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Effects of soil moisture and treatment volume on bentazon mobility in soil

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Abstract – Soil moisture affects the leaching behaviour of pesticides by inducing their physical entrapment in the soil structure. Columns containing soil aggregates were dampened to specific initial moisture levels. Bentazon was dripped onto surface aggregates in different volumes. The columns were then percolated after an equilibration period. Soil water from the columns was divided arbitrarily among mobile and immobile regions in order to describe the herbicide redistribution processes in the soil. When the soil was dry before treatment, bentazon losses by mass flow were 1.5 to 4 times higher than in wet conditions. Between application time and percolation, any water present in the porous matrix might favour pesticide diffusion towards immobile water regions as well as adsorption into and onto soil aggregates, preventing its leaching. The use of large solution volumes of the pesticide modifies surface soil moisture, suppressing any difference in behaviour between dry and wet soil application conditions.

bentazon / diffusion / leaching / mobile water / immobile water

1. INTRODUCTION

Summer pesticide application might be done in dry or wet soil conditions. According to the soil’s moisture state, pesticide spread over a given land surface might either crystallise on contact with dry soil or dissolve on wet soil. After periods of low or no rainfall, the pesticide would not be able to evolve or to move in the soil surface layer, whereas in wet conditions, the pesticide could interact with soil particles. Thus, the weather conditions preceding pesticide application may considerably affect the behaviour of the product in the soil.

Several authors have studied soil moisture effect at the time of treatment on solute movement in soil. Rao et al. (1974) mentioned, in field experiments, later and lesser picloram leaching when applied on air-dry soil (evaporation) compared with wet soil. The opposite was observed when studying mobile herbicides’ (isoproturon and mecrocrop) transfer through a loamy clay soil (Brown et al., 1995a). In a soil column experiment, Shipitalo et al. (1990) pointed out the effect on solute (bromide, strontium and atrazine) availability to transfer of slight precipitation occurring prior to heavy rainfall. Laboratory studies have also displayed the variability of pesticide movement depending on the humidity level of the soil (Geissbuhler et al., 1963; Green et al., 1968). A recent work (Beulke et al., 2004) concluded that variations in moisture level had no effect on isoproturon leaching and availability in soil water. These studies were all based on soil columns that had been dampened to a greater or lesser extent at the time of applying the pesticide and brought to field capacity before leaching.

Water resource management requires an understanding of the mechanisms and original conditions responsible for pesticide pollution. In order to comprehend solute movement in soils, Van Genuchten and Wierenga (1976) conceptualised soil as a porous medium composed of two distinct but connected continua. Liquid pore volume is divided into two regions: a region of mobile water occupying the coarse pores of soil (macropores); and an immobile region, referring to intra-aggregate porous space (meso-, micro- and nanopores). Solutes move by convective flow through the mobile region (White et al., 1986; Hagerman et al., 1989; Brown et al., 1995b), whereas intra-aggregate space is accessible mainly by molecular diffusion (Nye, 1979; Koch and Flühler, 1993) but also by convection (Jury and Flühler, 1992). The extent and redistribution of pesticides by water are controlled both by soil and compound characteristics and by environmental conditions, among which climatic conditions predominate (Yaron et al., 1996).

This work aims to determine the effects of soil initial moisture and the mode of pesticide application on product redistribution in mobile and immobile regions. Small columns filled with soil aggregates were saturated with water to distinct moisture levels and treated with bentazon. We varied the formulation and

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application volumes of the bentazon solution. Bentazon is a herbicide, used for cereal protection, chosen in this study as a model because of its high solubility in water (Abernathy and Wax, 1973; Huber and Otto, 1994) and its weak adsorption on soil constituents (Grey et al., 1996; Boivin et al., 2004). By successive stages of percolation and centrifugations, we analysed the amounts of bentazon transferred by mass flow and the concentration spread in the immobile water compartments. The experiment was carried out in six soils with contrasting physico-chemical properties.

2. MATERIALS AND METHODS

2.1. Soil columns

For this study, five agricultural soils were selected in the Lorraine region of Northeastern France and one in Brittany (Western France), each representing a major textural class (clay, loam or sand) and a high or low organic matter level (Tab. I). Soil retention capacity (RC) was determined for each soil (Tab. I) by the pressure method, whereby RC corresponds to the maximal amount of capillary water retained by a soil (Duchauzour, 1970).

