

The muscle tissue and its relationship to beef production Isabelle Cassar-Malek

▶ To cite this version:

Isabelle Cassar-Malek. The muscle tissue and its relationship to beef production. Master. Master Science des Aliments - Mention Nutrition, Santé, Aliments, Semestre 3 (UE3 - Biologie intégrée et physiologie des muscles), France. 2018, 57 p. hal-02785482

HAL Id: hal-02785482 https://hal.inrae.fr/hal-02785482

Submitted on 16 Jul2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

The muscle tissue and its relationship to beef production



Isabelle Cassar-Malek

isabelle.cassar-malek@inra.fr

Inra UMR Herbivores 63122 Saint-Genès-Champanelle







Myogenesis of Cattle - Challenges







Beef production "Construction" of beef quality

<u>A model for humans</u> (Gibbs and Weinstock , 2002)





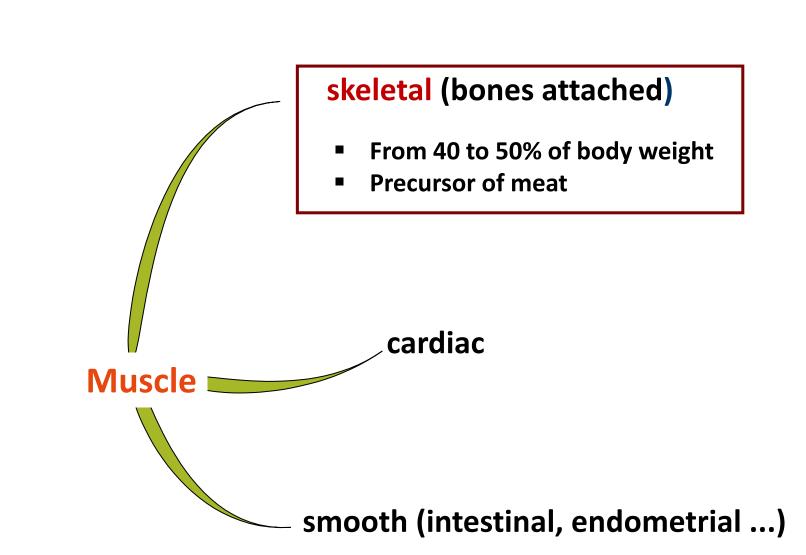
The word "muscle" comes from the Latin mus / musculus meaning "little mouse"



Terminology

- Myo = muscle
 - (myofibril, myoblast, myogenesis, myotome)
- <u>Sarco</u> = flesh (sarcolemma)

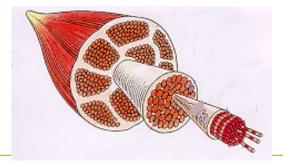
Muscle tissues



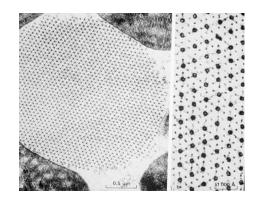
Skeletal muscle

- movement (locomotion, manipulation): voluntary
- posture and body position
- stability of the joints (tendons)
- reserve of proteins
- role in the oxidation of nutrients
- maintenance of body <u>temperature</u> (85% body heat, chill)

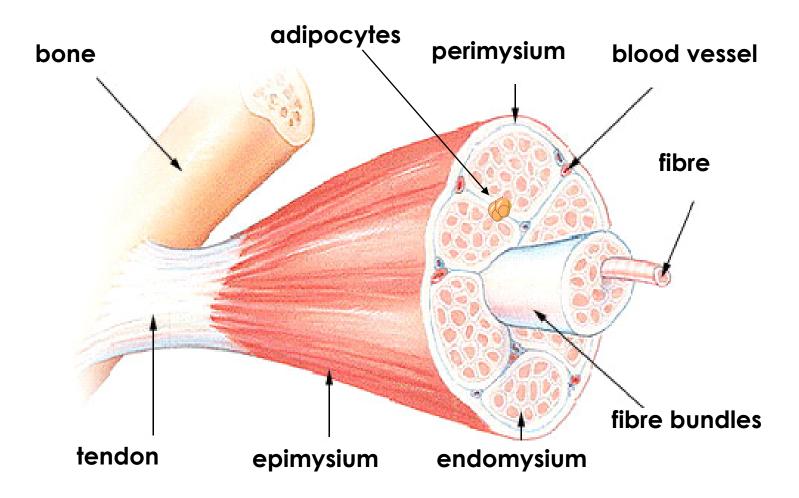
STRUCTURE OF THE SKELETAL MUSCLE



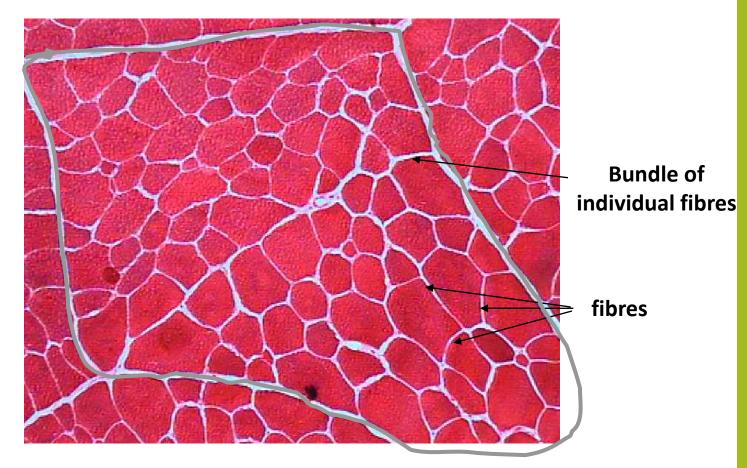




Macroscopic structure



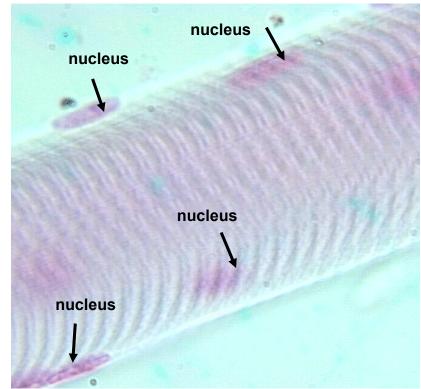
Microscopic structure



Histological section (bovine muscle)

