



## Genetic mapping of resistance factors to Melampsora sp in Populus sp: a review

Véronique Jorge, Alois Bresson, Arnaud Dowkiw, Patricia Faivre-Rampant,  
Brigitte Viguier, Marc Villar, Vedrana Vlainic, Elise Albert, Christopher  
Mangel, Redouane El Malki, et al.

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# BioForA



# Genetic mapping of resistance factors to *Melampsora* sp in *Populus* sp

A review



Véronique Jorge

## CONTRIBUTORS to the present work :

V. Jorge, A. Bresson, A. Dowkiw, P. Faivre-Rampant, B. Viguier, M. Villar, V. Vlainic, E. Albert, C. Mangel, R. El-Malki, M. Pegard, V. Segura, V. Guérin, C. Ridel, C. Bastien

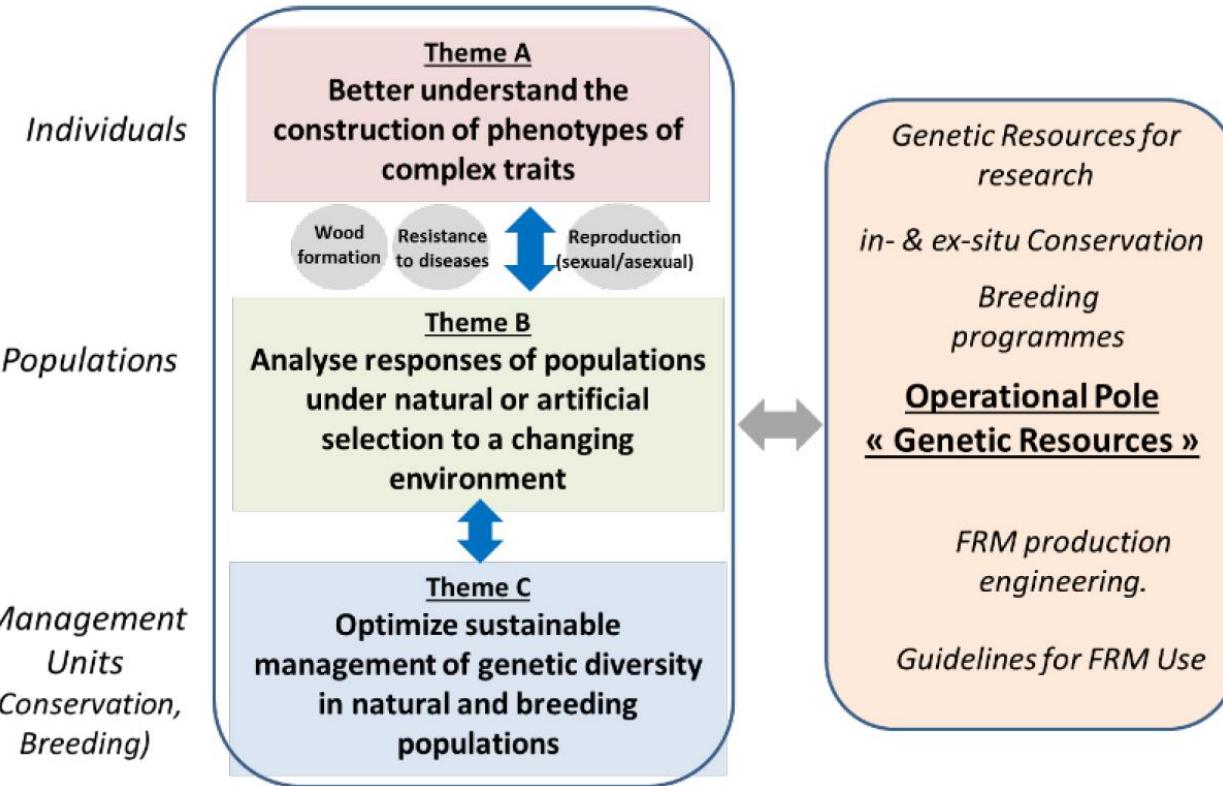
INRA Experimental unit GBFOR



# Unit BioForA

## Our research objective

Management of forest genetic resources for sustainable timber production in a changing climate context



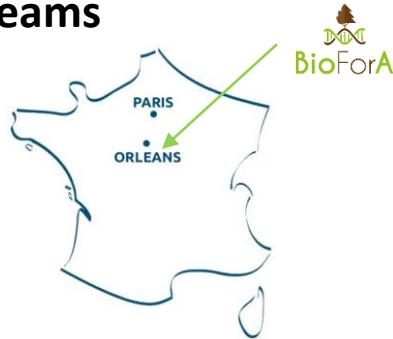
**Species studied :** Poplars, Douglas Fir, Scot Pine, wild cherry, larches, ash



- **Joint unit between INRA and ONF** (French National Forest Service)



- ~50 persons (40 perm.)
- 4 teams





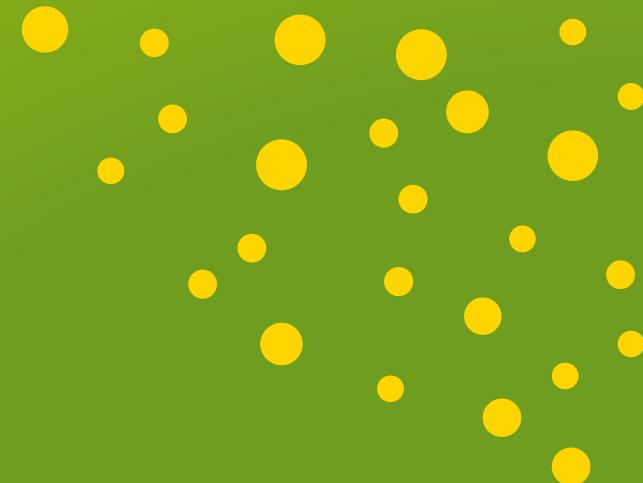
# SUMMARY



1. Poplar rust pathosystems
2. Questions and strategies to identify R factors
3. How to measure resistance components ?
4. Variability for resistance components
5. Genetic mapping: from QTL to QTN
6. Comparative genomics
7. Conclusion: potential uses for breeding

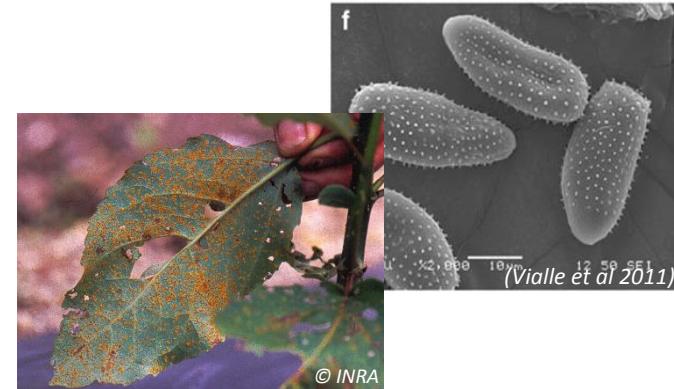
1

## Poplars / rusts pathosystems

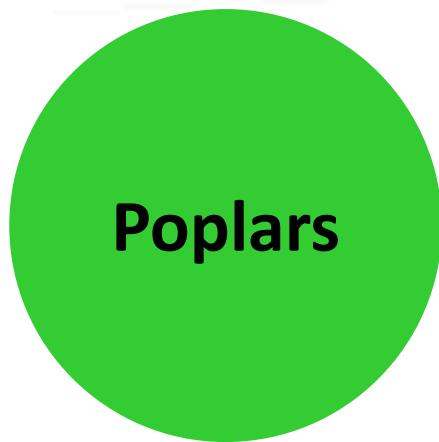


# Poplar rusts

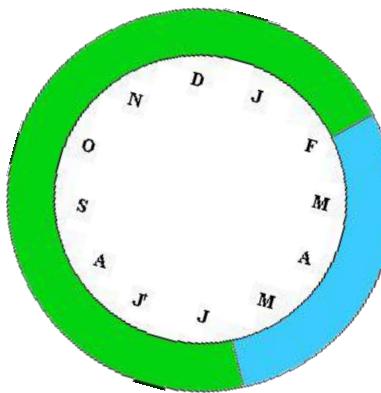
- Causes by *Melampsora spp*
- Biotrophic, heterotic cycle.
- Losses in biomass :
  - 32% per year (Dowkiw, 2003)
  - 57% (Steenackers et al. 1996)
- Resistance/tolerance is as major breeding issue
- **Qualitative resistances** deployed in hybrid cultivars overcame



# A complex biological cycle ...



Asexual reproduction  
Epidemics

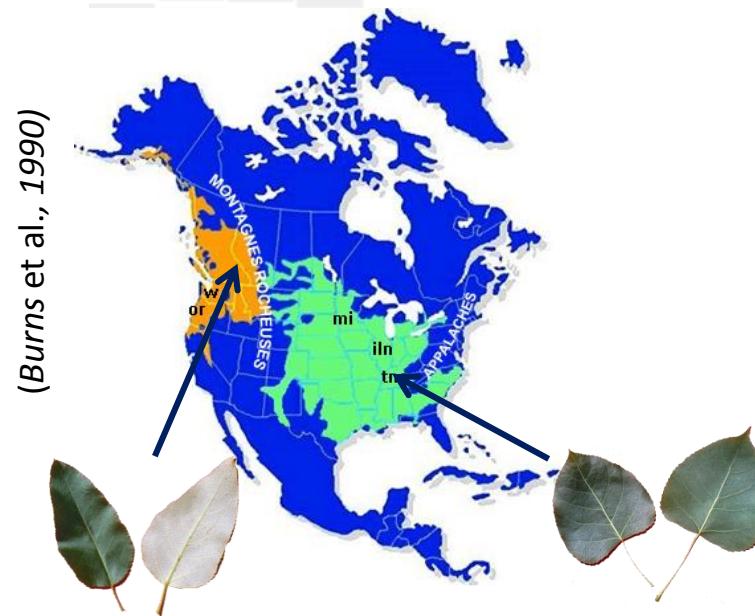


... favouring rapid pathogen evolution

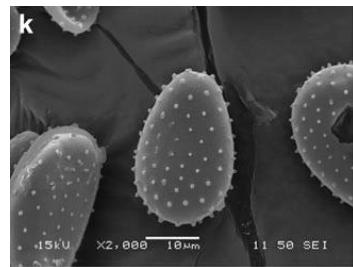
Sexual reproduction



## Natural poplars-foliar rusts pathosystems

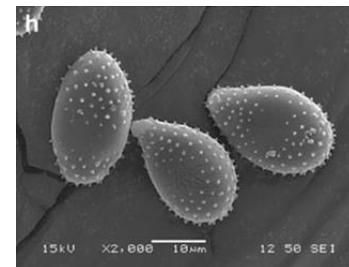


*Populus trichocarpa*  
*Melampsora occidentalis*

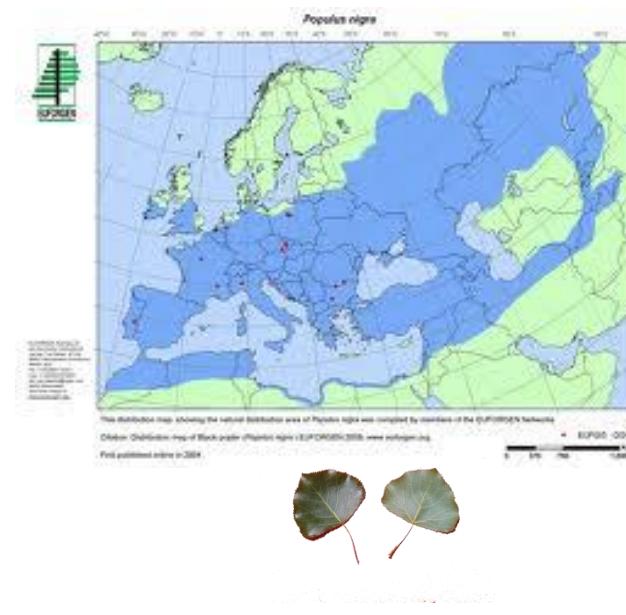


(Vialle et al 2011)

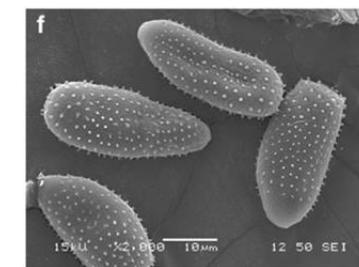
*Populus deltoides*  
*Melampsora medusae*  
*f.sp. deltoideae*



(Vialle et al 2011)



*Populus nigra*  
*Melampsora larici-populina*



(Vialle et al 2011)

# Poplar/rust « exapted » pathosystems



- Use of *Populus* exotic species => showdown of host and pathogen that did not **co-evolved**.
- The interaction is **compatible**



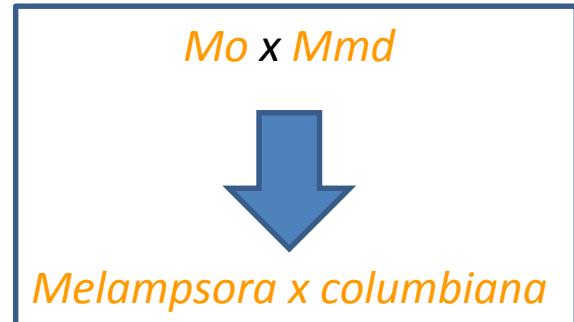
## Exapted pathosystem

(Gould et Vrba, 1982; Newcombe 1998)

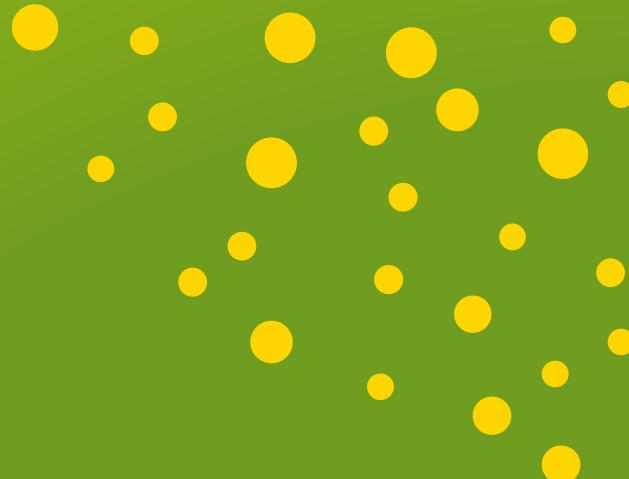
	<i>Melampsora occidentalis</i>	<i>Melampsora medusae f.sp. deltoideae</i>	<i>Melampsora larici-populina</i>
<i>Populus trichocarpa</i>	natural	exapted	exapted
<i>Populus deltoides</i>	?	natural	exapted
<i>Populus nigra</i>	?	exapted <sup>a</sup>	natural

	<i>Pt</i>	<i>Pd</i>	<i>Pn</i>
<i>Pt</i>	○	○	○
<i>Pd</i>	○	○	○
<i>Pn</i>	○	✗	○

Possible interspecific hybridation both in host and pathogen



<sup>a</sup> Pinon et Frey, 2005, ? : no published data



2

## Questions and strategies to identify R factors



## ➤ Questions

- What is the variability for resistance in the different type of pathosystems (natural/exapted)?
- What is the genetic determinism (loci and genes)?

