



HAL
open science

Characterization of *Vitis vinifera* NPR1 homologs involved in the regulation of Pathogenesis-Related gene expression

Gaëlle Le Henanff, Thierry Heitz, Pere Mestre, Jérôme Mutterer, Bernard Walter, Julie Chong

► To cite this version:

Gaëlle Le Henanff, Thierry Heitz, Pere Mestre, Jérôme Mutterer, Bernard Walter, et al.. Characterization of *Vitis vinifera* NPR1 homologs involved in the regulation of Pathogenesis-Related gene expression. *BMC Plant Biology*, 2009, 9 (54), 14 p. 10.1186/1471-2229-9-54 . hal-02660254

HAL Id: hal-02660254

<https://hal.inrae.fr/hal-02660254>

Submitted on 30 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Research article

Open Access

Characterization of *Vitis vinifera* NPR1 homologs involved in the regulation of Pathogenesis-Related gene expression

Gaëlle Le Henanff¹, Thierry Heitz², Pere Mestre³, Jérôme Mutterer², Bernard Walter¹ and Julie Chong*¹

Address: ¹Laboratoire Vigne, Biotechnologies et Environnement (LVBE, EA3991), Université de Haute Alsace, 33 rue de Herrlisheim, 68000 Colmar, France, ²Département Réseaux Métaboliques chez les Végétaux, IBMP du CNRS (UPR2357), 12 rue du général Zimmer, 67000 Strasbourg, France and ³Laboratoire de Génétique et Amélioration de la Vigne, INRA et Université de Strasbourg (UMR1131), 28 rue de Herrlisheim, 68000 Colmar, France

Email: Gaëlle Le Henanff - gaelle.le-henanff@uha.fr; Thierry Heitz - thierry.heitz@ibmp-ulp.u-strasbg.fr; Pere Mestre - mestre@colmar.inra.fr; Jérôme Mutterer - jerome.mutterer@ibmp-ulp.u-strasbg.fr; Bernard Walter - bernard.walter@uha.fr; Julie Chong* - julie.chong@uha.fr

* Corresponding author

Published: 11 May 2009

Received: 10 February 2009

BMC Plant Biology 2009, 9:54 doi:10.1186/1471-2229-9-54

Accepted: 11 May 2009

This article is available from: <http://www.biomedcentral.com/1471-2229/9/54>

© 2009 Le Henanff et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Grapevine protection against diseases needs alternative strategies to the use of phytochemicals, implying a thorough knowledge of innate defense mechanisms. However, signalling pathways and regulatory elements leading to induction of defense responses have yet to be characterized in this species. In order to study defense response signalling to pathogens in *Vitis vinifera*, we took advantage of its recently completed genome sequence to characterize two putative orthologs of NPR1, a key player in salicylic acid (SA)-mediated resistance to biotrophic pathogens in *Arabidopsis thaliana*.

Results: Two cDNAs named VvNPR1.1 and VvNPR1.2 were isolated from *Vitis vinifera* cv Chardonnay, encoding proteins showing 55% and 40% identity to Arabidopsis NPR1 respectively. Constitutive expression of VvNPR1.1 and VvNPR1.2 monitored in leaves of *V. vinifera* cv Chardonnay was found to be enhanced by treatment with benzothiadiazole, a SA analog. In contrast, VvNPR1.1 and VvNPR1.2 transcript levels were not affected during infection of resistant *Vitis riparia* or susceptible *V. vinifera* with *Plasmopara viticola*, the causal agent of downy mildew, suggesting regulation of VvNPR1 activity at the protein level. VvNPR1.1-GFP and VvNPR1.2-GFP fusion proteins were transiently expressed by agroinfiltration in *Nicotiana benthamiana* leaves, where they localized predominantly to the nucleus. In this system, VvNPR1.1 and VvNPR1.2 expression was sufficient to trigger the accumulation of acidic SA-dependent Pathogenesis-Related proteins PR1 and PR2, but not of basic chitinases (PR3) in the absence of pathogen infection. Interestingly, when VvNPR1.1 or AtNPR1 were transiently overexpressed in *Vitis vinifera* leaves, the induction of grapevine PR1 was significantly enhanced in response to *P. viticola*.

Conclusion: In conclusion, our data identified grapevine homologs of NPR1, and their functional analysis showed that VvNPR1.1 and VvNPR1.2 likely control the expression of SA-dependent defense genes. Overexpression of VvNPR1 has thus the potential to enhance grapevine defensive capabilities upon fungal infection. As a consequence, manipulating VvNPR1 and other signalling elements could open ways to strengthen disease resistance mechanisms in this crop species.

Background

Grapevine (*Vitis vinifera*) is a major fruit crop worldwide that is susceptible to many microbial infections, especially by fungi, thus requiring an intensive use of phytochemicals. The economic costs and negative environmental impact associated with these applications led to search for alternative strategies involving activation of the plant's innate defense system. In order to efficiently limit the losses due to diseases, it is therefore necessary to have a thorough knowledge of grapevine disease resistance mechanisms.

Plants have developed a two-layered innate immune system for defense against pathogens. Primary innate immunity, the first line of defense of plants, is achieved through a set of defined receptors, that recognize conserved microbe-associated molecular patterns [1]. In order to defend themselves against pathogens that can suppress primary defense mechanisms, plants have developed a secondary defense response that is triggered upon recognition of race-specific effectors. Resistance proteins monitor these effectors and subsequently trigger secondary defense responses that often culminate in localized cell death response associated with additional locally induced defense responses, that block further growth of the pathogen [1]. After recognition of the invading microorganism, induced resistance to different types of pathogens is achieved through a network of signal transduction pathways in which the small molecules salicylic acid (SA), jasmonic acid (JA) and ethylene (ET) act as secondary messengers [2]. These regulators then orchestrate the expression of sets of downstream defense genes encoding antimicrobial proteins or enzymes catalyzing the production of defense metabolites. Plant resistance to biotrophic pathogens is classically believed to be mediated through SA signalling [3]. SA accumulation as well as the coordinated expression of *Pathogenesis Related (PR)* genes encoding small proteins with antimicrobial activity are also necessary to the onset of Systemic Acquired Resistance (SAR) in plants. SAR is a plant immune response that establishes a broad spectrum resistance in tissues distant from the site of primary infection [4].

In the past years, considerable progress has been made in the model plant *Arabidopsis thaliana* in identifying genes that affect regulation of defense gene expression. Several key plant defense regulators especially involved in the SA signalling pathway have been cloned and characterized [4]. The *npr1* mutant was isolated in a genetic screen for plants that failed to express *PR2* gene after SAR induction [5]. NPR1 (Nonexpressor of *PR* genes 1) has been identified as a key positive regulator of the SA-dependent signalling pathway and is required for the transduction of the SA signal to activate *PR* gene expression and Systemic Acquired Resistance [5]. The *NPR1* gene was cloned in 1997 and shown as encoding a novel protein containing ankyrin repeats involved in

protein-protein interactions [6]. *NPR1* is constitutively expressed and levels of its transcripts increased only two-fold following SA treatment, suggesting that it is regulated at the protein level [7]. Indeed, *NPR1* activity is regulated by redox systems which have been recently identified [8]. Inactive *NPR1* is present as cytosolic disulfide-bound oligomers in the absence of pathogen attack. Following SA induction, oligomeric *NPR1* is reduced to active monomers [9]. *NPR1* monomers are translocated to the nucleus where they interact with the TGA class of basic leucine zipper transcription factors, leading to the expression of SA-dependent genes [3,9]. Recent studies have also involved WRKY transcription factors in SA defense responses downstream or in parallel with *NPR1* [10].

In Arabidopsis, the *NPR1*-dependent SA pathway controls the expression of *PR1*, *PR2* (β -1.3-glucanase) and *PR5* (thaumatin-like) genes. In contrast, induction of distinct defense genes encoding the defensin PDF1.2 and the *PR3* (basic chitinase) proteins is controlled by JA/ET dependent pathways [2].

Originally, the *npr1* mutant was thought to be only deficient in SA-mediated defense. However, it was shown that *NPR1* plays a role in other defense signalling pathways. In *npr1*, the establishment of Induced Systemic Resistance (ISR) in leaves by non-pathogenic root rhizobacteria is blocked. Interestingly, this resistance response is independent of SA but requires ET and JA signalling [11]. Apart from *NPR1*, Arabidopsis genome contains five *NPR1*-related genes called *AtNPR2* to *AtNPR6* [12]. Members of the *AtNPR* family encode proteins sharing two domains involved in mediating protein-protein interactions: the Broad Complex, Tramtrack and Bric a brac/Pox virus and Zinc finger (BTB/POZ) domain in the N-terminal and the Ankyrin Repeat Domain (ARD) in the middle of the protein. Whereas *AtNPR1* to *AtNPR4* have been implicated in signalling of defense responses, *AtNPR5* and *AtNPR6* (called *AtBOP1* and *AtBOP2*) form a distinct group involved in the regulation of developmental patterning of leaves and flowers [13].

AtNPR1 has been over-expressed in Arabidopsis, rice, tomato and wheat, resulting in enhanced bacterial and fungal resistance [7,14-16]. Moreover, homologs of *AtNPR1* have been cloned and characterized in several crop plants including rice [17], apple [18], banana [19] and cotton [20]. In rice, over-expression of *OsNPR1* conferred disease resistance to bacterial blight, but also enhanced herbivore susceptibility in transgenic plants [17]. Similarly, over-expression of the *Malus NPR1* in two apple cultivars resulted in activation of *PR* genes and enhanced resistance to *Erwinia amylovora* and to two important fungal pathogens of apple [18].

In grapevine, many studies described the induction of *PR* proteins and the production of stilbenes after infection

[21,22]. However, signalling pathways and regulatory elements leading to the induction of these responses remain to be characterized in this species. Recently, two genes encoding transcription factors of the WRKY family and potentially involved in grapevine resistance to pathogens have been characterized. Overexpression of *VvWRKY1* and *VvWRKY2* in tobacco conferred reduced susceptibility to different types of fungi [23,24].

