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EVALUATION OF A NEW BIOLOGICAL PEST CONTROL METHOD AGAINST THE PALM BORER, PAYSANDISA ARCHON USING OOPHAGUS PARASITOIDS

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SUMMARY

As part of the European project Palm Pprotect (2012-2014, 3 million \in), which aims to eradicate the two palm pests: *Rhynchophorus ferrugineus* and *Paysandisia archon*, the Mediterranean Forest and Entomology Experimental Unit of INRA PACA intends to develop a new biological pest control method against the palm borer *P. archon*. The egg stage of this moth is the most accessible. On one hand, a screening of several potentially effective parasitoids of the genus *Trichogramma* will be performed against the moth. On the other hand, collects of *P. archon* eggs will be conducted in order to observe if parasitoids, naturally present in the environment, would have succeeded in establishing themselves, particularly through the importation of palm trees in the Mediterranean region. The first results are reported and discussed.

Key words: Trichogramma, Paysandisia archon, biological pest control, oophagous parasitoids, palm tree.

RESUME

En tant que partenaire du projet européen Palm Protect (2012-2014, 3 millions d'euros) qui vise à éradiquer les deux ravageurs des palmiers *Rhynchophorus ferrugineus* et *Paysandisia archon*, l'unité Expérimentale Entomologie et Forêt Méditerranéenne INRA PACA propose de développer une nouvelle méthode biologique de contrôle contre le papillon palmivore *P. archon*. Les œufs sont le stade le plus accessible. D'une part, un criblage de plusieurs parasitoïdes potentiels du genre *Trichogramma* sera réalisé. D'autre part, une collecte d'oeufs de *P. archon* sera effectuée pour observer si des parasitoïdes naturellement présents dans le milieu ambiant pourraient réussir à s'installer, en particulier via l'importation de palmiers de la région méditerranéenne. Les premiers résultats sont décrits et discutés.

Mots clés : *Trichogramma*, *Paysandisia archon*, lutte biologique, parasitoïdes oviphages, palmier.

INTRODUCTION

Since 2009, the palm borer *Paysandisia archon* (Burmeister, 1880) is classified as a quarantine pest in France (Huguet and Fouilleux, 2010). Native from South America (Argentina, Brazil, Paraguay, Uruguay), it was first reported in France in the regions of Hyères and Toulon and would have been imported via palm trees (*Trithrinax campestris*) from Argentina in the 1990s (Montagud Alario, 2004). *P. archon* larvae cause significant damages on palm trees by feeding on the stipe. Moreover, the pest is able to attack different palm species as *Chamaerops humilis, Livistona chinensis, L. decipiens, Sabal spp., Phoenix canariensis, P. dactylifera, Trachycarpus fortunei* and *Washingtonia filifera* (Sarto i Monteys, 2005).

Human activities have greatly contributed to the expanding of the pest. In France, the moth has been observed for the first time in the Var region. Now, it can be seen throughout South of France from the "Alpes-Maritimes" to the "Pyrénées-Orientales". It is also found in many European countries as Italy, Spain, Greece, Slovenia, Switzerland and Cyprus (EPPO, 2011).

The lack of controls and vigilance during trade allowed the moth to gain ground. Furthermore, it appears that *P. archon* can travel large distances by flying. Its biology makes detection and control difficult: larvae remain endophagous throughout their life, the adults do not feed and are active during the day, females do not secrete long range pheromones (Sarto i Monteys *et al.*, 2012), making attractant-based trapping presently impossible. A new research is in progress (CNRS) concerning pheromones secretion by the male butterflies.

Mechanical control methods such as bagging or the use of birdlime have been advised as well as the use of chemicals. Regarding biological pest control, tests were performed and means established. The use of nematodes *Steinernema carpocapsae* as biopesticide, or products based on the entomopathogenic fungi *Beauveria bassiana*, are known to control the larvae. To date, no attempt of control using parasitoids or predators have been introduced.

In Argentina, natural enemies such as birds or *Ichneumonidae* parasites are likely controlling the pest (Sarto i Monteys and Aguilar, 2005).

The INRA PACA MFEU aims to find new natural enemies, particularly oophagous parasitoids.