## ➤ Strategies/approaches

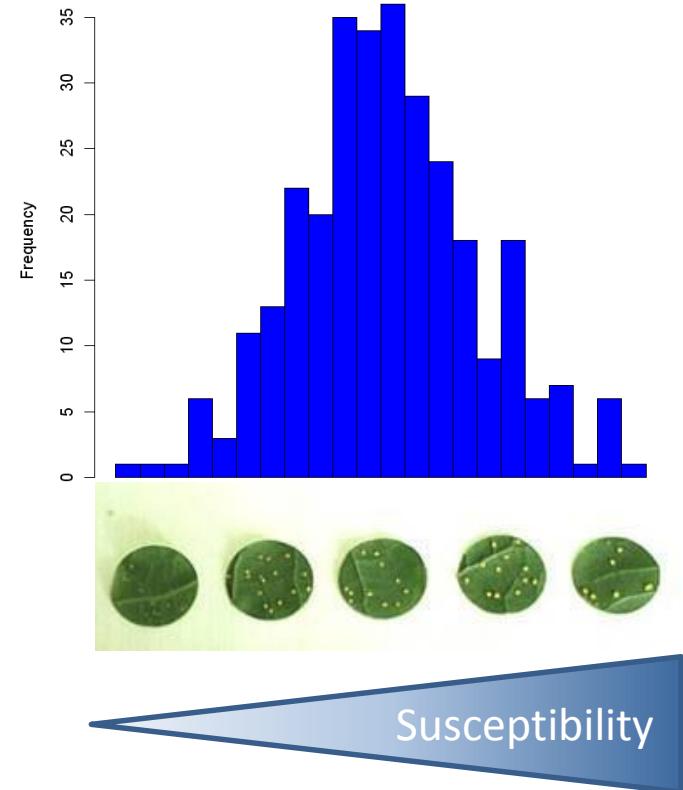
- Identify resistance components
- Evaluate variability for these components
- Map genetic factors : from QTLs to QTNs.
- Comparative genomics

# A simple classification of resistance types

**Qualitative resistance**  
**Gene for gene hypothesis**  
 (Flor, 1942)

		Pathogen
Plant	Avr/Avr Avr/vir	Vir/vir
R/R or R/r	+HR	
r/r		

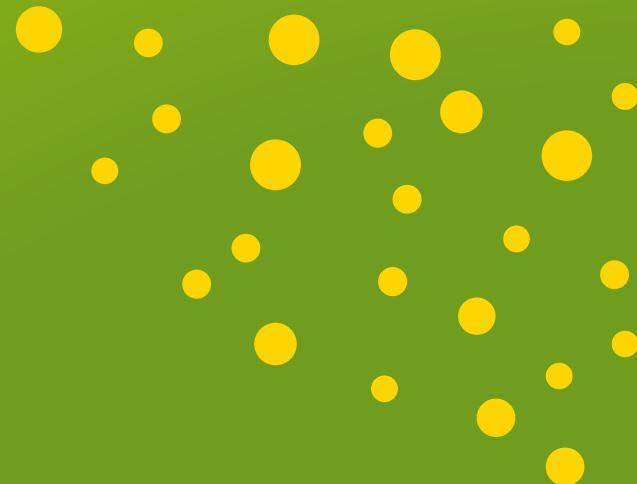
**Quantitative resistance**



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# 3

## **How to measure resistance components ?**



Plant	Pathogen	
	Avr/avr Avr/vir	Vir/vir
R/R or R/r		
r/r		

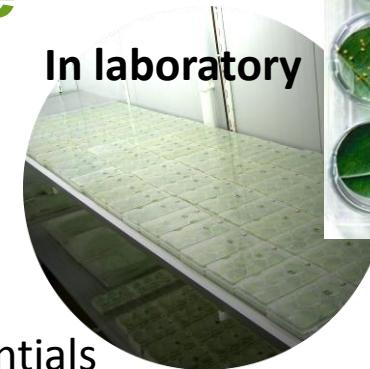
# Qualitative resistance

Specific



Control and characterise  
Pathogen variability

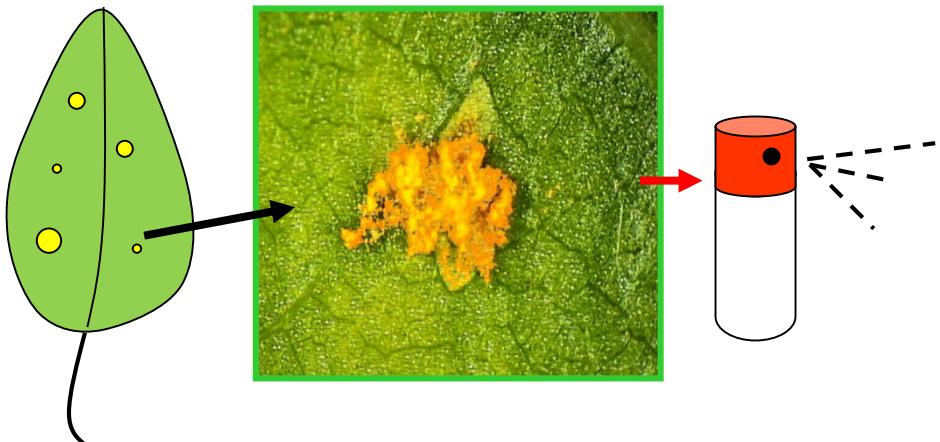
In laboratory



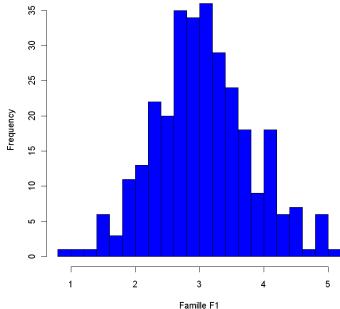
Host differentials

cv. 'Ogy'	R1
cv. 'Candidans'	R2
cv. 'Brabantica'	R3
cv. 'Unal'	R4
cv. 'Rap'	R5
87B12	R6
cv. 'Beaupré'	R7
cv. 'Hoogvorst'	R8

1 uredia = 1 clone of uredospores



Qualitative resistances / virulences well documented in exapted pathosystem :  
Also for TxD hybrids/Mxc (Newcombe et al. 2001) *P. deltoides/Mlp*



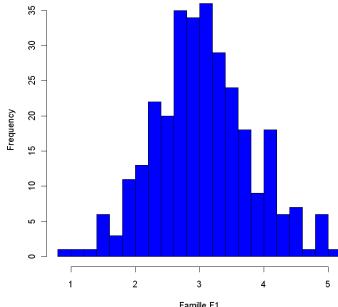
## Epidemiological components of quantitative resistance

In laboratory



Field



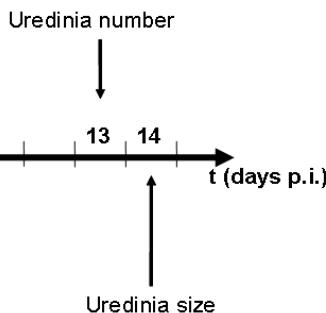


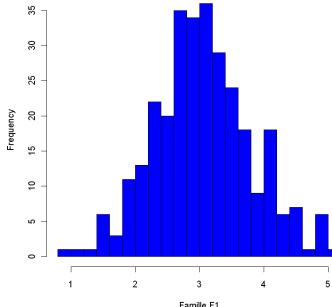
## Epidemiological components of quantitative resistance

In laboratory



Field





## Epidemiological components of quantitative resistance

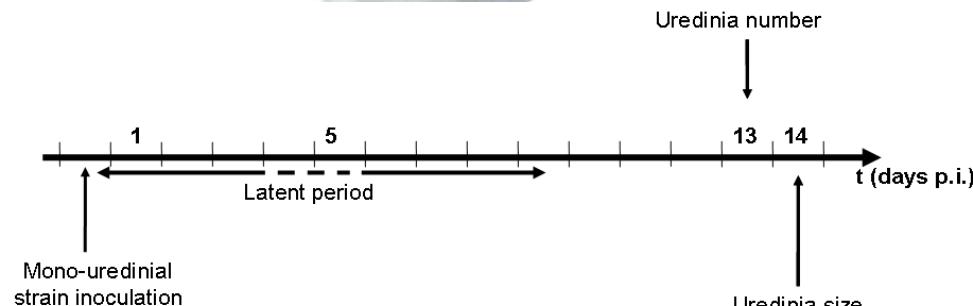
In laboratory



Field



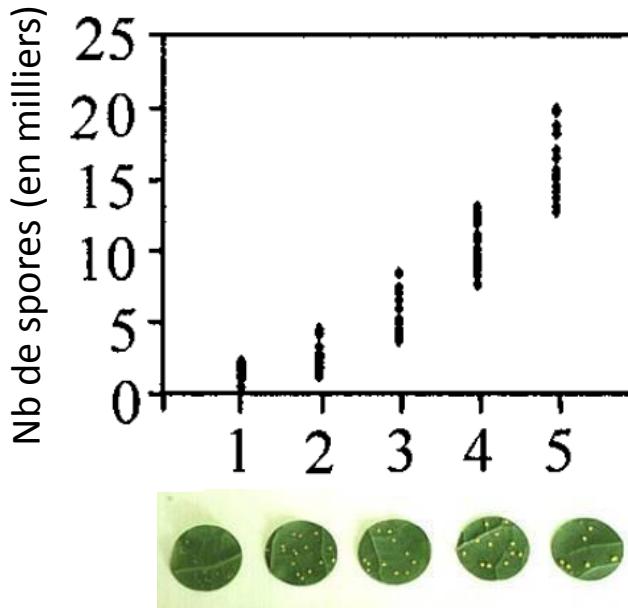
Rust score on the most infected leaf



## Epidemiological components of quantitative resistance

In laboratory

1 infection cycle

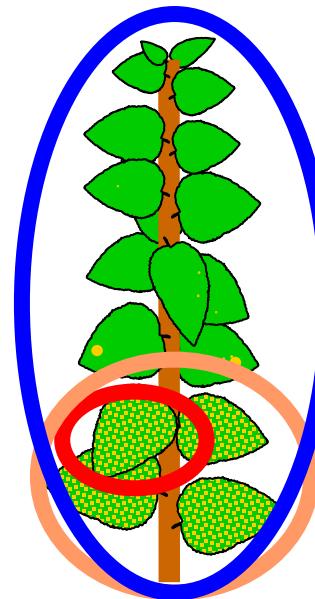


(Dowkiw et al 2003)

Measure of interaction on  
the pathogen

Field

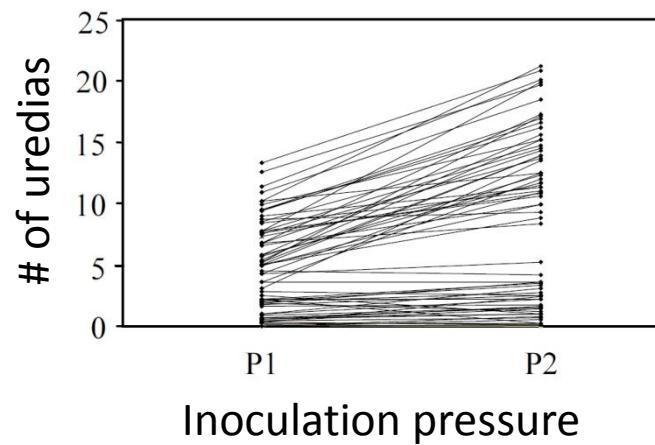
several infection cycles



Measure of interaction on  
the host plant  
(symptoms, leaf losses)

## Epidemiological components of quantitative resistance

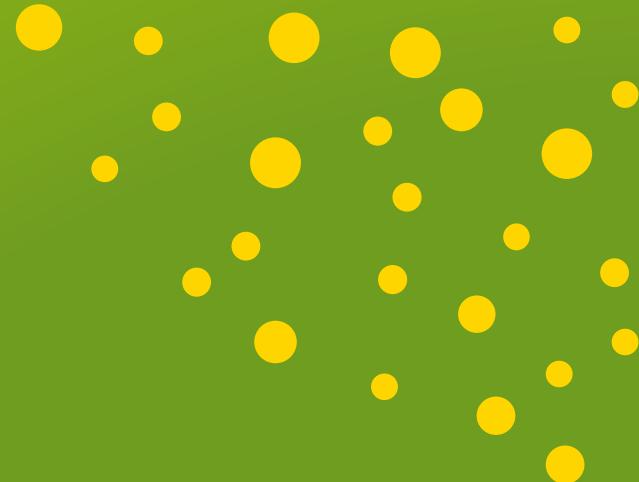
- Conditions of inoculation, scales to be tuned depending on the host and on level of infection:
  - hybrids/pure species
  - Qualitative R / quantitative R
  - period of evaluation (field)
  - age of plants
- We are sure about susceptibility not resistance



(Dowkiw et al 2003)

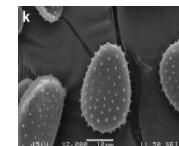
— 4

## Variability for resistance components



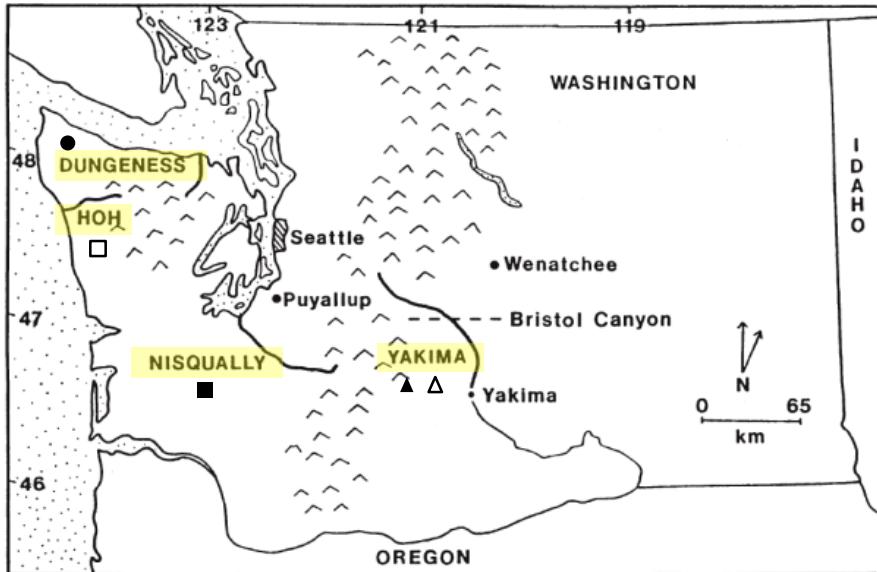
## Natural pathosystems

*Populus trichocarpa*



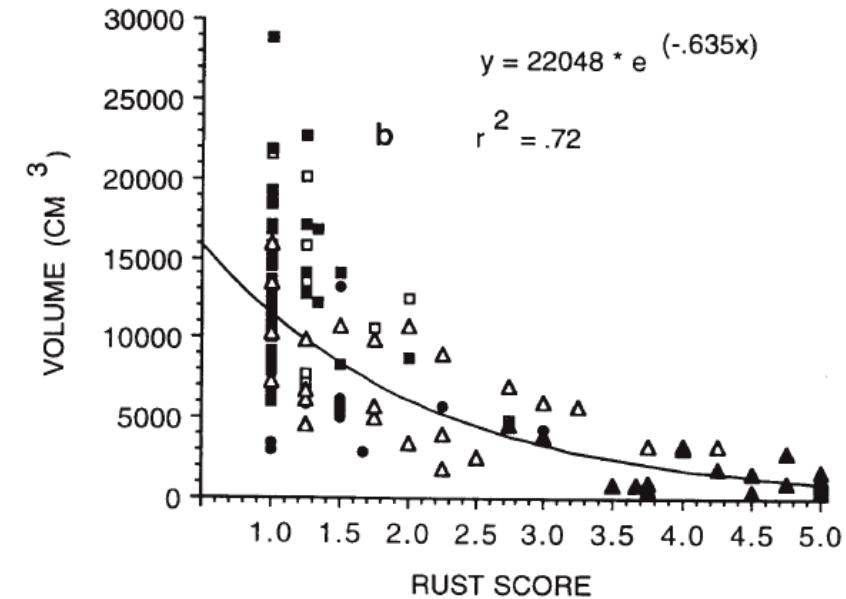
*Melampsora occidentalis*

(Vialle et al 2011)



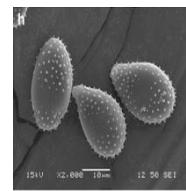
5 provenances, 126 clones  
Washington state

- Field evaluation
- Highly resistant genotypes
- Variability geographically structured