Recent completion of *Vitis vinifera* genome sequencing in a highly homozygous genotype and in a heterozygous grapevine variety has led to the identification of putative resistance genes and defense signalling elements [25,26]. Based on conserved domain analyses, the grape genome was found to contain a number of genes showing a nucleotide binding site (NBS) and a carboxy-terminal leucine-rich repeat (LRR) typical of resistance (R) genes [26]. Besides putative R genes, the grape genome contains several candidate genes encoding putative signalling components for disease response, with similarity to Arabidopsis EDS1, PAD4, NDR1 and NPR1 [26]. A possible role of the two grapevine regulatory elements sharing sequence similarity to the Arabidopsis SA signalling components NDR1 and EDS1 was recently described by our group [21].

Given the pivotal role of AtNPR1 in plant defense, we decided to take advantage of data from grapevine EST databases and genome sequencing to identify two genes encoding proteins with similarity to AtNPR1, that we called *VvNPR1.1* and *VvNPR1.2*. Expression of these genes was studied after treatment with benzothiadiazole (BTH, a SA analog) and after inoculation of two resistant or susceptible *Vitis* species with *Plasmopara viticola*, the causal agent of downy mildew. Nuclear localization of *VvNPR1.1* and *VvNPR1.2* was demonstrated by expressing GFP fusions. To get further insight into *VvNPR1* function, the two genes were transiently overexpressed in both *N. benthamiana* and *Vitis vinifera* leaves and consequences on *PR* gene induction were studied.

Results

Identification and sequence analysis of two NPR1-like genes in *Vitis vinifera*

At the beginning of this study, the grapevine genome was not entirely sequenced. The nucleic acid sequence of *AtNPR1* (At1g64280) was used to search an EST database of abiotically stressed *Vitis vinifera* cv Chardonnay leaves (EST Analysis Pipeline, ESTAP, [27]). Two ESTs with significant similarity to *AtNPR1* were identified. Sequence comparison of these two EST with data from grapevine genome sequencing project [28] enabled us to obtain the two full-length cDNAs, named *VvNPR1.1* (GSVIVT00016536001) and *VvNPR1.2* (GSVIVT00031933001). Amino acid sequence comparison of *VvNPR1.1* and *VvNPR1.2* showed that the two pro-

teins display 47% identity and 66% similarity. Completion of *V. vinifera* genome sequencing has revealed only two genes related to "defense" *AtNPRs* (K. Bergeault, unpublished results).

Amino acid sequence comparisons showed that *VvNPR1.1* has a higher identity with *AtNPR1* (55% identity and 75% similarity) than *VvNPR1.2* (40% identity and 61% similarity with *AtNPR1*). *VvNPR1.1* and *VvNPR1.2* were also compared to *NPR1* homologs in different plant species. Phylogenetic analysis (Figure 1A) reveals that *VvNPR1.1* groups closely with tobacco and tomato *NPR1* proteins (86% and 85% similarity respectively), with *NPR1* from monocots and with *AtNPR1* and *AtNPR2*. *VvNPR1.2* forms a discrete group with *NPR1* from apple (87% similarity), *AtNPR3* and *AtNPR4*.

VvNPR1.1 and *VvNPR1.2* encode putative proteins of 584 and 587 amino acids respectively (Figure 1B). According to PROSITE tool [29], *VvNPR1.1* and *VvNPR1.2* are predicted to have the same overall organization as members of the *AtNPR* family, with an amino terminal BTB/POZ domain and a central ankyrin repeat domain (Figure 1B). In addition, the carboxy terminal domains of *VvNPR1.1* and *VvNPR1.2* are rich in basic amino acids typical of nuclear localization signals (NLS, Figure 1C). Kinkema *et al.* [30] showed that five residues in the C-terminus of *AtNPR1* are essential for its nuclear translocation and constitute the NLS1. Four of these five amino acids are conserved in *VvNPR1.1* (Figure 1C), whereas some lysine residues have turned into arginine in *VvNPR1.2*. Basic amino acids of the second NLS in *AtNPR1* have been shown to be not necessary for nuclear targeting [30] and are less conserved among the different homologs even in the two grapevine proteins (Figure 1C).

VvNPR1.1 and *VvNPR1.2* expression following BTH treatment in grapevine leaves

In Arabidopsis, *AtNPR1* is constitutively expressed and can be further stimulated by SA or 2,6-dichloroisonicotinic acid (INA) treatment and by infection with *Hyaloperonospora parasitica* [31]. In order to study the expression profile of the two grapevine *NPR1* genes, detached leaves of *Vitis vinifera* cv Chardonnay were treated with a solution of BTH (a SA analog). We also monitored the expression of a grapevine *PR1* gene, a SAR marker, whose sequence is the most closely related to Arabidopsis SA-dependent *PR1* (GSVIVT_00038575001, [28]). As shown in Figure 2, *VvPR1* expression was strongly stimulated by BTH as soon as 12 h posttreatment compared to water-treated leaves where *VvPR1* expression was almost undetectable. *VvNPR1.1* was constitutively expressed in water-treated leaves, but expression was only slightly upregulated by BTH treatment (Figure 2). Interestingly, *VvNPR1.2*, whose expression was also detectable in con-

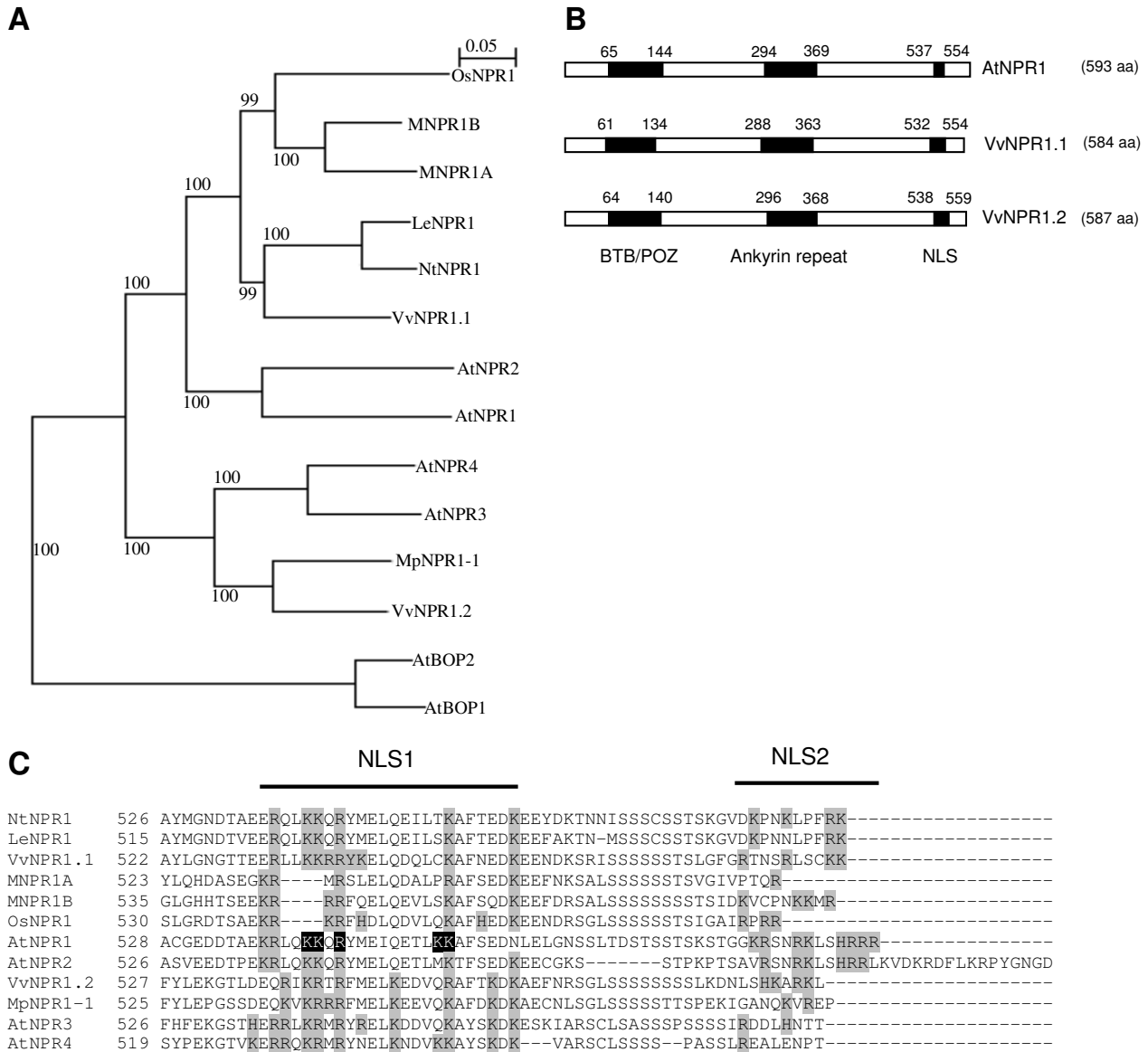


Figure 1
Comparison of VvNPR1.1 and VvNPR1.2 with other NPR1 homologs and members of Arabidopsis thaliana NPR family. (A) Phylogenetic tree generated with the Phylo_win program using the neighbour-joining method [44]. Sequence alignment was previously realized using the ClustalW tool. Accession numbers are: AtNPR1 (Atlg64280), AtNPR2 (At4g26120), AtNPR3 (At5g45110), AtNPR4 (At4g19660), AtBOPI (At3g57130), AtBOP2 (At2g41370), Nicotiana tabacum (NtNPR1, AAM62410.1), Oryza sativa cv. japonica (OsNPR1, AAX18700.1), Lycopersicon esculentum (LeNPR1, AAT57637.1), Musa acuminata (MNPR1A, ABI93182.1; MNPR1B, ABL63913.1), Malus × domestica (MpNPR1-1, ACC77697.1) and Vitis vinifera (Genoscope accession numbers: VvNPR1.1, GSVIVP00016536001; VvNPR1.2, GSVIVP00031933001). Bootstrap values based on 500 replicates are indicated beside the branches. (B) Schematic representation comparing the structure of AtNPR1, VvNPR1.1 and VvNPR1.2, including the positions of the BTB/POZ domain, the ankyrin repeat domain (ARD) and the nuclear localization signals (NLS). (C) Multiple alignment of putative nuclear localization signals (NLS) at C-terminus of NPRs from different plant species. Basic amino acids are highlighted in grey and residues essential for AtNPR1 nuclear localization [30] are highlighted in black.

