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Daisy Bougard, Maxime Belondrade, Christiane Segarra, Vincent Béringue,

Joliette Coste

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We found that PrP226* indeed accumulates in aggregates during the disease, but is also present naturally in brain in minute amounts. With our ELISA test we were able to follow the distribution of this fragment. PrP226* is most likely to accumulate in cerebellum, followed by cortical regions, but very rarely in spinal cord or olfactory bulb.¹⁰ Comparison of the results for PrP226* and PrP^{Sc} revealed that the distribution of both proteins correlates well, and strongly indicates that PrP226* is part of PrP^{Sc} aggregates (Lukan and Černilec, et al. under revision).

References

- Notari S, Strammiello R, Capellari S, Giese A, Cescatti M, Grassi J, Ghetti B, Langeveld JP, Zou WQ, Gambetti P, et al. Characterization of truncated forms of abnormal prion protein in Creutzfeldt-Jakob disease. J Biol Chem 2008; 283:30557-65; PMID:18753138; http://dx.doi.org/10.1074/jbc.M801877200
- Zanusso G, Farinazzo A, Prelli F, Fiorini M, Gelati M, Ferrari S, Righetti PG, Rizzuto N, Frangione B, Monaco S. Identification of distinct N-terminal truncated forms of prion protein in different Creutzfeldt-Jakob disease subtypes. J Biol Chem 2004; 279:38936-42; PMID:15247220; http://dx.doi.org/10.1074/jbc.M405468200
- Zou WQ, Capellari S, Parchi P, Sy MS, Gambetti P, Chen SG. Identification of novel proteinase K-resistant C-terminal fragments of PrP in Creutzfeldt-Jakob disease. J Biol Chem 2003; 278:40429-36; PMID:12917418; http://dx.doi.org/10.1074/jbc. M308550200
- Chesebro B, Trifilo M, Race R, Meade-White K, Teng C, LaCasse R, Raymond L, Favara C, Baron G, Priola S, et al. Anchorless prion protein results in infectious amyloid disease without clinical scrapie. Science 2005; 308:1435-9; PMID:15933194
- Chesebro B, Race B, Meade-White K, Lacasse R, Race R, Klingeborn M, Striebel J, Dorward D, McGovern G, Jeffrey M. Fatal transmissible amyloid encephalopathy: a new type of prion disease associated with lack of prion protein membrane anchoring. PLoS Pathog 2010; 6:e1000800; PMID:20221436; http://dx.doi.org/10.1371/ journal.ppat.1000800
- Stöhr J, Watts JC, Legname G, Oehler A, Lemus A, Nguyen HO, Sussman J, Wille H, DeArmond SJ, Prusiner SB, et al. Spontaneous generation of anchorless prions in transgenic mice. Proc Natl Acad Sci U S A 2011; 108:21223-8; PMID:22160704; http://dx.doi.org/10.1073/pnas.1117827108
- Kosmač M, Koren S, Giachin G, Stoilova T, Gennaro R, Legname G, Serbec VČ. Epitope mapping of a PrP(Sc)-specific monoclonal antibody: identification of a novel C-terminally truncated prion fragment. Mol Immunol 2011; 48:746-50; PMID:21176851; http://dx.doi.org/10.1016/j.molimm.2010.11.012
- Curin Serbec V, Bresjanac M, Popovic M, Pretnar Hartman K, Galvani V, Rupreht R, Cernilec M, Vranac T, Hafner I, Jerala R. Monoclonal antibody against a peptide of human prion protein discriminates between Creutzfeldt-Jacob's disease-affected and normal brain tissue. J Biol Chem 2004; 279:3694-8; PMID:14593100; http://dx.doi. org/10.1074/jbc.M310868200
- Dvorakova E, Vranac T, Janouskova O, Černilec M, Koren S, Lukan A, Nováková J, Matej R, Holada K, Čurin Šerbec V. Detection of the GPI-anchorless prion protein fragment PrP226* in human brain. BMC Neurol 2013; 13:126; PMID:24063733; http://dx.doi.org/10.1186/1471-2377-13-126

P.224: Evaluation of assays intended for the diagnosis of variant CJD

Jillian K Cooper, Kaetan Ladhani, and Philip D Minor

CJD Research and Resource Centre, National Institute for Biological Standards and Control (Medicines and Healthcare Products Regulatory Agency); South Mimms, Hertfordshire, UK

Keywords: variant CJD, diagnostic test, evaluation

Tests intended for the diagnosis of potentially life threatening disorders are listed in Annex IIA of the EU directive and have

associated minimum requirements outlined in a common technical specification. Variant CJD diagnostic tests were added to Annex IIA in 2011.

