



HAL
open science

Genome-wide analysis in response to N and C identifies new regulators for root AtNRT2 transporters

Sandrine Ruffel, Valentin Chaput, Jonathan Przybyla-Toscano, Ian Fayos, Catalina Ibarra, Tomas Moyano, Cécile Fizames, Pascal Tillard, Jose Antonio O'brien, Rodrigo A. Gutierrez, et al.

► **To cite this version:**

Sandrine Ruffel, Valentin Chaput, Jonathan Przybyla-Toscano, Ian Fayos, Catalina Ibarra, et al.. Genome-wide analysis in response to N and C identifies new regulators for root AtNRT2 transporters. 2019. <hal-02787649>

HAL Id: hal-02787649

<https://hal.inrae.fr/hal-02787649v1>

Preprint submitted on 5 Jun 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



HAL Authorization

1 **Short Title:** Systems biology approach to study NRT2s regulation

2

3 **Article title:**

4 Genome-wide Analysis In Response to N and C Identifies New Regulators for root AtNRT2
5 Transporters ¹

6

7 **Authors:**

8 Sandrine Ruffel^{a,2}, Valentin Chaput^{a,2}, Jonathan Przybyla-Toscano^a, Ian Fayos^a, Catalina
9 Ibarra^b, Tomas Moyano^b, Cécile Fizames^a, Pascal Tillard^a, Jose Antonio O'Brien^c, Rodrigo A.
10 Gutiérrez^b, Alain Gojon^a and Laurence Lejay^{a,3,4}

11

12 ^aBPMP, CNRS, INRA, Montpellier SupAgro, University Montpellier, Place Viala, 34060
13 Montpellier cedex, France

14 ^bDepartamento de Genética Molecular y Microbiología, Facultad de Ciencias Biológicas,
15 Millennium Institute for Integrative Biology, FONDAP Center for Genome Regulation,
16 Pontificia Universidad Católica de Chile, Santiago 8331150, Chile

17 ^cDepartamento de Genética Molecular y Microbiología, Facultad de Ciencias Biológicas.
18 Departamento de Fruticultura y Enología, Facultad de Agronomía e Ingeniería Forestal.
19 Pontificia Universidad Católica de Chile, Santiago 8331150, Chile

20

21 **One sentence summary:**

22 Identification of three transcription factors involved in the regulation of NRT2s transporters
23 using a systems biology approach and NRT2.1 as target gene in response to combinations of
24 N/C treatments

¹ This work was supported by an international grant from the French Research Agency (ANR) and Comision Nacional de Investigacion Cientifica y Tecnologica (CONICYT) (ModelN ANR-09-BLAN-0395).

² These authors contributed equally to this work

³ Author for contact: laurence.lejay@inra.fr

⁴ Senior author.

The author responsible for distribution of materials integral to the findings presented in this article in accordance with the policy described in the Instructions for Authors (www.plantphysiol.org) is: Laurence Lejay (Laurence.lejay@inra.fr)

SR performed the transcriptomic experiments the analysis and the generation of the gene regulatory network. SR and VC obtained and performed the experiments to characterise the mutants along with JPT, IF, PT and LL. JAO'B with CI performed Y1H experiments. TM, CF and RG performed bioinformatics and statistical analysis for the gene regulatory network and the interaction of the transcription factors with the NRT2 promoters. LL, SR, AG and RG designed the experiments. LL, SR and AG wrote the manuscript.

25 **Abstract**

26 In *Arabidopsis thaliana*, the High-Affinity Transport System (HATS) for root NO₃⁻ uptake
27 depends mainly on four NRT2 transporters, namely NRT2.1, NRT2.2, NRT2.4 and NRT2.5.
28 The HATS is the target of many regulations to coordinate nitrogen (N) acquisition with the N
29 status of the plant and with carbon (C) assimilation through photosynthesis. At the molecular
30 level, C and N signaling pathways have been shown to control gene expression of the *NRT2*
31 transporters. Although several regulators of these transporters have been identified in
32 response to either N or C signals, the response of *NRT2* genes expression to the interaction of
33 these signals has never been specifically investigated and the underlying molecular
34 mechanisms remain largely unknown. To address this question we used an original systems
35 biology approach to model a regulatory gene network targeting *NRT2.1*, *NRT2.2*, *NRT2.4* and
36 *NRT2.5* in response to N/C signals. Our systems analysis of the data highlighted the potential
37 role of three putative transcription factors, TGA3, MYC1 and bHLH093. Functional analysis
38 of mutants combined with yeast one hybrid experiments confirmed that all 3 transcription
39 factors are regulators of *NRT2.4* or *NRT2.5* in response to N or C signals.

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58 **Introduction**

59 As all living organisms, plants must integrate internal and external signals to adapt to
60 fluctuating environmental conditions. This is particularly the case concerning mineral
61 nutrition, because most nutrients display dramatic changes in external availability, whereas
62 their internal concentrations must be kept within a limited range to be compatible with
63 physiological processes. Accordingly, root nutrient uptake systems are finely tuned by
64 regulatory mechanisms activated by local signaling of external nutrient availability and
65 systemic signaling of the nutrient status of the whole plant (Schachtman and Shin, 2007).
66 Furthermore, acquisition of the various nutrients has to be coordinated to remain consistent
67 with the global chemical composition of plant tissues and with the fact that most nutrients
68 contribute to the synthesis of biomolecules with a relatively strict elemental stoichiometry
69 (e.g., C, N and S for amino acids). Therefore, the signaling pathways that are specific for the
70 different nutrients must interact to ensure this coordination. Although coordinated regulation
71 of uptake systems for different nutrients have been clearly demonstrated at the physiological
72 level, the underlying molecular mechanisms remain largely obscure (Schachtman and Shin,
73 2007). The cross-talks between N and C signaling mechanisms are certainly those that have
74 been most often investigated (Coruzzi and Zhou, 2001; Nunes-Nesi et al., 2010; Ruffel et al.,
75 2014), first because N and C are the two mineral nutrients plants require in largest quantities,
76 and also because they connect two key functions of plants as autotrophic organisms, *i.e.*,
77 photosynthesis and assimilation of inorganic nitrogen. Moreover, the importance of N/C
78 signaling interaction is dramatically illustrated by the fact that most N-responsive genes in
79 *Arabidopsis* are actually regulated by C/N interaction (Gutierrez et al., 2007).

80 The nitrogen nutrition of most herbaceous plants relies on the uptake of nitrate (NO_3^-), which
81 is ensured in root cells by two classes of transport systems. The High-Affinity Transport
82 System (HATS) is predominant in the low range of NO_3^- concentrations (up to \sim ca 1 mM),
83 whereas the Low-Affinity Transport System (LATS) makes an increasing contribution to total
84 NO_3^- uptake with increasing external NO_3^- concentration (Crawford and Glass, 1998). In all
85 species investigated to date, genes encoding the various transporter proteins involved in either
86 HATS or LATS have mostly been identified in the *NRT2* and *NPF* (formerly *NRT1/PTR*)
87 families, respectively (Nacry et al., 2013; O'Brien et al., 2016). The respective roles of HATS
88 and LATS in the total NO_3^- acquisition by the plant are still a matter of debate. However, field
89 studies suggest that even in agricultural conditions, the HATS has a major contribution over
90 the whole developmental cycle (Malagoli et al., 2004; Garnett et al., 2013). Both the structure

91 and regulation of the HATS have been extensively studied in *Arabidopsis thaliana*. In this
92 species, almost all the HATS activity depends on four NRT2 transporters, namely NRT2.1,
93 NRT2.2, NRT2.4 and NRT2.5 (Filleur et al., 2001; Kiba et al., 2012; Lezhneva et al., 2014),
94 which all require an interaction with the NAR2.1 protein to be active in NO₃⁻ transport (Kotur
95 et al., 2012). Under most conditions, NRT2.1 is the main contributor to the HATS (Cerezo et
96 al., 2001; Filleur et al., 2001). However, NRT2.4 and NRT2.5 display a very high-affinity for
97 NO₃⁻ and are important for taking up this nutrient when present at very low concentration
98 (<50 μM) in the soil solution (Kiba et al., 2012; Lezhneva et al., 2014). Furthermore, unlike
99 NRT2.1 and NRT2.4, NRT2.5 does not require the presence of NO₃⁻ to be expressed, and is
100 therefore considered crucial for ensuring the initial uptake of NO₃⁻ as soon as it appears in the
101 external medium (Kotur and Glass, 2015).

102 Most interestingly, the HATS has been shown to be the target of almost all regulations
103 governing root NO₃⁻ acquisition in *Arabidopsis* (Nacry et al., 2013), and this is associated
104 with control of *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* expression at the mRNA level. In
105 particular, previous reports have shown that *NRT2.1* is induced both by N starvation (Lejay et
106 al., 1999; Cerezo et al., 2001; Gansel et al., 2001), and by light and sugars, indicating
107 coordination with photosynthesis (Lejay et al., 1999; Lejay et al., 2003). This makes *NRT2.1*
108 a very relevant model gene for investigating the interaction between N and C signalling
109 networks in roots. This also holds true for *NRT2.4* (Lejay et al., 2008; Kiba et al., 2012), but
110 not for *NRT2.5*, which until now has only been reported to be up-regulated by N starvation
111 (Lezhneva et al., 2014). For these reasons, and also due to its high functional importance as
112 the main component of the HATS, *NRT2.1* has been extensively investigated to unravel its
113 regulatory mechanisms. Accordingly, a quite significant number of genes have been found to
114 encode regulators of *NRT2.1* expression, such as *LBD37-39* (Rubin et al., 2009), *TGA1* and
115 *TGA4* (Alvarez et al., 2014), *NLP6* and *NLP7* (Marchive et al., 2013; Guan et al., 2017),
116 *NRG2* (Xu et al., 2016), *BT1-2* (Araus et al., 2016), *NRT1.1* (Munos et al., 2004), *CIPK8* (Hu
117 et al., 2009), *HNI9/IWS1* (Widiez et al., 2011) and *HY5* (Chen et al., 2016). Most of these
118 genes contribute to the regulation of *NRT2.1* expression in response to changes in N
119 provision. The only exception is *HY5*, which encodes a transcription factor reported to ensure
120 long-distance signalling of the stimulation of *NRT2.1* expression in roots by illumination of
121 the shoot. Strikingly, none of the above regulators were shown to be involved in the cross-talk
122 between N and C signalling pathways. Even more surprising, the response of *NRT2.1*
123 expression itself (as well as those of the other *NRT2s*) to the interaction of N and C signals

124 was not specifically investigated. As a consequence, the molecular mechanisms responsible
125 for the coordinated regulation of the NO_3^- HATS by N and C status of the plant are unknown.
126 Our study aimed at filling this gap. Therefore, using *NRT2.1* as a marker gene to identify
127 relevant combinations of N/C treatments, we developed a systems biology approach based on
128 genome-wide transcriptome analysis in roots of *Arabidopsis* plants to model a regulatory gene
129 network targeting *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* in response to N/C signals. This
130 highlighted the potential role of three putative transcription factors, TGA3, MYC1 and
131 bHLH093 in controlling the expression of these transporter genes. Functional analysis of loss-
132 of-function mutants confirmed that all 3 transcription factors are regulators of *NRT2.4* or
133 *NRT2.5* in response to N or C signals. Furthermore, yeast one hybrid experiments confirmed
134 that at least TGA3 and MYC1 are able to bind *NRT2.4* and *NRT2.5* promoters.

135

136

137 **Results**

138 **Regulation of root nitrate transporters by interaction between nitrogen and light** 139 **provision**

140 We wished to determine whether induction of *NRT2.1* by N starvation is dependent on light,
141 and conversely if *NRT2.1* induction by light is dependent on the availability of NO_3^- (Figure
142 1A and Figure 1B). In order to reveal possible interactions between C and N signalling
143 pathways for the regulation of *NRT2.1*, we performed two different sets of experiments. In the
144 first set of experiments, plants were starved for N for up to 72h either in the dark or at three
145 different light intensities, $50 \mu\text{mol m}^{-2}\text{s}^{-1}$ (LL), $250 \mu\text{mol m}^{-2}\text{s}^{-1}$ (IL) and $800 \mu\text{mol m}^{-2}\text{s}^{-1}$
146 (HL) (Figure 1A). In the second set of experiments, plants were treated with 10mM NO_3^- ,
147 1mM NO_3^- or no N and transferred during 8h from the dark to HL conditions (Figure 1B).

148 In LL and IL conditions, *NRT2.1* expression was, as expected, induced when plants were
149 starved for N even if both the kinetic and the level of induction were different depending on
150 light intensity (Figure 1A). When plants were kept in the dark, *NRT2.1* expression was not
151 induced by N starvation but it remained very low both on 10mM NO_3^- and on N free solution.
152 More surprisingly, the induction of *NRT2.1* expression by N starvation was also almost
153 completely abolished when plants were treated in HL conditions. However, under HL *NRT2.1*
154 mRNA levels were always high, even under repressive conditions such as 10mM NO_3^- . This
155 unexpected result is specific of *NRT2.1* since *NRT2.2*, *NRT2.4* and *NRT2.5*, known to be also
156 induced by N starvation in roots (Li et al., 2007; Kiba et al., 2012; Lezhneva et al., 2014), are
157 still regulated by N starvation in HL (Supplemental Figure 1A). However, just like *NRT2.1*,
158 *NRT2.2*, *NRT2.4* and *NRT2.5* were not regulated by N starvation in the absence of light.
159 These data confirm the need of light for the regulation by N starvation of root NO_3^-
160 transporters. It also suggests that the mechanisms involved in *NRT2.1* regulation by N
161 starvation are somewhat different from the mechanisms involved in the regulation of *NRT2.2*,
162 *NRT2.4* and *NRT2.5*.