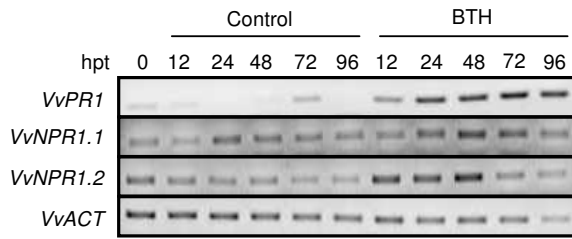


Figure 2
Expression patterns of VvNPR1.1 and VvNPR1.2 upon BTH treatment. Detached leaves of *Vitis vinifera* cv Chardonnay were sprayed with a solution of BTH (80 mg.L⁻¹) or water as control. Samples were collected at different time points. Hpt: hours post treatment; 0: untreated leaves at the beginning of the experiment. Actin (*VvACT*) was used as an internal control. Primer sequences are listed in table 1.

rol leaves, was further induced by BTH and peaked between 12 to 48 h after treatment (Figure 2). These results show that, as observed in *Arabidopsis*, *VvNPR1.1* and *VvNPR1.2* are constitutively expressed in grapevine and that *VvNPR1.2* expression can be further enhanced by a SAR inducer.

Expression patterns of VvNPR1.1 and VvNPR1.2 during compatible and incompatible interactions with *Plasmopara viticola*

We have next investigated whether the expression of *VvNPR1.1* and *VvNPR1.2* could be modulated after pathogen infection and whether their expression was differentially affected during compatible or incompatible interactions. Grapevine and related species exhibit a wide spectrum of resistance to the biotrophic pathogen *Plasmopara viticola*, the downy mildew agent. Two different *Vitis* species, the resistant *Vitis riparia* cv Gloire de Montpellier and the susceptible *Vitis vinifera* cv Chardonnay, were challenged with *Plasmopara viticola* or water as control. The expression patterns of *VvNPR1.1* and *VvNPR1.2* were determined after inoculation using real-time quantitative PCR. The expression of each gene after inoculation was calculated as fold induction compared to H₂O-inoculated leaves at the same time point as described by Pfaffl *et al* [32].

Five days after inoculation with *P. viticola*, a number of necrotic spots were observed on leaves of the resistant species *V. riparia*, whereas sporangia covered almost the entire leaf surface of the susceptible *V. vinifera* (data not shown). Expression of a stilbene synthase gene (*VvSTS*) was determined as a positive control of defense gene induction by *P. viticola* infection. As expected, *P. viticola* inoculation triggered *VvSTS* expression in both susceptible and tolerant *Vitis* species (Figure 3A). However, *VvSTS*

expression was enhanced much earlier in resistant *V. riparia*, where transcripts began to accumulate 12 h after inoculation and were stimulated about 20-fold at 2 days. In contrast, maximal induction of *VvSTS* expression was measured 5 days after inoculation in *V. vinifera* cv Chardonnay (Figure 3A). Thus, *VvSTS* transcript accumulation was delayed in susceptible *V. vinifera* cv Chardonnay compared to resistant *V. riparia*.

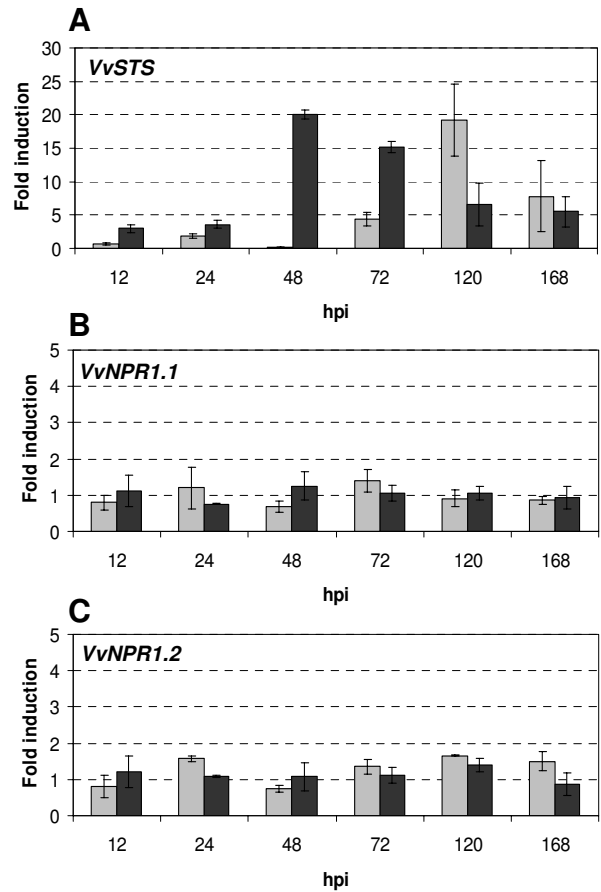


Figure 3
Expression patterns of VvNPR1.1 and VvNPR1.2 during a compatible or an incompatible interaction between grapevine and *Plasmopara viticola*. Leaves of plantlets of *Vitis vinifera* cv Chardonnay (grey bars) and *Vitis riparia* cv Gloire de Montpellier (dark bars) were inoculated with *Plasmopara viticola* (1.5 × 10⁵ spores mL⁻¹). Control leaves were sprayed with water. Leaves were collected at different time points as indicated. Hpi: Hours post inoculation. Transcript levels of each gene (Stilbene synthase *VvSTS* (A); *VvNPR1.1* (B); *VvNPR1.2* (C)) were normalized to actin transcript levels. The fold induction indicates normalized expression levels in inoculated leaves compared to normalized expression levels observed in water-treated leaves at the same time point. Expression ratio at the beginning of the experiment (0) is set to 1. Mean values and standard deviations were obtained from 2 duplicate experiments.

Transcript accumulation of *VvNPR1.1* and *VvNPR1.2* was then quantified after *P. viticola* infection. As shown in Figure 3B and 3C, no significant change in the expression of these two genes was detectable for either genotype. Other studies from our group have shown that constitutive expression of *VvNPR1.1* and *VvNPR1.2* was also not affected by infection with *Botrytis cinerea* or with *Pseudomonas syringae* pv *pisi* (data not shown). Taken together, expression studies suggest that *VvNPR1.1* and *VvNPR1.2* are not regulated at transcriptional level upon pathogen infection.

Subcellular localization of *VvNPR1.1* and *VvNPR1.2*

The amino acid sequences of both *VvNPR1.1* and *VvNPR1.2* were found to contain a putative nuclear localization signal (NLS1) in the C terminus of the protein (Figure 1C). To determine the subcellular localization of *VvNPR1.1* and *VvNPR1.2*, the coding regions of *VvNPR1.1*, *VvNPR1.2*, and *AtNPR1* were fused to 5'-terminus of *eGFP* under the control of the CaMV 35S promoter. The resulting constructs were introduced into *Nicotiana benthamiana* following transient transformation by agroinfiltration. Leaf sectors of agroinfiltrated *N. benthamiana* were observed 3 days after infiltration for GFP fluorescence by confocal microscopy (Figure 4). GFP fluorescence levels were comparable with the 3 constructions studied. Control leaves expressing free GFP yielded a weak fluorescence predominantly visible in the cytoplasm (Figure 4A and 4B). As described previously [30], the *AtNPR1*-GFP fusion protein fluorescence strongly labelled the nucleus (Figure 4C and 4D). Consistent with the presence of the NLS1, *VvNPR1.1*-GFP and *VvNPR1.2*-GFP fusion proteins were localized to the nucleus and to a lesser extent to the cytoplasm both in mesophyll and epidermal cells (Figure 4E and 4F). Localization of GFP fluorescence to nucleus was further observed in cells from peeled epidermis transiently transformed with *VvNPR1.1* (Figure 4G and 4H). Treatment of *N. benthamiana* leaves with SA 48 h before observation did not influence the localization of the fusion proteins (data not shown).

Transient expression of *VvNPR1.1* and *VvNPR1.2* in *N. benthamiana* triggers the accumulation of acidic PR1 and PR2 but not of PR3

To investigate if *VvNPR1.1* and *VvNPR1.2* could control the expression of PR genes (especially the *PR1* gene), PR protein accumulation was analyzed after transient expression of *AtNPR1*-GFP, *VvNPR1.1*-GFP and *VvNPR1.2*-GFP. Leaves of *N. benthamiana* were analyzed 3 days after agroinfiltration for PR protein production by Western blot with anti sera raised against tobacco PR proteins. PR proteins were undetectable in untreated leaves (Figure 5). Transient expression of *AtNPR1*-GFP, *VvNPR1.1*-GFP and *VvNPR1.2*-GFP was sufficient to trigger accumulation of acidic PR1, in contrast to expression of empty vector (encoding free GFP) which produced no signal (Figure 5).

