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The Metabolic Clearance Rate of Estradiol-17β in Rainbow Trout, *Salmo gairdneri* R., Estimated by Both Single Injection and Constant Infusion Methods: Increase during Oocyte Maturation

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A dual cannulation of free-swimming rainbow trout is used to estimate the metabolic clearance rate (MCR) of Estradiol-17β (E2-17β) by both single injection and constant infusion methods. It is shown that E2-17β MCR changes significantly during the progress of oogenesis, mainly at the end of the sexual cycle. The same changes in MCR and very similar values are found with both single injection and constant infusion methods: MCR is stable (28.8 ml/hr/kg) from the postovulation period (throughout endogenous vitellogenesis) to early exogenous vitellogenesis. It decreases significantly during advanced exogenous vitellogenesis (18.7 ml/hr/kg) and increases clearly at the onset of oocyte maturation (40.9 ml/hr/kg). A direct relationship between MCR and plasma E2-17β occurs: Plasma E2-17β levels increase (advanced exogenous vitellogenesis) when MCR decreases. Then estradiol decline takes place at the same time that MCR reaches its highest values (oocyte maturation). An increase in MCR is probably one event required to allow the establishment of an appropriate hormonal environment for oocyte maturation.

The best known biological activity of estradiol-17β (E2-17β, 1,3,5(10)-estratrien-3,17β-diol) in fish is the hepatic stimulation of vitellogenin synthesis (for review see Ng and Idler, 1983). However, a role played by estradiol on meiotic maturation and ovulation has also been suggested recently. Estradiol-17β lowers the efficiency of the maturational gonadotropin (GtH) *in vitro* to induce intrafollicular oocyte maturation in rainbow trout (Jalabert, 1975), and reduces the output by ovarian follicles of 17α-hydroxy-20β-dihydroprogesterone (17α,20β-OH-P,4-pregnen-17α,20β-diol-3-one), the most potent steroid inducer of this maturational process (Fostier *et al*., 1973; Jalabert, 1976; Jalabert and Fostier, 1984a). Furthermore, a negative estrogen feedback on the pituitary has been shown at the migratory germinal vesicle stage, probably preventing the ovulation surge of GtH in trout (Bommelaer *et al*., 1981), although this does not seem to be true in goldfish (Pankhurst and Stacey, 1985). Therefore, the vitellogenic steroid appears to be an antimaturational hormone (Jalabert and Fostier, 1984b) and the decrease in its plasma levels may be an obligatory event prior to spontaneous ovulation. Such decline has been recorded in various salmonid species (Fostier *et al*., 1978; Jalabert *et al*., 1978; Fostier and Jalabert, 1982; Scott *et al*., 1983; Sower and Schreck, 1982; Van der Kraak *et al*., 1984; Kagawa *et al*., 1983) and correlated with a decrease of the ovary's ability to secrete estradiol-17β (Zohar, 1982; Kagawa *et al*., 1983), hypothesized to be basically a fall in aromatase activity (Young *et al*., 1983). However, very little attention has been given to the clearance of the hormone from the blood which plays a significant role in the regulation of its plasma level. Most of the research on clearance of steroids in fish concerns corticosteroids (Idler *et al*., 1963; Donaldson and Fagerlund, 1968, 1969; Fagerlund and Donaldson, 1969, 1970; Idler and Truscott, 1969; Owen and Idler, 1972; Henderson *et al*., 1974; Leloup-Hatey,
1974, 1976; Goodman and Butler, 1976; Truscott et al., 1977; Patino et al., 1985) but very little deals with sexual steroids.