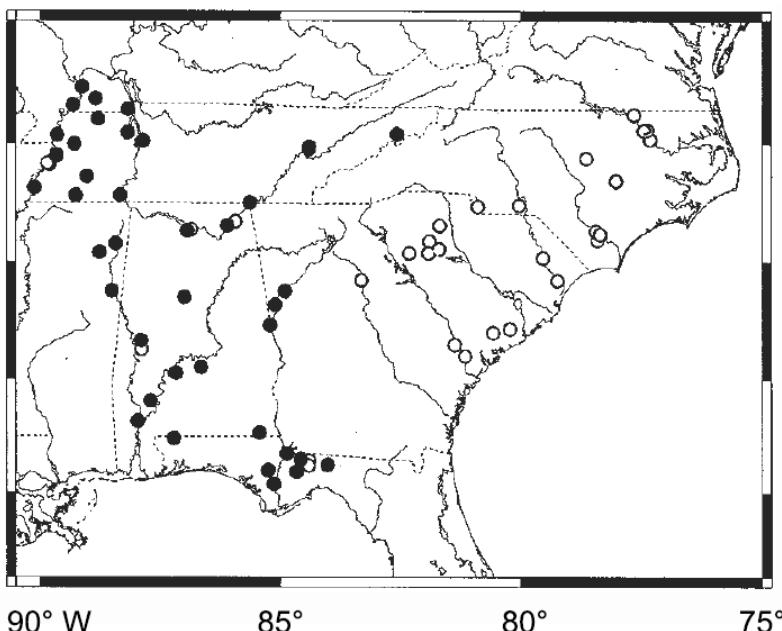


## Natural pathosystems

*Populus deltoides*

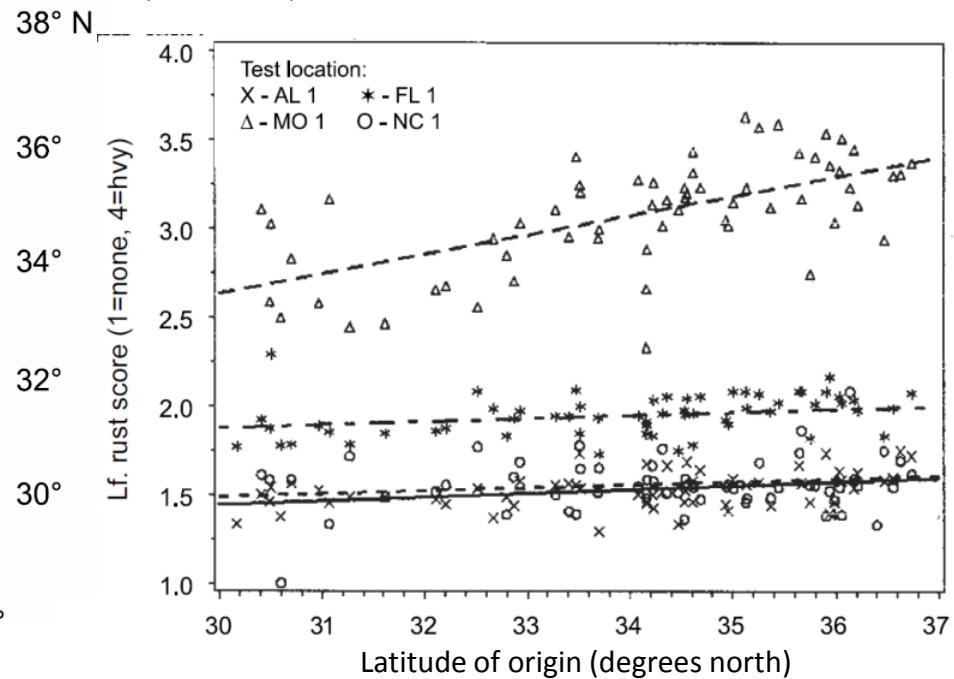


*Melampsora medusae*  
f.sp. *deltoideae*



62 provenances

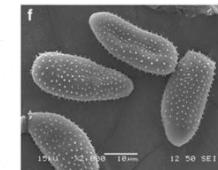
Eastern USA  
4 clonal trials



➤ Variability geographically structured

## Natural pathosystems

*Populus nigra*



*Melampsora larici-populina*

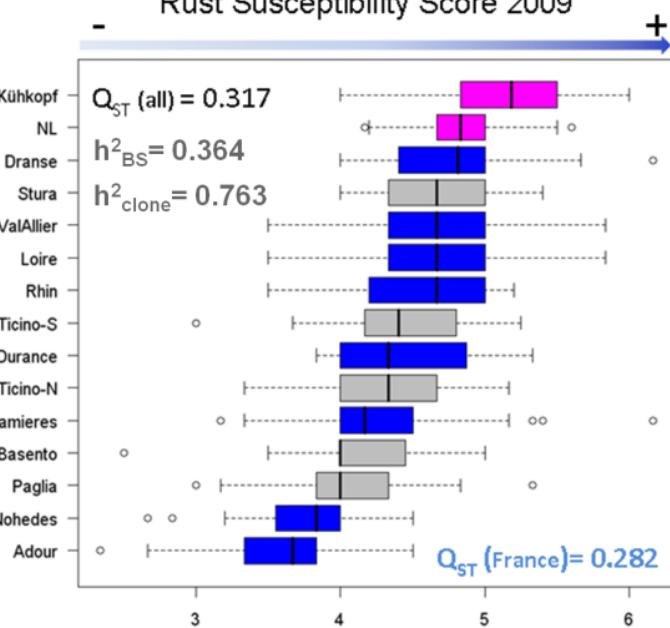


14 metapopulations, 1141 clones

Western Europe

(Vialle et al 2011)

Rust Susceptibility Score 2009





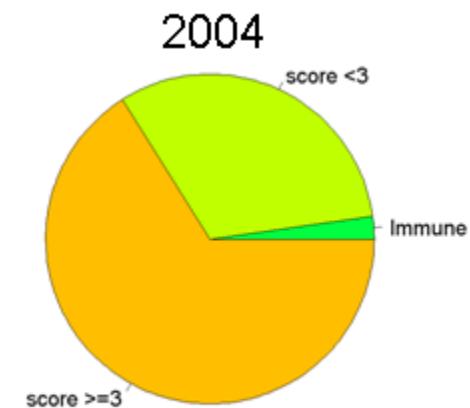
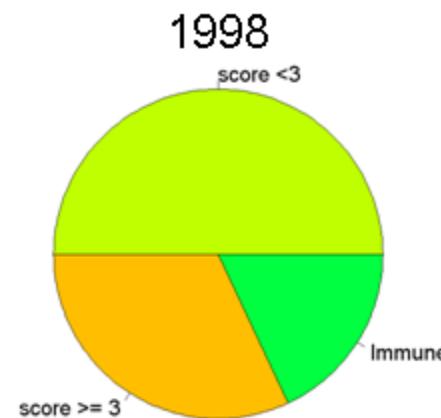
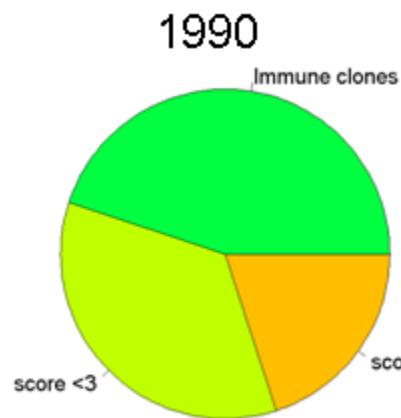
## Exapted pathosystems

*Populus deltoides*



*Melampsora larici-populina*

(Vialle et al 2011)

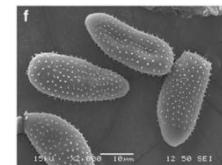


545 *P.deltoides* genotypes – Field rust susceptibility (Orléans)

- Qualitative resistances
- Erosion of resistance over time

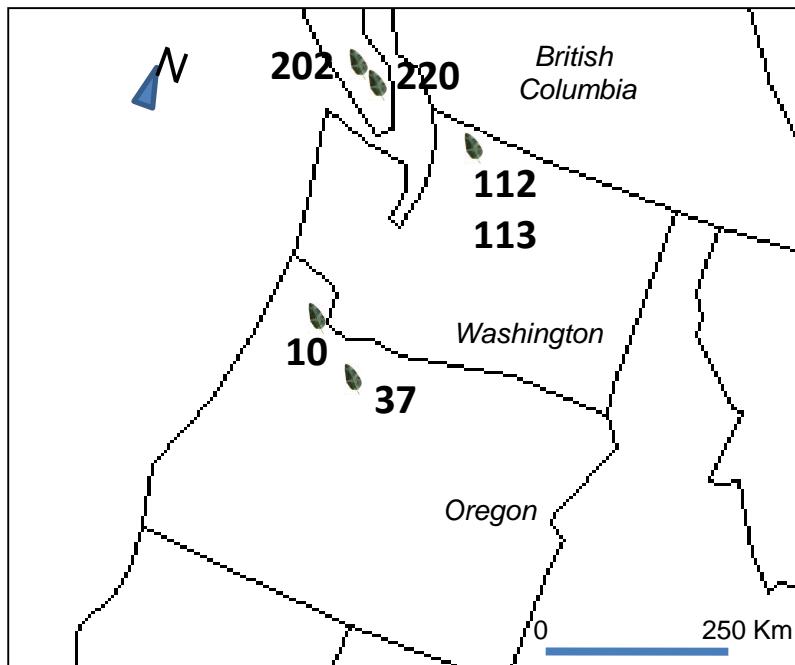
## Exapted pathosystems

*Populus trichocarpa*



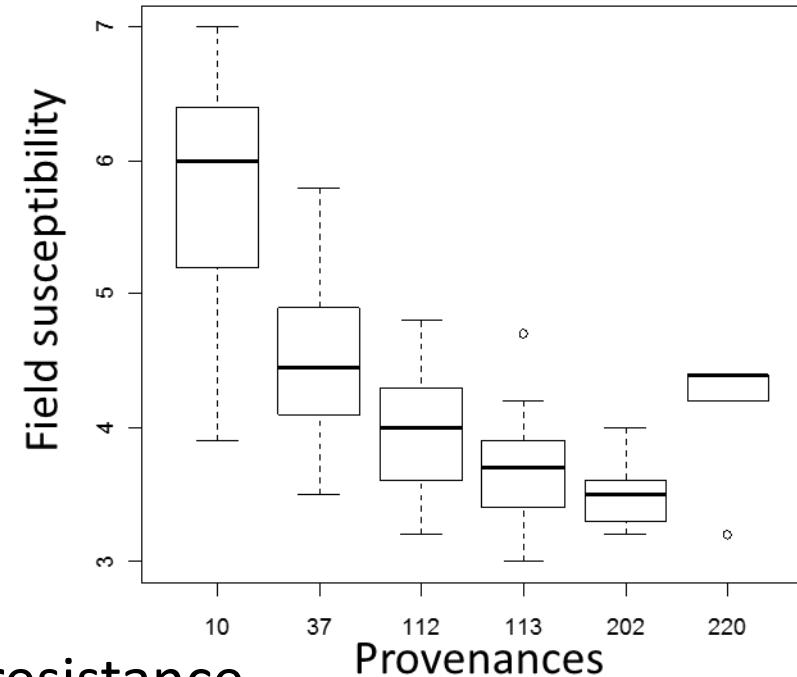
*Melampsora larici-populina*

(Vialle et al 2011)



6 provenances, 102 clones  
Clonal trial, Orléans (F)

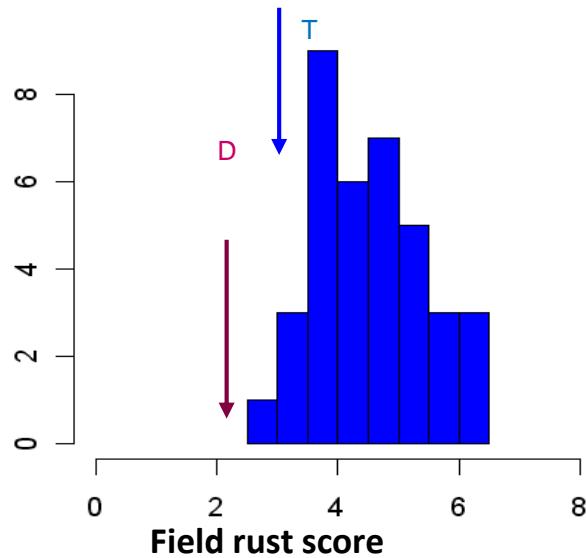
- Quantitative resistance
- Strong geographic structuration
- Idem *P. trichocarpa* / Mxc (La Mantia et al 2013)



## Exapted pathosystems

### Case of interspecific hybridization

Hybrides **DxT / DxN** vs **Mlp**

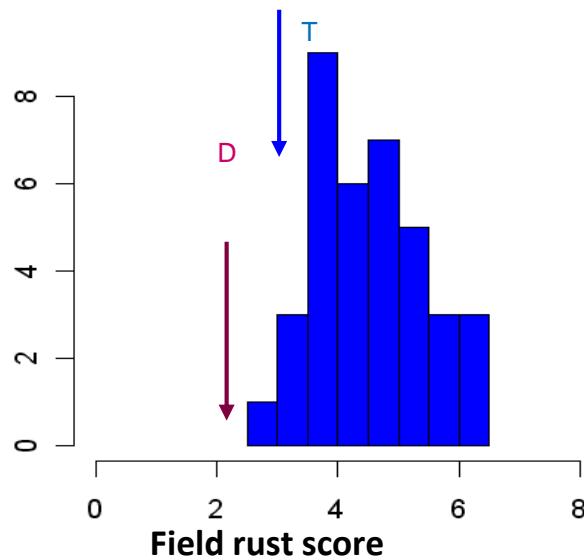


Negative heterosis .....

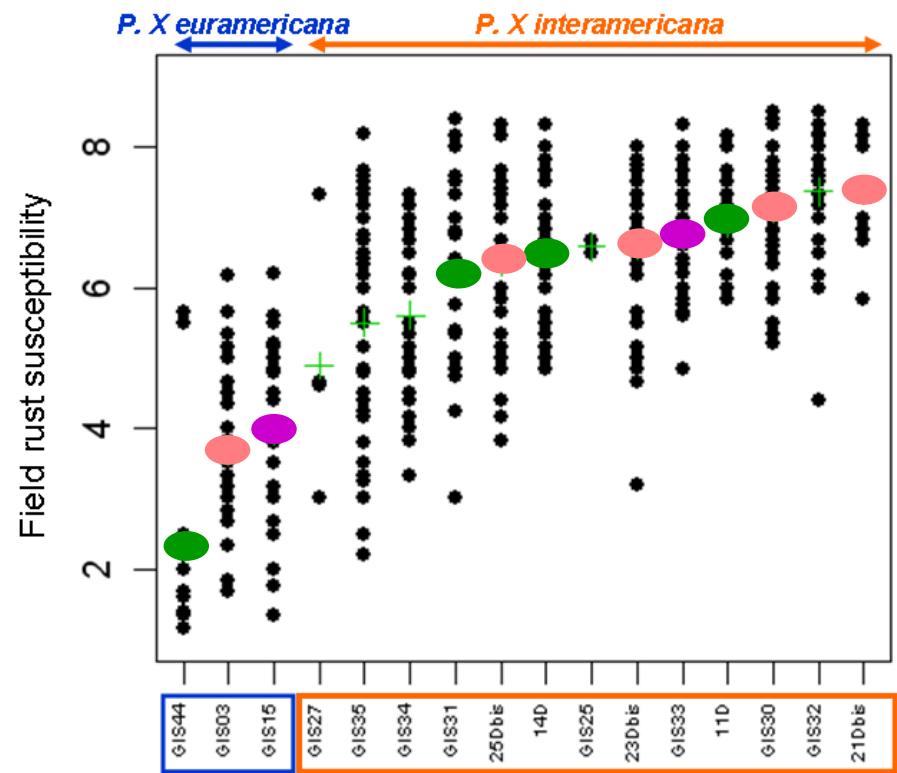
# Exapted pathosystems

## Case of interspecific hybridization

Hybrides **DxT / DxN** vs **Mlp**



Negative heterosis ....