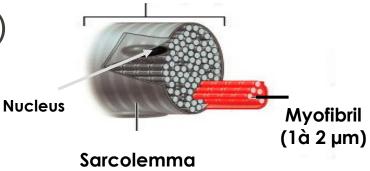
Muscle fibres

- <u>differentiated</u> cells
- From 75 to 90% of muscle volume
- cylindrical
- unbranched
- <u>multinucleated</u>
- length: up to 60 cm
- diameter: 10 to 100 microns
- striated

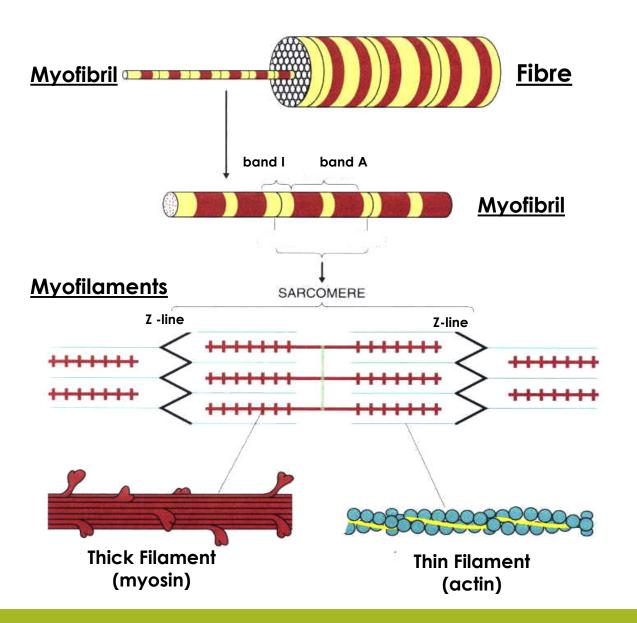


Cellular organisation

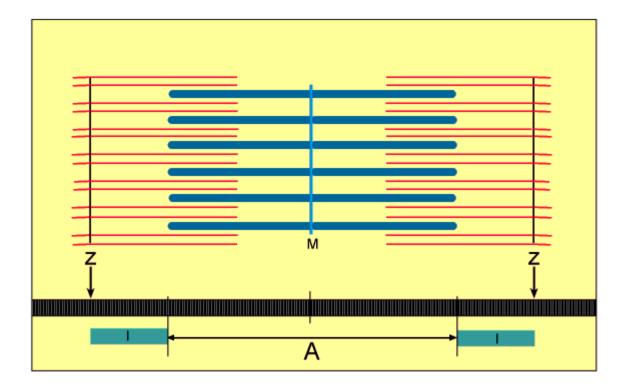
- membrane (sarcolemma)
- cytoplasm (<u>sarcoplasm</u>): incl. glycogen, myoglobin
- post-mitotic <u>nuclei</u>
- mitochondria
- endoplasmic <u>reticulum</u>: differentiated (<u>sarcoplasmic</u>)
- cytoskeleton (myofibrils)