Two methods can be used to estimate the metabolic clearance rate (MCR) of a hormone: (1) the analysis of the disappearance curve of the labeled hormone from the blood after a single rapid injection, and (2) the measurement of steroid entering the blood, once steady state is reached, by means of a constant infusion (Tait, 1963). A continuous infusion of tritiated testosterone was done through the branchial venous system by Fletcher et al. (1969), to estimate testosterone MCR in sexually mature male and female skate, Raja radiata. Blood samples were withdrawn via a second cannula which was put in the caudal artery. Recently, Querat et al. (1982, 1985) chose the single injection method to determine estradiol MCR in immature silver eels; injection and sampling were done through a cannula inserted into the pectoral artery. To our knowledge, these are the only two studies on sexual steroid MCR in fish. Others works deal with steroid accumulation in various target tissues (Schreck, 1973; Myers and Avila, 1980) or with the problem of the residues’ half-life after anabolic treatments (Johnstone et al., 1978; Fagerlund and McBride, 1978; Fagerlund and Dye, 1979; Lone and Matty, 1981; Johnstone et al., 1983) but the methods used here were not appropriate for MCR estimations.

The following study was performed to measure the estradiol metabolic clearance rate in female rainbow trout, at various stages of ovogenesis, mainly during the preovulation period. Both single injection and constant infusion methods were used and compared. For this, the single cannulation technique (Zohar, 1980; Bry and Zohar, 1980) was modified to obtain a dual cannulation of free-swimming trout.

MATERIALS AND METHODS

Animals. Three-year-old female rainbow trout (1.3 to 2 kg) were used, most of them belonging to a “winter strain” spawning in November–December. A few animals of a “spring strain,” spawning in March–April, were also used in the study with the constant infusion method. Fish were kept in a recirculating water system (at 12 ± 2°C) and submitted to a natural photoperiod.

At the end of the experiments the fish were killed and weighed. Ovary fragments were fixed in Bouin–Holland solution and stained by Regaud’s hematoxylin and Heidenheim’s Azan (Gabe, 1968).

When feasible, the oocyte maturation stage was estimated by observation with a stereomicroscope (Jalabert and Fostier, 1984b).

The following stages were identified:

- Advanced endogenous vitellogenesis: Vitellin vesicles (stained blue with Azan) and perinuclear lipoidic globules appeared.
- Beginning of exogenous vitellogenesis: Vitellin granules stained with orange-G were visible in the peripheral cytoplasmic area.
- Advanced exogenous vitellogenesis: The vitellin granules spread toward the middle of the cytoplasm, coalesced, and grew in size. The vitellin vesicles transformed into cortical vesicles.
- Preovulation stages were defined according to Jalabert and Fostier (1984b):
  - Subperipheral germinal vesicle (SPGV): The germinal vesicle began to appear at the oocyte periphery and could be observed as a diffuse clear spot at the center of the dark cytoplasm.
  - Germinal vesicle breakdown (GVBD).
- Post-ovulation period: These fish ovulated between the day of cannulation and the end of the constant infusion.

Cannulation. The dorsal aorta catheterization was performed as described earlier by Zohar (1980). Briefly, fish were maintained anesthetized on an operation table, and their gills were perfused with 300 ppm phenoxyethanol solution. An 80-cm-long catheter filled with 0.7% saline solution containing lithium heparinate (150 IU/ml) was inserted into the dorsal aorta at the midline of the buccal roof, slightly anterior to the first gill bar. For the caudal vessel catheterization, a metal needle with an inner diameter closely fitting the outer diameter of the cannula (Clay–Adams PE-50) was fixed on a 5-ml syringe. The needle was inserted just under the lateral line, a little behind the adipose fin at about a 20° angle between the skin and needle. Penetration of the vessel was indicated by blood entering into the syringe after pumping slightly. A 120-cm-long catheter filled with heparinate solution (150 IU/ml) was placed into the needle and inserted 8 cm into the blood vessel; then the guiding needle was gently withdrawn. The cannula was sutured to the skin behind the penetration point and passed along the body through a subcutaneous tunnel made with a 20-cm-long metal needle; then it was sutured to the skin on its head near the cannula coming from the...
injected steroid was partially diluted with plasma from tubing. Thus, the 500 µl used for the single injection into the experimental animal to prevent the adsorption to was composed of half 0.7% saline solution and half plasma; and in the constant infusion experiment trout balanced solution, without any glucose or antibiotic (Jalabert and Fostier, 1984a), was used with 1% plasma.