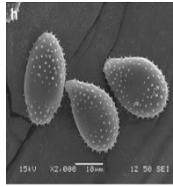
16  $F_1$  families involving common *P. deltoides* female parents

# To sum up about variability !

## Natural pathosystems



*P. trichocarpa* (Dunlap and Stettler, 1998)  
*Mo*



*P. deltoides*  
*Mmd*

(Land and Jeffreys, 2006)  
 (Nelson and Tauer, 1986)  
 (Hamelin et al 1993, 1992)



*P. nigra*  
*Mlp*

(Bastien et al, in prep.)  
 (Légonnet et al, 1999)

(Vialle et al 2011)



*P. deltoides*  
*P. trichocarpa*  
*Mlp, Mxc*

- Some genotypes strongly resistant, no qualitative resistance
- Quantitative resistance strongly geographically structured

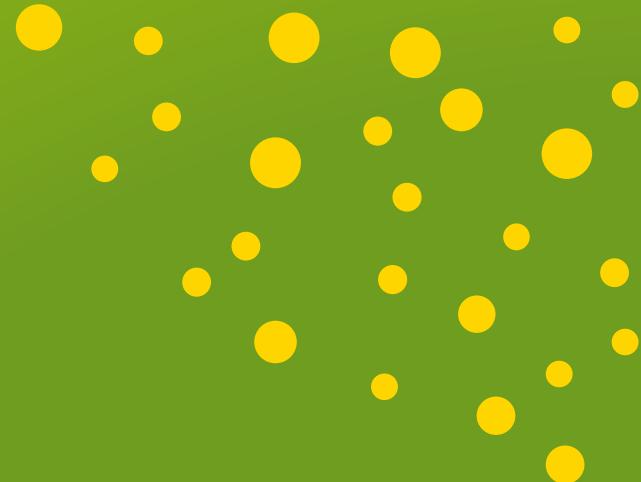
## Exapted pathosystems

- Qualitative resistances in *P. deltoides* almost all overcomed after 15 years
- Quantitative resistance in both sp., structured in *P. trichocarpa*

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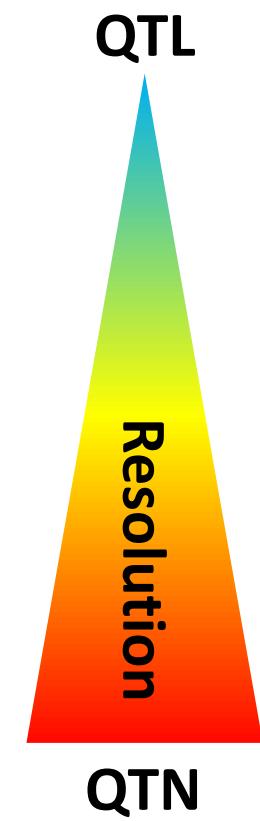
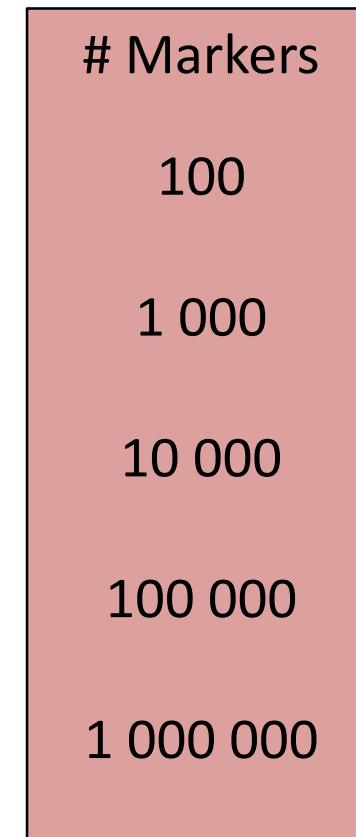
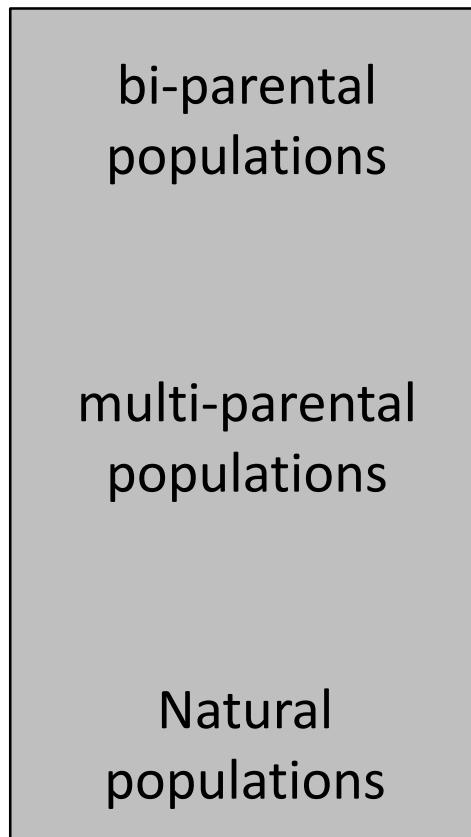
# 5

## Genetic mapping: from QTL to QTN



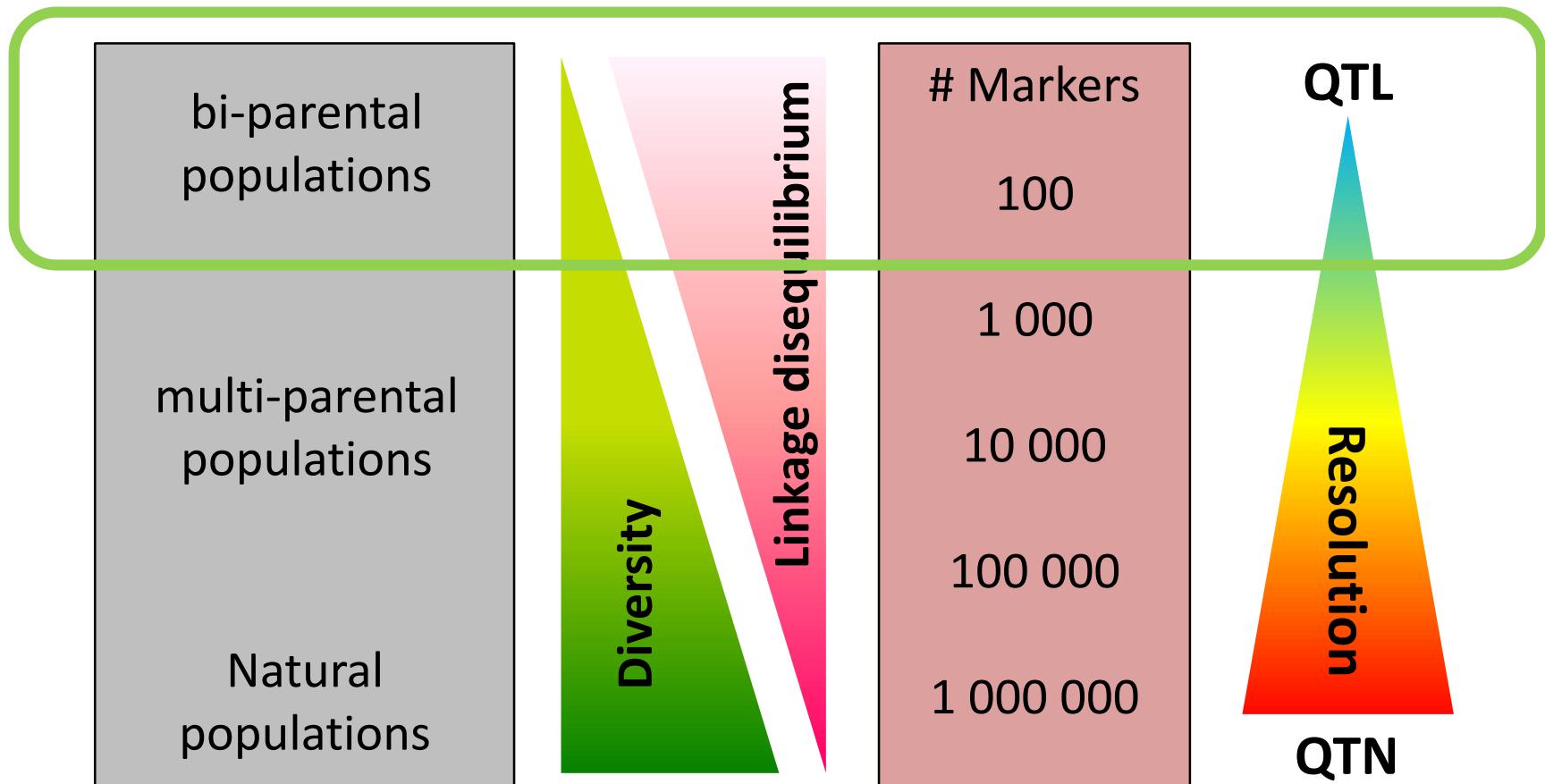
## Strategies to study genetic architecture of resistance

### From QTLs to QTNs



## Strategies to study genetic architecture of resistance

### From QTLs to QTNs

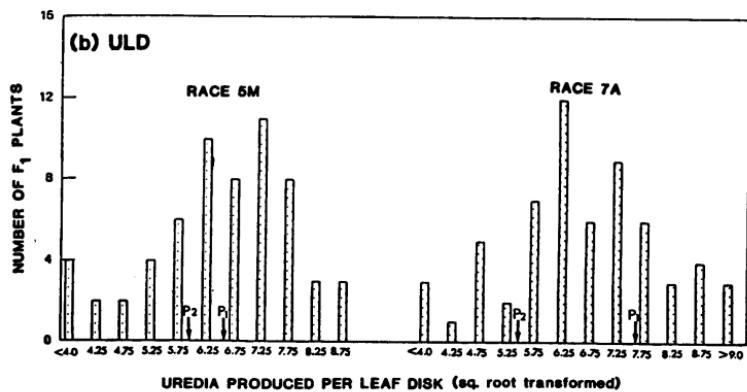


# Natural pathosystem

bi-parental populations

## *P. deltoides* x *P. deltoides* F1

2 strains **Mmd**



(Prakash & Heather, 1989)

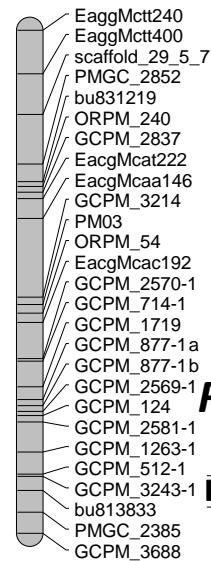
- Qualitative resistance (locus *Lrd1* ; Tabor et al, 2000)
- Genetic variability for quantitative resistance (Prakash & Haether 1989a)
- Interaction genotype/strain (Prakash & Heather 1989b, Tabor et al 2000)

*No map localisation*

## *P. nigra* x *P. nigra* F1

5 strains **Mlp**

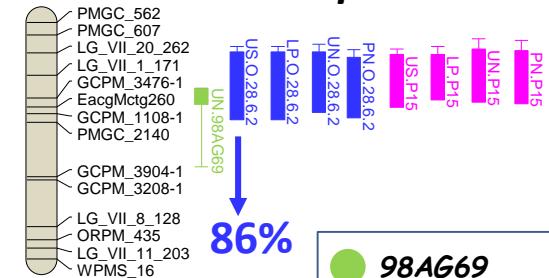
Chro 1



**RMLp1**



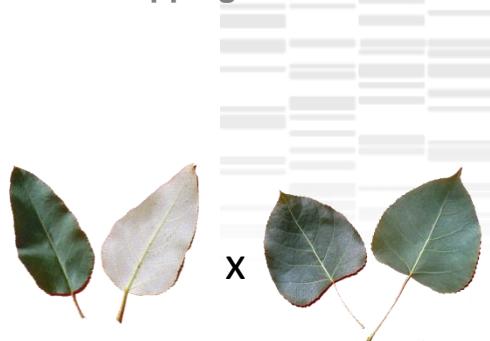
Chro 7



86%

61% ↓ 12%

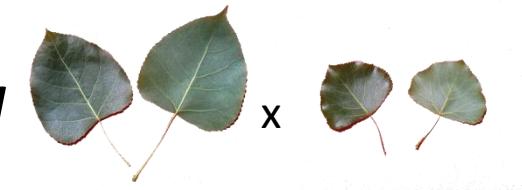
- 20 QTLs (2% - 86.2% variance explained)
- 2 major QTLs (chro 1 & 7, RMLp1, RMLp2)
- QTLs and their effect are strain specific (no QTL for OSSAS.28.5.6).
- Colocalisation with R-genes clusters
- « HR » mainly controlled by 1 locus



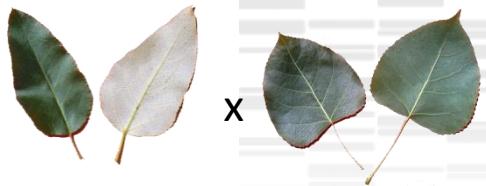
## Exapted pathosystem

bi-parental populations

**Mainly hybrid families used**



Family	Type	Taxon	Challenged against
Family 331	F2	<i>P. trichocarpa</i> x <i>P. deltoides</i>	<i>Mmd</i> <i>Mlp</i>
Family 545	F1	<i>P. trichocarpa</i> x <i>P. deltoides</i>	<i>Mcx</i>
Family 54	F1	<i>P. deltoides</i> x <i>P. trichocarpa</i>	<i>Mlp</i>
Family 87001	F1	<i>P. deltoides</i> x <i>P. trichocarpa</i>	<i>Mlp</i>
Family 87002	F1	<i>P. deltoides</i> x <i>P. nigra</i>	



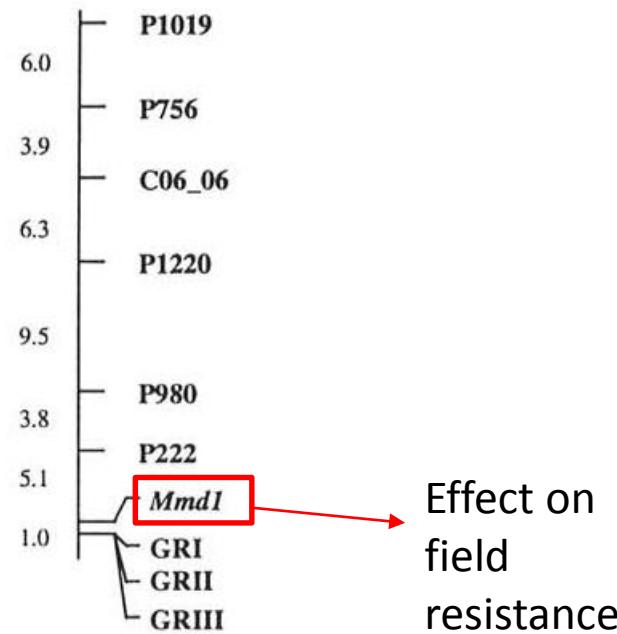
# Exapted pathosystem

bi-parental populations

Family 331, *P. trichocarpa* x *P. deltoides*

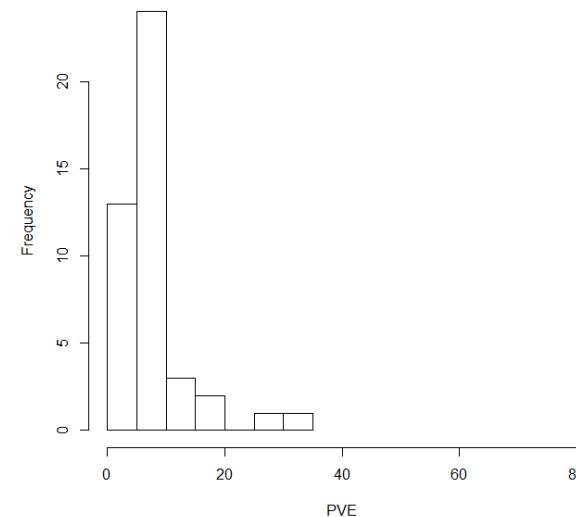
## 1 *Mmd* strain & field assessment

### Chromosome 4

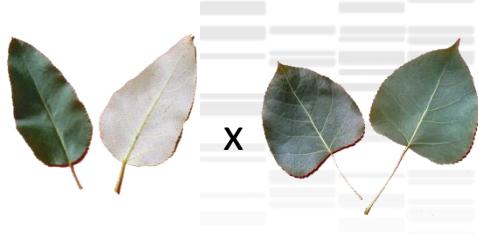


## 5 *Mlp* strains

- 17 regions (44 QTLs)
- The largest on chro 15
- Strain specific QTLs



The qualitatives R against *Mmd* is not working against *Mlp* !

