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# Accounting for Linkage Disequilibrium in genome scans for selection without individual genotypes: the local score approach

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- 1 Motivations
- 2 The FLK & hapFLK approaches
- 3 The local score approach
- 4 Simulation results
- 5 Examples
- 6 Conclusions

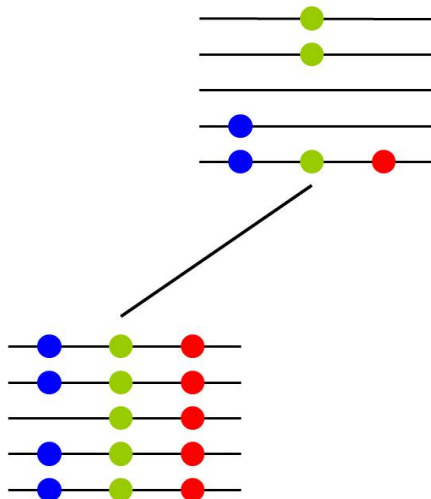
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# Genome scans for selection

- Most genomic regions are neutral, but some of them are (or have been) under selection (natural or artificial).
- Detecting the regions under selection is important for theory (evolution) and applications (medicine, agronomy).
- Genome wide scans for selection now possible from dense genotyping (SNP chips) or sequencing (NGS) data.
- Focus on positive (adaptive) selection.

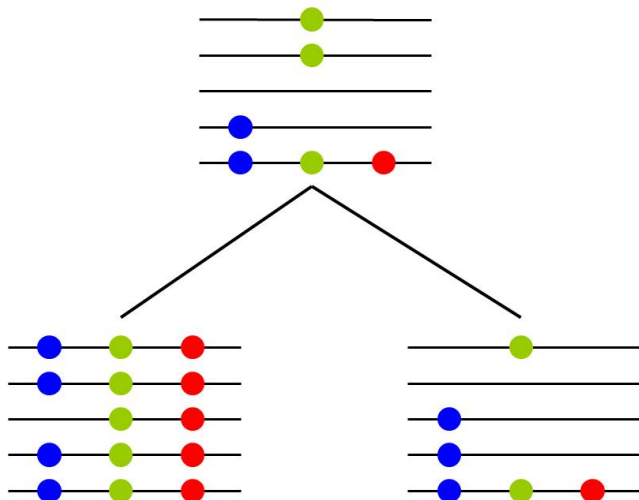
# Population differentiation approach

Look for markers with contrasted allele frequencies between populations.

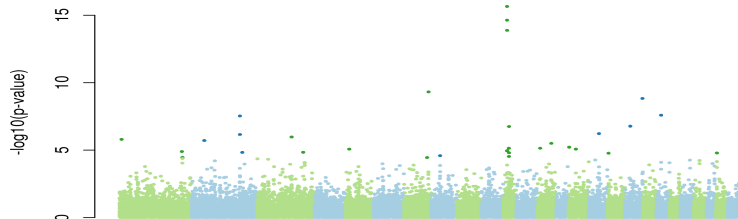


# Population differentiation approach

Look for markers with contrasted allele frequencies between populations.



# Linkage Disequilibrium (LD) helps!



- **Single-marker statistics** have a **large variance**, high values can be reached just by chance due to drift.
- Due to LD, markers in the **neighborhood of a selected locus** also show **elevated differentiation** between populations.

→ Account for LD in selection scans by:

- 1 using haplotype tests
- 2 looking for clusters of markers with high differentiation

- **Cut the genome into fixed windows** and computes a summary of the single-marker statistics within each window.
- **Summarize each window** using:
  - the average of single-marker statistics (Weir *et al*, 2005).
  - the number of markers exceeding a given threshold (Myles *et al*, 2008).
  - the number of markers differentially fixed between populations (Johansson *et al*, 2010).
- **Individual genotypes not required** (pooled sequencing).
- Limitations:
  - How to choose **window size**? the **single-marker threshold**?
  - How to decide that a **window** is **under selection**?

→ **Overcome these issues** using the statistical **local score** theory.

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$p = (p_1, \dots, p_i, \dots, p_n)$ : allele frequencies at one SNP in several populations.

$\bar{p}$  and  $s_p^2$ : observed mean and variance of  $p$ .

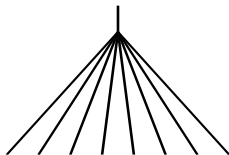
$$F_{ST} = \frac{s_p^2}{\bar{p}(1-\bar{p})}$$

- $H_0$  : “neutral evolution” (genetic drift)  
vs  $H_1$  : “positive selection in one (or more) population ”.
- $H_0$  rejected if  $F_{ST}$  too large.

