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Towards the structural screening of microbial ecosystems : selected projects and MetaFoldScan presentation

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Gwenaëlle André-Leroux. Towards the structural screening of microbial ecosystems : selected projects and MetaFoldScan presentation. Séminaire, Dec 2018, Montevideo, Uruguay. hal-02785774

HAL Id: hal-02785774

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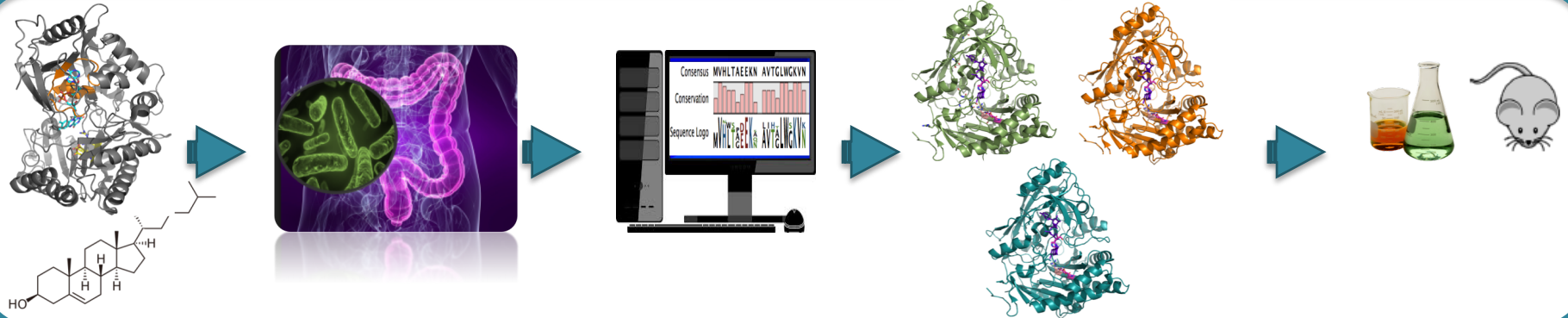
Submitted on 4 Jun 2020

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Towards the Structural Screening of Microbial Ecosystems

Selected projects and MetaFoldScan presentation

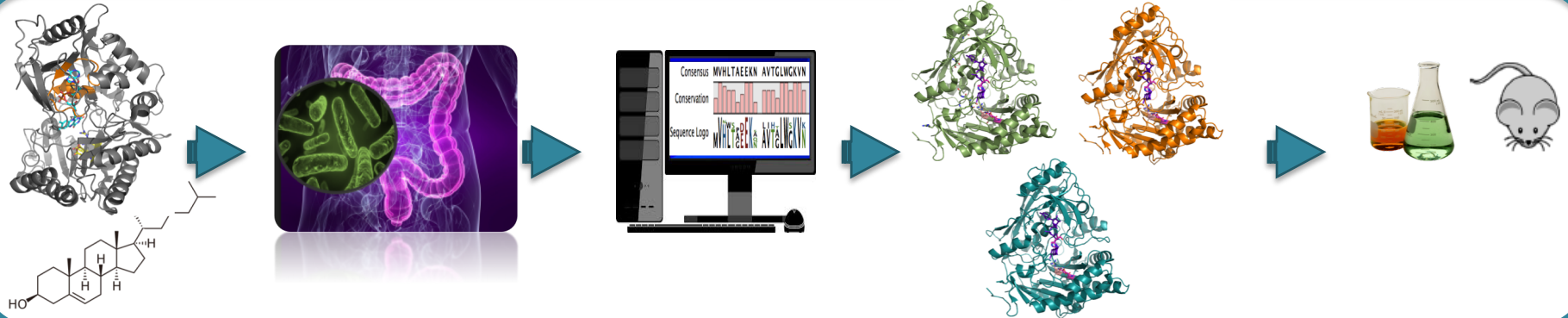


Gwenaëlle André-Leroux

Montevideo University of Sciences December 2018

Towards the Structural Screening of Microbial Ecosystems

Selected projects and MetaFoldScan presentation

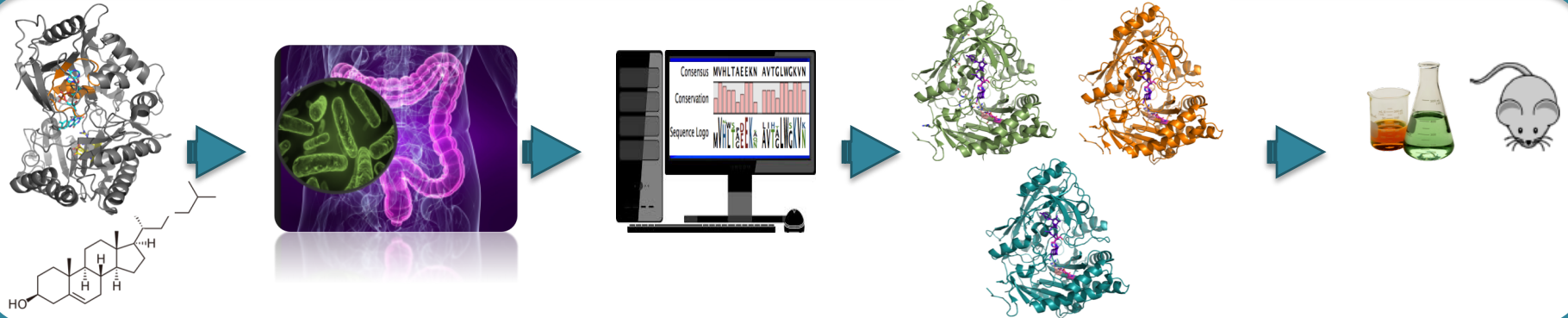


Outline

- Cholesterol conversion in the gut microbiota: the mystery enzyme(s) ?
- MetaFoldScan project : 3D screening of the gut microbiota
- Conclusions and perspectives

Towards the Structural Screening of Microbial Ecosystems

Selected projects and MetaFoldScan presentation



Outline

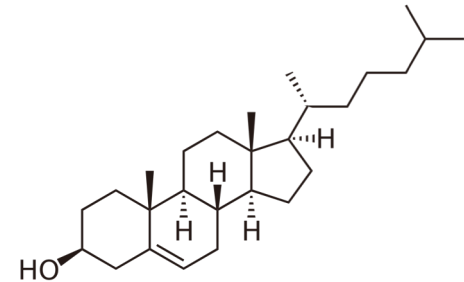
- Cholesterol conversion in the gut microbiota: the mystery enzyme(s) ?
- MetaFoldScan project : 3D screening of the gut microbiota
- Conclusions and perspectives

★ Cholesterol :

- Is a sterol that plays a central role as cell-membrane agent.
- Is precursor of steroid hormone (progesterone, testosterone, cortisone) and bile salt.
- Originates 30% from diet vs 70 % from bile and desquamated sterol of the gut epithelium.

★ Cholesterol level and gut microbiota :

- Can be metabolized by colonic bacteria.
- The gut microbiota reduces cholesterol to coprostanol.
- Neomycin impacts serum cholesterol and fecal sterol in hypercholesterolemic patients.



★ Cholesterol is absorbed by human intestine, not coprostanol :

- Cholesterol conversion relates to abundance of bacteria with cholesterol reducing activities.
- Gérard et al, 2004, Veiga et al, 2005
- Cholesterol conversion to coprostanol could result in lower cholesterolemia.

★ One strain from hog-sewage lagoon:

Gram+ coccobacillus *Eubacterium coprostanoligenes* ATCC 51222T Freier *et al*, 1993

Oral administration of *E. copro* → significant decrease of plasma cholesterol in dietary induced hypercholesterolemic rabbits Li *et al*, 1995

★ One from « high converter » human:

Isolate from the *Bacteroides* phylum, *Bacteroides* sp. Strain D8. Gram- anaerobie. Gérard *et al*, 2007

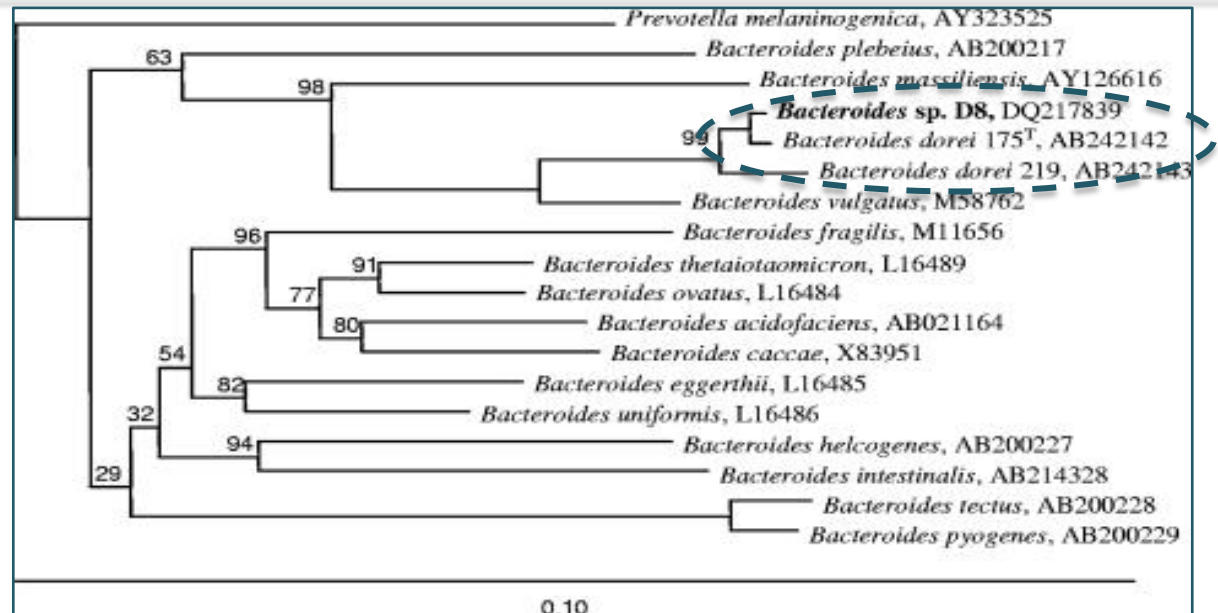
Phylogenetic tree construction → this strain clusters in an independant clade with the two isolates of *Bacteroides dorei* species.

Intestinal microbiota
>1,000 genera of bacteria.

2 phyla

Bacteroidetes

Firmicutes



Phylogenetically close

Phylogenetically remote

| Strain | Medium | SBM | BCM |
|---------------------------------|--------|-----|-----|
| <i>B. Dorei 175^T</i> | | - | - |
| <i>B. Dorei D8</i> | | + | - |
| <i>E. coprostanoligenes</i> | | - | + |

SBM : Standard Brain Medium
 BCM: Basal Cholesterol Medium

Coll. C. Juste and P. Gérard, Micalis



Metabolic pathway for cholesterol to coprostanol conversion in *B. dorei D8*?



Which are the enzymes of the catabolic pathway that degrade cholesterol into coprostanol?

★ One strain from hog-sewage lagoon:

Gram+ coccobacillus *Eubacterium coprostanoligenes* ATCC 51222T Freier *et al*, 1993

Oral administration of *E. copro* → significant decrease of plasma cholesterol in dietary induced hypercholesterolemic rabbits Li *et al*, 1995

★ One from « high converter » human:

Isolate from the *Bacteroides phylum*, *Bacteroides sp. Strain D8*. Gram- anaerobie. Gérard *et al*, 2007

Phylogenetic tree construction → this strain clusters in an independant clade with the two isolates of *Bacteroides dorei* species.

Microbial genes or enzymes involved in cholesterol metabolism in the gut are unknown.

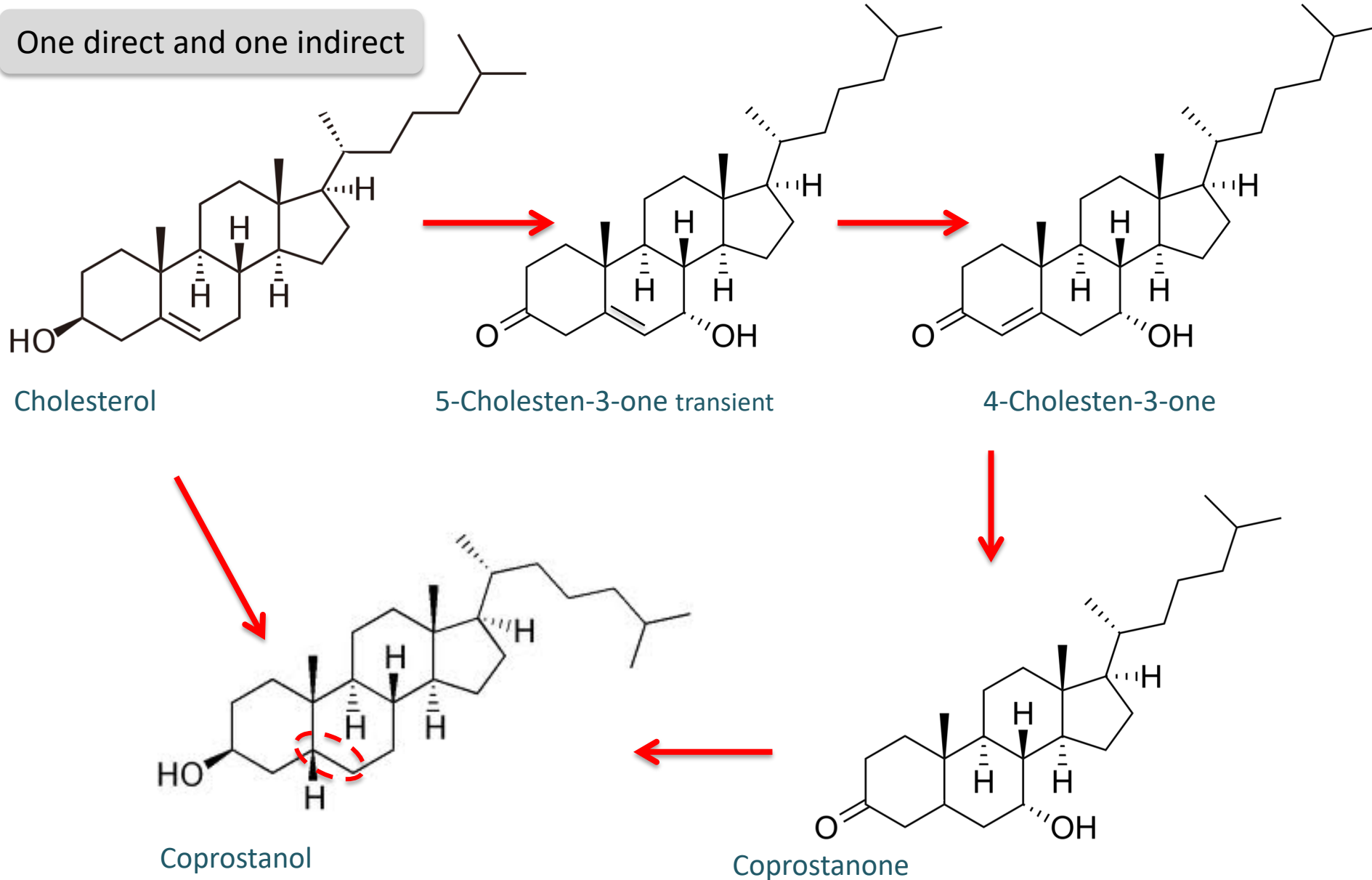
However ...

Genomes of *B. Dorei*D8 & *B. Dorei*175^T sequenced and assembled at MaIAGE

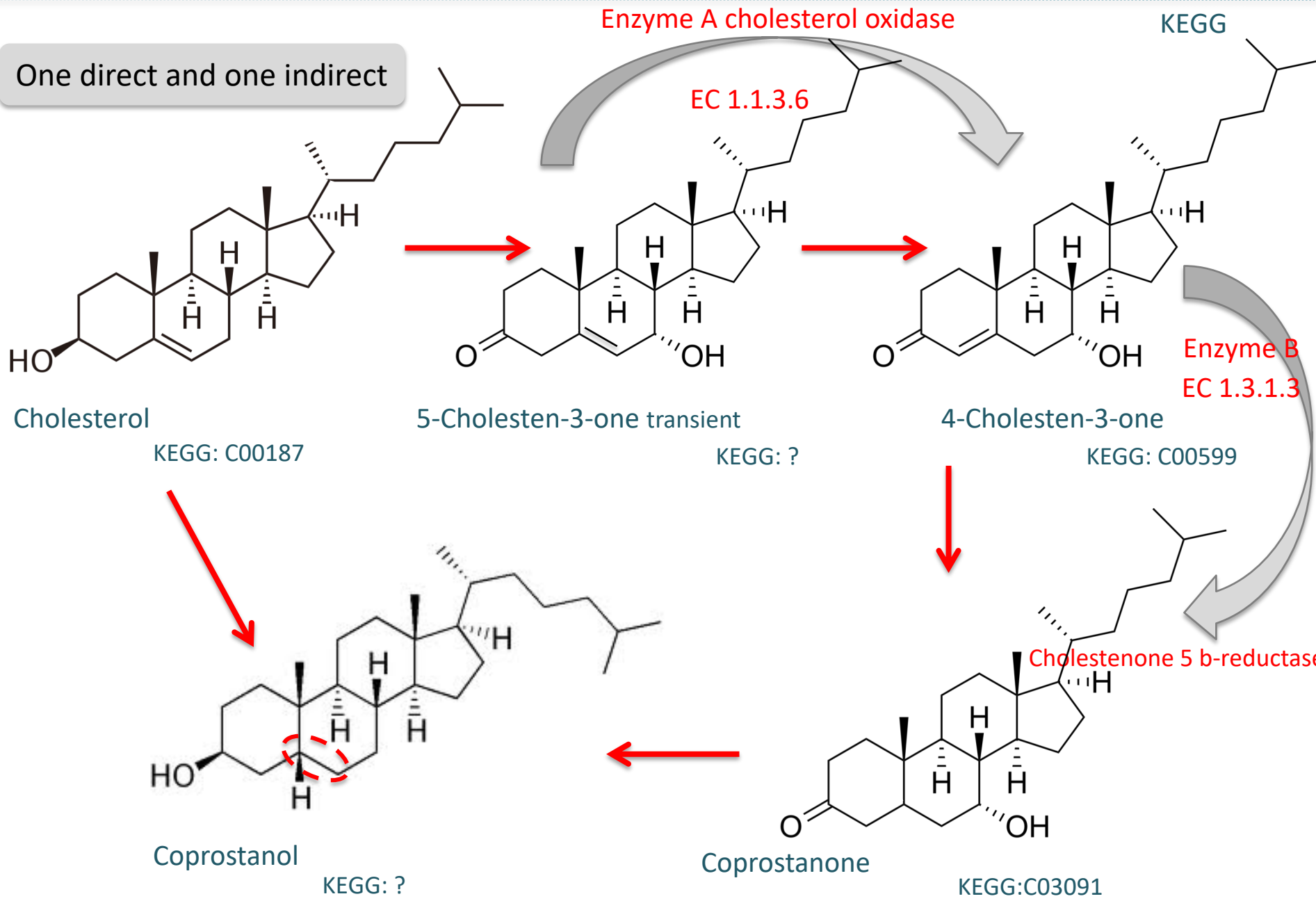
Valentin Loux

One direct and one indirect

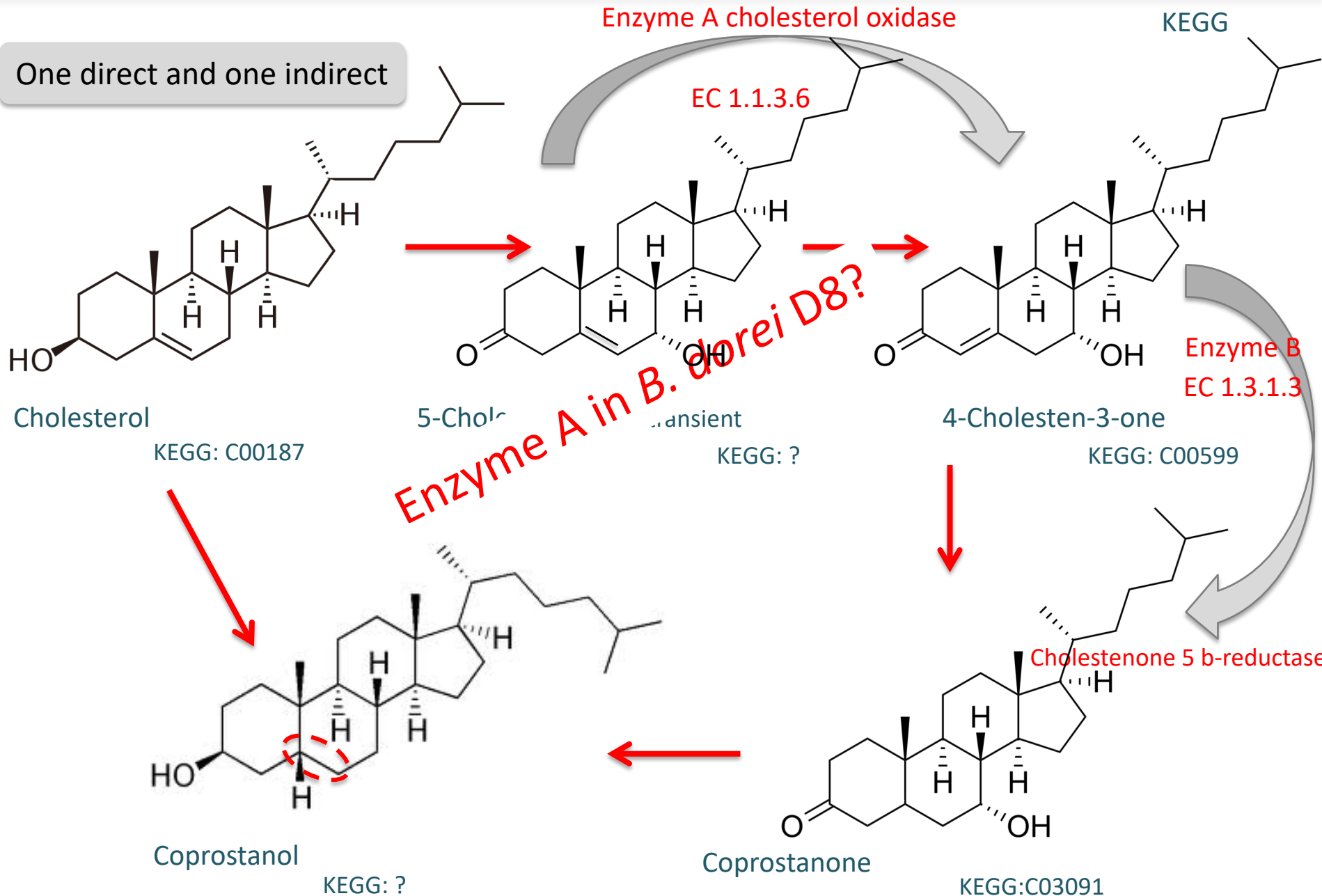
KEGG



One direct and one indirect

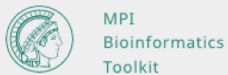


One direct and one indirect



Server of protein structure prediction

<http://toolkit.tuebingen.mpg.de/hhpred>



Search Alignment Sequence Analysis 2ary Structure 3ary Structure Classification Utils
 HHblits HHpred HMMER PatternSearch ProtBLAST/PSI-BLAST

HHpred

| ID | Date | Tool | |
|---------|------|------|---|
| 7456410 | | HHPR | X |
| 1681937 | | HHPR | X |
| 5162681 | | HHPR | X |
| 6311127 | | HHPR | X |
| 8160192 | | HHPR | X |

Input Parameters

Enter a protein sequence/multiple sequence alignment in FASTA/CLUSTAL format

To create a structural model of your query protein, run a HHpred search with the HH-suite database, select the top-scoring template(s) and click on 'Create model using template(s)'. You can also generate a PIR file that can be subsequently submitted to MODELLER.

