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Effects of various inhibitors including carboxin on *Botrytis cinerea* mitochondria isolated from mycelium

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Summary — Due to the development of resistant strains, the practical control of *Botrytis cinerea* remains a problem. Little is known about its respiratory chain and this paper describes the preparation and the properties of *B. cinerea* mitochondria with the aim of testing new antifungal compounds. NADH, succinate and α -ketoglutarate as substrates provided good oxygen consumption. Respiratory control ratios were high and showed that electron transport was well coupled to oxidative phosphorylation. The phosphorylation efficiencies (ADP/O) obtained with all 3 substrates were compatible with theoretical values. The addition of cytochrome *c* did not greatly enhance the oxygen consumption due to ascorbate: the quality of the outer mitochondrial membrane was good enough to prevent externally added cytochrome *c* from interacting easily with the respiratory chain. These observations indicate that the mitochondria isolated sustain a high level of structural and functional integrity. This study showed also that they were devoid of the alternative pathway resistant to antimycin A and cyanide, but that they exhibited a rotenone-insensitive respiratory route. The action and the effective doses of oligomycin, carbonylcyanide *m*-chlorophenylhydrazone (CCCP) and carboxyatractyloside on *B. cinerea* mitochondria were similar to those observed with plant mitochondria. At 10 μ M, carboxin completely inhibited succinate oxidation in mitochondria from *B. cinerea* but failed to have any effect on NADH and α -ketoglutarate oxidation. This toxicity level was comparable to that reported by other authors with members of the fungal class Basidiomycetes.

respiration inhibitors / rotenone insensitive route / carboxin / alternative pathway / *B. cinerea*

Résumé — Effets d'inhibiteurs dont la carboxine sur les mitochondries de *Botrytis cinerea* isolées de mycélium. Du fait de l'apparition de phénomènes de résistance aux produits anti-*Botrytis*, la lutte contre la pourriture grise demeure un problème. La chaîne respiratoire de *B. cinerea*, dont on connaît peu de chose, pourrait constituer une cible intéressante pour découvrir des produits de remplacement. Cet article décrit une méthode simple pour préparer des mitochondries de *B. cinerea* et précise leurs caractéristiques. Le NADH, le succinate et l' α -cétoglutarate sont des substrats qui stimulent fortement la consommation d'oxygène. Les contrôles respiratoires sont élevés et prouvent que le transport des électrons est bien couplé à la phosphorylation oxydative. Les rapports ADP/O obtenus avec les 3 substrats sont compatibles avec les valeurs théoriques. L'addition de cytochrome *c* n'augmente pas notablement la consommation en oxygène provoquée par l'ascorbate : la qualité de la membrane externe des mitochondries est suffisante pour empêcher le cytochrome *c* de réagir sur la chaîne respiratoire. Ces observations montrent que les mitochondries isolées présentent une bonne intégrité tant sur le plan structural que fonctionnel. Cette étude montre aussi qu'elles ne possèdent pas une voie alternative résistante à l'antimycine A et au cyanure mais présentent une voie insensible à la roténone. Les effets et les niveaux d'activité de l'oligomycine, du carbonylcyanide *m*-chlorophénylhydrazone (CCCP) et du carboxyatractyloside sur les mitochondries de *B. cinerea* ont été similaires à ceux généralement observés avec les mitochondries végétales. A 10 μ mol.l⁻¹, la carboxine inhibe complètement l'oxydation du succinate par les mitochondries de *B. cinerea* mais n'a aucun effet sur les oxydations du NADH et de l' α -cétoglutarate. Ce niveau de toxicité est comparable à celui relevé par d'autres auteurs avec des champignons appartenant à la classe des Basidiomycètes.

inhibiteur de la respiration / voie résistante à la roténone / carboxine / voie alternative / *B. cinerea*

* Correspondence and reprints

Abbreviations: EDTA: ethylenediaminetetraacetic acid; BSA: bovine serum albumin; RCR: respiratory control ratio; ADP/O ratio: micromoles adenosine diphosphate/microatoms oxygen utilized; TPP: thiamine pyrophosphate; TMPD: *N, N, N', N'*-tetramethyl-*p*-phenylenediamine; TTFA: thenoyltrifluoroacetone; CCCP, carbonylcyanide *m*-chlorophenylhydrazone.