Typically tests are evaluated for sensitivity using a large number of clinically relevant samples, for vCJD this is not possible due to the small number of clinical cases and the very limited number of relevant samples. To help ensure fair and appropriate access to the small number of rare samples held at the UK CJD Resource Centre its oversight committee have developed a process to assess sensitivity and specificity (http://www.nibsc.org/ Spotlight/CJD_Resource_Centre/CJD_Tests.aspx).

This process involves analysis of 1. vCJD tissue homogenates spiked into the relevant blood component (to determine analytical sensitivity) 2. Blood components from experimentally or naturally infected animals (diagnostic sensitivity) 3. Blood components from normal donor samples (for diagnostic specificity).

Each stage of the evaluation process is reviewed by the oversight committee and if minimum requirements are met samples from vCJD clinical cases are provided for testing.

Two test developers have completed the evaluation process and tested blood components from clinical cases of vCJD held at the UK CJD Resource Centre.¹

References

 Cooper JK, Ladhani K, Minor P. Comparison of candidate vCJD in vitro diagnostic assays using identical sample sets. Vox Sang 2012; 102:100-9; PMID:22126309; http://dx.doi.org/10.1111/j.1423-0410.2011.01525.x

P.225: A newly developed assay for the presymptomatic detection of prions in blood

Daisy Bougard,¹ Maxime Belondrade,¹ Christiane Segarra,¹ Vincent Béringue,² and Joliette Coste¹

¹Laboratoire TransDiag, Sécurité Transfusionnelle et Innovation Diagnostique, Etablissement Français du Sang Pyrénées Méditerranée; Montpellier, France; ²INRA UR892 Virologie Immunologie Moléculaires; Jouy-en-Josas, France

Introduction. Prion diseases or Transmissible Spongiform Encephalopathies (TSEs) are neurodegenerative diseases including variant Creutzfeldt-Jakob disease (vCJD) in humans. The central event of these diseases would be the conformational change of a normal cellular protein PrP^C into an infectious form PrP^{TSE}. It is now evident that TSEs are transmissible by blood transfusion and this has raised concerns that a reservoir of infectious asymptomatic people could exist in the blood donor population. A recent prevalence study based on stored lymphoreticular samples analyses led to en estimation of 1 in 2000 persons being potentially infected by vCJD prions in the United Kingdom. Until now, no screening test could detect the infectious agent in human blood before the onset of clinical signs of disease.

The objective of this study is to develop a sensitive and specific test that would enable the detection of PrP^{TSE} in the blood during the presymptomatic phase of TSE.

Materials and Methods. The detection assay comprises three major steps: (1) a ligand-coated bead pre-analytical step in order

to concentrate PrP^{TSE} from the different blood components and to remove inhibitory factors which can interfere with the amplification; (2) a PrP^{TSE} amplification by serial PMCA using transgenic mouse brain homogenate as substrate and (3) a specific detection of the amplified PrP^{TSE} by immuno-blotting after partial proteinase K digestion. The sample volume has been optimized for 500 µL of plasma and for 25 to 50 µL of buffy-coat. Whole blood samples from infected sheep collected during preclinical and clinical phases of scrapie were processed in buffy-coat, white blood cells (WBC) and plasma. Blood from humanised transgenic mice infected by vCJD was also tested in relation with the duration post inoculation.

Results. PMCA assay allowed detection of PrP^{TSE} in: (1) the WBC of four sheep at the acute phase of scrapie with a 100% sensitivity and specificity, (2) in the plasma and buffy coat collected in the asymptomatic phase of the disease (3) in the blood of vCJD-infected transgenic mice before the occurrence of clinical signs.

Conclusions. The expected level of sensitivity for the detection of prion in the blood was reached. This assay is currently evaluated as a confirmatory detection test for the presence of the vCJD agent in human blood. The next step will be to perform prevalence studies by analysing panels of at-risk populations.

P.226: Neurodegeneration and neurogenesis in the gastrointestinal tract of prion infected mice

Laura Ellett,¹ Amanda Quattrocelli,¹ Richard Schregle,¹ Ewan Chan,¹ Gene Venables,² John B Furness,² and <u>Victoria A Lawson¹</u>

¹Department of Pathology; The University of Melbourne, Parkville, VIC Australia; ²Department of Anatomy and Neuroscience; The University of Melbourne, Parkville, VIC Australia

The presence of disease associated PrP^{Sc} in the gastrointestinal tract of patients affected by prion diseases represents a source of disease transmission through surgical procedures and may contribute to symptoms affecting their quality of life.

