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Evaluating Resistance to Bt Toxin Cry1Ab by F₂ Screen in European Populations of *Ostrinia nubilalis* (Lepidoptera: Crambidae)

H. ENGELS,¹ D. BOURGUET,² L. CAGÁN,³ B. MANACHINI,⁴ I. SCHUPHAN,¹ T. J. STODOLA,⁵
A. MICOUD,⁶ C. BRAZIER,⁶ C. MOTTET,⁶ AND D. A. ANDOW⁵

ABSTRACT The large-scale cultivation of transgenic crops producing *Bacillus thuringiensis* (Bt) toxins have already lead to the evolution of Bt resistance in some pest populations targeted by these crops. We used the F₂ screening method for further estimating the frequency of resistance alleles of the European corn borer, *Ostrinia nubilalis* (Hübner) (Lepidoptera: Crambidae), to Bt maize, *Zea mays* L., producing the Cry1Ab toxin. In France, Germany, and Italy, 784, 455, and 80 lines of European corn borer were screened for resistance to Mon810 maize, respectively. In Slovakia, 26 lines were screened for resistance to the Cry1Ab toxin. The cost of F₂ screen performed in the four countries varied from US\$300 to \$1,300 per line screened. The major difference in cost was mostly due to a severe loss of univoltine lines during the screen in Germany and Slovakia. In none of the screened lines did we detect alleles conferring resistance to Mon810 maize or to the Cry1Ab toxin. The frequency of resistance alleles were $<1.0 \times 10^{-3}$, $<1.6 \times 10^{-3}$, $<9.2 \times 10^{-3}$, and $<2.6 \times 10^{-2}$ in France, Germany, Italy, and Slovakia, with 95% probability, respectively. The average detection probability over all lines was $\approx 90\%$. Making the assumption that European corn borer populations in these countries belong to the same genetic entity, the frequency of alleles conferring resistance to the Cry1Ab produced by the Mon810 maize in western and central Europe was 1.0×10^{-4} , with a 95% confidence interval of $0-3.0 \times 10^{-4}$.

KEY WORDS European corn borer, Bt maize, Mon810, resistance management, HDR strategy

Transgenic crops producing *Bacillus thuringiensis* (Bt) toxins—referred to as Bt crops—are widely planted in the United States, India, Argentina, Brazil, Canada, China, and Spain (James 2009). For example, maize (*Zea mays* L.), which provides control of lepidopteran and coleopteran maize pests, has been planted on >200 million ha during the last decade, corresponding to 30.5% of the global biotech area (James 2009). Large-scale cultivation of these Bt crops is exerting a tremendous selection pressure on target pest species, which prove to evolve field resistance (Matten et al. 2008; Kruger et al. 2009; Tabashnik et al. 2008, 2009), diminishing the benefits of these crops.

The risk of target pests becoming resistant to Bt crops has led to the development of several strategies

to manage transgenic crops (Gould 1998, Roush 1998, Shelton et al. 2002, Vacher et al. 2003, Zhao et al. 2005, Tuytuynov et al. 2008). Among these the most widely accepted and implemented strategy is the “high-dose/refuge” (HDR) strategy (Alstad and Andow 1995). This strategy has two components. First, the Bt crops need to produce Bt toxin at high concentration, with the expectation that this concentration would be sufficient to kill most if not all heterozygous individuals. The other part of this strategy is refuges—the occurrence of non-Bt plants, crops, or weeds, preserving a pool of susceptible individuals. The HDR is expected to work best when high dose of toxin is expressed in plant tissue, resistance alleles are rare (initial frequencies $<10^{-3}$), and functionally recessive and refuges are close to Bt plants, so that nearly all resistant survivors from the Bt plants will mate with susceptible individuals from refuge plants to produce heterozygous offspring that cannot survive on Bt plants the following generation (Alstad and Andow 1995).

Several techniques have been used to estimate the frequency of resistance alleles in natural populations, including in-field screening (Venette et al. 2000) and screening of field-collected egg masses or larvae (Siegfried et al. 2007). When resistance alleles have not been identified and are believed to be rare and recessive, Andow and Alstad (1998, 1999) and Andow and Ives (2002) suggest that the most efficient method

¹ Institute for Environmental Research (Biologie V), Aachen University, 52074 Aachen, Germany.

² Corresponding author: Centre de Biologie pour la Gestion des Populations (CBGP), UMR INRA-IRD-CIRAD-Montpellier SupAgro, Campus International de Baillarguet, 34988 Montferrier sur Lez, France (e-mail: bourguet@supagro.inra.fr).

³ Department of Plant Protection, Slovak Agricultural University, 94976 Nitra, Slovakia.

⁴ Department of Animal Biology University of Palermo, 90123 Palermo, Italy.

⁵ Department of Entomology, University of Minnesota, St. Paul, MI 55108.

