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# 11Genetic parameters and QTL analysis of $\delta^{13}$ C and ring width 12in maritime pine

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# 1**Abstract**

2Classical quantitative genetics and quantitative trait dissection analysis (QTL) 3approaches were used in order to investigate the genetic determinism of wood 4cellulose carbon isotope composition ( $\delta^{13}$ C, a time integrated estimate of water use 5efficiency) and of diameter growth and their relationship on adult trees (15 years) of 6a forest tree species (maritime pine). We used a half diallel experimental set-up to (1) 7estimate heritabilities for  $\delta^{13}$ C and ring width and (2) to decompose the phenotypic  $8\delta^{13}C$  / growth correlation into its genetic and environmental components. We found 9considerable variation for  $\delta^{13}$ C (range of over 3‰) and for ring width (range of over 105 mm) and significant heritabilities (narrow sense 0.17 / 0.19 for  $\delta^{13}$ C and ring width, 11 respectively, 100% additivity). The significant phenotypic correlation between  $\delta^{13}$ C 12and ring width was not determined by the genetic component, but was attributable to 13environmental components. Using a genetic linkage map of a full-sib family, four 14significant and four suggestive QTLs were detected for  $\delta^{13}$ C, the first for  $\delta^{13}$ C in a 15 forest tree species, as far as known to the authors. Two significant and four 16suggestive QTLs were found for ring width. No co-location of QTLs was found 17between  $\delta^{13}$ C and growth.

# 18Keyword Index :

19stable isotope, <sup>13</sup>C, growth, water use efficiency, heritability, quantitative trait, Pinus 20pinaster, tree rings

# 1 Introduction

2In given environmental conditions, trees with high water-use efficiency at the leaf 3level (intrinsic WUE, defined as the ratio of net CO<sub>2</sub> assimilation rate *A* to stomatal 4conductance for water vapour *g*) can maintain higher growth rates under water 5limited conditions than trees with lower WUE (Sun *et al.* 1996; Nguyen-Queyrens *et* 6*al.* 1998). Measurements of plant carbon isotope composition ( $\delta^{13}$ C) provide time-7integrated estimates of WUE (Farquhar, O'Leary & Berry 1982; Farquhar & 8Richards 1984) that can be applied to adult trees (Zhang & Marshall 1994; Zhang & 9Marshall 1995; Guehl *et al.* 1995; Sun *et al.* 1996). Assessments of differences in 10WUE among- and within-tree species are facilitated by the crown- and time-11integrative nature of tree ring  $\delta^{13}$ C.

12Forest tree species are known to be among the most polymorphic species of the flora 13(Hamrick, Godt & Sherman-Broyles 1992). Genotypic differences in leaf  $\delta^{13}$ C of 14conifer species were found among provenances in common garden studies (Zhang, 15Marshall & Jaquish 1993; Zhang *et al.* 1994; Zhang *et al.* 1995; Guehl *et al.* 1995; 16Nguyen-Queyrens *et al.* 1998). It has been suggested that provenance differences of 17 $\delta^{13}$ C might be determined by differences in stomatal sensitivity to changes in vapour 18pressure deficit (Zhang *et al.* 1995) and/or differences in plant hydraulic 19characteristics (Guehl *et al.* 1995). However, differences of  $\delta^{13}$ C among genetic 20families within provenances of *Picea mariana* (Mill.) were found to be mainly 21determined by differences in photosynthetic capacity (Johnsen & Major 1995; Major 22& Johnsen 1996). Similar indications were obtained for maritime pine (*Pinus* 23*pinaster* Ait.) by Guehl *et al.* (1995).

24Tree growth is an important goal for forest tree breeding programs. To avoid 25inadvertent negative selection for growth when selecting for high WUE, it is 26important to know if  $\delta^{13}$ C and growth are genetically linked. Positive but weak 27phenotypic relationships between  $\delta^{13}$ C and height or diameter growth (Flanagan & 28Johnsen 1995; Johnsen *et al.* 1999; Nguyen-Queyrens *et al.* 1998) have been found 29among trees within different forest tree species. Genetic parameters calculated for 30physiological or morphological traits can disentangle phenotypic relationships into 31genotypic and environmental components. Johnsen *et al.* (1999) found strong genetic 32correlations between  $\delta^{13}$ C and tree height or tree diameter. They concluded that *A* 33was determining  $\delta^{13}$ C and growth performance and thus constituted probably the link 34between the two traits. However, since  $\delta^{13}$ C as an indicator of WUE could be either 35controlled by *A* and / or by *g*, there is not necessarily a strong relationship between *A* 36and  $\delta^{13}$ C. This suggests that the existence of a genetic correlation between  $\delta^{13}$ C and 37growth is depending on the factor by which WUE is controlled.