Query : Fasta sequence

[Paste Example](#) [Upload File](#)

Align two sequences or MSAs

Select HH-Suite Database

× PDB_mmCIF70_11_May

Proteomes

Custom JobID

Submit Job

Protein data bank

Profile-profile comparison tools:

- Calculated from a MSA of related sequences collected from Hhblits (>Psi-blast+).
- Matrix of similarity scores calculated from frequency of aa at the corresponding positions in the MSA.

Profile contains more information than a sequence.

Result

Hits: Rank of « matching » pdb proteins

- Prediction of MSA profile with SS for each protein of *B. Dorei 175^T* & *B. dorei D8* genomes
- Compare each protein with a bank of cholesterol oxidase « Enzyme A » to be assessed

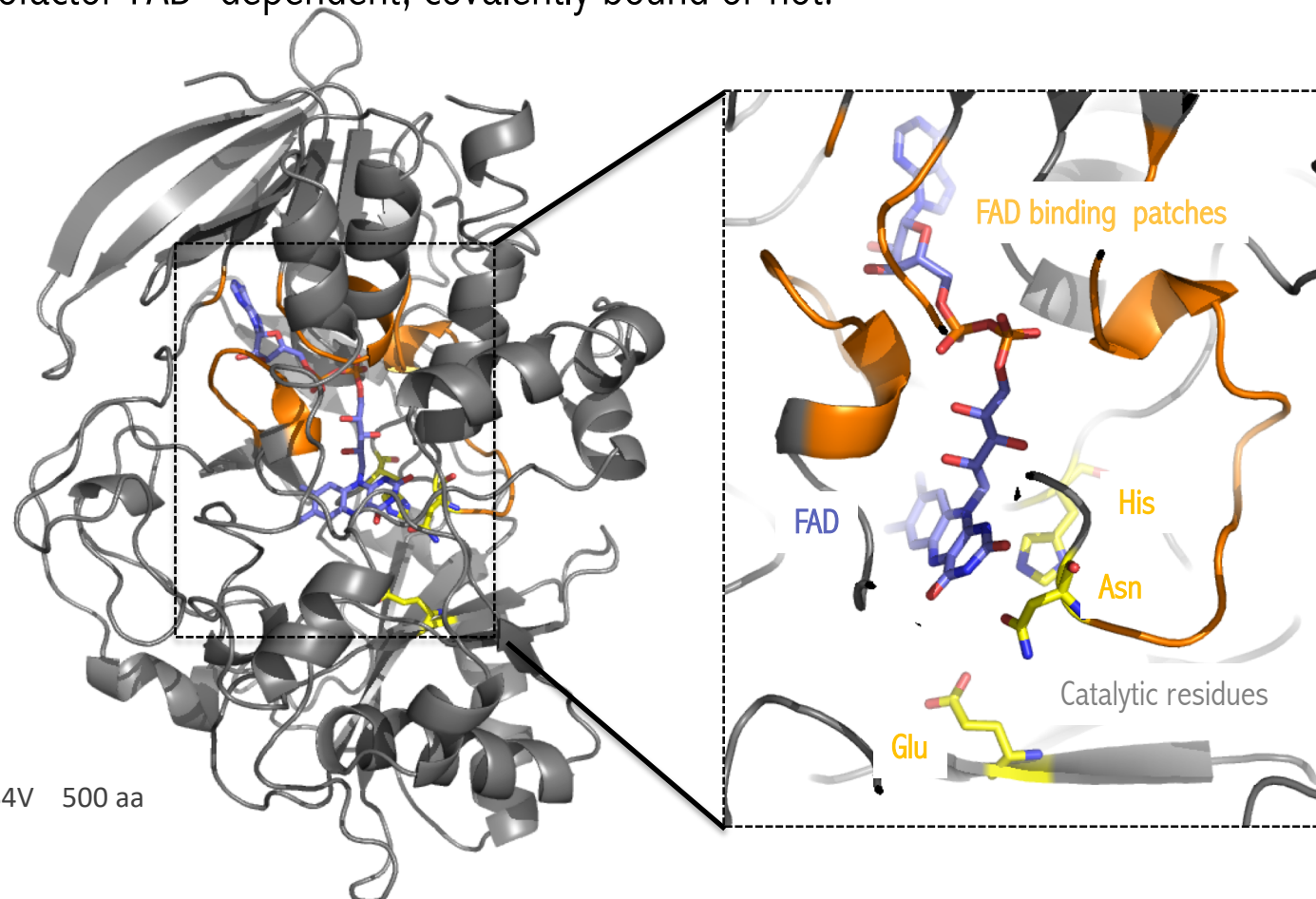
- 1- Split of each fasta sequence in the genome -> more than 3,400 for each genome of *B. dorei*
- 2- Multiple sequence alignment for each fasta: 'HHblits' > PSI-Blast
- 3- Prediction & addition of SS elements: 'adssl.pl'
- 4- Profiling & comparaison with the 3D template « bank »: 'HHBlitsdb.pl' & 'HHsearch'
- 5- Extraction, ranking and analyse of scores.
- 6- Analyse of selected sequences of proteins. Annotation? Fonction?
- 7- *In fine*, a list of cholesterol oxidase putative proteins to test *in vitro* / *in vivo*



The 3D template « bank » is critical

Cholesterol oxidase:

- monomeric flavoenzyme catalyzes oxidation and isomerization into cholest-4-en-3-one.
- two folds exist, cofactor-FAD- dependent, covalently bound or not.



1B4V. pdb & fasta

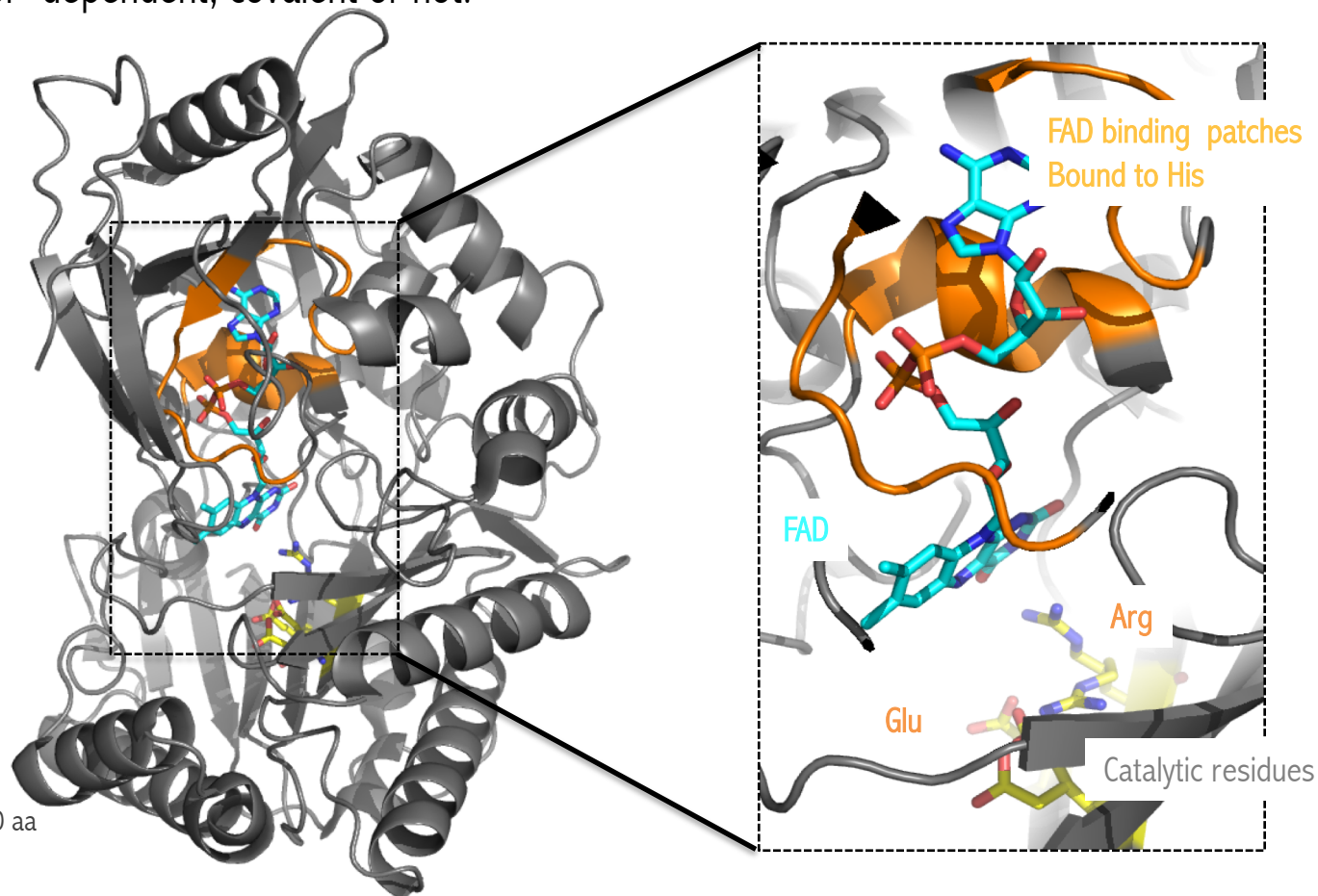
Cholesterol oxydase from *Streptomyces* Yue et al, 1999 - 1.5 Å

1COY.pdb & fasta

Cholesterol oxydase from *R. hoagii ex B. steroliticum* Li et al, 1993 - 1.8 Å

Cholesterol oxidase:

- monomeric flavoenzyme catalyzes oxidation and isomerization into cholest-4-en-3-one.
- two folds exist, cofactor- dependent, covalent or not.



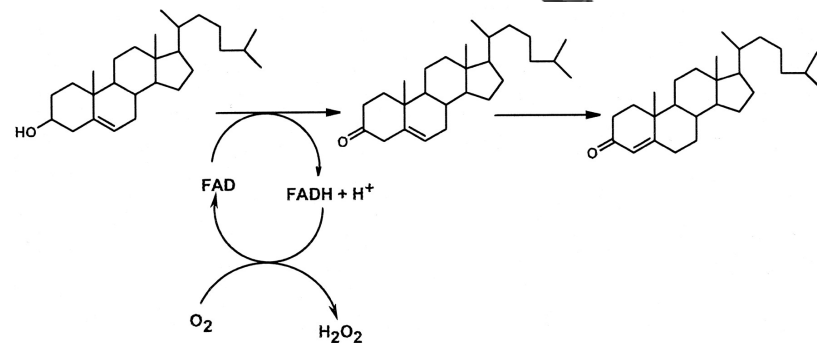
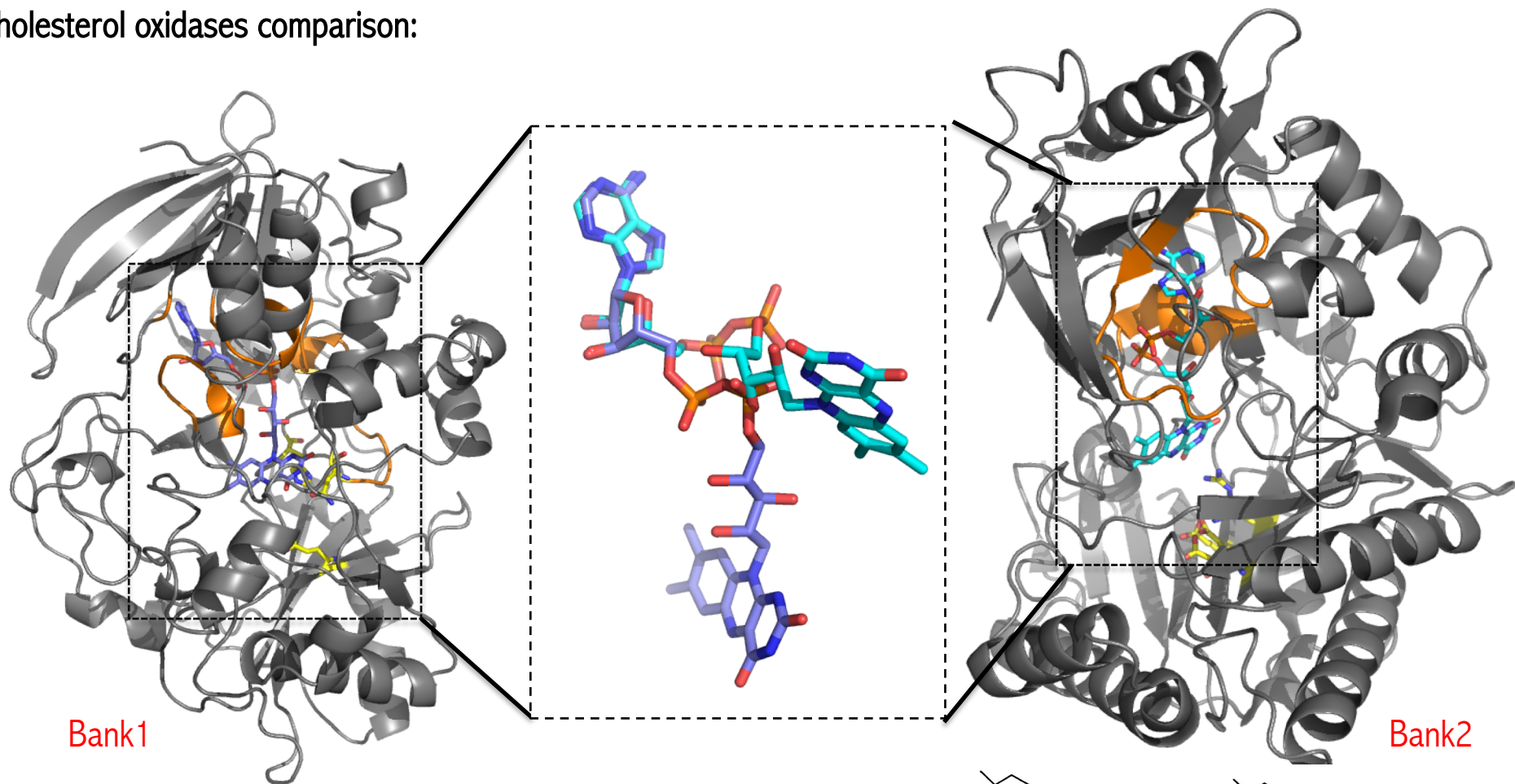
3JS8.pdb & fasta

1119.pdb & fasta

Cholesterol oxydase from *Chromobacterium sp. DS-1* Sagermann et al, 2010 - 1.5 Å

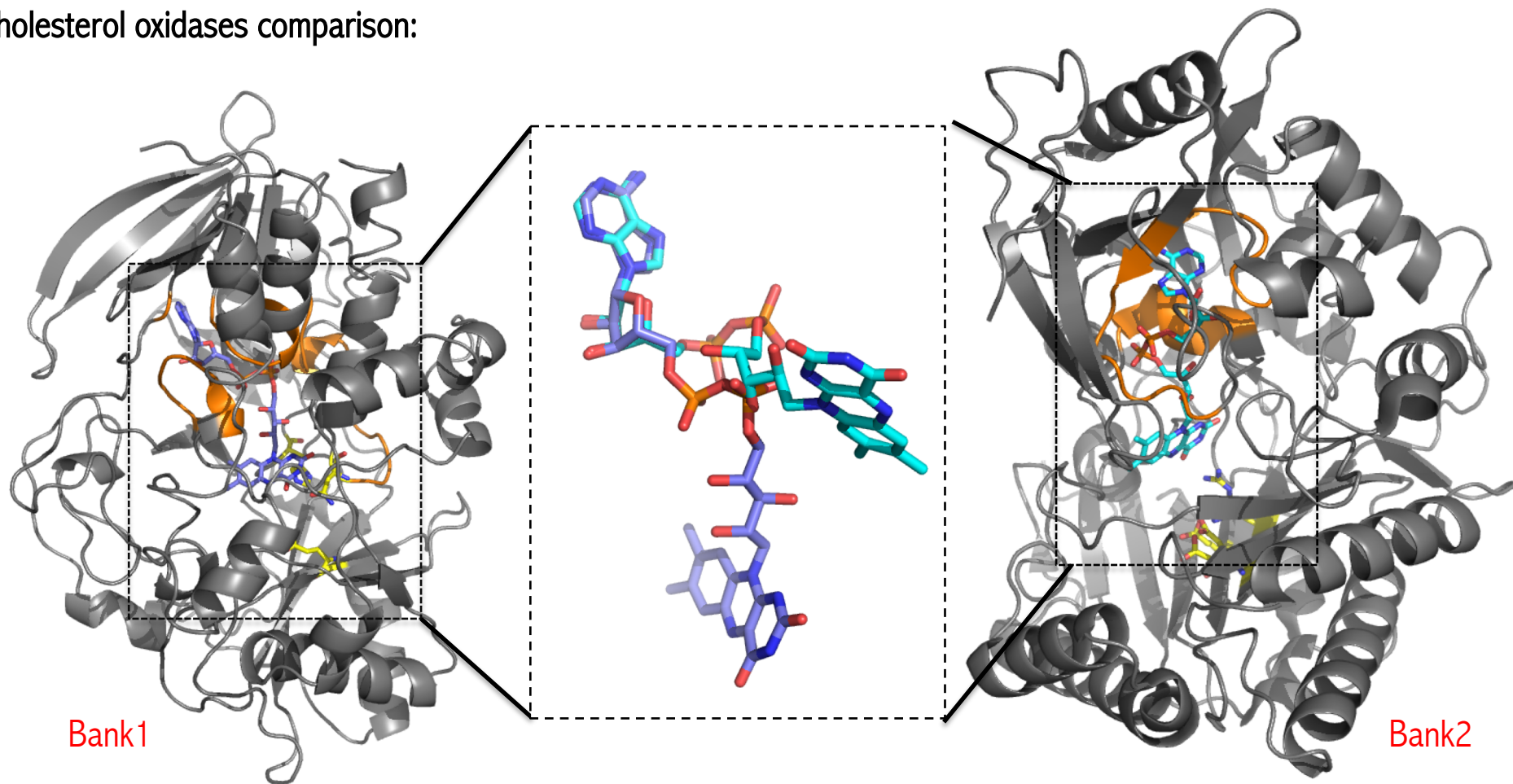
Cholesterol oxydase from *B. steroliticum* Coulombe et al, 2001 - 1.7 Å

Cholesterol oxidases comparison:



- ★ 3D structures distinct
- ★ Catalytic sites different
- ★ Binding patches divergent
- ★ FAD binding topologies contrasting despite same redox role

Cholesterol oxidases comparison:



- ★ 3D structures distinct
- ★ Catalytic sites different
- ★ Binding patches divergent
- ★ FAD binding topologies contrasting despite same redox role



Structural detection with 2 banks required

★ *B. dorei*D8 Sorted by Probability – Bank1

| | | | | |
|-----|------|--------|--------|---|
| 37 | 92.9 | 5=32 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01957.fasta.a3m.hhm.out.hhmakeBK1 |
| 33 | 93.3 | 2=31 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_03587.fasta.a3m.hhm.out.hhmakeBK1 |
| 32 | 94.2 | 3=29 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00872.fasta.a3m.hhm.out.hhmakeBK1 |
| 241 | 97.1 | 209=32 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00845.fasta.a3m.hhm.out.hhmakeBK1 |
| 479 | 97.2 | 441=38 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_04063.fasta.a3m.hhm.out.hhmakeBK1 |
| 179 | 97.5 | 142=37 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01842.fasta.a3m.hhm.out.hhmakeBK1 |
| 49 | 97.6 | 11=38 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_02168.fasta.a3m.hhm.out.hhmakeBK1 |
| 256 | 97.6 | 195=61 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_03122.fasta.a3m.hhm.out.hhmakeBK1 |
| 35 | 97.6 | 2=33 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01099.fasta.a3m.hhm.out.hhmakeBK1 |
| 35 | 97.8 | 2=33 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00348.fasta.a3m.hhm.out.hhmakeBK1 |
| 37 | 97.9 | 4=33 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_04051.fasta.a3m.hhm.out.hhmakeBK1 |
| 37 | 98.0 | 2=35 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00992.fasta.a3m.hhm.out.hhmakeBK1 |
| 66 | 98.1 | 29=37 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_04457.fasta.a3m.hhm.out.hhmakeBK1 |
| 37 | 98.2 | 1=36 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01766.fasta.a3m.hhm.out.hhmakeBK1 |
| 40 | 98.2 | 3=37 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00293.fasta.a3m.hhm.out.hhmakeBK1 |
| 41 | 98.2 | 7=34 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01761.fasta.a3m.hhm.out.hhmakeBK1 |
| 163 | 98.7 | 104=59 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00882.fasta.a3m.hhm.out.hhmakeBK1 |
| 118 | 98.9 | 81=37 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_01104.fasta.a3m.hhm.out.hhmakeBK1 |
| 208 | 99.1 | 133=75 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00352.fasta.a3m.hhm.out.hhmakeBK1 |
| 237 | 99.1 | 175=62 | 1B4V:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_03708.fasta.a3m.hhm.out.hhmakeBK1 |
| 280 | 99.3 | 211=60 | 1COY:A | /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS/BANK1/bdd8_00254.fasta.a3m.hhm.out.hhmakeBK1 |