⁶ Unité Résistance aux Produits Phytosanitaires, AFSSA, 69364 Lyon, France.

is an F₂ screen. This technique preserves genetic variation among isofemale lines and concentrates resistance alleles into homozygous genotypes in the F₂ generation. F₂ screens have been used for the European corn borer, *Ostrinia nubilalis* (Hübner) (Lepidoptera: Crambidae) (Andow et al. 1998, 2000; Bourguet et al. 2003, Stodola et al. 2006). Unlike F₂ screens performed on other pest species (He et al. 2001, Génissel et al. 2003, Wenes et al. 2006, Downes et al. 2007, Huang et al. 2007, Xu et al. 2009) no major Bt resistance alleles, i.e., alleles conferring a level of resistance sufficiently high to allow survival and development of homozygous individuals on Bt crops, have been detected in the European corn borer. Accordingly, all laboratory selections (Bolin et al. 1999, Chaufaux et al. 2001) or field monitoring using discriminating doses of toxin (Farinos et al. 2004, Siegfried et al. 2007) performed on this pest failed to reveal such major Bt resistance alleles. Still, the F₂ screen allowed an estimation of the frequency of resistance alleles in natural populations, which has been valuable for resistance management planning.

The goal of this study is to continue the investigation of Bt resistance alleles in European European corn borer populations by using the F₂ screening method. Andow et al. (1998, 2000), Stodola et al. (2006), and Bourguet et al. (2003) performed F₂ screens on U.S. Corn Belt and French populations of this pest. Here, we extend the investigation in western and central Europe to determine whether 1) the high-dose refuge strategy could be implemented with success for managing the cultivation of Bt maize expressing the Cry1Ab toxin such as the Mon810 and 2) the cost of the F₂ screen makes it a sustainable method to monitor the evolution of Bt resistance in European populations of the European corn borer.

Materials and Methods

F₂ Screen

The F₂ screen protocol for *O. nubilalis* was adapted from Andow and Alstad (1998). Mated females are considered to be the preferred stage for initiating an F₂ screen, but many variant methods have been proposed (Bentur et al. 2000, Zhao et al. 2002, Stodola and Andow 2004, Stodola et al. 2006). For our F₂ screens either mated females, pair-mated adults originating from field-collected late instars, pair-mated adults originating from egg masses collected in the field or large field-collected egg masses were used. For the pair-mated adults from egg masses, only one adult was taken from an egg mass for pair-mating. For the large field-collected egg masses, only large (>60 eggs) egg masses were used to set F₁ families from an isofemale line, bypassing the need to establish the P₁ generation. In France, lines were exclusively started from paired adults of field-collected diapausing larvae. In Italy and Slovakia, wild-mated females were used to establish isofemale lines. From each isofemale line, the F₁ offspring were reared and sib-mated and the F₂ neonates were screened either on Bt maize (hybrids expressing

the Mon810 transformation event and therefore producing a truncated Cry1Ab toxin) leaves or diet containing the Cry1Ab toxin. Starting from the two P₁ adults, each isofemale line allowed us to characterize at least four haplotypes. By sib-mating the F₁ generation, 1/16 of the F₂ individuals were expected to be homozygous for the resistance allele if one of the P₁ adults carried such an allele.

Insect Collection, Rearing, and Screening

France. Insect Collection. Pest insects were collected from 1999 to 2004 in four different regions in France: Aquitaine, Centre, Midi-Pyrénées, and Rhône-Alpes. By comparison, the F₂ reported by Bourguet et al. (2003) was performed on populations collected in Midi-Pyrénées only. All European corn borers were collected as diapausing larvae on maize stalks during the autumn or early winter. Only one larva was taken per stalk to minimize the probability that siblings were collected. Each isofemale line was started by pairing a virgin male and a virgin female obtained after breaking diapause (for details, see Bourguet et al. 2003). Each pair of female and male was placed at 25°C, 70% RH, and a photoperiod of 16:8 (L:D) h in a small plastic container equipped with papersheets on top for oviposition. All pairs were held until enough F₁ egg masses were collected or females died.

F₁ Rearing and F₂ Screening. F₁ egg masses hatched on average after 6 d. For each line, F₁ larvae were reared at 25°C, 70% RH, and a photoperiod of 16:8 (L:D) h in a large plastic box (13 by 11 cm) filled with the artificial maize diet described by Gahukar (1975). Diet was added or changed twice a week. The larval development before pupation took on average 30 d. F₁ adults were transferred to a new large plastic box equipped with papersheets on top for oviposition (Bourguet et al. 2003). The F₂ generation of each isofemale line was screened for resistance by inoculating neonates on leaves of at least two different Bt maize plants (Elgina, event Mon810). These leaves were evaluated for surviving larvae at 5 d after inoculation. All lines with survivors were scored as potential positives.