38Adaptive traits, like  $\delta^{13}$ C, are characterised by high phenotypic variation among and 39within populations of forest tree species (Meinzer *et al.* 1992; Zhang *et al.* 1993; 40Zhang *et al.* 1995; Flanagan *et al.* 1995; Nguyen-Queyrens *et al.* 1998). Moreover, 41high heritabilities for  $\delta^{13}$ C have been found for non-woody (Matus, Slinkard & Van 42Kessel 1995; Asay, Johnson & Palazzo 1998) and woody species (Johnsen *et al.* 431999). The development of genetic mapping (Tanksley 1993) has made it possible to 44localize genetic factors controlling quantitative traits (QTLs, Quantitative Trait 45Loci). High heritability of a trait is a favourable factor for quantitative trait dissection 46analysis. 1In crop plant breeding, improvement of WUE has been an important aim and 2therefore the first QTLs for  $\delta^{13}$ C were detected in tomato (Martin *et al.* 1989).

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2therefore the first QTLs for  $\delta^{13}$ C were detected in tomato (Martin *et al.* 1989). 3Mansur *et al.* (1993) found in a preliminary investigation of  $\delta^{13}$ C on soybean one 4large genomic region that could be responsible for as much as 53% of the observed 5variation. In a study of three weeks old barley plants (Pakniyat *et al.* 1997), twelve 6AFLP markers were detected for  $\delta^{13}$ C, two of these markers alone accounted for 753.2% of the variation. QTLs for water use efficiency, as measured by the ratio of 8dry weight to water used, were found in soybean (Mian *et al.* 1996; Mian, Ashley & 9Boerma 1998).

10Detection of QTL on woody species, however, is still in development, due to long 11generation time and therefore the lack of controlled crosses. Genetic maps have often 12to be constructed from F1 full-sib progenies. Carlson *et al.* (1991) were the first to 13show that RAPD primers could be screened for informative markers segregating in a 141:1 ratio in diploid tissue of full-sib progenies. Grattapaglia and Sederoff (1994) 15extended this idea in constructing parental maps of an interspecific eucalyptus hybrid 16family in a mapping strategy named "two-way pseudo-testcross". It was further used 17in conifers (Kubisiak *et al.* 1996, Arcade *et al.* 2000) with RAPDs and AFLPs. For 18maritime pine the genome coverage required for linkage map construction and QTL 19analysis was achieved by using RAPD markers (Plomion *et al.*1995a; Plomion, 200'Malley & Durel 1995b) and AFLP analysis (Costa *et al.* 2000).

21Our objectives were: 1) To estimate the variability and heritability of  $\delta^{13}$ C and ring 22width in a forest tree species (*Pinus pinaster* Ait.) using a half-diallel experimental 23design. 2) To investigate the phenotypic correlation between  $\delta^{13}$ C and growth. 3) To 24separate the phenotypic correlation between  $\delta^{13}$ C and growth into a genetic and an 25environmental component. 4) To dissect  $\delta^{13}$ C and ring width into mendelian inherited 26components (quantitative trait dissection analysis) using a *Pinus pinaster* Ait. full-sib 27family. 5) To compare QTLs for  $\delta^{13}$ C with QTLs for growth.

#### 1Material and Methods

# 2Half Diallel

3A twelve by twelve half-diallel of maritime pine (Pinus pinaster Ait.) was used to 4estimate the variability, heritability and genetic correlations among the studied traits. 5Parental trees were crossed in 1980 and seeds from the controlled crosses sown in a 6nursery in spring 1982 and planted in autumn 1982. The 12 parents were trees 7phenotypically selected for stem growth and straightness in the local provenance of 8the Landes de Gascogne. The half-diallel was located in Cestas (Gironde, France, 944°N44' 0°W44') on a semi-humid podzolic soil. Spacing was 4 m between rows and 101.1 m between individual trees, i.e. 2272 trees/ha. No selfed crosses were analysed, 11therefore the half-diallel consisted of 66 families (12 female and 11 male parents) of 125 to 15 individuals each. Three families were not available, therefore only 63 13 families were analysed. A parentage test was performed using 3 microsatellites 14(Gerber et al. 2000) confirming the authenticity of the progenies used in the half 15diallel. The experimental design consisted of 74 incomplete randomised blocks (the 16 large number of blocs is due to the fact that the presented half-diallel is part of a 17much larger complete diallel). For the present study, 564 trees were cut in march 181997 (trees were fifteen years old). Disks were sampled, dried in a greenhouse and 19analysed for carbon isotopic composition ( $\delta^{13}$ C) and ring width as described below.