163 The second set of experiments confirmed the strong interaction between C/N signals as it
164 revealed that the level of N nutrition affects the regulation of *NRT2.1* expression by light
165 (Figure 1B). Indeed, when plants were starved for N during 48h, *NRT2.1* expression was
166 much less induced by light as compared to plants grown on 10 or 1mM NO_3^- (Figure 1B).
167 Among other root NO_3^- transporters, only *NRT2.2* and *NRT2.4* were induced by light and their
168 level of induction seemed to be also dependent on N nutrition (Supplemental Figure 1B).
169 However, in contrast to *NRT2.1*, the level of expression of both *NRT2.2* and *NRT2.4* was high

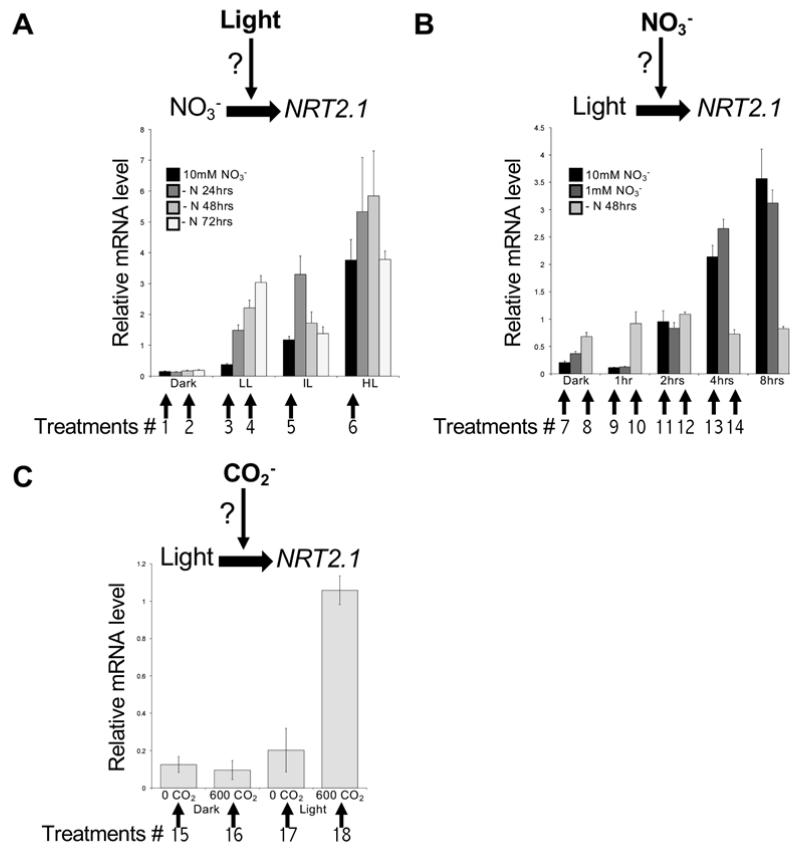


Figure 1. Interaction between Nitrogen and Light/Carbon provision modulates *NRT2.1* mRNA accumulation in roots.

A. Different light regimes modulate *NRT2* regulation in roots of plants experiencing from high NO₃⁻ provision (10 mM) to N deprivation (-N). The light regimes encompass dark, low light intensity (50 μmol m⁻² s⁻¹; LL), intermediate light intensity (250 μmol m⁻² s⁻¹; IL) and high light intensity (800 μmol m⁻² s⁻¹; HL). Plants were supplied with NO₃⁻ 10 mM one week ahead the experiment and acclimated for 24 hours in the different light regimes before applying the N deprivation for 24, 48 or 72 hr.

B. Different N provisions modulate *NRT2* regulation in roots of plants experiencing a dark to light transition. The N provisions encompass 10mM NO₃⁻, 1mM NO₃⁻ (for 72 hr) and N deprivation for 48 hr (-N). Plants are kept in the dark (*i.e.*, 40hr) before transition to high light intensity (800 μmol m⁻² s⁻¹; HL) and roots are collected at time 0 (Dark) and 1, 2, 4 and 8 hr after light transition.

C. Regulation of *NRT2* by photosynthesis activity. Plants are grown in regular NO₃⁻ regime (1mM) and intermediate light intensity until they are transferred for 4 hr in a CO₂-deprived atmosphere (0ppm) or in high CO₂-supplied atmosphere (600ppm), either in the dark or in the light.

In these 3 experimental conditions, roots have been collected to assess *NRT2.1* mRNA accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Expression pattern of *NRT2.1* across the 35 conditions tested (16 in A, 15 in B and 4 in C) has driven the choice of 18 conditions to investigate gene reprogramming associated to the regulation of NO₃⁻ transport. These 18 conditions are indicated with arrows and numbers on the x-axis of the 3 *NRT2.1* bar graphs (Each arrow corresponds to one condition with 2 independent biological repeats constituted of a pool of approx. 10 plants each).

170 when plants were starved for N and low when plants were grown on 1 or 10mM NO₃⁻
 171 (Supplemental Figure 1B). For *NRT2.4*, it confirms that this transporter is more sensitive to
 172 high N repression than *NRT2.1* (Kiba et al., 2012). The same result was obtained for *NRT2.5*,
 173 whose expression is barely detectable on either 10mM or 1mM NO₃⁻ (Supplemental Figure

174 1B). However, concerning regulation by light, even when *NRT2.5* expression was high in N
175 starved plants, light did not induce but rather seemed to repress *NRT2.5* mRNA accumulation
176 after 8h in the light (Supplemental Figure 1B).

177 In a previous study, we showed that expression of *NRT2.1* and *NRT2.4* is induced by light
178 only in the presence of CO₂ in the atmosphere, suggesting that light regulation of these genes
179 corresponds to a control exerted by photosynthesis (Lejay et al., 2008). As in the rest of our
180 study we used micro-array experiments to look for genes involved in the regulation of root
181 NO₃⁻ transporters by photosynthesis, it was important for us to be able to discriminate between
182 genes regulated by light itself or by photosynthesis. To do so, we performed a third set of
183 experiments where plants were transferred from dark to light for 4h in an atmosphere
184 containing 0 or 600ppm CO₂. The results confirmed (i) that both *NRT2.1* and *NRT2.4* are only
185 induced by light in the presence of CO₂ and (ii) that *NRT2.5* is not induced by light or
186 photosynthesis as suggested by our previous experiment (Figure 1C and Supplemental Figure
187 1C).

188

189 **Gene network for the regulation of root nitrate transporters by light and N starvation**

190 The experiments performed above allowed us to reveal interesting interactions between C and
191 N regulation of root NRT2 NO₃⁻ transporters. We took advantage of this experimental design
192 to develop a systems biology approach aiming at inferring a gene regulatory network
193 underlying the interactions between N and C signals in the regulation of root NO₃⁻
194 transporters. Due to the central position of *NRT2.1* as regulatory target affecting N
195 acquisition, we used it as a focus gene around which to find associated gene networks.

196 We performed Affymetrix microarrays on selected combinations of light and N treatments,
197 which were found discriminant for regulation of *NRT2.1*. Altogether, we chose 18 treatments
198 labelled with numbered arrows in Figure 1. These 18 treatments correspond to 3 sets of
199 conditions representative of (i) the light-dependent induction of *NRT2.1* expression in
200 response to N starvation, (ii) the light induction of *NRT2.1* on 10mM NO₃⁻ and (iii) the
201 specific regulation of *NRT2.1* by photosynthesis and not by light itself. For each treatment, 2
202 independent biological replicates were generated and used for Affymetrix ATH1 microarray
203 hybridization.

204 Regulation of *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* gene expression in response to N
205 starvation and light/photosynthesis was similar on microarrays as compared to the results
206 obtained by quantitative PCR in Figure 1 and Supplemental Figure 1 (Supplemental Figure
207 2). These results also confirmed that these four NO₃⁻ transporters are the main *NRT2s*

208 expressed in roots. *NRT2.3*, *NRT2.6* and *NRT2.7* showed very low expression levels on the
209 microarrays under our experimental conditions. It is also noteworthy that *NRT2.1* was the
210 most highly expressed member of the family among the 7 *NRT2*s (5 to 50 fold higher
211 expression as compared to *NRT2.2*, *NRT2.4*, *NRT2.5*) (Supplemental Figure 2).

212 To find gene regulatory networks that could integrate N and C signalling and thus control
213 *NRT2.1* expression, we defined 5 different subsets of conditions addressing the regulation by
214 N on one side and by C on the other side, as described in Figure 2A. Genes defined as
215 regulated by N-deprivation like *NRT2.1* are differentially regulated by N provision in
216 conditions 1 to 4 in experiment 1, where *NRT2.4* is also found regulated and in conditions 7 to
217 14 in experiment 2, where *NRT2.2* and *NRT2.5* were also found regulated. To select the most
218 robust genes regulated by N provision only the intersection between the 2 groups was
219 isolated. In addition to *NRT2.1*, the intersection defines a set of 33 genes including the 2
220 transcription factors *TGA3* (At1g22070) and *MYC1* (At4g00480). On another hand, genes
221 considered as regulated by C provision like *NRT2.1* are differentially regulated by light
222 intensity in conditions 1, 3, 5, and 6 in experiment 1, by light time exposure in conditions 9,
223 11 and 13 in experiment 2 and by photosynthesis in conditions 15 to 18 in experiment 3.
224 Similarly, to narrow down the specificity of gene regulation by C factor, only common genes
225 to at least 2 experiments were isolated. This core set corresponds to 142 genes including
226 *NRT2.1* but also 2 others transcription factors *bHLH093* (At5g65640) and *ARR14*
227 (At2g01760) (Figure 2A).

228 Next, we only focused on the 174 genes that showed a response to N starvation (34 genes)
229 and/or C provision (142 genes); *NRT2.1* being the common gene between the 2 responsive
230 gene lists together with a *Kinesin3* gene (At5g54670-ATK3) coding for a microtubule motor
231 protein. The possible connection of the 4 transcription factors with *NRT2.1* and the other
232 genes was determined by a Gene Networks analysis performed on the VirtualPlant platform
233 (Katari et al., 2010). The generated network contains 124 gene nodes. These genes are
234 connected to each other by 260 edges, representing regulatory relationships such as predicted
235 transcription factor-target gene interactions (Figure 2B). Regulatory interactions were
236 proposed based on detection of at least one predicted binding site for a given transcription
237 factor within the promoter region of the target gene as done previously (Gutierrez et al.,
238 2008). According to the parameters used, 50 genes out of the 174 are not connected to any
239 other genes in the network (See Material and Methods for details about the parameters).
240 Among these 50 genes, the transcription factor *ARR14* was excluded due, for instance, to a
241 low level of correlation between this gene and *NRT2.1* expression patterns. However, *TGA3*,

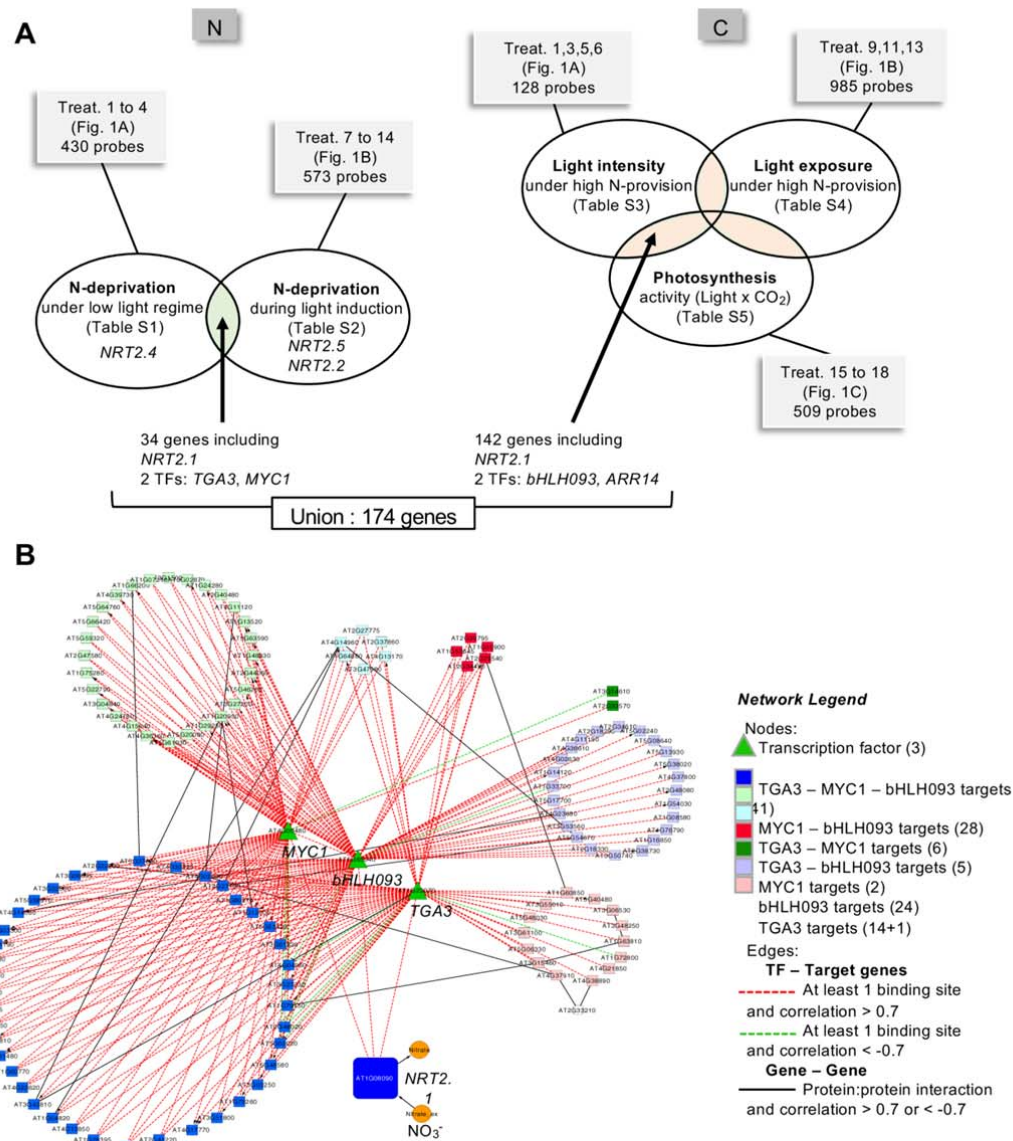


Figure 2. Gene expression multi-analysis driven by *NRT2.1* expression pattern combined to an integrative analysis identified a candidate gene regulatory network connected to the NO_3^- transport system.