Following intracerebral inoculation of wildtype mice with prions, PrP^{Sc} was detected in the gastrointestinal tract during symptomatic disease (20 weeks post infection). PrP^{Sc} accumulation in the ileum was associated with high levels of infectivity, caspase activation, loss of neuronal subpopulations in the myenteric plexus and the loss of enteric glial cell integrity.^{1, 2} In the current study a time course analysis indicated that PrP^{Sc} accumulation was detected in the ileum within 3 weeks of intracerebral inoculation, with highest levels detected 6-10 weeks post-inoculation, at which time mice demonstrated no neurological symptoms of disease. There was a quantitative decrease in PrP^{Sc} load 13 weeks post infection suggestive of a shedding of PrP^{Sc} from the gastrointestinal tract and a potential source of environmental contamination.

A significant loss of neurofilament (medium) immunoreactive neurons, reflective of intrinsic sensory neurons, was detected 10 weeks post inoculation and coincident with high PrP^{Sc} loads. This was followed by the apparent recovery of this neuronal population between weeks 13 and 17 post-inoculation and coincident with reduced PrP^{Sc} loads. These results suggest that the accumulation of PrP^{Sc} may directly contribute to the loss of this neuronal population and that a reduction in PrP^{Sc} burden enabled its regeneration.

We had previously observed the qualitative loss of neuronal nitric oxide synthase (nNOS) immunoreactive neurons, a marker of inhibitory motor neurons, in conjunction with regions of distorted glial fibrillary acidic protein (GFAP) immunoreactivity, indicative of enteric glial cell derangement at the terminal stages of disease. We now report a significant decrease in nNOS immunoreactive neurons between 10 and 17-week post inoculation and evidence of enteric glial cell derangement as early as 6 weeks post inoculation. These results support the view that glial cell dysfunction in the gastrointestinal tract of prion affected animals precedes and contributes to the loss of nNOS immunoreactive neurons.

This study has identified PrP^{sc} accumulation and glial cell dysfunction as mediators of prion induced neurodegeneration of specific neuronal subpopulations within the gastrointestinal tract and the potential for regeneration of neuronal populations following a reduction in PrP^{sc} load. The effect of neuronal loss on gastrointestinal dysfunction will be presented.

References

- Lawson VA, Furness JB, Klemm HM, Pontell L, Chan E, Hill AF, Chiocchetti R. The brain to gut pathway: a possible route of prion transmission. Gut 2010; 59:1643-51; PMID:21071583; http://dx.doi.org/10.1136/gut.2010.222620
- Drew SC, Haigh CL, Klemm HM, Masters CL, Collins SJ, Barnham KJ, Lawson VA. Optical imaging detects apoptosis in the brain and peripheral organs of prion-infected mice. J Neuropathol Exp Neurol. 2011; 70:143-50; PMID: 21343883; http://dx.doi. org/10.1097/NEN.0b013e3182084a8c.

P.227: Selective binding of high molecular mass assemblies of amyloid β-peptide to prion protein in patients with Alzheimer's disease

Frank Dohler,¹ Diego Sepulveda-Falla,¹ Susanne Krasemann,¹ Hermann Altmeppen,¹ Hartmut Schlueter,² Inga Zerr,³ Jakob Matschke,¹ and <u>Markus Glatzel</u>¹

¹Institute of Neuropathology, University Medical Centre Hamburg-Eppendorf; Hamburg, Germany, ²Mass Spectrometric Proteomics Group Institute of Clinical Chemistry, University Medical Centre Hamburg-Eppendorf; Hamburg, Germany; ³Department of Neurology, National TSE Reference Centre, Georg-August University Goettingen; Goettingen, Germany

In Alzheimer's disease the generation of oligomeric species of amyloid β -peptide is causal to disease initiation and progression. Oligomeric species of amyloid β -peptide bind to the N-terminus of plasma membrane-bound cellular prion protein (PrP^c). This binding may be associated to synaptic degeneration. Composition of bound amyloid β -peptide oligomers, binding domains within PrP^c and modifiers of this binding have mostly been studied in cell culture or murine models of Alzheimer's disease. Our