★ *B. dorei*D8 Sorted by Probability – Bank2

| | | | | |
|------|---|---------|----------|--|
| 5.9 | - | 212-241 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_01995.fasta.a3m.hhm.out.hhmakeBK2- |
| 6.8 | - | 5-19 | - 1I19:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_01730.fasta.a3m.hhm.out.hhmakeBK2- |
| 7.0 | - | 26-52 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_00608.fasta.a3m.hhm.out.hhmakeBK2- |
| 7.2 | - | 237-266 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_01989.fasta.a3m.hhm.out.hhmakeBK2- |
| 7.3 | - | 5-40 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_01278.fasta.a3m.hhm.out.hhmakeBK2- |
| 7.7 | - | 135-164 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_03492.fasta.a3m.hhm.out.hhmakeBK2- |
| 7.7 | - | 9-28 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_00561.fasta.a3m.hhm.out.hhmakeBK2- |
| 8.2 | - | 233-262 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_04010.fasta.a3m.hhm.out.hhmakeBK2- |
| 8.3 | - | 9-28 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_03809.fasta.a3m.hhm.out.hhmakeBK2- |
| 9.1 | - | 9-26 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_02053.fasta.a3m.hhm.out.hhmakeBK2- |
| 9.9 | - | 9-28 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_00931.fasta.a3m.hhm.out.hhmakeBK2- |
| 15.5 | - | 364-406 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_02744.fasta.a3m.hhm.out.hhmakeBK2- |
| 18.0 | - | 6-23 | - 3JS8:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_02160.fasta.a3m.hhm.out.hhmakeBK2- |
| 99.9 | - | 18-190 | - 1I19:A | - /projet/mig/work/vmartin/D8/D8.batch4.20150203/RESULTS-BANK2/bdd8_03618.fasta.a3m.hhm.out.hhmakeBK2- |

★ *B. dorei* 175^T

Sorted by Prob > 95 and size cut off > 50 amino acid residues

Véronique Martin

99.4 % 70 aa bacdor_01751.fasta.a3m.hhm.out.hhmakeBK1 → Hypothetical protein
99.1 % 75 aa bacdor_01928.fasta.a3m.hhm.out.hhmakeBK1 → L-aspartate oxidase
99.1 % 62 aa bacdor_02089.fasta.a3m.hhm.out.hhmakeBK1 → Succinate Dehydrogenase flavoprotein
97.5 % 61 aa bacdor_03886.fasta.a3m.hhm.out.hhmakeBK1 → Pyridine nucleotide oxido reductase

★ *B. dorei* 175^T

Sorted by Prob > 95 and size > 50 amino acid residues

99.9 % 172 aa bacdor_03524.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetyl muramate dehydrogenase

★ *B. dorei* 175^T Sorted by Prob > 95 and size cut off > 50 amino acid residues

99.4 % 70 aa baccor_01751.fasta.a3m.hhm.out.hhmakeBK1 → Hypothetical protein
99.1 % 75 aa baccor_01928.fasta.a3m.hhm.out.hhmakeBK1 → L-aspartate oxidase
99.1 % 62 aa baccor_02089.fasta.a3m.hhm.out.hhmakeBK1 → Succinate Dehydrogenase flavoprotein
97.5 % 61 aa baccor_03886.fasta.a3m.hhm.out.hhmakeBK1 → Pyridine nucleotide oxido reductase

★ *B. dorei* 175^T Sorted by Prob > 95 and size > 50 amino acid residues

99.9 % 172 aa baccor_03524.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetyl muramate dehydrogenase

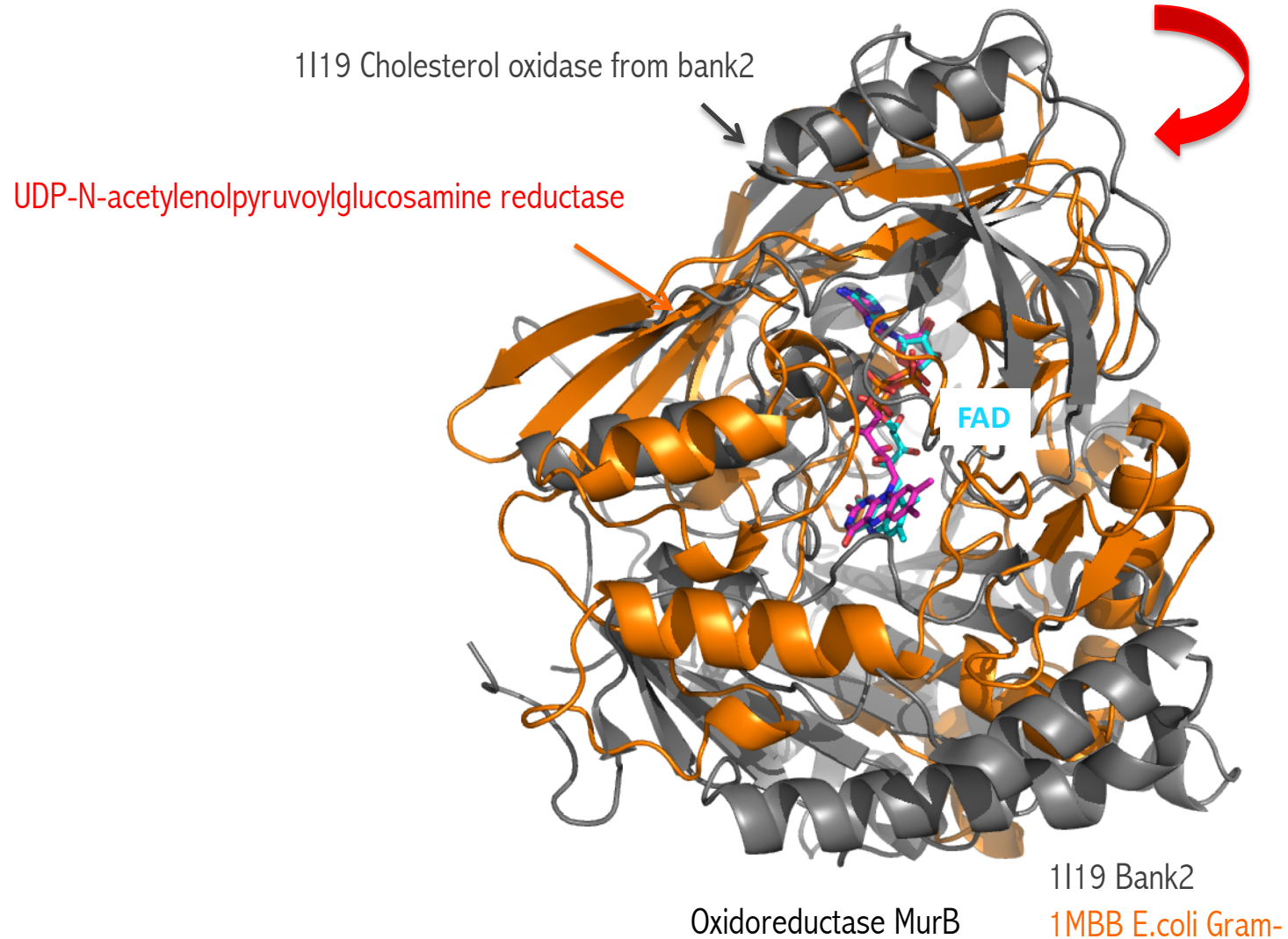
=

99.9 % 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase

★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

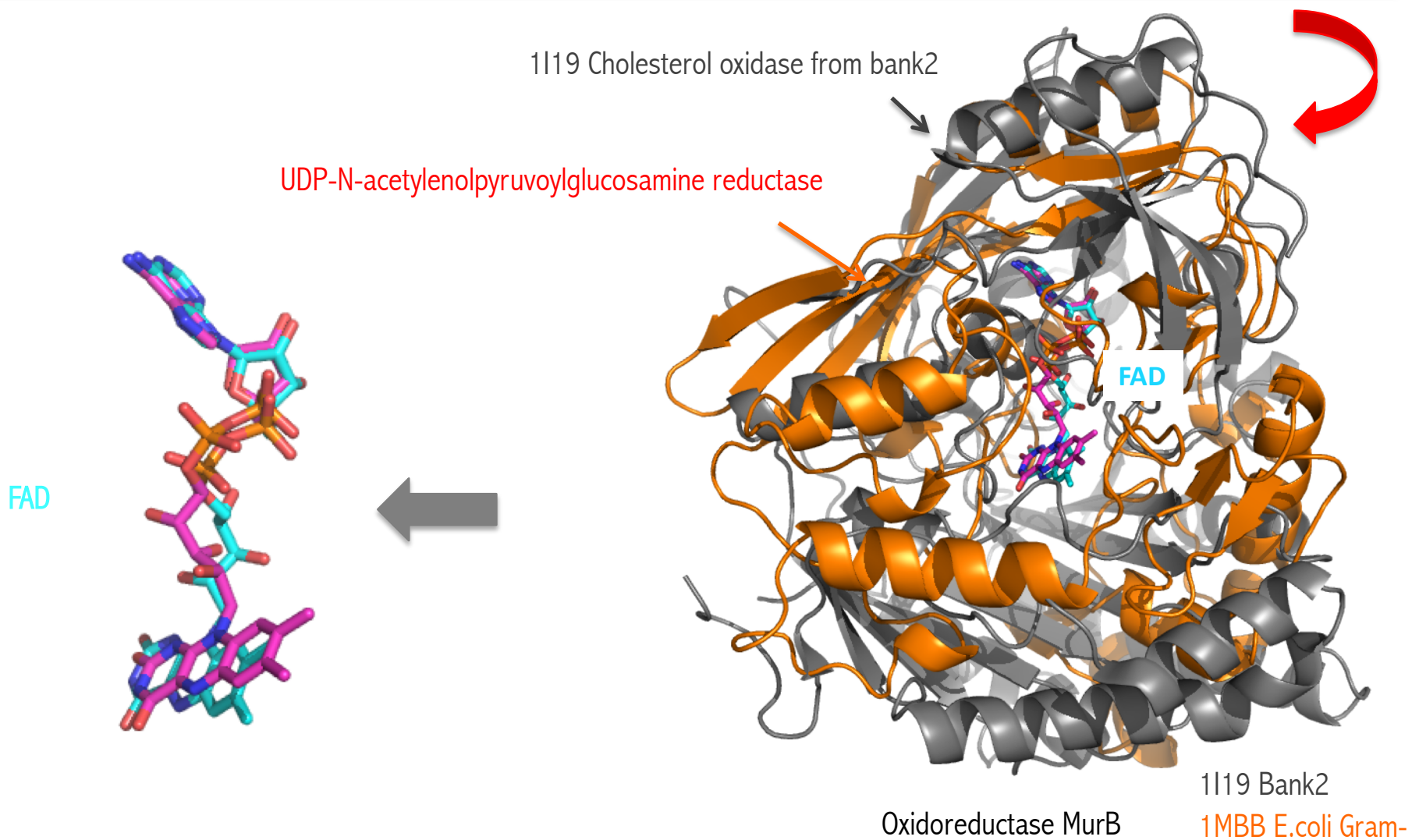
99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

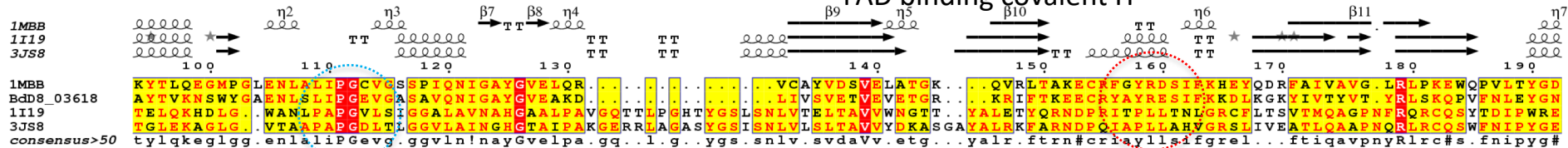
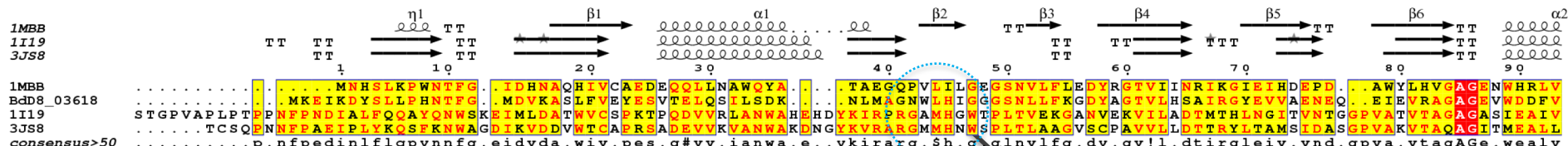
99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



★ *B. dorei*D8

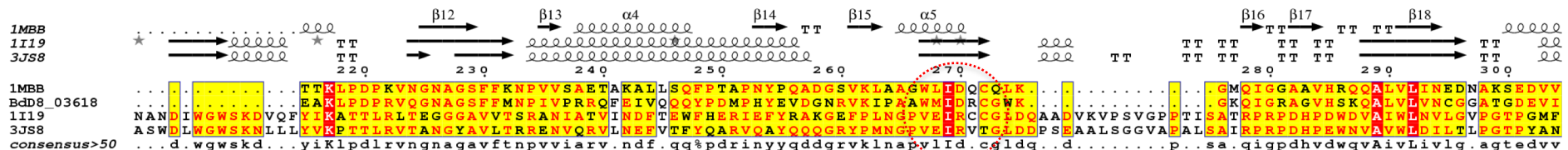
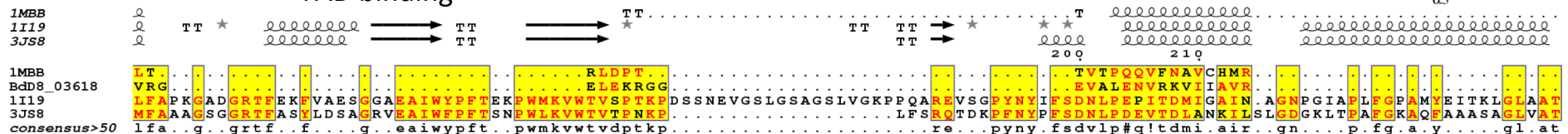
Sorted by Prob > 95 and size cut off > 50 amino acid residues

99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase

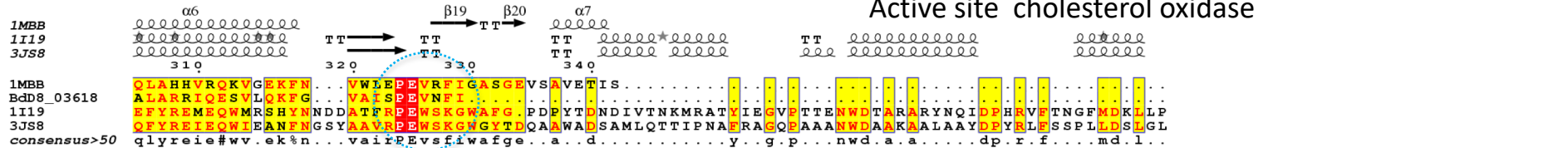


FAD binding

Active site oxido-reductase



Active site cholesterol oxidase

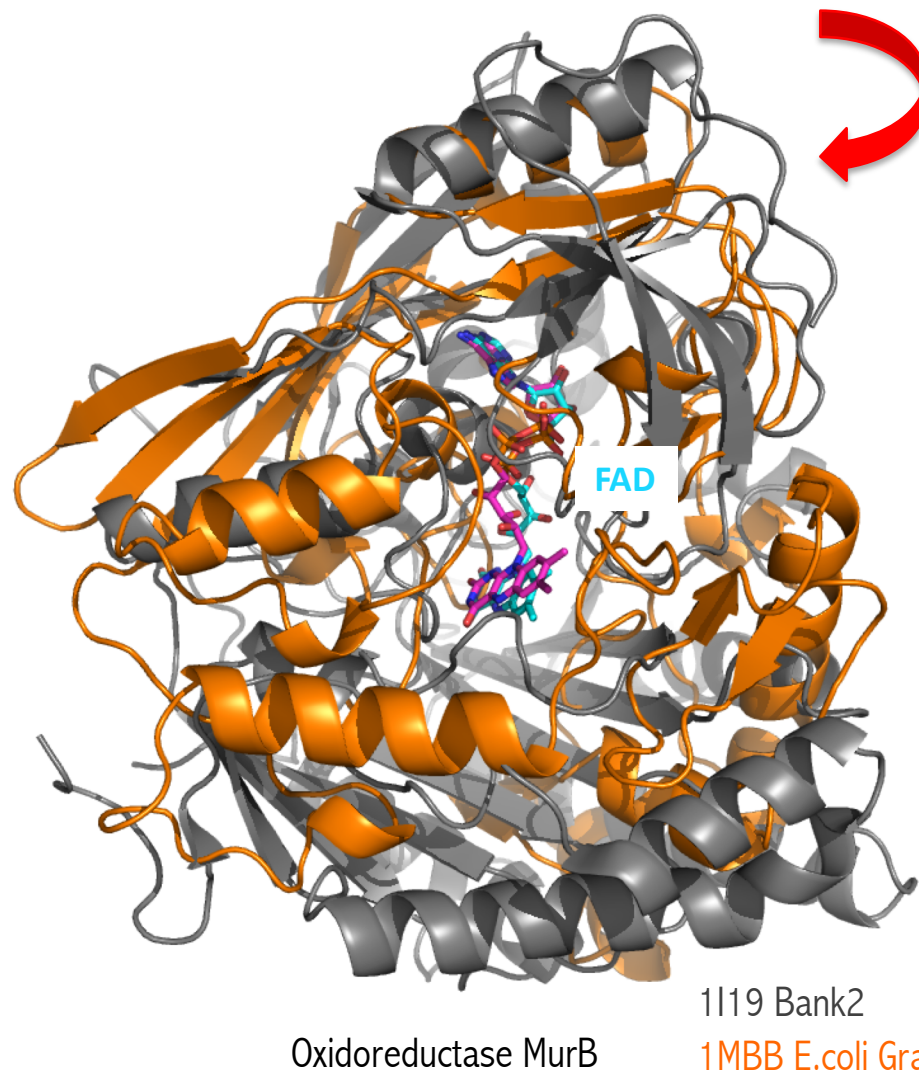
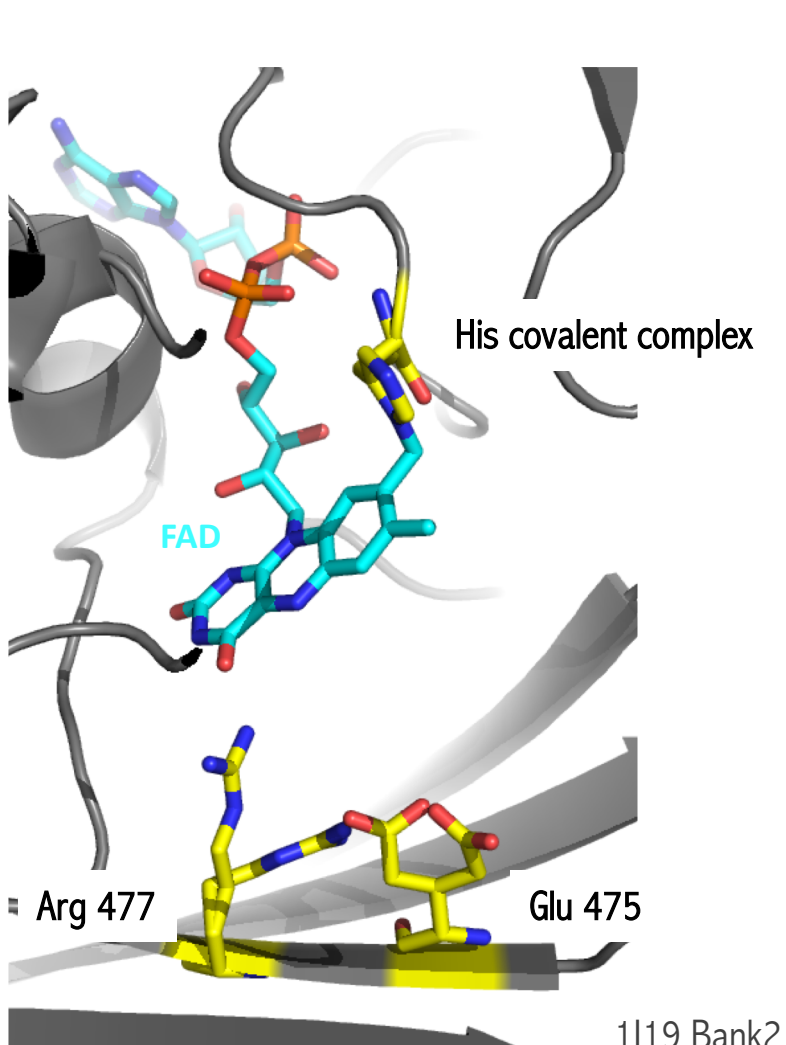


