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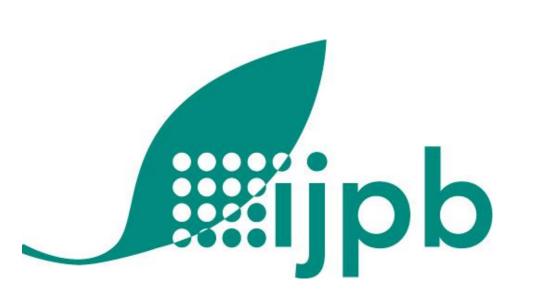
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Schematic organization of LD from yeast

(Buchanan *et al.*, 2000)

Potential partners of Ypt7p



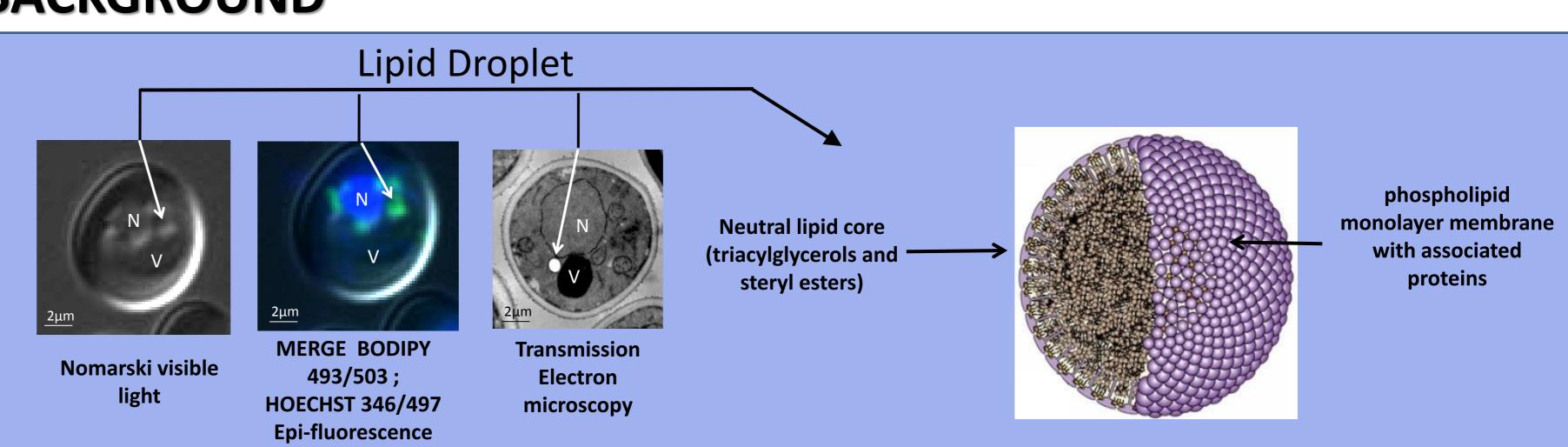
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BACKGROUND

In yeast, storage lipids are maintained in the cytoplasm in specialized organelles called lipid droplets (LDs) (Murphy et al., 2012). These structures consist mainly of a core of neutral lipids (triacylglycerols and/or steryl esters) enclosed in a monolayer of phospholipids. They contain a number of associated proteins, involved in many metabolisms, signaling and trafficking. Indeed, LD is now known to be a complex dynamic organelle. Many studies have reported interactions between LDs and intracellular organelles, like endoplasmic reticulum, early endosome, peroxisome and mitochondria (Zehmer et al., 2009). We have previously shown, in *S. cerevisiae*, the relationship between vacuole and LDs, and described first evidences of the involvement of the Rab GTpase Yp7p in the dynamics of LDs.



