



HAL
open science

Transmissible Spongiform Encephalopathies rapid post mortem tests in goats: Strategies to improve the efficiency of the active surveillance statutory programme

Daniela Meloni, Elena Bozzetta, Jan P. M. Langeveld, Martin H. Groschup, Wilfred Goldmann, Olivier Andréoletti, Isabelle Lantier, Lucien J. M. van Keulen, Alex Bossers, Elsa Manzardo, et al.

► To cite this version:

Daniela Meloni, Elena Bozzetta, Jan P. M. Langeveld, Martin H. Groschup, Wilfred Goldmann, et al.. Transmissible Spongiform Encephalopathies rapid post mortem tests in goats: Strategies to improve the efficiency of the active surveillance statutory programme. International Prion Congress 2014, May 2014, Trieste, Italy. Landes Bioscience, Prion, 8 (S1), 2014, 10.4161/pri.29370 . hal-02744039

HAL Id: hal-02744039

<https://hal.inrae.fr/hal-02744039>

Submitted on 3 Jun 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

This article was downloaded by: [138.102.135.242]

On: 12 March 2015, At: 05:20

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Prion

Publication details, including instructions for authors and subscription information:
<http://www.tandfonline.com/loi/kprn20>

Prion Diseases in Animals

Published online: 01 Apr 2014.



[Click for updates](#)

To cite this article: (2014) Prion Diseases in Animals, Prion, 8:sup1, 59-109, DOI: [10.4161/pri.29370](https://doi.org/10.4161/pri.29370)

To link to this article: <http://dx.doi.org/10.4161/pri.29370>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Versions of published Taylor & Francis and Routledge Open articles and Taylor & Francis and Routledge Open Select articles posted to institutional or subject repositories or any other third-party website are without warranty from Taylor & Francis of any kind, either expressed or implied, including, but not limited to, warranties of merchantability, fitness for a particular purpose, or non-infringement. Any opinions and views expressed in this article are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor & Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

It is essential that you check the license status of any given Open and Open Select article to confirm conditions of access and use.

ancillary target for scrapie selection. Moreover, our findings support the idea that AGER is involved in the mechanism of neuronal dysfunction associated with prion diseases.

P.105: Transmissible Spongiform Encephalopathies rapid post mortem tests in goats: Strategies to improve the efficiency of the active surveillance statutory programme

Daniela Meloni,¹ Elena Bozzetta,¹ Jan PM Langeveld,² Martin H Groschup,³ Wilfred Goldmann,⁴ Olivier Andréoletti,⁵ Isabelle Lantier,⁶ Lucien JM Van Keulen,² Alex Bossers,² Elsa Manzardo,¹ Maria C Cavarretta,¹ Daniela Loprevite,¹ Maria Gabriella Perrotta,⁷ Danilo Pitardi,¹ Francesco Ingravalle,¹ Simone Peletto,¹ Silvia Colussi,¹ and Pier Luigi Acutis¹

¹Istituto Zooprofilattico Sperimentale del Piemonte Liguria e Valle d'Aosta; Turin, Italy; ²Central Veterinary Institute of Wageningen UR; Lelystad, The Netherlands; ³Friedrich-Loeffler Institut, Federal Research Institute for Animal Health; Insel Riems, Germany; ⁴Roslin Institute and R(D)SVS University of Edinburgh; Roslin, Midlothian, UK; ⁵UMR INRA ENVT 1225 Interactions Hotes Agents Pathogènes, ENVT; Toulouse, France; ⁶INRA IASP, Centre INRA de Tours; Nouzilly, France; ⁷Ministero della Salute; Rome, Italy