A. Venn diagrams identifying common genes regulated by N provision on low light condition and dark to light transition (34 genes) or regulated by light/carbon (142 genes). The union of these gene lists defines a population of 174 genes, including 4 transcription factors.

B. The core set of 174 genes differentially expressed has been structured into a Gene Regulatory Network using the Gene Networks analysis tool in VirtualPlant software (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>) (32). The network includes 124 nodes (genes) and 260 edges connecting genes. The nodes have been organized according to their connection to the 3 transcription factors *MYC1*, *TGA3* and *bHLH093* and are detailed in the Network Legend. *ARR14* is excluded from the network due to its lack of connectivity to other nodes according to the edges selected to generate the network.

242 MYC1 and bHLH093 have all predicted regulatory interactions with *NRT2.1* plus 40 other
 243 genes of the network (indicated in blue in Figure 2B). The network predicts also that only one
 244 or only two of these transcription factors putatively regulate the 79 remaining genes (one gene
 245 being connected to the network by predicted protein-protein interaction with 2 TGA3-targets).

246 Nevertheless, almost all sub-networks are interconnected through protein-protein interaction
247 prediction, suggesting possible coordination within the network at the whole.

248

249 **Regulation of *MYC1*, *TGA3* and *bHLH093* in response to C and N**

250 The gene regulatory network we obtained revealed 3 main transcription factors: MYC1 and
251 TGA3 which were found to be co-regulated with *NRT2.1* in response to N starvation and
252 bHLH093 which was found to be co-regulated with *NRT2.1* in response to
253 light/photosynthesis. In order to validate their regulation, we measured gene expression by
254 QPCR across all the conditions performed in experiment 1 and 2 (Figure 3A). The results
255 confirmed that expression of *TGA3* and *MYC1* genes is induced 2- to 3-fold after transferring
256 the plants to a N-free solution, especially under LL or HL conditions. Furthermore, similar to
257 *NRT2.1*, *MYC1* regulation of gene expression requires the presence of light (Figure 3A and
258 Supplemental Figure 3). The results also confirmed that *bHLH093* gene expression is only
259 induced by light (between 3- and 4-fold after 8h of HL), independent of N nutrition. This is
260 supported by the fact that *bHLH093* is not regulated by N starvation (Figure 3A and
261 Supplemental Figure 3). On the contrary, *MYC1* and *TGA3* genes are not only regulated by N
262 starvation, but their expression is also induced by light, especially in plants starved for N
263 (Figure 3A and Supplemental Figure 3). Like for *NRT2.1*, putative *cis*-binding elements for
264 TGA3, MYC1 and bHLH093 were also found in the promoters of *NRT2.2*, *NRT2.4* and
265 *NRT2.5* (Figure 3B). Furthermore, DNA affinity purification sequencing (DAP-seq)
266 experiments recently performed by O'Malley et al. (2016) confirmed that TGA3 binds in
267 silico to the promoter of *NRT2.1*, *NRT2.2* and *NRT2.4* (Figure 3C). Unfortunately, no data are
268 available for MYC1 and bHLH093 in this work. Altogether, these results suggest that the
269 transcription factors we identified are involved in regulation of several root *NRT2s*.

270 To our knowledge, the transcription factors TGA3, MYC1 and bHLH093 have not
271 been isolated in previous transcriptomic approach as candidates for regulation of root NO₃⁻
272 transporters. In order to understand why they have not been found before we looked at the
273 expression pattern of the known regulatory elements for *NRT2.1* in our experimental set up.
274 The results show that the known regulators for *NRT2.1* were not co-regulated with *NRT2.1*
275 expression in our conditions (Figure 4). This was also the case for *HY5*, a transcription factor
276 recently identified as involved in the regulation of *NRT2.1* by light/photosynthesis (Chen et
277 al., 2016). In our hands, this transcription factor was only induced by light independently of
278 the presence of CO₂ and therefore not by photosynthesis like *NRT2.1* (Figure 4). As most of
279 the previous transcriptomic experiments were performed to study the signalling pathways

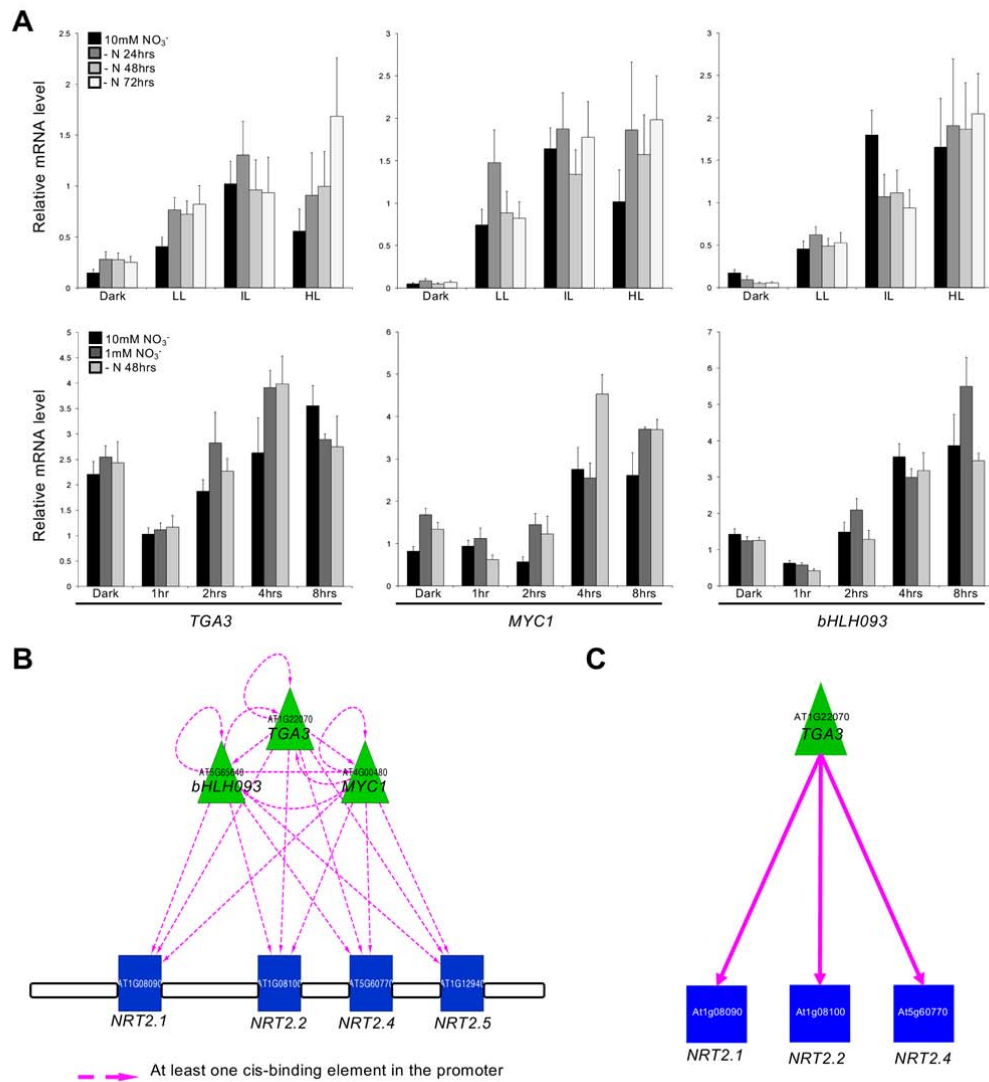


Figure 3. *TGA3*, *MYC1* and *bHLH093* are candidate transcription factors for the control of the expression of *NRT2* gene family.

A. Gene expression analysis of the 3 candidate transcription factors in the extended set of Nitrogen/Carbon combinations confirms correlation with *NRT2.1* regulation. Expression patterns have been determined by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene).

B. *NRT2.2*, *NRT2.4* and *NRT2.5* as well as *NRT2.1* display putative cis-binding elements for the 3 transcription factors in their promoter region. The gene network has been done using the Gene Networks analysis tool in VirtualPlant software (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>) (32); only Regulated Edges box and One Binding Site option has been selected in this case.

C. *TGA3* binds *in silico* with the promoter of *NRT2.1*, *NRT2.2* and *NRT2.4*. The analysis has been done using the Plant Cistrome Database (<http://neomorph.salk.edu/PlantCistromeDB/>) (63).

280 involved in short-term induction by NO₃⁻, we also looked at the regulation of *TGA3*, *MYC1*
 281 and *bHLH093* in those conditions (Supplemental Figure 4). We chose the transcriptomic
 282 experiments performed by Wang et al. (2004). In this study WT plants and the null mutant for
 283 nitrate reductase (NR) were treated with 5mM KNO₃ for 2h and compared to control plants

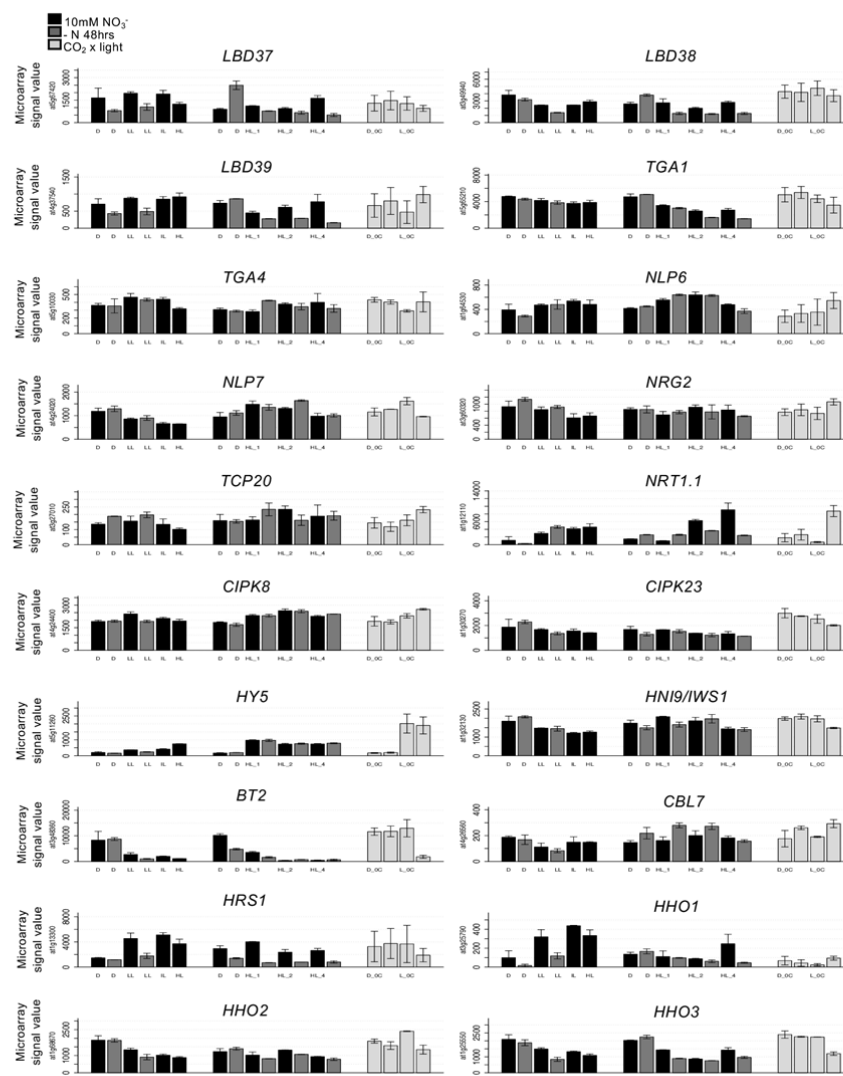


Figure 4. Most of the genes previously determined as *NRT2s* regulators do not display expression patterns similar to the patterns of the 3 candidate transcription factors in the set of Nitrogen and Light/Carbon combinations.

Graphs display the expression pattern of the 16 genes extracted from the whole transcriptomic dataset. Data are organized according to the multi-analysis (*i.e.*, S1 to S5, Figure 2). LBD37, LBD38, LBD39 represent the expression of genes involved in NO_3^- uptake (*NRT2.1* and *NRT2.5*) and assimilation, likely mimicking the effects of N organic compounds (21). TGA1, TGA4, NLP6, NLP7, NRG2, NRT1.1, CIPK8, CIPK23 are required for the NO_3^- -dependent induction of *NRT2.1* (22, 23, 25, 27, 28, 45, 64, 65). TCP20 and HNI9/IWS1 are involved into *NRT2.1* regulation controlled by systemic signaling (29, 66). BT2 represses expression of *NRT2.1* and *NRT2.4* genes under low NO_3^- conditions (26). CBL7 regulates *NRT2.4* and *NRT2.5* expression under N-starvation conditions (40). HY5 has been recently identified as a regulator of *NRT2.1* by mediating light promotion of NO_3^- uptake (30). HRS1, HHO1, HHO2 and HHO3 are repressors of *NRT2.4* and *NRT2.5* expression under high N conditions (41, 42).