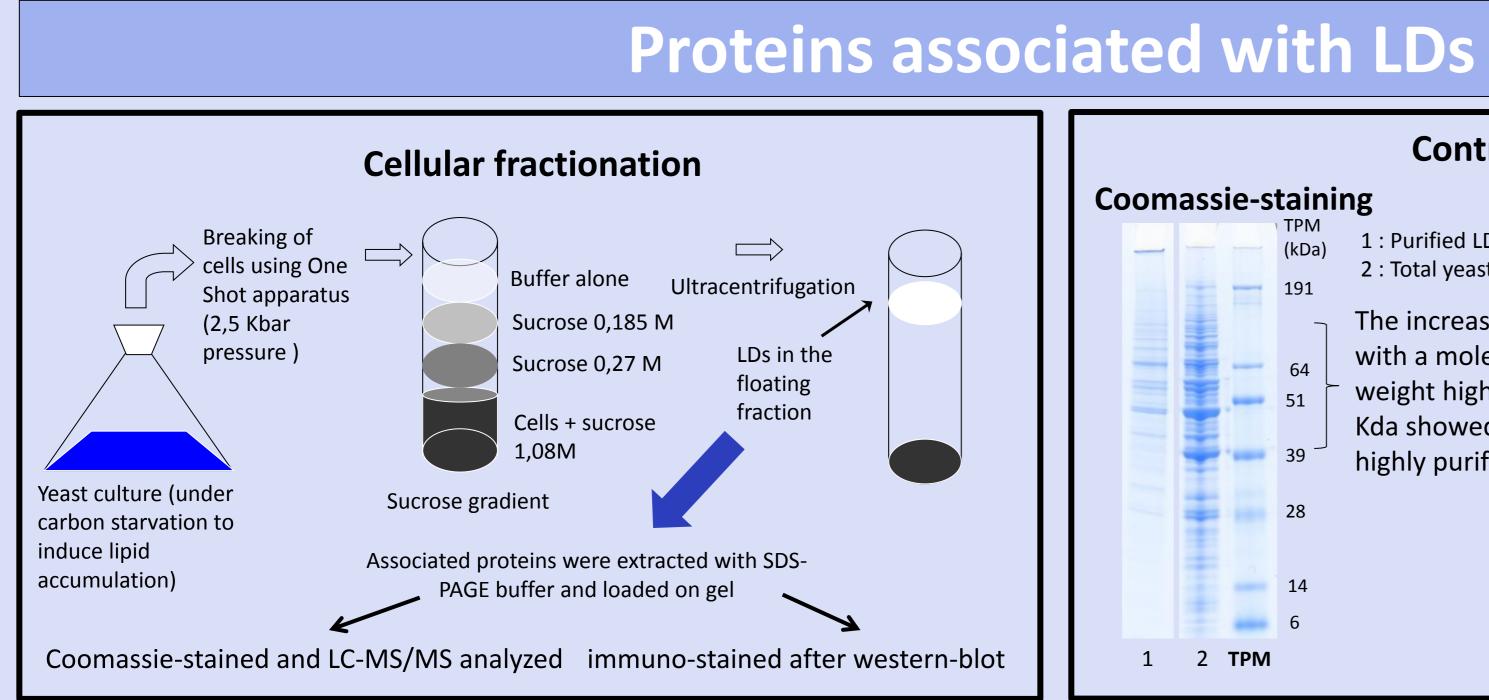
OBJECTIVES

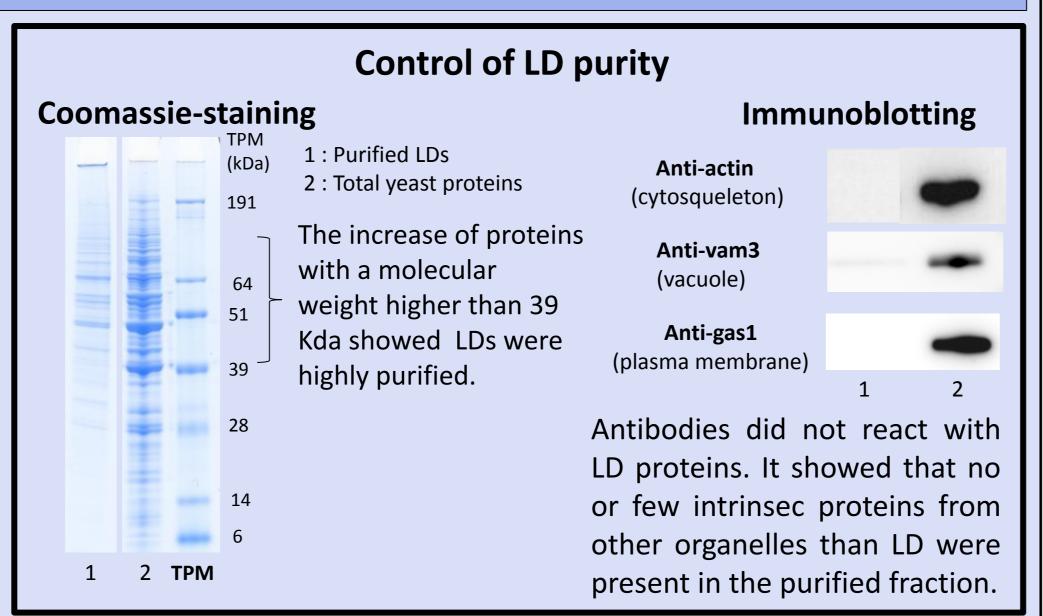
Microscopic observations of LDs from *S*.

cerevisiae (BY4741); V: vacuole; N: Nucleus

The aim of this study was to investigate the involvement of Ypt7p in LD dynamics by identifying its potential partners on LDs and by exploring the trafficking pathways which could be involved in these dynamics.