In the first case 2–7 µCi/kg body wt of [3H]E2-17β was injected and in the second case 0.26–0.98 µCi/hr/kg body wt was given.

Steroid extraction and purification. In the constant infusion experiment, plasma aliquots were treated with 300–600 µl of a tissue solubilizer (NCS, Amersham, U.K.) to measure the total plasma radioactivity. To the remaining part of this plasma, 5 µg of E2-17β was added (Steraloids) as carrier. All samples were extracted twice with 2 ml cyclohexane–ethylacetate (50:50); then an aliquot of the total extractable radioactivity was counted, and the remaining extract was chromatographed on Sephadex LH-20 columns. Two different-sized columns were used: 0.5 × 12 cm in the single injection treatment with benzene–methanol (85:15) as eluant (Foster et al., 1978) and 0.5 × 8 cm in the constant infusion method with dichloromethane–methanol (95:5). The estrone and estradiol fractions were collected and their radioactivity measured. For the constant infusion samples, the estradiol fraction was then purified by high-pressure liquid chromatography (HPLC, Waters, pumps M 45 and 6000 A) using a reverse-phase system: Bondapack C-18 column (Waters, diameter 4 mm, length 30 cm) eluted with methanol:water (57:43). Estrogens were detected by their absorption at 280 nm (Detector Waters 440).

The constancy of the [3H]/[14C] ratio after three successive crystallizations in hexane:ethanol was used as an identification criterion (Axelrod et al., 1965) after addition of 10 µg of the cold E2-17β.

Radioactivity measurements. Organic phases were dried and dissolved in methanol, and a 50-µl aliquot was added with 5 ml OCS scintillator. Plasma digested by the tissue solubilizer was completed with 5 ml liquid scintillation cocktail (Picofluor, Packard). Radioactivity was counted on an Intertechnique SL 4000 counter (France) and expressed in disintegrations per minute (dpm). The tritium content of the sample was corrected for losses during purification, using the [14C] recovery.

Data analysis. Single injection: if R is the total radioactivity cleared (the total radioactivity injected) and x(t) the plasma radioactive concentration according to time, the metabolic clearance rate is defined (Tait, 1963) by

\[
\text{MCR} = \frac{R}{\int_0^t x \, dt}
\]

The curve of the hormone disappearance from the blood can be described as the sum of three exponentials. The first one, constructed with the very first plots (within the first minute = two to three first samples), was attributed to the tracer dilution in the...
plasma; thus \( x(t) \) was described by a double exponential: \( x(t) = Ae^{-\alpha t} + Be^{-\beta t} \) and then

\[
\text{MCR} = \frac{\alpha B}{\beta A + \alpha B}
\]

\( \alpha, \beta, A, \) and \( B \) were calculated by linear regression analysis of the plasma disappearance curve after logarithmic transformation. **Constant infusion:** At the steady state, when the plasma radioactive concentration \( x \) has become stable, with the infusion at a constant rate \( r \), counterbalancing exactly metabolism of the labeled steroid, then \( \text{MCR} = \frac{r}{x} \) ml of plasma per time unit. MCR was calculated for each fish from the mean of all the radioactive concentrations during the steady state. Then it was expressed in Table 2 as a mean \( \pm \) SE. In both cases, MCR was calculated per kilogram of body weight.

**Statistics.** MCR values of studied oogenesis stages were compared with Wilcoxon–Mann and Whitney nonparametric tests (Snedecor and Cochran, 1971).

**RESULTS**

**Steady-State Characteristics in Animals under Constant Infusion**

A steady state was reached within 1 to 4 hr in the constant infusion method. Then plasma concentrations of radioactivity remained stable (Figs. 1 and 2) until the end of infusion (22 to 50 hr).