Ultra-structure



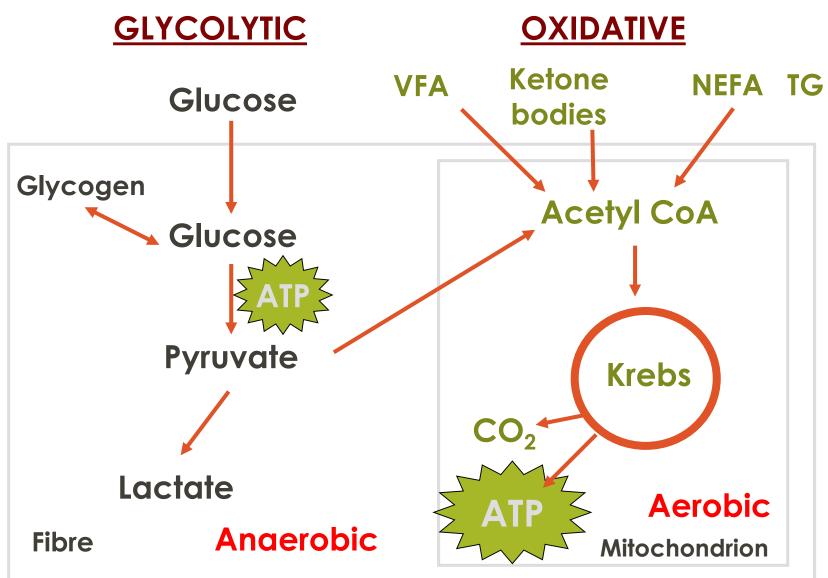
The sarcomer

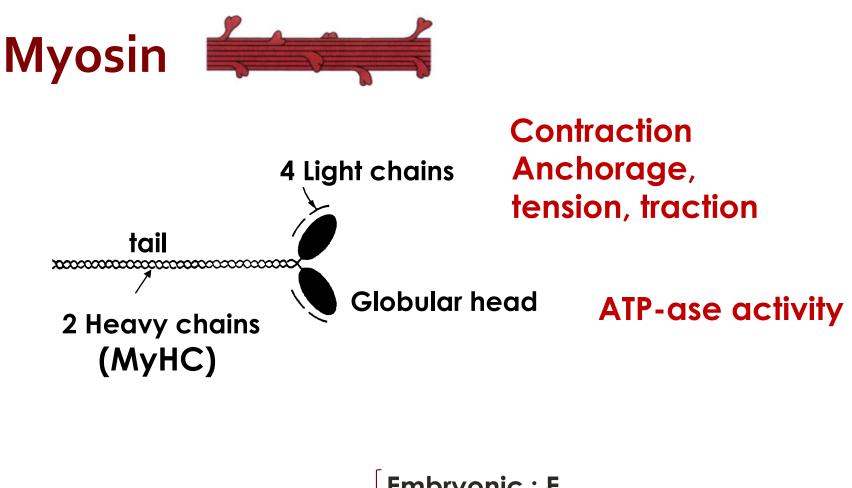


13

CONTRACTILE AND METABOLIC PROPERTIES

Metabolic pathways





<u>Several isoforms</u>

Embryonic : E Neonatal or fetal : F Slow: I Fast: IIa, IIb, IIx

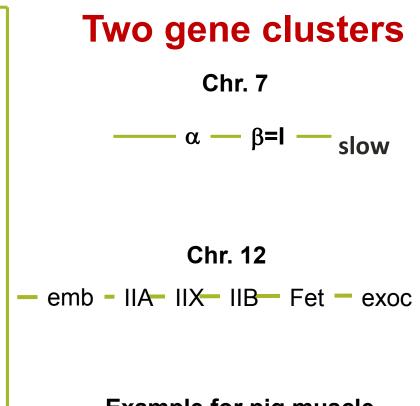
MyHC isoforms

« Adult » isoforms

I (slow) IIA IIX (fast) IIB

Developmental isoforms

Embryonic Fetal α-cardiac Extra occular (exoc)



Example for pig muscle

Fibre types

| CLASSIFICATION | 1 | IIA | IIX |
|----------------------|-----------|------------------|------------|
| Speed of contraction | Slow | Fast | Fast |
| Metabolism | Oxidative | Oxido-glycolytic | Glycolytic |
| Fatigue resistance | High | High | Low |
| Glycogen Content | Low | High | High |
| Lipid Content | High | High | Low |
| Vascularization | High | High | Low |

| ₹ | I/IIA 🔁 | | IIX |
|---|---------|--------|-----|
| | Hybrid | Hybrid | |

Fibres types / physical activity



Sprint 너

- Fast IIX fibres
- using glycogen





Jogging, ski, distance runner ⇒

- Slow type I fibres
- using lipids

Alternating sprint/endurance ⇒

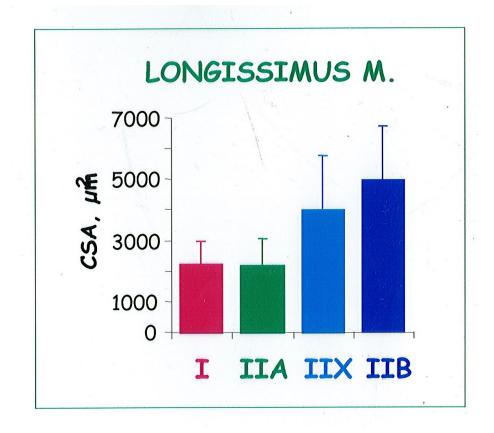
- Fast oxido-glycolytic IIA fibres
- Using both carbohydrates and lipids
- Adaptated to rythm changes

Classification according to fibre type

| Туре | I | IIA | IIX | |
|-------------------------|----|-----|-----|---------|
| | | | | |
| « Mean value » | 20 | 25 | 55 | |
| m. Diaphragma | 55 | 45 | 0 | (red) |
| m. Longissimus thoracis | 25 | 25 | 50 | |
| m. Semitendinosus | 15 | 25 | 60 | (white) |

In cattle, according to Totland et al. (1991), Picard et al (2002)

Cross section area of the fibres



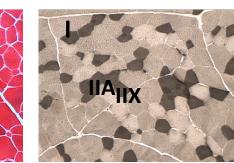
Mean cross section area of fibres

Azorubine

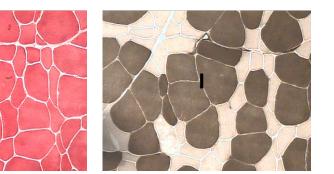
TΒ

RA

ATPase



| <u>Mean</u> a | area of typ | <u>e I fibres</u> |
|---------------|-------------|-------------------|
| ТВ | RA | signif. |
| 1725 | 3957 μm | 2 *** |



TB: I<IIA<IIX RA: I>IIA>IIX

In cattle TB: m. *triceps femoris* Ra: m. *rectus abdominis*

Oury et al., 2009

CLASSIFICATION OF MUSCLE FIBRES

Methods

DIFFERENT METHODS BASED ON

- functional tests (speed of contraction)
- metabolic criteria (the type of energy metabolism)
 - speed of energy utilization during contraction
 - o main source of energy

1- In situ techniques

tissues 🔿 section

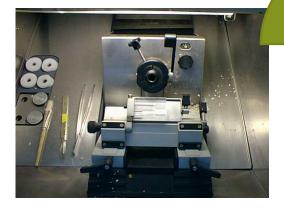
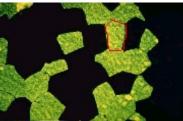


Image analysis

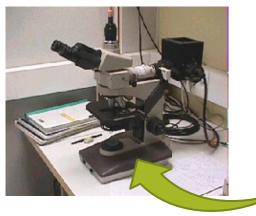
Detection



enzymatic (SDH activity)



immunology (slow MyHC)



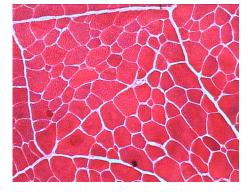
Histochemistry for fibre typing

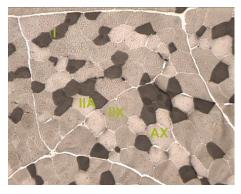
Detection of three pure fibres and hybrid fibres in cattle muscle

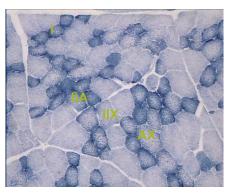
Azorubine

ATPase

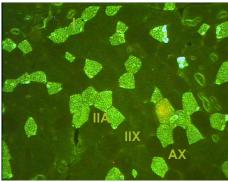
SDH



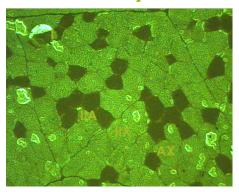




slow MyHC

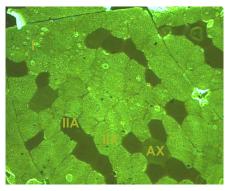


5B9 Alexis



fast MyHC

S5 15F4 (Agrobio)