Germany. Insect Collection. European corn borer sampling took place in 2003–2005 at four different German locations. During the summer, adults were collected using light trap cages. Moths were brought to the laboratory and transferred in pairs to small cages covered with waxed paper for oviposition. Caging females together with a male was to ensure the mating of females that might not have mated in the field. Late instars of *O. nubilalis* were collected during fall 2003 and 2004 from Bonn and Heilbronn. Only one larva was taken from a single maize plant to avoid the collection of brothers and sisters originated from the same egg mass. Larvae were kept in groups of 15 individuals in petri dishes and provided with an artificial European corn borer diet based on Wyniger (1974) and Farinos et al. (2004) and held at 25°C, 70% RH, and continuous day light to try to prevent diapause. Pupation time varied strongly between indi-

viduals and many larvae did not pupate at all. Once a week the dishes were checked for pupae, which were removed and males and females were kept separately in plastic boxes until the adults emerged. After emergence, males and females were paired. Moths were provided with a solution of 10% honey in water and held at 25°C, 70% RH, and a photoperiod of 16:8 (L:D) h. Rearing procedures of F_1 larvae, F_1 adults, and F_2 screening on Bt maize has been performed using identical protocol described below, regardless of the initiation method of the line.

F_1 Rearing and F_2 Screening. After mating the P_1 adults in the cages, egg masses were collected daily, placed in petri dishes with European corn borer diet and held at 25°C, 70% RH, and a photoperiod of 16:8 (L:D) h until the blackened head capsules were visible within the eggs. To synchronize the offspring of a female, egg masses were held at 10°C until the female had finished oviposition. The egg masses from a single female were combined and larvae were reared at 25°C, 70% RH, and a photoperiod of 16:8 (L:D) h. Larvae were transferred weekly to new dishes containing fresh diet, and after the second larval instar the density was limited to 15 individuals per dish. F_1 pupae from different families were kept separately, and emerged adults were transferred daily to F_1 family cages for sib-mating. Moths were provided with a solution of 10% honey in water to maximize the oviposition rate.

Neonates hatching from F_2 egg masses were screened on excised Bt maize (Novelis, event Mon810) leaves. F_2 screens were performed in plastic boxes (10 by 10 cm). Five to 10 European corn borer egg masses were transferred into each box together with six to eight leaves of Bt maize tissue over a moistened filter paper. Every three days, new Bt leaves were added and the filter paper was remoistened. The boxes were held in the same environmental conditions as the F_1 generation. All dishes were checked for surviving larvae after 7–10 d. Resistance was only assumed when larvae survived beyond the second larval instar with additional evidence of feeding on Bt maize leaves.

Italy. Insect Collection. Adult European corn borer females were collected in Italy from 2003 to 2004 in three different regions (Lombardia, Piemonte, and Veneto). Wild-mated females had a life span of ≈ 6 d. They were individually held in cylindrical mesh cages with paper on top for oviposition. To synchronize the emergence of first instars, egg masses were collected daily and were stored at 13°C and 85% RH until eggs were blackheaded. Larvae and moths were reared at 25°C, high relative humidity, and a photoperiod of 16:8 (L:D) h.

F_1 Rearing and F_2 Screening. Larvae hatched after 3–4 d. They were reared in plastic dishes with artificial diet (Lozzia and Manachini 2003), added when necessary. After ≈ 30 d, larvae pupated and pupae were stored in white folded paper. If necessary pupae were synchronized and held at 13°C with wet cotton until 30 pupae were collected. F_1 adults were transferred to family cages for sib-mating and were provided with a water-honey solution. Egg masses were collected

from paper placed on top of the cages. Hatching F_2 larvae were screened on excised Bt maize (Elgina, event Mon810) leaves or Cry1Ab toxin.

Slovakia. Insect Collection. Wild-mated European corn borer females were collected by sweep netting during 2004 and 2005 at Komjatice and Nitra. Those females had a lifespan from 2 to 4 d and were held in small mesh cages with paper on top for oviposition. Eggs were collected daily, stored in petri dishes at 85% RH and 10°C until the last egg laying of each female. The eggs were stored at 25°C for the next few days until the occurrence of black heads of future larvae. Black headed egg masses were placed on pieces of aluminum foil and added on the diet (≈ 150 ml) in canning jars. In 2004, the artificial diet described by Wyniger (1974) was used in the experiment. A simplified artificial diet described by Nagy (1970) was used in 2005. Larvae and moths were reared at 25°C and 70% RH.

F_1 Rearing and F_2 Screening. F_1 egg masses were transferred to petri dishes. Larvae hatched after 3–4 d with an estimated proportion of $\approx 50\%$. One hundred larvae per dish were reared in 250-ml canning jars with diet. Pupae were collected daily from dishes and were placed in F_1 family cages, where they emerged after ≈ 7 d. Egg masses were collected from paper hanging inside the cages. Neonates hatching from F_2 egg masses were screened on diet with Cry1Ab toxin. We performed the F_2 screen using the Cry1Ab toxin (provided by Johannes A. Jehle, Agricultural Service Centre for Rhineland-Palatinate, Germany) laid on the top of the diet at a concentration of $2 \mu\text{g}/\text{cm}^2$.