#### 20Full-sib family

21A three-generation outbred pedigree comprising 202 fifteen-year-old trees was used 22to study the genetic architecture of the studied traits, i.e. the number, genome 23location and effect of Quantitative Trait Loci. The four grand parents were trees 24phenotypically selected for stem growth and straightness in the local provenance of 25the Landes de Gascogne and grafted in clonal archives. These grand parents were 26tested in a polycross progeny test and classified according to their breeding value as 27"Vigor +" (for vigorous trees) and "Vigor -" (for less vigorous trees). Each of the 28parental trees is the result of the cross of one "Vigor +" and one "Vigor -" 29grandparent. The two parental trees were crossed in 1980 and seeds from the 30controlled cross-sown in spring 1982. They produced progeny seedlings that were 31planted in autumn 1982. The family was located in Malente (Gironde, France, 44° N 3230' 0° W 47') on a semi-humid podzolic soil. Spacing was 4 m between rows and 1.1 33m between individual trees, i.e. 2272 trees / ha. The trees were felled in March 1997 34and stem discs were cut, dried in a greenhouse and analysed for  $\delta^{13}$ C and ring width. 35From all the analysed families, 16 trees with no visible growth in the last four years 36before harvest were removed from the analysis.

# 37Ring width measurements

38Wood subsamples were taken from four positions on the circumference of the stem 39discs for the last four years of growth (1993 to 1996). As the trees were cut in march 401997, this includes any growth during winter 96/97 utilising reserve material from 41the summer 1996 growth period. Two different methods were used for ring width 42measurements. For the half diallel experiment, ring width was measured using the 43indirect X ray-method first described by Polge (1966). For the full-sib family, the 44ring widths were measured at the four sampling points on the circumference using a 1semi-automatic system consisting of a digitising tablet linked to a computer 2(precision 0.1 mm standard deviation). The width was averaged for each ring. To 3make the ring width data comparable to the  $\delta^{13}$ C measurements, for each tree the 4mean growth was calculated for the years 1993 to 1996 (mean ring width: MRW), 5using an arithmetic mean.

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# 7Isotope measurements

8The  $\delta^{13}$ C was measured of a bloc of four rings, which represents a ring-width 9weighted mean of the  $\delta^{13}$ C of each ring. The sampled blocks of wood were cut by 10hand into small pieces, pre-ground in a centrifugal mill (Tecator, Cyclotech 1093 11Sample Mill, Höganäs, Sweden) and milled to a fine powder in a ball mill (Retsch, 12MM2000, Haan, Germany). Cellulose was extracted after an acidic acid /nitric acid 13procedure described in Brendel, Iannetta & Stewart (2000). In brief, the method uses 14a concentrated nitric acid / 80% acetic acid on-to-ten dilution (0.2 cm<sup>3</sup> in 2 cm<sup>3</sup>) to 15digest lignin, proteins and hemicelluloses in 50 mg of powdered wood sample. The 16digested molecule fragments are then washed out using ethanol, remainders of acid 17are removed during a water wash. The samples are dried chemically with a pure 18ethanol / acetone progression and physically in a vacuum centrifugal evaporator 19(speed vac) at 100 hPa for 2 h. The original protocol (Brendel et al. 2000) was 20modified to include two extraction cycles, a 0.5 molar NaOH wash replacing the 21water wash to remove acids more thoroughly and prolonging the ethanol washes to 5 22min at 60 °C. For  $\delta^{13}$ C analysis, 1 mg cellulose subsamples were combusted and 23analysed for <sup>13</sup>C composition using a continuous flow isotope ratio mass 24spectrometer (Delta S, Finnigan MAT, Bremen, Germany). Carbon isotope 25composition was calculated relative to the Pee Dee Belemnite standard as (Craig 261957):

$$27\,\delta^{13}C = \frac{R_{sa} - R_{sd}}{R_{sd}} \times 1000[\%]$$
(1),

28where  $R_{sa}$  and  $R_{sd}$  are the <sup>13</sup>C/<sup>12</sup>C ratios of the sample and the standard, respectively. 29The discrimination between the  $\delta^{13}$ C of atmospheric CO<sub>2</sub> ( $\delta_{air} \approx -8\%$ ) and the  $\delta^{13}$ C of 30plant material ( $\delta_{plant}$ ) was calculated as (Farquhar *et al.* 1984) :

$$31^{\Delta} = \frac{\delta_{air} - \delta_{plant}}{1 + \delta_{plant} / 1000} \quad (2).$$

32Intrinsic WUE was estimated from discrimination using a modified equation from 33(Farquhar *et al.* 1982) :

$$34WUE = \frac{A}{g} = \frac{c_a}{1.6} \left(\frac{b-\Delta}{b-a}\right) \left[\frac{\text{mol CO}_2}{\text{mol H}_2\text{O}}\right] \quad (3),$$

1where  $c_a$  is the atmospheric CO<sub>2</sub> concentration (estimated as 360·10<sup>-6</sup> mol mol<sup>-1</sup>), *b* is 2the net fractionation caused by carboxylation (27‰) and  $\Delta$  is the discrimination 3between the  $\delta^{13}$ C of atmospheric CO<sub>2</sub> and the  $\delta^{13}$ C of cellulose (Equation 2).