284 treated with 5mM KCl for 2h. The data sets allowed the authors to determine the genes that
 285 respond specifically to NO_3^- in both WT and NR-null plants. The results show that, as
 286 expected, *NRT2.1*, *NRT2.2* and *NRT2.4* are induced by NO_3^- while *NRT2.5* seems to be
 287 repressed (Supplemental Figure 4A). In the same time, most of the known regulators for

288 *NRT2.1* are also induced by NO₃⁻ except *NLP7* and *TCP20*, two transcription factors which
289 have not been isolated using transcriptomic approaches (Supplemental Figure 4B). On the
290 contrary, in the same conditions, our three transcription factors, *TGA3*, *MYC1* and *bHLH093*
291 were not regulated by NO₃⁻ supply neither in WT nor NR-null plants. All these results
292 reinforced the originality of our experimental set up and explain why we found new
293 candidates that have never been isolated in previous transcriptomic experiments.

294

295 **Role of MYC1, TGA3 and bHLH093 in the regulation of NRT2s root nitrate** 296 **transporters**

297 To determine if MYC1, TGA3 and bHLH093 are involved in regulation of *NRT2* root NO₃⁻
298 transporters we used two independent insertion mutants for each of the transcription factors:
299 *tga3.2*, *tga3.3* for TGA3, *myc1.2*, *myc1.3* for MYC1 and *bHLH093.1*, *bHLH093.5* for
300 bHLH093. As both *TGA3* and *MYC1* were found to be regulated by N starvation, we also
301 produced a double mutant, *tga3.2/myc1.2*, to test a potential additive effect of those
302 transcription factors on the regulation of *NRT2s*. In addition, to reinforce our conclusions
303 concerning the role of bHLH093, we also produced an overexpressing line by transforming
304 the *bhlh093.1* mutant with a *35S::bHLH093* construct. The measurement of *MYC1*, *TGA3* and
305 *bHLH093* expression level confirmed an almost complete absence of their transcripts in their
306 respective mutants and a strong overexpression of *bHLH093* in the overexpressing line
307 (Supplemental Figure 5A and B).

308 As expected for a role of TGA3 and MYC1 in the regulation of *NRT2s* by N starvation, the
309 induction of both *NRT2.4* and *NRT2.5* is overall reduced in *tga3* and *myc1* mutants compared
310 to wild type plants, especially after 72h of N starvation (Figure 5A). This lower induction in
311 response to N starvation is stronger in the double mutant *tga3.2/myc1.2* and is observed in
312 that case consistently after 24h, 48h and 72h of N starvation for *NRT2.4* and *NRT2.5* and after
313 48h for *NRT2.2*. It suggests that TGA3 and MYC1 are not redundant and that both factors
314 may function as transcriptional activators under low N conditions. This result is supported by
315 the fact that neither the level of expression nor the regulation of *MYC1* in *tga3* mutants and of
316 *TGA3* in *myc1* mutants are affected compared to wild type plants (Supplemental Figure 5A).
317 However, surprisingly, MYC1 and TGA3 do not affect the regulation of *NRT2.1* in the same
318 conditions (Figure 5A). In agreement with a role of MYC1 and TGA3 in the regulation of
319 *NRT2.4* and *NRT2.5*, Y1H experiments show that both transcription factors are able to bind to
320 the promoter of these two transporters (Figure 5B).

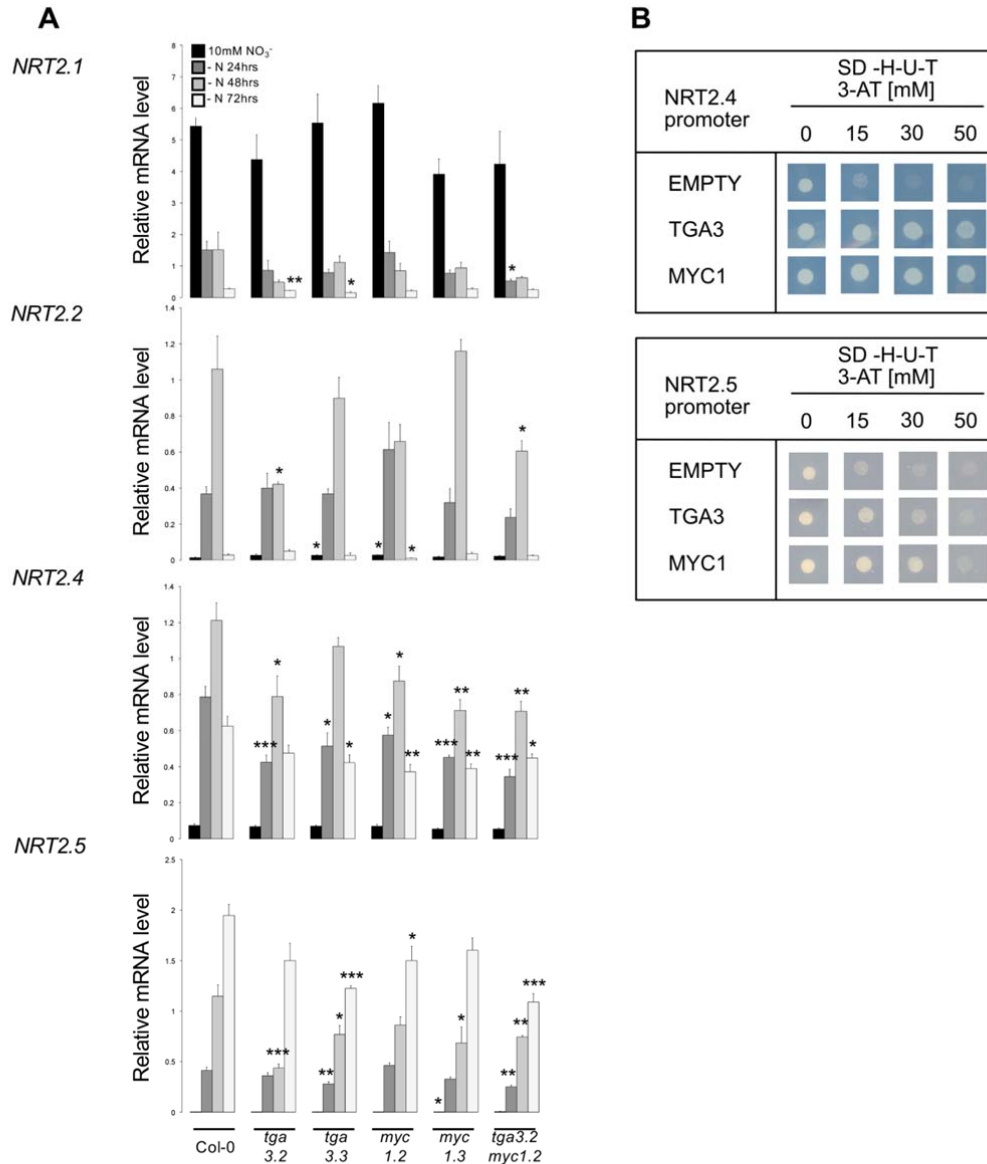


Figure 5. TGA3 and MYC1 are required for *NRT2.4* and *NRT2.5* full induction during N-deprivation.

A. Characterization of the knock-out mutants for TGA3 (*tga3.2* and *tga3.3*), MYC1 (*myc1.2* and *myc1.3*) and the TGA3/MYC1 double mutants (*tga3.2 myc1.2*). The plants were supplied with NO_3^- 10 mM one week ahead the experiment and acclimated for 24 hr in high light conditions ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$) before applying the N deprivation for 24, 48 or 72 hr. Roots have been collected to assess *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* mRNA accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Values are means of three biological replicates \pm SD. Differences between WT and the KO mutants are significant at * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ (Student's *t* test).

B. Characterization of TGA3 and MYC1 interaction with *NRT2.4* and *NRT2.5* promoters in a Y1H assay. Yeast cells were grown on SD-H-U-T minimal media without histidine (H), uracil (U), tryptophan (T) and containing 3- amino-1,2,4-triazole (3AT) at 0, 15, 30 and 50 mM. Interaction between the transcription factors and the promoters results in HIS3 reporter activation in contrast to the empty vector that does not interact.

321 Out of the three *NRT2s*, which are induced by light, *NRT2.4* and to a lower extent *NRT2.1*,
 322 have a significant lower induction after 4h and 8h of light in the *bHLH093* mutants as
 323 compared to wild type plants (Figure 6A and 6B). Conversely, the induction by light of both
 324 *NRT2.1* and *NRT2.4* is higher in the *35S::bHLH093* plants (Figure 6B). Interestingly, this

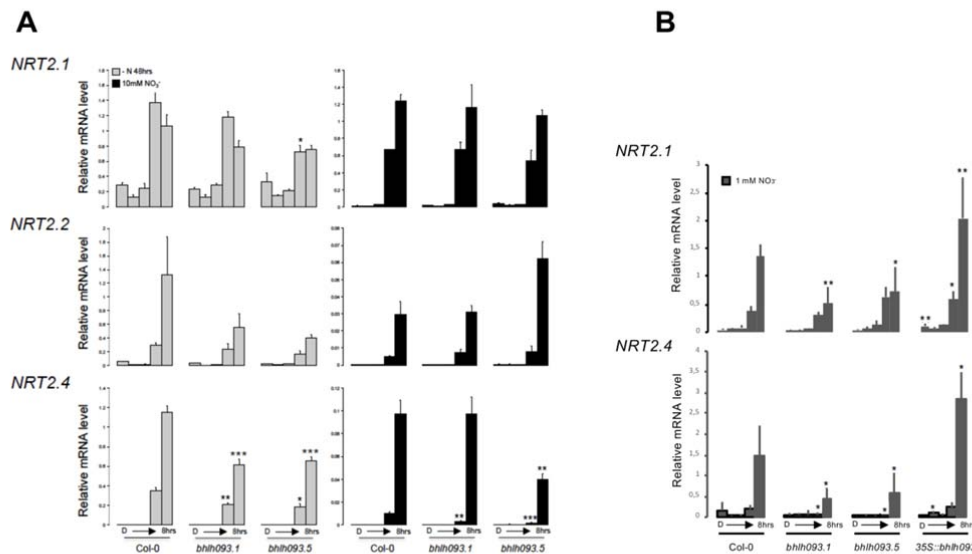


Figure 6. *bHLH093* is required for *NRT2.4* full induction by light in N-deprivation condition.

A. Characterization of the knock-out mutants for *bHLH093* (*bHLH093-1* and *bHLH093-5*) on 0N or 10mM NO_3^- . The plants were either starved for N for 48 hr (light gray bars) or supplied with NO_3^- 10 mM one week ahead the experiment (black bars) and were kept in the dark 40 hr before transition to high light intensity ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$) during 1h, 2h, 4h and 8h. Roots have been collected to assess *NRT2.1*, *NRT2.2* and *NRT2.4* mRNA accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Values are means of three biological replicates \pm SD. Differences between WT (Col-0) and the KO mutants are significant at $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ (Student's *t* test).

B. Characterization of the knock-out (*bHLH093-1* and *bHLH093-5*) and the over-expressor (*35S::bHLH093*) mutants for *bHLH093* on 1mM NO_3^- . The plants were grown on 1mM NO_3^- and were kept in the dark 40 hr before transition to intermediate light intensity ($250 \mu\text{mol m}^{-2} \text{s}^{-1}$; IL) during 1h, 2h, 4h and 8h. Roots have been collected to assess *NRT2.1* and *NRT2.4* mRNA accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Values are means of three biological replicates \pm SD. Differences between WT (Col-0) and the mutants are significant at $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ (Student's *t* test).

325 phenotype seems to depend on the amount of NO_3^- in the nutritive solution since the effect of
 326 *bHLH093* is preferentially seen when plants are starved for N or on 1mM NO_3^- and is almost
 327 absent when plants are grown on 10mM NO_3^- (Figure 6A and 6B).