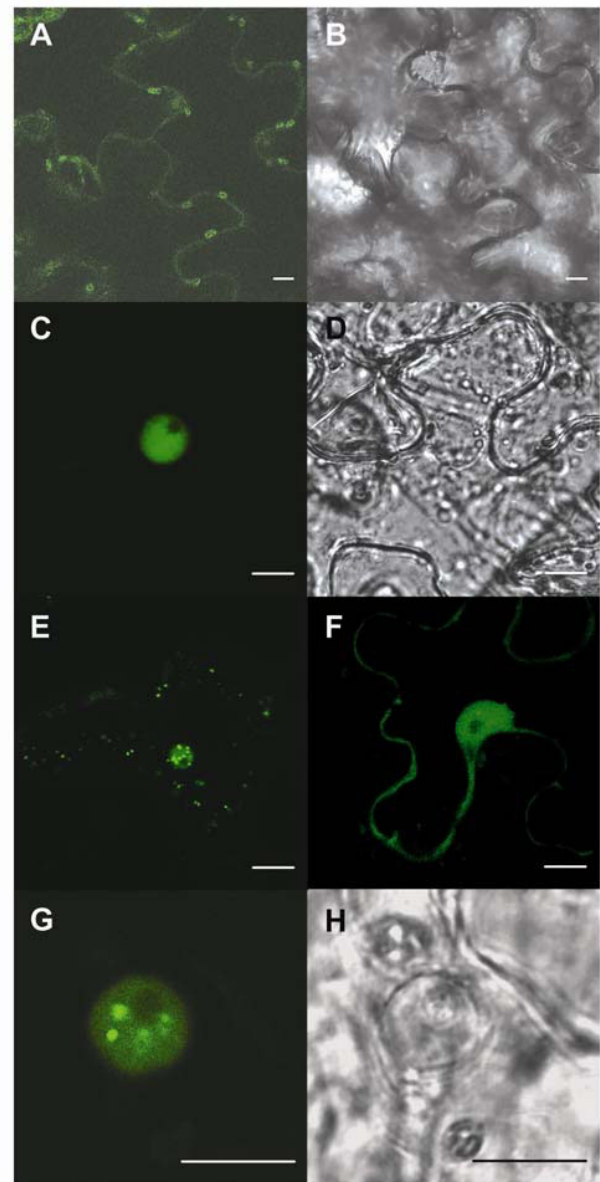


Figure 4
Subcellular localization of *VvNPR1.1* and *VvNPR1.2*.
N. benthamiana leaves were infiltrated with *A. tumefaciens* GV3101 containing empty vector (pK7FWG2) encoding free GFP (A, B), or *AtNPR1* (C, D), *VvNPR1.1* (E, G, H), and *VvNPR1.2* (F) in pK7FWG2. Confocal images were captured 3 days after infiltration. GFP images (A, C, E, F, G) and differential contrast images (B, D, H) of *N. benthamiana* epidermal cells were compared to show the subcellular localization of GFP, *AtNPR1*-GFP, *VvNPR1.1*-GFP and *VvNPR1.2*-GFP. Bar = 10 μ M.

In order to determine if another marker of the SA pathway could be enhanced by *VvNPR1* expression, the same analysis was performed to detect acidic β -1.3 glucanase (PR2). Agroinfiltration of vector alone triggered the expression of PR2 compared to infiltration with H₂O (Figure 5). How-

ever, transient expression of *AtNPR1-GFP*, *VvNPR1.1-GFP* and *VvNPR1.2-GFP* induced a stronger accumulation of PR2 compared to infiltration with empty vector (Figure 5). In order to determine if PR protein induction by *AtNPR1* and *VvNPR1* is specific of SA signalling, we analyzed the accumulation of basic chitinase (PR3), a SA-independent marker whose expression is controlled by the JA/ET pathway in Arabidopsis [2]. Anti-PR3 serum recognized two proteins of 32 and 34 kDa corresponding to the two basic chitinase isoforms described in tobacco [[33], Fig 5]. Similarly to PR2, agroinfiltration with empty vector triggered the expression of PR3 compared to infiltration with H₂O (Figure 5). However, in contrast to PR1 and PR2, expression of *AtNPR1-GFP*, *VvNPR1.1-GFP* and *VvNPR1.2-GFP* did not modify significantly PR3 accumulation compared to empty vector (Figure 5).

Similar results concerning PR protein expression were observed after infiltration of *N. benthamiana* with *Agrobacterium* harbouring the coding regions of *AtNPR1*, *VvNPR1.1* and *VvNPR1.2* under the control of the 35S

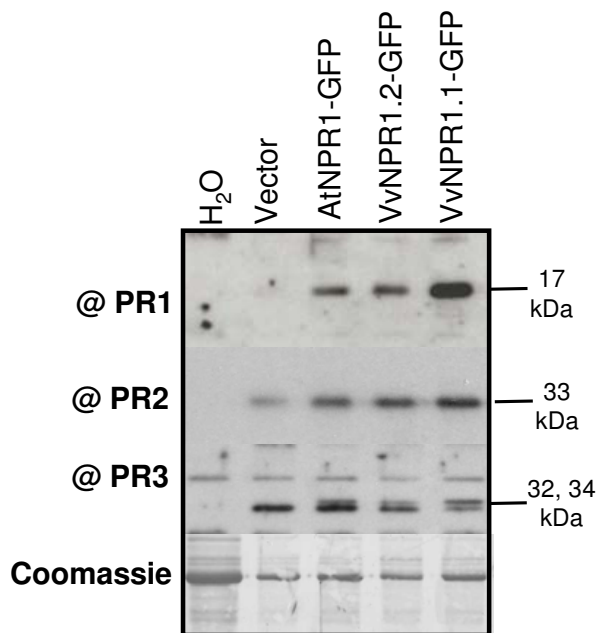


Figure 5
Induction of PR1 and PR2 accumulation in *N. benthamiana* by transient expression of *VvNPR1.1* and *VvNPR1.2*. *N. benthamiana* leaves were infiltrated with water (H₂O) or *A. tumefaciens* GV3101 containing *VvNPR1.1*, *VvNPR1.2*, or *AtNPR1* in pK7FWG2 or empty vector. Leaves were harvested 3 days after agroinfiltration. Soluble proteins were extracted, submitted to SDS-PAGE and probed with sera against tobacco PR1, PR2 or basic chitinases (PR3).

CaMV promoter in a pBinplus vector devoid of GFP (data not shown).

Transient expression of *AtNPR1* and *VvNPR1.1* in grapevine leaves enhances accumulation of *VvPR1* transcripts

Heterologous expression in *N. benthamiana* showed that *VvNPR1.1* and *VvNPR1.2* were able to trigger the accumulation of acidic PR1 and PR2 in the absence of pathogen inoculation. To evaluate the effect of *VvNPR1* expression in a homologous system (*Vitis vinifera*), we used a recently described protocol of transient gene expression by vacuum agroinfiltration in grapevine [34]. *AtNPR1* and *VvNPR1.1*, which is the most closely related to *AtNPR1*, were transiently expressed in leaves of *V. vinifera* cv Syrah, a genotype showing high efficiency of transient expression [34]. Gene expression was first analyzed 3 days after agroinfiltration. Grapevine leaves were also later inoculated with *P. viticola* 3 days after agroinfiltration and analyzed 2 days after oomycete inoculation. To confirm that *AtNPR1* and *VvNPR1.1* were expressed in agroinfiltrated grapevine leaves, we monitored the accumulation of full length transgene-derived mRNAs of *AtNPR1* and *VvNPR1.1* by RT-PCR as shown in Figure 6. No PCR amplification was revealed when omitting the reverse transcription step (data not shown).

Real time quantitative PCR was used to study the expression of *VvPR1* and *VvSTS* in grapevine leaves expressing *AtNPR1* and *VvNPR1.1*, 3 days after agroinfiltration. As shown in Figure 7A, infiltration with empty vector stimulated the expression of *VvPR1*, probably because of the agroinfiltration stress. Interestingly, in leaves expressing *AtNPR1* and *VvNPR1.1*, a stronger increase in *VvPR1* transcript accumulation was measured (Figure 7A). In contrast, no significant increase in *VvSTS* transcript accumulation was measured in leaves expressing *AtNPR1* and *VvNPR1.1* compared to H₂O-infiltrated leaves (Figure 7B). In another experiment, we inoculated grapevine leaves with *P. viticola* 3 days after agroinfiltration and analyzed gene expression 2 days after inoculation. *VvPR1* expression was induced by fungal infection as expected. Consistent with the results obtained in uninoculated leaves, *VvPR1* stimulation in infected leaves was clearly higher in leaves expressing *AtNPR1* and *VvNPR1.1* than in leaves preinfiltrated with control *Agrobacterium* (Figure 7C). Although *VvSTS* expression was stimulated 3 fold by infection, no significant effect on its expression was observed when leaves were preinfiltrated with the different constructs (Figure 7D).

Together, these results show that transient expression of both *AtNPR1* and *VvNPR1.1* in *Vitis vinifera* is able to enhance expression of a grapevine defense gene known to be controlled by the SA signalling pathway in model plants.

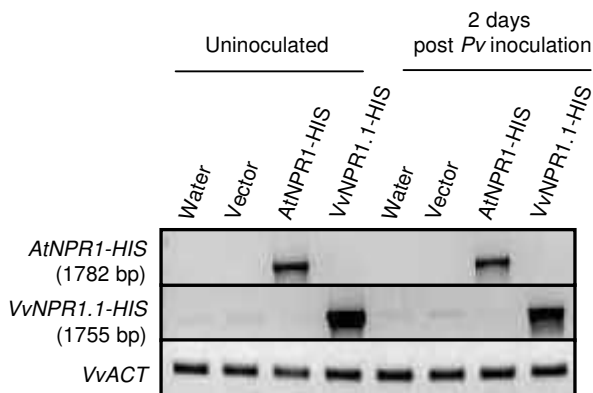


Figure 6
Detection of *AtNPR1* and *VvNPR1.1* transgene expression in grapevine leaves. Leaves from *in vitro* grown *V. vinifera* cv Syrah were infiltrated with *A. tumefaciens* transformed with pBIN+ carrying *AtNPR1* or *VvNPR1.1*. Control plants were infiltrated with water. Infiltrated leaves were challenged with *P. viticola* 3 days after agroinfiltration. Total RNAs were extracted 3 days after agro-infiltration (uninoculated) and 2 days after *P. viticola* inoculation. Full-length mRNA from each transgene was specifically amplified after reverse transcription with primers listed in table 1. *VvACT* was used as internal control.