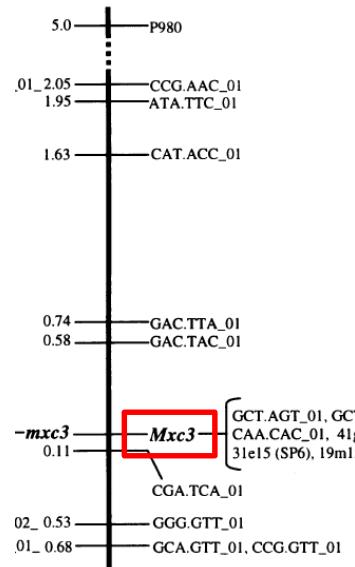


# Exapted pathosystem

bi-parental populations

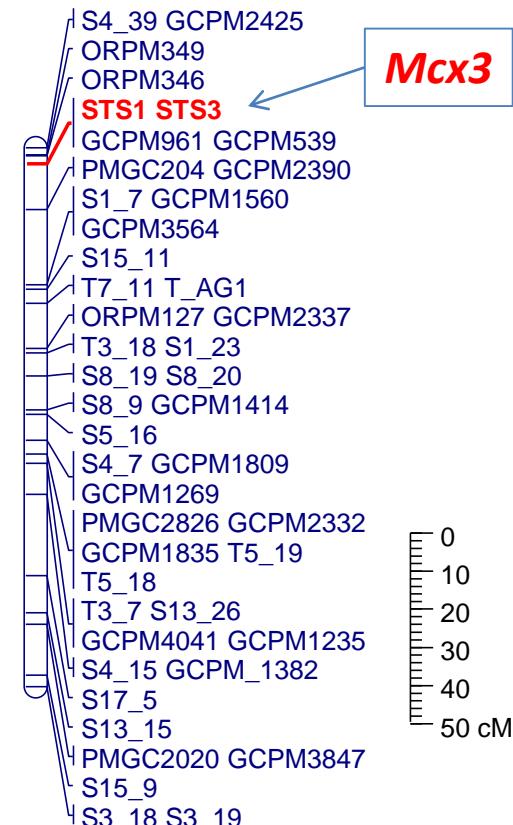
Family 545, *P. trichocarpa* x *P. deltoides*

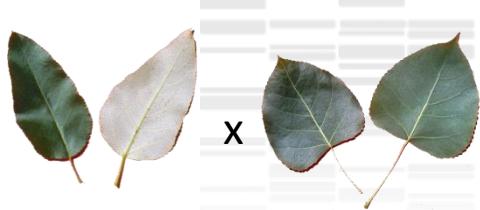
1 *Mcx* strain



Confirmed location on  
Chromosome 4

Fam 13

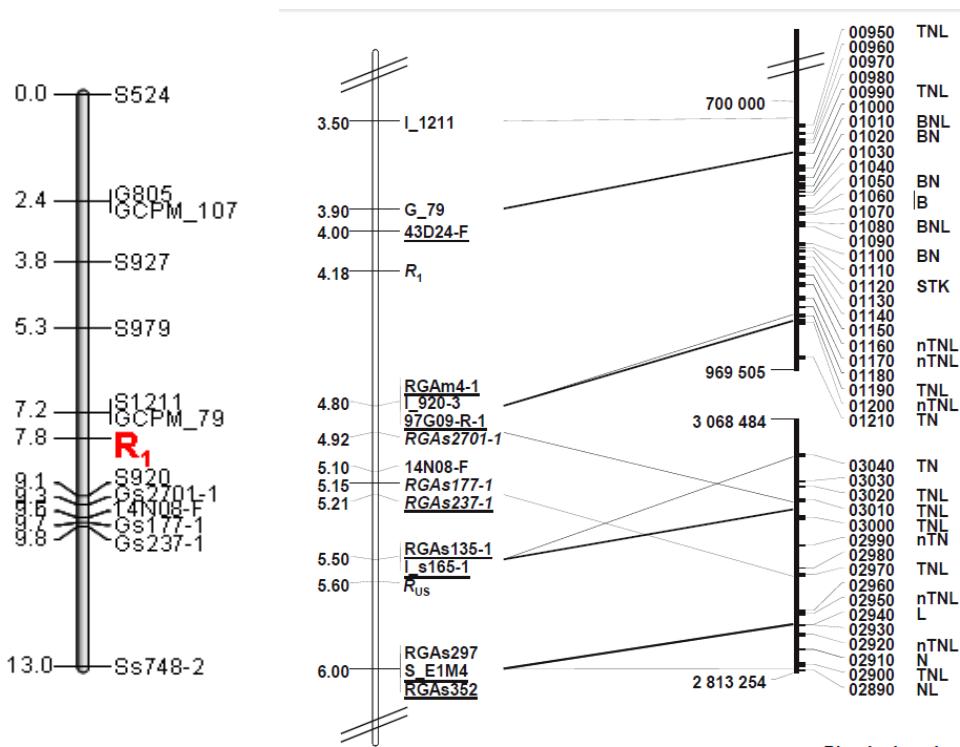




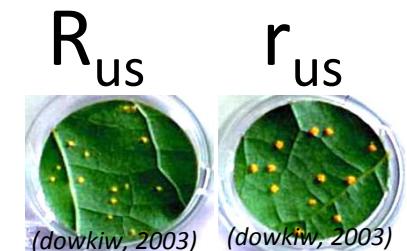
# Exapted pathosystem

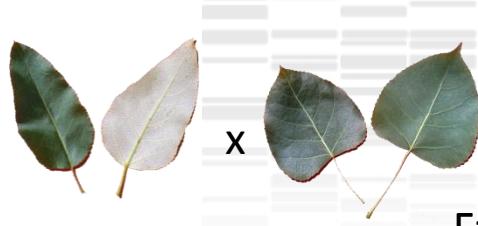
## bi-parental populations

Family 54 *P. deltoides* x *P. trichocarpa*  
5 *Mlp* & field assessment



- 2 major factors on Chromosome 19  
 $R_1$  inherited from *P. deltoides*  
 $R_{US}$  from *P. trichocarpa*
- Several minor QTLs
- Specificity
- Fine and physical mapping, cloning  $R_{US}$
- In major R-gene cluster





# Exapted pathosystem

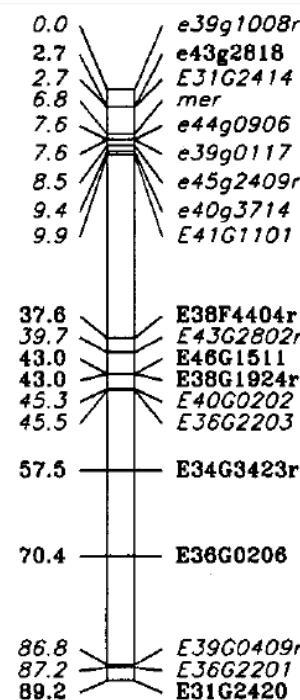
bi-parental populations

Family 87001 *P. deltoides* x *P. trichocarpa*

Family 87002 *P. deltoides* x *P. nigra*

3 **Mlp** strains

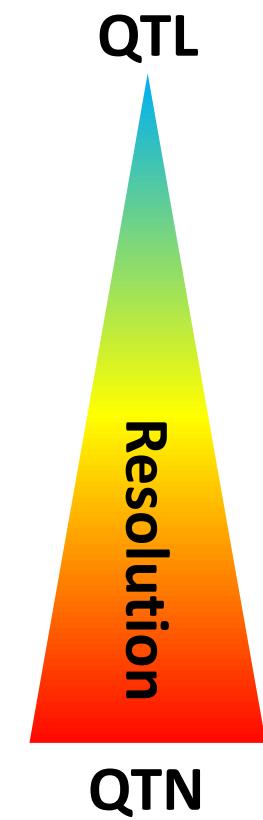
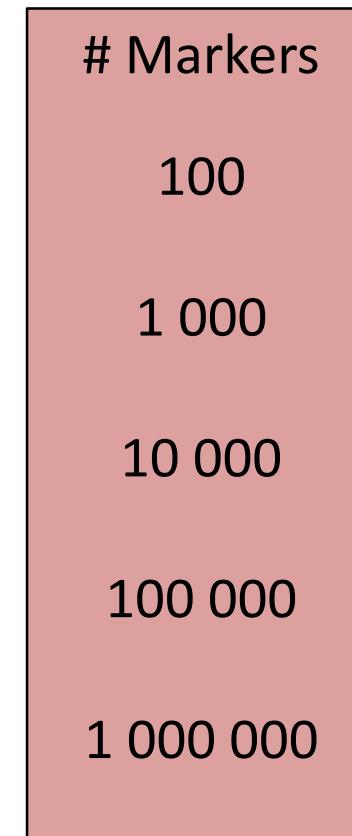
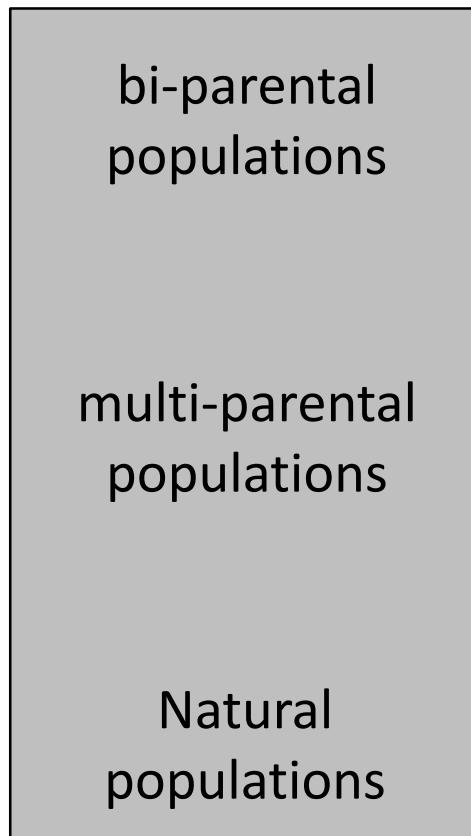
## Chromosome 19



**Mer** locus inherited from *P. deltoides*  
Fine and physical mapping, cloning failed

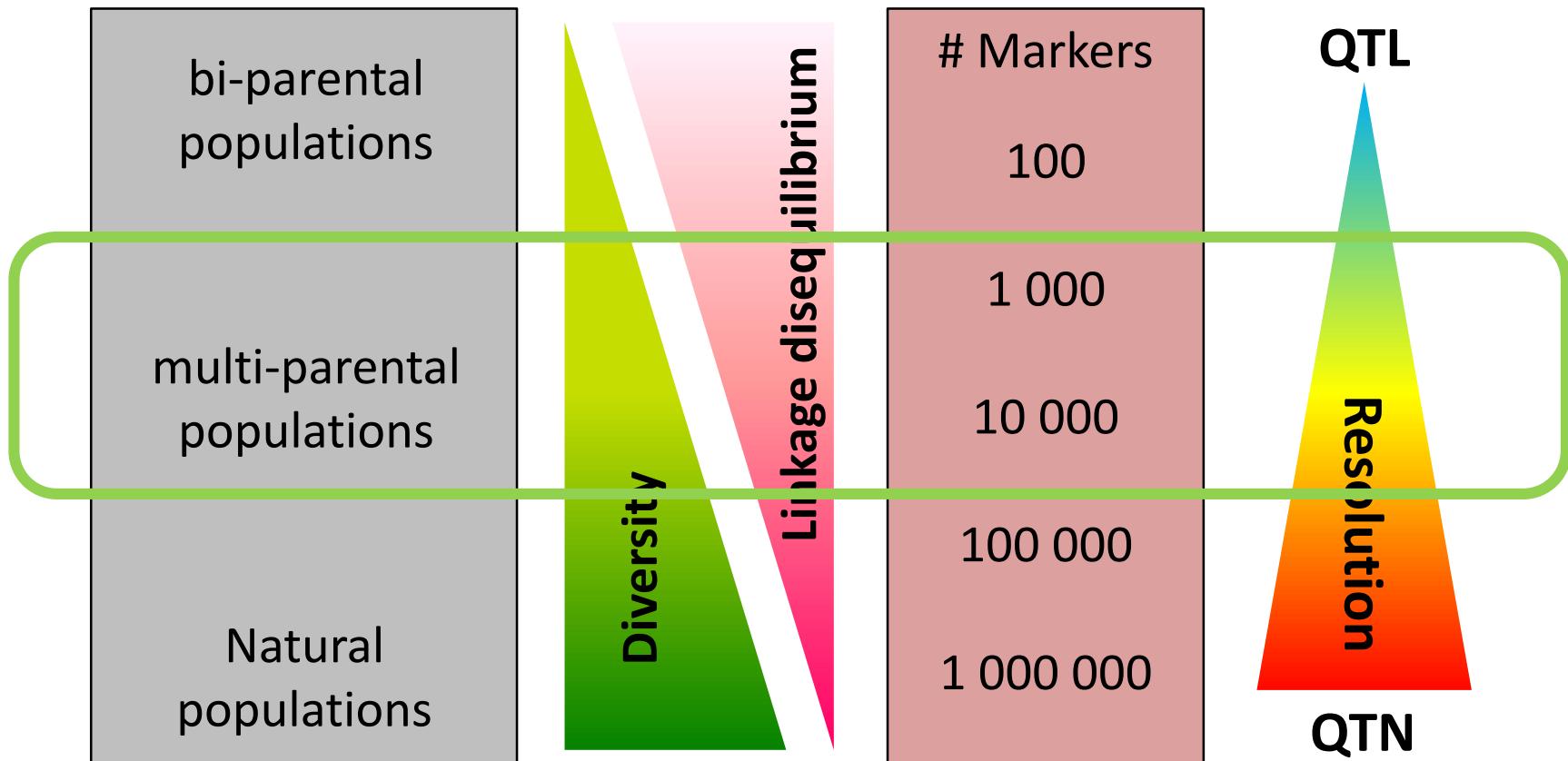
## Strategies to study genetic architecture of resistance

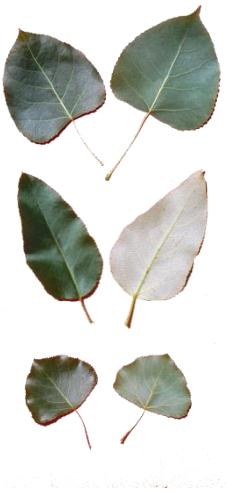
### From QTLs to QTNs



## Strategies to study genetic architecture of resistance

### From QTLs to QTNs





### 14 x 14 factorial mating design

	D 5	T 4	N 5
D 5			
T 4			
N 5	X		

118 FS families

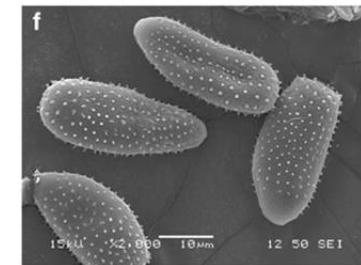
3480 clones

INRA  
1988-2000

## Natural and exapted pathosystem multiparental populations

*Melampsora larici-populina*

VS.



(Vialle et al 2011)

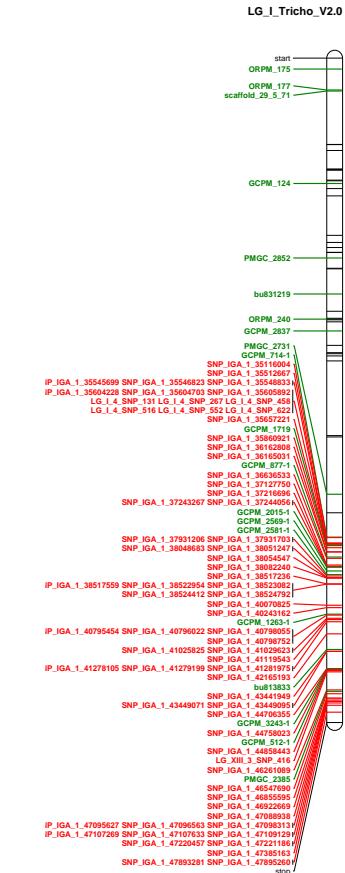
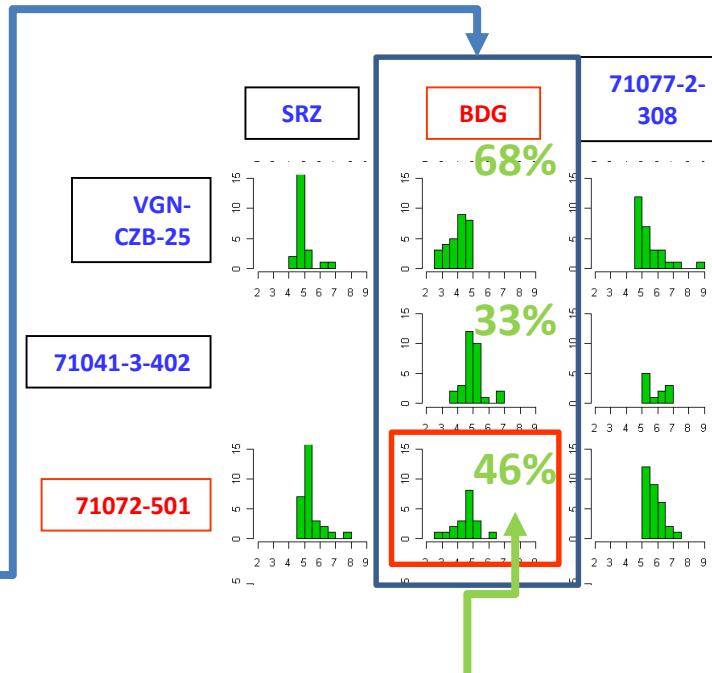
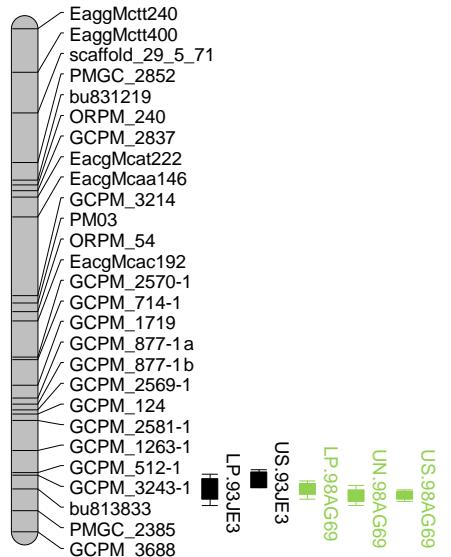
- Several characterized strains
- Field trials