**Identification of the Plasma Radioactivity**

In blood samples collected after the single \( E_2-17\beta \) injection, 95% of tritiated counts, recovered with solvent extraction, could be attributed to estradiol, after chromatography on Sephadex LH-20 columns. Thus, the disappearance curves of the total extractable radioactivity from the plasma and those of radioactive estradiol were parallel and almost superimposed (Fig. 3).

In samples obtained in the constant infusion method, 90% of the total plasma radioactivity was represented by unconjugated

---

**FIG. 1.** \( E_2-17\beta \) plasma radioactivity during constant infusion of rainbow trout at two different oogenesis stages: advanced endogenous (○) and advanced exogenous (●) vitellogenesis.

**FIG. 2.** \( E_2-17\beta \) plasma radioactivity during constant infusion of rainbow trout at two different oogenesis stages: oocyte maturation (○) and ovulation (●).

**FIG. 3.** Plasma disappearance curves of total extractable radioactivity (●, cyclohexane:ethylacetate, 50:50) and radioactive estradiol (○, purified by Sephadex LH-20 chromatography) after injection of one dose of \([2,4,6,7-\text{H}]\)estradiol in cardiovascular system.
CHANGES IN ESTRADIOL-17β MCR IN TROUT

steroids (extractable radioactivity). These latter samples, after chromatography on HPLC using the 14C activity to assess the recovery, were composed of 90% E2-17β.

The identification of estradiol was confirmed for two constant infusion experiments after column chromatography (normal pressure chromatography on Sephadex LH-20 and HPLC on C-18 Bondapack) by the constancy of the 14C/3H ratio through successive crystallizations (Table 1).

For three animals treated with constant infusion, estrone fractions, obtained after chromatography on Sephadex LH-20 column, were measured as representing 7 to 25% of estradiol (Fig. 4).

Plasma Disappearance of Radioactive Estradiol after Single Injection

The patterns of the E2-17β disappearance curves in the three different oocyte developmental stages were similar (Fig. 5). These curves could be described by the sum of two exponentials.

Metabolic Clearance Rate

Single injection. MCR values (Table 2) decreased from the beginning of exogenous vitellogenesis (23.1 to 34.8 ml/hr/kg) up to advanced exogenous vitellogenesis stages (18.5 to 23.8 ml/hr/kg). Highest values were found during the first stages of oocyte maturation (32.8 to 45.2 ml/hr/kg).

Constant infusion. MCR remained constant from the postovulation period up to the beginning of exogenous vitellogenesis (21 to 35.5 ml/hr/kg; mean = 27.9 ml/hr/kg); it fell to the lowest value (14 ml/hr/kg) during advanced exogenous vitellogenesis. Then a clear rise occurred during oocyte maturation (40.7 to 45 ml/hr/kg).

Statistical Test

Data from both methods were comparable, and thus they were gathered to test statistically (Wilcoxon–Mann and Whitney nonparametric tests) the differences between the various stages. The MCR values measured at oocyte maturation were tested against those measured at the postovulation period up to advanced exogenous vitellogenesis or up to the beginning of exogenous vitellogenesis. The values at the oocyte maturation stage were always highly significantly greater than the others. Furthermore, the values of advanced exogenous vitellogenesis were significantly lower than those of the stages extending from the postovulation period up to the beginning of exogenous vitellogenesis.