Statistical Analysis

Results from each F_2 screen were analyzed for the expected resistance allele frequency ($E[p_R]$) with its 95% credibility interval and the probability of a false negative (P_{No}). Bayesian analysis was used to estimate $E[p_R]$ and credibility intervals, which allow conclusions about the populations sampled (Andow and Alstad 1998, 1999; Schneider 1999). A custom program compiled in C⁺⁺ was used to compute P_{No} . It depended on the number of F_1 males and F_1 females, the number of F_2 offspring screened per isofemale line, and the nonscreen mortality of F_2 larvae. Analytical methods for calculating P_{No} are described in Stodola and Andow (2004).

Cost of F_2 Screens

We estimated the fixed and variable costs for the F_2 screens performed in each country. We took into account the salaries for scientific and nonscientific staff, personnel for insect collection and rearing, travel costs for field sampling, and all expenses for consumables and associated infrastructure, including cage construction. The overall cost was divided by the number of isofemale lines tested, providing an average cost per screened line.

Table 1. Number of lines progressing through the F₂ screen experiment

Yr	Location	P ₁ lines started	P ₁ mated ^a	F ₁ larvae produced	F ₁ adults produced	Screened
F ₂ screen lines started from wild-mated female						
2003	Germany	37	24	24	15	9
	Italy	69		47	46	22
2004	Germany	118	61	61	60	60
	Italy	189		147	112	58
	Slovakia	426	131	79	53	16
2005	Slovakia	753	99	51	50	10
Lines started from paired adults of diapausing larvae						
1999	France	152	78	74	62	52
2000	France	347	229	229	127	104
2001	France	339	226	143	116	95
2002	France	461	295	190	180	169
2003	France	750	541	311	310	186
2004	France	329	223	193	190	178
	Germany	≈150	100	100	100	93
2005	Germany	144	144	144	144	140
Lines started from single large F ₁ egg mass						
2005	Germany	200 ^b		167	167	153

^a Line was considered mated if a female laid fertile eggs.

^b Number of F₁ egg masses collected for this purpose.

Results

Multiple methods for starting F₂ screen lines have been used from 1999 to 2005 in the four countries involved in this study. Despite the methodological variation, all methods provided comparable results.

In France, several lines were lost before they were screened because of unmated P₁ females and hatching problems of the F₁ egg masses. The proportion of hatched F₁ eggs varied widely from 20 to 80%. In Germany, the losses were due to several factors. P₁ moths died before oviposition or did not mate, F₁ egg masses did not hatch or collected larvae did not pupate. Losses of collected larvae and adults were high and varied from 43 to 75%. The highest losses of adults were due to their transport to the laboratory. The lowest losses were from starting the lines with large egg masses, which circumvented the need to establish a P₁ generation. In Italy, the main losses during the F₂ screen were due to unmated P₁ and F₁ females. The proportion of hatched F₁ eggs was only 56%. Finally, in Slovakia, the massive loss of lines was due to both unmated females and larvae that remained in dia-

pause. The proportion of hatched F₁ eggs was not exactly recorded, but it was estimated to be ≈50%.

In total, 784, 455, 80, and 26 isofemale lines were screened in France, Germany, Italy, and Slovakia, respectively (Table 1). For all these lines, there were no surviving F₂ larvae. Thus, major resistance alleles to Cry1Ab were detected in none of the European populations tested in this study (Table 2). Hence, E[P_R] = 3.0 × 10⁻⁴, 4.0 × 10⁻⁴, 3.1 × 10⁻³ and 9 × 10⁻³ in France, Germany, Italy, and Slovakia, respectively (Table 3). The detection probability associated with these estimates varied from 84 to 98% (Table 3).

The cost for the F₂ screen was ≈US\$300, \$700, \$400, and \$1,300 per screened line in France, Germany, Italy, and Slovakia, respectively.

Discussion

Frequency of Alleles Conferring Cry1Ab Resistance. The F₂ generations of the lines tested so far in U.S. and French populations of the European corn borer were all susceptible to the concentration of Cry1Ab toxin produced by Mon810 Bt maize. Similarly, none of 1,345 lines screened in the current study contained alleles allowing European corn borer larvae to actively feed on Mon810 maize. Hence, the frequency of such alleles are <1.0 × 10⁻³, <1.6 × 10⁻³, <9.2 × 10⁻³, and <2.6 × 10⁻² in France, Germany, Italy, and Slovakia, with 95% probability, respectively.