# Natural pathosystem

## multiparental populations

### Chromosome 1



- In each family, markers explaining the variation for US
- The region of the QTL has been densify in markers (72 SNP on 11 Mb)
- Validation for strain 98AG69

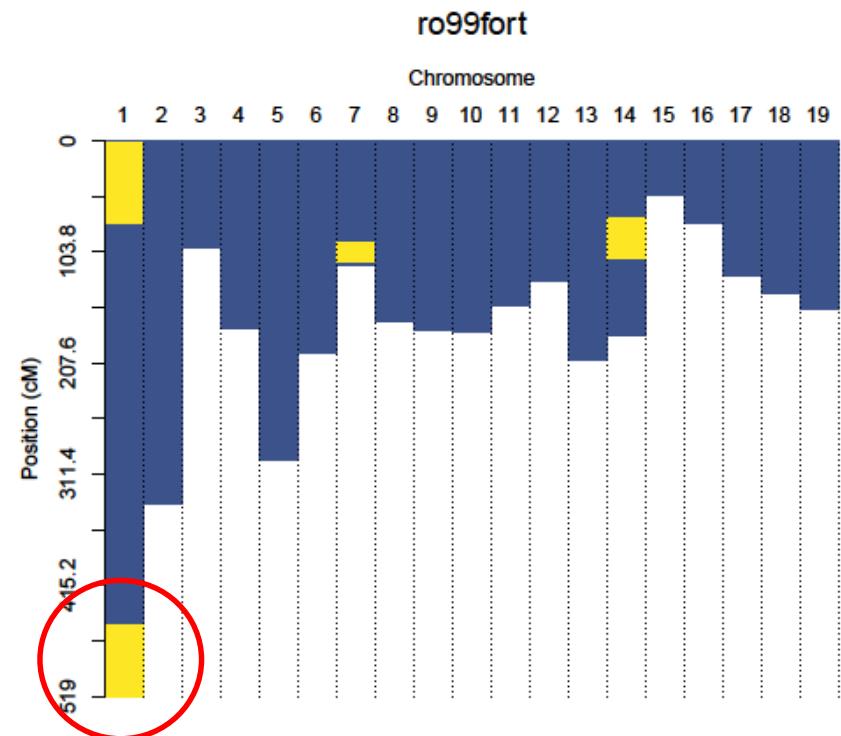
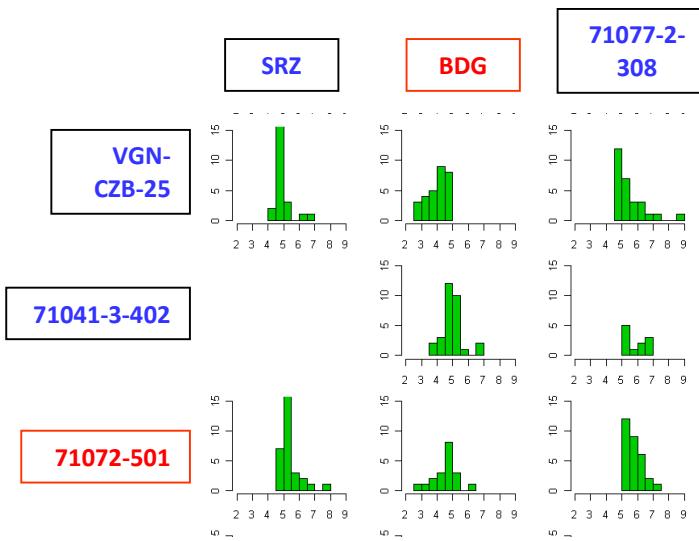


# Natural pathosystem

multiparental populations

QTL mapping in complex pedigree using FlexQTL (Bink et al. Univ. Wageningen)

## Rust scoring in field trial



Previously detected

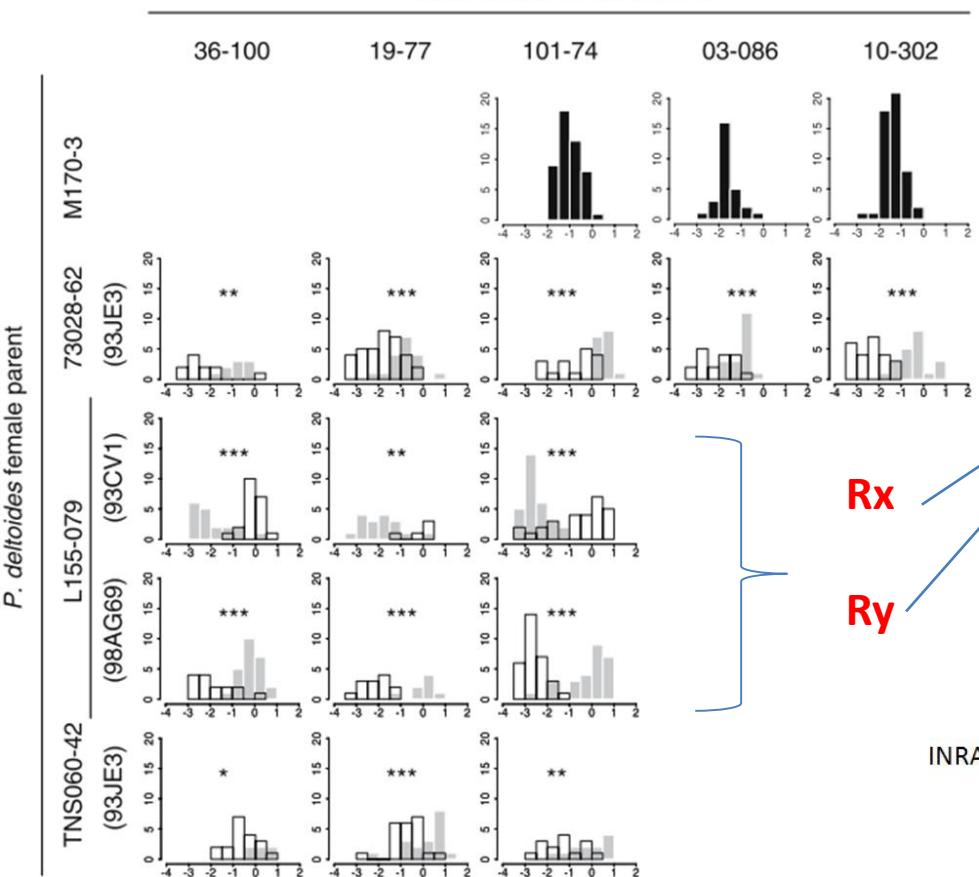
(Mangel et al, unpublished)

# Exapted pathosystem

## multiparental populations

2 *Mlp* strains

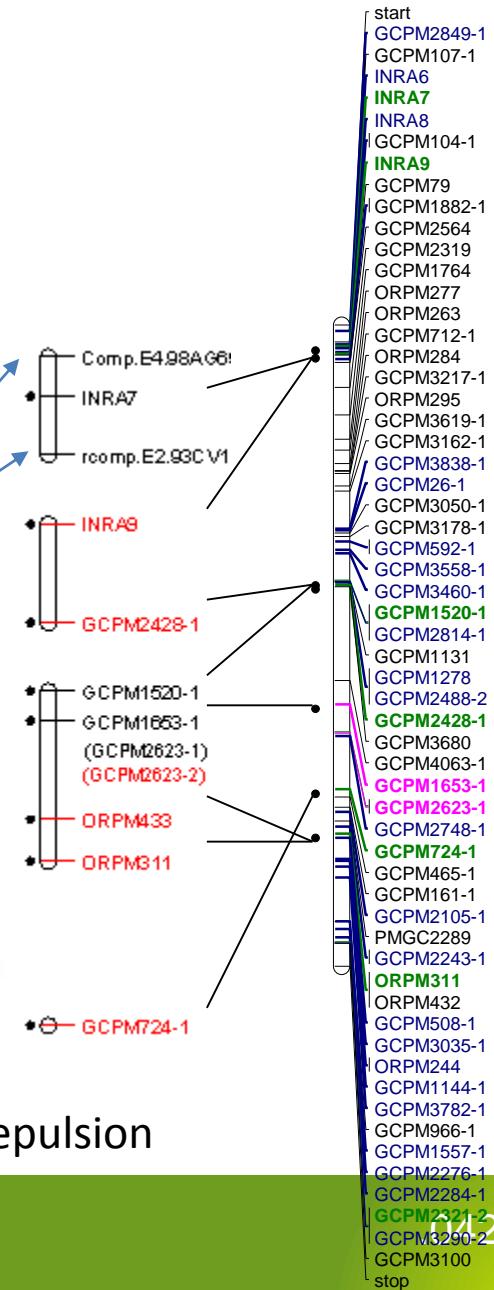
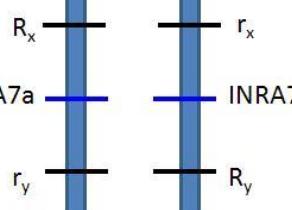
*P. trichocarpa* male parent



R1  
(slide 31)

Rx  
Ry

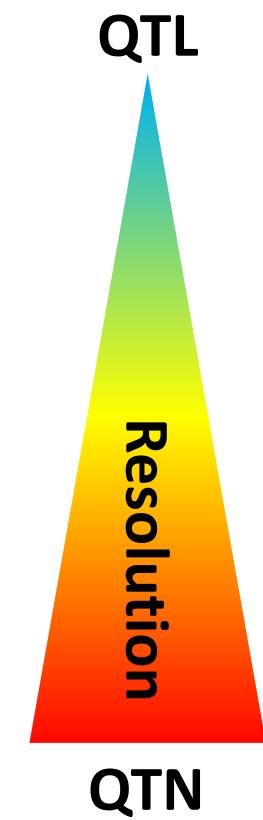
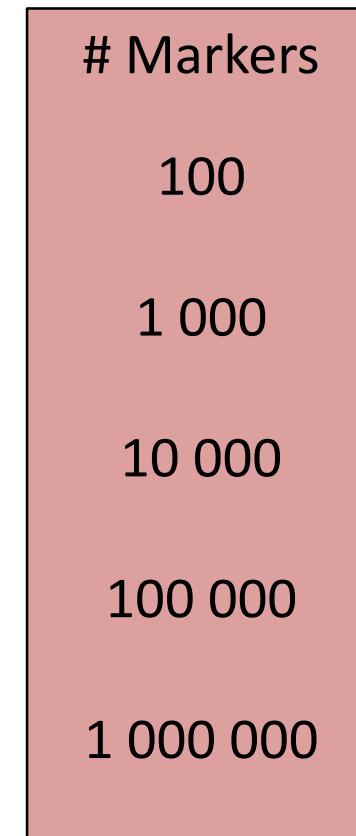
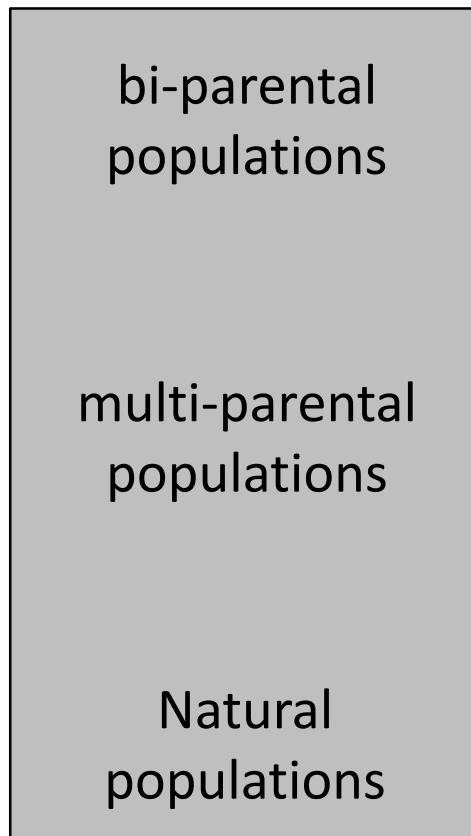
L155-079



2 additional loci linked in repulsion

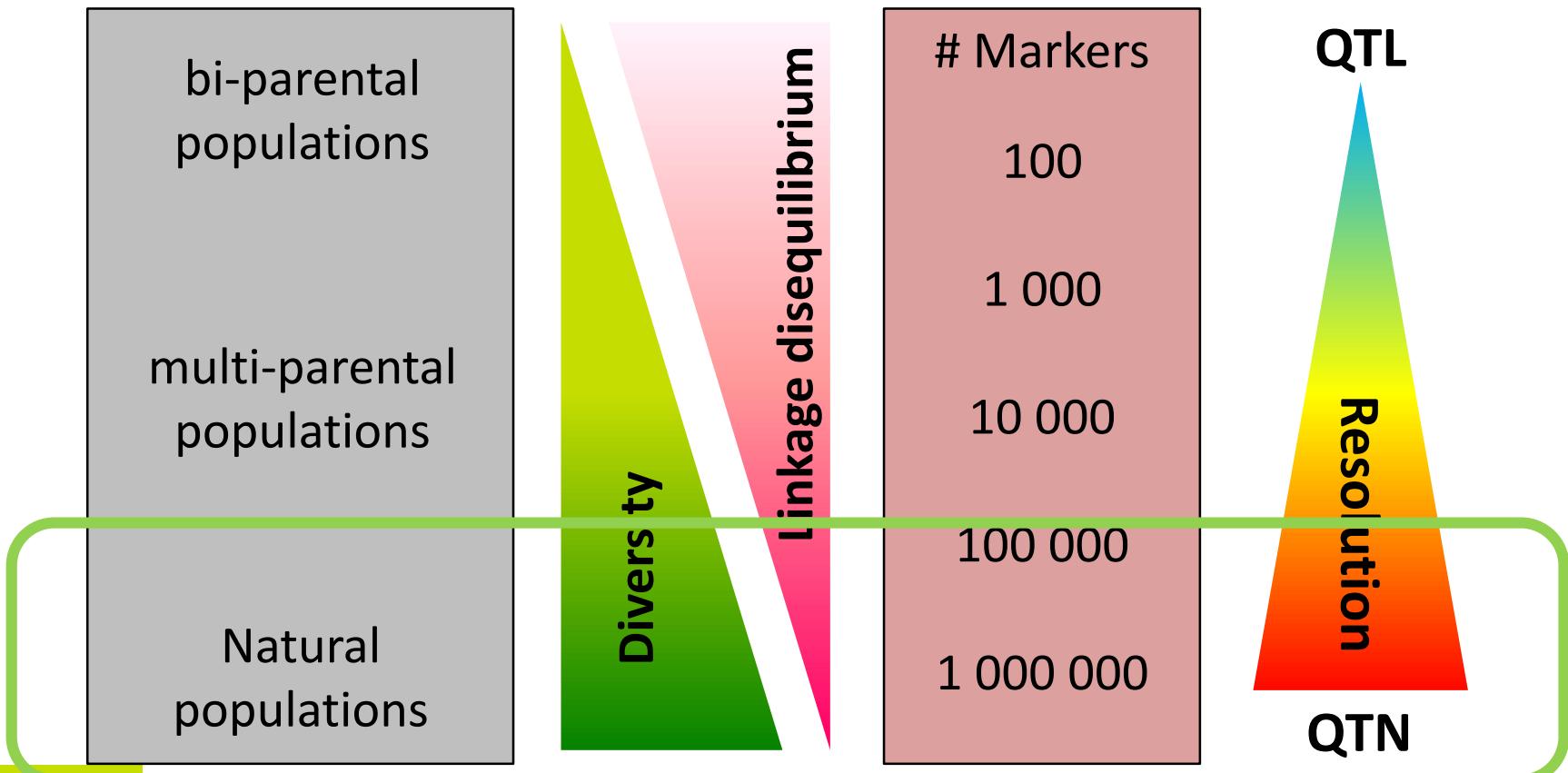
## Strategies to study genetic architecture of resistance