328

329

330

331

332

333

334 **Discussion**

335 **Interaction between nitrogen and light provision affect regulation of *NRT2.1* expression**

336 As part of its central physiological role, the root NO_3^- HATS is a main target of the C/N
337 regulatory networks ensuring the necessary integration of both, N acquisition by roots and C
338 acquisition by shoots. The HATS regulation by N starvation has been well characterised in
339 previous studies, especially through the study of *NRT2.1* expression. Split-root experiments
340 have demonstrated that this regulation relies on systemic signaling pathways (Gansel et al.,
341 2001), and underlying molecular mechanisms have recently been unraveled (Ohkubo et al.,
342 2017). On the other hand, *NRT2.1* expression is also dramatically induced by light and sugars
343 through an Oxidative Pentose Phosphate Pathway (OPPP)-dependent signaling mechanism
344 (Lejay et al., 1999; Lejay et al., 2003; Lejay et al., 2008; de Jong et al., 2014). Over the past
345 decade, the importance of signal interaction for the regulation of gene expression has become
346 more and more obvious and especially for C/N regulation (Palenchar et al., 2004; Gutierrez et
347 al., 2007; Krouk et al., 2009). However, the details of how this interaction affects regulation
348 of *NRT2.1* expression in response to combined N/C treatments were unknown. Our results
349 clearly show that the interplay of N and C signaling mechanisms has a major role as light
350 conditions can totally suppress N regulation of *NRT2.1* expression, and vice versa (Figure 1A
351 and 1B). Similar to the case for inorganic N assimilation, it seems that low sugars inhibit
352 *NRT2.1* expression, overriding signals from N metabolism (Stitt et al., 2002; Nunes-Nesi et
353 al., 2010). Surprisingly, the regulation of *NRT2.1* by N starvation is not only abolished when
354 plants are treated in the dark. It happens also under high light conditions (Figure 1A).
355 However, in that case, the level of *NRT2.1* expression is always high, even on normally
356 repressive conditions like 10 mM NO_3^- , while in the dark the level of *NRT2.1* stays low,
357 independently of the level of N. One model to explain these results is that enhancement of
358 growth due to combination of high light and high NO_3^- supply results in a sustained high N
359 demand for growth, relieving the feedback repression normally associated with high NO_3^-
360 supply. This model is supported by a recent metabolomics analysis performed on *Arabidopsis*
361 *thaliana* under diverse C and N nutrient conditions (Sato and Yanagisawa, 2014). Taken
362 together, these results clearly support the idea that the control of *NRT2.1* expression involves
363 a complex network of interactions between signals emanating from N and C metabolisms.
364 However, this level of complexity seems to be rather specific for *NRT2.1*. In contrast to
365 *NRT2.1*, expression of *NRT2.2*, *NRT2.4* and *NRT2.5* is always repressed on 10 mM NO_3^- ,
366 independent of light levels (Supplemental Figure 1A). It should be noted that in the N

367 starvation experiments plants are transferred on a media with no N. This leads to the variation
368 of two factors, the N status of the plants, which decreases when plants are starved for N, and
369 the presence of NO_3^- in the nutritive solution, which is suppressed by the transfer to N-free
370 solution. Concerning the regulation of *NRT2.2*, *NRT2.4* and *NRT2.5* it is not known which
371 one of these two factors is predominant since their expression was only measured in N
372 starvation experiments (Kiba et al., 2012; Lezhneva et al., 2014; Kotur and Glass, 2015). It is
373 thus possible that *NRT2.2*, *NRT2.4* and *NRT2.5* are only regulated locally by NO_3^- and not by
374 systemic signals of N demand. This idea is supported by the work of (Ma et al., 2015)
375 showing that the regulation of *NRT2.4* and *NRT2.5* by N starvation depends on CBL7, which
376 is specifically induced by NO_3^- deficiency. Moreover, NIGT/HRS1s have been shown to act
377 as transcriptional repressor of *NRT2.4* and *NRT2.5* upon NO_3^- treatment (Kiba et al.,
378 2018)Safi et al. 2018). Local regulation by NO_3^- would explain why these transporters, unlike
379 *NRT2.1*, are always repressed when plants are on 10 mM NO_3^- , regardless of the light
380 conditions (Supplemental Figure 1A).

381

382 **Identification of three new candidates for regulation of *NRT2* genes using a systems** 383 **biology approach**

384 Over the past few years, transcriptomic approach and systems biology have been powerful
385 tools to identify new regulatory elements involved in N signaling (For review (Medici and
386 Krouk, 2014; Vidal et al., 2015). For root *NRT2s* genes and HATS activity in *Arabidopsis*, it
387 enabled the identification of CIPK23 and CIPK8 in response to NO_3^- , LBDs transcription
388 factors in response to high N and BT2, a negative regulator of *NRT2.1* and *NRT2.4* under low
389 N conditions (Figure 7) (Ho et al., 2009; Hu et al., 2009; Rubin et al., 2009; Araus et al.,
390 2016). For C and N signaling, previous microarray studies in response to transient treatments
391 with NO_3^- , sucrose or NO_3^- plus sucrose have been used to reveal, at the level of the genome,
392 the existence of interaction between C and N signaling (Wang et al., 2003; Price et al., 2004;
393 Scheible et al., 2004; Wang et al., 2004; Gutierrez et al., 2007; Huang et al., 2016). In
394 *Arabidopsis*, over 300 genes have been found differentially expressed by combined C:N
395 treatments compared to C or N treatments (Palenchar et al., 2004). However, because of the
396 number of genes affected by C and/or N regulation and the complex interactions between the
397 signalling pathways, none of these studies have led so far to the identification and the
398 validation of new regulatory elements. The unexpected regulations of root *NRT2s* and
399 especially of *NRT2.1* in our experimental set-up offer an interesting opportunity to find genes
400 more specifically involved in the regulation of root NO_3^- transporters by C and/or N, and to

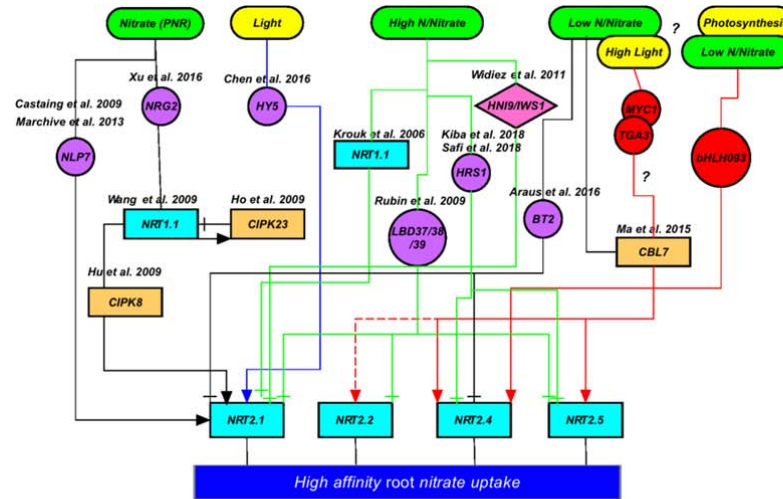


Figure 7. Schematic representation of the known regulatory elements for the regulation of root high-affinity NO_3^- transporters in response to external NO_3^- , the N status of the plant and light/photosynthesis.

Purple circles represent the transcription factors identified in previous studies while red circles represent the transcription factors identified in our study.

401 build a gene network model integrating regulators responding to N and/or C signals. As
 402 compared to previous transcriptomic approaches on N and C signaling in plants, we were able
 403 to narrow down the number of candidate genes by (i) using *NRT2.1* as a specific target and
 404 (ii) integrating the data from several Affymetrix microarrays to find gene networks co-

405 regulated with the expression of *NRT2.1* in response to different combinations of light and N
406 treatments.

407 Therefore, the gene regulatory network includes only three transcription factors, *bHLH093*,
408 *MYC1* and *TGA3* (Figure 2B). *bHLH093* was found co-regulated with *NRT2.1* in response to
409 light through photosynthesis because, like *NRT2.1*, it is not induced by light in the absence of
410 CO₂ (Supplemental Figure 3). *MYC1* and *TGA3* were found co-regulated with *NRT2.1* in
411 response to N starvation. The analysis of their level of expression across all the experiments
412 revealed that *TGA3* and *MYC1* are induced by N starvation but especially in LL and HL
413 conditions, while *bHLH093* seems overall induced by light no matter what the level of N
414 (Figure 3A). Furthermore, *MYC1* is also clearly induced by light (Figure 3A and
415 Supplemental Figure 3). Taken together these results support the validity of our approach to
416 find regulatory elements affected by C and/or N signalling and which are thus candidates for
417 the regulation by C/N interaction. Interestingly, none of these three transcription factors was
418 found involved in the regulation of root NO₃⁻ transporters by previous studies. One
419 explanation to this relates to the fact that the expression of *bHLH093*, *MYC1* and *TGA3* is not
420 responsive to the induction by NO₃⁻, which was by far the major environmental change
421 investigated by previous studies (Supplemental Figure 4A). Conversely, none of the
422 regulatory genes identified in previous studies was found with our approach. Indeed, most of
423 them are not affected by N starvation and/or by light (Figure 4). The only exception is *HY5*,
424 which encodes a recently identified mobile transcription factor involved in the regulation of
425 *NRT2.1* by sugar signals (Chen et al., 2016) and that is not found co-regulated with *NRT2.1* in
426 our analysis. This is explained by the fact that, unlike *NRT2.1*, we found *HY5* induced by light
427 even in the absence of CO₂ in our dataset (Figure 4). It indicates that expression of *HY5* does
428 not depend of the production of sugars through photosynthesis and is directly regulated by
429 light. The role of *HY5* in light signalling and not in C signalling is supported by previous
430 studies showing that *HY5* works downstream phytochrome signalling (Quail, 2002; Li et al.,
431 2010). Taken together, these results suggest that *NRT2.1* would be induced by both a light
432 component dependent on *HY5* and a C component dependent on the OPPP (Lejay et al.,
433 2008; de Jong et al., 2014; Chen et al., 2016). Accordingly, both Lejay et al. (2008) and Chen
434 et al. (2016) found that induction of *NRT2.1* by light is higher as compared to the addition of
435 sucrose in the dark. Furthermore, there is still an induction of *NRT2.1* expression by
436 increasing supply of sucrose in the mutant *hy5* (Chen et al., 2016).

437 ***bHLH093*, *MYC1* and *TGA3*, three transcription factors involved in regulation of**
438 ***NRT2.4* and *NRT2.5* gene expression**

439 The use of mutants validated our approach and showed that bHLH093 has mainly a role in the
440 induction by light of *NRT2.4*, while MYC1 and TGA3 affect induction by N starvation of
441 both *NRT2.4* and *NRT2.5* and in a more modest way *NRT2.2* (Figure 5A and Figure 6).
442 Furthermore, Y1H experiments support the fact that MYC1 and TGA3 are direct regulators of
443 *NRT2.4* and *NRT2.5* as already suggested for TGA3 and *NRT2.4* by the results obtained by
444 O'Malley et al. (2016) (Figure 3 and Figure 5B). Conversely, Chromatin
445 Immunoprecipitation (ChIP) experiments, using plants expressing bHLH093 fused to GFP,
446 failed to reveal a robust interaction with the promoter of *NRT2.4* (data not shown). It suggests
447 that bHLH093 is an indirect regulator and that it is rather involved in the signalling pathway
448 governing the regulation of *NRT2.4* and in a lesser extend *NRT2.1* by photosynthesis.
449 As represented in Figure 7, most of the regulatory elements identified to date concern the
450 primary NO₃⁻ response (PNR), with only three elements involved in the repression by high N
451 or high NO₃⁻ and one in the induction by light. Along with CBL7, MYC1 and TGA3 seem
452 thus to be part of an independent signalling pathway involved in the induction of root NO₃⁻
453 transporters in response to low N, while bHLH093 is, to our knowledge, the first element
454 involved in a regulatory mechanism linked to photosynthesis (Ma et al., 2015). As discussed
455 above, the role of these transcription factors in the regulatory mechanisms involved in C/N
456 interactions is also supported by our results. Indeed the role of bHLH093 in the regulation by
457 light seems to be dependent of the level of N and the role of MYC1 and TGA3 seems to be
458 stronger in high light conditions (Figure 5A and Figure 6).
459 However, surprisingly, none of these 3 transcription factors affect strongly the regulation of
460 *NRT2.1*, that we used as a target gene in our systems biology approach. This result could
461 indicate that the regulatory mechanisms differ between the four *NRT2* genes involved in the
462 HATS. Indeed, *NRT2.1* is regulated by at least 4 different mechanisms (local induction by
463 NO₃⁻ and repression by high NO₃⁻, systemic repression by N metabolites and induction by C),
464 while *NRT2.4* is regulated by C and N starvation and *NRT2.5* only by N starvation.
465 Furthermore, our experimental setup revealed obvious complex interactions between N and C
466 signalling for *NRT2.1*, which do not exist for *NRT2.2*, *NRT2.4* and *NRT2.5*. As discussed
467 above, if *NRT2.2*, *NRT2.4* and *NRT2.5* are only repressed by NO₃⁻ and not by N metabolites,
468 MYC1 and TGA3 could be involved in a NO₃⁻-specific signalling pathway upregulating the
469 very high-affinity transporters (*NRT2.4* and *NRT2.5*) when the external NO₃⁻ concentration
470 becomes too low to be efficiently taken up by *NRT2.1*. Previous results support a role, for at
471 least TGA3, in a NO₃⁻-specific signalling pathway. Indeed, TGA3 is part of a family of 7
472 genes in *Arabidopsis thaliana* and two of them, TGA1 and TGA4, have already been

473 involved in the induction of *NRT2.1* and *NRT2.2* in response to NO_3^- (Alvarez et al., 2014).
474 Taken together these results and our findings suggest that this family of transcription factors
475 could participate in a more general way to the regulation of root NO_3^- transporters by NO_3^- .
476 Concerning *MYC1* there is no direct evidence to support its role in a NO_3^- signalling pathway
477 (Bruex et al., 2012).
478 Since *NRT2.1* and *NRT2.4* are both regulated by C through OPPP, it was even more
479 surprising to find that the absence of *bHLH093* affects mainly the induction by light of
480 *NRT2.4* compared to *NRT2.1* (Figure 6) (Lejay et al., 2008). However, the role of *bHLH093*
481 seems to be dependent on the level of N since it plays a significant role in the regulation of
482 *NRT2.4* only under low N conditions, whereas the induction of *NRT2.1* by light is mostly seen
483 in this experiment under high N conditions (10mM NO_3^-). It could explain why, in those
484 conditions, *bHLH093* mutation does not affect the regulation of *NRT2.1* by light, while in the
485 second experiment, where plants were grown on a moderate level of NO_3^- (1mM), *NRT2.1* is
486 well induced by light and the NO_3^- concentration could be low enough to reveal the impact of
487 *bHLH093* on *NRT2.1* regulation (Figure 6B). To our knowledge, the role of *bHLH093* in the
488 roots and in response to light has never been characterised before. The only information
489 concerns a role in flowering promotion under non-inductive short-day conditions through the
490 gibberellin pathway (Sharma et al., 2016).