#### 4

# 5Estimation of genetic parameters

6The normality of the distribution of the traits for both experimental set-ups was 7tested using Smirnov-Kolmogorov test. While  $\delta^{13}$ C was normally distributed, a small 8distortion from the normality was observed for ring width (p-value = 0.01). 9However, this distortion was considered to be too small to necessitate an adjustment.

10Analyses of variance for block and family effects in the half diallel were carried out 11with the OPEP software (Baradat 1989; Baradat & Labbé 1995) according to the 12following model derived from the "Henderson III" model (Searle 1971):  $13Y_{ijk} = \mu + B_i + F_j + \varepsilon_{ijk}$  (4)

14where  $Y_{ijk}$  is the value of the trait for the individual k belonging to the family j, 15located in the block i,  $B_i$  is the fixed effect of the ith block,  $F_j$  is the random effect of 16the jth family and  $\varepsilon_{ijk}$  is the random residual comprising : individual deviation from 17family mean and family x block interactions. When block and family effects were 18significant, data were adjusted to the block effect, prior to the decomposition of 19family effect. The half diallel analysis was carried out with OPEP using the model 20presented below, it is derived from the simplification of the random diallel model 21described by Garretsen & Keuls (1977) (Baradat & Desprez-Lousteau 1997) which is 22adapted to non orthogonal trials with reciprocal crosses :

$$23Y_{ijk} = \mu + a_i + a_j + s_{ij} + \varepsilon_{ijk}$$

$$(5),$$

24where Y<sub>ijk</sub> is the value of the trait for the individual k corresponding to the cross 25between the male i and the female j, ai (aj) is the general combining ability (GCA) of 26the  $i^{th}$  ( $j^{th}$ ) parent,  $s_{ij}$  is the specific combining ability (SCA) of the cross between the 27ith and the jth parent and  $\varepsilon_{ijk}$  is the residual term. The additive and dominance 28 variances are:  $\sigma_A^2 = 4\sigma_a^2$  and  $\sigma_D^2 = 4\sigma_s^2$ , whereas the phenotypic variance is :  $29\sigma^2_P = \sigma^2(Y_{ijk}) = 2\sigma^2_a + \sigma^2_s + \sigma^2_\epsilon$ . The narrow and broad sense heritabilities were 30calculated as  $h_{ns}^2 = \sigma_A^2 / \sigma_P^2$  and  $h_{bs}^2 = (\sigma_A^2 + \sigma_D^2) / \sigma_P^2$ , respectively. The percentage 31of additivity is calculated as the additive variance divided by the sum of additive plus 32dominance variances:  $\sigma_A^2 / (\sigma_A^2 + \sigma_D^2)$ . Genetic and environmental correlations were 33computed with OPEP using a multi-trait analysis of variance and covariance: (1) 34"estimated" genetic and environmental correlations were calculated according to the 35additive and dominance effects assessed in the random model (Equation 5; parental 36level), and (2) "predicted" correlations were assessed from the individual breeding 37values of each tree (individual level; figures 2 a-c). However, as the estimated and 38the predicted correlations gave similar results, only the correlation coefficients based 39on the prevalent "estimated" results were used in the discussion. Standard errors of 40 estimates of heritabilities were computed using the robust Jackknife method (Lebart 41et al. 1979).

# 1QTL detection

1

2Two genetic maps corresponding to the female and male parents of the full-sib 3family were established using AFLP markers, genotyped on a subset of 90 F1. The 4whole mapping population was further genotyped with evenly spaced markers to 5increase the statistical power of QTL detection (Chagné *et al.* submitted). In order to 6reduce the intra-trial environmental background noise, the data were adjusted for the 7block effect. We used the two-way pseudo-test cross mapping strategy to construct 8the linkage map (Grattapaglia & Sederoff 1994). Twelve linkage groups were found 9for the female map, equalling the number of chromosomes for *Pinus pinaster*. For 10the male map 15 linkage groups were detected, however the combination of the male 11and the female maps into a consensus map using  $\frac{3}{4}/\frac{1}{4}$ -segregating markers yields 12 12linkage groups for each parent. For QTL analysis however, only the  $\frac{1}{2}/\frac{1}{2}$ -segregating 13markers could be used.