In Europe, the population genetic structure is not fully understood yet. Populations of *Ostrinia* feeding on mugwort and hop in France belong to *O. scapularis*, a sibling species of the European corn borer (Frolov et al. 2007). Conversely, no or low genetic differentiation has been detected among European corn borer populations collected on maize throughout France (Bourguet et al. 2000, Malausa et al. 2007) or among several European corn borer populations—also collected on maize—from France, Germany, Italy, Slovakia, Spain, and Greece (Gaspers 2009) and among Italian European corn borer population feeding on rice, *Oryza sativa* L., and on maize (Manachini et al. 2007). Similarly, there is no clear genetic differentiation between European corn borer populations feeding on maize in western and central Europe (Gaspers 2009). Consequently, in western and central Europe, *O. nubilalis* probably constitute more or less a single

Table 2. Results from F₂ screen and subsequent rescreening of positive lines^a

Country	Yr	Screen generation	Screening method	Partial positives	Full positives
France	1999–2004	F ₂	Bt maize	0	0
Germany	2003–2005	F ₂	Bt maize	0	0
Italy	2003	F ₂	Diet, incorporated	— ^b	— ^b
	2004	F ₂	Bt maize	0	0
Slovakia	2004	F ₂	Diet, surface	3	0
	2005	F ₂	Diet, surface	0	0
	2005	F ₃	Bt maize, incorporation to diet	0	0

^a A full positive result required many larvae feeding and growing to at least the third instar on the screening media. A partial positive required a rating ≥ 2 on the Bolin scale (when tested on Bt maize), evidence of feeding and tunneling (when tested on diet treated with Bt), or larvae surviving to the second instar on either Bt maize or diet treated with Bt.

^b Could not interpret results due to problems with toxin.

Table 3. Expected frequencies of major recessive resistance alleles in various European populations of *O. nubilalis*, with experimentwise probability of detecting a resistance allele if one were present in the line

Location	Yr	Lines screened	E _[Pr] (95% CI)	Experimentwise detection prob.
France	1999–2004	784	0.0003 (0–0.0010)	0.950
	1999–2004 ^a	1,595	0.0002 (0–0.0005)	0.872
Germany	2003–2005	455	0.0004 (0–0.0016)	0.975
Italy	2003–2004	80	0.0031 (0–0.0092)	0.914
Slovakia	2004–2005	26	0.0090 (0–0.026)	0.837 ^b

^a Includes data previously published in Bourguet et al. (2003).

^b The screen results from 10 of these lines could not be interpreted due to problems with the toxin. They are not included in further calculations of E_[Pr] and P_{No}.

panmictic population so that resistance management and monitoring could be considered at this large scale.

If we considered populations of France, Germany, Italy, and Slovakia as part of the same population, then we can pool the results of the various F₂ screens reported here as well as those published previously by Bourguet et al. (2003). Over these four countries, a total of 2,156 lines of European corn borer have been screened from 1999 to 2004 (Table 3) with no evidence of resistance to Bt maize in any of those lines. The expected frequency of resistance over these 2,156 lines is 1.0×10^{-4} with a 95% CI of $0-3.0 \times 10^{-4}$ and an experiment-wise detection probability of $\approx 90\%$. This provides the most extensive monitoring data on resistance in any target insect pest species, including all transgenic insecticidal crops and all insecticides. It indicates that resistance is rare enough for a successful deployment of the HDR strategy throughout western Europe and possibly also in central Europe, although additional F₂ screens and further investigation of the genetic structure from this area must be performed before any firm conclusion.

Cost of F₂ Screen. The F₂ screen is a complex, time- and labor-consuming method that requires skilled staff experienced in insect rearing and susceptibility testing. Its cost strongly depends on the number of isofemales tested in a row and, more importantly, on the proportion of lines that are ultimately screened for susceptibility. Hence, the cost varied from \approx US\$300 (for the screens performed in France) to \approx US\$1,300 per line (for the screens performed in Slovakia). The cost of US\$700 per line for the screens performed in Germany is similar to the F₂ screen expenses of Andreadis et al. (2007) on *S. nonagrioides*. However, this latter pest is considerably more difficult to rear than *O. nubilalis*.

The highest costs found in Slovakia (US\$1,300) and in Germany (US\$700) are mostly due to the presence, in these two countries, of univoltine populations (Cañ 1998). These univoltine individuals are genetically constrained (Stengel and Schubert 1982), and a large fraction of the larvae do not pupate in the laboratory, strongly reducing the number of lines that can be tested. Univoltine populations also occur in France, but they are mostly located in north-eastern France. Localities where sampling took place in France were typically not in the northeast, and probably had a low percentage of univoltine larvae.