491

492 **Materials and Methods**

493 **Plant Material**

494 *Arabidopsis thaliana* genotypes used in this study were the wild-type Col-0 ecotype and
495 mutants obtained from the Salk Institute: *tga3.2* (Salk_081158), *tga3.3* (Salk_088114),
496 *myc1.2* (Salk_057388), *myc1.3* (Salk_006354), *bHLH093.1* (Salk_121082) and *bHLH093.5*
497 (Salk_104582).

498 In all experiments plants were grown hydroponically under non sterile conditions as described
499 by Lejay et al. (1999). Briefly, the seeds were germinated directly on top of modified
500 Eppendorf tubes filled with pre-wetted sand. The tubes were then positioned on floating rafts
501 and transferred to tap water in a growth chamber under the following environmental
502 conditions: light/dark cycle of 8 h/16 h, light intensity of $250 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, temperature of
503 $22/20^\circ\text{C}$, and RH of 70%. After 1 week, the tap water was replaced with a complete nutrient
504 solution. The experiments were performed on plants grown on 1 mM NO_3^- as N source. The
505 other nutrients were added as described by Lejay et al. (1999). The plants were allowed to

506 grow for 3 additional weeks before the experiments. Nutrient solutions were renewed weekly
507 and on the day before the experiments.

508

509 **Treatments**

510 Two different sets of experiments were performed to (i) study the impact of light on the
511 regulation of NO_3^- transporter genes in the roots by N starvation, and (ii) study the impact of
512 the N status of the plants on the regulation of these genes by light.

513 In the first set of experiments 4 weeks old plants were transferred on a solution containing 10
514 mM NO_3^- . After one week the plants were transferred in the morning either in continuous
515 dark or in a light/dark cycle at three different light intensities (50, 250 and 800 $\mu\text{moles}\cdot\text{h}^{-1}\cdot\text{m}^{-2}$)
516 and starved for N during 24h, 48h and 72h, by replacing NO_3^- with CaCl_2 2.5 mM and
517 K_2SO_4 2.5 mM.

518 In the second set of experiments 4 weeks old plants were transferred on a solution containing
519 10 mM NO_3^- . They were then pre-treated during 3 days on nutrient solution containing
520 contrasted level of N: (i) no N, (ii) 1 mM NO_3^- or (iii) 10 mM NO_3^- . After 32h in the dark the
521 plants were transferred to light for 1h, 2h, 4h and 8h under three different light intensities (50,
522 250 and 800 $\mu\text{moles}\cdot\text{h}^{-1}\cdot\text{m}^{-2}$).

523 The dependence of the expression of NO_3^- transporter genes on photosynthesis was
524 investigated by modifying the CO_2 concentration in the atmosphere. After a pretreatment of
525 40 h in the dark, plants grown on 1mM NO_3^- were placed for 4 h in the light ($\square 150 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$)
526 or in the dark in a 240-L, airtight plexiglass chamber connected to a computerized
527 device for controlling temperature, humidity, and CO_2 concentration in the atmosphere
528 (Atelliance Instruments; see Delhon et al. (1996) for details). The CO_2 concentration in the
529 atmosphere was held constant during the treatments at 0 or 600 $\mu\text{L L}^{-1}$.

530 All experiments were repeated two or three times.

531

532 **RNA Extraction and Gene Expression Analysis**

533 Root samples were frozen in liquid N_2 in 2-mL tubes containing one steel bead (2.5 mm
534 diameter). Tissues were disrupted for 1 min at 30 s^{-1} in a Retsch mixer mill MM301
535 homogenizer (Retsch, Haan, Germany). Total RNA was extracted from tissues using TRIzol
536 reagent (Invitrogen, Carlsbad, CA, USA). Subsequently 4 μg of RNA were treated with
537 DNase (DNase I, SIGMA-ALDRICH, USA) following the manufacturer's instructions.
538 Reverse transcription was achieved in the presence of Moloney murine leukemia virus reverse
539 transcriptase (Promega, Madison, WI, USA) after annealing with an anchored oligo(dT)₁₈

540 primer as described by Wirth et al. (2007). The quality of the cDNA was verified by PCR
541 using specific primers spanning an intron in the gene *APTR* (At1g27450) forward 5'-
542 CGCTTCTTCTCGACACTGAG-3'; reverse 5'-CAGGTAGCTTCTTGGGCTTC-3'.
543 Gene expression was determined by quantitative real-time PCR (LightCycler; Roche
544 Diagnostics, Basel, Switzerland) with the kit LightCycler FastStart DNA Master SYBR Green
545 I (Roche Diagnostics, Basel, Switzerland) according to the manufacturer's instructions with 1
546 μ l of cDNA in a total volume of 10 μ l. The amplifications were performed as described
547 previously by Wirth et al. (2007). All the results presented were standardized using the
548 housekeeping gene Clathrin (At4g24550). Gene-specific primer sequences were: NRT2.1
549 forward, 5'-ACAAGGGCTAACGTGGATG-3'; NRT2.1 reverse, 5'-
550 CTGCTTCTCCTGCTCATTCC-3'; NRT2.2 forward, 5'-GCAGCAGATTGGCATGCATTT-
551 3'; NRT2.2 reverse, 5'-AAGCATTGTTGGTTGCGTTCC-3'; NRT2.4 forward, 5'-
552 GAACAAGGGCTGACATGGAT -3'; NRT2.4 reverse, 5'- GCTTCTCGGTCTCTGTCCAC
553 -3'; NRT2.5 forward, 5'-TGTGGACCCTCTTCCAAAAA-3'; NRT2.5 reverse, 5'-
554 TTTGGGGATGAGTCGTTGTGG-3'; MYC1 forward, 5'-AACCTTAACGACTCTGTG-3';
555 MYC1 reverse, 5'-CCGCAACTATGTAGTCTCTG-3'; TGA3 forward, 5'-
556 CTCTCAGAAAGTGTGGC-3'; TGA3 reverse, 5'-CATATACGAGGAGATGAGTG-3';
557 bHLH093 forward, 5'-AGCTTGAAGGCCAACC-3'; bHLH093 reverse, 5'-
558 GCTCTTTCATGTAATCTATGGCA-3'; Clathrin forward, 5'-
559 AGCATACTGCGTGCAAAG-3'; Clathrin reverse, 5'-TCGCCTGTGTCACATATCTC-
560 3'.

561

562 **Acquisition of Genome-Wide Expression and Statistical Analysis**

563 Genome-wide expression was determined using Affymetrix ATH1 GeneChip expression
564 microarrays according to manufacturer's instructions. To do so, biotinylated cRNA was
565 synthesized from 200 ng of total RNA from *Arabidopsis* roots. Affymetrix data were
566 normalized in R (<http://www.r-project.org/>) using MAS5.

567 Then, normalized data were subjected to different statistical analyses, all centered on *NRT2.1*
568 expression pattern but including various sets of microarray data among the whole data set. As
569 a first approach to build a gene network involved in the regulation of root NO₃⁻ transporters,
570 we examined genes displaying expression pattern correlated to *NRT2.1* expression pattern
571 across the entire dataset. A R² coefficient cut-off above 0.8 or below -0.8 led to the
572 identification of 79 AGIs displaying an expression pattern correlated to *NRT2.1*, including 77
573 genes positively correlated with *NRT2.1* (Table S1). Among these 79 genes, none of them

574 displays a function related to gene regulation but rather related to metabolic activity and more
575 precisely to carboxylic acid metabolic process as, for example, the *Glutamate synthase 2* gene
576 (Supplemental Figure 6A). Moreover, a hierarchical clustering of the treatments according to
577 the expression pattern of these genes clearly revealed that their response is largely driven only
578 by the light/carbon factor, putting aside any possible regulation by N provision (Supplemental
579 Figure 6B). Therefore, we determined that a global analysis of the entire data set was not
580 relevant to identify regulators of NO_3^- transport integrating C and N availability and that a
581 finest analysis of gene expression in different subsets of treatments will be more powerful.
582 The list of genes regulated by N-deprivation specifically under low light regime was
583 determined by a t.test analysis ($p.\text{value}<0.05$) between conditions 3 and 4. All genes also
584 found regulated between conditions 1 and 2 based on the same analysis are removed from this
585 list (Figure 1, Table S1). Genes regulated by N-deprivation during light induction are
586 determined by a 2 ways ANOVA using Nitrogen as one factor (presence = conditions
587 7,9,11,13 / absence = conditions 8,10,12,14) and Light as the second factor (no Light =
588 conditions 7,8 / 1hr-light = conditions 9,10 / 2hr-light = conditions 11,12 / 4hr-light =
589 conditions 13,14). Genes of interest are regulated by the interaction of the 2 factors
590 ($p.\text{value}<0.05$) and display a similar regulation by N from dark to 2hr-light as observed for
591 *NRT2.1* (Figure 1, Table S2). Genes regulated by light intensity under high N-provision and
592 by light time exposure under high N-provision are both determined by a linear modeling of
593 gene expression across light intensity (conditions 1,3,5,6) or time exposure (conditions
594 7,9,11,13) using a R^2 above 0.9 ($p.\text{value}$ is below 0.003) (Figure 1, Tables S3 and S4).
595 Finally, genes regulated by photosynthesis activity are determined by a 2 ways ANOVA
596 using CO_2 level as one factor (0ppm = conditions 15,17 / 600ppm = conditions 16,18) and
597 Light as the second factor (Dark = conditions 15,16 / Light = conditions 17,18). To narrow
598 down the list of *NRT2.1*-like genes, only those passing post-hoc Tukey tests comparing
599 conditions 18 to all 3 others ($p.\text{value}<0.05$) and displaying a ratio >2 or <0.5 are selected
600 (Figure 1, Table S5).

601

602 **Visualization of gene connectivity by clustering and gene network analysis**

603 Heat map hierarchical cluster of gene expression and samples was generated with the MeV
604 software using Pearson correlation as distance metric and Average as linkage method
605 (www.tm4.org) (Saeed et al., 2003). The Gene Network was generated with the VirtualPlant
606 1.3 software (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>) (Katari et al., 2010). The
607 connectivity of the nodes is based on 5 categories corresponding to literature data, post-

608 transcriptional regulation, protein:protein interactions, transcriptional regulation and regulated
609 edges meaning transcription factor - target relationship based at least on one binding site in
610 the promoter of the target gene. Two nodes are linked by an edge if they fall in any of these
611 categories combined to an expression pattern correlated at a $R^2 > 0.7$ or < -0.7 . Visualization of
612 the gene regulatory network has been performed with Cytoscape (<http://www.cytoscape.org/>)
613 (Shannon et al., 2003). Node properties have been modified to reveal connectivity with the 3
614 transcription factors and highlight *NRT2.1* position within the network.

615

616 **Y1H Assays**

617 For the generations of the plasmids for promoter analysis by Y1H, particular promoter
618 fragments of NRT2.4 (1968bp), NRT2.5 (1692bp) were first amplified by PCR with
619 overlapping ends as described by Gibson et al. (2009). For the bait, the pMW2 and pMW3
620 vectors were used (Deplancke et al., 2006). pMW vectors were amplified by PCR with
621 overlapping ends as a single sequence (pMW2) or as 2 independent sections (pMW2). Final
622 vectors were made as described by Gibson et al., 2009. The Y1H prey vectors for TGA3 and
623 MYC1 transcriptions factors were a kind gift from Franziska Turck (Castrillo et al., 2011).
624 All the fragments generated for all constructs were validated by DNA sequencing.

625 The Y1H assay was performed according to protocol described by Grefen (2014) with minor
626 modifications. Briefly, the vectors pMW2-NRT2.4, pMW3-NRT2.4, pMW2-NRT2.5,
627 pMW3-NRT2.5 were first linearized with restriction enzymes. For pMW2 vectors BamH1
628 (NEB) was used and for pMW3 vectors Xho1 (NEB). The resulting linearized constructs
629 were subsequently co-integrated into the yeast strain: YM4271 as described by Grefen (2014).
630 The transformed yeast strains were tested for autoactivation and the selected colonies with the
631 higher sensitivity to 3-AT were then transformed with the construct pDEST-AD-TGA3 or
632 pDEST-AD-MYC1 or pDEST-AD (Empty vector). Empty vector was included as a negative
633 control. Resulting yeast were dropped on selection media (SD -His-Ura-Trp) supplemented
634 with increasing concentrations of 3-AT (0, 15, 30, 50, 80, 100 mM). Yeast growth was
635 verified after 48h.

636

637 **Acknowledgments**

638

639 We thank members of the lab in France and Chile for discussion.

640

641

642 **Figure legends**

643

644 **Figure 1.** Interaction between Nitrogen and Light/Carbon provision modulates *NRT2.1*
645 mRNA accumulation in roots. A, Different light regimes modulate *NRT2* regulation in roots
646 of plants experiencing from high NO_3^- provision (10 mM) to N deprivation (-N). The light
647 regimes encompass dark, low light intensity ($50 \mu\text{mol m}^{-2} \text{s}^{-1}$; LL), intermediate light intensity
648 ($250 \mu\text{mol m}^{-2} \text{s}^{-1}$; IL) and high light intensity ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$; HL). Plants were supplied
649 with NO_3^- 10 mM one week ahead the experiment and acclimated for 24 hours in the different
650 light regimes before applying the N deprivation for 24, 48 or 72 hr. B, Different N provisions
651 modulate *NRT2* regulation in roots of plants experiencing a dark to light transition. The N
652 provisions encompass 10mM NO_3^- , 1mM NO_3^- (for 72 hr) and N deprivation for 48 hr (-N).
653 Plants are kept in the dark (*i.e.*, 40hr) before transition to high light intensity ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$;
654 HL) and roots are collected at time 0 (Dark) and 1, 2, 4 and 8 hr after light transition. C,
655 Regulation of *NRT2* by photosynthesis activity. Plants are grown in regular NO_3^- regime
656 (1mM) and intermediate light intensity until they are transferred for 4 hr in a CO_2 -deprived
657 atmosphere (0ppm) or in high CO_2 -supplied atmosphere (600ppm), either in the dark or in the
658 light. In these 3 experimental conditions, roots have been collected to assess *NRT2.1* mRNA
659 accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene).
660 Expression pattern of *NRT2.1* across the 35 conditions tested (16 in A, 15 in B and 4 in C) has
661 driven the choice of 18 conditions to investigate gene reprogramming associated to the
662 regulation of NO_3^- transport. These 18 conditions are indicated with arrows and numbers on
663 the x-axis of the 3 *NRT2.1* bar graphs (Each arrow corresponds to one condition with 2
664 independent biological repeats constituted of a pool of approx. 10 plants each).