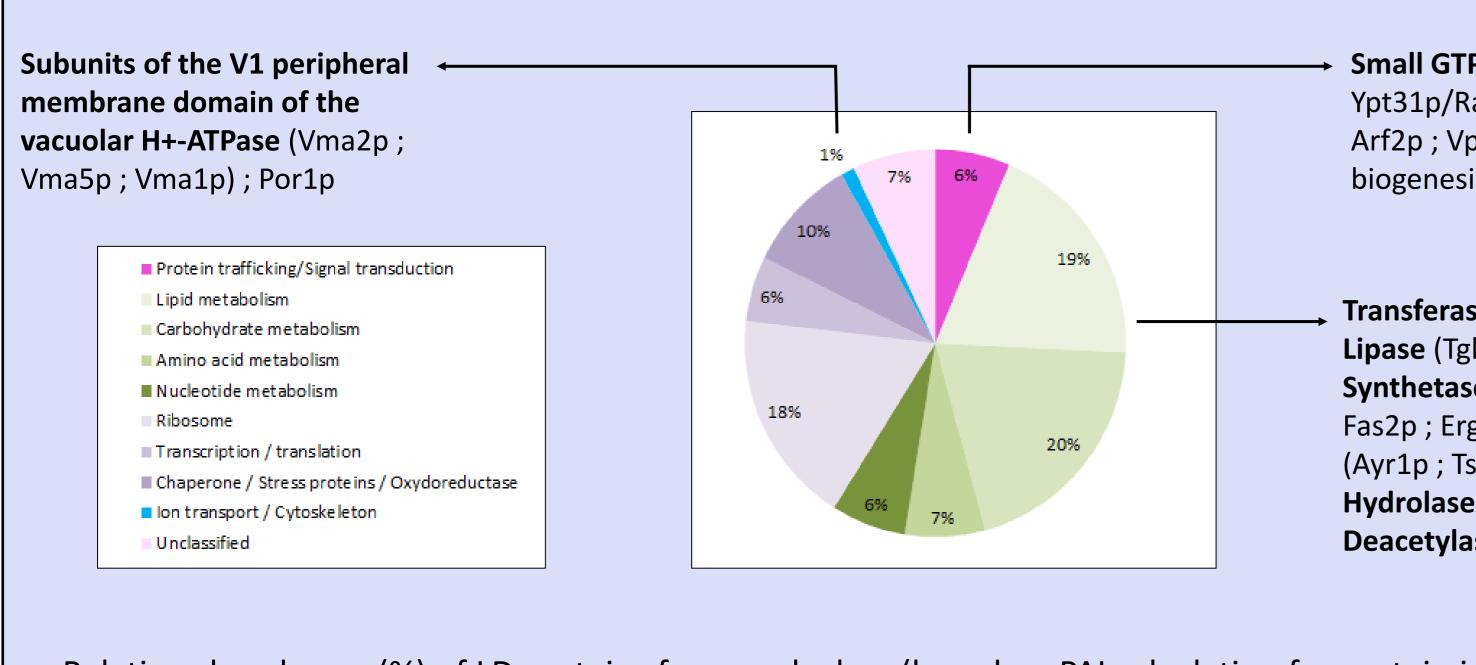
RESULTS





Proteomic of purified LDs from WT *S. cerevisiae*

151 LD-associated proteins were classified according to their functionality (SGB database : http://www.yeastgenome.org). Many metabolism enzymes, especially lipid metabolism, were found showing important biological processes were carried at its surface. Several membrane-trafficking proteins were also identified showing that these proteins were functionally associated with LDs.



Small GTPase Rab Family (Ypt7p/Rab7; Ypt1p/Rab1; Ypt31p/Rab11 ; Sec4p/Rab8) ; GTPase family (Arf1p ; Arf2p; Vps1p, Sar1p); Nus1p, Gvp36p (vacuole biogenesis); Rer2p (ER protein sorting)

Transferase (erg6p; Slc1p; Loa1p; Pdr16p; Eht1p); **Lipase** (Tgl1p; Tgl3p; Tgl4p; Tgl5p; Pgc1p; Yju3p); Synthetase (Faa4p; Faa1p; Faa3p; Fat1p; Fas1p; Fas2p; Erg7p; Erg20p); Esterase (Tgl1p); Reductase (Ayr1p; Tsc10p; Erg27p); **Epoxidase** (Erg1p); **Hydrolase** (Yeh1p); **Deshydrogenase** (Hfd1p); **Deacetylase** (Say1p)

Relative abundance (%) of LD proteins from each class (based on PAI calculation for protein identified at least by 2 mass-spectra in LC-MS/MS analysis). Yeasts were at the exponential state.

