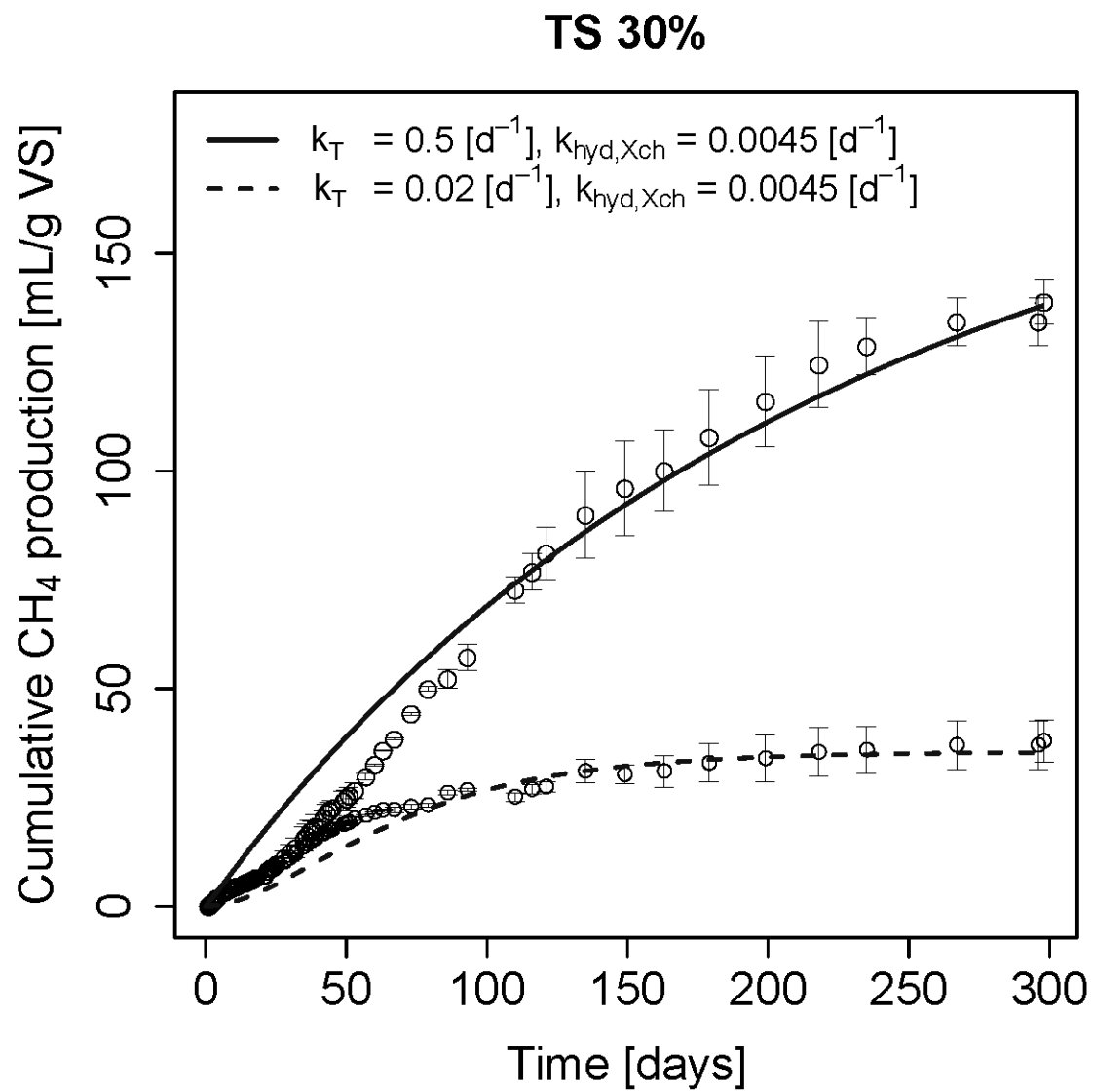


Figure4