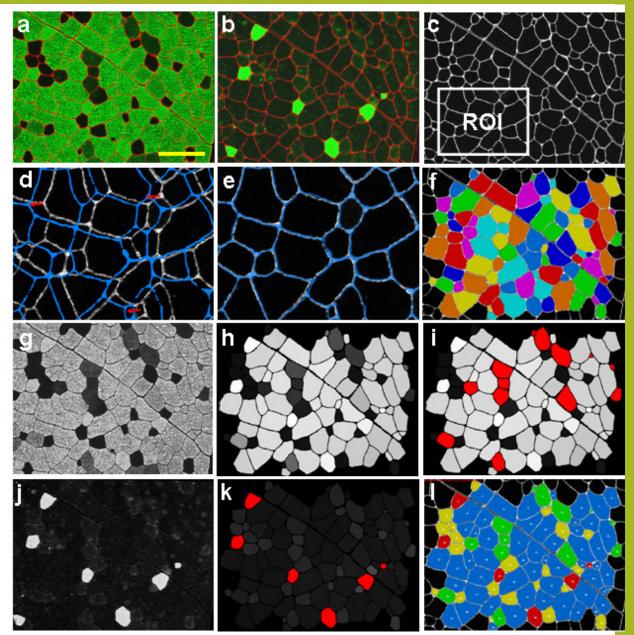
MyCH I + IIx

S5 8H2 (Agrobio)

lmage analysis

on high number of fibres

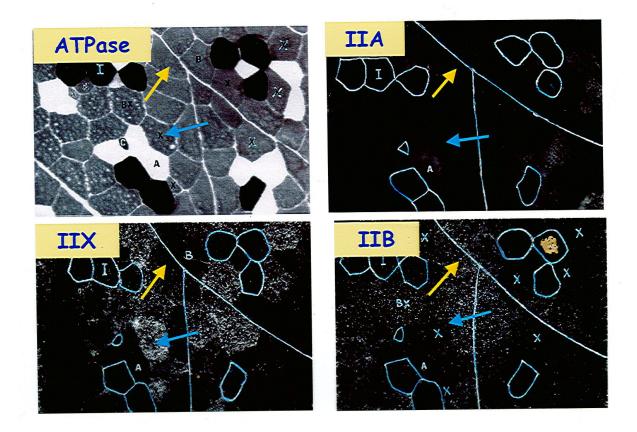
- % of each fibre type
- area
- % of area



Meunier et al., 2010, Histochem²⁷Cell Biol

MyHC in situ hybridization

in pig longissimus muscle (100 kg BW)



Lefaucheur et al. (1998)

2- From muscle homogenates

Electrophoretic separation of MyHC isoforms



SDS PAGE

Talmadge and Roy (1995) modified by Picard et al. (2011)

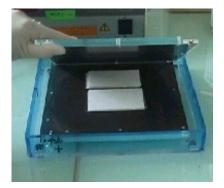
Western-blot

proteins

Electrophoresis



Transfert



Immuno-detection

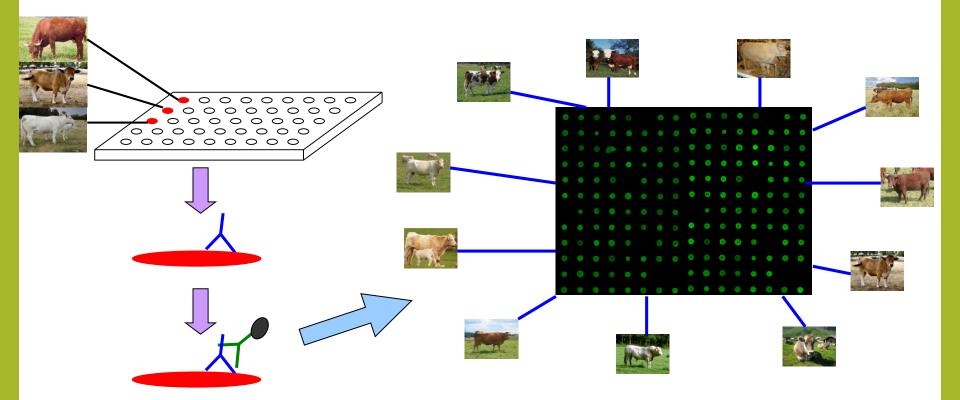
Bovine myoblast culture



desmin

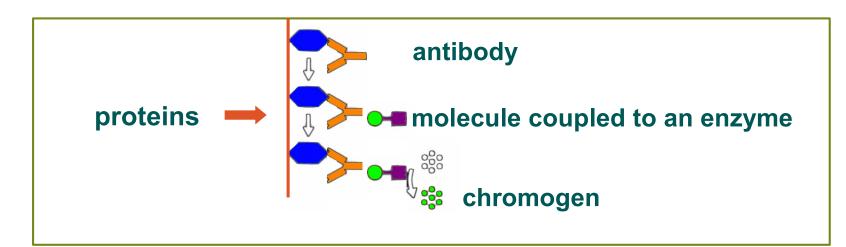
D4 D6 D8 D10

High-throughput protein analysis

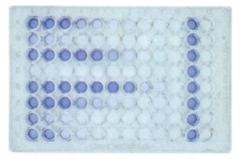


Dot-blot technology: analyse up to 96 samples simultaneously simplification of the western blot method