Discussion

In order to characterize defense response signalling components in grapevine, we identified two homologs of *AtNPR1* in *Vitis vinifera* cv Chardonnay. Our study provides the first elements for the functional characterization of *VvNPR1*.

Expression studies of *VvNPR1.1* and *VvNPR1.2* showed that these genes are constitutively expressed and that expression can be further enhanced by treatment with BTH, a SA analog. Induction of *NPR1* genes by treatment with SA or its analogs has been described in a number of plant species including Arabidopsis, mustard, apple, rice, banana and cotton [4,17-20,35]. Interestingly, *VvNPR1.2* is the most responsive to BTH induction and forms a phylogenetically related group with *MpNPR1*, *AtNPR3* and *AtNPR4* which are also highly induced by BTH or INA (another SA analog) respectively [18,36]. In rice, it has been shown that *OsNPR1* is more rapidly induced in the incompatible interactions leading to resistance than in the compatible interactions leading to disease [17]. Similarly, *MNPR1A* from banana was induced earlier and to higher levels after infection in a *Fusarium oxysporum* tolerant cultivar than in a sensitive one [19]. To evaluate if *VvNPR1* expression could be differentially regulated during compatible or incompatible interactions between *Vitis* species and *Plasmopara viticola*, we examined the expression of both genes after inoculation of susceptible *Vitis vinifera* cv

Chardonnay or resistant *Vitis riparia* cv Gloire de Montpellier with downy mildew. The expression of a gene encoding a stilbene synthase, an enzyme involved in the synthesis of phytoalexins, which is known to be stimulated by *P. viticola* infection was also studied as a positive control. We detected a faster induction of *STS* gene expression after inoculation of the resistant genotype (*Vitis riparia*), consistent with an earlier induction of defense genes in incompatible versus compatible interactions [37]. However, no significant changes in transcript levels were detected for both *VvNPR1.1* and *VvNPR1.2* after infection with downy mildew. Overall, the constitutive expression of *VvNPR1* and the absence of transcriptional regulation after pathogen infection suggest that *VvNPR1* activity is regulated at the protein level in grapevine, as previously described in Arabidopsis [4].

In order to address *VvNPR1* function, particularly its sub-cellular localization and its ability to regulate defense gene expression, we first used an heterologous system for transient expression by agroinfiltration of *N. benthamiana* leaves. This method has been described as a rapid and efficient system for the *in vivo* analysis of plant transcription factors and promoters of *PR* genes [38]. The predicted amino acid sequences of *VvNPR1.1* and *VvNPR1.2* were found to contain a putative nuclear localization signal (NLS1) in their C terminus. Consistently, transiently expressed *VvNPR1*-GFP and *AtNPR1*-GFP fusion proteins were localized predominantly to the nucleus, even in the absence of the SAR inducer SA. Constitutive nuclear localization was also revealed by transient expression of *AtNPR1*-GFP after bombardment of epidermal onion cells [30]. By contrast, in stable transformants, exclusive nuclear localization of *AtNPR1*-GFP, which is required for activation of *PR* gene expression, was triggered only after treatment with a SAR inducer or infection with a pathogen [30]. Similarly, Arabidopsis lines overexpressing *AtNPR1* under the control of the constitutive 35S CaMV promoter and grown under non-inducing conditions have not revealed an increase in the basal level of *PR* genes, indicating that *AtNPR1* is essentially inactive in the absence of pathogen infection. *NPR1*-overexpressing plants will thus not activate SA-dependent defense responses until they are challenged with a pathogen [7].

In this study, we showed by transient expression that *VvNPR1.1* and *VvNPR1.2* are functional in triggering the accumulation of acidic *PR1* and *PR2* in *N. benthamiana*. This effect was obtained in the absence of an exogenous inducer and correlated with the nuclear localization of *VvNPR1.1* and *VvNPR1.2*. It is likely that agroinfiltration of *N. benthamiana* leaves itself induces a biotic stress that activates responses related to SAR, including targeting of *NPR1* proteins to the nucleus. This hypothesis is supported by a higher basal level of *PR* proteins in empty vector-agroinfiltrated leaves compared to leaves infiltrated

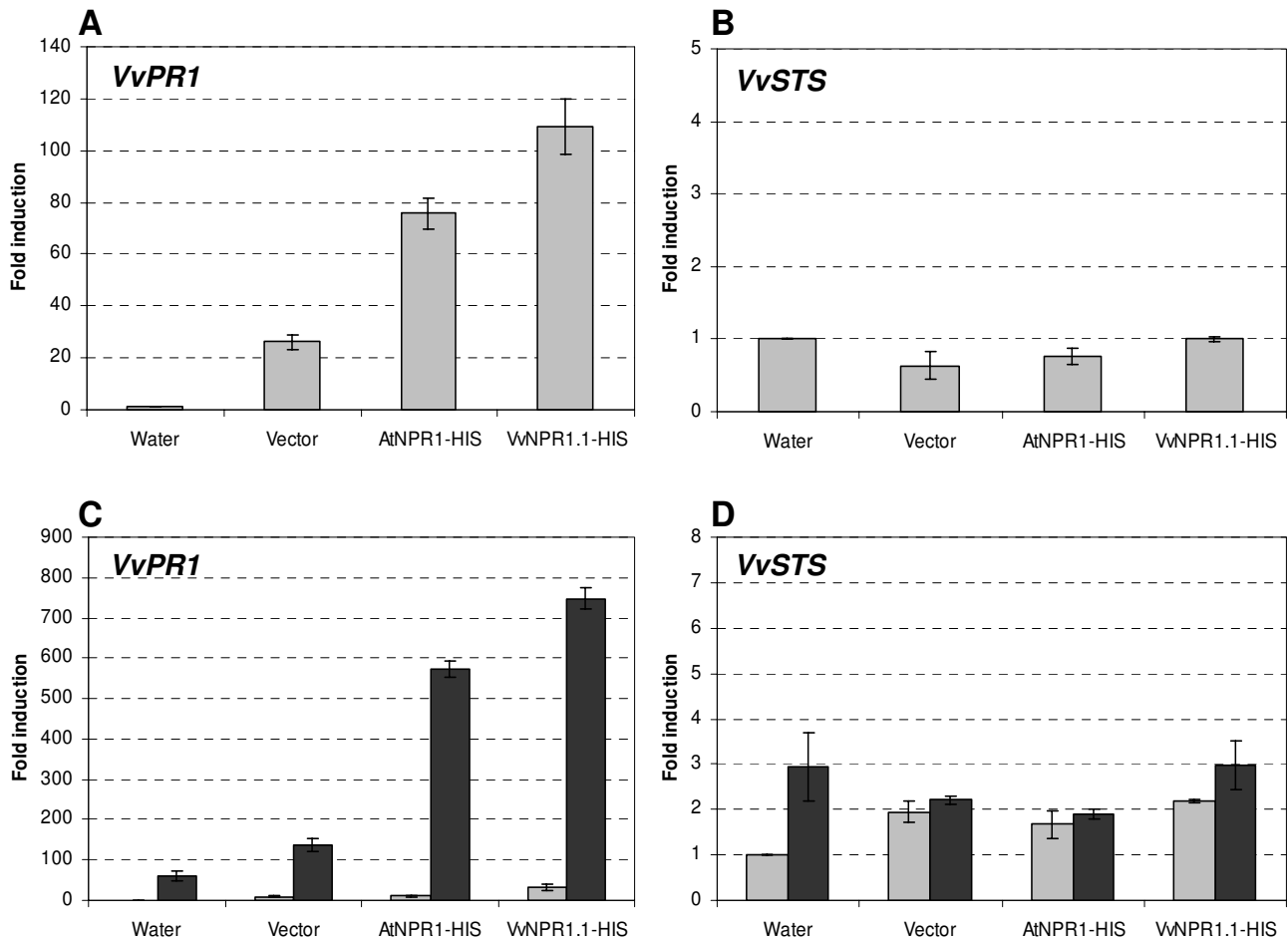


Figure 7

Expression of VvPR1 and VvSTS after transient overexpression of AtNPR1 and VvNPR1.1 in grapevine leaves. (A, B) Expression levels of VvPR1 (A) and VvSTS (B), in uninoculated leaves, 3 days after agro-infiltration. (C, D) Expression levels of VvPR1 (C) and VvSTS (D) in uninoculated and inoculated leaves. Leaves were infiltrated with *Agrobacterium* carrying the different constructs and expression of VvPR1 and VvSTS was analyzed 3 days later (grey bars). Three days after agroinfiltration, leaves were inoculated with *P. viticola* and expression of genes of interest was analyzed 2 days after inoculation (black bars). Fold induction indicates expression levels in agroinfiltrated leaves compared to the expression in non-inoculated water-infiltrated leaves, which is set to 1. Mean values and standard deviations were obtained from 2 duplicate experiments.

with water (Figure 5). Similarly, it has been reported that *Agrobacterium*-mediated transient assays of stress-inducible PR promoters have relatively high levels of GUS activity in water and mock-treatments [38]. Finally, it appears that both grapevine NPR1 are active in *N. benthamiana*, in agreement with the ability of AtNPR1 to activate defense responses in other plant species such as rice and wheat [14,16]. Induction of PR protein accumulation was rather specific of defense markers that have been demonstrated to be SA-specific in tobacco [39]. Conversely, NPR1 expression had no significant effect on basic chitinase (PR3) accumulation. In Arabidopsis, PR3 represents an SA-independent marker whose expression is controlled by the JA/ET pathway [2]. Moreover, class I basic chitinase

expression is activated by overexpression of an ethylene-responsive transcription factor (ERF) in tobacco cells [40].