## Lewontin et Krakauer (LK) test (1973)

$$LK = \frac{n-1}{\bar{F}_{ST}} F_{ST}$$

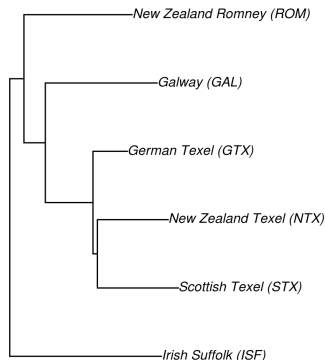
- **LK distribution under  $H_0$  is  $\chi^2$  with  $n - 1$  degrees of freedom.**
- But, only true if populations have a **star like phylogeny with equal population sizes.**



# FLK test (Bonhomme *et al*, 2010)

Extension of LK accounting for

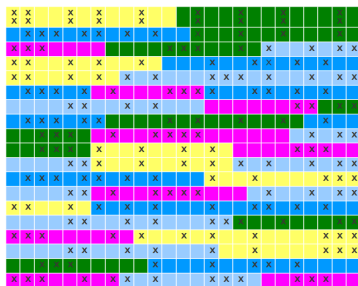
- differences in effective size between populations.
- differences in correlations between population pairs.



(first estimated from genome wide data)

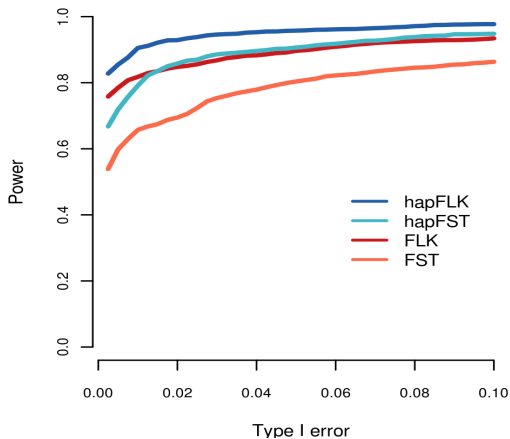
# hapFLK test (Fariello *et al*, 2013)

- **Define local haplotypes** around each SNP position using the model of Scheet and Stephens (2006).



- **Compute haplotype frequencies** in each population.
- Apply FLK, considering haplotypes as alleles.

# Detection power



4 populations with hierarchical structure, 1 under selection.

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- For each marker  $m$ , define the **score**:

$$X_m = -\log_{10}(p_m) - \xi$$

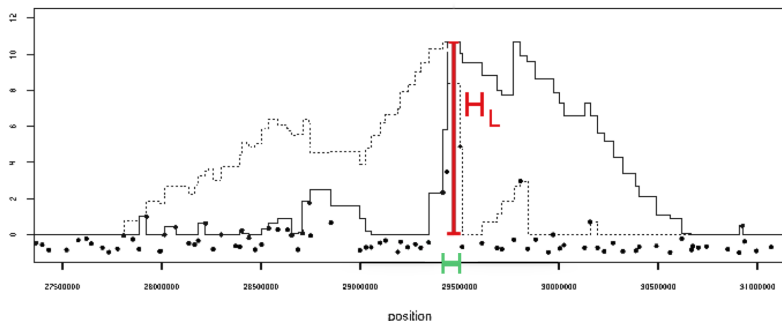
$p_m$  p-value of a test for selection,  $\epsilon$  fixed threshold.

- Low p-value =  $H_0$  (neutral evolution) unlikely = high score.
- **Cumulate scores** using the so-called **Lindley process**:

$$h_0 = 0, \quad h_m = \max(0, h_{m-1} + X_m)$$

- Look for **local maxima of the Lindley process**, which are associated to **genomic regions** that are **enriched in high scores / low p-values**.
- Here  $p_m$  is the p-value of FLK.

# Example



- The Lindley process (black line) has several **excursions above 0** (local maxima).
- The **global maximum** ( $H_L$ ) is called the **local score**.
- Each excursion is associated to an interval enriched in high scores (in green).