14For QTL analysis, MultiQTL software (A. Korol, http://www.multiqtl.com) was used. 15In a first step, QTLs were detected by interval mapping using a LOD threshold of 1.5 16and a one-QTL-model (one QTL per linkage group). In a second step, these QTLs 17were taken as co-factors (composite interval mapping, CIM; introduced by Jansen & 18Stam 1994 and Zeng 1994), allowing individual QTL to be detected independently to 19the background noise. In a third step, a two QTL model (Korol *et al.* 1998) using 20CIM was applied, first testing if two QTLs are significant and then testing if two 21QTLs are more significant than one QTL. Standard deviations for the positions of the 22QTLs were calculated using a bootstrap method.

23As there are difficulties involved when using asymptotic approximations of LOD 24statistics(fixed LOD level) for QTL detection (Doerge & Churchill 1996), a 25permutation approach was used to determine appropriate significance thresholds. 26Two theoretical critical thresholds were considered, the first corresponding to a per 27linkage group type I error of 5% allowing the detection of "suggestive" QTL and the 28second corresponding to a genome wise type I error of 5% allowing the detection of 29"significant" QTL. Theoretical critical threshold corresponding to a genome wise 30type I error of 5% were calculated for each chromosome taking into account the 31number of markers in each chromosome. If  $\alpha_m$  is the critical threshold at the marker 32 level corresponding to a 5% genome wise type I error, the  $\alpha_c$  (critical threshold at the 33chromosome level) for a chromosome comprising n markers would be:  $34\alpha_c = 1 - (1 - \alpha_m)^n$ . These theoretical thresholds were compared to the thresholds 35associated with the LOD obtained by CIM at the chromosome level after 1000 36permutations of the data. The proportion of phenotypic variance explained by each 37QTL was estimated using the coefficient of determination (R<sup>2</sup>, estimated by CIM, 381000 permutations), which is based on the partial correlation of a putative QTL with 39the trait adjusted for cofactors in the multi-locus model.

# 1**Results**

# 2Trait distributions

3Means, ranges and variabilities for  $\delta^{13}$ C were very similar between the half diallel 4and the full-sib experimental designs (Table 1, Figure 1). For MRW (mean ring 5width) growth was higher in the half diallel by 0.66 mm and also the range of 6observed values was larger (Table 1). However, the coefficient of variation was 7slightly higher for the full-sib experimental design than for the half diallel. This was 8also true when the coefficients of variation for MRW were calculated using the block 9effect adjusted data (data not shown).

# 10Half diallel

11Analysis of variance including family and block effect (Equation 4) indicated 12variation among families for both  $\delta^{13}$ C and MRW, which justified the decomposition 13of the family effect according to Equation 5. Taking into account the significant 14block effect for  $\delta^{13}$ C, data were adjusted prior to the genetic decomposition. The 15narrow sense heritabilities were highly significant (p < 0.005) for  $\delta^{13}$ C and for MRW 16and close in their values (Table 2). No dominance effects were detected for these two 17traits, thus narrow sense and broad sense heritabilities are equal and additivities are 18100%.

19The phenotypic correlation between mean ring width and  $\delta^{13}$ C associated faster 20growth with less negative  $\delta^{13}$ C values (higher WUE) and was significant with a 21coefficient of correlation of r = 0.45 (Table 3 and Figure 2a). The correlation was not 22significant (Table 3) for the genetic component (additive effect), whereas the 23environmental component was highly significant with a strong correlation coefficient 24(r = 0.52).

# 25Full-sib family

26A significant positive phenotypic correlation between MRW and  $\delta^{13}$ C of the full-sib 27 family (r = 0.39; p < 0.005; Figure 3) was observed. For  $\delta^{13}$ C, eight OTLs were 28 found on seven linkage groups (chromosomes) and for MRW six QTLs on four 29linkage groups (Table 4). Using the one-QTL model, six QTLs were detected for  $30\delta^{13}$ C and two for MRW. With the two-QTL model, one pair of QTLs was found for  $31\delta^{13}$ C and two pairs for MRW. For  $\delta^{13}$ C, two of the QTLs detected with the one-QTL 32model and the QTL-pair detected with the two-QTL model and for MRW one QTL-33pair are "significant QTLs" at a probability corresponding to a 5% genome wise type 34I error. All other detected QTLs are "suggestive QTLs" at a probability 35corresponding to a 5% chromosome type I error. For  $\delta^{13}$ C, QTLs were detected on 36the male and the female maps, however not on the same chromosomes. A multi-locus 37model, including the male and female maps, explained 51.4% of the phenotypic 38variation of  $\delta^{13}$ C, the major QTL at chromosome 6 alone explaining 12.4%. For 39MRW, no QTLs were found on the female map and a multi-locus model for the male 40map explained 42.9% of the observed phenotypic variation. No co-localisation for a 41QTL of  $\delta^{13}$ C and of MRW was found.