After being air-dried, soil samples (surface layer 5–15 cm) were sieved between 2 and 5 mm so that their structure resembled that of an aggregated seedbed. Small soil columns were made by filling Whatman Autovials® (diameter 2 cm × 6.5 cm height, Fig. 1) with dry soil aggregates. Aggregate columns were either left to dry or dampened to 50% or 100% of soil retention capacity. When the pesticide was applied in a large volume of solution (cf. Sect. 2.2.2), the theoretical water volume required to dampen the soil to 100% of RC was adapted.

2.2. Pesticide

2.2.1. Pesticide characteristics

Bentazon (3-isopropyl-1H-2,1,3-benzothiadiazine-4(3H)one-2,2-dioxide) is a weak acid (pKa 3.3) selected for its high water solubility (570 mg·L⁻¹ at pH 7, 22 °C) and its weak adsorption on soil (mean Kd value 0.6 cm³·g⁻¹) (Huber and Otto, 1994). Specific bentazon adsorptions by the 6 soils studied were previously measured using standard equilibration techniques (Boivin, 2003). Their Kd values are reported in Table II.

2.2.2. Pesticide application

The wet columns were left to equilibrate for a 16 h period and then [phenyl-U-¹⁴C] bentazon (radiochemical purity >99%; specific activity 6.69 MBq·mg⁻¹, BASF, Germany) was added (0.3–0.4 µg according to the daughter solution preparation) to each soil column. In a first experiment concerning the six soils, bentazon was applied to the top aggregates in 0.02 mL.

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**Table I.** Selected top soil properties of the six soils used in the study.

<table>
<thead>
<tr>
<th>Soil series</th>
<th>Classification¹</th>
<th>Textural class(%)</th>
<th>pH_water</th>
<th>Organic Matter (%)</th>
<th>RC²</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Clay</td>
<td>Silt</td>
<td>Sand</td>
<td></td>
</tr>
<tr>
<td>Bouzule clay</td>
<td>vertic stagnic cambisol</td>
<td>41.1</td>
<td>48.6</td>
<td>10.3</td>
<td>6.9</td>
</tr>
<tr>
<td>Villey clay</td>
<td>stagnic cambisol</td>
<td>53.6</td>
<td>39.7</td>
<td>6.7</td>
<td>6.7</td>
</tr>
<tr>
<td>Bouzule loam</td>
<td>stagnic luvisol</td>
<td>30.9</td>
<td>50</td>
<td>19.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Kerlavic loam sand</td>
<td>dystic cambisol</td>
<td>17.7</td>
<td>45.2</td>
<td>37.1</td>
<td>5.3</td>
</tr>
<tr>
<td>Villey sand</td>
<td>fluvic stagnic cambisol</td>
<td>11.4</td>
<td>22.5</td>
<td>66.1</td>
<td>5.8</td>
</tr>
<tr>
<td>Corcieux sand</td>
<td>eutric cambisol</td>
<td>10.4</td>
<td>19</td>
<td>70.6</td>
<td>6.2</td>
</tr>
</tbody>
</table>


**Figure 1.** Experimental column (Whatman Autovial®) filled with dry soil aggregates: an autosampler vial can be attached to the tip.

**Table II.** Bentazon adsorption characteristics by the 6 soils studied (Boivin, 2003).

<table>
<thead>
<tr>
<th>Soil series</th>
<th>Kd</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bouzule clay</td>
<td>1.2</td>
</tr>
<tr>
<td>Villey clay</td>
<td>1.4</td>
</tr>
<tr>
<td>Bouzule loam</td>
<td>1.4</td>
</tr>
<tr>
<td>Kerlavic loam sand</td>
<td>1.9</td>
</tr>
<tr>
<td>Villey sand</td>
<td>1.4</td>
</tr>
<tr>
<td>Corcieux sand</td>
<td>1.3</td>
</tr>
</tbody>
</table>
Effects of soil moisture and treatment volume on bentazon mobility in soil

2.3. Experimental method

2.3.1. Leaching study

The treated columns were sealed with parafilm and stored for 6 days at room temperature (20 ± 2 °C). No water was added during this period. Subsequently, all the columns were eluted with distilled water using a peristaltic multi-channel pump (Ismatec model mp25) at an arbitrary flow rate of 1 mm·min⁻¹, fixed with the intention of facilitating the exchanges between mobile and immobile waters as well as processes of solubilisation and adsorption/desorption. We aimed to add a total of 16.5 mL – equivalent to an intense rainfall in spring or summer – to each column, when both the soil-dampening stage (concerning exclusively wet columns) and percolation stage were taken together. The dry columns were percolated with 16.5 mL. The columns initially dampened to 50% or 100% of RC were percolated with a volume equivalent to the difference between 16.5 mL and the volume of water already applied during the soil-dampening stage.