665

666 **Figure 2.** Gene expression multi-analysis driven by *NRT2.1* expression pattern combined to
667 an integrative analysis identified a candidate gene regulatory network connected to the NO_3^-
668 transport system. A, Venn diagrams identifying common genes regulated by N provision on
669 low light condition and dark to light transition (34 genes) or regulated by light/carbon (142
670 genes). The union of these gene lists defines a population of 174 genes, including 4
671 transcription factors. B, The core set of 174 genes differentially expressed has been structured
672 into a Gene Regulatory Network using the Gene Networks analysis tool in VirtualPlant
673 software (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>) (Katari et al., 2010). The network
674 includes 124 nodes (genes) and 260 edges connecting genes. The nodes have been organized
675 according to their connection to the 3 transcription factors *MYC1*, *TGA3* and *bHLH093* and

676 are detailed in the Network Legend. *ARR14* is excluded from the network due to its lack of
677 connectivity to other nodes according to the edges selected to generate the network.

678

679 **Figure 3.** *TGA3*, *MYC1* and *bHLH093* are candidate transcription factors for the control of
680 the expression of *NRT2* gene family. A, Gene expression analysis of the 3 candidate
681 transcription factors in the extended set of Nitrogen/Carbon combinations confirms
682 correlation with *NRT2.1* regulation. Expression patterns have been determined by RT-QPCR
683 (relative accumulation to *Clathrin* housekeeping gene). B, *NRT2.2*, *NRT2.4* and *NRT2.5* as
684 well as *NRT2.1* display putative cis-binding elements for the 3 transcription factors in their
685 promoter region. The gene network has been done using the Gene Networks analysis tool in
686 VirtualPlant software (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>) (Katari et al., 2010);
687 only Regulated Edges box and One Binding Site option has been selected in this case. C,
688 *TGA3* bounds *in silico* with the promoter of *NRT2.1*, *NRT2.2* and *NRT2.4*. The analysis has
689 been done using the Plant Cistrome Database (<http://neomorph.salk.edu/PlantCistromeDB>)
690 (O'Malley et al., 2016).

691

692 **Figure 4.** Most of the genes previously determined as *NRT2s* regulators do not display
693 expression patterns similar to the patterns of the 3 candidate transcription factors in the set of
694 Nitrogen and Light/Carbon combinations. Graphs display the expression pattern of the 20
695 genes extracted from the whole transcriptomic dataset. Data are organized according to the
696 multi-analysis (*i.e.*, S1 to S5, Figure 2). LBD37, LBD38, LBD39 repress the expression of
697 genes involved in NO_3^- uptake (*NRT2.1* and *NRT2.5*) and assimilation, likely mimicking the
698 effects of N organic compounds (Rubin et al., 2009). *TGA1*, *TGA4*, *NLP6*, *NLP7*, *NRG2*,
699 *NRT1.1*, *CIPK8*, *CIPK23* are required for the NO_3^- -dependent induction of *NRT2.1* (Munos et
700 al., 2004; Castaings et al., 2009; Ho et al., 2009; Hu et al., 2009; Konishi and Yanagisawa,
701 2013; Marchive et al., 2013; Alvarez et al., 2014; Xu et al., 2016). *TCP20* and *HNI9/IWS1*
702 are involved into *NRT2.1* regulation controlled by systemic signaling (Widiez et al., 2011;
703 Guan et al., 2014). *BT2* represses expression of *NRT2.1* and *NRT2.4* genes under low NO_3^-
704 conditions (Araus et al., 2016). *CBL7* regulates *NRT2.4* and *NRT2.5* expression under N-
705 starvation conditions (Ma et al., 2015). *HY5* has been recently identified as a regulator of
706 *NRT2.1* by mediating light promotion of NO_3^- uptake (Chen et al., 2016). *HRS1*, *HHO1*,
707 *HHO2* and *HHO3* are repressors of *NRT2.4* and *NRT2.5* expression under high N conditions
708 (Kiba et al., 2018)Safi et al. 2018)

709

710 **Figure 5.** TGA3 and MYC1 are required for *NRT2.4* and *NRT2.5* full induction during N-
711 deprivation. A, Characterization of the knock-out mutants for TGA3 (*tga3.2* and *tga3.3*),
712 MYC1 (*myc1.2* and *myc1.3*) and the TGA3/MYC1 double mutants (*tga3.2 myc1.2*). The
713 plants were supplied with NO_3^- 10 mM one week ahead the experiment and acclimated for 24
714 hr in high light conditions ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$) before applying the N deprivation for 24, 48 or
715 72 hr. Roots have been collected to assess *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* mRNA
716 accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Values
717 are means of three biological replicates \pm SD. Differences between WT and the KO mutants
718 are significant at $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ (Student's *t* test). B, Characterization
719 of TGA3 and MYC1 interaction with *NRT2.4* and *NRT2.5* promoters in a Y1H assay. Yeast
720 cells were grown on SD-H-U-T minimal media without histidine (H), uracil (U), tryptophan
721 (T) and containing 3- amino-1,2,4-triazole (3AT) at 0, 15, 30 and 50 mM. Interaction between
722 the transcription factors and the promoters results in HIS3 reporter activation in contrast to
723 the empty vector that does not interact.

724

725 **Figure 6.** *bHLH093* is required for *NRT2.4* full induction by light in N-deprivation condition.
726 A, Characterization of the knock-out mutants for *bHLH093* (*bHLH093-1* and *bHLH093-5*) on
727 0N or 10mM NO_3^- . The plants were either starved for N for 48 hr (light gray bars) or supplied
728 with NO_3^- 10 mM one week ahead the experiment (black bars) and were kept in the dark 40 hr
729 before transition to high light intensity ($800 \mu\text{mol m}^{-2} \text{s}^{-1}$) during 1h, 2h, 4h and 8h. Roots
730 have been collected to assess *NRT2.1*, *NRT2.2* and *NRT2.4* mRNA accumulation by RT-
731 QPCR (relative accumulation to *Clathrin* housekeeping gene). Values are means of three
732 biological replicates \pm SD. Differences between WT (Col-0) and the KO mutants are
733 significant at $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ (Student's *t* test). B, Characterization of
734 the knock-out (*bHLH093-1* and *bHLH093-5*) and the over-expressor (*35S::bHLH093*)
735 mutants for *bHLH093* on 1mM NO_3^- . The plants were grown on 1mM NO_3^- and were kept in
736 the dark 40 hr before transition to intermediate light intensity ($250 \mu\text{mol m}^{-2} \text{s}^{-1}$; IL) during
737 1h, 2h, 4h and 8h. Roots have been collected to assess *NRT2.1* and *NRT2.4* mRNA
738 accumulation by RT-QPCR (relative accumulation to *Clathrin* housekeeping gene). Values
739 are means of three biological replicates \pm SD. Differences between WT (Col-0) and the
740 mutants are significant at $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ (Student's *t* test).

741

742 **Figure 7.** Schematic representation of the known regulatory elements for the regulation of
743 root high-affinity NO_3^- transporters in response to external NO_3^- , the N status of the plant and

744 light/photosynthesis. Purple circles represent the transcription factors identified in previous
745 studies while red circles represent the transcription factors identified in our study.

746

747 **Supplemental Figure 1.** Interaction between Nitrogen and Light/Carbon provision modulates
748 mRNA accumulation in roots of most of the *NRT2* family members.

749

750 **Supplemental Figure 2.** Expression pattern of *NRT2* family genes in the set of Nitrogen and
751 Carbon/Light combinations as determined by Arabidopsis Affymetrix ATH1 microarray
752 hybridization.

753

754 **Supplemental Figure 3.** Expression pattern of *TGA3*, *MYC1* and *bHLH093* transcription
755 factors in the set of Nitrogen and Carbon/Light combination as determined by *Arabidopsis*
756 Affymetrix ATH1 microarrays hybridization.

757

758 **Supplemental Figure 4.** *TGA3*, *MYC1* and *bHLH093* display expression pattern different
759 than most of the known regulators of *NRT2* genes in response to NO_3^- .

760

761 **Supplemental Figure 5.** Expression pattern of *TGA3*, *MYC1* and *bHLH093*.

762

763 **Supplemental Figure 6.** Biomaps and hierarchical clustering of the 79 most correlated genes
764 to *NRT2.1* expression across all experiments.

765

766 **Supplemental Table 1.** List of 430 probes regulated by N deprivation under low light regime
767 only.

768

769 **Supplemental Table 2.** List of 573 probes regulated by the interaction between nitrogen and
770 light.

771

772 **Supplemental Table 3.** List of 128 probes linearly regulated by light intensity.

773

774 **Supplemental Table 4.** List of 985 probes linearly regulated during light induction.

775

776 **Supplemental Table 5.** List of 509 probes regulated by the interaction between light and
777 CO_2 .

778

779 **Supplemental Table 6.** List of 80 probes coregulated based on *NRT2.1* expression and
780 pearson correlation.

781

782

Parsed Citations

Alvarez JM, Riveras E, Vidal EA, Gras DE, Contreras-Lopez O, Tamayo KP, Aceituno F, Gomez I, Ruffel S, Lejay L, Jordana X, Gutierrez RA (2014) Systems approach identifies TGA1 and TGA4 transcription factors as important regulatory components of the nitrate response of Arabidopsis thaliana roots. Plant J. 80: 1-13

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Araus V, Vidal EA, Puelma T, Alamos S, Mieulet D, Guiderdoni E, Gutierrez RA (2016) Members of BTB Gene Family of Scaffold Proteins Suppress Nitrate Uptake and Nitrogen Use Efficiency. Plant Physiol. 171: 1523-1532

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Bruex A, Kainkaryam RM, Weckowski Y, Kang YH, Bernhardt C, Xia Y, Zheng X, Wang JY, Lee MM, Benfey P, Woolf PJ, Schiefelbein J (2012) A gene regulatory network for root epidermis cell differentiation in Arabidopsis. PLoS Genet. 8: e1002446

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Castaigns L, Camargo A, Pocholle D, Gaudon V, Texier Y, Boutet-Mercey S, Taconnat L, Renou JP, Daniel-Vedele F, Fernandez E, Meyer C, Krapp A (2009) The nodule inception-like protein 7 modulates nitrate sensing and metabolism in Arabidopsis. Plant J. 57: 426-435

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Castrillo G, Turck F, Leveugle M, Lecharyn A, Carbonero P, Coupland G, Paz-Ares J, Onate-Sanchez L (2011) Speeding cis-trans regulation discovery by phylogenomic analyses coupled with screenings of an arrayed library of Arabidopsis transcription factors. PLoS One 6: e21524

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Cerezo M, Tillard P, Filleur S, Munos S, Daniel-Vedele F, Gojon A (2001) Major alterations of the regulation of root NO₃⁻ uptake are associated with the mutation of Nrt2.1 and Nrt2.2 genes in Arabidopsis. Plant Physiol. 127: 262-271

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Chen X, Yao Q, Gao X, Jiang C, Harberd NP, Fu X (2016) Shoot-to-Root Mobile Transcription Factor HY5 Coordinates Plant Carbon and Nitrogen Acquisition. Curr. Biol. 26: 640-646

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Coruzzi GM, Zhou L (2001) Carbon and nitrogen sensing and signaling in plants: emerging "matrix effects". Curr. Opin. Plant Biol. 4: 247-253

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Crawford NM, Glass ADM (1998) Molecular and physiological aspects of nitrate uptake in plants. Trends Plant Sci. 3: 389-395

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

de Jong F, Thodey K, Lejay LV, Bevan MW (2014) Glucose elevates NITRATE TRANSPORTER2.1 protein levels and nitrate transport activity independently of its HEXOKINASE1-mediated stimulation of NITRATE TRANSPORTER2.1 expression. Plant Physiol. 164: 308-320

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Delhon P, Gojon A, Tillard P, Passama L (1996) Diurnal regulation of NO₃⁻ uptake in soybean plants IV. Dependence on current photosynthesis and sugar availability to the roots. J. Exp. Bot. 47: 893-900

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Deplancke B, Mukhopadhyay A, Ao W, Elewa AM, Grove CA, Martinez NJ, Sequerra R, Doucette-Stamm L, Reece-Hoyes JS, Hope IA, Tissenbaum HA, Mango SE, Walhout AJ (2006) A gene-centered C. elegans protein-DNA interaction network. Cell 125: 1193-1205

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Filleur S, Dorbe MF, Cerezo M, Orsel M, Granier F, Gojon A, Daniel-Vedele F (2001) An Arabidopsis T-DNA mutant affected in Nrt2 genes is impaired in nitrate uptake. FEBS Lett. 489: 220-224

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Gansel X, Munos S, Tillard P, Gojon A (2001) Differential regulation of the NO₃⁻ and NH₄⁺ transporter genes AtNrt2.1 and AtAmt1.1 in Arabidopsis: relation with long-distance and local controls by N status of the plant. Plant J. 26: 143-155