# 1 Discussion

# 2Trait distributions

3Standard deviations and ranges were similar for half diallel as well as for the full-sib 4(Table 1). For  $\delta^{13}$ C the coefficients of variation were nearly the same, whereas the 5coefficient of variation for mean ring width was higher for the full-sib than for the 6half diallel (Table 1). Variability of traits might be expected to be higher in a half 7diallel with twelve parental trees, than in one full-sib family. However, it has to be 8taken into account that parental trees for both experiments were selected from the 9same provenance (Landes, Gascogne) and therefore might be genetically close. 10Further, with a polygenic complex trait such as  $\delta^{13}$ C, due to transgression, even 11parents with only a small difference in a measured trait can produce offspring with 12 extreme values (Prioul *et al.* 1997). For  $\delta^{13}$ C, the average of family variation of the 13half diallel (0.55‰ standard deviation within a range of 0.0‰ to 1.0‰ standard 14deviations) was similar to the variation found for the 186 trees of the full-sib family 15(0.63‰ standard deviation). Similarly, for MRW, the average of standard deviations 16 within half-diallel families was 0.94 mm, close to the 0.99 mm standard deviation 17 found for the full-sib family. Using Equations 2 and 3, the measured  $\delta^{13}$ C values 18transformed into a range of WUE of 67 µmol CO<sub>2</sub> / mol H<sub>2</sub>O to 100 µmol / mol for 19the half-diallel and a range of 65  $\mu$ mol / mol to 95  $\mu$ mol / mol for the full-sib. This 20 represents for the half diallel and the full-sib a variation from one to one-and-a half 21times the WUE.

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# 23Heritabilities and quantitative trait dissection analysis

24The heritabilities for mean ring width and  $\delta^{13}$ C were found to be significant, similar 25between the two traits and of rather moderate value. Therefore selective crossings 26can improve growth and WUE. The heritabilities for ring width are comparable to 27values found in the literature for maritime pine or other conifers. Danjon (1994) 28found for maritime pine trees from the same provenance as used in the present study 29narrow sense heritabilities for diameter growth ranging from 0 to 0.45 for different 30experimental set-ups (40 to 100 half or full-sib families). Blada (1999) found for a 31*Pinus cembra* L. 10x10 full-diallel narrow sense heritabilities for diameter from 0.23 32to 0.32 and broad sense heritabilities from 0.50 to 0.59.

33For  $\delta^{13}$ C there are not yet any publications known to the authors that estimated 34heritability for maritime pine, and there are only a few publications of estimates of 35heritability for  $\delta^{13}$ C for other species. Narrow sense heritability estimates by Johnsen 36*et al.* 1999 for *Picea mariana* are lower for diameter growth (0.14) than for  $\delta^{13}$ C 37(0.54). For non-woody species, heritabilities for  $\delta^{13}$ C can be high (broad sense 38heritabilities for *Lens culinaris* Medikus 0.73, Matus *et al.* 1995 and for *Agropyron* 39*desertorum* (Fischer ex Link) Schultes 0.90, Asay *et al.* 1998), however it was shown 40that water stress could reduce the heritability of  $\delta^{13}$ C (Ehdaie & Waines 1994, 41Johnson *et al.* 1990). An explanation for the moderate heritabilities found in the 42present study could therefore be the integrative properties of  $\delta^{13}$ C measured on 43cellulose of several rings, together with the possibility of frequent water stress. The 44present study was located in the south-west of France, where summer drought is 1common (Nguyen-Queyrens *et al.* 1998). The half diallel was created from the 2descendants of trees selected for growth vigour. This might have restricted the 3genetic base compared to natural populations and hence lowered the detectable 4heritability of growth.

5Existing QTLs for maritime pine were localized for traits related to growth (Plomion, 6Durel & O'Malley 1996, Gerber, Lascoux & Kremer 1997). We were able to provide 7here the first example of QTL observations for  $\delta^{13}$ C in a forest tree species. The four 8significant QTLs found for  $\delta^{13}$ C explained nearly one-third of the phenotypic 9variation observed for this trait. Several experiments (Prioul *et al.* 1997) have shown 10that even for complex traits, such as growth or carbon isotope discrimination, the 11expected number of major loci is quite small, a small number of genetic factors is 12predominantly determining a quantitative trait. No co-localisations of QTLs for  $\delta^{13}$ C 13and QTLs for MRW were observed, suggesting no common genetic control for these 14two traits. However, underestimation of number of QTL is inherent to the 15methodology of QTL detection.