In Italy and in southern France, populations are mainly bivoltine. This offers two advantages. First, all larvae reared at 25°C with a photoperiod of 16:8 (L:D) h pupate ≈ 10 d after molting to the fifth instar. Hence, most P₁ lines producing offspring are likely to be tested for their susceptibility to Bt maize. The second advantage is the occurrence of two European corn borer flights during the summer. This increases the window of time during which wild-mated females can be collected and screened. Indeed, compared with diapausing larvae, which can be stored at 4°C for several months, wild moths must be handled immediately during the days after their capture. Because all lines must be managed more or less at the same time, this limits the number of lines that can be started. The extended window of time offered by the bivoltine populations could be very interesting when a limited fraction of wild females produce fertile egg masses like—a situation encountered in Slovakia but not during the previous F₂ screens performed in France (Bourguet et al. 2003).

Finally, in Germany, the relatively high cost per line is partly due to the low—sometimes very low—density of European corn borer in maize field. Indeed, the low levels of European corn borer infestation increased the costs of sampling. Conversely, these costs were lower in Italy, notably in northern Italy and southern France where maize fields are often heavily infested by *O. nubilalis*. Noteworthy, European countries and regions suffering strong bivoltine infestations are those where Bt maize is more likely to be planted and where European corn borer resistance to those maize are more likely to be selected.

In conclusion, the F₂ screen is a valuable technique for screening the susceptibility of European corn borer populations to Bt toxins and Bt maize. Isofemale lines can be started using either wild-moths captured within or around maize fields during the summer or adults emerging from diapausing larvae collected during the fall–winter. In studies performed so far on European corn borer (Andow et al. 1998, 2000; Bourguet et al. 2003; Stodola et al. 2006; this study), the experiment-wise detection probability was often higher than 90% and even reached 99%. However, the current study reveals some limitations of the F₂ screen. The screen must occur during the second laboratory generation. Hence, it requires the complete development of at least one generation in the

laboratory. In the German and Slovakian screens, most larvae originated from a univoltine population and did not pupate in the laboratory. As a result, many lines were lost, greatly increasing the cost per line screened. A cost >US\$500 per univoltine line is certainly too high for any kind of monitoring program. Hence, although F₂ screen is probably the best option for resistance monitoring in European regions where maize fields are infested by bivoltine or multivoltine populations of *O. nubilalis*, this question remains open in other regions where maize fields host univoltine populations of this pest. Additional research on breaking diapause in univoltine populations is needed before F₂ screens can be used effectively in these populations.

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References Cited

Alstad, D. N., and D. A. Andow. 1995. Managing the evolution of insect resistance to transgenic plants. *Science* 268: 1894–1896.

Andow, D. A., and D. N. Alstad. 1998. F₂ Screen for rare resistance alleles. *J. Econ. Entomol.* 91: 572–578.

Andow, D. A., and D. N. Alstad. 1999. Credibility interval for rare resistance allele frequencies. *J. Econ. Entomol.* 94: 755–758.

Andow, D. A., and A. R. Ives. 2002. Monitoring and adaptive resistance management. *Ecol. Appl.* 12: 1378–1390.

Andow, D. A., D. N. Alstad, Y.-H. Pang, P. C. Bolin, and W. D. Hutchison. 1998. Using an F₂ screen to search for resistance alleles to *Bacillus thuringiensis* toxin in European corn borer (Lepidoptera: Crambidae). *J. Econ. Entomol.* 91: 579–584.

Andow, D. A., D. M. Olson, R. L. Hellmich, D. N. Alstad, and W. D. Hutchison. 2000. Frequency of resistance to *Bacillus thuringiensis* toxin Cry1Ab in an Iowa population of European corn borer (Lepidoptera: Crambidae). *J. Econ. Entomol.* 93: 26–30.

Andreadis, S. S., F. Alvarez-Alfageme, I. Sanchez-Ramos, T. J. Stodola, D. A. Andow, P. G. Milonas, M. Savopoulou-Soultani, and P. Castañera. 2007. Frequency of resistance to *Bacillus thuringiensis* toxin Cry1Ab in Greek and Spanish populations of *Sesamia nonagrioides* (Lepidoptera: Noctuidae). *J. Econ. Entomol.* 100: 195–201.

Bentur, J. S., D. A. Andow, M. B. Cohen, A. M. Romena, and F. Gould. 2000. Frequency of alleles conferring resistance to a *Bacillus thuringiensis* toxin in a Philippine population of *Scirpophaga incertulas* (Lepidoptera: Pyralidae). *J. Econ. Entomol.* 93: 1515–1521.

Bolin, P. C., W. D. Hutchison, and D. A. Andow. 1999. Long-term selection for resistance to *Bacillus thuringien-*

sis CryIA(c) endotoxin in a Minnesota population of European corn borer (Lepidoptera: Crambidae). *J. Econ. Entomol.* 92: 1021–1030.

Bourguet, D., M.-T. Bethenod, N. Pasteur, and F. Viard. 2000. Gene flow in the European corn borer *Ostrinia nubilalis*: implications for the sustainability of transgenic insecticidal maize. *Proc. R. Soc. Lond. B.* 267: 117–122.