FAD binding

★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

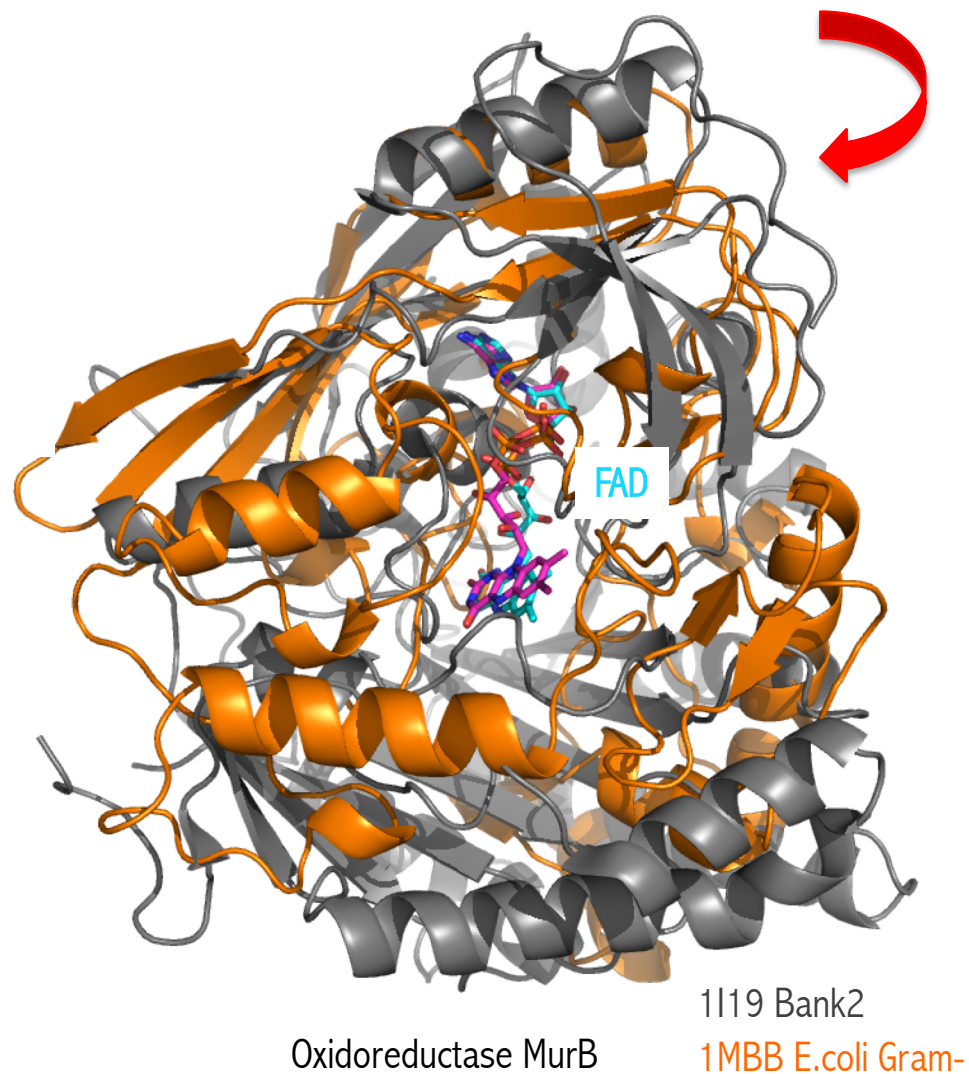
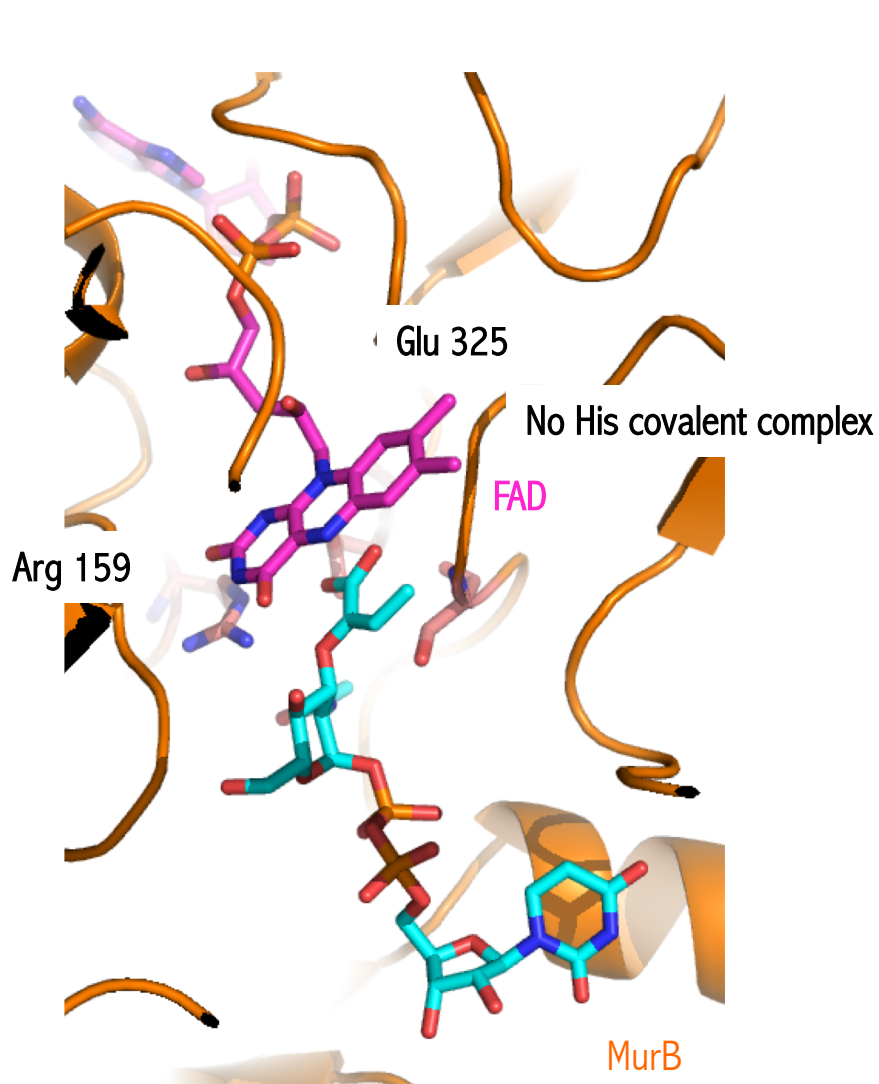
99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

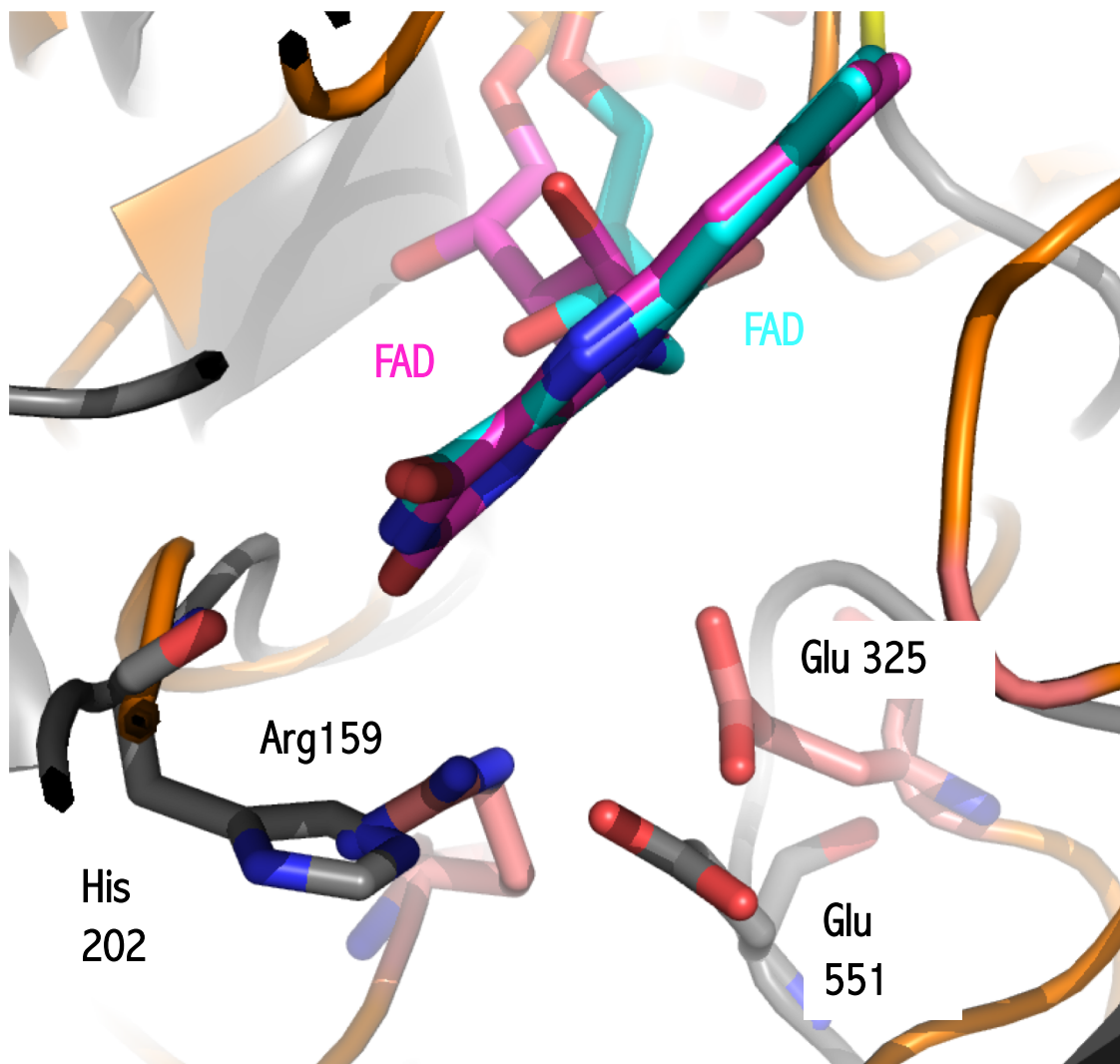
99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

99.9 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



1119 Bank2
1MBB MurB

★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

- 99.4 % 70 aa baccor_01751.fasta.a3m.hhm.out.hhmakeBK1 → Hypothetical protein
- 99.1 % 75 aa baccor_01928.fasta.a3m.hhm.out.hhmakeBK1 → L-aspartate oxidase
- 99.1 % 62 aa baccor_02089.fasta.a3m.hhm.out.hhmakeBK1 → Succinate Dehydrogenase flavoprotein
- 97.5 % 61 aa baccor_03886.fasta.a3m.hhm.out.hhmakeBK1 → Pyridine nucleotide oxido reductase

★ *B. dorei* 175^T

Sorted by Prob > 95 and size > 50 amino acid residues

- 99.9 % 172 aa baccor_03524.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetyl muramate dehydrogenase

=

- 99.9 % 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



Cloning and expression tests in progress.

Then, purification, activity test and crystallization assays

Cholesterol oxidases are secreted. This contains a signal peptide.

★ *B. dorei*D8

Sorted by Prob > 95 and size cut off > 50 amino acid residues

- 99.4 % 70 aa baccor_01751.fasta.a3m.hhm.out.hhmakeBK1 → Hypothetical protein
- 99.1 % 75 aa baccor_01928.fasta.a3m.hhm.out.hhmakeBK1 → L-aspartate oxidase
- 99.1 % 62 aa baccor_02089.fasta.a3m.hhm.out.hhmakeBK1 → Succinate Dehydrogenase flavoprotein
- 97.5 % 61 aa baccor_03886.fasta.a3m.hhm.out.hhmakeBK1 → Pyridine nucleotide oxido reductase

★ *B. dorei* 175^T

Sorted by Prob > 95 and size > 50 amino acid residues

- 99.9 % 172 aa baccor_03524.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetyl muramate dehydrogenase

=

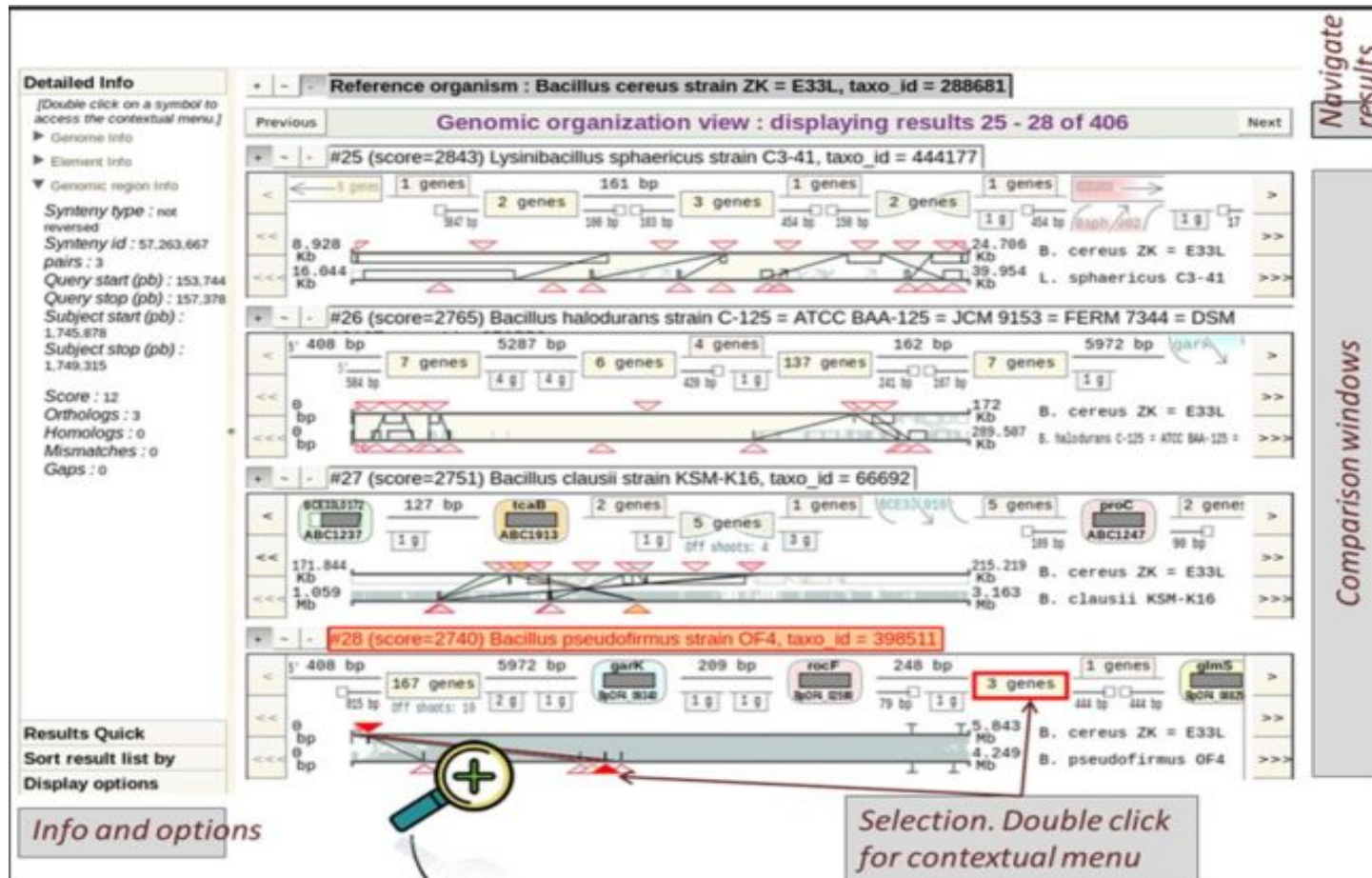
- 99.9 % 172 aa bdd8_003618.fasta.a3m.hhm.out.hhmakeBK2 → UDP-N-acetylenolpyruvoylglucosamine reductase



Analysis of synthenies by Thomas Lacroix using Insyght.

★ <http://genome.jouy.inra.fr/Insyght>

➡ Powerful tool for genes comparison and synteny inferences



Detailed Info
[Double click on a symbol to access the contextual menu.]
▶ Genome info
▶ Element info
▼ Genomic region info
Synteny type : not reversed
Synteny id : 57.263.667
pairs : 3
Query start (pb) : 153.744
Query stop (pb) : 157.378
Subject start (pb) : 1.745.878
Subject stop (pb) : 1.749.315
Score : 12
Orthologs : 3
Homologs : 0
Mismatches : 0
Gaps : 0

Results Quick
Sort result list by
Display options

Reference organism : Bacillus cereus strain ZK = E33L, taxo_id = 288681

Genomic organization view : displaying results 25 - 28 of 406

#25 (score=2843) Lysinibacillus sphaericus strain C3-41, taxo_id = 444177

#26 (score=2765) Bacillus halodurans strain C-125 = ATCC BAA-125 = JCM 9153 = FERM 7344 = DSM

#27 (score=2751) Bacillus clausii strain KSM-K16, taxo_id = 66692

#28 (score=2740) Bacillus pseudofirmus strain OF4, taxo_id = 398511

Navigation: Previous, Next, Navigate results

Comparison windows: Comparison windows

Info and options: Info and options

Selection: Selection. Double click for contextual menu

★ <http://genome.jouy.inra.fr/Insyght>

➡ Powerful tool for genes comparison and syntenic inferences

Syntenies : co-localization of homologous loci across species

Comparison 2 vs 2 with a reference genome.

No synteny. No clear information

The screenshot shows the Insyght web interface for comparing five Corynebacterium strains. The strains listed are:

- #1 Corynebacterium efficiens (strain: DSM 44549 = NBRC 100395 = JCM 11189 = YS-314 = AJ 12310)
- #3 Corynebacterium efficiens (strain: DSM 44549 = NBRC 100395 = JCM 11189 = YS-314 = AJ 12310)
- #4 Corynebacterium diphtheriae (strain: NCTC 13129 = ATCC 700971)
- #5 Agrobacterium vitis (strain: S4 = ATCC BAA-846)
- #6 Burkholderia cenocepacia (strain: H12424)

The interface displays gene models for various genes, including dnaA, recF, and others, across these strains. A 'Comparison windows' sidebar is visible on the right. A contextual menu is open at the bottom, showing options like 'Show other homologs' and 'View types'.

Info and options Contextual menu: browse multiple homologies per gene

★ Direct introduction of the COPROSTANOL standard by DIMS-MS/MS

Samples resuspended in 500 mL of Chloroform/ Methanol (50/50)

Diluted to 1/100eme before analysis

- Ions detection corresponding to COPROSTANOL
- DIMS-MS/MS method in positive ionisation mode