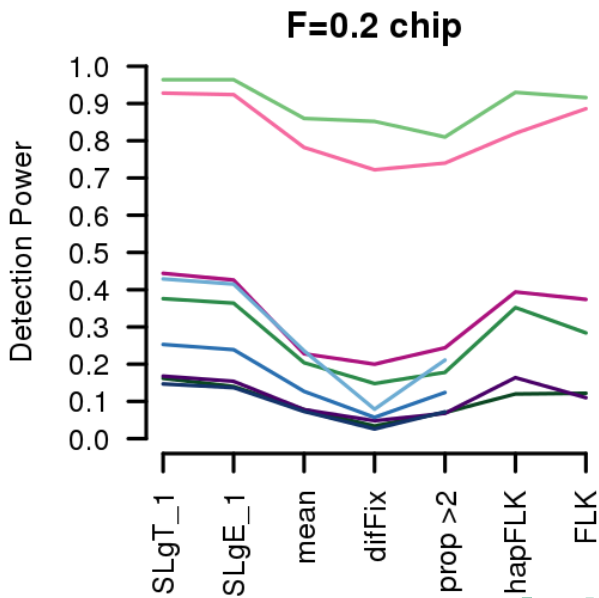
- **p-value threshold** in  $\log_{10}$  scale.
- Ex:  $\xi = 2$  cumulates p-values below  $10^{-2}$ .
- For **high**  $\xi$ , only **most significant markers** contribute:
  - similar to single point approach.
  - **strong selection**.
- For **low**  $\xi$ , more markers contribute:
  - **longer** intervals.
  - **recent selection**.

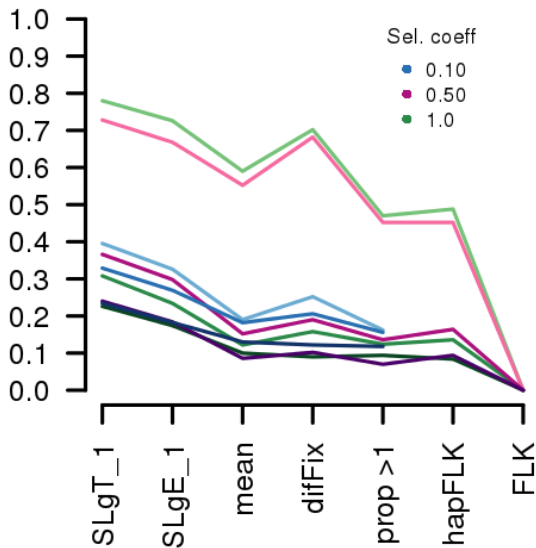
- How likely is a given excursion under neutrality?
- Depends on:
  - the number of markers in the sequence ( $M$ ).
  - the correlation between scores ( $\rho$ ).
- We provided **two approaches** allowing to **compute significance thresholds for excursions** :
  - 1 analytical formula:** valid if single-marker p-values are uniform under neutrality.
  - 2 re-sampling approach:** valid for all datasets, but requires some computing time.

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# Simulation procedure

- **Two populations** with same effective size, one neutral and **one under selection**.
- Genomic region of **10Mb** with **one selected site**.
- Several statistics compared, in different scenarios.
- **Detection threshold** of each statistic such that selection is detected in 5% of the **neutral samples** (type I error 5%).
- For the local score, also computed using our re-sampling approach  
→ observed type I error 6%.
- Tuning parameters (window size,  $\xi$  ...) chosen to optimize detection power.

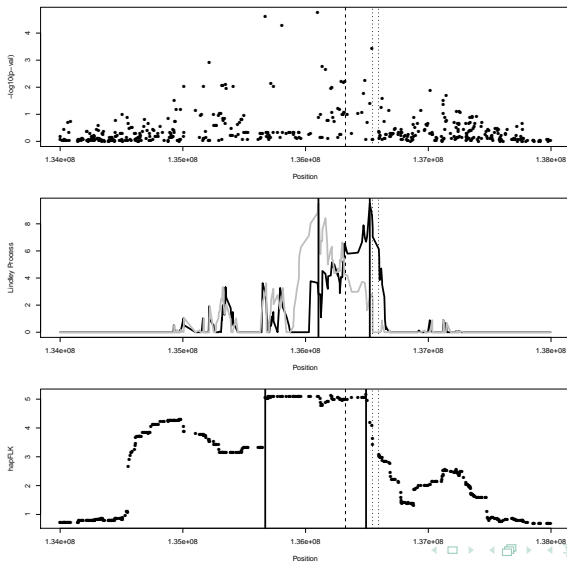


**F=0.4 sequence**

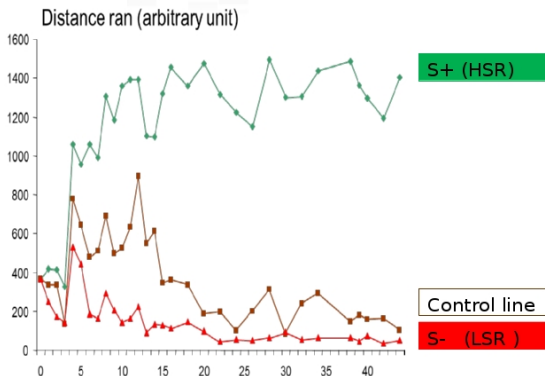
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# Lactase region in Humans

Test of selection based on HapMap genotypes (Europea and Asia).