ELISA assay



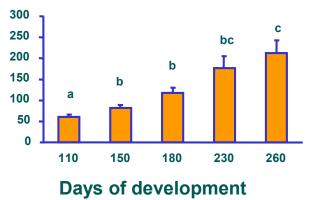
Detection

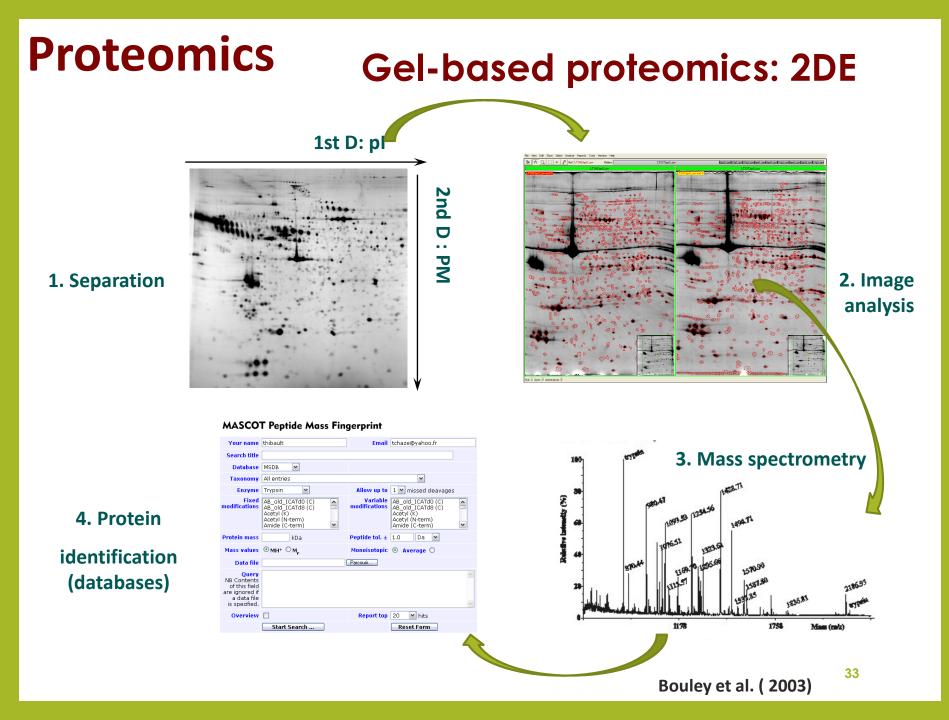


Optical density



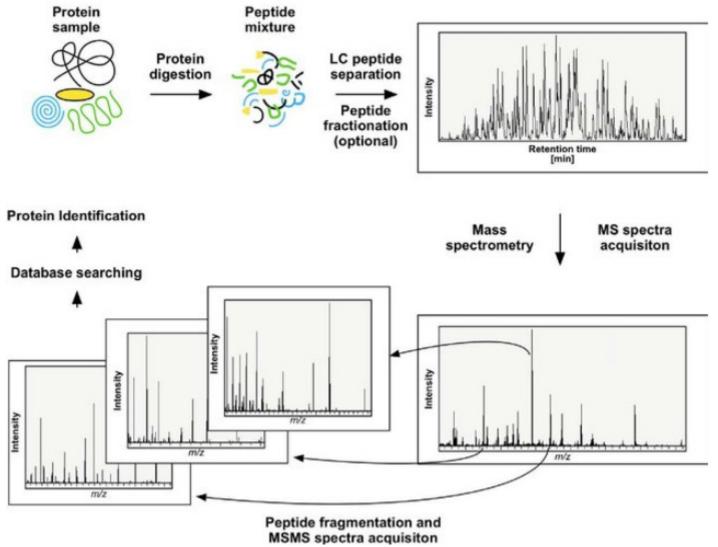
FABP content





Proteomics

Shotgun: nano LC MS/MS



From Vojtech Tambor

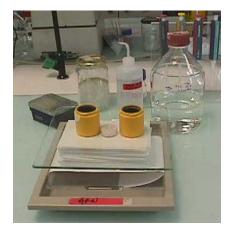
Gene expression

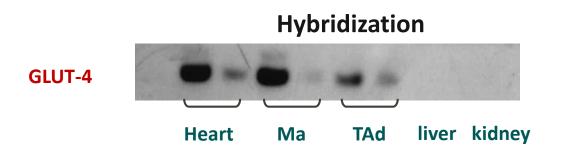
RNA

Northern-blotting to detect specific RNA molecules among a mixture of RNA

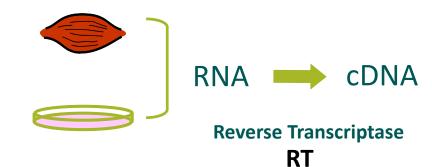
Electrophoresis

Transfert



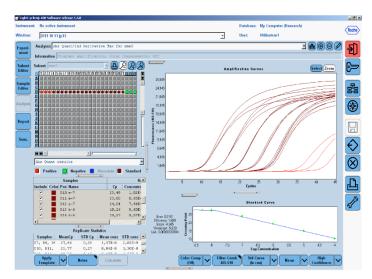


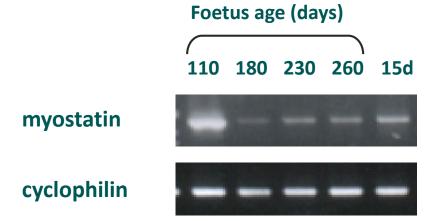
qRT-PCR



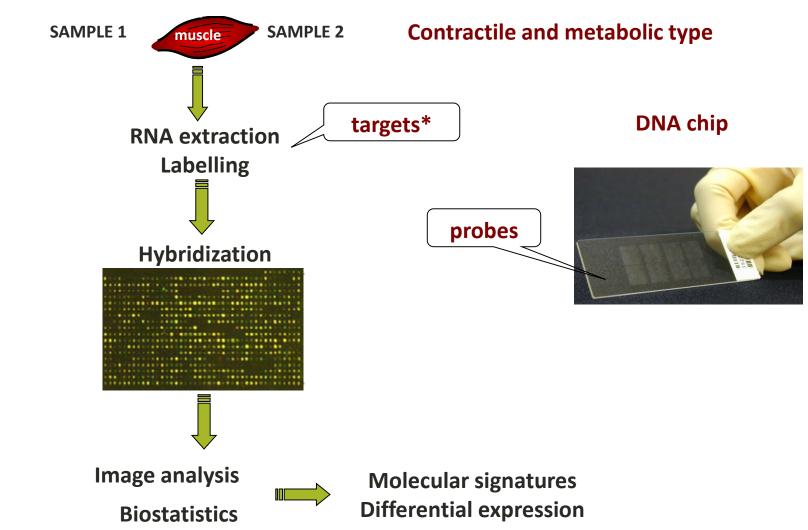


PCR



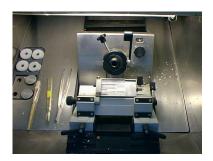


Transcriptomics



Choosing the most accurate technique

- Reveal the contractile type by immuno-histochemistry using anti-MyHC antibodies and the SDH activity on serial sections (Picard and al., 1998): distinguish the hybrid fibres, and get information on the cross section area of each fibre type
- Not relevant for high throughput phenotyping (large numbers of animals)
- Detect contractile type using electrophoresis of MyHC and metabolic type by assaying metabolic enzymes





IIB OR NOT IIB?