Catherine. Juste
Fabienne Beguet-Crespel

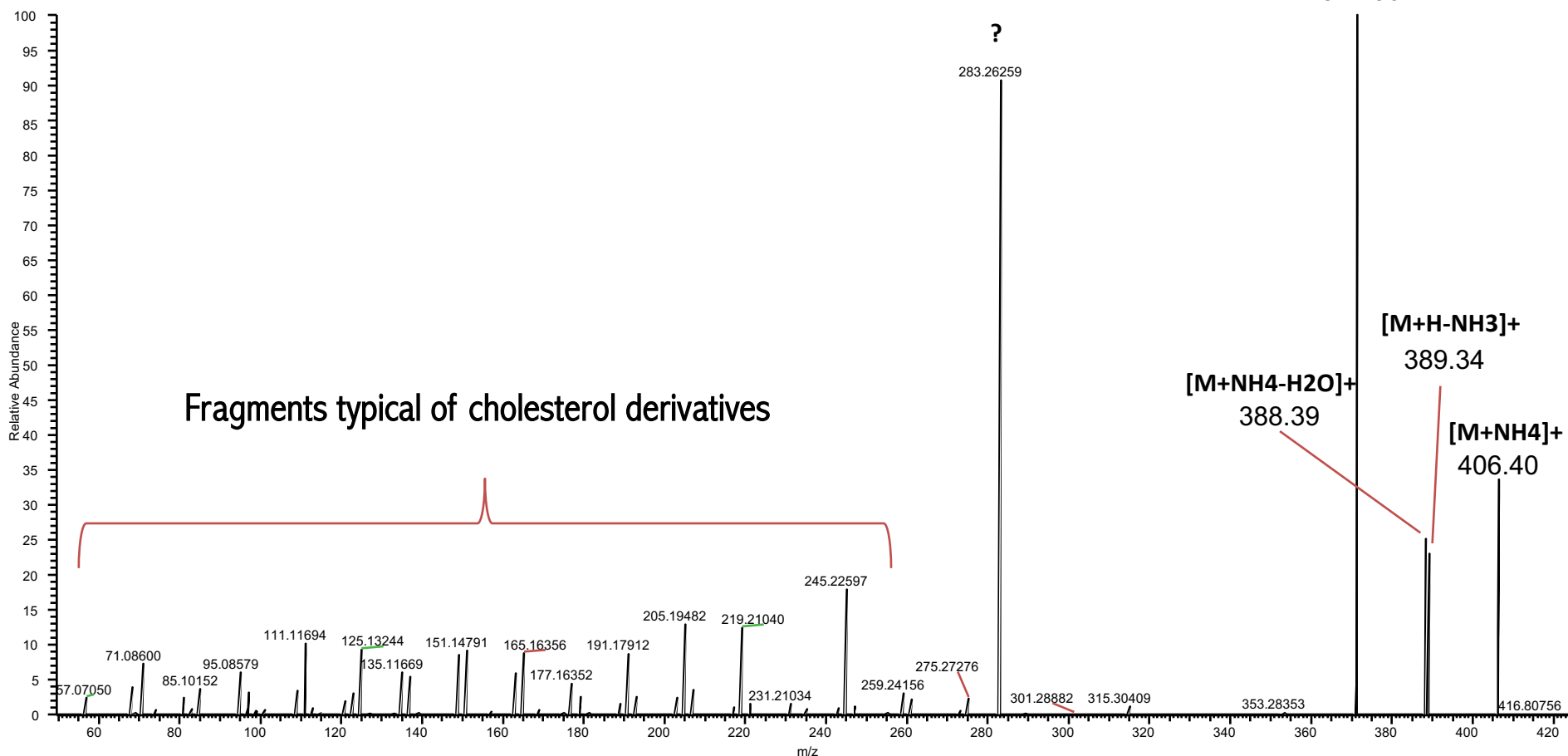
Coprostanol

$[M+H-H_2O]^+$
371.36

$[M+H-NH_3]^+$
389.34

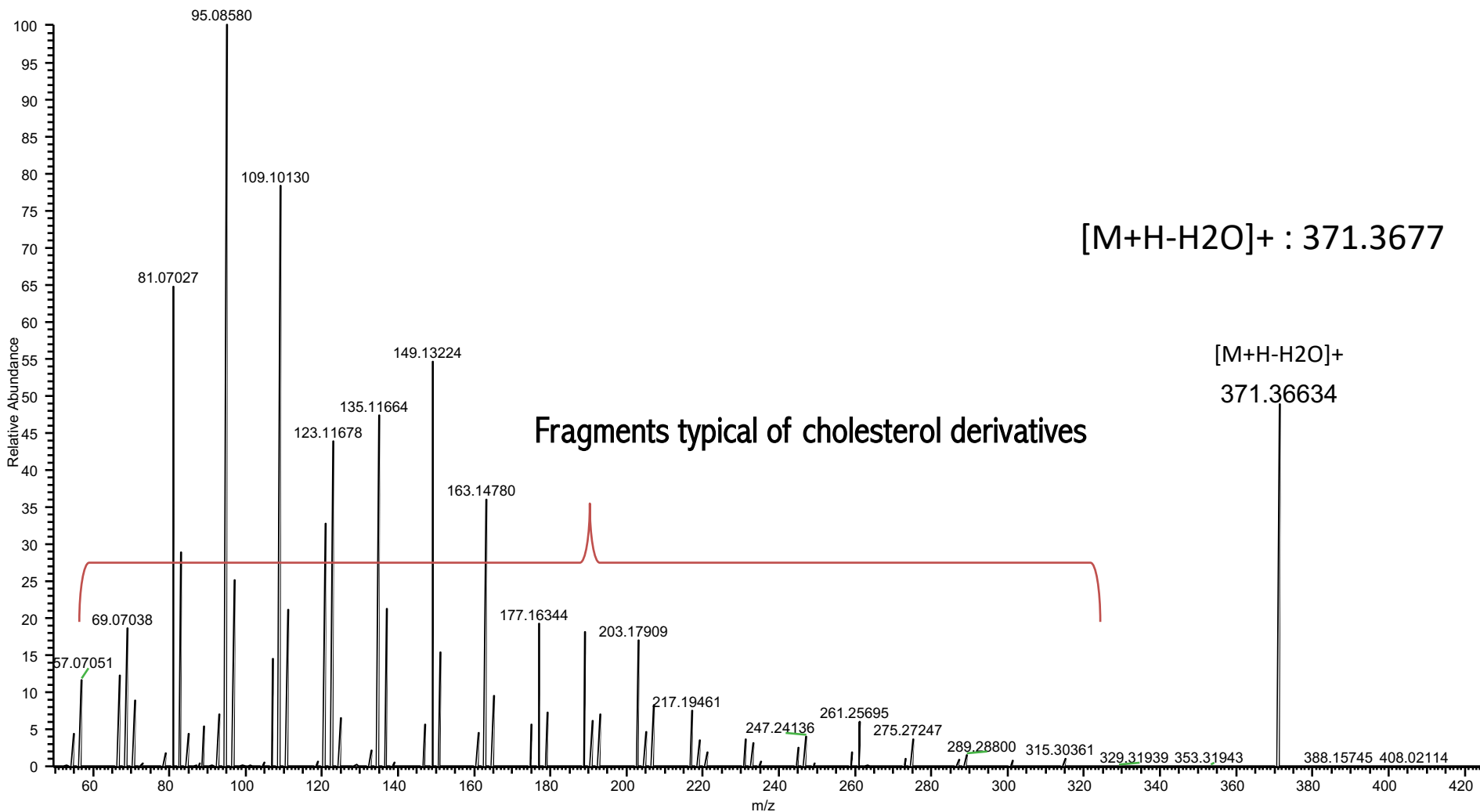
$[M+NH_4-H_2O]^+$
388.39

$[M+NH_4]^+$
406.40



- Ions detection corresponding to COPROSTANOL
- DIMS-MS/MS method in positive ionisation mode

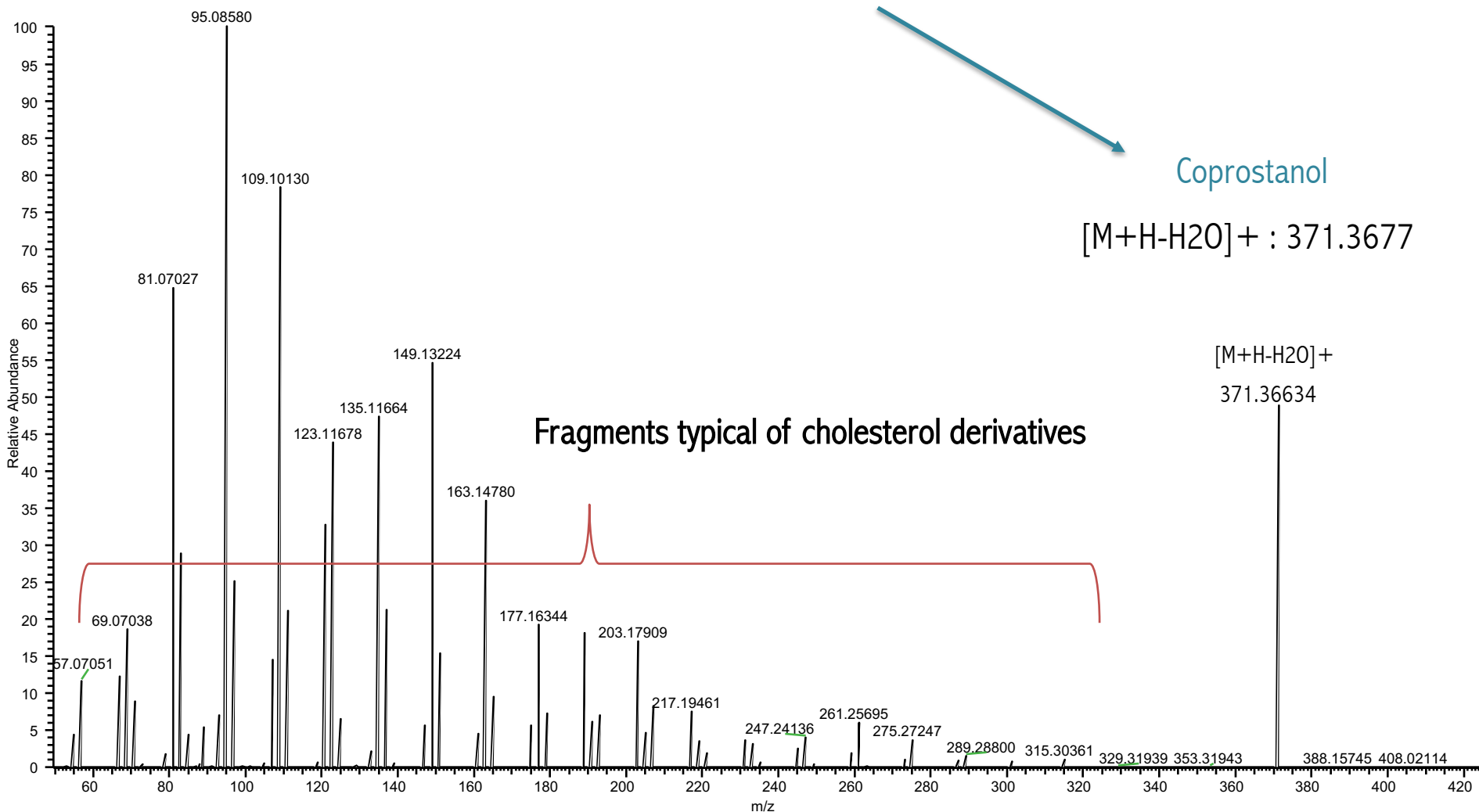
Catherine. Juste
Fabienne Beguet-Crespel



- Ions detection corresponding to COPROSTANOL
- DIMS-MS/MS method in positive ionisation mode

Catherine. Juste
Fabienne Beguet-Crespel

Coprostanol is present & preferentially detected



- ★ Analysis of candidates- Comparison within cholesterol degrading genomes and others
Comparison between *E. coprostanoligenes* (2017), *B. Dorei D8*, *Parabacteroides distasonis* shown to lower cholesterolemia while producing coprostanol. PhD thesis Aline Potiron.
- ★ Annotation comparator's analysis
- ★ Genomic organisation's analysis



One hit Interesting: Analysis in progress with Thomas Lacroix.
Present in degrading bacteria, absent in non degrading bacteria



Annotated as lipolytic protein G-D-S-L family, phospholipase, major facilitator superfamily

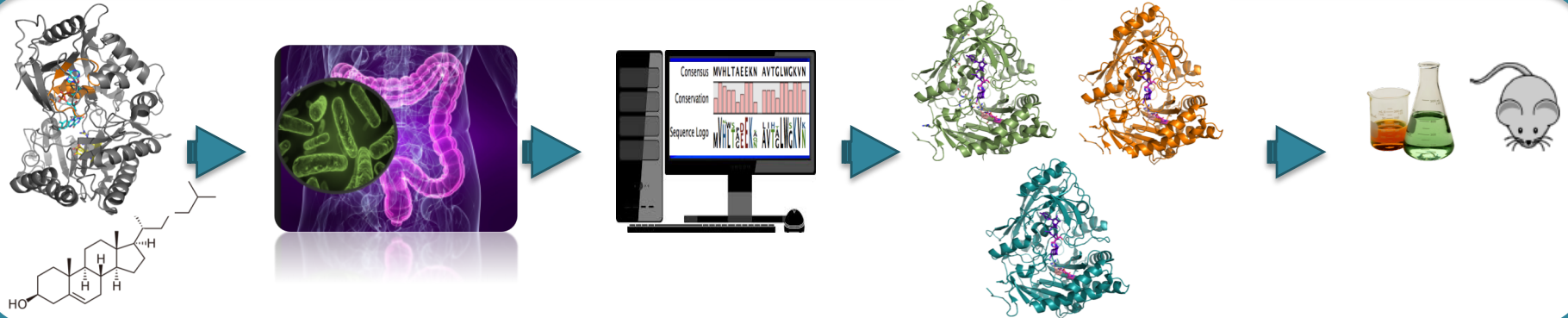


Cholesterol oxidase with a Ser His Asp conserved catalytic site plus the oxyanionic hole identified

Homology modeling and docking of cholesterol in progress

- ➔ No significant difference between the two genomes. Similar enzymes are found.
- ➔ Transcriptomic analysis of both chromosome and plasmids of *B. dorei* 175^T & *B. dorei* D8?
- ➔ Activity test and 3D structure validation of our cholesterol-oxidase hit in bank2.
- ➔ Genomic & transcriptomic analysis of both chromosome & plasmids of *E. coprostanoligenes*.
- ➔ Can we rule out some synergy among bacteria of the microbiota?
- ➔ Promising hit coming from entire genomes comparison: cholesterol esterase function and pathway.

Towards the Structural Screening of Microbial Ecosystems



Outline

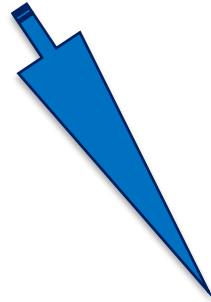
- Cholesterol conversion in the gut microbiota: the mystery enzyme(s) ?
- **MetaFoldScan project : 3D screening of gut microbiota**
- Conclusions and perspectives

The gut microbiota and our cells: intimate interaction

- 100 000 billions of bacteria
- 4 phyla
- ~1000 species



Pionneur



Lactobacille



Bifidobacteria



Faecalibacterium prausnitzii

« Symbiontes »



Clostridium difficile

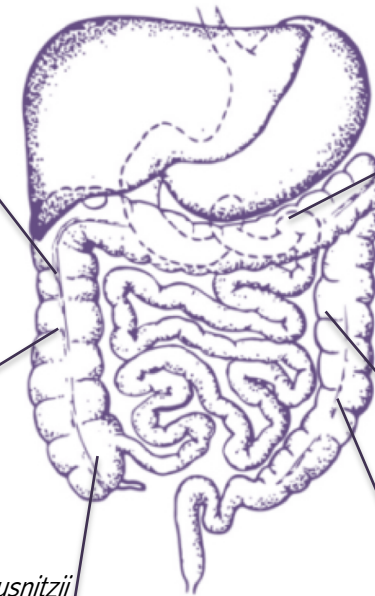


Enterococcus faecalis



Campylobacter

« Pathobiontes »



Catalog of 10 millions genes



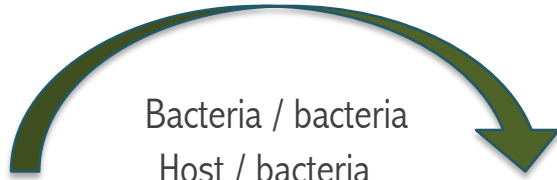
Universel : core genome

Identity card

Bidirectional dialog: language based on signaling

The gut microbiota and our cells: intimate interaction

Dialog

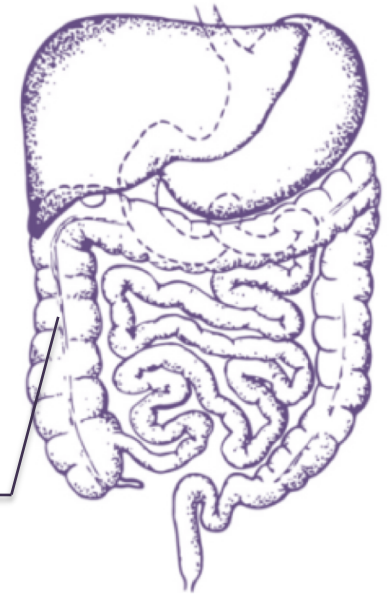


Gene diversity
Composition in bacteria

Symbiose

Health markers

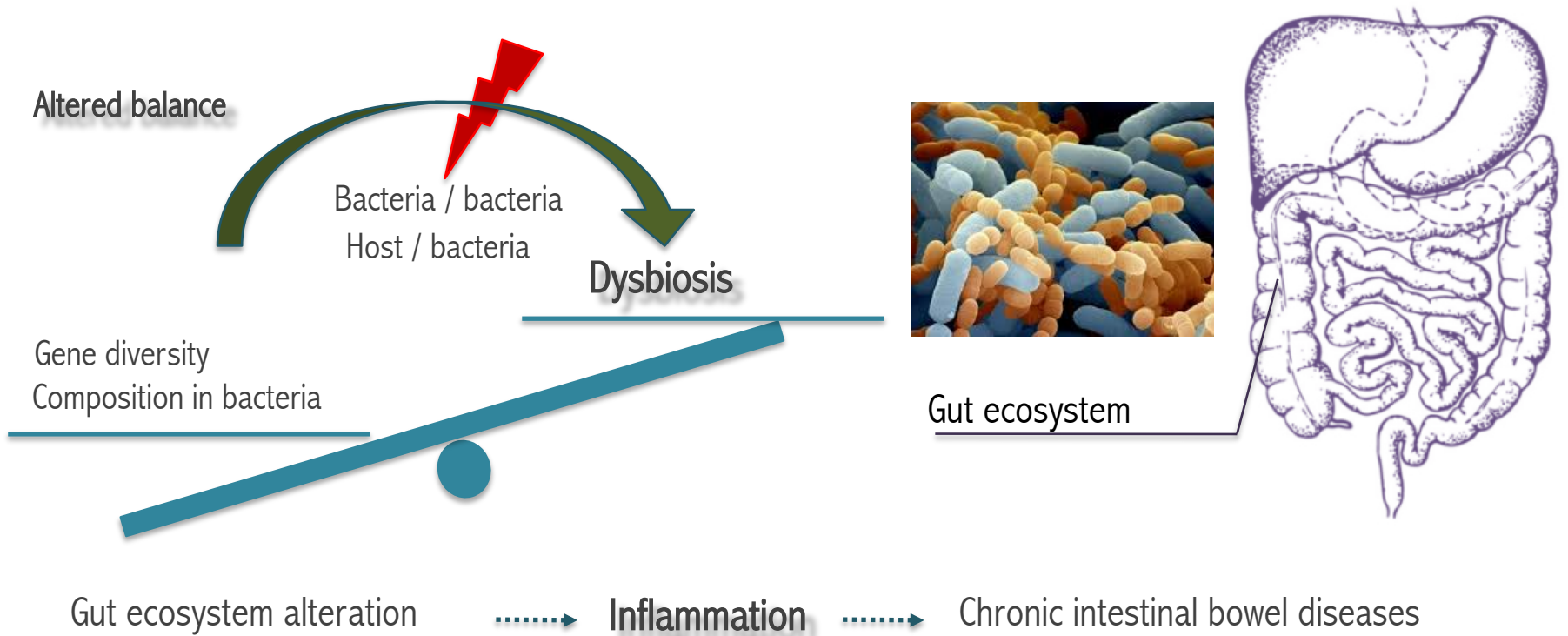
Homeostasis



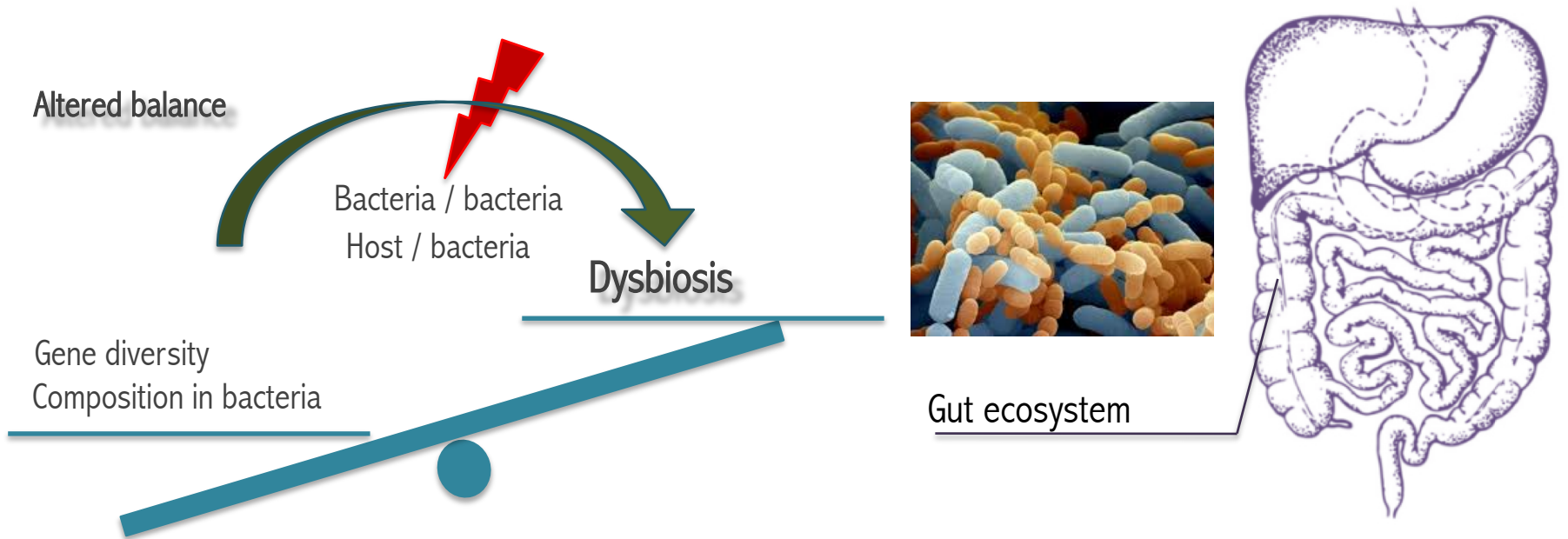
Gut ecosystem

Key interface cells/food/bacteria

Decipher the molecular mechanisms responsible for this dialog



Decipher the molecular mechanisms responsible for this dialog



Gut ecosystem alteration Inflammation Chronic intestinal bowel diseases

Catalogue of 10 millions of genes

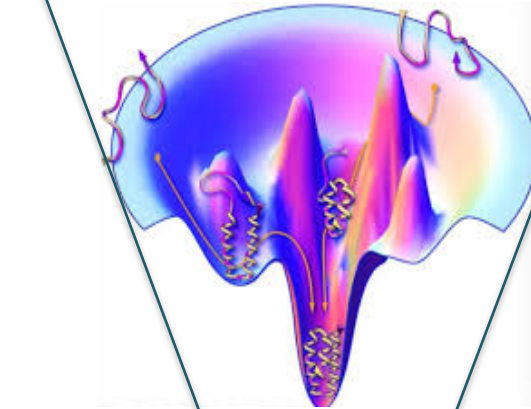
Metadata to explore and target



Investigate the molecular mechanisms of this balance

- - More than 10 millions prokaryotic genes
- - 40% protein sequences of unknown function
- - Function can be inferred from structure
- - Assessing the fold is possible
- - Fishing out the right fold is crucial !