SDS-PAGE Co-immunoprecipitation Anti-GFP Silver nitrate SDS-PAGE of eluted partners and LC/MS-MS analysis of selected band + LDs from GFP-YPT7 strain (a) (1) GFPm (*) from assay (1) Elution compared to controls (2, + soluble extract from GFP strain (2) partners of GFP and 3, unspecific linkage on + LDs from WT strain (3) agarose) Δ : GFP-Ypt7p • : Soluble GFP 1 2 3 1 2 3 Selected list of potential Ypt7p partners identified by LC/MS-MS **Potential Function Main localization** partners BAR-domain protein; Lipid binding; Eisosome The identification Endocytose Gdi1p, a Unknown LD known partner of Pet10p Ypt7p, validated Subunit H of the V1 peripheral membrane Vacuole, mb the experiment. domain of the vacuolar H+-ATPase Ergosterol biosynthesis Golgi, cytoplasm **GDP** dissociating inhibitor; known partner **Secretory pathway** Functional validation with mutants Staining of neutral lipids with Bodipy 493/503 Fatty-acid (FA) analysis **Epi-fluorescence** of mutant strains of selected Ypt7p partners % of variation of total FA / WT (GC) kes1∆ +6% WT $vma13\Delta$ -6% lsp1∆ pet10∆ -8% vma13∆ Strain vma13<u>⊿</u> + 25 % Fluorescent signal 310 +/- 30.2 187 +/- 3.5 (485/518) recorded using (+65%) Fluoroskan Average number of LDs per Vma13p, good 6.36 +/- 2.57 cell (EPI-fluorescence 3.71 +/- 1.69 (+71%)* candidate? observation) * : pvalue < 0,05 calculated on 173 cells using student test