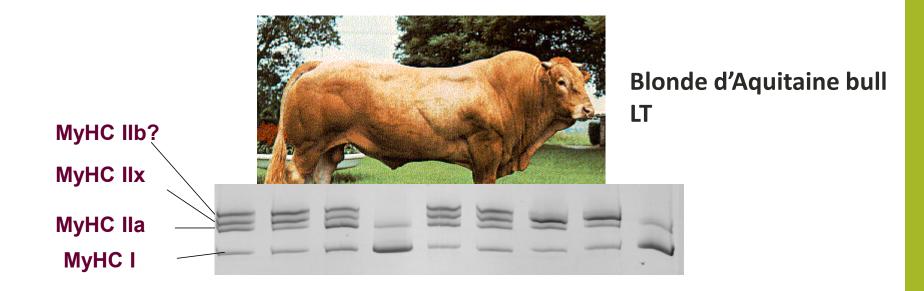
Myosin IIB

MyHC-IIb is the predominant motor protein in most skeletal muscles of rats and mice

The mRNA for this isoform is only expressed in a very small subset of specialized muscles in adult large mammals, including humans.

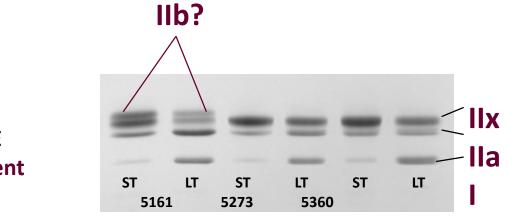
Is IIb MyHC expressed in cattle?

• For many authors, IIb MyHC (MYH4, BTA 19) would not be expressed in cattle muscles, but extraoccular muscle:



A fourth MyHC isoform in cattle

 A particular MyHC in Blonde d'Aquitaine bulls with common ancestor



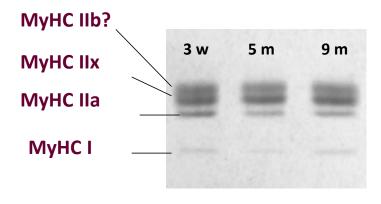
| SDS | -PAGE |
|-------------|--------------|
| 5-8% | gradient |

| llb? | <u>35%</u> | <u>18%</u> |
|------|------------|------------|
| llx | 41% | 23% |
| lla | 20% | 37% |
| 1 | 4% | 22% |

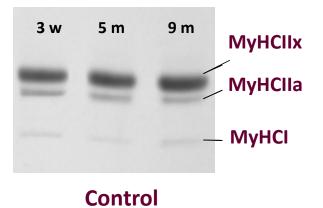


A MyHC expressed in post-natal muscle

- This MyHC is detected at all studied ages
- Onset as soon as fœtal life?



Bull with this MyHC



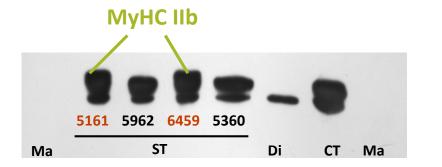
Identification

Immuno-detection

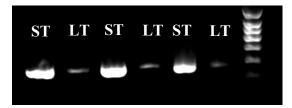
with an antibody specific of the fast MyHC (IIa, IIx, IIb)



Primers designed in the 5'-UTR of the MyHC IIb transcript



Ma : *masseter*, slow (I), Di: *diaphragma* (I +IIa), CT: *cutaneus trunci* (IIa+ IIx)



Amplification of a cDNA fragment and sequencing

This isoform is the IIb MyHC (encoded by MYH4)

Picard and Cassar-Malek, 2009

Abundance of the transcript

Specific regulation of MYH4 expression in cattle?

RT-PCR using primers for 5'-UTR MyHC IIb (Chikuni et al, 2004)

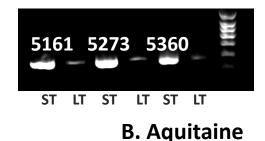
AMPLIFICATION



Charolais

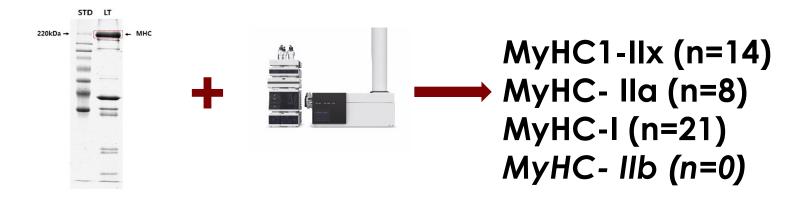
~300 pb in ST

Transcripts observed in all 11 young bulls but the protein was present in 2 only (ST BA)



What's next?

Kim (2014) by using electrophoresis and nano LC-MS/MS of MyHCs did not observed unique peptides of MyHC IIb in Hanwoo Steer LT muscle



However this has to be tested on our samples!

Conclusion



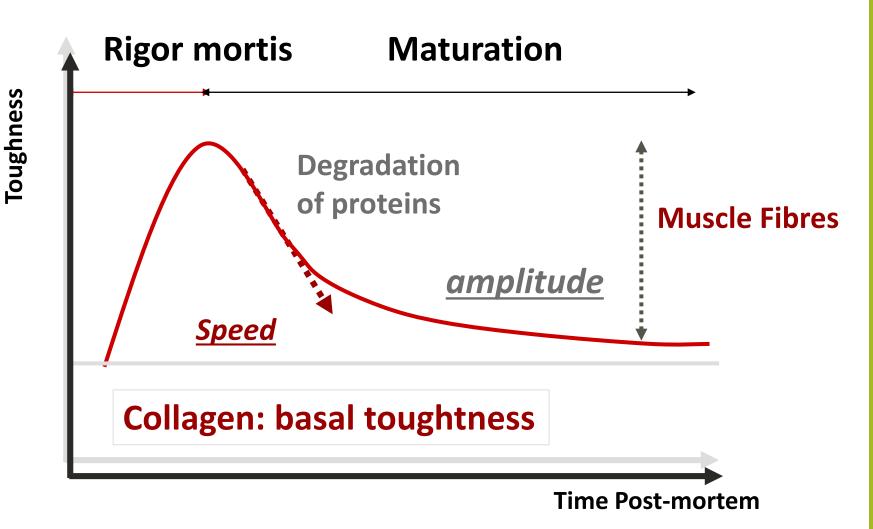
IN CATTLE

- Three types of fibres I, IIA, IIX
- and a fourth one IIB in some french bovines
- with a variable frequency between breeds
- 6% in Charolais, 35% in Blonde d'Aquitaine, 45% in Limousin