All the mobile water was collected and weighed at the end of the percolation. ¹⁴C-bentazon concentration was measured immediately after percolation by liquid scintillation (Packard Tri-Car 460 CD): 1 mL was counted in 10 mL Ultima Gold Scintillator-Packard. This aimed both at characterising the fraction of soil water and bentazon available by convective transport, through inter-aggregate space, during leaching events, and at quantifying bentazon transfer.

2.3.2. Immobile water content characterisation

The columns (Whatman® autovial) consisted of a barrel with a perforated base covered by a glass microfibre filter allowing a sampler vial to be attached (Fig. 1). After percolation, the columns were centrifuged (Beckman Avanti Centrifuge J25) successively at 333 g (5 min) and 7244 g (7 min). Previous experiments had shown that longer times of centrifugation did not allow any further water to be collected and were destructive for the experimental device. Each corresponding water fraction was collected in a sampler vial attached to the base tip of the column. Both fraction volumes were determined by weighing and the ¹⁴C-bentazon concentration was determined by liquid scintillation counting. The purpose of this was to estimate the water storage capacity of two different immobile water compartments and to characterise the bentazon concentration therein. Data (volume and herbicide content) concerning the residual water fraction (a third immobile region) remaining after centrifugations were inferred from weighing and calculation. In order to check bentazon residual amounts in the soil by measurement, combustion of air-dried and pulv-erised soil samples was performed in parallel after centrifugations, for several columns of each treatment. No significant differences appeared between calculated and measured data.

2.4. Statistical analysis

The effects of initial soil aggregate moisture and the application volume on bentazon amounts transferred into mobile water and the bentazon levels in immobile regions were tested through a variance analysis (ANOVA).

3. RESULTS AND DISCUSSION

3.1. Water compartments of aggregate columns

The total soil solution volumes recovered after the percolation and centrifugation stages are presented in Table III. No significant difference in volume was noted between dry and wet (50 or 100% of RC) columns. The proportion of the volume of the different water regions (percolated water, 333 g extracted water and 7244 g extracted water) in relation to the total water collected is also reported in Table III.

3.2. Bentazon mass flow transfer behaviour

3.2.1. Small volume application

Table IV presents the amounts of bentazon transferred by mass flow in mobile water, for the six soils studied, according to initial soil moisture (dry, dampened to 50% of RC, or dampened to 100% of RC).

When bentazon was applied to dry soil aggregates, bentazon amounts in mobile water accounted for 42.1% (Bouzule loamy soil) to 58.1% (Villey clay soil) of the initial dose. No statistical differences between soils were revealed by variance analysis (Tab. IV). Conversely, bentazon application on wet soil led to a significant decrease in amounts exported by macroporal flow for each soil: 1.5 to 4 times less herbicide was measured in percolates, with the unexplained exception of Villey sandy soil. Moreover, bentazon mass transfer remained unaltered by aggregate water saturation level (50 or 100% of RC).

3.2.2. Large volume application

Bentazon exportations by mass flow, according to soil initial moisture (dry or 100% of RC) when a 50 times greater (1 mL)
volume is applied of an aqueous bentazon solution are presented in Table V. For the three soils, percolated bentazon amounts ranged from 17.8% to 29.5%, when soil aggregates were dry previous to pesticide application, and from 17.9% to 26.1% when they were wet. It would seem that an increased treatment volume limited the initial soil moisture effect we observed previously. Indeed, for each of the three soils (Bouzule clay, Bouzule loam and Corcieux sand), the same herbicide amounts were recovered in mobile water, whatever the initial aggregate water saturation.

### Table III. Total soil solution recovered after leaching and centrifugations of dry soil columns (low volume of treatment) and proportion of the different water compartments (means of 4 replicates).

