

Adaptation of Listeria monocytogenes to temperature: exploration of intracellular subproteome through shotgun proteomics

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Listeria monocytogenes is a foodborne pathogen

(listeriosis) in susceptible patients. Most human

consumption of refrigerated ready-to-eat foods.

Although initial contamination levels in foods are

usually low, the ability of these bacteria to survive

and multiply at low temperatures allows it to reach

levels high enough to cause disease. It is able to

grow in a broad spectrum of temperatures, between

proteins (i) could be related to the adaptation of *L.*

monocytogenes at different temperatures and (ii)

differently expressed according to

Protein extraction from cultures in planktonic mode

of growth was performed in stationary phase after

48h, 20h and 16h at 10°, 25° and 37°C, respectively.

For sessile cells (biofilms), protein extraction was

carry out in mid-log phase after 24h, 8h and 4h at

10°, 25° and 37°C, respectively. In the cellular

fractionation method (Figure 1) cells were washed

twice in Tris-EDTA. Pellet was resuspended in 1 ml

TE and bacterial cells were broken using a cell

disrupter by applying 2.5 kBar pressure. Insoluble

materials containing cell walls were removed by

centrifugation. After trypsin digestion of proteins,

peptides separation and identification were

MS/MS) and MS data were analysed by Progenesis

The first preliminary results from cells in biofilm at

mid-log phase and planktonic cells at stationary

phase allowed to identify 845 and 954 proteins

respectively. After analysis of variance, 14 proteins

from the biofilm samples and 15 from planktonic

samples were statistically significant (a p-value of

<0.05 was considered significant). Many of the

QI (Figure 2).

Results

by shotgun proteomics (LC-QTOF-

planktonic versus biofilm mode of growth.

Materials and Methods

appear

to

be

Adaptation of Listeria monocytogenes to temperature: exploration of intracellular subproteome through shotgun proteomics







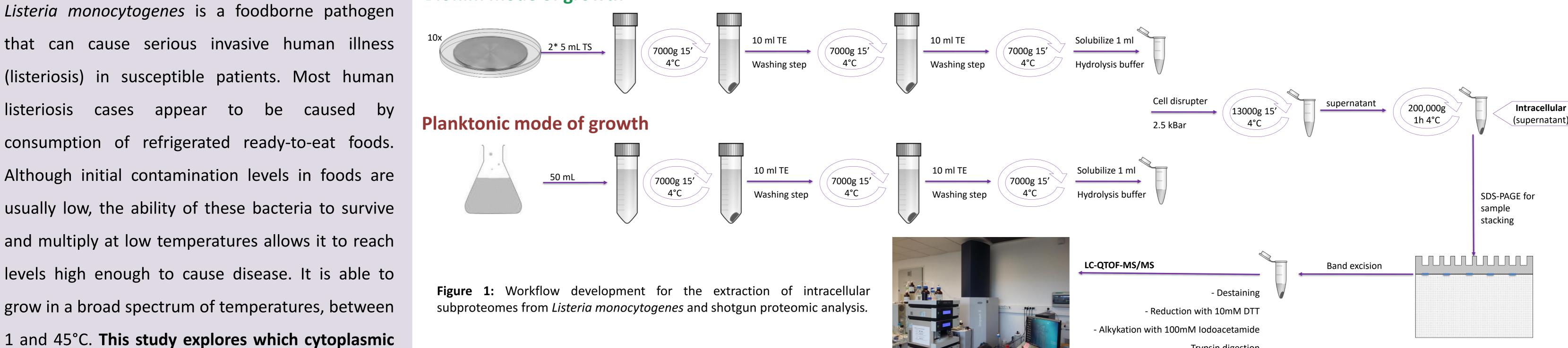
caused by



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Fractionation and LC-MS/MS workflow Introduction

Biofilm mode of growth



Tiago Santos¹, Christophe Chambon², Didier Viala², Julia Esbelin¹ and Michel Hébraud^{1,2}