Bourguet, D., J. Chaufaux, M. Seguin, C. Buisson, J. L. Hinton, T. J. Stodola, P. Porter, G. Cronholm, L. L. Buschman, and D. A. Andow. 2003. Frequency of alleles conferring resistance to Bt maize in French and US corn belt populations of *Ostrinia nubilalis*. *Theor. Appl. Genet.* 106: 1225–1233.

Cagán, L. 1998. Voltinism of the European corn borer, *Ostrinia nubilalis* Hbn., in Slovakia. *Plant Prot. Sci.* 34: 81–86.

Chaufaux, J., M. Séguin, J. J. Swanson, D. Bourguet, and B. D. Siegfried. 2001. Chronic exposure of the European corn borer (Lepidoptera: Crambidae) to Cry1Ab *Bacillus thuringiensis* toxin. *J. Econ. Entomol.* 94: 1564–1570.

Downes, S., R. J. Mahon, and K. M. Olsen. 2007. Monitoring and adaptive resistance management in Australia for Bt-cotton: current status and future challenges. *J. Inv. Pathol.* 95: 208–213.

Farinos, G. P., M. de la Poza, P. Hernandez-Crespo, F. Ortega, and P. Castañera. 2004. Resistance monitoring of field populations of the corn borers *Sesamia nonagrioides* and *Ostrinia nubilalis* after 5 years of Bt maize cultivation in Spain. *Entomol. Exp. Appl.* 110: 23–30.

Frolov, N. A., D. Bourguet, and S. Ponsard. 2007. Reconsidering the taxonomy of several *Ostrinia* species in the light of reproductive isolation: a tale for E. Mayr. *Biol. J. Linn. Soc.* 91: 49–72.

Gahukar, R. T. 1975. Nouvelles techniques adoptées pour l'élevage d'*Ostrinia nubilalis* Hübner sur milieu artificiel. *Ann. Zool. Ecol. Anim.* 7: 491–498.

Gaspers, C. 2009. The European corn borer (*Ostrinia nubilalis*, Hbn.), its susceptibility to the Bt-toxin Cry1F, its pheromone races and its gene flow in Europe in view of an insect resistance management. Dissertation RWTH, Aachen University, Germany.

Génissel, A., S. Augustin, C. Courtin, G. Pilate, P. Lorme, and D. Bourguet. 2003. Initial frequency of alleles conferring resistance to *Bacillus thuringiensis* poplar in a field population of *Chrysomela tremulae*. *Proc. R. Soc. Lond. B.* 270: 791–797.

Gould, F. 1998. Sustainability of transgenic insecticidal cultivars: integrating pest genetics and ecology. *Annu. Rev. Entomol.* 43: 701–726.

He, D. J., J. L. Shen, W. J. Zhou, and C. F. Gao. 2001. Using F₂ genetic method of isofemale lines to detect the frequency of resistance alleles to *Bacillus thuringiensis* toxin from transgenic Bt cotton in cotton bollworm (Lepidoptera: Noctuidae). *Cotton Sci.* 13: 105–108.

Huang, F., B. R. Leonard, and D. A. Andow. 2007. Sugarcane borer (Lepidoptera: Crambidae) resistance to transgenic *Bacillus thuringiensis* maize. *J. Econ. Entomol.* 100: 164–171.

James, C. 2009. Global status of commercialized biotech/GM. ISAAA Brief, no. 36. International Service for the Acquisition of Agri Biotech Applications, Ithaca, NY.

Kruger, M., J.B.J. Van Rensburg, and J. Van den Berg. 2009. Perspective on the development of stem borer resistance to Bt maize and refuge compliance at the Vaalharts irrigation scheme in South Africa. *Crop Prot.* 28: 684–689.

Lozzia, G. C., and B. Manachini. 2003. Susceptibility of *Ostrinia nubilalis* Hübner (Lepidoptera: Crambidae) to *Bacillus thuringiensis* var. *kurstaki*. *Bull. Insect* 50: 215–219.