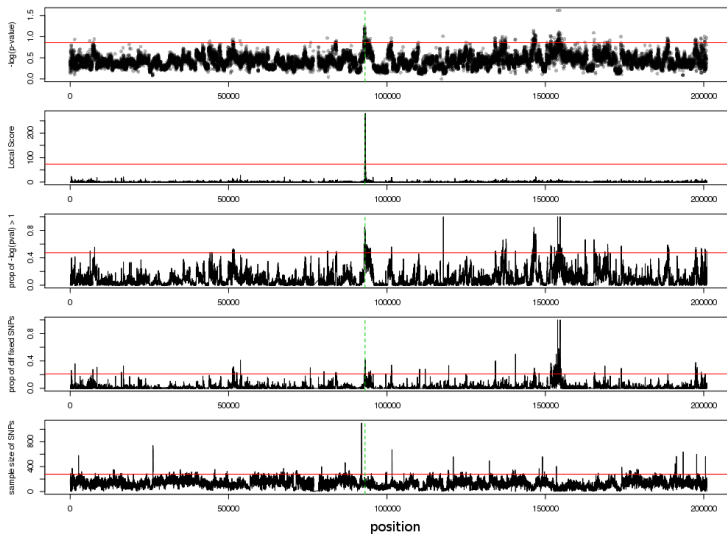


# Divergent selection experiment on behaviour in Quail



- Pooled DNA from each line sequenced at generation 50
- Strong drift ( $F = 0.4$ ).

# Selection scan on chromosome 1



## Significant regions genome-wide

Chr.	Position	L (kb)	Genes
1	92,963,481-93,182,440	219	NSUN3, <b>ARL13B</b>
2	1,584,033-1,688,400	104	VIPR1
3	61,586,217-61,604,464	19	ECHDC1, RNF146
3	75,088,250-75,170,494	82	MMS22L
4	11,412,372-11,452,609	40	GLOD5
4	90,953,044-91,008,245	56	<b>CTNNA2</b>
6	35,234,870-35,336,720	102	FOXI2, <b>PTPRE</b>
6	6,311,718-6,644,395	333	UBE2D1, CISD1, <b>IPMK</b>
10	17,825,157-17,825,227	0.07	
25	1,296,647-1,296,706	0.059	

Genes **in bold** have been associated to **autistic disorders** or **behavioral traits** in Humans.

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# Detecting selection using the local score

- Accounts for **LD without individual genotypes**.
- One single tuning parameter,  $\xi$ , with intuitive interpretation.  
 $\xi = 1$  recommended for detection power.
- **Statistical significance** of candidate regions easy to compute.
- **Increased detection power** compared to single-marker, window-based or haplotype-based tests.
- Convincing results on 2 real datasets with different features.
- Can be applied to **any single-marker test providing p-values**, for selection scans or **any other context**.
- Ref: Fariello *et al*, Molecular Ecology 2017.

## Quail husbandry and sampling:

- **Cécile Arnould & Christine Leterrier**, Unité de Physiologie de la Reproduction et des Comportements, INRA Tours
- **Julien Recoquillay**, Unité de Recherches Avicoles, INRA Tours
- **David Gourichon**, Pôle d'Expérimentation Avicole, INRA Tours

## Computing Facilities:

- Genotoul bioinformatics platform Toulouse Midi-Pyrénées.

## DNA preparation and sequencing:

- **Olivier Bouchez & Gérald Salin**, GeT-PlaGe Genotoul, INRA Toulouse
- **Sophie Leroux & Frédérique Pitel**, GenPhySE, INRA Toulouse

## Bioinformatic and statistic analyses:

- **Patrice Dehais**, SIGENAE, INRA Toulouse
- **David Robelin & Thomas Faraut**, GenPhySE, INRA Toulouse

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- Supervised by Lounès Chikhi (Evolution et Diversité Biologique) and Olivier Mazet (INSA).
- Influence of population structure on past population size estimation (Mazet *et al*, Heredity 2017).