126,000 known structures



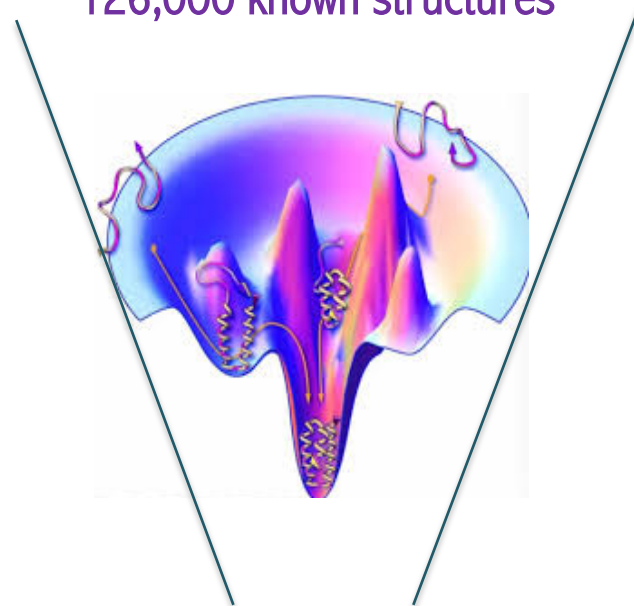
Only 1400 3D folds

- - More than 10 millions prokaryotic genes
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- - Function can be inferred from structure
- - Assessing the fold is possible
- - Fishing out the right fold is crucial !



Fold prediction informs on the function

126,000 known structures



Only 1400 3D folds

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- - Assessing the fold is possible
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Fold prediction informs on the function

Metagenomic



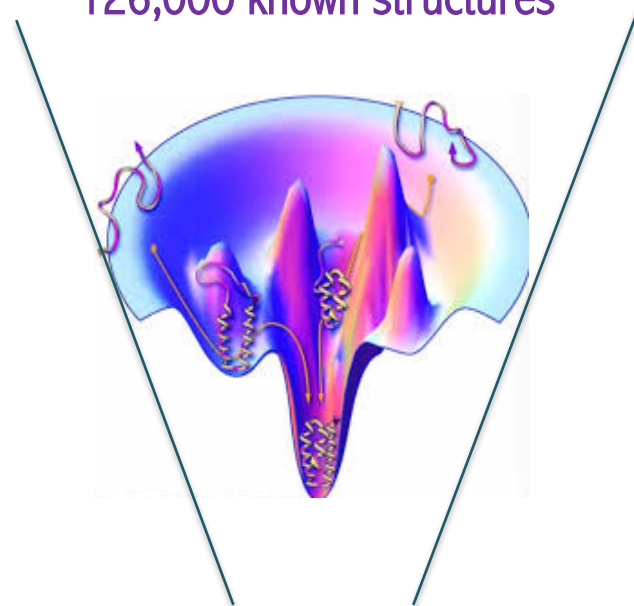
Functional metagenomic

Catalogues of sequences

Fold detection and prediction accurate

Computer compatible with big data

126,000 known structures



Only 1400 3D folds

- - More than 10 millions prokaryotic genes
- - 40% protein sequences of unknown function
- - Function can be inferred from structure
- - Assessing the fold is possible
- - Fishing out the right fold is crucial !



Fold prediction informs on the function

Metagenomic



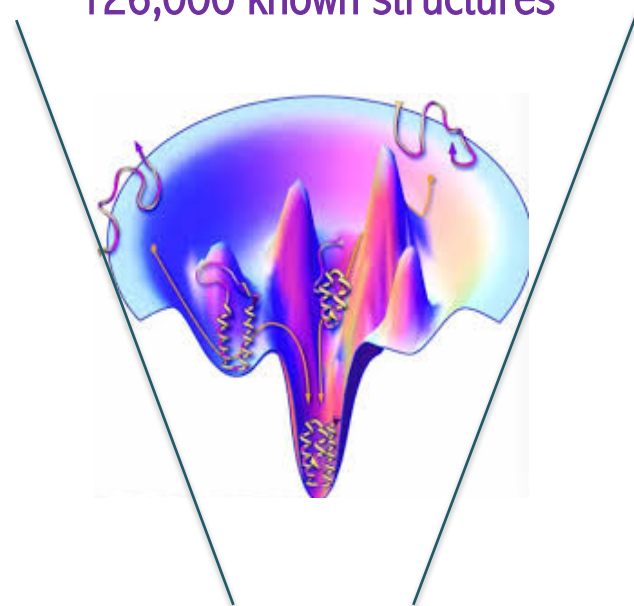
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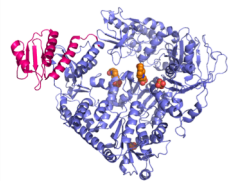
126,000 known structures



Only 1400 3D folds

Comprehensive detection and 3D modeling = StructurOmics

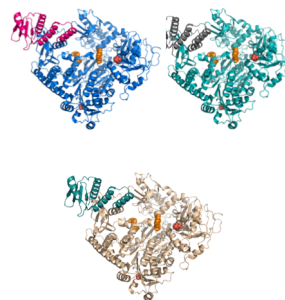
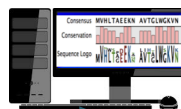
Target protein



Relevance



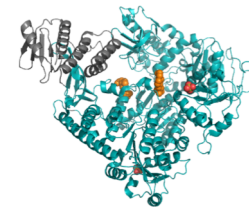
Scan microbiota



Hits identification



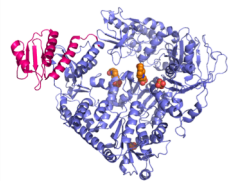
Validation



Biomarker

- Biological property
- Therapeutical interest
- Structural challenge

Target protein

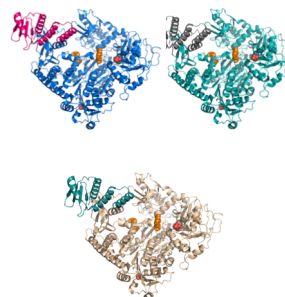
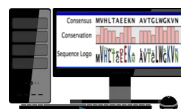


Relevance

- Biological property
- Therapeutical interest
- Structural challenge



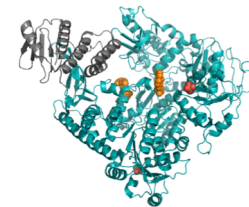
Scan microbiota



Hits identification



Validation



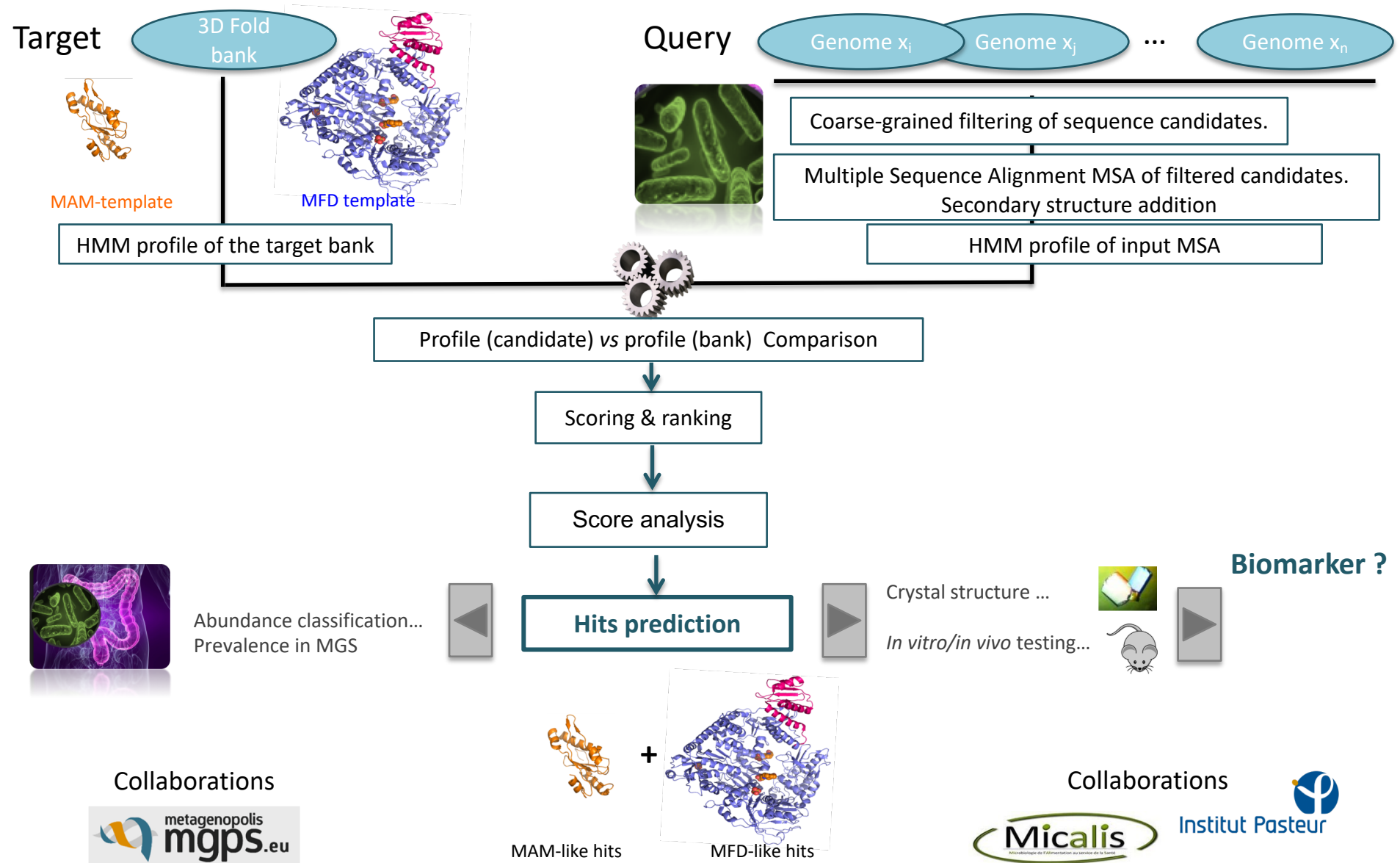
Biomarker

Preliminary data:

- Histone-like Nucleotide Structuring protein in *Enterococcus faecalis*
- Penicillin binding protein in *Bacillus subtilis*
- Cholesterol oxidase for *Bacteroides dorei* and *dorei D8*



- ❖ Experimental data in progress
- ❖ Scaling up to full microbiota requested



To tackle both methodological and biological issues

MAM

JM. Chatel

Microbial Anti-inflammatory Molecule

- Unknown biological function
- Small 180 residues
- Structurally not characterized
- Rare, found in *F. prausnitzii*
- *F. Prausnitzii* 5% of the microbiota, IBD
Crohn Disease, Ulcerative Colitis,
- Fishing from homology modeling
- Associated with probiotic properties

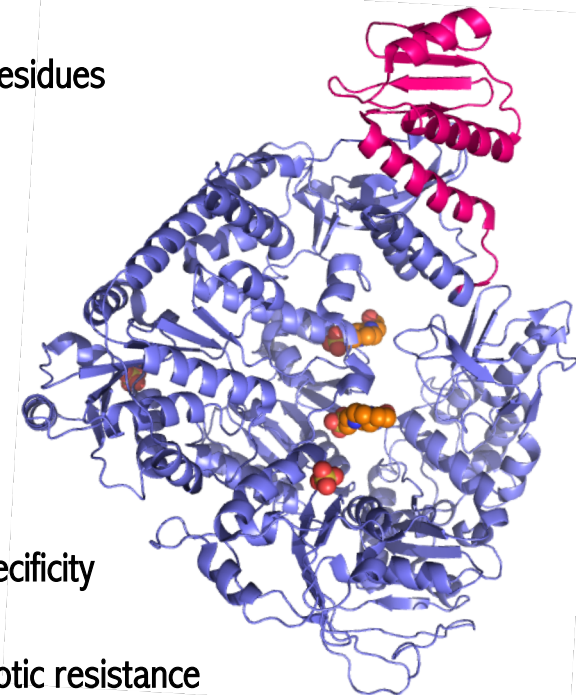


MFD

N. Rama Rao

Mutation Frequency Decline

- Involved in bacterial DNA repair, in virulence
- Multi-domains 1150 residues
- Solved structure
- Ubiquitous, Gram+/-
- Promiscuity
- Molecular basis of specificity
- Associated with antibiotic resistance



Cohort of 124 European individuals, healthy and obese, danish and spanish
 Nordic & Mediterranean faecal DNA

Vol 464 | 4 March 2010 | doi:10.1038/nature08821

nature

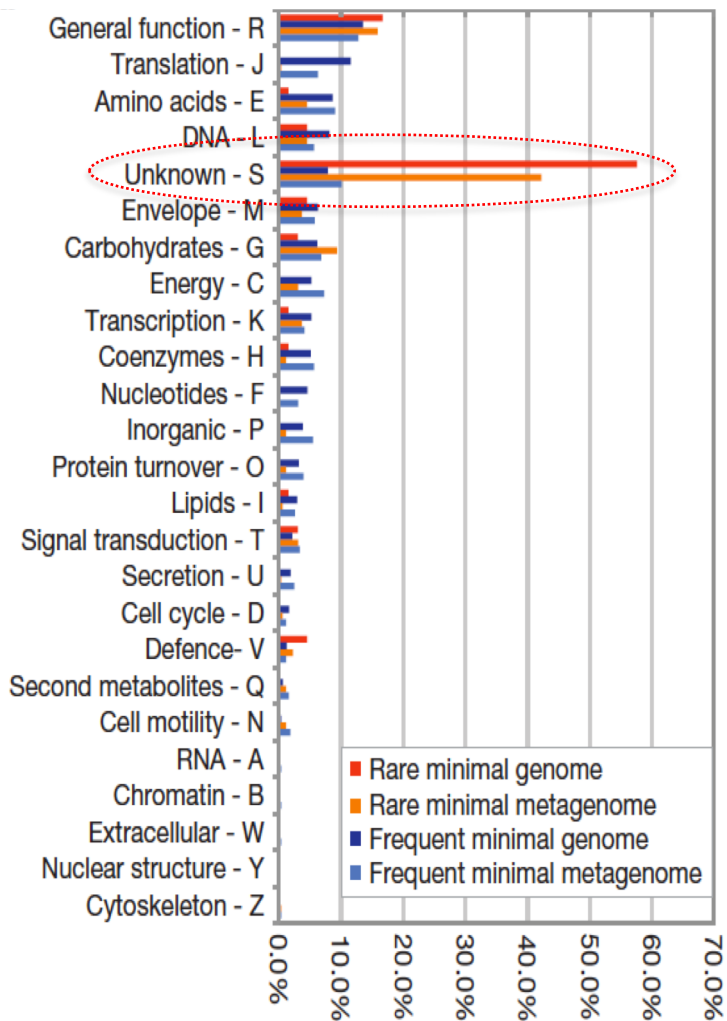
A human gut microbial gene catalogue established by metagenomic sequencing

Junjie Qin^{1*}, Ruiqiang Li^{1*}, Jeroen Raes^{2,3}, Manimozhiyan Arumugam², Kristoffer Solvsten Burgdorf⁴, Chaysavanh Manichanh⁵, Trine Nielsen⁴, Nicolas Pons⁶, Florence Levenez⁶, Takuji Yamada², Daniel R. Mende², Junhua Li^{1,7}, Junming Xu¹, Shaochuan Li¹, Dongfang Li^{1,8}, Jianjun Cao¹, Bo Wang¹, Huiqing Liang¹, Huisong Zheng¹, Yinlong Xie^{1,7}, Julien Tap⁶, Patricia Lepage⁶, Marcelo Bertalan⁹, Jean-Michel Batto⁶, Torben Hansen⁴, Denis Le Paslier¹⁰, Allan Linneberg¹¹, H. Bjørn Nielsen⁹, Eric Pelletier¹⁰, Pierre Renault⁶, Thomas Sicheritz-Ponten⁹, Keith Turner¹², Hongmei Zhu¹, Chang Yu¹, Shengting Li¹, Min Jian¹, Yan Zhou¹, Yingrui Li¹, Xiuqing Zhang¹, Songgang Li¹, Nan Qin¹, Huanming Yang¹, Jian Wang¹, Søren Brunak⁹, Joel Doré⁶, Francisco Guarner⁵, Karsten Kristiansen¹³, Oluf Pedersen^{4,14}, Julian Parkhill¹², Jean Weissenbach¹⁰, MetaHIT Consortium†, Peer Bork², S. Dusko Ehrlich⁶ & Jun Wang^{1,13}

To understand the impact of gut microbes on human health and well-being it is crucial to assess their genetic potential. Here we describe the Illumina-based metagenomic sequencing, assembly and characterization of 3.3 million non-redundant microbial genes, derived from 576.7 gigabases of sequence, from faecal samples of 124 European individuals. The gene set, ~150 times larger than the human gene complement, contains an overwhelming majority of the prevalent (more frequent) microbial genes of the cohort and probably includes a large proportion of the prevalent human intestinal microbial genes. The genes are largely shared among individuals of the cohort. Over 99% of the genes are bacterial, indicating that the entire cohort harbours between 1,000 and 1,150 prevalent bacterial species and each individual at least 160 such species, which are also largely shared. **We define and describe the minimal gut metagenome and the minimal gut bacterial genome in terms of functions present in all individuals and most bacteria, respectively.**

- - 3,3 millions of genes , 99,1 % of bacterial origin
- - Minimal gut bacterial genome : 1,000 to 1,150 prevalent bacterial species
- - Each individual harbors at least 160 such species

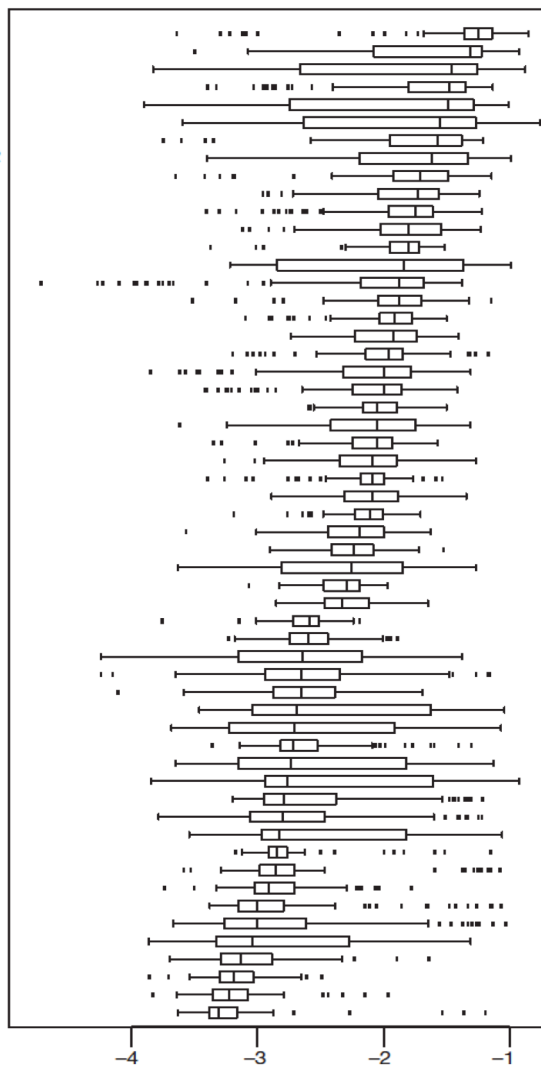
Minimal gut genome: functions necessary for a bacterium to thrive in a gut context



Core genome: 57 bacterial species present in > 90 of individuals (prevalence and consensus)

➔ Mostly Bacteroidetes & Firmicutes

- Bacteroides uniformis*
- Alistipes putredinis*
- Parabacteroides merdae*
- Dorea longicatena*
- Ruminococcus bromii* L2-63
- Bacteroides caccae*
- Clostridium* sp. SS2-1
- Bacteroides thetaiotaomicron* VPI-5482
- Eubacterium hallii*
- Ruminococcus torques* L2-14
- Unknown sp. SS3 4
- Ruminococcus* sp. SR1 5
- Faecalibacterium prausnitzii* SL3 3
- Ruminococcus lactaris*
- Collinsella aerofaciens*
- Dorea formicigenerans*
- Bacteroides vulgatus* ATCC 8482
- Roseburia intestinalis* M50 1
- Bacteroides* sp. 2_1_7
- Eubacterium siraeum* 70 3
- Parabacteroides distasonis* ATCC 8503
- Bacteroides* sp. 9_1_42FAA
- Bacteroides ovatus*
- Bacteroides* sp. 4_3_47FAA
- Bacteroides* sp. 2_2_4
- Eubacterium rectale* M104 1
- Bacteroides xylanisolvens* XB1A
- Coprococcus comes* SL7 1
- Bacteroides* sp. D1
- Bacteroides* sp. D4
- Eubacterium ventriosum*
- Bacteroides dorei*
- Ruminococcus obeum* A2-162
- Subdoligranulum variabile*
- Bacteroides capillosus*
- Streptococcus thermophilus* LMD-9
- Clostridium leptum*
- Holdemania filiformis*
- Bacteroides stercoris*
- Coprococcus eutactus*
- Clostridium* sp. M62 1
- Bacteroides eggerthii*
- Butyrivibrio crossotus*
- Bacteroides fingoldii*
- Parabacteroides johnsonii*
- Clostridium* sp. L2-50
- Clostridium nexile*
- Bacteroides pectinophilus*
- Anaerotruncus colihominis*
- Ruminococcus gnavus*
- Bacteroides intestinalis*
- Bacteroides fragilis* 3_1_12
- Clostridium asparagiforme*
- Enterococcus faecalis* TX0104
- Clostridium scindens*
- Blautia hansenii*