- Malausa, T., A. Dalecky, S. Ponsard, P. Audiot, R. Streiff, Y. Chaval, and D. Bourguet. 2007. Genetic structure and gene flow in French populations of two *Ostrinia* taxa: host races or sibling species? *Mol. Ecol.* 16: 4210–4222.
- Manachini, B., A. Zanardo, and A. Spada. 2007. Using AFLP to investigate the genetic similarity of *Ostrinia nubilalis* (Lepidoptera: Crambidae) infesting rice and corn. *Proceedings IV Temperate Rice Conference*: 350–351.
- Matten, S. R., G. P. Head, and H. D. Quemada. 2008. How governmental regulation can help or hinder the integration of Bt crops within IPM programs, pp. 27–39. *In* J. Romeis, A. M. Shelton, and G. G. Kennedy (eds.), *Integration of insect resistant genetically modified crops within IPM programs*. Springer, New York.
- Nagy, B. 1970. Rearing of the European corn borer (*Ostrinia nubilalis* Hbn.) on a simplified artificial diet. *Acta Phytopathol. Acad. Sci. Hung.* 5: 73–79.
- Roush, R. T. 1998. Two-toxin strategies for management of insecticidal transgenic crops: can pyramiding succeed where pesticide mixtures have not? *Phil. Trans. R. Soc. Lond. B* 353: 1777–1786.
- Schneider, J. C. 1999. Confidence interval for Bayesian estimates of resistance allele frequencies. *J. Econ. Entomol.* 92: 755.
- Shelton, A. M., J. Z. Zhao, and R. T. Roush. 2002. Economic, ecological, food safety, and social consequences of the deployment of Bt transgenic plants. *Annu. Rev. Entomol.* 47: 845–881.
- Siegfried, B. D., T. Spencer, A. L. Crespo, N. P. Storer, G. P. Head, E. D. Owens, and D. Guyer. 2007. Ten years of monitoring for Bt resistance in the European corn borer: what we know, what we don't know and what we can do better. *Am. Entomol.* 53: 208–214.
- Stengel, M., and G. Schubert. 1982. Etude comparative de la vitesse de croissance et de la sensibilité à la photopériode de deux races de pyrales du maïs (*Ostrinia nubilalis* Hübner, Lepidoptera, Pyralidae) et de leurs hybrides. *Agronomie* 2: 989–994.
- Stodola, T. J., and D. A. Andow. 2004. F₂ screen variations and associated statistics. *J. Econ. Entomol.* 97: 1756–1764.
- Stodola, T. J., D. A. Andow, A. R. Hyden, J. L. Hinton, J. J. Roark, L. L. Buschman, P. Porter, and G. B. Cronholm. 2006. Frequency of resistance to *Bacillus thuringiensis* toxin Cry1Ab in southern U.S. Corn Belt population of European corn borer (Lepidoptera: Crambidae). *J. Econ. Entomol.* 99: 502–507.
- Tabashnik, B. E., A. J. Gassmann, D. W. Crowder, and Y. Carrière. 2008. Insect resistance to Bt crops: evidence versus theory. *Nat. Biotechnol.* 26: 199–202.
- Tabashnik, B. E., J. B. J. Van Rensburg, and Y. Carrière. 2009. Field-evolved insect resistance to Bt crops: definition, theory, and data. *J. Econ. Entomol.* 102: 2011–2025.
- Tuytuynov, Y., E. Zhadanovskaya, D. Bourguet, and R. Arditi. 2008. Landscape refuges delay resistance of the European corn borer to Bt-maize: a demo-genetic dynamic model. *Theor. Pop. Biol.* 74: 138–146.
- Vacher, C., D. Bourguet, F. Rousset, C. Chevillon, and M. E. Hochberg. 2003. Modelling the spatial configuration of refuges for a sustainable control of pests: a case study of Bt cotton. *J. Evol. Biol.* 16: 378–387.
- Venette, R. C., W. D. Hutchinson, and D. A. Andow. 2000. An in-field screen for early detection and monitoring of insect resistance to *Bacillus thuringiensis* in transgenic crops. *J. Econ. Entomol.* 93: 1055–1064.
- Wenes, A.-L., D. Bourguet, D. A. Andow, C. Courtin, G. Carré, P. Lorme, L. Sanchez, and S. Augustin. 2006. Frequency and fitness cost of resistance to *Bacillus thuringiensis* in *Chrysomela tremulae* (Coleoptera: Chrysomelidae). *Heredity* 97: 127–134.
- Wyniger, R. 1974. *Insektenzucht. Methoden der Zucht und Haltung von Insekten und Milben im Laboratorium*. Verlag Eugen Ulmer, Stuttgart, Germany.
- Xu, Z., F. Li, J. Chen, F. Huang, D. A. Andow, Y. Wang, Y. C. Zhu, and J. Shen. 2009. Using an F₂ screen to monitor frequency of resistance alleles to Bt cotton in field populations of *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae). *Pest Manag. Sci.* 65: 391–397.
- Zhao, J. Z., Y. X. Li, H. L. Collins, and A. M. Shelton. 2002. Examination of the F₂ screen for rare resistance alleles to *Bacillus thuringiensis* toxins in the diamondback moth (Lepidoptera: Plutellidae). *J. Econ. Entomol.* 95: 14–21.
- Zhao, J. Z., J. Cao, H. L. Collins, S. L. Bates, R. T. Roush, E. D. Earle, and A. M. Shelton. 2005. Concurrent use of transgenic plants expressing a single and two *Bacillus thuringiensis* genes speeds insect adaptation to pyramided plants. *Proc. Natl. Acad. Sci. U.S.A.* 102: 8426–8430.
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