Tests were set up and collects carried out with two distinct objectives:

 \rightarrow In the laboratory, a screening of different *Trichogramma* strains was performed. Strains with physiological and morphological benefits were previously selected. The strains were then raised for several generations on hosts laying bigger eggs than the host *Ephestia kuehniella* usually used. Trichogramma produced on big eggs were then tested on *P. archon* eggs.

 \rightarrow In the field, collects of *P. archon* eggs have started in order to look for native oophagous parasitoids which would naturally parasitized eggs of this butterfly. The oldest sites attacked will be preferred. The aim is to raise those parasitoids on large lepidopteran eggs and then, to study their potential parasitism on *P. archon* eggs

This article informs the first results obtained by INRA PACA during the first year of Palm Protect program.

MATERIAL AND METHODS

REARING OF TRICHOGRAMMA

Selected strains

Nineteen strains of trichogramma belonging to 9 species and taken from the INRA PACA collection (Figure 1) were selected on biological criteria that could facilitate their potential on big eggs of *P. archon*:

- Strains fertility, or their ability to parasitize a large number of eggs.

- Originated host from which the strains have been collected before rearing on *E. kuehniella* eggs. Strains from biggest host eggs were selected.

- Others benefits such as large ovipositor (which could help to penetrate chorion of big eggs more easily), or the ability to parasitize on high vegetation layers, have also been taken into account.

The different selected strains are shown in Table 1. Those

Picture : Triaden Picture : Trichogramma

Figure 1 : Trichogramma collection of INRA PACA Collection de trichogrammes à l'INRA PACA

strains, initially reared on *Ephestia kuehniella* eggs, were reared during several generations on *Philosamia ricini* (Drury) (Lep., *Saturniidae*), and *Bombyx mori* (L.) (Lep., *Bombycidae*) eggs, in order to accustom the parasitoids on larger eggs and stronger chorion.

Table 1: Species / strains of trichogramma initially selected.Espèces / souches de trichogrammes pré-sélectionnées

Specie / strain	Author
Trichogramma chilonis SB	Ishii
<i>Trichogramma chilonis</i> Th	Ishii
Trichogramma chilonis Ca	Ishii
Trichogramma chilonis P	Ishii
Trichogramma chilonis C	Ishii
Trichogramma cordubensis Pt	Vargas & Cabello
Trichogramma cordubensis Et	Vargas & Cabello
Trichogramma cordubensis Ct	Vargas & Cabello
Trichogramma pretiosum U	Riley
Trichogramma pretiosum HA	Riley
Trichogramma pretiosum HB	Riley
Trichogramma dendrolimi I	Matsumura
Trichogramma dendrolimi L	Matsumura
Trichogramma buesi C	Voegelé
Trichogramma exiguans U	Pinto & Platner
<i>Trichogramma embryophagum</i> It	Hartig
Trichogramma embryophagum I	Hartig
Trichogramma euproctidis A	Girault
Trichogramma daumaleae C	Dugast & Voegelé

Tests on alternative new host P. ricini and B. mori

The rearing of *Philosamia ricini* (Figure 2) was undertaken, in order to provide big eggs necessary for trichogramma rearing. The first larvae were ordered at the OPIE association. The breeding was then maintained on privet leaves over several generations, in order to have available adults and eggs for maintaining trichogramma rearing. Tubes containing 60 eggs of *P. ricini* for about 30 trichogramma were used. The eggs renewal was performed

by migration of the emerging trichogramma in a new tube (7.5 \times 1.2 cm) with honey, for each generation of each



Figure 2 : Philosamia ricini

strain. In the same way, a *B. mori* rearing was undertaken and the selected trichogramma were bred on those eggs.

Tests on the pest P. archon

Strains with the best parasitism on big eggs were tested on *Paysandisia archon* eggs in tubes and fed with honey. The pest eggs were regularly provided by our partner CIRAD of Montpellier. Several tests have been done following different modalities showed in Figure 3.