INTRODUCTION

Botrytis cinerea, an economically important phytopathogenic fungus, causes grey mould in various plants. Due to the development of resistant strains (Leroux and Clerjeau, 1985), its practical control remains a problem especially in vineyards. Although the morphological and biochemical characteristics of *B. cinerea* have been studied extensively, little is known about its respiratory chain. However, the respiratory chain of *B. cinerea* could be an interesting target for alternative fungicides. So, this paper describes the preparation and the properties of *B. cinerea* mitochondria. Then, the effects of carboxin, an antifungal carboxamide used for control of several Basidiomycetes is studied on these organelles.

METHODS

Growth

The *B. cinerea* strain was a wild-type strain sensitive to dicarboximide, aromatic hydrocarbon and benzimidazole fungicides (Fritz *et al*, 1977; Leroux and Fritz, 1984). Conidia were obtained from 15-d cultures grown on potato-dextrose agar slopes in test tubes. Mycelial colonies were prepared by seeding 8.10^5 conidia ml⁻¹ in the following media: 10 g glucose or 10 g starch; 2 g KH₂PO₄; 1.5 g K₂HPO₄; 0.5 g Mg SO₄, 7H₂O; 1 g (NH₄)₂SO₄; 2 g yeast extract and deionized water made up to 1 l, pH 6.2. Incubation took place at 23 °C on a rotary shaker at 150 rpm for 26 h.

Mitochondrial preparation

The young hyphae were harvested by filtration, washed twice with refrigerated distilled water and finally with the isolation medium which consisted of 0.5 mol.l⁻¹ sucrose, 5 mmol.l⁻¹ cysteine, 1 mmol.l⁻¹ EDTA, 0.3% (w/v) BSA, 0.1 mol.l⁻¹ sodium phosphate buffer, pH 7.4. All subsequent operations were performed in the cold (0–4 °C). The hyphae were suspended in a 3-fold volume of isolation medium and disrupted for 3 x 3 s with a Multi-Moulinette mixer (Moulinex, France). The suspension of disrupted hyphae was then filtered in < 3 s through 2 layers of non-woven cloth lining round the basket of an electric juice extractor (Moulinex, France). The mitochondria were collected from the filtrate by differential centrifugation, as a fraction sedimenting between 6 000 g for 15 min and 10 000 g for 20 min. The mitochondrial pellet was washed in 0.5 mol.l⁻¹ sucrose, 1 mmol.l⁻¹

EDTA, 0.3% (w/v) BSA, 0.1 mol.l⁻¹ sodium phosphate buffer, pH 7.4 and centrifuged again at 10 000 g for 20 min. The final mitochondrial suspension was carefully resuspended with a small brush in 250 µl of washing medium and kept on ice until use. Protein was determined by the procedure of Hartree (1972).

Measurement of oxygen uptake

Oxygen consumption was measured polarographically at 25 °C with a Clark-type oxygen electrode (Yellow Spring Instrument, USA) fitted to the Oxygraph model KIC (Gilson, USA). The standard reaction medium contained 0.5 mol.l⁻¹ sucrose, 10 mmol.l⁻¹ KCl, 5 mmol.l⁻¹ Mg Cl₂, 1 mmol.l⁻¹ EDTA, 0.1% (w/v) BSA, 10 mmol.l⁻¹ sodium phosphate buffer, pH 7.2. The final volume, including the 50 µl of mitochondrial suspension (0.2–0.4 mg protein), was 2 ml. Respiratory rates were calculated from a recorded trace taking into account that the maximal concentration of oxygen in air-saturated reaction medium was 207 µmol.l⁻¹. This value was found with our mitochondrial fraction in limiting concentrations of NADH (Estabrook and Mackler, 1957). RCR and ADP/O ratios were calculated as described by Estabrook (1967). All substrates were solubilized in the reaction medium and when needed in spite of the buffer, the pH was adjusted to 7.2 with small volumes of 1 N or 6 N KOH. Final concentrations of substrates were 10 mmol.l⁻¹ α-ketoglutarate (plus 0.2 mmol.l⁻¹ NAD⁺ and 0.3 mmol.l⁻¹ TPP), 10 mmol.l⁻¹ succinate (plus 0.2 mmol.l⁻¹ ATP, 1 min before succinate), 1 mmol.l⁻¹ NADH. All of the inhibitors were dissolved in ethanol except NaCN which was dissolved in water. The ethanol volumes used in each experiment were without effect on mitochondrial respiration.