<table>
<thead>
<tr>
<th>Soil series</th>
<th>Total soil solution (mL)</th>
<th>Leachate (% of total)</th>
<th>SD</th>
<th>333 g water extracted (% of total)</th>
<th>SD</th>
<th>7244 g water extracted (% of total)</th>
<th>SD</th>
<th>Residual water (% of total)</th>
<th>SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bouzule clay</td>
<td>15.8</td>
<td>63.9</td>
<td>0.5</td>
<td>5.7</td>
<td>0.7</td>
<td>7.2</td>
<td>0.4</td>
<td>23.2</td>
<td>0.5</td>
</tr>
<tr>
<td>Villey clay</td>
<td>16.6</td>
<td>68.3</td>
<td>1.7</td>
<td>5.9</td>
<td>0.3</td>
<td>6.7</td>
<td>0.2</td>
<td>18.0</td>
<td>0.2</td>
</tr>
<tr>
<td>Bouzule loam</td>
<td>16.4</td>
<td>71.1</td>
<td>0.7</td>
<td>4.4</td>
<td>0.4</td>
<td>6.4</td>
<td>0.3</td>
<td>18.0</td>
<td>0.1</td>
</tr>
<tr>
<td>Kerlavic loam sand</td>
<td>16.0</td>
<td>75.6</td>
<td>2.4</td>
<td>3.5</td>
<td>0.6</td>
<td>8.1</td>
<td>0.4</td>
<td>12.8</td>
<td>0.7</td>
</tr>
<tr>
<td>Villey sand</td>
<td>16.3</td>
<td>82.6</td>
<td>1.5</td>
<td>3.2</td>
<td>0.2</td>
<td>5.6</td>
<td>0.4</td>
<td>8.5</td>
<td>0.3</td>
</tr>
<tr>
<td>Corcieux sand</td>
<td>16.2</td>
<td>76.5</td>
<td>10.0</td>
<td>1.8</td>
<td>1.0</td>
<td>7.6</td>
<td>0.4</td>
<td>14.1</td>
<td>0.5</td>
</tr>
</tbody>
</table>

Standard Deviation (SD) indicates error between the replicates.

### Table IV. Small volume of treatment: mean mobile water bentazon amounts per column after 6 days of equilibration.

<table>
<thead>
<tr>
<th>Soil series</th>
<th>dry</th>
<th>50% of RC</th>
<th>100% of RC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>leached amounts (% of initial dose)</td>
<td>SD</td>
<td>leached amounts (% of initial dose)</td>
</tr>
<tr>
<td>Bouzule clay</td>
<td>45.7a</td>
<td>0.7</td>
<td>33.4b</td>
</tr>
<tr>
<td>Villey clay</td>
<td>58.1a</td>
<td>14.3</td>
<td>22.5b</td>
</tr>
<tr>
<td>Bouzule loam</td>
<td>42.1a</td>
<td>2.8</td>
<td>11.2b</td>
</tr>
<tr>
<td>Kerlavic loam sand</td>
<td>49.9a</td>
<td>10.0</td>
<td>30.4b</td>
</tr>
<tr>
<td>Villey sand</td>
<td>53.9a</td>
<td>5.5</td>
<td>48.8a</td>
</tr>
<tr>
<td>Corcieux sand</td>
<td>52.3a</td>
<td>6.5</td>
<td>22.0b</td>
</tr>
</tbody>
</table>

Letters a, b and c refer to variance analysis of initial moisture effect on bentazon exportations by soil (the same letters indicate no significant differences between treatments). Standard Deviation (SD) indicates error between the replicates.

### Table V. Large volume of treatment: mean mobile water bentazon amounts per column after 6 days of equilibration.

<table>
<thead>
<tr>
<th>Soil series</th>
<th>dry</th>
<th>100% of RC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>leached amounts (% of initial dose)</td>
<td>SD</td>
</tr>
<tr>
<td>Bouzule clay</td>
<td>29.5</td>
<td>4.0</td>
</tr>
<tr>
<td>Bouzule loam</td>
<td>17.8</td>
<td>2.0</td>
</tr>
<tr>
<td>Corcieux sand</td>
<td>20.5</td>
<td>2.1</td>
</tr>
</tbody>
</table>

No statistical differences were revealed by the variance analysis of initial moisture effect on bentazon exportations by soil. Standard Deviation (SD) indicates error between the replicates.

### 3.3. Bentazon redistribution in immobile water compartments

#### 3.3.1. Small volume application

The bentazon distribution in the various water compartments of three soils selected from the panel (Bouzule clay, Bouzule loam and Corcieux sand), after percolation and centrifugations of treated columns by a weak volume on dry aggregates, is represented in Figure 2a. The product is divided into the distinct soil water regions (mobile and immobile regions). Mean bentazon concentrations for the three soils in the three immobile water regions, meso-, microporal and resid-ual, were, respectively, 18.2 ± 2.5 µg·L⁻¹, 23.2 ± 6.8 µg·L⁻¹ and 42.4 ± 10.2 µg·L⁻¹.