Discussion

Biofilm mode of growth

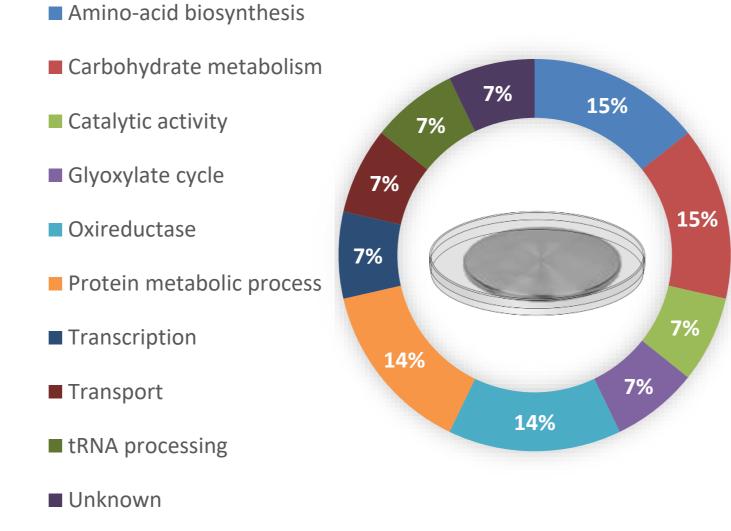


Figure 2: Distribution by biological process of the 14 statistical significant identified proteins from biofilm cultures and extraction at the mid-log phase.

■ Cell redox homeostasis ■ Enzymatic activity Hydrolase Oxireductase activity Transcription

Planktonic mode of growth

- Trypsin digestion

■ tRNA methylation

Unknown

Figure 3: Distribution by biological process of the 15 statistical significant identified proteins from planktonic cultures and extraction at the stationary phase.

- The inosine-5'-monophosphate dehydrogenase guab was identified with higher abundance at 37°C. The overexpression of guaB in L. monocytogenes could reflect a particular need for purines in surviving cells where DNA is being repaired.
- Lmo0135, which was also more abundant at 37°C, has a peptide transporter activity shown to be required for *L. monocytogenes* virulence and acid resistance.
- The ATP-dependent protease hslV is important for managing protein levels and directing stress responses and here it was detected in higher abundance in the 10°C condition.
- Glycerol-3-phosphate dehydrogenase glpD is known to be involved in glycerol uptake and metabolism, this oxireductase was more abundant in the 37°C extraction.

Table 1: Statistical significant identified proteins of interest from biofilm cultures and extraction at the mid-log phase.

| Gene | Acession | Highest | Lowest | Protein | Function | Subcellular |
|---------|----------|-----------|-----------|----------------------|-----------------------------|-------------|
| | | mean | mean | | | location |
| | | condition | condition | | | |
| guaB | Q926Y9 | Condition | Condition | Inosine-5'- | Catalytic activity playing | Cytoplasm |
| | | 37°C | 25°C | monophosphate | a role in the regulation of | |
| | | | | dehydrogenase | cell growth | |
| Lmo0135 | Q8YAJ0 | Condition | Condition | Lmo0135 | Peptide transporter | Periplasmic |
| | | 37°C | 25°C | | activity | space |
| HSLV | Q8Y7J9 | Condition | Condition | ATP-dependent | metal ion binding | Cytoplasm |
| | | 10°C | 25°C | protease subunit | | |
| glpD | Q8Y7I4 | Condition | Condition | Glycerol-3-phosphate | Oxireductase | Cytosol |
| | | 10°C | 37°C | dehydrogenase | | |

FtsK protein is a cell division protein located at the cell septa and which coordinates chromosomal dimer segregation during cell division resulting in the separation of the daughter cells. Universal stress proteins (Usp) of L. monocytogenes are up regulated by σB. Usps are proteins that accumulate in cells during stationary phase and during a variety of stress conditions (heat shock, ultraviolet light, ethanol stress etc.) causing growth arrest in cells. Furthermore, Usp proteins are required for the management of DNA damage and are induced by mutations in the FtsK protein. In *L. monocytogenes*, FtsK expression is σB-dependent, suggesting coordinated expression with Usps to prevent chromosomal damage during cell separation. In this analysis, from stationary phase, this DNA translocase was more abundant at 37°C.

Table 2: Statistical significant identified protein of interest from planktonic cultures and extraction at the stationary phase.

| Gene | Accession | Highest | Lowest | Description | Function | Cellular | | | |
|------|-----------|-----------|-----------|-----------------|-------------|----------|--|--|--|
| | | mean | mean | | | location | | | |
| | | condition | condition | | | | | | |
| ftsK | Q8Y7A3 | Condition | Condition | DNA translocase | DNA binding | Cell | | | |
| | | 37°C | 25°C | | | membrane | | | |

identified proteins are connected to basic cell functions (Figure 2 and 3) but some are related to the temperature adaptation (Table 1 and 2). Among the thermoregulated proteins, some are particularly

overexpressed at 37°C, the temperature at which *L.* monocytogenes is virulent, and others at low temperature condition, as those prevailing in food workshops.

Conclusions

More biological replicates and shotgun analyses of intracellular subproteomes will complement these preliminary results so that comparisons of protein expression throughout several conditions (temperatures, mode / phase of growth) feed databases and help to model regulatory circuitry that drive adaptation of L. monocytogenes to harsh conditions. An ongoing surfaceome study in these different conditions will also contribute for these goal.