In order to gain further information on VvNPR1 activity in a homologous system, we used a recently described method of *Agrobacterium*-mediated transient gene expression in *Vitis vinifera* [34]. This system circumvents the time consuming process of generating stable transgenic lines in grapevine. In this study, we provide a first example of successful use of *Agrobacterium*-mediated transient expression for functional analysis of signalling elements in grapevine. AtNPR1 and VvNPR1.1 were successfully expressed at relatively high level in leaves of *V. vinifera* cv Syrah after agroinfiltration. Transient expression of these two signal-

Table 1: Sequence of primers used for semi-quantitative RT-PCR in grapevine

Gene	Accession number	Forward Primer 5' → 3'	Reverse Primer 5' → 3'
VvACT	AF369524 ^a	TGCTATCCTTCGTCTTGACCTTG	GGACTTCTGGACAACGGAATCTC
VvPR1	GSVIVT0003857500 ^b	GGAGTCCATTAGCACTCCTTTG	CATAATTCTGGGCGTAGGCAG
VvNPR1.1	GSVIVT0001653600 ^b	GGAATTCGATGTTGGGTACG	GCAACCTTGTCAGAATGTCC
VvNPR1.2	GSVIVT0003193300 ^b	GCCGTACGGTAAGGTTGGAT	GAGCCTTCCCGATGAAGTTG

^a Genbank accession number^b Genoscope Grape Genome Browser number

ling genes resulted in increased *VvPR1* gene expression in both uninoculated and in *P. viticola* inoculated leaves. In inoculated tissues, the expected stimulation of *PR1* expression by *P. viticola* was observed; however, *PR1* expression was further enhanced in infected leaves overexpressing *AtNPR1* or *VvNPR1.1*. It is likely that the activity of the NPR1 proteins is enhanced by *P. viticola* inoculation. Moreover, it appeared that *VvNPR1.1* had a stronger activity than *AtNPR1* on induction of *PR1* expression in grapevine.

Transient expression in *N. benthamiana* and *V. vinifera* shows that *VvNPR1.1* and *VvNPR1.2* have a positive activity on the expression of *PR1* and *PR2* genes (Figure 5). It is thus likely that as in other plant species, *VvNPR1* controls the expression of a set of SA-responsive defense genes in grapevine. However, it remains to be determined if *VvNPR1.1* and *VvNPR1.2* perform different functions in grapevine defense. Arabidopsis genome contains 3 additional genes closely related to *AtNPR1*, which are likely involved in plant defense responses [36], and 2 other more distant genes, *AtBOP1* (*AtNPR5*) and *AtBOP2* (*AtNPR6*), with functions in the control of growth asymmetry in leaf and floral patterning [13]. Among NPRs involved in plant defense, phylogenetic analysis revealed that *AtNPR1* and *AtNPR2* form a subgroup, whereas

AtNPR3 and *AtNPR4* form a distinct pair [36]. Interestingly, grapevine genome sequencing revealed only two genes related to "defense" *AtNPRs*. *VvNPR1.1* belongs to the subgroup comprising *AtNPR1* and *AtNPR2*, and *VvNPR1.2* forms a distinct subgroup with *AtNPR3*, *AtNPR4* and *MpNPR1-1* from apple (Figure 1). Curiously, a hallmark of this second subgroup is a high inducibility of gene expression by BTH or its analogs [[18,36] and this study]. Different members of the *AtNPR* family appear to mediate different functions in plant defense. *AtNPR1* has been identified as a key positive regulator of SA-dependent gene expression that is required for SAR establishment as well as for basal resistance to virulent pathogens [4]. On the other hand, *AtNPR3* and *AtNPR4* have been proposed to act as negative regulators of plant defense, since the double *npr3npr4* mutant shows elevated basal *PR1* expression and enhanced resistance to virulent bacterial and oomycete pathogens [36]. However, the negative regulation of defense mechanisms by *AtNPR3* and *AtNPR4* is in contradiction with another study where *npr4* single mutants were shown to be more susceptible to the virulent bacterial pathogen *Pseudomonas syringae* pv. tomato DC3000 [12]. In this study, *AtNPR4* was also implicated in the regulation of JA-inducible genes and in the crosstalk between the SA- and the JA-dependent signalling pathways [12]. Even if *VvNPR1.2* is closely related to

Table 2: Sequence of primers used for real-time PCR in grapevine

Gene	Accession number	Forward Primer 5' → 3'	Reverse Primer 5' → 3'
VvACT	AF369524 ^a	TCCTGTGGACAATGGATGGA	CTTGATCCCTCAGCACCTT
VvSTS	DQ366301 ^a	CATCAAGGGTGCTATGCAGGT	TCAGAGCACACCACAAGAACTCG
VvPR1	GSVIVT0003857500 ^b	GGAGTCCATTAGCACTCCTTTG	CATAATTCTGGGCGTAGGCAG
VvNPR1.1	GSVIVT0001653600 ^b	GACCACAACCGAGCTTCTTGATCT	ATAATCTGGGCTCTTTCCGCATT
VvNPR1.2	GSVIVT0003193300 ^b	GCAGGAAACAAACAAGGACAGGAT	CAGCCATTGTTGGTGAAGAGATTG

^a Genbank accession number^b Genoscope Grape Genome Browser number

AtNPR3 and AtNPR4, it is likely not acting as a negative regulator of defense genes since its expression in *N. benthamiana* resulted in enhanced PR1 and PR2 accumulation. Moreover, VvNPR1.2 is closely related to MpNPR1-1, whose overexpression led to activation of PR genes and resistance to bacterial and fungal pathogens in apple [18] (Figure 1). Therefore, phylogenetic analysis is not sufficient to predict a positive or negative control of defense responses for a given member of the NPR family. However, it is likely that the two NPR1 homologs identified in grapevine do not perform fully overlapping functions. Overexpression or silencing of the two genes in grapevine will help to clarify their respective role in resistance to different pathogens in the future.

Conclusion

We show here that genome sequence resources combined with transient expression in heterologous and homologous systems allow to obtain rapidly functional information on grapevine genes. The upregulation of acidic PR1 and PR2 expression by VvNPR1 both in *N. benthamiana* and *Vitis vinifera* strongly suggests that VvNPR1 is a component of the SA defense signalling pathway in grapevine. This implies the existence of highly conserved mechanisms for regulation of defense gene expression among plant species. As a consequence, overexpression of VvNPR1 and other signalling elements has the potential to enhance disease resistance in this crop species. Further work will concentrate on the search for transcription factors interacting with the two VvNPR1 proteins in grapevine, and on the analysis of pathogen tolerance in *npr1* mutant and wild type Arabidopsis overexpressing VvNPR1.1 and VvNPR1.2.

Methods

Biological material

Vitis vinifera cv Chardonnay 96 and *Vitis riparia* cv Gloire de Montpellier were obtained from ENTAV (Etablissement National Technique pour l'Amélioration de la Viticulture, Le Grau du Roi, France). *Vitis vinifera* cv Syrah was provided by INRA (Colmar, France). These clones were propagated on MS medium supplemented with 20 g.L⁻¹ sucrose and 0.7% bacto-agar in a growth chamber at 25°C, under a 16/8 h photoperiod.

Four-week-old *in vitro* plantlets of *Vitis vinifera* cv Chardonnay and *Vitis riparia* were transferred to potting soil (Fertiligène, NFU 44-571) inside a closed translucent propagator under saturating humidity for 7 days. Plantlet acclimatization was realized by gradually raising the propagator's lid. Plants were grown in potting soil for 3 weeks (22°C, 16/8 h photoperiod, 70% humidity) before use for treatments or pathogen inoculation. Eight-week old *in vitro*-grown plants from *Vitis vinifera* cv Syrah were used for *Agrobacterium* infiltration experiments.

Nicotiana benthamiana plants were grown in potting soil under a 16/8 h photoperiod for 2 weeks prior to be used for *Agrobacterium* infiltration. *Plasmopara viticola* was kindly provided by Sabine Merdinoglu (INRA, Colmar, France).

Treatment of plants with chemicals and pathogens

Detached leaves of *Vitis vinifera* cv Chardonnay were sprayed with a BTH solution (80 mg.L⁻¹, Bion, Syngenta Agro AG, Dielsdorf, Switzerland). Control leaves were sprayed with water. Leaves were maintained in sealed Petri dishes on humid Whatmann 3 MM paper, collected at different time points and immediately frozen in liquid nitrogen.

For *Plasmopara viticola* inoculation, *Vitis vinifera* cv Chardonnay and *Vitis riparia* plantlets were placed in a closed translucent propagator. Abaxial leaf surfaces were sprayed with freshly collected sporangia propagated on *V. vinifera* cv Chardonnay and resuspended in water at approximately 1.5 × 10⁵ spores.mL⁻¹. Inoculated plants were placed in a growth chamber at 21°C under obscurity for 24 h, then under a 16/8 h photoperiod for 6 days. Inoculation of *Vitis vinifera* cv Syrah was performed by spraying 10⁴ spores.mL⁻¹ on detached leaves of agroinfiltrated *in vitro*-cultured plantlets that were maintained in sealed Petri dishes on humid Whatmann paper under conditions described above. Leaves were collected at different time points and immediately frozen in liquid nitrogen.

Cloning of VvNPR1.1 and VvNPR1.2

The nucleic acid sequence of Arabidopsis NPR1 was used to search an EST database of abiotically stressed leaves of *V. vinifera* cv Chardonnay (EST Analysis Pipeline, ESTAP [27], using BLASTN. Two EST with significant similarity to AtNPR1 were identified. Full-length cDNAs were reconstituted by searching the Genoscope database of grapevine genome sequencing with the two EST previously identified [28]. Full-length cDNAs of VvNPR1.1 (GSVIVT00016536001) and VvNPR1.2 (GSVIVT00031933001) were amplified from reverse-transcribed RNA from SA-treated Chardonnay leaves using Pfx DNA polymerase (Invitrogen). AtNPR1 cDNA was amplified from reverse-transcribed cDNA from *Arabidopsis thaliana* Col-0 leaves.