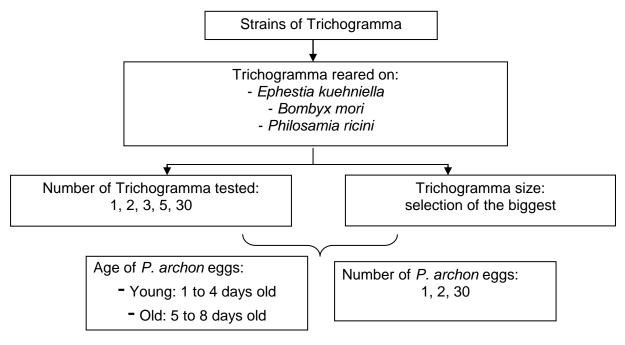


Figure 3 : Modalities of tests on *P. archon* eggs.

In a first time, emergence of trichogramma from *P. archon* eggs proposed to the parasitoids for oviposition was then examined.

COLLECTS OF OOPHAGUS PARASITOIDS

First collects on the field were done. The aim was to let incubate the collected eggs until emergence of any oophagous parasitoids. Parasitoids will be then grouped by type and then reared on large eggs.

FIRST RESULTS

TESTS ON BIG EGGS

Rearing results on Philosamia ricini eggs

Concerning trichogramma rearing on *P. ricini* eggs, the different strains initially selected all have been raised on this host over several generations. The Trichogramma size has significantly increased in the first generation.

Rearing results on Bombyx mori eggs

Breeding of the different trichogramma strains on *B. mori* eggs was also a success. However, although the size of trichogramma from these eggs is greater than those reared on *E. kuehniella*, they remain significantly smaller than those produced on P. ricini eggs.

Screening on Paysandisia archon eggs

To date, no emergence of trichogramma could have been observed on eggs of *P. archon*. The eggs have hatched, giving a larva, or aborted. Whatever the host, trichogramma females showed no real interest in the moth eggs.

COLLECTS

First results of collects for *P. archon* eggs evidenced difficulty of finding eggs *in situ*. Other collects are nevertheless considered in areas where *P. archon* attacks are denser.

DISCUSSION

Trichogramma are widely used in biological pest control around the world. They are rather generalist parasites capable of parasiting many host eggs with variable shapes and sizes. Indeed, trichogramma are observable on a large number of moth family, they can be found on *Sphingidae, Noctuidae, Tortricidae, Saturniidae*, etc. depending on the species of trichogramma (Consoli, 2010).

The first step of our protocol, which covers trichogramma rearing on *P. ricini* and *B. mori* eggs showed positive results. These parasitoids were able to develop over several generations on these hosts, with a larger size. Females were found interested in these eggs from their first contact with them. In addition, laboratory production has supply fresh eggs that are more attractive for trichogramma.

Unfortunately, in this study, we were unable to demonstrate their ability to parasitize *P. archon* eggs. Several factors come into play in the choice of oviposition and can cause a rejection from the female:

- egg size,
- egg resistance to the ovipositor,
- chemical factors,

- age of the egg: a too old egg reduces the attractiveness of trichogramma and decreases their success of parasitism.

- secretion of toxins (Consoli, 2010).

P. archon eggs have a very characteristic shape (rice grain) which differs greatly from the original host of rearing. Moreover, they are very large (about 1.5 mm wide and 4.5 mm long).

Also, the selected alternative hosts for rearing was perhaps not the better choice. Additional assays have to be realized on others lepidopteran host families. Indeed, trichogramma gain experience at the larval stage regarding their host preference. This preference goes toward a host from which they originate (Consoli, 2010) and as there is no native Castniidae in France, phylogenetically or physiologically close families will be used for our next experiments. These assumptions could be the cause of lack of interest of tested trichogramma toward *P. archon* eggs.

CONCLUSION AND PERSPECTIVES

To date, parasitism tests on *P. archon* eggs using different trichogramma strains have not yielded positive results. However, the interest to continue the breeding of these parasitoids on other major lepidopteran hosts before testing on *P. archon* eggs remains. Families potentially close of Castniidae will be preferred. An European trichogramma specie (*Trichogramma gicai*), naturally more robust than those we usually harvest will be sought from the spring of 2013.

A new campaign for collecting *P. archon* eggs in heavy infested areas is already planned. Similarly, *P. archon* eggs will be placed in different climate and environmental zones, chosen in order to trap any native oophagous parasitoids potentially adapted to this pest eggs. Thus, many new avenues are still developing in the field of oophagous parasitoids.

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