CONCLUSION and ACKNOWLEDGMENT

The proteome of purified LDs from WT S. cerevisiae showed several membrane-trafficking proteins, whom Ypt7p, and many subunits of the V1 peripheral membrane domain of the vacuolar H+ ATPase. One other V1 subunit of this pump, Vma13p, was identified by Co-IP as a potential partner of Ypt7p. Its deletion had a great effect on LD dynamics. As its deletion is also known to abolish the homeostasis and the acidification of the vacuole, the effect on LDs could be a consequence of these dysfunctionments. Anyway, this showed a tight link between LD dynamics and vacuole. Thus, we hypothesized that accumulation of LDs in the cytosol of VMA13 mutants could be due to modifications of interactions between LDs and vacuole, involving Ypt7p and Vma13p, in the context of defects in vacuole-linked homeostasis. This remains to be confirmed by other interacting tools, like yeast double-hybrid.

We also showed that deletion of subunits of the HOPS complex lead to accumulation of LDs in the cells. Thus, this showed this tethering complex, known to be involved in fusion processes, is involved in LD dynamics.

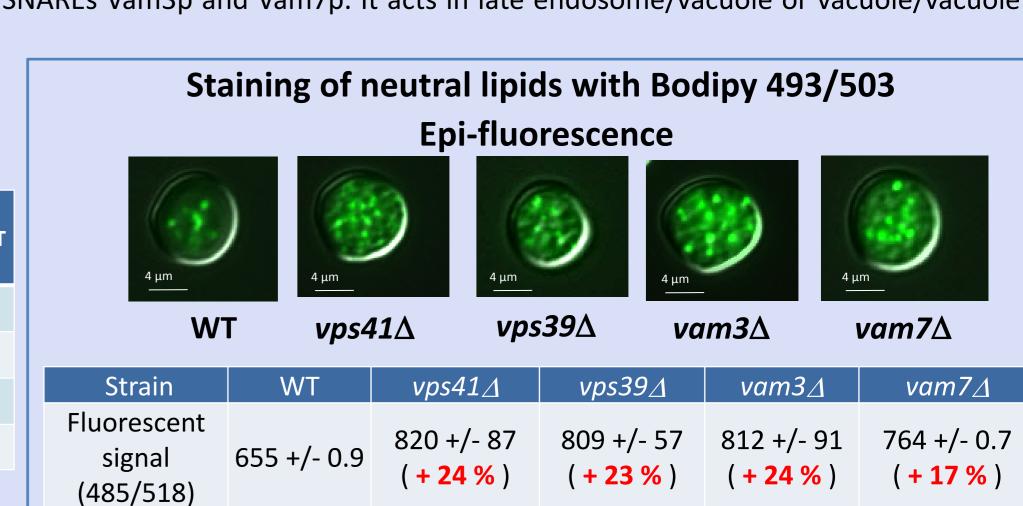
Altogether, these results lead us to hypothesize that LDs could interact with vacuole, via a hemi-fusion mechanism involving Ypt7p, as Rab effector, the HOPS complex and the V-ATPase.

We greatly acknowledge C. Ungermann (a) for the gift of the GFP-MYC-YPT7 strain, H. Riezman (Univ basel, Biozentrum, CH-4056 Basel, Switzerland) for the gift of Anti-Gas1 antibodies and A. Guillot, from PAPPSO (Jouy-en-Josas, France), for the quality of the LC-MS/MS analysis.

Is HOPS complex involved in LD dynamics?

Ypt7p is the Rab effector of the multisubunit tethering HOPS complex, which involves specifically Vps41p, Vps39p and the interacting SNAREs Vam3p and Vam7p. It acts in late endosome/vacuole or vacuole/vacuole membrane fusion.

FA analysis of mutant strains of HOPS pathway % of variation of total FA / WT Strain (GC) vps41∆ + 10 % $vps39\Delta$ + 27 % + 30 % $vam3\Delta$ $vam7\Delta$ + 19 %



Vma13p: Potential partner of Ypt7p, involves in LD dynamics

HOPS complex involves in LD dynamics