When treatment was carried out with a small volume on wet aggregates (Fig. 2b), the bentazon levels divided between the fine porosity compartments, for the same soils, were statistically lower than with initially dry aggregates: 7.3 ± 3 µg·L⁻¹ and 10.5 ± 5.8 µg·L⁻¹ in meso- and microporal waters. Because no effect of soil water saturation was revealed by variance analysis, only data relative to soil dampened to 100% of RC are presented in Figure 2b. It can be noticed that a majority of the herbicide remains in the residual water (48.5 ± 1.5 µg·L⁻¹, 89.5 ± 7.7 µg·L⁻¹ and 98.6 ± 7.6 µg·L⁻¹, respectively, for the Bouzule clay, the Bouzule loam and the Corcieux sandy soils).
3.3.2. Large volume application

Adding bentazon in greater volumes (1 mL), as practised in laboratory studies based on the experimental model, tended to reduce the initial moisture effect previously observed between dry and wet soil. Redistribution profiles proved to be similar for Bouzule loam and Corcieux soils, whatever their initial humidity level (dry or wet), as shown in Figure 3. In the case of Bouzule clay soil, treatment in wet conditions promoted bentazon redistribution in residual water to the detriment of meso- and microporal waters.

3.4. Significance

The soil solution in columns of aggregates was divided arbitrarily into several regions. A first region constituted temporary water filling, in water-saturated soil conditions, of the spaces between aggregates (macroporosity). This mobile water obeys the laws of gravity. The water extracted by centrifugation at 333 g filled medium-size pores, whereas water extracted at 7244 g filled the finest soil pores. Because they are strongly retained, water from these two compartments is considered to be immobile. Water remaining in the column after centrifugations would correspond to immobile water located within soil nanopores and/or water adsorbed on soil constituents. A pesticide introduced into the system would be likely to be split up into the different types of water by convection and/or diffusion processes.

The results of the study show that bentazon redistribution within soil porosity may be significantly affected by the pesticide application mode and by the initial soil humidity level at the time of pesticide treatment. When dissolved in a weak dichloromethane volume and dripped on dry aggregates, bentazon is quickly crystallised on aggregate surfaces, because of
the almost instantaneous volatilisation of the solvent. During the equilibration period, the immobilised product was unlikely to undergo either physical, chemical or biological phenomena because of the maintained dry state of the aggregates. The compound is then re-solubilised and transported through the porous network by water percolation. Capillary forces developed by micropores carry the water and bentazon away into fine porosity, even if the phenomenon is limited by the presence of air (Youngs et al., 1994). Bentazon is simultaneously transported by convection in macroporosity. Once inside the immobile water regions, exchanges of herbicide with mobile water are limited by water flow velocity; little time is left for diffusion to take place (Hartley, 1964). High concentrations may be diffused in capillary spaces. Bentazon in mobile water, during its spread in soil porosity, could be subject to a quick adsorption phenomenon (Boivin et al., 2004). Despite the short contact time, a part of the initial pesticide amount would be retained on external sorption sites of the soil particles. A direct transfer of pesticide in microcrystal form cannot be excluded. Amounts transferred by mass flow represent half of the applied dose. The “small volume application on dry soil” combination preserves a relative availability of the product to leaching and confers a direct transfer of pesticide on dry soil aggregates in a 50 times greater volume of aqueous solution used for treatment directly dampened the column’s surface aggregates (without previous solubilisation). As a result, a part of the product is carried away through the aggregates by the creation of locally immobile regions that will not participate in diffusive exchanges with mobile water. Transfer in mobile water may be thus considerably reduced. In a laboratory study, Shipitalo et al. (1990) also showed weaker atrazine leaching in the case of soil blocks receiving low rainfall prior to a significant leaching rainfall, than in the case of dry soil.

4. CONCLUSION

Initial soil moisture at the time of pesticide treatment influenced bentzon mass transfer behaviour when application was carried out according to field conditions. After 6 days of equilibration, according to the soil studied, previous soil dampening had the effect of reducing the amounts leached from 35 to 77%, compared with a treatment performed with a low volume on dry soil. In our experimental conditions, on wet soils, the decrease in bentazon availability to mass transfer was attributed mainly to product redistribution by diffusion towards immobile water spaces, where it is likely to be physically trapped. In such application conditions, pesticide leaching might also be reduced by its adsorption onto and into soil aggregates. On a field scale, such phenomena might be expected: the probability of a soluble pesticide being quickly transported by mass flow towards soil depth might be potentially lower when soil is wet at the time of treatment or when a low rainfall carries it away or favours its diffusion within aggregates. Moreover, this study allowed us to highlight the fact that pesticide application technique (volume) in a laboratory experiment was meaningful where transfer results are concerned. Adding a larger 1 mL volume had the effect of dividing bentazon within soil fine porosity, thereby reducing its availability to transfer and suppressing any difference in behaviour between dry and wet soil application conditions.
Acknowledgments: This work was financially supported by INRA and Arvalis.

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