Relative abundance (log₁₀)

| | | 265 | 9,00E-93 | 100.00 | 100 | 100 | 135 | 0 | 0 | 1 | 135 | 1 | 135 | |
|----|--|------------------------|----------|----------|--------|---------|--------|----------|---------|---|-----|-----|------|------|
| | | bitscore | evalue | pident | qcovs | qcovhsp | length | mismatch | gapopen | | | | | |
| 1 | Faecalibacterium_prausnitzii_A2-165 NCBI | WP_005932151.1 | | | | | | | | | | | | |
| 2 | MAM_wt | tr R6SBT0 R6SBT0_9BACE | 23.5 | 6.4 | 47.826 | 17 | 17 | 23 | 10 | 1 | 14 | 36 | 624 | 644 |
| 3 | MAM_wt | tr D4JK97 D4JK97_9FIRM | 28.5 | 0.013 | 45.000 | 15 | 15 | 20 | 11 | 0 | 1 | 20 | 9 | 28 |
| 4 | MAM_wt | tr R7EPB3 R7EPB3_9BACE | 26.6 | 0.55 | 41.667 | 24 | 24 | 36 | 18 | 1 | 52 | 84 | 185 | 220 |
| 5 | MAM_wt | tr C3QB13 C3QB13_9BACE | 29.3 | 0.063 | 40.625 | 24 | 24 | 32 | 19 | 0 | 88 | 119 | 95 | 126 |
| 6 | MAM_wt | tr D4C9H4 D4C9H4_9CLOT | 27.3 | 0.30 | 40.541 | 27 | 27 | 37 | 21 | 1 | 73 | 108 | 128 | 164 |
| 7 | MAM_wt | tr D4KBR2 D4KBR2_9FIRM | 75.9 | 1.02e-19 | 35.570 | 98 | 98 | 149 | 72 | 4 | 1 | 132 | 1 | 142 |
| 8 | Faecalibacterium_prausnitzii_SL3_3 | tr R7B146 R7B146_9BACE | 25.0 | 1.2 | 34.694 | 36 | 36 | 49 | 31 | 1 | 15 | 63 | 119 | 166 |
| 9 | MAM_wt | tr D4L4M8 D4L4M8_9FIRM | 24.3 | 1.9 | 34.483 | 21 | 21 | 29 | 19 | 0 | 75 | 103 | 516 | 544 |
| 10 | MAM_wt | tr C3QPR5 C3QPR5_9BACE | 25.8 | 1.1 | 34.375 | 24 | 24 | 32 | 21 | 0 | 88 | 119 | 95 | 126 |
| 11 | MAM_wt | tr R5TKN8 R5TKN8_9FIRM | 26.2 | 0.50 | 34.286 | 26 | 26 | 35 | 22 | 1 | 48 | 82 | 205 | 238 |
| 12 | MAM_wt | tr C0X1L4 C0X1L4_ENTFL | 26.6 | 0.39 | 34.211 | 27 | 27 | 38 | 24 | 1 | 14 | 50 | 150 | 187 |
| 13 | MAM_wt | tr D1PS54 D1PS54_9FIRM | 24.6 | 2.2 | 34.043 | 34 | 34 | 47 | 27 | 3 | 44 | 89 | 105 | 148 |
| 14 | MAM_wt | tr R6P689 R6P689_9CLOT | 24.3 | 1.8 | 33.333 | 27 | 27 | 36 | 24 | 0 | 56 | 91 | 98 | 133 |
| 15 | MAM_wt | tr A6LD60 A6LD60_PARD8 | 24.3 | 4.4 | 33.333 | 36 | 36 | 57 | 26 | 2 | 31 | 78 | 382 | 435 |
| 16 | MAM_wt | tr B0G4V7 B0G4V7_9FIRM | 25.4 | 1.1 | 32.500 | 29 | 29 | 40 | 26 | 1 | 68 | 106 | 150 | 189 |
| 17 | MAM_wt | tr C0D4Q5 C0D4Q5_9FIRM | 25.0 | 3.3 | 32.394 | 53 | 53 | 71 | 42 | 3 | 16 | 86 | 189 | 253 |
| 18 | MAM_wt | tr R7JPH0 R7JPH0_9BACT | 24.6 | 1.3 | 32.258 | 23 | 23 | 31 | 21 | 0 | 6 | 36 | 123 | 153 |
| 19 | MAM_wt | tr E4VUA2 E4VUA2_BACF | 27.7 | 0.35 | 31.818 | 33 | 33 | 44 | 30 | 0 | 1 | 44 | 280 | 323 |
| 20 | MAM_wt | tr A7VJ13 A7VJ13_9CLOT | 25.0 | 1.3 | 31.507 | 53 | 53 | 73 | 44 | 3 | 2 | 72 | 121 | 189 |
| 21 | MAM_wt | tr B0NGV9 B0NGV9_CLOS | 26.2 | 0.68 | 31.373 | 38 | 38 | 51 | 33 | 1 | 4 | 54 | 61 | 109 |
| 22 | MAM_wt | tr Q5M488 Q5M488_STRT2 | 26.9 | 0.13 | 31.111 | 33 | 33 | 45 | 24 | 2 | 36 | 80 | 83 | 120 |
| 23 | MAM_wt | tr R5U2R3 R5U2R3_9BACE | 28.5 | 0.12 | 30.909 | 41 | 41 | 55 | 36 | 1 | 76 | 130 | 47 | 99 |
| 24 | MAM_wt | tr A4EBV5 A4EBV5_9ACTN | 27.3 | 0.21 | 30.612 | 34 | 34 | 49 | 30 | 2 | 57 | 102 | 292 | 339 |
| 25 | MAM_wt | tr B0P145 B0P145_9CLOT | 32.7 | 0.004 | 30.337 | 66 | 66 | 89 | 49 | 5 | 9 | 97 | 1241 | 1316 |
| 26 | Clostridium_sp._SS2-1 | tr D4LSX8 D4LSX8_9FIRM | 24.3 | 2.2 | 30.303 | 24 | 24 | 33 | 23 | 0 | 21 | 53 | 78 | 110 |
| 27 | MAM_wt | tr R7FZ59 R7FZ59_9FIRM | 25.0 | 0.99 | 30.159 | 41 | 41 | 63 | 37 | 1 | 23 | 78 | 160 | 222 |
| 28 | MAM_wt | tr B0PHX3 B0PHX3_9FIRM | 23.5 | 4.7 | 29.787 | 34 | 34 | 47 | 31 | 2 | 3 | 48 | 173 | 218 |
| 29 | MAM_wt | tr R6X7N3 R6X7N3_9PORP | 24.3 | 3.3 | 29.545 | 61 | 61 | 88 | 53 | 4 | 4 | 86 | 547 | 630 |
| 30 | MAM_wt | tr A5Z5Q6 A5Z5Q6_9FIRM | 27.7 | 0.19 | 29.412 | 47 | 47 | 68 | 42 | 2 | 66 | 128 | 345 | 411 |
| 31 | MAM_wt | tr B9YAA0 B9YAA0_9FIRM | 28.1 | 0.15 | 29.268 | 30 | 30 | 41 | 29 | 0 | 1 | 41 | 68 | 108 |
| 32 | MAM_wt | tr B7B871 B7B871_9PORP | 25.0 | 1.9 | 28.846 | 32 | 32 | 52 | 24 | 3 | 54 | 96 | 204 | 251 |
| 33 | MAM_wt | tr P96214 P96214_MYCTU | 25.4 | 1.4 | 28.571 | 38 | 38 | 56 | 35 | 1 | 52 | 102 | 61 | 116 |
| 34 | MAM_wt | tr R6LDT4 R6LDT4_9FIRM | 24.3 | 1.5 | 28.070 | 39 | 39 | 57 | 36 | 1 | 49 | 100 | 17 | 73 |
| 35 | MAM_wt | tr C9L847 C9L847_BLAHA | 24.3 | 1.9 | 27.869 | 41 | 41 | 61 | 38 | 2 | 5 | 59 | 79 | 139 |
| 36 | MAM_wt | tr D4JTH7 D4JTH7_9FIRM | 23.9 | 2.3 | 27.778 | 40 | 40 | 54 | 39 | 0 | 15 | 68 | 1 | 54 |
| 37 | MAM_wt | tr D4M698 D4M698_9FIRM | 26.6 | 0.46 | 27.692 | 47 | 47 | 65 | 45 | 1 | 72 | 134 | 187 | 251 |
| 38 | MAM_wt | tr B6VXT6 B6VXT6_9BACE | 26.6 | 0.72 | 27.536 | 44 | 44 | 69 | 41 | 1 | 76 | 135 | 77 | 145 |
| 39 | MAM_wt | tr C0X1L4 C0X1L4_ENTFL | 26.6 | 0.39 | 34.211 | 27 | 27 | 38 | 24 | 1 | 14 | 50 | 150 | 187 |



Analysis



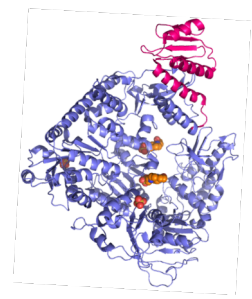
No other hit except in *Faecalibacterium Prausnitzii SL3_3*

MAM strictly restricted to a niche of *F. Prausnitzii species*

| | | | bitscore | evalue | pident | qcovs | qcovhsp | length | mismatch | gapopen | |
|----|-----------------------------|-----------------------------|----------|-----------|--------|-------|---------|--------|----------|---------|----|
| 2 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A0A0D6MVR3 A0A0D6MVR3_A | 773 | 0.0 | 39.464 | | 95 | 95 | 1120 | 642 | 15 |
| 3 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R7JPZ2 R7JPZ2_9BACT | 572 | 0.0 | 44.162 | | 78 | 57 | 668 | 361 | 4 |
| 4 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B0PG14 B0PG14_9FIRM | 660 | 0.0 | 36.837 | | 92 | 92 | 1094 | 641 | 18 |
| 5 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R5UYH1 R5UYH1_9BACE | 542 | 9.11e-175 | 41.740 | | 74 | 57 | 678 | 374 | 5 |
| 6 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B6W1Q4 B6W1Q4_9BACE | 561 | 0.0 | 40.387 | | 80 | 61 | 723 | 405 | 6 |
| 7 | 2EYQ:A PDBID CHAIN SEQUENCE | tr E5WUF0 E5WUF0_9BACE | 538 | 1.75e-173 | 41.654 | | 74 | 57 | 677 | 376 | 5 |
| 8 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R6S1S3 R6S1S3_9BACE | 544 | 6.91e-176 | 41.802 | | 74 | 57 | 677 | 375 | 5 |
| 9 | 2EYQ:A PDBID CHAIN SEQUENCE | tr E4VYD7 E4VYD7_BACFG | 544 | 1.05e-175 | 42.097 | | 73 | 57 | 677 | 373 | 5 |
| 10 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B3CGR2 B3CGR2_9BACE | 543 | 2.44e-175 | 41.740 | | 74 | 57 | 678 | 375 | 5 |
| 11 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R7B293 R7B293_9BACE | 650 | 0.0 | 37.827 | | 83 | 83 | 994 | 552 | 14 |
| 12 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C3QPJ5 C3QPJ5_9BACE | 539 | 1.63e-173 | 41.445 | | 74 | 57 | 678 | 376 | 5 |
| 13 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C6Z4G4 C6Z4G4_9BACE | 558 | 0.0 | 40.249 | | 80 | 61 | 723 | 406 | 6 |
| 14 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A0A0M1W4E3 A0A0M1W4E3_9 | 560 | 0.0 | 40.387 | | 80 | 61 | 723 | 405 | 6 |
| 15 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C3QBP8 C3QBP8_9BACE | 537 | 7.85e-173 | 41.298 | | 74 | 57 | 678 | 377 | 5 |
| 16 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A0A108TBR9 A0A108TBR9_BA | 534 | 5.30e-172 | 41.298 | | 74 | 57 | 678 | 377 | 5 |
| 17 | 2EYQ:A PDBID CHAIN SEQUENCE | tr Q8AB59 Q8AB59_BACTN | 536 | 1.41e-172 | 40.855 | | 74 | 57 | 678 | 380 | 5 |
| 18 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R7EH57 R7EH57_9BACE | 539 | 2.31e-173 | 41.003 | | 74 | 57 | 678 | 380 | 5 |
| 19 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A6L2L5 A6L2L5_BACV8 | 558 | 0.0 | 40.249 | | 80 | 61 | 723 | 406 | 6 |
| 20 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C9LCA3 C9LCA3_BLAHA | 576 | 0.0 | 44.250 | | 72 | 60 | 687 | 375 | 2 |
| 21 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R5L9D1 R5L9D1_9FIRM | 593 | 0.0 | 46.311 | | 72 | 55 | 637 | 337 | 2 |
| 22 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C0CUY2 C0CUY2_9FIRM | 601 | 0.0 | 46.330 | | 78 | 57 | 654 | 345 | 2 |
| 23 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R6P1H2 R6P1H2_9CLOT | 578 | 0.0 | 46.271 | | 80 | 57 | 657 | 339 | 5 |
| 24 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R6PNN2 R6PNN2_9CLOT | 578 | 0.0 | 45.723 | | 69 | 56 | 643 | 340 | 3 |
| 25 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B0NCF3 B0NCF3_CLOSV | 570 | 0.0 | 44.428 | | 71 | 59 | 682 | 364 | 3 |
| 26 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A7VI89 A7VI89_9CLOT | 656 | 0.0 | 35.992 | | 88 | 88 | 1053 | 619 | 11 |
| 27 | 2EYQ:A PDBID CHAIN SEQUENCE | tr D4C8Z1 D4C8Z1_9CLOT | 674 | 0.0 | 37.367 | | 87 | 87 | 1033 | 597 | 12 |
| 28 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B0NYB3 B0NYB3_9CLOT | 596 | 0.0 | 47.088 | | 73 | 52 | 601 | 314 | 1 |
| 29 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A4E9V3 A4E9V3_9ACTN | 534 | 1.92e-171 | 42.857 | | 76 | 58 | 672 | 370 | 6 |
| 30 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R6LB70 R6LB70_9FIRM | 634 | 0.0 | 37.151 | | 93 | 93 | 1074 | 612 | 16 |
| 31 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R5WLH5 R5WLH5_9FIRM | 477 | 7.11e-158 | 50.316 | | 41 | 41 | 475 | 229 | 3 |
| 32 | 2EYQ:A PDBID CHAIN SEQUENCE | tr B0G3N8 B0G3N8_9FIRM | 583 | 0.0 | 43.629 | | 82 | 63 | 722 | 387 | 6 |
| 33 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R7FNN1 R7FNN1_9FIRM | 637 | 0.0 | 37.681 | | 89 | 89 | 1035 | 568 | 14 |
| 34 | 2EYQ:A PDBID CHAIN SEQUENCE | tr C0X941 C0X941_ENTFL | 684 | 0.0 | 36.750 | | 92 | 92 | 1083 | 636 | 15 |
| 35 | 2EYQ:A PDBID CHAIN SEQUENCE | tr R6G373 R6G373_9FIRM | 654 | 0.0 | 36.794 | | 91 | 91 | 1098 | 600 | 22 |
| 36 | 2EYQ:A PDBID CHAIN SEQUENCE | tr D4JLK0 D4JLK0_9FIRM | 226 | 1.79e-63 | 34.483 | | 34 | 34 | 406 | 247 | 6 |
| 37 | 2EYQ:A PDBID CHAIN SEQUENCE | tr D4JVX9 D4JVX9_9FIRM | 562 | 0.0 | 46.026 | | 75 | 52 | 604 | 320 | 3 |
| 38 | 2EYQ:A PDBID CHAIN SEQUENCE | tr A5Z5K5 A5Z5K5_9FIRM | 672 | 0.0 | 36.886 | | 90 | 90 | 1079 | 612 | 21 |
| 39 | 2EYQ:A PDBID CHAIN SEQUENCE | tr D4KAB8 D4KAB8_9FIRM | 564 | 0.0 | 46.384 | | 71 | 55 | 636 | 333 | 4 |

Post-analysis

- ➔ Mfd ubiquitous as expected
- ➔ Modeling of Mfd complex functional cycle that associates distinct functions with conformations
- ➔ Focus on *B. cereus* strains clinical and not to identify some mark of pathogeny-NO stress



Galaxy is a scientific workflow, data integration and data analysis platform to render computational biology accessible to research scientists who do not have computer programming or system administration knowledge.

Galaxy is an open, web-based platform:

- **Accessible:** users without programming experience can easily specify parameters and run tools and workflows.
- **Reproducible:** galaxy captures information so that any user can reproduce a complete computational analysis.
- **Transparent:** users share and publish analyses *via* the web, compute interactive, web-based a complete analysis.

The screenshot displays the Galaxy web interface with several key components highlighted:

- Tools available:** A blue arrow points to the 'Tools' sidebar on the left, which lists various tools including 'Migale Tools', 'ProteoRE', and multiple instances of 'MetaFoldScan'.
- Submission form or view of input/output files:** An orange arrow points to the central configuration panel for 'MetaFoldScan 5'. This panel includes:
 - Protein(s) of interest:** A list of FASTA files: 91: MFD Chaîne A.fasta, 76: MFD.fasta, 56: Bacteroides_caccae.fasta, 55: Acetobacter_aceti_NBRC_14818.fasta, and 53: .fasta.
 - Query File (genome):** A text input field with a 'Select/Unselect all' checkbox.
 - Minimum sequence size:** A text input field containing '100'.
 - Maximum sequence size:** A text input field containing '700'.
 - Execute:** A blue button with a checkmark icon.
- History of results:** A purple arrow points to the 'History' panel on the right, which shows a list of past analyses. The most recent entry is '91: MFD Chaîne A / Acetobacter_aceti_NBR C_14818.faa', which is highlighted in green and shows '1 sequences' in 'fasta' format.

Galaxy / Dev-Migale | Analyse de données | Workflow | Données partagées | Visualisation | Aide | Utilisateur | Using 15.8 GB

Tools

- search tools
- ProteoRE
- MiRDeep2
- SNIPlay
- MetaFoldScan**
- MetaFoldScan MetaFoldScan,
- MetaFoldScan 2 MetaFoldScan,
- MetaFoldScan 3 MetaFoldScan,
- MetaFoldScan 4 MetaFoldScan,
- MetaFoldScan 4 (1 CPU) MetaFoldScan,
- MetaFoldScan 5 MetaFoldScan,
- MetaFoldScan 4 (test) MetaFoldScan,
- MetaFoldScan 6 MetaFoldScan,
- MetaFoldScan Results Filtering
- MetaFoldScan Results Filtering

Protein(s) of interest

20: ORF_VH1_dani_wt.faa
1: dorei-2prot.fasta

Reference protein

Will you select a genome from your history or use a built-in index?

Use a built-in genome index **Query genomes**

Select genome

Select/Unselect all

- Test_Genome
- Test2_Genome
- Acetobacter_aceti_NBRC_14818
- Alistipes_putredinis
- Anaerotruncus_colihominis
- Bacteroides_caccae
- Bacteroides_dorei_5_1_36_D4
- Bacteroides_doreiDSM17855
- Bacteroides_eggerthii
- Bacteroides_finegoldii
- Bacteroides_fragilis_3_1_12
- Bacteroides_intestinalis
- Bacteroides_ovatus_ATCC_8483
- Bacteroides_pectinophilus
- Bacteroides_sp._2_2_4
- Bacteroides_sp._4_3_47FAA
- Bacteroides_sp._9_1_42FAA
- Bacteroides_sp.D1
- Bacteroides_stercoris
- Bacteroides_thetaiotaomicron_VPI-5482
- Bacteroides_uniformis
- Bacteroides_vulgatus_ATCC_8482
- Bacteroides_xylanisolvans_XB1A
- Blautia_hansenii
- butyrate-producing_bacterium_SS3-4
- Butyrivibrio_crossotus
- Clostridium_asparagiforme
- Clostridium_leptum
- Clostridium_nexile
- Clostridium_scindens
- Clostridium_sp._L2-50
- Clostridium_sp._M62_1
- Clostridium_sp._SS2-1
- Collinsella_aerofaciens
- Coprococcus_comes
- Coprococcus_eutactus
- Dorea_formicigenerans_ATCC_27755
- Dorea_longicatena
- Enterococcus_faecalis_TX010
- Eubacterium_hallii
- Eubacterium_rectale_M104_1
- Eubacterium_siraeum_70_3
- Eubacterium_ventriosum
- Faecalibacterium_prausnitzii_SL3_3
- Holdemania_filiformis
- Mycobacterium_tuberculosis

History

Rechercher des données

MetaFoldScan
10 shown, 6 deleted
15.83 GB

- 30: Log
- 29: ORF_VH1_dani_wt / Bacteroides_thetaiotaomicron_VPI-5482 & butyrate-producing_bacterium_SS3-4
- 28: Log
- 27: ORF_VH1_dani_wt / Faecalibacterium_prausnitzii_SL3_3
- 22: ORF_VH1_dani_wt / Bacteroides_thetaiotaomicron_VPI-5482 & butyrate-producing_bacterium_SS3-4
- 21: ORF_VH1_dani_wt / Faecalibacterium_prausnitzii_SL3_3

MetaFoldScan 5 MetaFoldScan, (Galaxy Version 1)
Options

Protein(s) of interest

📄
📁

- 91: MFD Chaîne A.fasta
- 76: MFD.fasta
- 56: Bacteroides_caccae.fasta
- 55: Acetobacter_aceti_NBRC_14818.fasta
- 53: .fasta

Query File (genome)

Select/Unselect all

ace

Acetobacter_aceti_NBRC_14818

100

Maximum sequence size

700

✓ Execute

METAFOLDSCAN - ...
DESCRIPTION

Work in progress

Promising as a web tool to screen a couple of genomes or more

Associates accuracy and autonomy

MetaFoldScan 5 MetaFoldScan, (Galaxy Version 1)
Options

Protein(s) of interest

📄
📁

- 91: MFD Chaîne A.fasta
- 76: MFD.fasta
- 56: Bacteroides_caccae.fasta
- 55: Acetobacter_aceti_NBRC_14818.fasta
- 53: .fasta

Query File (genome)

Select/Unselect all

Acetobacter_aceti_NBRC_14818

Maximum sequence size

✓ Execute

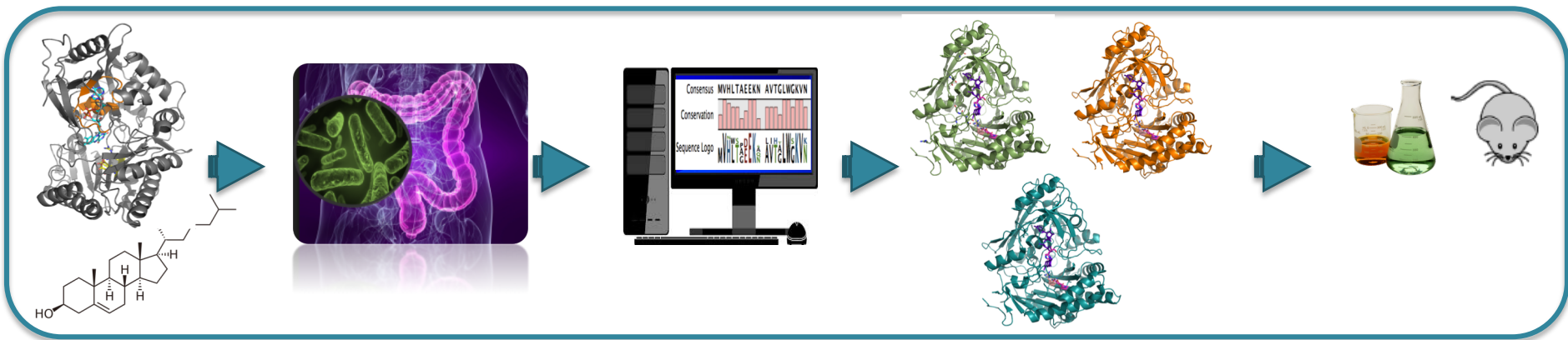
METAFOLDSCAN - ... *Work in progress*

DESCRIPTION

Main hurdle : scaling up from core genome to 10 millions of genes

→ **Filtering**: size (number of residues), signal peptide, secretome, etc.