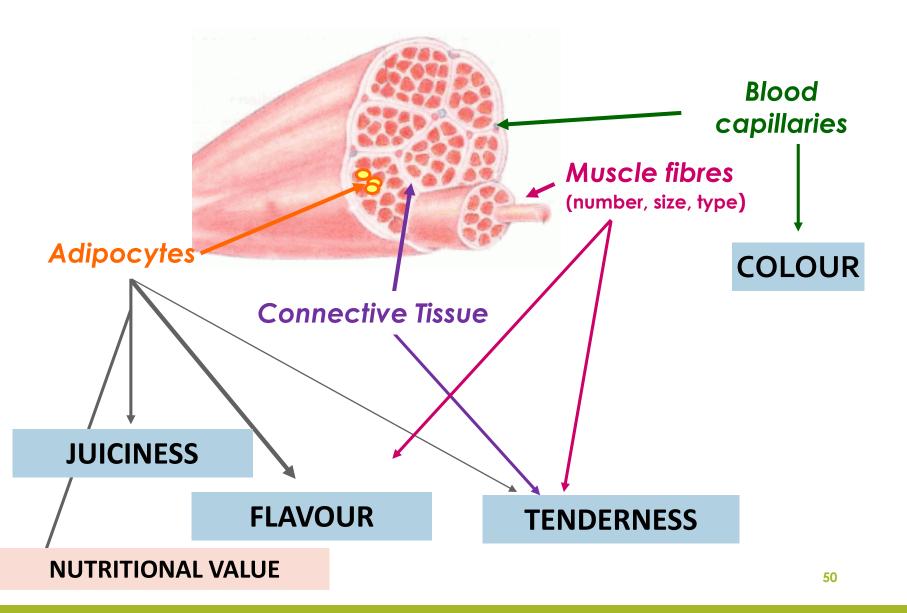
FROM MUSCLE TO MEAT



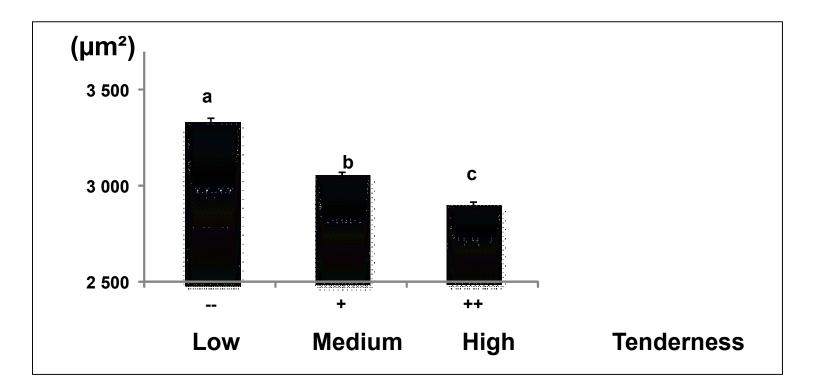
Conversion of muscle into meat



How muscle biochemistry affects Beef quality



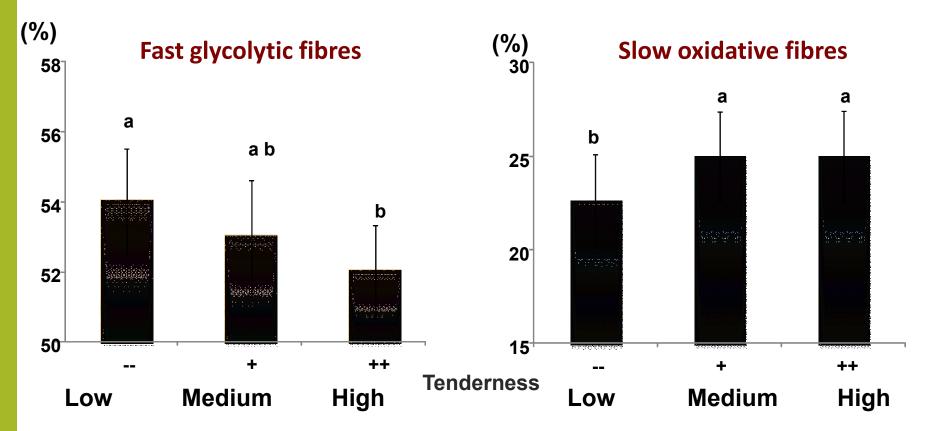
Cross sectional area of fibres/quality



For *Longissimus thoracis* muscle (LT), the tenderest muscles have the lowerest cross sectional area of fibres

Chriki et al., ICoMST 2011

Fibre type/quality



For *LT,* the muscles the less tender have the higher % of IIX fibres and the lower % of slow I fibres

Chriki et al., ICoMST 2011

Muscle properties according to the breed

- Muscle mass + collagen + intra-muscular fat

+ Muscle mass - collagen - intra-muscular fat

| + red | + white |
|-----------|------------|
| slow | fast |
| oxydative | glycolytic |



Dairy breeds

Hardy breeds

Beef breeds

Research for the meat sector

- For beef cattle research, a main objective is to control both the development of muscles and qualities of the meat, with specific attention towards tenderness (the top priority quality attribute).
- Variability in beef tenderness originates from genetic polymorphisms and modulation of gene expression according to rearing conditions.
- Beef tenderness is a complex phenotype (post-mortem expression). Identification of relevant markers at the DNA or protein level is ongoing.
- The next challenge is to integrate the knowledge and develop detection tests for desirable animals to ensure proper breeding programmes or management systems.