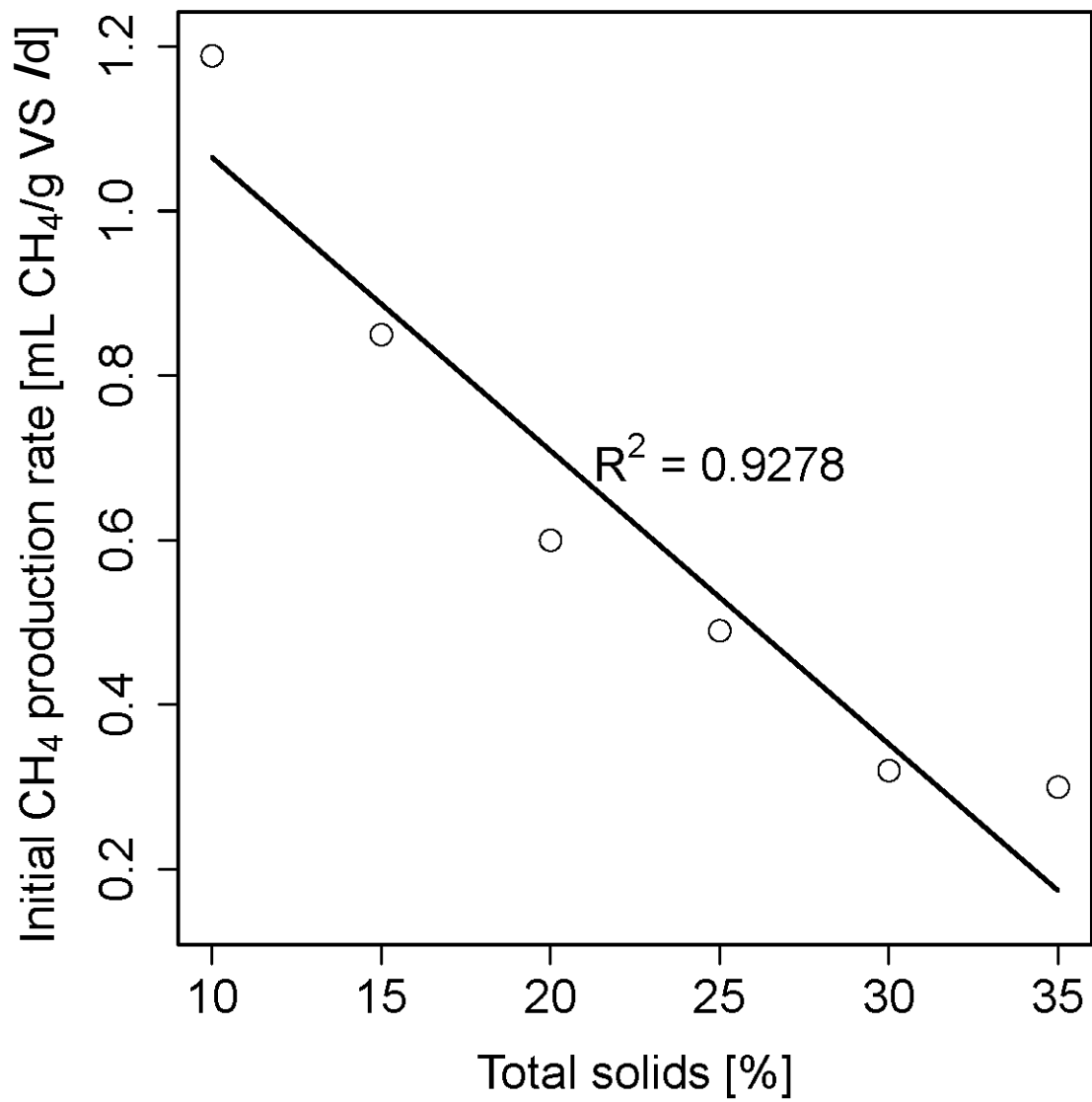




Figure5