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# 17Relationships between $\delta^{13}$ C and MRW

18The phenotypic correlations between  $\delta^{13}$ C and growth (ring width) found for the half 19diallel and full-sib experimental designs are significant with moderate coefficients of 20correlation (Figure 2a, Figure 3, Table 3) and the estimated regressions are similar in 21slope and intercept. Among trees in the same environmental conditions, this suggests 22that an increased growth relates to a higher WUE. Depending on a plants' 23physiology, a difference in WUE could be predominantly determined by stomatal 24conductance and/or by assimilation rate. The Farquhar model of carbon isotope 25discrimination (Farquhar, Ehleringer & Hubick 1989) predicts that an increasing 26photosynthetic capacity will decrease  $\Delta$ . Positive as well as negative correlations 27have been found between photosynthesis and growth (Johnsen et al. 1995), however 28when assuming a positive correlation between photosynthetic capacity and growth, a 29positive correlation between  $\delta^{13}$ C and growth could suggest a predominantly 30 assimilation rate based control of  $\delta^{13}$ C. This is in agreement with results for black 31spruce: differences among families were found to be mainly determined by 32differences in photosynthesis (Johnsen et al. 1995), whereby differences in 33photosynthesis were rather the result of non-stomatal limitations than of stomatal 34limitation (Major *et al.* 1996). If  $\delta^{13}$ C would be controlled by stomatal conductance, 35the Farquhar model predicts a negative correlation between  $\delta^{13}$ C and growth. 36Therefore, the positive correlation between  $\delta^{13}$ C and growth suggests that the 37variation of WUE among the measured trees is rather controlled by assimilation than 38by stomatal conductance. This was the case for the half diallel as well as for the full-39sib family.

40The calculated genetic and environmental correlations between  $\delta^{13}$ C and MRW 41indicate that the phenotypic correlation is mainly based on environmental influence. 42This result is in agreement with the lack of colocalisations between QTLs for  $\delta^{13}$ C 43and for MRW. The bloc effect was included in the estimation of the genetic 44parameters, hence the environmental correlation is probably due to micro-45environmental influences on each individual tree. As the model also accounts for any 1type of genetic effect, including intra-family genetic variation, the observed large 2environmental variation therefore suggests for the two measured traits a high 3sensitivity to micro-environmental conditions. This also suggests for growth and 4water use efficiency a high non-genetic placticity to adjust to environmental 5conditions. The strong environmental correlation that was found for the two traits is 6therefore probably due to a substantial environmental influence of less negative  $\delta^{13}$ C 7with increased growth and *vice versa*.

8The non-significant genetic correlation in the presented study is in opposition to the 9strong genetic correlation between  $\delta^{13}$ C and tree growth (height and diameter) found 10for black spruce (Johnsen et al. 1999). Several factors might have contributed to this 11discrepancy. 1) Johnsen et al. (1999) suggested assimilation rate as common control 12 for  $\delta^{13}$ C and growth. However, correlations between assimilation rate and growth 13 found in the literature range from negative over non significant to positive 14 relationships (Johnsen *et al.* 1995). Therefore, even if  $\delta^{13}$ C is determined by 15assimilation rate, if growth is not determined by assimilation rate, then there might 16be no correlation between  $\delta^{13}$ C and growth. 2) Furthermore, genetic control was 17rather moderate for both traits, which might have lowered the significance of a 18genetic correlation. 3) It has also to be taken into account that in the present study 19carbon isotope discrimination measured on cellulose of main stem wood was 20compared to the average diameter growth of four years, whereas Johnsen et al. 21(1999) compared  $\delta^{13}$ C of needle material with height or trunk diameter. These 22complex traits might include the action of a number of slightly different genes and a 23common genetic control might exist for the combination of needle  $\delta^{13}$ C to height or 24diameter and not for the combination of trunk cellulose to mean annual growth of the 25same growth period.

26The lack of a genetic correlation between  $\delta$ 13C and MRW found in the half-diallel is 27in agreement with the lack of co-localizations between QTLs for  $\delta$ 13C and for MRW 28found in the full-sib experimental design. Albeit the lack of a genetic correlation, 29both traits were found to be heritable and significant QTLs were detected. This opens 30new perspectives for the investigation of the genetic determinism of water use 31efficiency and the identification of groups of genes involved in drought responses.

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# 1Tables

2Table 1 Means, standard deviations (SD), coefficients of variation (SD / mean), 3ranges (maximum – minimum) and probability (p-value in %; \* significant at 5% 4level; ns not significant) of block and family effects (Equation 4) of the two 5experimental designs for  $\delta^{13}$ C (in [%]) and mean ring width (MRW: in [mm])

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bexperimental designs for 0 °C (in [700]) and mean ring what (interv), in [inin])								
		Ν	Mean	SD	Coef. of	Range	Block	Family
					Var.		effect	effect
$\delta^{13}C$	Half diallel	564	-26.21	0.61	0.02	3.64	2.8*	2.3*
	Full-sib	186	-26.48	0.63	0.02	3.23	0.0*	
MRW	Half diallel	564	2.88	1.10	0.38	6.77	ns	0.2*
	Full-sib	186	2.11	0.99	0.47	5.15	0.9*	
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7Table 2 Half diallel: Heritabilities for  $\delta^{13}$ C and mean ring width (MRW) with 8standard deviations and 95% confidence interval in parentheses. As there were no 9dominance effects detected and therefore additivity is 100%, the narrow sense (ns) 10and broad sense (bs) heritabilities are equal.