DISCUSSION

The determination of the MCR of estradiol is necessary to understand the regulation of its plasma levels during ovogenesis. For such a study to be valid requires that the injected labeled hormone should not disturb the normal distribution and metabolism of the endogenous steroid (Tait, 1963). In the present work, single injection produced an initial plasma concentration of 0.5 to 1.5 ng of the labeled steroid per milliliter of blood. This value is below or equal to the endogenous hormone level. In the constant infusion method, the highest rate was 2.9 ng/hr for a fish weighing 2.1 kg. The blood volume of such a fish is estimated to

<table>
<thead>
<tr>
<th>Number</th>
<th>Sephadex LH-20</th>
<th>HPLC</th>
<th>First crystallization</th>
<th>Second crystallization</th>
<th>Third crystallization</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td>0.27</td>
<td>0.23</td>
<td>0.22</td>
<td>0.23</td>
<td>—</td>
</tr>
<tr>
<td>2</td>
<td>0.35</td>
<td>0.34</td>
<td>0.32</td>
<td>0.33</td>
<td>0.32</td>
</tr>
</tbody>
</table>
be 70 ml, according to a ratio of 3.5 to 4.5% of blood to body weight (Smith, 1966; Niki­

kinmaa et al., 1981). Thus without any epuration, the dilution of the labeled hor­

mone in the whole blood volume would produce an increase of 42 pg/ml in E$_2$-17$eta$ concentration, compared with a level of 200 pg/ml measured by RIA before the be-

ginning of the perfusion. In both cases, this variation is relatively low. Therefore, our experimental conditions permit the mea-

surement of MCR.

In both single injection and constant infu-

sion methods, the same MCR changes and very similar values were obtained; in both cases, plasma radioactivity plotted against time gave reproducible curve shapes. Thus, the two techniques seem to be adequate for MCR measurements, and the sum of two exponentials can reason-

ably be used to fit the plasma E$_2$-17$eta$ disappearance curve after single injection.

Most of the nonconjugated radioactivity can be attributed to E$_2$-17$eta$ both in single injection and constant infusion samples, as also was reported in the immature eel by Querat et al. (1985) after Celite chromatog-

raphy. However, E$_2$-17$eta$ has been identified by following the constancy of the $^3$H/$^{14}$C ratio during successive crystallizations in only a few of our samples. Plasma radioac-

tivity decreased rapidly; thus, the amount of labeled steroid in each sampling became too small for these purification stages. Nevertheless, Sephadex LH-20 chromatog-

raphy appears sufficient for attaining a high degree of purity (Table 1).

Mammal and hen E$_2$-17$eta$ MCR data are 30 to 100 times (Reed and Murray, 1979) and 125 to 200 times (Tsang and Grunder, 1984) higher, respectively, than ours. By contrast, the MCR of immature eel (Querat et al., 1985) is about 40 times lower than that of our values for the trout. These discrepancies in MCR among different fish species, mammals, and birds might be ex-

plained, at least in part, by specific thermoregulation modes or specific sex ster-

oid-binding protein (SBP) systems. Ther-

moregulation modes may play a significant role in these specific rates as temperature in fish is known to influence hepatic and ovarian metabolic activities (Kime and Saksena, 1980; Manning and Kime, 1984). Specific SBP systems suggest another pos-

sible explanation. Sandberg et al. (1966)
TABLE 2
Metabolic Clearance Rate (ml/hr/kg) of E₂-17β in Rainbow Trout at Various Stages of Oogenesis Estimated by Both Single Injection and Constant Infusion Methods

<table>
<thead>
<tr>
<th>Method</th>
<th>Advanced endogenous vitellogenesis</th>
<th>Beginning of exogenous vitellogenesis</th>
<th>Advanced exogenous vitellogenesis</th>
<th>Oocyte maturation</th>
<th>Post-ovulation period</th>
</tr>
</thead>
<tbody>
<tr>
<td>Constant infusion</td>
<td>25.2 ± 0.5</td>
<td>31.5 ± 0.6</td>
<td>27.2 ± 0.9</td>
<td>31 ± 1.5</td>
<td>45 ± 1.7</td>
</tr>
<tr>
<td>Single injection</td>
<td>34.8</td>
<td>23.1</td>
<td>18.5 ± 0.3</td>
<td>23.8 ± 0.4</td>
<td>45.2 ± 0.5</td>
</tr>
<tr>
<td>Mean/oocyte stage</td>
<td>27.9</td>
<td>28.8</td>
<td>18.7 ± 0.3</td>
<td>40.9 ± 0.5</td>
<td>26.8</td>
</tr>
</tbody>
</table>