RESULTS

Mitochondrial preparation

The amount of mycelium and the ratio of mycelium/isolation medium were critical for cell disruption. Usually 30–40 g (wet weight) of mycelium with 100 ml of isolation medium gave the best results with our mixer. The disruption time was also very critical for mitochondrial integrity. We obtained the better coupled mitochondria with a disruption time of 3 x 3 s (a longer time provided more mitochondrial protein but mitochondria were not as well coupled). In the above conditions, mitochondrial protein yield was 1–2 mg. The media chosen to prepare mitochondria were as simple as possible but the presence of BSA and EDTA always proved essential, particularly in the respiration medium.

Respiratory properties

When *B cinerea* was grown in the glucose medium, only exogenous NADH and succinate gave a well coupled respiration. This was not the case for endogenous NADH-linked substrates like α -ketoglutarate. To obtain well coupled respiration, not only with exogenous NADH and succinate but also with α -ketoglutarate, *B cinerea* had to be grown in the starch medium. Table I summarizes the results obtained for oxygen consumption, respiratory control and ADP/O ratios of mycelial mitochondria from *B cinerea*. RCR values showed that electron transport in *B cinerea* mitochondria was well coupled to oxidative phosphorylation. Good phosphorylation efficiencies (ADP/O ratios) were obtained with all three substrates. ADP/O ratios suggested that 2 coupling sites were utilized with exogenous NADH and succinate while 3 coupling sites were used with α -ketoglutarate.

Inhibitors of electron transport and oxidative phosphorylation

For all tested substrates, oxidation was completely inhibited by addition of 1 $\mu\text{mol.l}^{-1}$ antimycin A or 0.5 mmol.l^{-1} NaCN (fig 1a,b), which are both known to be effective inhibitors of electron transport, between cytochromes *b* and *c1* and at cytochrome *a a3*, respectively. This indicated that *B cinerea* mitochondria did not exhibit an alternative pathway resistant to antimycin A and cyanide as observed in *Aspergillus oryzae*, *Neurospora crassa* or *Ustilago maydis* (Kawakita, 1971; Lambowitz and Slayman, 1971; Sherald and Sisler, 1972). As expected, the inhibitory effect of antimycin A was shunted by the addition of 0.2 mmol.l^{-1} TMPD, an artificial electron mediator between cytochromes *b* and *c1* (Lee *et al*, 1967).

Rotenone at 10 $\mu\text{mol.l}^{-1}$ inhibited α -ketoglutarate oxidation by up to 30% but failed to have any effect on NADH and succinate oxidation. At 20 $\mu\text{mol.l}^{-1}$ and over, the inhibitory effect of rotenone on α -ketoglutarate oxidation never exceeded 40%. At the same time, the ADP/O ratio decreased to a value indicating that only 2 coupling sites were still functioning (fig 1b). A complete inhibition was obtained with further addition of antimycin A. Such results suggest that *B cinerea* mitochondria have a rotenone-insensitive respiratory route, like higher plant mitochondria (Douce, 1985).

We checked over the activity of TTFA and carboxin (5,6-dihydro-2-methyl-1,4-oxathiin-3-carboxanilide), as specific inhibitors of succinate oxidation at the level of complex II (White *et al*, 1983). While it was necessary to use 2 mmol.l^{-1} TTFA to completely inhibit succinate oxidation, only 10 $\mu\text{mol.l}^{-1}$ of carboxin were sufficient. 1 $\mu\text{mol.l}^{-1}$ carboxin inhibited succinate oxidation by up to 68% (not shown). This efficiency is similar to that reported by Ulrich and Mathre (1972) in *U maydis* mitochondria.

At 15 $\mu\text{mol.l}^{-1}$, oligomycin, an inhibitor of oxidative phosphorylation, added to state 3 (during which the oxidative phosphorylation occurs) decreased the oxidation of all substrates tested to a level close to the respiration normally observed during state 4 (during which no ATP is formed). As found with higher plant mitochondria (Ikuma and Bonner, 1967) this inhibition was released by 3 $\mu\text{mol.l}^{-1}$ of a known uncoupler like CCCP and subsequent addition of ADP had no effect (fig 1a). Three $\mu\text{mol.l}^{-1}$ CCCP added alone during state 4 strongly increased (200%) oxygen consumption (not shown). In all cases a further addition of 1 $\mu\text{mol.l}^{-1}$ antimycin A completely inhibited the oxygen consumption generated by CCCP (fig 1a).

Table I. Respiratory properties of mitochondria isolated from mycelia of *B cinerea*.