### From QTLs to QTNs



## Strategies to study genetic architecture of resistance

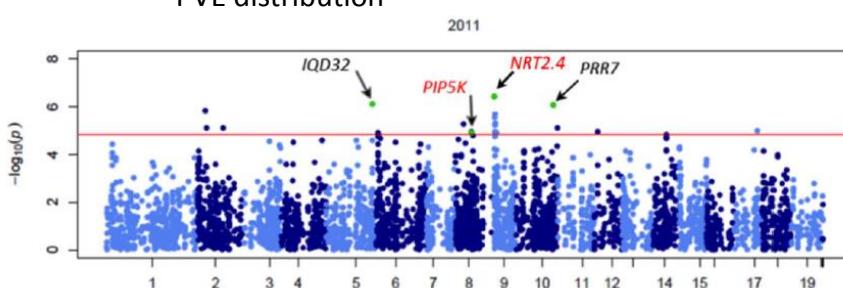
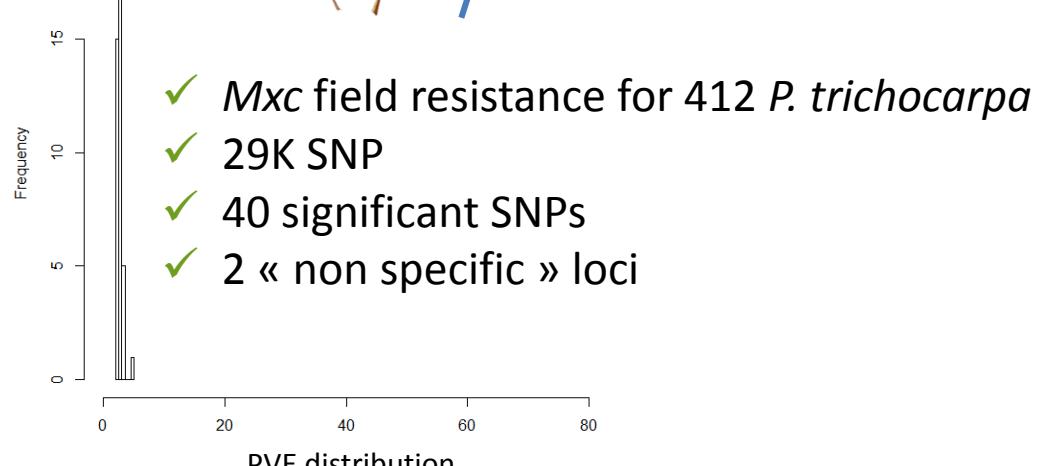
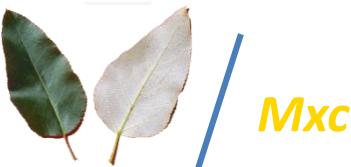
### From QTLs to QTNs



# Natural populations

## GWAS

LaMantia et al. PlosONE 2013

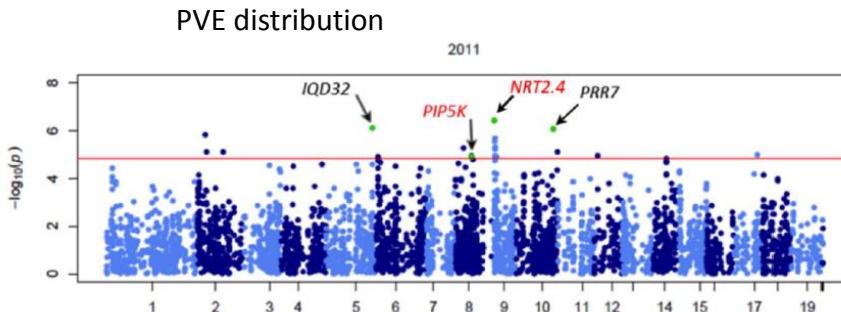
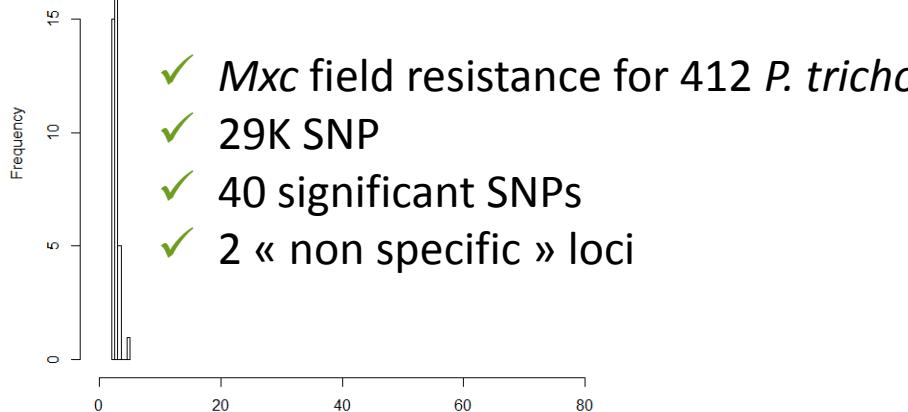
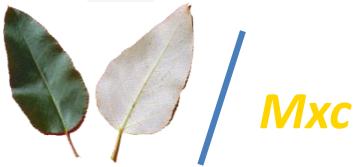


- ✓ 3 sign. SNP match with differentially expressed genes in *P. trichocarpa* x *P. deltoides* cv. Beaupré/Mlp interaction

# Natural populations

## GWAS

LaMantia et al. PlosONE 2013

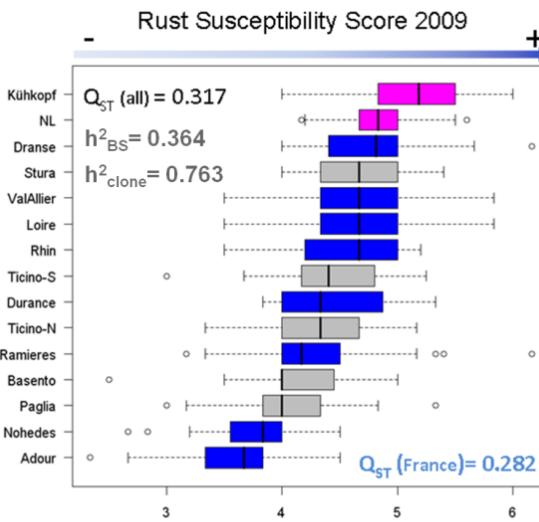


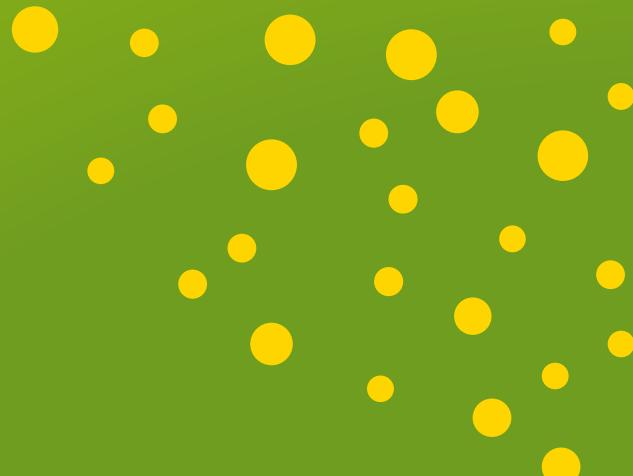
- ✓ 3 sign. SNP match with differentially expressed genes in *P. trichocarpa* x *P. deltoides* cv. Beaupré/*Mlp* interaction

Bastien et al, in prep



- ✓ Lab & Field resistance for 1100 *P. nigra*
- ✓ 8K SNP (focused on genes & candidate regions)
- ✓ To be done ...





# 6 Comparative genomics

# Sum up !

## Loci and QTL identification (« Heat map »)

### 4 mapping pedigrees

Family 331 : TDxTD

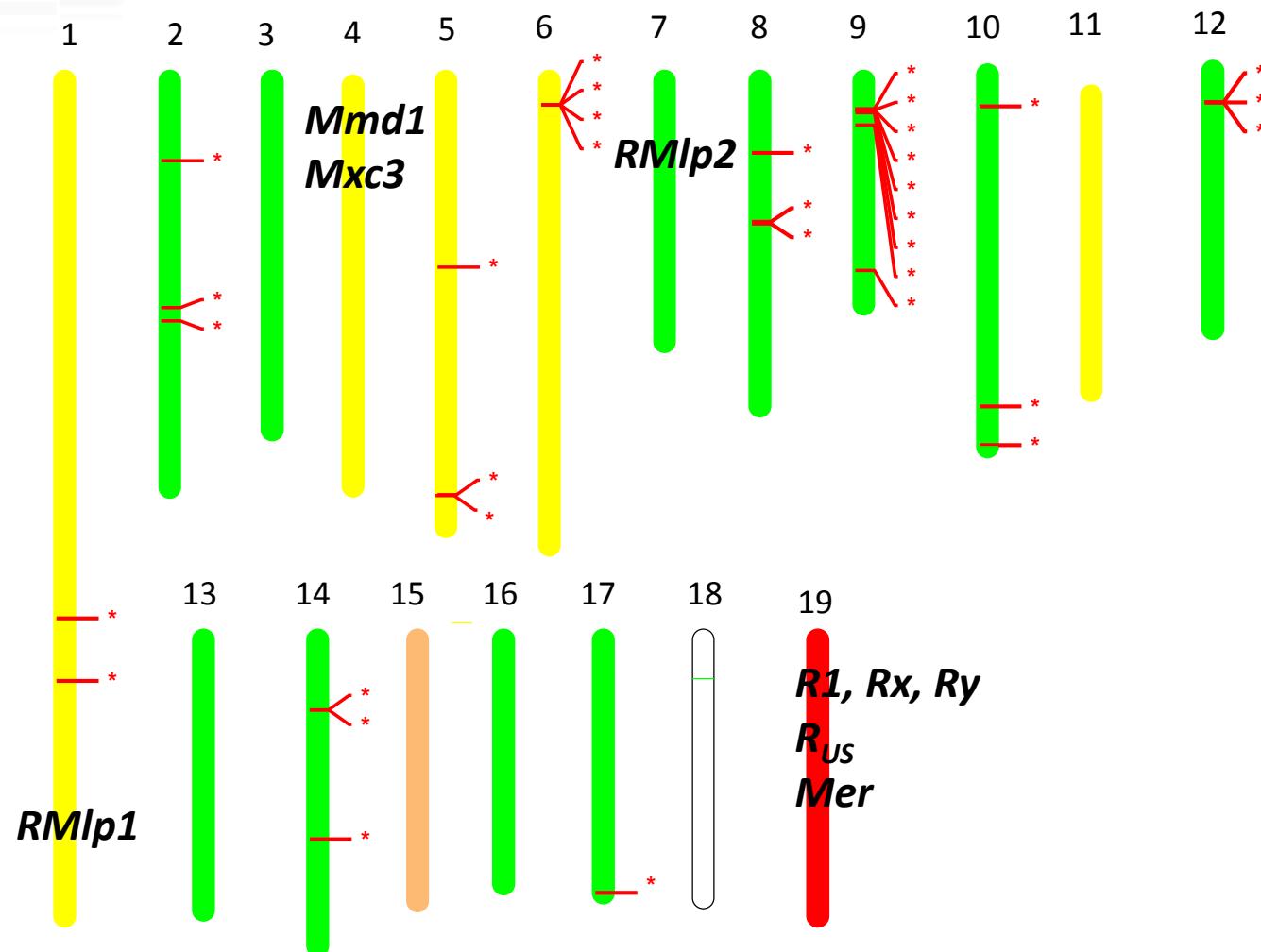
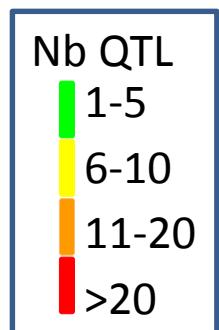
Family 54: DxT