Annex X to Regulation (EC) No 999/2001 lists the approved rapid post mortem tests which may be used within the framework of the EU monitoring programmes. Little is known regarding the efficiency of different rapid methods on goats. The authors compared the performance of the *IDEXX HerdChek*[®] BSE-scrapie, the *Bio-Rad*[®] *TeSeE*TM *SAP* and the *Bio-Rad*[®] *TeSeE*TM *Sheep/Goat* tests over 96 central nervous system goat tissues including 20 experimentally obtained BSE positives, 41 scrapie positives and 35 negative samples prepared following the European reference laboratory standard method of homogenate preparation (50% w/v protocol). The relation between age, prion type, PNRP genotype and test performance were further investigated. Clear differences in relative diagnostic sensitivity were shown, being the *IDEXX HerdChek*[®] BSE-scrapie the best performing system setting its 95% IC sensitivity at 100%. Interestingly, *BioRad*[®] *TeSeE*TM *Sheep/Goat* test showed a 95% IC sensitivity of 85% on BSE positive samples and an optimistic 82.9% on scrapie ones (considering the only “suspect” result as positive), while *BioRad*[®] *TeSeE*TM *SAP* reached only a 75% sensitivity on BSE samples but a 90.2% on scrapie positives. The reproducibility was not absolute as some discrepant results in any combination of tests was shown, nevertheless, the lower limit of the 95% CI was always above 0.78 considering the samples as a whole, and always above 0.74 when the stratification by PrP^{Sc} profile was applied, values consistent with a substantial-good-fair to good agreement. All the tests correctly identified the 35 negative samples, showing a 100% specificity. Our findings on BSE samples do not suggest any contribution of the original inocula (bovine or goat BSE) on the test results whereas the mean age of the natural scrapie positive animals set at 70 months, confirming the hypothesis that the

sensitivity of detection using brainstem appeared to be dependent on the age of tested individuals. The studies on the correlation between rapid test performance and genotype is ongoing. Considering the current lack of genetic selection for eradicating/controlling classical goat scrapie at population level, the efficiency of the surveillance in place would be crucially dependent on the system used to identify infected flocks.

Acknowledgments. Funded by EU project “goatBSE” FOOD-CT-2006-36353 and Italian Ministry of Health Project 2005 (IZSPLV 14/05 RC).

P.106: In vitro generated mouse prion protein causes brain neurodegeneration in FVB/N female mice

Seyed Ali Goldansaz,¹ Dagnachew Hailemariam,¹ Nathalie Daude,² David Wishart,³ and Burim N Ametaj¹

¹Department of Agricultural, Food & Nutritional Science; University of Alberta; Edmonton, AB Canada; ²Centre for Prions and Protein Folding Diseases; University of Alberta; Edmonton, AB Canada; ³Departments of Biological Sciences and Computing Science, University of Alberta; Edmonton, AB Canada

Recently we reported that lipopolysaccharide (LPS) from *Escherichia coli* 0111:B4 was able to instantly convert mouse prion protein into a beta-rich isoform (moPrP^B) resistant to proteinase K digestion under normal physiological conditions. In this study we tested whether subcutaneously (sc) administered moPrP^B (29-232) is able to cause prion-like pathology in vivo. Six groups of 15 FVB/N female mice each were treated sc with: (1) Saline, (2) LPS (0.1 µg/g of body weight), (3) moPrP^B (45 µg/mice), (4) LPS+moPrP^B, (5) RML (Rocky Mountain Laboratory scrapie prions at 10⁷ ID units), and (6) LPS+RML. Saline and LPS were administered over a period of 6wk using ALZET[®] osmotic minipumps (ALZET, Cupertino, CA) implanted sc, whereas moPrP^B and RML were administered as one time sc injection at the beginning of LPS infusion. Five animals from each treatment group were euthanized at 11wk post-inoculation (pi), with no clinical signs of prion disease. The rest of the mice were left to develop clinical signs associated with prion disease until terminal sickness. Hematoxylin & eosin (H&E) and PrP^{Sc}-stainings were conducted from brain tissues to determine presence of vacuolation and PrP^{Sc} accumulation, respectively. Western blot (WB) analysis and scrapie cell assay (SCA) using L929 cells were also conducted to evaluate presence of resistant PrP (PrP-res). All treatment groups, except for saline, showed mild brain vacuolation at 11wk pi in the cerebral cortex (Cc), thalamus (Th), midbrain (Mb), and cerebellum (Cr) and mild PrP^{Sc} accumulation only in the LPS+RML treatment. Terminally sick mice exhibiting clinical signs of prion disease from the LPS, moPrP^B, and LPS+moPrP^B treated groups showed widespread brain neurodegeneration in the Cc, Th, Mb, and Cr comparable to positive controls. Computer evaluated vacuolation of terminally sick mice showed numerically larger size vacuoles in the Cr and Mb brain regions of LPS, moPrP^B, LPS+PrP^B, and LPS+RML treated mice