Substrates	State 3 respiration rates ($\text{nmol O}_2 \text{ min}^{-1} (\text{mg protein})^{-1}$)	Respiratory control ratios (RCR)	ADP/O ratios
NADH	157 \pm 53	3.6 \pm 0.3	1.23 \pm 0.07
Succinate	115 \pm 40	2.7 \pm 0.8	1.31 \pm 0.08
α -Ketoglutarate	55 \pm 16	3.5 \pm 0.5	2.21 \pm 0.14

Note : The respiratory activities were measured polarographically at 25 °C as described in *Methods*. The ADP concentration was 100 $\mu\text{mol.l}^{-1}$ and reactions were carried with 0.2 to 0.4 mg mitochondrial protein. Mean values \pm SEs are given for 5 determinations.

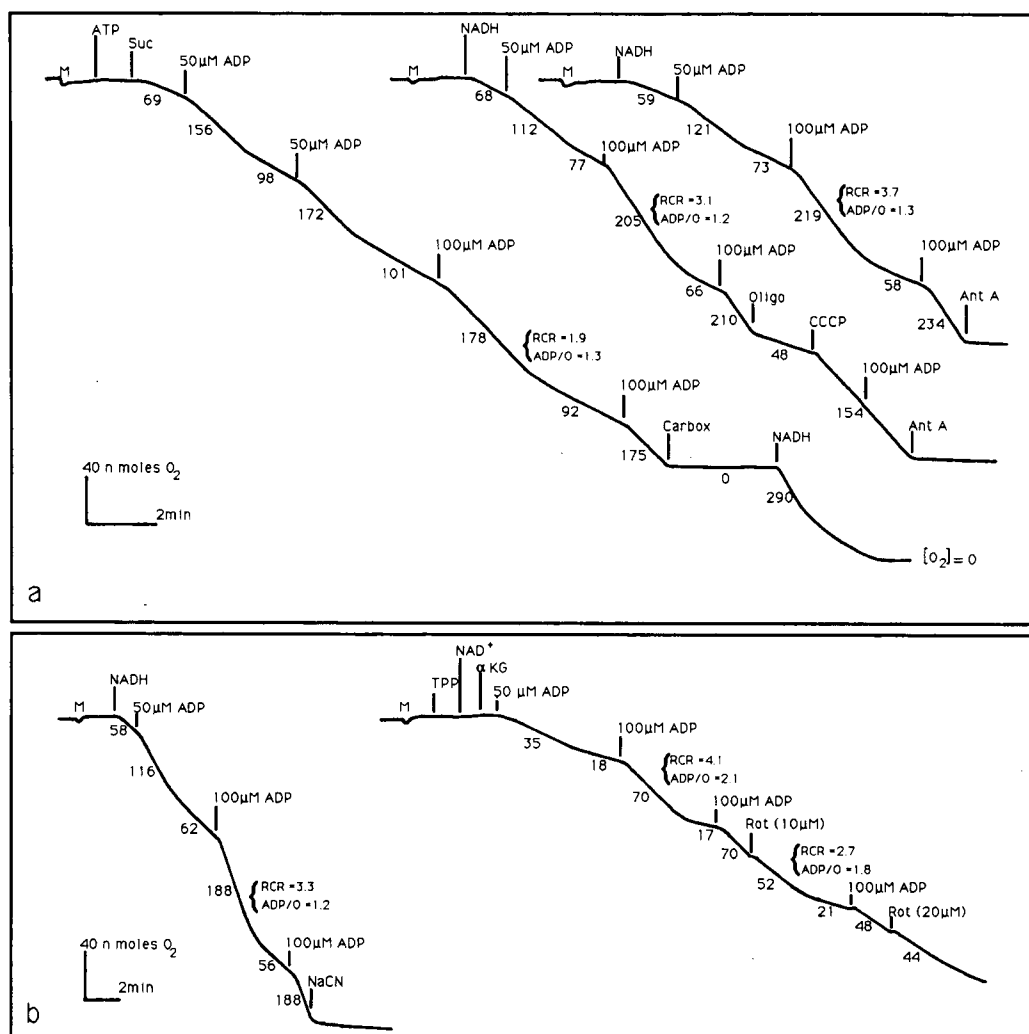


Fig 1. Oxidation of NADH, succinate (Suc) and α -ketoglutarate (α KG) by mitochondria isolated from mycelia of *B cinerea*. Concentrations of substrates were: a) antimycin A (Ant A), $1 \mu\text{mol.l}^{-1}$; carboxin (Carbox), $10 \mu\text{mol.l}^{-1}$; CCCP, $3 \mu\text{mol.l}^{-1}$ and oligomycin (Oligo), $15 \mu\text{mol.l}^{-1}$; b) rotenone (Rot), 10 to $20 \mu\text{mol.l}^{-1}$ and NaCN, 0.5mmol.l^{-1} . The numbers beside each trace show the rate of oxygen uptake in $\text{nmol O}_2 \text{min}^{-1} (\text{mg protein})^{-1}$; the reactions were carried out at 25°C with $0.2\text{--}0.4 \text{mg protein}$.