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Garnett T, Conn V, Plett D, Conn S, Zanghellini J, Mackenzie N, Enju A, Francis K, Holtham L, Roessner U, Boughton B, Bacic A, Shirley N, Rafalski A, Dhugga K, Tester M, Kaiser BN (2013) The response of the maize nitrate transport system to nitrogen demand and supply across the lifecycle. *New Phytol.* 198: 82-94

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Gibson DG, Young L, Chuang RY, Venter JC, Hutchison CA, 3rd, Smith HO (2009) Enzymatic assembly of DNA molecules up to several hundred kilobases. *Nat. Methods* 6: 343-345

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Grefen G (2014) The Split-Ubiquitin System for the Analysis of Three-Component Interactions. *Arabidopsis Protocols*, Humana Press, Totowa, NJ: 659-678

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Guan P, Ripoll JJ, Wang R, Vuong L, Bailey-Steinitz LJ, Ye D, Crawford NM (2017) Interacting TCP and NLP transcription factors control plant responses to nitrate availability. *Proc Natl Acad Sci U S A* 114: 2419-2424

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Guan P, Wang R, Nacry P, Breton G, Kay SA, Pruneda-Paz JL, Davani A, Crawford NM (2014) Nitrate foraging by Arabidopsis roots is mediated by the transcription factor TCP20 through the systemic signaling pathway. *Proceedings of the National Academy of Sciences of the United States of America* 111: 15267-15272

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Gutierrez RA, Lejay LV, Dean A, Chiaromonte F, Shasha DE, Coruzzi GM (2007) Qualitative network models and genome-wide expression data define carbon/nitrogen-responsive molecular machines in Arabidopsis. *Genome Biol.* 8: R7

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Gutierrez RA, Stokes TL, Thum K, Xu X, Obertello M, Katari MS, Tanurdzic M, Dean A, Nero DC, McClung CR, Coruzzi GM (2008) Systems approach identifies an organic nitrogen-responsive gene network that is regulated by the master clock control gene CCA1. *Proc Natl Acad Sci U S A* 105: 4939-4944

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Ho CH, Lin SH, Hu HC, Tsay YF (2009) CHL1 functions as a nitrate sensor in plants. *Cell* 138: 1184-1194

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Hu HC, Wang YY, Tsay YF (2009) AtCIPK8, a CBL-interacting protein kinase, regulates the low-affinity phase of the primary nitrate response. *Plant J.* 57: 264-278

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Huang A, Sang Y, Sun W, Fu Y, Yang Z (2016) Transcriptomic Analysis of Responses to Imbalanced Carbon: Nitrogen Availabilities in Rice Seedlings. *PLoS One* 11: e0165732

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Katari MS, Nowicki SD, Aceituno FF, Nero D, Kelfer J, Thompson LP, Cabello JM, Davidson RS, Goldberg AP, Shasha DE, Coruzzi GM, Gutierrez RA (2010) VirtualPlant: a software platform to support systems biology research. *Plant Physiol.* 152: 500-515

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Kiba T, Feria-Bourrellier AB, Lafouge F, Lezhneva L, Boutet-Mercey S, Orsel M, Brehaut V, Miller A, Daniel-Vedele F, Sakakibara H, Krapp A (2012) The Arabidopsis nitrate transporter NRT2.4 plays a double role in roots and shoots of nitrogen-starved plants. *Plant Cell* 24: 245-258

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Kiba T, Inaba J, Kudo T, Ueda N, Konishi M, Mitsuda N, Takiguchi Y, Kondou Y, Yoshizumi T, Ohme-Takagi M, Matsui M, Yano K, Yanagisawa S, Sakakibara H (2018) Repression of Nitrogen Starvation Responses by Members of the Arabidopsis GARP-Type Transcription Factor NIGT1/HRS1 Subfamily. *Plant Cell* 30: 925-945

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Konishi M, Yanagisawa S (2013) Arabidopsis NIN-like transcription factors have a central role in nitrate signalling. *Nat Commun* 4: 1617

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Kotur Z, Glass AD (2015) A 150 kDa plasma membrane complex of AtNRT2.5 and AtNAR2.1 is the major contributor to constitutive high-affinity nitrate influx in Arabidopsis thaliana. Plant Cell Environ 38: 1490-1502

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Kotur Z, Mackenzie N, Ramesh S, Tyerman SD, Kaiser BN, Glass AD (2012) Nitrate transport capacity of the Arabidopsis thaliana NRT2 family members and their interactions with AtNAR2.1. New Phytol. 194: 724-731

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Krouk G, Tranchina D, Lejay L, Cruikshank AA, Shasha D, Coruzzi GM, Gutierrez RA (2009) A systems approach uncovers restrictions for signal interactions regulating genome-wide responses to nutritional cues in Arabidopsis. PLoS Comput Biol 5: e1000326

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Lejay L, Gansel X, Cerezo M, Tillard P, Muller C, Krapp A, von Wiren N, Daniel-Vedele F, Gojon A (2003) Regulation of root ion transporters by photosynthesis: functional importance and relation with hexokinase. The Plant Cell 15: 2218-2232

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Lejay L, Tillard P, Lepetit M, Olive F, Filleur S, Daniel-Vedele F, Gojon A (1999) Molecular and functional regulation of two NO₃- uptake systems by N- and C-status of Arabidopsis plants. Plant J. 18: 509-519

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Lejay L, Wirth J, Pervent M, Cross JM, Tillard P, Gojon A (2008) Oxidative pentose phosphate pathway-dependent sugar sensing as a mechanism for regulation of root ion transporters by photosynthesis. Plant Physiol. 146: 2036-2053

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Lezhneva L, Kiba T, Feria-Bourrellier AB, Lafouge F, Boutet-Mercey S, Zoufan P, Sakakibara H, Daniel-Vedele F, Krapp A (2014) The Arabidopsis nitrate transporter NRT2.5 plays a role in nitrate acquisition and remobilization in nitrogen-starved plants. Plant J. 80: 230-241

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Li J, Li G, Gao S, Martinez C, He G, Zhou Z, Huang X, Lee JH, Zhang H, Shen Y, Wang H, Deng XW (2010) Arabidopsis transcription factor ELONGATED HYPOCOTYL5 plays a role in the feedback regulation of phytochrome A signaling. Plant Cell 22: 3634-3649

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Li W, Wang Y, Okamoto M, Crawford NM, Siddiqi MY, Glass AD (2007) Dissection of the AtNRT2.1:AtNRT2.2 inducible high-affinity nitrate transporter gene cluster. Plant Physiol. 143: 425-433

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Ma Q, Tang RJ, Zheng XJ, Wang SM, Luan S (2015) The calcium sensor CBL7 modulates plant responses to low nitrate in Arabidopsis. Biochem. Biophys. Res. Commun. 468: 59-65

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Malagoli P, Laine P, Le Deunff E, Rossato L, Ney B, Ourry A (2004) Modeling nitrogen uptake in oilseed rape cv Capitol during a growth cycle using influx kinetics of root nitrate transport systems and field experimental data. Plant Physiol. 134: 388-400

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Marche C, Roudier F, Castaigns L, Brehaut V, Blondet E, Colot V, Meyer C, Krapp A (2013) Nuclear retention of the transcription factor NLP7 orchestrates the early response to nitrate in plants. Nature Communications 4: 1713

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Medici A, Krouk G (2014) The primary nitrate response: a multifaceted signalling pathway. J. Exp. Bot. 65: 5567-5576

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Munos S, Cazes C, Fizames C, Gaymard F, Tillard P, Lepetit M, Lejay L, Gojon A (2004) Transcript profiling in the chl1-5 mutant of Arabidopsis reveals a role of the nitrate transporter NRT1.1 in the regulation of another nitrate transporter, NRT2.1. Plant Cell 16: 2433-2447

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Nacry P, Bouguyon E, Gojon A (2013) Nitrogen acquisition by roots: physiological and developmental mechanisms ensuring plant adaptation to a fluctuating resource. Plant Soil 370: 1-29

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Nunes-Nesi A, Fernie AR, Stitt M (2010) Metabolic and signaling aspects underpinning the regulation of plant carbon nitrogen interactions. Mol. Plant 3: 973-996

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

O'Brien JA, Vega A, Bouguyon E, Krouk G, Gojon A, Coruzzi G, Gutierrez RA (2016) Nitrate Transport, Sensing, and Responses in Plants. Mol. Plant 9: 837-856

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

O'Malley RC, Huang SS, Song L, Lewsey MG, Bartlett A, Nery JR, Galli M, Gallavotti A, Ecker JR (2016) Cistrome and Epicistrome Features Shape the Regulatory DNA Landscape. Cell 165: 1280-1292

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Ohkubo Y, Tanaka M, Tabata R, Ogawa-Ohnishi M, Matsubayashi Y (2017) Shoot-to-root mobile polypeptides involved in systemic regulation of nitrogen acquisition. Nature Plants 3: 17029

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Palenchar PM, Kouranov A, Lejay LV, Coruzzi GM (2004) Genome-wide patterns of carbon and nitrogen regulation of gene expression validate the combined carbon and nitrogen (CN)-signaling hypothesis in plants. Genome Biol 5: R91

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Price J, Laxmi A, St Martin SK, Jang JC (2004) Global transcription profiling reveals multiple sugar signal transduction mechanisms in Arabidopsis. Plant Cell 16: 2128-2150

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Quail PH (2002) Photosensory perception and signalling in plant cells: new paradigms? Curr. Opin. Cell Biol. 14: 180-188

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Rubin G, Tohge T, Matsuda F, Saito K, Scheible WR (2009) Members of the LBD family of transcription factors repress anthocyanin synthesis and affect additional nitrogen responses in Arabidopsis. Plant Cell 21: 3567-3584

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Ruffel S, Gojon A, Lejay L (2014) Signal interactions in the regulation of root nitrate uptake. J. Exp. Bot. 65: 5509-5517

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Saeed AI, Sharov V, White J, Li J, Liang W, Bhagabati N, Braisted J, Klapa M, Currier T, Thiagarajan M, Sturn A, Snuffin M, Rezantsev A, Popov D, Ryltsov A, Kostukovich E, Borisovsky I, Liu Z, Vinsavich A, Trush V, Quackenbush J (2003) TM4: a free, open-source system for microarray data management and analysis. BioTechniques 34: 374-378

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Sato S, Yanagisawa S (2014) Characterization of metabolic states of Arabidopsis thaliana under diverse carbon and nitrogen nutrient conditions via targeted metabolomic analysis. Plant Cell Physiol. 55: 306-319

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Schachtman DP, Shin R (2007) Nutrient sensing and signaling: NPKS. Annu. Rev. Plant Biol. 58: 47-69

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Scheible WR, Morcuende R, Czechowski T, Fritz C, Osuna D, Palacios-Rojas N, Schindelasch D, Thimm O, Udvardi MK, Stitt M (2004) Genome-wide reprogramming of primary and secondary metabolism, protein synthesis, cellular growth processes, and the regulatory infrastructure of Arabidopsis in response to nitrogen. Plant Physiol. 136: 2483-2499

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T (2003) Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 13: 2498-2504

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Sharma N, Xin R, Kim DH, Sung S, Lange T, Huq E (2016) NO FLOWERING IN SHORT DAY (NFL) is a bHLH transcription factor that promotes flowering specifically under short-day conditions in Arabidopsis. Development 143: 682-690

Pubmed: [Author and Title](#)

Google Scholar: [Author Only Title Only Author and Title](#)

Stitt M, Muller C, Matt P, Gibon Y, Carillo P, Morcuende R, Scheible WR, Krapp A (2002) Steps towards an integrated view of nitrogen metabolism. J. Exp. Bot. 53: 959-970

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Vidal EA, Alvarez JM, Moyano TC, Gutierrez RA (2015) Transcriptional networks in the nitrate response of Arabidopsis thaliana. Curr. Opin. Plant Biol. 27: 125-132

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Wang R, Okamoto M, Xing X, Crawford NM (2003) Microarray analysis of the nitrate response in Arabidopsis roots and shoots reveals over 1,000 rapidly responding genes and new linkages to glucose, trehalose-6-phosphate, iron, and sulfate metabolism. Plant Physiol. 132: 556-567

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Wang R, Tischner R, Gutierrez RA, Hoffman M, Xing X, Chen M, Coruzzi G, Crawford NM (2004) Genomic analysis of the nitrate response using a nitrate reductase-null mutant of Arabidopsis. Plant Physiol. 136: 2512-2522

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Widiez T, El Kafafi el S, Girin T, Berr A, Ruffel S, Krouk G, Vayssières A, Shen WH, Coruzzi GM, Gojon A, Lepetit M (2011) High nitrogen insensitive 9 (HNI9)-mediated systemic repression of root NO₃⁻ uptake is associated with changes in histone methylation. Proceedings of the National Academy of Sciences of the United States of America 108: 13329-13334

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Wirth J, Chopin F, Santoni V, Viennois G, Tillard P, Krapp A, Lejay L, Daniel-Vedele F, Gojon A (2007) Regulation of root nitrate uptake at the NRT2.1 protein level in Arabidopsis thaliana. J. Biol. Chem. 282: 23541-23552

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)

Xu N, Wang R, Zhao L, Zhang C, Li Z, Lei Z, Liu F, Guan P, Chu Z, Crawford NM, Wang Y (2016) The Arabidopsis NRG2 Protein Mediates Nitrate Signaling and Interacts with and Regulates Key Nitrate Regulators. The Plant Cell 28: 485-504

Pubmed: [Author and Title](#)

Google Scholar: [Author Only](#) [Title Only](#) [Author and Title](#)