For subcellular localization, the AtNPR1, VvNPR1.1 and VvNPR1.2 coding sequences were cloned by Gateway (Invitrogen) recombination reactions into the pK7FWG2 vector [41], upstream of eGFP.

For transient expression in *N. benthamiana* and grapevine leaves, full-length AtNPR1, VvNPR1.1 and VvNPR1.2 cDNAs were cloned between the CaMV 35S promoter and the 35S terminator sequences of the pUCAP-intron vector

[42]. This vector contains an intron between the promoter and the terminator sequence, which was excised and replaced by *NPR1* cDNA sequences. A six histidine tag coding region was added to the 3' end of each cDNA in order to facilitate detection of transgene product. The cassette containing *AtNPR1*, *VvNPR1.1* and *VvNPR1.2* between the CaMV 35S promoter and the 35S terminator was excised by *AscI/PacI* digestion and cloned into the pBINplus vector [43].

Sequence alignment and phylogenetic analysis

Protein sequence alignment was realized using the ClustalW program. The phylogenetic tree was constructed with the Phylo_win program [44], using the Neighbor-Joining method. Bootstrap values were obtained from 500 replicates.

Gene expression analysis by semi-quantitative PCR and real-time quantitative PCR

RNA extraction and DNase I treatment were performed as described in Chong et al. [21]. Reverse transcription was performed on 0.5 µg of RNA with the iScript cDNA synthesis kit (Biorad), according to the manufacturer's instructions.

Semi-quantitative RT-PCR was performed by using recombinant Taq DNA polymerase (Invitrogen, Cergy Pontoise, France). Control reactions to normalize RT-PCR were done with primers derived from grapevine actin sequences. PCR on serial dilutions of cDNA were performed at 55°C and 29 cycles to define semi-quantitative conditions that resulted in amplification linear to RNA amounts. The experiments were performed twice with similar results. Primers used for PCR are listed in table 1.

For real time PCR, reactions were carried out on the iCycler system (Biorad, Marnes-la-Coquette, France). PCR reactions were carried out in triplicates in a reaction buffer containing 1× iQ SYBR® Green Supermix, 0.2 mM of forward and reverse primers and 10 ng of reverse transcribed RNA in a final volume of 25 µl. Thermal cycling conditions were: 30s at 95°C followed by 40 cycles of 15s at 94°C, 30s at 60°C and 30s at 72°C. Acquisition temperatures were 83°C for *VvPR1* and 77°C for *VvACT1*, *VvSTS*, *VvNPR1.1* and *VvNPR1.2*. The calibration curve for each gene was obtained by performing real-time PCR with serial dilutions of the cloned cDNA fragment (from 10² to 10⁸ cDNA copy number). The specificity of the individual PCR amplification was checked using a heat dissociation curve from 55 to 95°C following the final cycle of the PCR. The results obtained for each gene of interest at each time point were normalized to the expression of a reference gene (*VvACT1*) and fold induction compared to H₂O treatment was calculated as described by Pfaffl et al [32]. Mean values and standard deviations were obtained from

2 duplicate experiments and are representative of 2 independent experiments. Primers used for real-time quantitative PCR are listed in table 2.

Transient expression in tobacco and grapevine leaves

For transient expression in tobacco leaves, we used *Agrobacterium tumefaciens* GV3101 transformed with pK7FWG2 or pBINplus carrying *AtNPR1*, *VvNPR1.1* or *VvNPR1.2*. An overnight culture of bacteria containing the appropriate construct was resuspended in the same volume of 10 mM MgCl₂. Bacterial suspension's concentration was adjusted to OD600 = 0.5 with 10 mM MgCl₂. Acetosyringone (200 µM final) was added to the bacterial suspension prior tobacco leaf infiltration using a syringe without needle.

Transient expression in grapevine leaf experiments was realized as described in Santos-Rosa et al. [34]. *A. tumefaciens* C58CI culture transformed with pBINplus carrying *AtNPR1-His* or *VvNPR1.1-His* was prepared as described [34]. Detached leaves from 8- to 10 week-old grown *V. vinifera* cv Syrah were submerged abaxial face down in cylindrical flasks (40 mL) containing 7 mL of bacterial culture. Leaves were covered by a disk of Miracloth. Flasks were then placed into a dessicator. Vacuum was applied for 2 min at 15 mm Hg with an oil-pump (GmbH, Type N035.3AN.18). Vacuum was applied twice for each leaf. Leaves were then placed in sealed Petri dishes on humid Whatmann paper for three days before harvest or inoculation with *Plasmopara viticola* as described in "Biological materials".

Subcellular localization of VvNPR1.1 and VvNPR1.2

AtNPR1, *VvNPR1.1* and *VvNPR1.2* in pK7FWG2 vector [41] were transiently transformed into *Nicotiana benthamiana* by agroinfiltration as described above. Agroinfiltrated leaf sectors were observed 3 days after infiltration. Images were acquired with a LSM510 confocal microscope (Carl Zeiss, software version AIM 4.2), using a 63×, 1.2 NA water immersion objective lens at 23°C. Fluorescence of free GFP or GFP fusion proteins was observed after excitation with a 488 nm laser line, using a 505–550 band-pass emission filter.

Immunoblot analysis of PR proteins

Foliar explants were harvested from *N. benthamiana* infiltrated with *Agrobacterium* carrying *AtNPR1*, *VvNPR1.1* and *VvNPR1.2* in pK7FWG2 vector, 3 days after infiltration.

Total soluble protein was extracted from leaves by grinding in liquid nitrogen and resuspending the powder in extraction buffer as described [30]. The protein concentration of the extract was determined with the Bio-Rad protein assay. SDS PAGE was carried out according to standard procedures with 10 µg of total proteins. Proteins

were electro-transferred on Immobilon P membranes (Millipore, Bedford, MA). Detection was realized with the immune-star chemiluminescent kit (Bio-Rad, Hercules, CA). The blots were probed by using polyclonal antisera raised against an acidic PR-1 isoform (PR 1b, [45]), a β -1.3 glucanase isoform (PR-2, [46]) and basic chitinases (PR-3, [47]) purified from tobacco. Polyclonal antisera were kindly provided by M. Legrand (IBMP, Strasbourg, France) and used at a 1:10 000 dilution. Protein loading was checked by Coomassie Blue staining of membranes. The experiment was performed twice with similar results.

Abbreviations

NPR1: non expressor of PR genes 1; SA: salicylic acid; BTH: benzothiadiazole; INA: 2.6-dichloroisonicotinic acid; JA: jasmonic acid; ET: ethylene; PR: pathogenesis related; SAR: systemic acquired resistance; BTB/POZ: broad complex, tramtrack and bric a brac/pox virus and zinc finger; ARD: ankyrin repeat domain; NLS: nuclear localization signal.

Authors' contributions

GLH carried out most of the experiments, *ie*, gene cloning and phylogenetic analyses, expression studies, transient expression in grapevine, and participated in transient expression in *N. benthamiana*.

TH participated in the design of the study, gave advices for GFP localization experiments and helped to draft the manuscript.

PM participated in the design of the study and helped for transient expression in *V. vinifera*.

JM did the confocal microscopy observations.

BW conceived the study and has done general supervision.

JC carried out transient expression in *N. benthamiana* and western blot analyses, performed conceptual and experimental design and drafted the manuscript.

All authors read and approved the final manuscript.

Acknowledgements

We are grateful to Sabine Merdinoglu (INRA, Colmar) for providing *Plasmodium viticola* isolate, and to Michel Legrand (IBMP, Strasbourg) for the anti-PR protein sera. Special thanks to Christophe Bertsch (LVBE, Colmar) for helpful discussions and to Pierrette Geoffroy (IBMP, Strasbourg) for taking care of *N. benthamiana* plants. This work was supported by the Université de Haute Alsace and by a doctoral fellowship from the French Ministry of Research to GLH.

References

- de Wit PJ: **How plants recognize pathogens and defend themselves.** *Cell Mol Life Sci* 2007, **64(21)**:2726-2732.