→ **Data storage**, including the temporary data generated crash after 22 genomes and 2 months of calculations



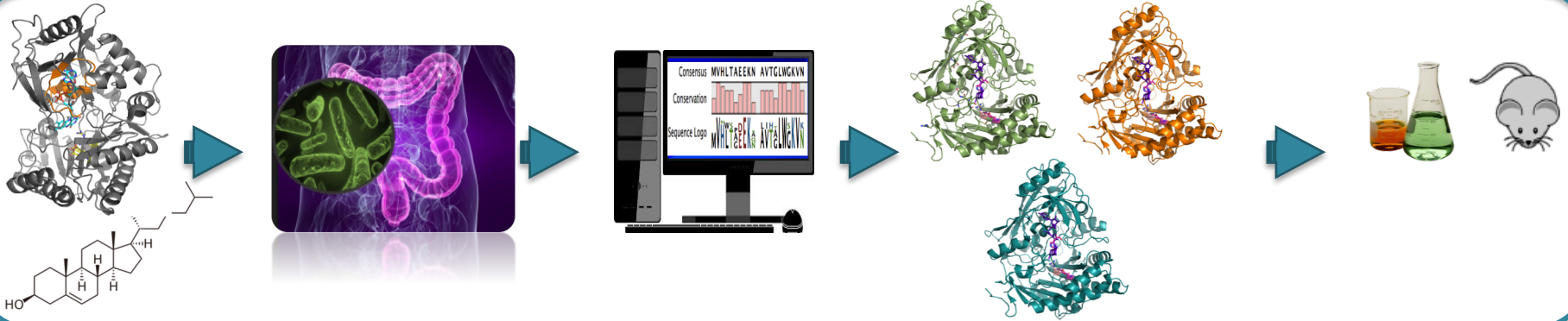
- Collaboration with B. Offmann –University of Nantes, France, for FORSA
- Structural annotation of protein sequences: from Hidden Markov Model profiling to alphabet protein
- FORSA scores the compatibility of any amino acid sequence with any fold represented in the form of protein blocks sequences (1D representation of 3D information).
- FORSA is highly scalable to large datasets
- FORSA uses both local and global threading through parallelising large amount of analyses.

➡ **One genome one protein one hour on a laptop**



Deployable, scalable to 1,000 genomes and more, early 2019

Towards the Structural Screening of Microbial Ecosystems



Outline

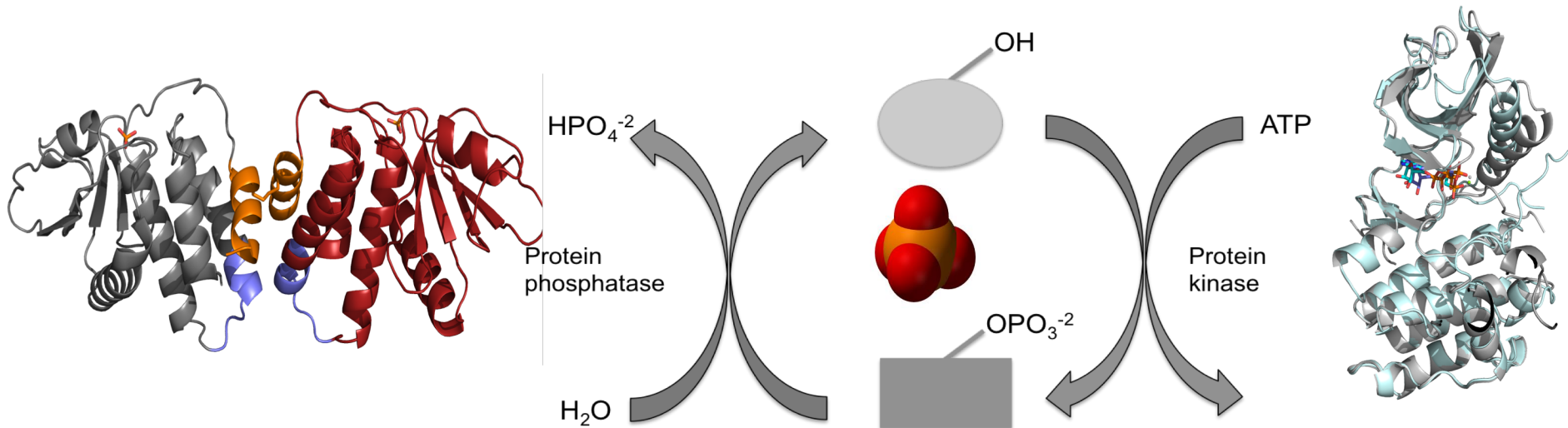
- Cholesterol conversion in the gut microbiota: the mystery enzyme(s) ?
- MetaFoldScan project : 3D screening of the gut microbiota
- **Conclusions and perspectives**

Collaborative projects: actors of the dialog gut /host

Cell signaling proteins



Responsible for the dialog
bacteria/bacteria et host/bacteria

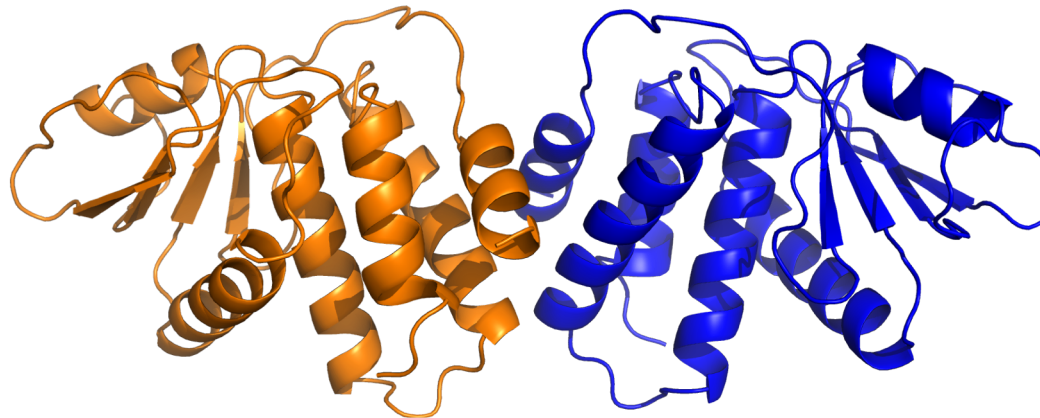


Dialog between in and outside: Tyr-phosphatase & kinases amplify a signal from the membrane

Collaborative project 1: identify and characterize Tyr-phosphatases

Dr. A. Villarino & Dr. M. Berois, Universidad de Montevideo, Uruguay

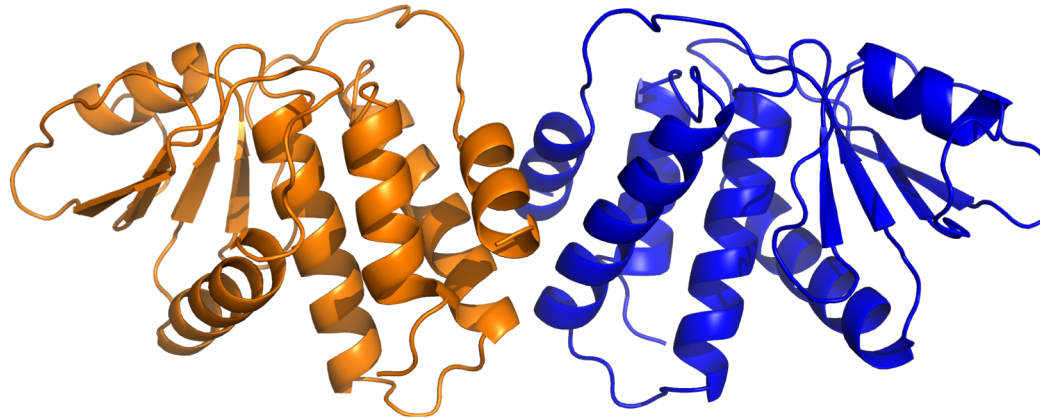
Tyr-Phosphatase






Collaborative project 1: identify and characterize Tyr-phosphatases



Dr. A. Villarino & Dr. M. Berois, Universidad de Montevideo, Uruguay



Tyr-Phosphatase



Intracellulaire pathogen ORF

-  Actif as a dimer
-  Hijacks the immune system
-  Dephosphorylates an essential cell mediator cellulaire essentiel

 2 CR1, 1AI, 2 Master
 MaIAGE, Migale

 2 PR, 2 Master
 Facultad de Ciencias, Montevideo, Uruguay

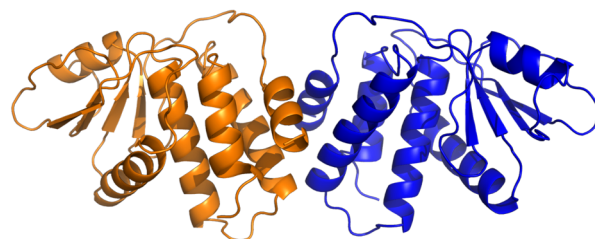
 Ecos-Sud Uruguay
 grants

Screen the gut microbiota to target Tyr-phosphatases

Dr. A. Villarino & Dr. M. Berois, Universidad de Montevideo, Uruguay

Antigène in the new Crohn detection kit: Tyr-phosphatase PtpA

Mycobacterium avium ssp paratuberculosis



viral Tyr-phosphatase

Bacteria



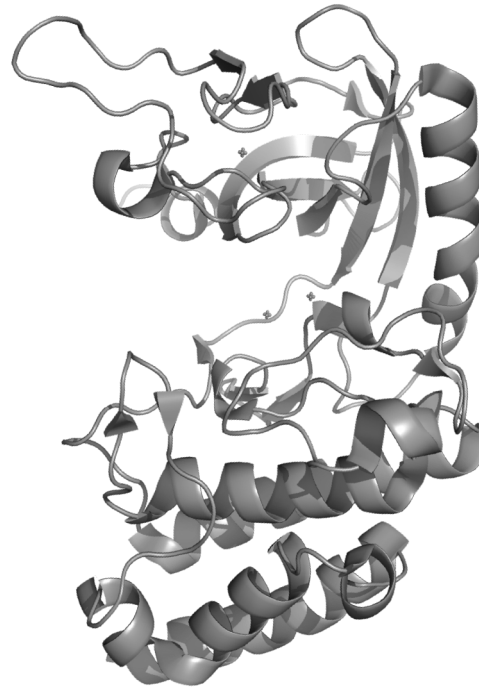
- Conservation
- Distribution
- Structure/fonction

Structural homologs of Tyr-phosphatases




Collaborative project 2: identify a pathway involved in metabolism



Dr. M.N. Lisa, IBR-Conicet, Rosario Argentina

Ser/Thr Protein Kinase



Mycobacterium tuberculosis

-  Central metabolism
-  Operon of 7 proteins
-  Detects nutrients availability

 1DR2,1 CR1, 1AI
 MalAGe, Migale

 1PI
 IBR, Rosario, Argentina

 **MEM**
CONICET


Grants

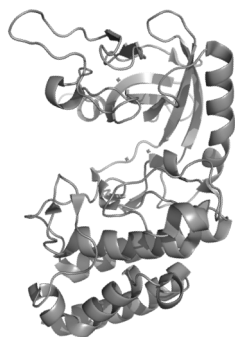
Deposited

Screen and target kinase PknG and its upstream and downstream partners

Dr. M.N. Lisa, IBR-Conicet, Rosario Argentina

Antigène in the new Crohn detection kit: Ser/Thr Protein Kinase PknG

Mycobacterium avium ssp paratuberculosis



Mycobacterium tuberculosis

Actinobactéries



- Conservation
- Distribution
- Structure/fonction

Structural homologs of Ser/Thr kinases and the 6 partners of the signaling pathway

Actinobacteria: 250 genomes, 3 months computation time, heat-map with R analysis

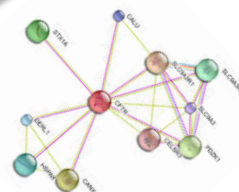
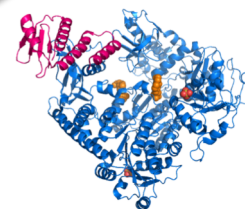
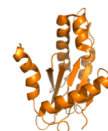
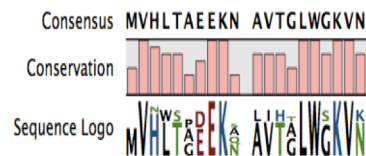
❖ **Strong positioning** at the interface structural bioinformatics & Biology

❖ **Innovative webserver** to achieve gut ecosystem exploration.

Needs scaling up & tuning (filters)

Experimental validation of the structural hits

❖ **High valorisation potential**



HTS of structures
 Structure –Function –Annotation
 Functional clusters and Pathways

Originality /prevalence of folds
 Disease/health -associated
 patterns of diversity



... Towards new targets with Health impact

... Towards other microbiotas

Acknowledgments



Véronique Martin

Jean-François Gibrat

Thomas Lacroix



Plateforme de BioInformatique - INRA Jouy en Josas

Sandra Derozier

Valentin Loux



Fabienne Béguet-Crespel

Jean-Marc Chatel

Philippe Gérard

Catherine Juste

Nalini Rama Rao



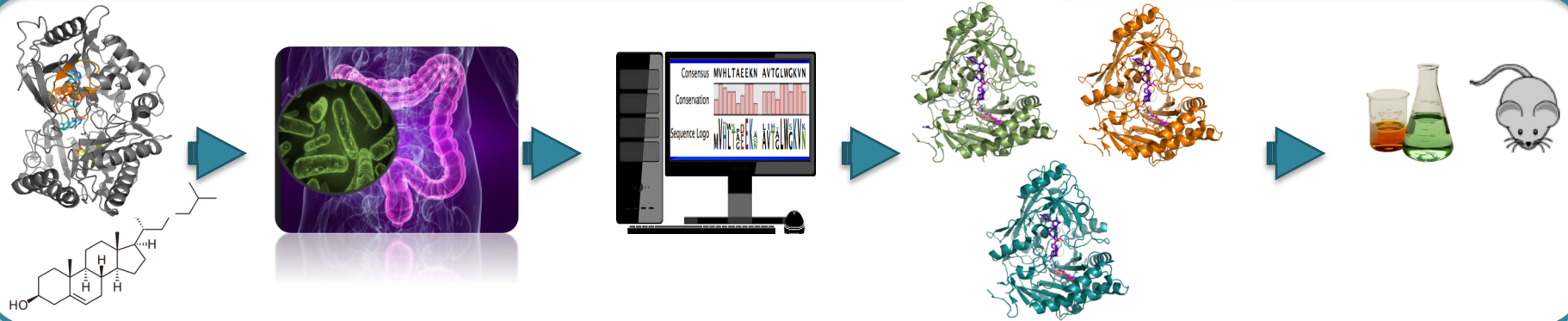
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Danilo Segovia

Dario Porley



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Bernard Offmann

Surbhi Dingra

MEM Metaprogram



Institut Pasteur

Mariano Martinez

Pedro Alzari

CONICET



María Natalia Lisa

Acknowledgments



Véronique Martin

Jean-François Gibrat

Thomas Lacroix



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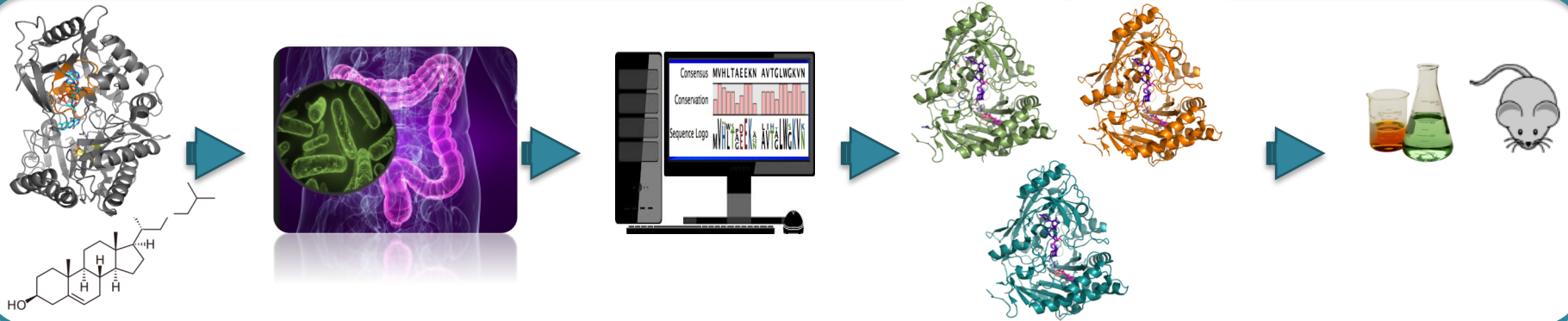
Andrea Villarino

Mabel Berois

Danilo Segovia

Dario Porley

And you for your kind attention ...



UNIVERSITÉ DE NANTES

Bernard Offmann

Surbhi Dingra

MEM Metaprogram



Institut Pasteur

Mariano Martinez

Pedro Alzari

CONICET



María Natalia Lisa

WP1

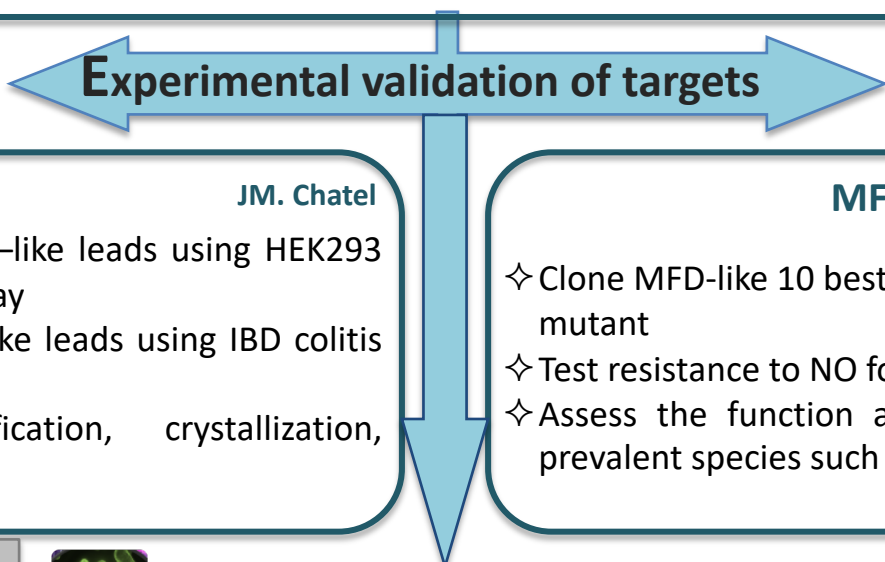


Bioinformatics development

Integration of MetaHit datasets, filtering, scanning and 3D fold detection

G. André-Leroux

- ✧ Set up of Metahit* browsing. Splitting of genomes. Filtering. Connection to pdb
- ✧ HMM profiling of MAM and MFD targets 3D candidates: from hits to leads
- ✧ Analysis of specificity, promiscuity, identification of associated pathways



WP2



MAM hits

JM. Chatel

- ✧ *In vitro* validation of MAM –like leads using HEK293 NFκB luciferase reporter assay
- ✧ *In vivo* validation of MAM-like leads using IBD colitis in mouse model
- ✧ MAM production, purification, crystallization, structural characterization

WP3

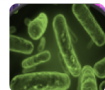


MFD hits

N. Rama Rao

- ✧ Clone MFD-like 10 best hits in *E.coli* deficient MFD mutant
- ✧ Test resistance to NO for “Complemented *E.coli* strains”
- ✧ Assess the function and specificity of MFD leads in prevalent species such as *Bacteroidetes* & *Firmicutes*

WP4



Applications - Impacts

Biologist friendly web-tool interface

V. Loux

- ✧ Production of command line pipeline distribution.
- ✧ Integration into Galaxy.
- ✧ Integration into training sessions within Migale Bioinformatics cycle
- ✧ Extension to other microbiotas

★ <http://genome.jouy.inra.fr/Insyght>

➡ Powerful tool for genes comparison and sytheny inferences

Sythenies : co-localization of homologous loci accross species

Comparison 2 vs 2 with a reference genome.

BDBH method Bi-Directional Best Hit i.e best reciprocal match between two proteomes.

Gene comparison. Sythenies visible

The screenshot shows the Insyght web interface for gene comparison. It displays a list of genes (dnaA, dnaN, recF, gyrB, etc.) and their sythenies across different species. The interface includes a 'Contextual menu' and 'Comparison windows'.

Info and options

Contextual menu: browse multiple homologies per gene