Biomarkers of tenderness

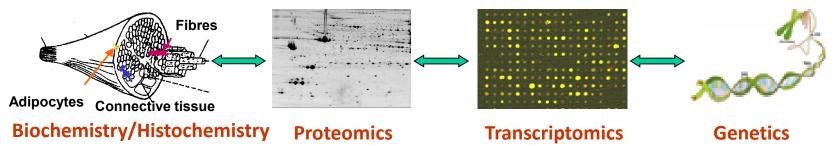
• The relationships between fibres' properties and tenderness is different according to the muscle



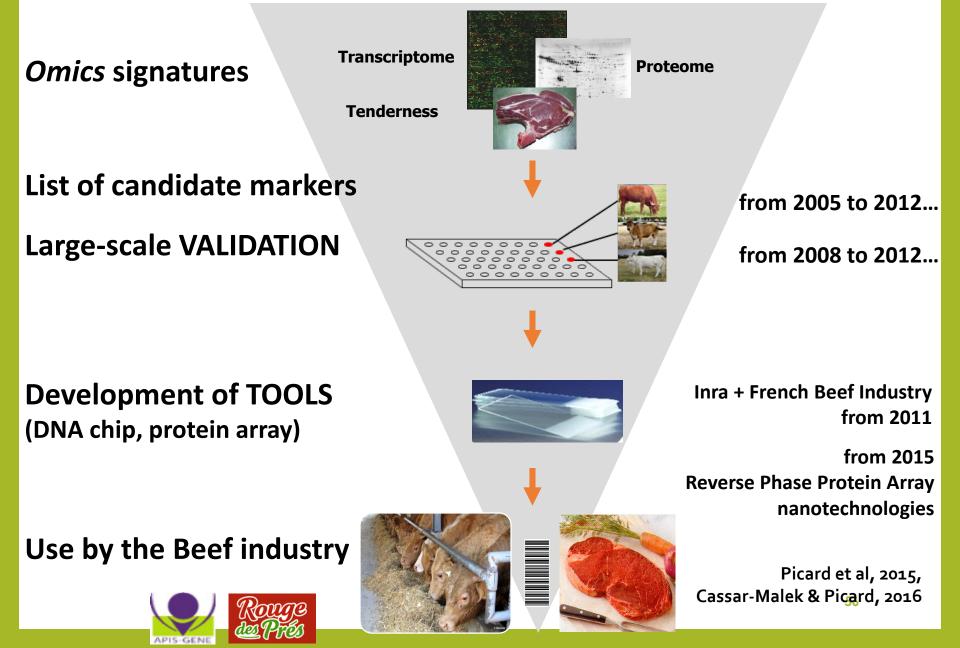
Difficult to establish general law

 Muscle characteristics (collagen, fibres, lipids) explain around 30% of tenderness variability

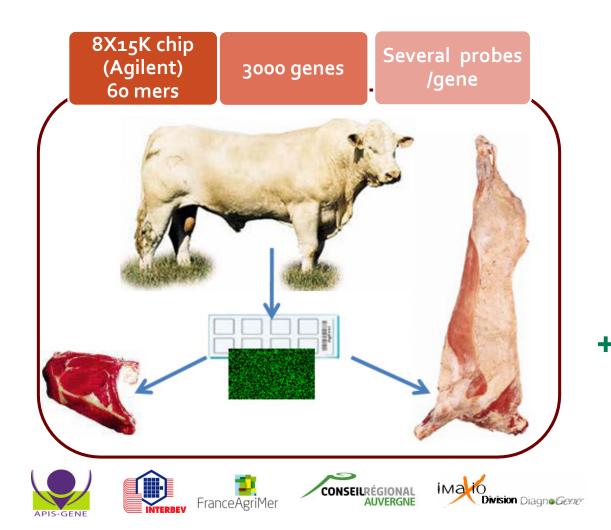
Need to identify other muscle characteristics
involved in tenderness by Genomics



From omics to prediction tools for the Beef sector



A "meat quality chip" for Beef prediction



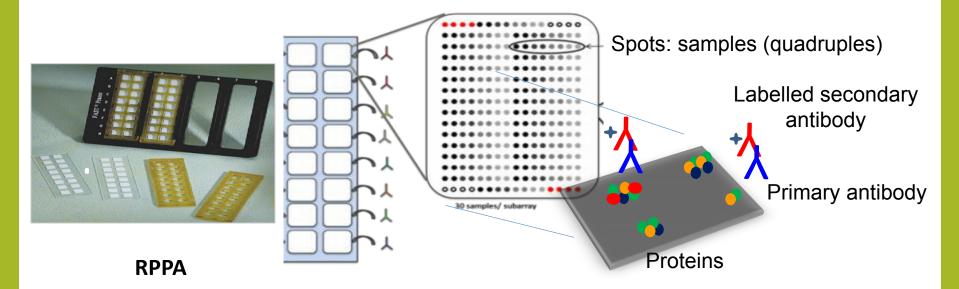


Development of turnkey software

Hocquette et al. 2012. BMC Veterinary Research, 8:135.

Towards high throughput screening of protein biomarkers

Reverse Phase Protein Arrays, a recent methodology



Quantification of the abundance of one biomarker in up to 500 samples

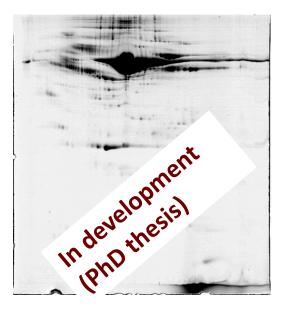
On going project + other methods (nano technologies)

Toward less invasive markers

Search for plasma markers

The plasma perfuses all tissues of the body and thus may contain information on physiological mechanisms and performance.

In livestock animals, plasma proteomics is a promising strategy to identify biomarkers of the potential of meat production.





Take-home messages

• The skeletal muscle tissue is a specialized tissue but heterogeneous in structure.

• The muscles are characterized by their contractile and metabolic properties (isoforms of contractile proteins, preferential metabolic pathway).

• They are involved in different types of movements (fast or slow, short or endurance effort).

• The muscle characteristics can explain only one third of variability in Beef quality (tenderness). Biomarkers are identified for development of « diagnostic » tools.