DISCUSSION

We have shown that it is necessary to grow *B cinerea* in a medium containing starch (instead of glucose) to obtain well coupled mitochondria with all substrates tested. Except for the fact that in the starch medium *B cinerea* was morphologically different and grew more slowly we have no satisfactory explanation for this phenomenon. However, for Schwitzguébel *et al* (1981) the growth conditions changed the density of mitochondria isolated from *N crassa* and this should reflect changes in lipid and/or protein composition of mitochondria.

Our results showed that washed mitochondria isolated from *B cinerea* mycelia, without purifica-

tion procedure, rapidly oxidize α -ketoglutarate, succinate and NADH with good respiratory control and ADP/O ratios, indicating that these organelles sustain a high level of structural and functional integrity. The addition of $30 \mu\text{mol.l}^{-1}$ cytochrome *c* did not greatly enhance the O_2 consumption due to 8mmol.l^{-1} ascorbate (results not shown). This agrees with the good integrity of our washed mitochondria (Neuburger *et al*, 1982).

We have shown that *B cinerea* mitochondria are devoid of the alternative pathway resistant to antimycin A and cyanide. However, by growing *N crassa* in the presence of antimycin A, cyanide or chloramphenicol, Lambowitz and Slayman (1971) induced the alternative pathway. With *B*

cinerea we did not attempt the same experiment, but we did not succeed in inducing the alternative pathway with sodium salicylate which has been described as calorigen in the genus *Arum* (Raskin *et al*, 1987).

With regard to the rotenone-insensitive respiratory route observed in *B cinerea* mitochondria, we have shown that it was different from the alternative pathway because it was completely inhibited by antimycin A.

Carboxin and, more generally, carboxamides are specifically valuable systemic fungicides for control of Basidiomycetes (smuts and rusts of cereals as well as rots caused by *Rhizoctonia solani*). However, carboxin is known to be an inhibitor of complex II of the electron transport system in a variety of organisms, including sensitive fungi (*U maydis*), tolerant fungi (*Fusarium oxysporum*, *Saccharomyces cerevisiae*), higher plants (*Phaseolus vulgaris*) and animals (pig heart, rat liver) (Mathre, 1971; Ulrich and Mathre, 1972). Even though published data indicate that succinate-ubiquinone reductase activities from Basidiomycetes fungi, mammalian sources and bacterium (*Micrococcus denitrificans*) are more sensitive than those from higher plants and the yeast *S cerevisiae*, the selectivity of carboxamides seems to be relative (Kuhn, 1984). Moreover, we have found that carboxin strongly inhibits succinate oxidation in *B cinerea* mitochondria, which is in accordance with the *in vitro* toxicity of this compound, for several Deuteromycetes including *Botrytis* sp, reported by Edgington and Barron (1967). If the present carboxamides failed to control *B cinerea* in the field, this is not due to a differential sensitivity of the target site or a lower accessibility of the fungal wall and plasmalemma to this site.

The reason for this is to be found among the obstacles encountered by systemic fungicides before reaching the fungus such as biological and non-biological conversion into non-fungitoxic substances. Chin *et al* (1969) reported that carboxin is oxidized to its inactive sulfoxide in water, soil and plants like wheat, barley and cotton. Maybe it could be possible to find particular structural analogs of carboxin which are effective in practice, among carboxamides.

With *B cinerea* mitochondria we can easily test whether a product is an inhibitor of the oxidative phosphorylation or an uncoupler by using respectively CCCP or oligomycin. With mitochondria from potato tubers, Macherel *et al* (1986) reported another uncoupling test using carboxyatracty-

loide, an inhibitor of adenine nucleotide transport. We had obtained the same results with *B cinerea* mitochondria and 10 $\mu\text{mol.l}^{-1}$ carboxyatractyloside instead of 15 $\mu\text{mol.l}^{-1}$ oligomycin (not shown).

If we except the alternative pathway, we have observed a respiratory chain similar to that encountered in the mitochondria of microorganisms (Lloyd, 1974) in *B cinerea* mitochondria. The washed mitochondria described in the present paper will allow us to accurately study the mode of action of fungicides on the respiratory chain of *B cinerea*.

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