- Thomma BP, Eggermont K, Penninckx IA, Mauch-Mani B, Vogelsang R, Cammue B, Broekaert WF: **Separate jasmonate-dependent and salicylate-dependent defense response pathways in Arabidopsis are essential for resistance to distinct microbial pathogens.** *Proc Natl Acad Sci USA* 1998, **95**:15107-15111.
- Loake G, Grant M: **Salicylic acid in plant defence – the players and protagonists.** *Curr Opin Plant Biol* 2007, **10(5)**:466-472.
- Dong X: **NPR1, all things considered.** *Curr Opin Plant Biol* 2004, **7(5)**:547-552.
- Cao H, Bowling SA, Gordon AS, Dong X: **Characterization of an Arabidopsis mutant that is nonresponsive to inducers of systemic acquired resistance.** *Plant Cell* 1994, **11**:1583-1592.
- Cao H, Glazebrook J, Clarke JD, Volko S, Dong X: **The Arabidopsis NPR1 gene that controls systemic acquired resistance encodes a novel protein containing ankyrin repeats.** *Cell* 1997, **88**:57-63.
- Cao H, Li X, Dong X: **Generation of broad-spectrum disease resistance by overexpression of an essential regulatory gene in systemic acquired resistance.** *Proc Natl Acad Sci USA* 1998, **95(11)**:6531-6536.
- Tada Y, Spoel SH, Pajerowska-Mukhtar K, Mou Z, Song J, Wang C, Zuo J, Dong X: **Plant immunity requires conformational changes of NPR1 via S-nitrosylation and thioredoxins.** *Science* 2008, **321(5891)**:952-956.
- Mou Z, Fan W, Dong X: **Inducers of plant systemic acquired resistance regulate NPR1 function through redox changes.** *Cell* 2003, **113(7)**:935-944.
- Eulgem T, Somssich IE: **Networks of WRKY transcription factors in defense signaling.** *Curr Opin Plant Biol* 2007, **10(4)**:366-371.
- Pieterse CM, van Wees SC, van Pelt JA, Knoester M, Laan R, Gerrits H, Weisbeek PJ, van Loon LC: **A novel signaling pathway controlling induced systemic resistance in Arabidopsis.** *Plant Cell* 1998, **10(9)**:1571-1580.
- Liu G, Holub EB, Alonso JM, Ecker JR, Fobert PR: **An Arabidopsis NPR1-like gene, NPR4, is required for disease resistance.** *Plant J* 2005, **41(2)**:304-318.
- Hepworth SR, Zhang Y, McKim S, Li X, Haughn GW: **BLADE-ON-PETIOLE-dependent signaling controls leaf and floral patterning in Arabidopsis.** *Plant Cell* 2005, **17(5)**:1434-1448.
- Chern MS, Fitzgerald HA, Yadav RC, Canlas PE, Dong X, Ronald PC: **Evidence for a disease-resistance pathway in rice similar to the NPR1-mediated signaling pathway in Arabidopsis.** *Plant J* 2001, **27(2)**:101-113.
- Lin WC, Lu CF, Wu JW, Cheng ML, Lin YM, Yang NS, Black L, Green SK, Wang JF, Cheng CP: **Transgenic tomato plants expressing the Arabidopsis NPR1 gene display enhanced resistance to a spectrum of fungal and bacterial diseases.** *Transgenic Res* 2004, **13(6)**:567-581.
- Makandar R, Essig JS, Schapaugh MA, Trick HN, Shah J: **Genetically engineered resistance to Fusarium head blight in wheat by expression of Arabidopsis NPR1.** *Mol Plant Microbe Interact* 2006, **19(2)**:123-129.
- Yuan Y, Zhong S, Li Q, Zhu Z, Lou Y, Wang L, Wang J, Wang M, Li Q, Yang D, et al.: **Functional analysis of rice NPR1-like genes reveals that OsNPR1/NH1 is the rice orthologue conferring disease resistance with enhanced herbivore susceptibility.** *Plant Biotechnol J* 2007, **5(2)**:313-324.
- Malnoy M, Jin Q, Borejsza-Wysocka EE, He SY, Aldwinckle HS: **Overexpression of the apple MpNPR1 gene confers increased disease resistance in Malus x domestica.** *Mol Plant Microbe Interact* 2007, **20(12)**:1568-1580.
- Endah R, Beyene G, Kiggundu A, Berg N van den, Schluter U, Kunert K, Chikwamba R: **Elicitor and Fusarium-induced expression of NPR1-like genes in banana.** *Plant Physiol Biochem* 2008, **46(11)**:1007-1014.
- Zhang Y, Wang X, Cheng C, Gao Q, Liu J, Guo X: **Molecular cloning and characterization of GhNPR1, a gene implicated in pathogen responses from cotton (Gossypium hirsutum L.).** *Biosci Rep* 2008, **28(1)**:7-14.
- Chong J, Le Henaff G, Bertsch C, Walter B: **Identification, expression analysis and characterization of defense and signaling genes in Vitis vinifera.** *Plant Physiol Biochem* 2008, **46(4)**:469-481.
- Jeandet P, Douillet-Breuil AC, Bessis R, Debord S, Sbaghi M, Adrian M: **Phytoalexins from the Vitaceae: biosynthesis, phytoalexin gene expression in transgenic plants, antifungal activity, and metabolism.** *J Agric Food Chem* 2002, **50(10)**:2731-2741.

23. Marchive C, Mzid R, Deluc L, Barrieu F, Pirrello J, Gauthier A, Corio-Costet MF, Regad F, Cailleteau B, Hamdi S, et al.: **Isolation and characterization of a *Vitis vinifera* transcription factor, VvWRKY1, and its effect on responses to fungal pathogens in transgenic tobacco plants.** *J Exp Bot* 2007, **58(8)**:1999-2010.
24. Mzid R, Marchive C, Blancard D, Deluc L, Barrieu F, Corio-Costet MF, Drira N, Hamdi S, Lauvergeat V: **Overexpression of VvWRKY2 in tobacco enhances broad resistance to necrotrophic fungal pathogens.** *Physiol Plant* 2007, **131(3)**:434-447.
25. Jaillon O, Aury JM, Noel B, Policriti A, Clepet C, Casagrande A, Choise N, Aubourg S, Vitulo N, Jubin C, et al.: **The grapevine genome sequence suggests ancestral hexaploidization in major angiosperm phyla.** *Nature* 2007, **449(7161)**:463-467.
26. Velasco R, Zharkikh A, Troggo M, Cartwright DA, Cestaro A, Pruss D, Pindo M, Fitzgerald LM, Vezzulli S, Reid J, et al.: **A high quality draft consensus sequence of the genome of a heterozygous grapevine variety.** *PLoS ONE* 2007, **2(12)**:e1326.
27. **EST Analysis Pipeline** [<http://staff.vbi.vt.edu/estap/>]
28. **Grape Genome Browser** [<http://www.genoscope.cns.fr/externe/GenomeBrowser/Vitis/>]
29. Hulo N, Bairoch A, Bulliard V, Cerutti L, Cuče BA, de Castro E, Lachaize C, Langendijk-Genevaux PS, Sigrist CJ: **The 20 years of PROSITE.** *Nucleic Acids Res* 2008, **36**:D245-249.
30. Kinkema M, Fan W, Dong X: **Nuclear localization of NPR1 is required for activation of PR gene expression.** *Plant Cell* 2000, **12(12)**:2339-2350.
31. Ryals J, Weymann K, Lawton K, Friedrich L, Ellis D, Steiner HY, Johnson J, Delaney TP, Jesse T, Vos P, et al.: **The Arabidopsis NIM1 protein shows homology to the mammalian transcription factor inhibitor I kappa B.** *Plant Cell* 1997, **9(3)**:425-439.
32. Pfaffl MW: **A new mathematical model for relative quantification in real-time RT-PCR.** *Nucleic Acids Res* 2001, **29(9)**:e45.
33. Legrand M, Kauffmann S, Geoffroy P, Fritig B: **Biological function of pathogenesis-related proteins: Four tobacco pathogenesis-related proteins are chitinases.** *Proc Natl Acad Sci USA* 1987, **84(19)**:6750-6754.
34. Santos-Rosa M, Poutaraud A, Merdinoglu D, Mestre P: **Development of a transient expression system in grapevine via agroinfiltration.** *Plant Cell Rep* 2008, **27(6)**:1053-1063.
35. Meur G, Budatha M, Gupta AD, Prakash S, Kirti PB: **Differential induction of NPR1 during defense responses in *Brassica juncea*.** *Physiol Mol Plant Pathol* 2006, **68**:128-137.
36. Zhang Y, Cheng YT, Qu N, Zhao Q, Bi D, Li X: **Negative regulation of defense responses in Arabidopsis by two NPR1 paralogs.** *Plant J* 2006, **48(5)**:647-656.
37. Kortekamp A: **Expression analysis of defence-related genes in grapevine leaves after inoculation with a host and a non-host pathogen.** *Plant Physiol Biochem* 2006, **44**:58-67.
38. Yang Y, Li R, Qi M: **In vivo analysis of plant promoters and transcription factors by agroinfiltration of tobacco leaves.** *Plant J* 2000, **22(6)**:543-551.
39. Kang MK, Park KS, Choi D: **Coordinated expression of defense-related genes by TMV infection or salicylic acid treatment in tobacco.** *Mol Cells* 1998, **8(4)**:388-392.
40. Nakano T, Nishiuchi T, Suzuki K, Fujimura T, Shinshi H: **Studies on transcriptional regulation of endogenous genes by ERF2 transcription factor in tobacco cells.** *Plant Cell Physiol* 2006, **47(4)**:554-558.
41. Karimi M, Inze D, Depicker A: **GATEWAY vectors for *Agrobacterium*-mediated plant transformation.** *Trends Plant Sci* 2002, **7(5)**:193-195.
42. La Camera S, Geoffroy P, Samaha H, Ndiaye A, Rahim G, Legrand M, Heitz T: **A pathogen-inducible patatin-like lipid acyl hydrolase facilitates fungal and bacterial host colonization in Arabidopsis.** *Plant J* 2005, **44(5)**:810-825.
43. van Engelen FA, Molthoff JW, Conner AJ, Nap JP, Pereira A, Stiekema WJ: **pBINPLUS: an improved plant transformation vector based on pBIN19.** *Transgenic Res* 1995, **4(4)**:288-290.
44. Galtier N, Gouy M, Gautier C: **SEAVIEW and PHYLO_WIN: two graphic tools for sequence alignment and molecular phylogeny.** *Comput Appl Biosci* 1996, **12(6)**:543-548.
45. Kauffmann S: **Les protéines PR (Pathogenesis-Related) du Tabac: des protéines impliquées dans les réactions de défense aux agents pathogènes. Isolement, propriétés sérologiques et activités biologiques.** Université Louis Pasteur, Strasbourg, Institut de Biologie Moléculaire des Plantes du CNRS; 1988.
46. Kauffmann S, Legrand M, Geoffroy P, Fritig B: **Biological function of 'pathogenesis-related' proteins: four PR proteins of tobacco have 1,3-beta-glucanase activity.** *Embo J* 1987, **6(11)**:3209-3212.
47. Heitz T, Jung J-L, Fritig B, Hahne G, Legrand M: **Local and systemic accumulation of PR proteins in tobacco plants infected with tobacco mosaic virus.** *Mol Plant Microbe Interact* 1994, **7**:776-779.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