	$h^2_{ns} = h^2_{bs}$	%additivity
δ <sup>13</sup> C [‰]	$0.17 \pm 0.06 \ (0.06 - 0.29)^*$	100%
MRW [mm]	$0.19 \pm 0.06 \ (0.06 - 0.31)^*$	100%
11* significant at the 59	% level	

13Table 3 Half diallel: Correlations between mean ring width and  $\delta^{13}$ C; r is the 14correlation coefficient on the family level, estimated by OPEP software with 15standard deviation and 95% confidence interval in parentheses; r' is the correlation 16coefficient estimated by linear regression analysis using the calculated individual tree 17breeding values which are shown in Figure 2.

	r	r'					
Phenotypic correlation	$0.45 \pm 0.057 \ (0.33 - 0.56)^*$	0.46*					
Genetic correlation	$0.27 \pm 0.21 \ (0.07 - 0.15) \ \text{NS}$	0.02 NS					
Environmental correlation	$0.52 \pm 0.16 (0.20 - 0.83)^*$	0.79*					

18\* significant at the 5% level; NS : not significant

1Table 4 Full sib: Results of the composite interval mapping analysis for  $\delta^{13}$ C and 2mean ring width (MRW) using MultiQTL software. The p-value associated with the 3LODs were calculated using 1000 permutations of the data, standard deviation of 4position (SD) was calculated using the bootstrap method (1000 permutations); in 5case of a significant QTL-pair for a chromosome, Lod and p-value are given for the 6tests of (I) two QTLs *versus* no QTLs and (II) two QTLs *versus* one QTL (difference 7of Lod for two and for one QTL).

											- 0	
	map	Chr <sup>a</sup>	N°	P	osition ± SD <sup>c</sup>		Lod	P-value <sup>°</sup>	Dir. <sup>e</sup>	R <sup>2</sup>	R <sup>2</sup> total	
δ <sup>13</sup> C	male	3a	149		4.3±13.2		1.78	0.021*	+	0.047		
	male	6	164		102.7±21.6		4.40	0.001***	+	0.124	0.260	
	male	8	85		0.0±15.7		1.85	0.021*	-	0.050	0.200	
	male	9	183		104.6±28.5		1.90	0.033*	-	0.047		
	female	2	84		209.1±56.4	[	2.30	0.019*	+	0.065		
	female	5	164		99.9±22.0		1.98	0.0033***	-	0.062	0.246	
	female	12	153	1:	0.0±36.7	1:	4.24	I: 0.002***	-	0.119	0.240	
				2:	135.4±27.1	II:	1.88	II: 0.036*	+			
MRW	male	2b	180	1:	47.2±15.9	1:	2.49	I:0.048*	-	0.181		
			153	2:	51.8±11.1	II:	1.57	II: 0.027*	+			
	male	5	145		56.8±33.4		2.41	0.022*	-	0.065	0 400	
	male	6	174	1:	7.5±31.0	1:	4.24	I : 0.001***	-	0.124	0.429	
			168	2:	133.7±22.0	II:	2.68	II: 0.002*	+			
	male	11	165		11.6±30.9		1.73	0.040*	+	0.059		
						-						

8ª chromosome ID

9<sup>b</sup> number of full-sibs with available data for QTL detection

10° Lod score peak position (from the top) of the chromosome (cM)

 $11^{\rm d}$  Probability for the null hypothesis of no QTL at the chromosome level

12 \*\*\* : probability corresponding to a 5% genome wise type I error (significant QTL)

13 \* : probability corresponding to a 5% chromosome type I error (suggestive QTL)

14<sup>e</sup> direction of influence of presence of the allele for each QTL

# 1Figure legends

**2Figure 1** Distribution of cellulose  $\delta^{13}$ C (raw data; not corrected for block effect) for 3a) the half diallel experiment and b) the full-sib experiment; parameters for the 4normal distributions as in Table 1

**Figure 2** Linear correlations between mean ring width (MRW) and  $\delta^{13}$ C for the half 6diallel experiment using data corrected with the individual tree breeding values 7(OPEP software): a) phenotypic, b) genetic (additive effects) and c) environmental 8correlations; all data are adjusted for block effect, data for genetic and environmental 9correlations are centred and standardized by the mean.

**Figure 3** Phenotypic linear correlation between mean ring width (MRW) and  $\delta^{13}$ C 11for the full-sib experiment; data adjusted for block effect





# 1Figure 3