Note. SPGV: subperipheral germinal vesicle; GVBD: germinal vesicle breakdown.

have shown that in mammals, the greater a steroid is bound, the less it will be metabolized; and recently, Petra et al. (1985) reported a decrease in the clearance rate of testosterone after the infusion of pure SBP. In fish steroid-binding capacities are higher than those for the mammalian binding system (Wingfield, 1980); it is the case for teleosts in which a sex steroid-binding protein has been described (trout: Fostier and Breton, 1975; eel: Querat et al., 1983). Thus, we can assume that differences in SBP characteristics among species would result in different MCR values. In the present study, MCR values were shown to change during oogenesis: these values were stable from the postovulation period throughout endogenous vitellogenesis and early exogenous vitellogenesis. They then decreased significantly during advanced exogenous vitellogenesis and increased clearly at the onset of oocyte maturation. Three stages of ovarian development were examined by both single injection and constant infusion methods; for each of them, MCR measurements were comparable (Table 2). Our data, derived from two methods, indicate that the previously described slow rise in circulating E₂-17β levels occurring at early vitellogenesis (Lambert et al., 1978; Whitehead et al., 1978; Scott et al., 1980; Van Bohemen and Lambert, 1981; Breton et al., 1983) is accompanied by a quite stable basal MCR (endogenous vitellogenesis 27.9 ml/hr/kg and beginning of exogenous vitellogenesis 28.8 ml/hr/kg). Since the MCR remained constant, in this early vitellogenic stage, the slight estrogen increase may have been the consequence of an augmentation of its secretion rate by the ovaries. Later, at the exogenous vitellogenesis and preovulation stages, when some of the most important events for the oocyte begin (vitellogenin synthesis and incorporation, hormonal environment changes), there is a direct relation between MCR and circulating E₂-17β levels. Variations in plasma E₂-17β levels are parallel to its MCR: the lowest MCR (range = 14–23.8 ml/hr/kg, mean = 18.7 ml/hr/kg) occurred during advanced exogenous vitellogenesis, an oogenic stage known for the highest plasma E₂-17β concentrations. So when the MCR decreased, E₂-17β circulating levels increased. In the same way, the highest MCR (range = 32.8–45.2 ml/hr/kg, mean = 40.9 ml/hr/kg) coincided with a decrease in estradiol during gametogenesis. A clear increase of the clearance activity is thus shown when the vitellogenic hormone is decreasing. Seasonal variations in the binding protein capacity might exist in the female rainbow trout, which would explain, at least partially, the variations in MCR. Differences in steroid MCR (testosterone) between males and females have been reported in skate (Fletcher et al., 1969) but, to our knowl-
edge, no MCR variations have been shown in relation to the development of gametogenesis in fish. Ng and Idler (1980) found that in flounder and in salmonids, hypophysectomy was followed by the disappearance of circulating SBP, which could be restored by treatment of both glycoproteic and nonglycoproteic gonadotropins. GtH is one component of the circulating hormonal environment submitted to important variations during the latest stages of oocyte development in rainbow trout; thus, it could underlie a possible regulation of these SBP capacities. In addition, hypophyseal hormones appear to play a role in the regulation of the metabolism of androstenedione by liver microsomes from juvenile rainbow trout (Hansson and Gustafsson, 1981). Such a regulation may exist for the hepatic metabolism of estradiol-17ß (Hansson and Rafter, 1983). Following the end of vitellogenesis, the decrease in E2-17ß is probably essential for the establishment of a hormone environment required for oocyte maturation (Jalabert and Fostier, 1984b).

Sexual steroids have been studied mainly with regard to their secretion or their circulating levels but more attention should be paid to their clearance, especially during periods of sudden or dramatic variations in their concentration.

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