87001,2 : DxN, DxT

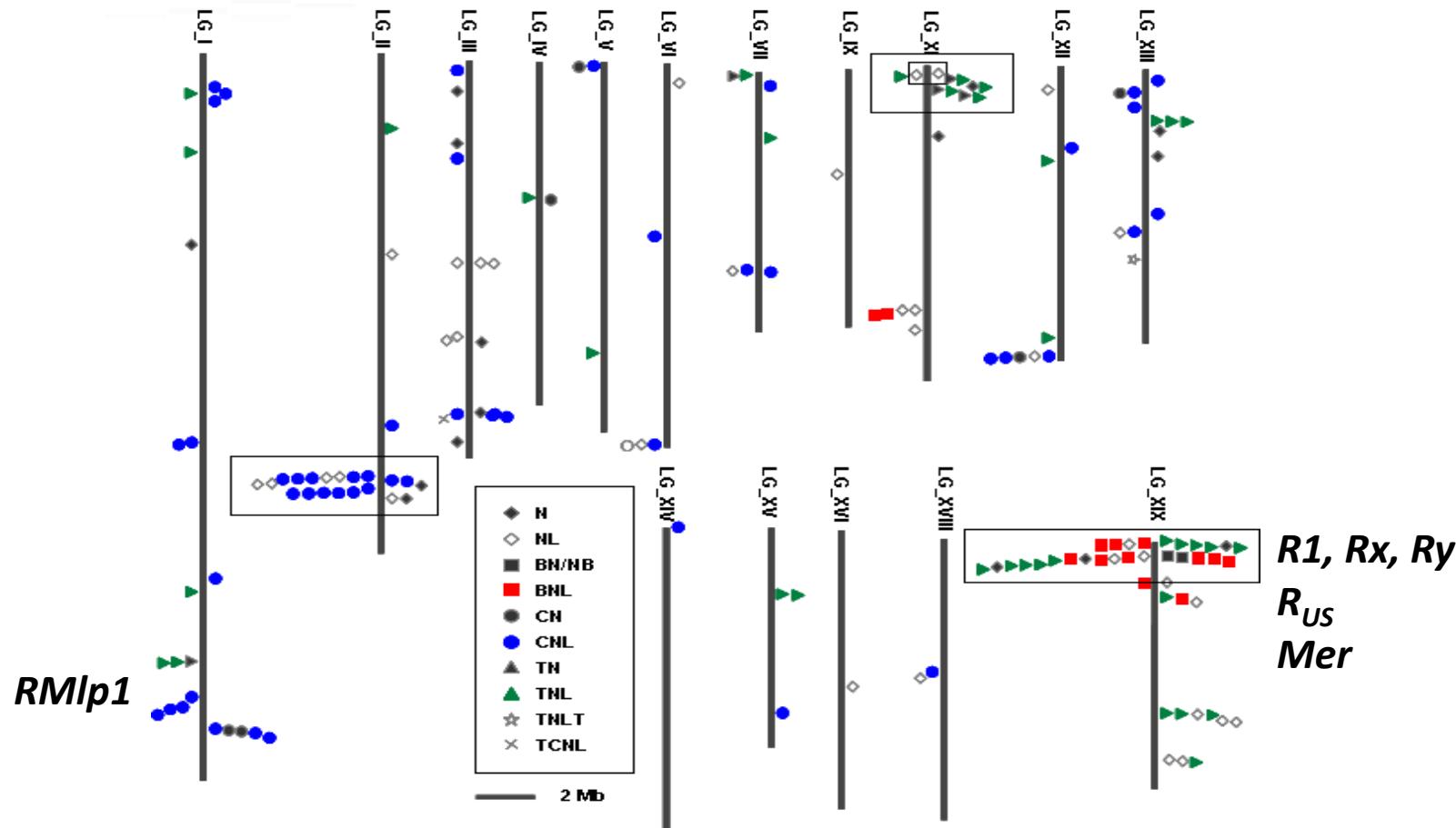
Family 1311: NxN

### *Factorial GWAS (\*)*

- Several components lab & field
- Several strains



# Some RGA clusters co-localise with resistance factors and QTLs described

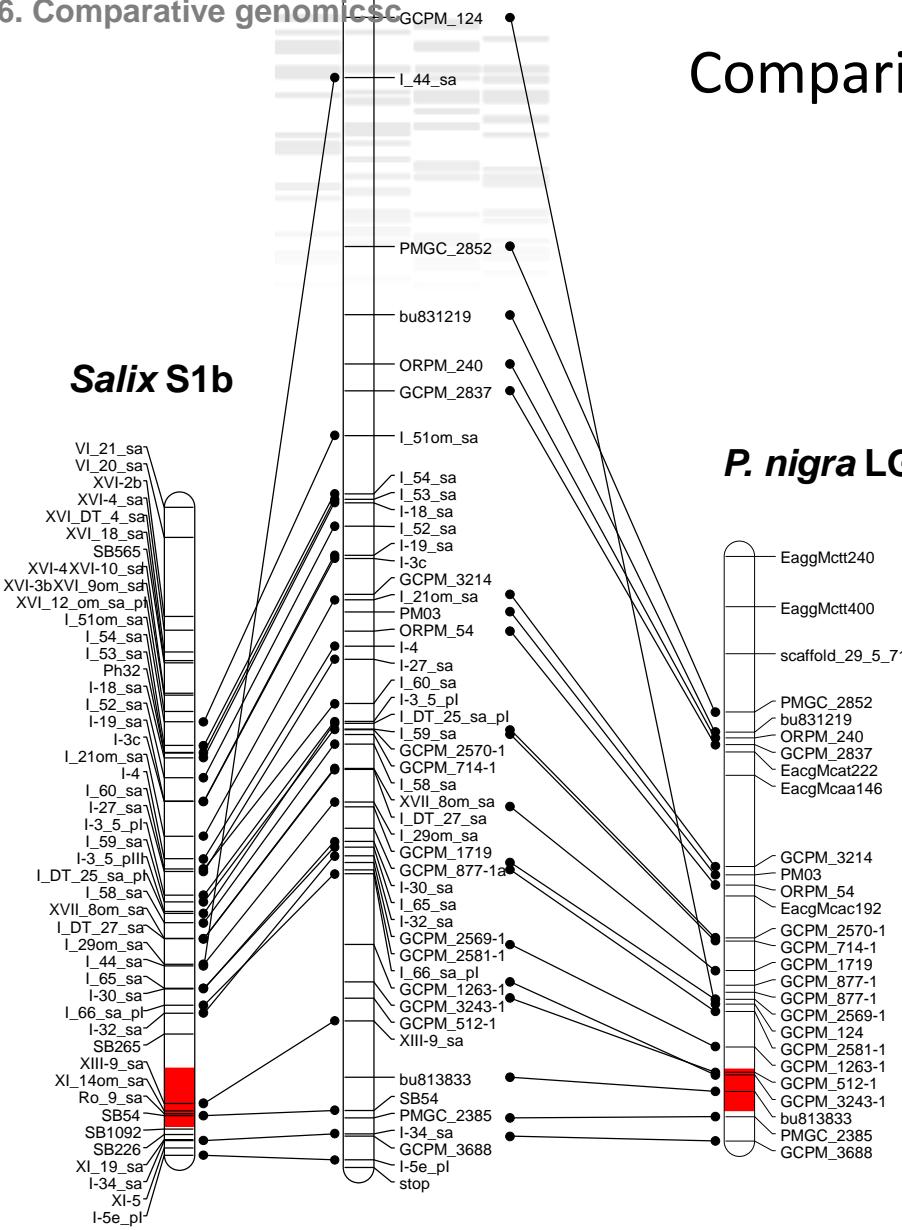
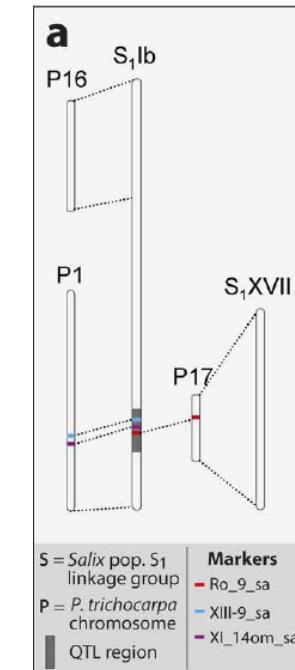


(Kolher et al 2008)

.049

# Comparison with *Salix* / *Mle* pathosystem

- Alignment of maps possible thanks to co-linearity between *Salix* and *Populus* genomes
- Genetic mapping show co-location
- But physical mapping and cloning not
- Some complex rearrangement between both genus

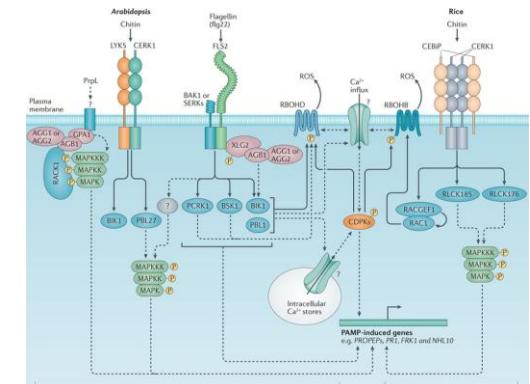
**Salix S1b****RMIp1**

(Samils et al, 2011)  
(Bertin et al 2016)

# Any pattern of loci identify with natural / exapted classification ?

Major Loci identified	<i>Melampsora occidentalis</i>	<i>Melampsora medusae f.sp. deltoideae</i>	<i>Melampsora larici-populina</i>
<i>Populus trichocarpa</i>	Natural ?	Exapted <i>Mmd1</i>	Exapted <i>Rus,</i>
<i>Populus deltoides</i>	?	Natural ?	Exapted <i>R1, Rx, Ry</i>
<i>Populus nigra</i>	?	Exapted <sup>a</sup> ?	Natural <i>RMIp1, RMIp2</i>

- Both control quantitative / qualitative resistance
- Both showed specificity
- No overlap between loci identified, expected ? =>



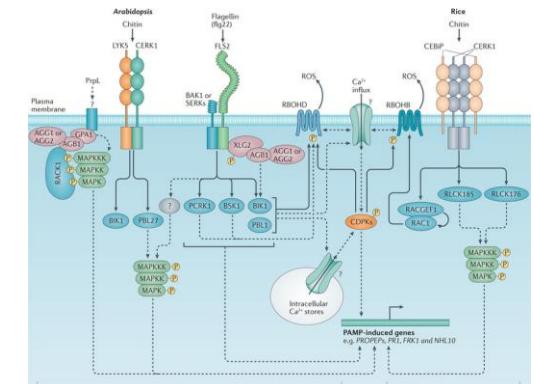
Couto & Zipfel, 2016

Nature Reviews | Immunology

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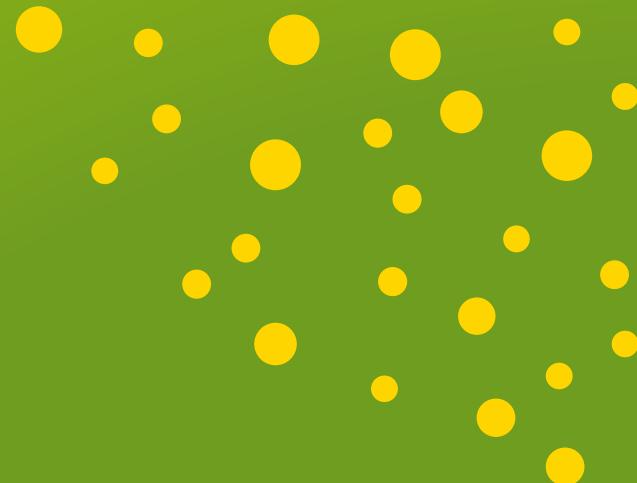
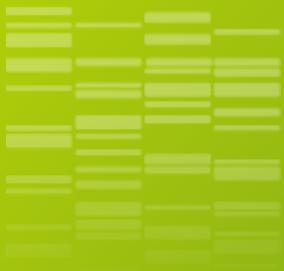
Couto & Zipfel, 2016

What are the function of exapted resistances in the natural area?

---

**7**

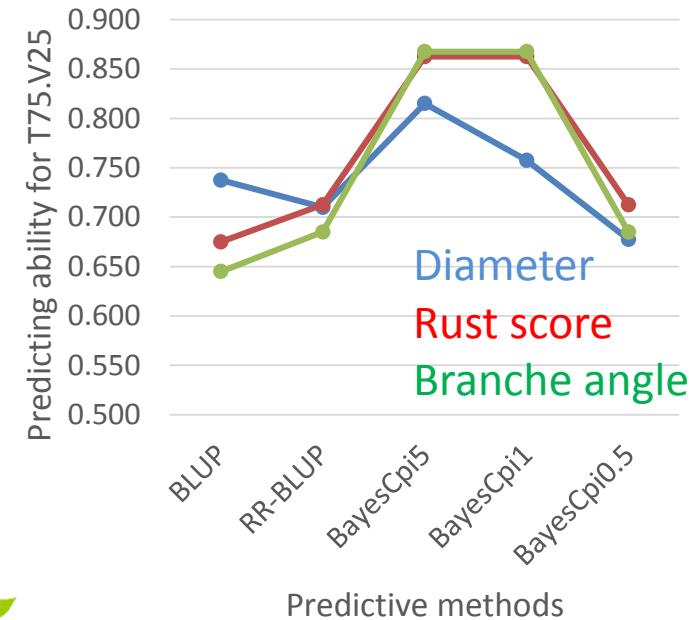
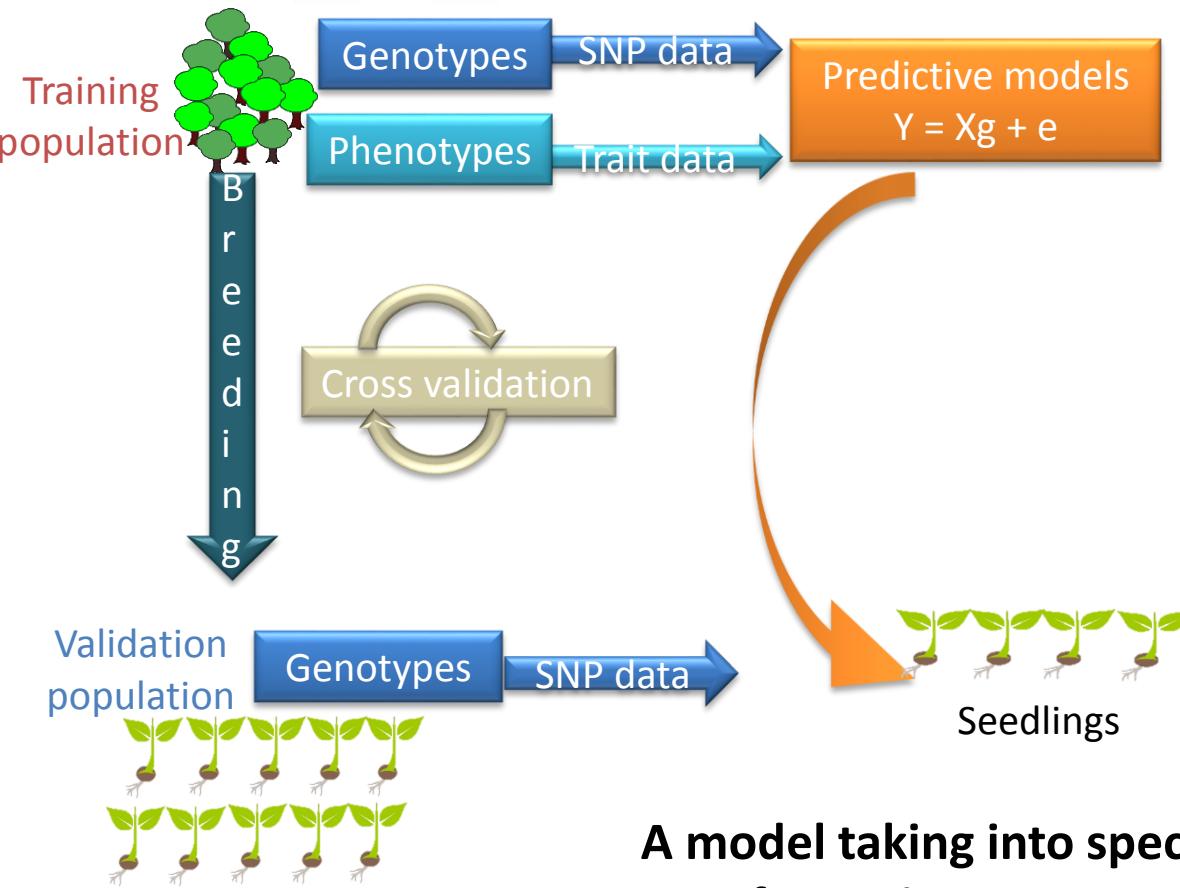
## Conclusion: potential uses for breeding





## A case study of genomic selection in *Populus nigra*

Which is the best predictive model for rust resistance ?



A model taking into specific marker effects on the trait.  
=> Information on genetic determinism is useful!

# Research in Poplar genetics and genomics at INRA (BioForA-Orleans & URGV-Evry)

*Cv 'italica'*



*P. nigra*



M. Villar

Natural populations

*Genetic diversity  
conservation  
dynamics of natural  
ecosystem*

*Cv 'italica'*



*P. nigra*



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M. Villar

*Other species and interspecific hybrids  
*P. deltoides, P. trichocarpa, DxN, DxT**

With focus on :

- *Resistance to Melampsora larici-populina*
- *Phenology*
- *Wood properties*

*Cv 'italica'*



*P. nigra*

M. Villar



**Natural populations**

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## *Populus Genomes*

*genome structure*

*Whole genome sequencing*

*SNP identification*

*HT genotyping*

*SSR*

**Molecular polymorphisms**

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P. Faivre-Rampant



V. Jorge

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V. Jorge



V. Segura

## *Genetic Maps and QTL detection* (*F1, factorial mating design*)

## *Association genetics in natural populations*

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Optimisation of breeding Strategies

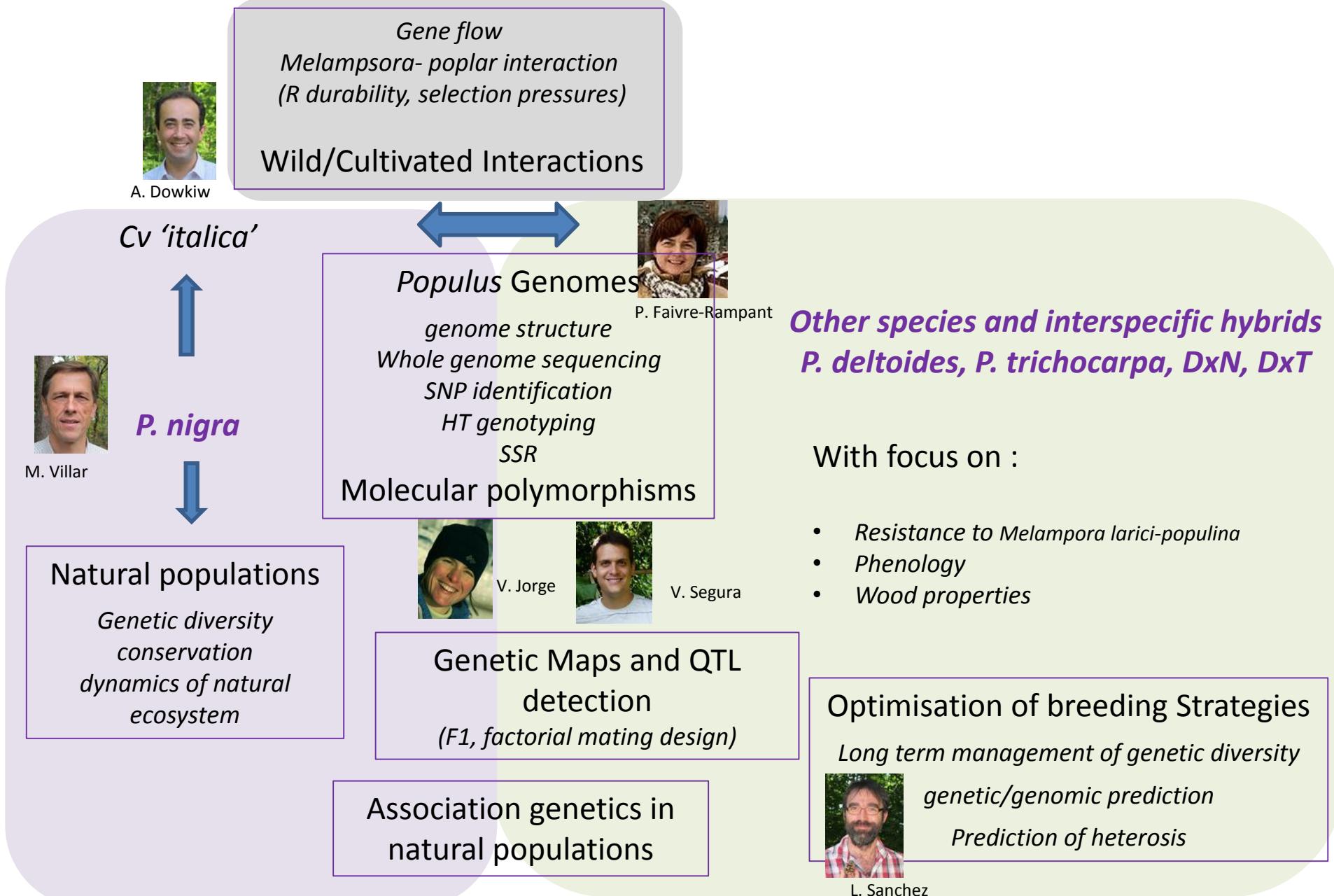
*Long term management of genetic diversity  
genetic/genomic prediction  
Prediction of heterosis*



L. Sanchez

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