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Hide and seek: hidden genetic variance contributing to the adaptive potential of selfing populations

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21 **Running title:** Partial self-fertilization and adaptation from standing genetic variation.

22

23

24 **Keywords:** self-fertilization, standing genetic variation, adaptation, cryptic genetic variation,
25 residual allogamy.

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40 **Abstract:** Standing genetic variation is considered a major contributor to the adaptive
41 potential of species. The low heritable genetic variation observed in self-fertilising
42 populations has led to the hypothesis that species with this particular mating system would be
43 less likely to adapt. However, a non-negligible amount of cryptic genetic variation for
44 polygenic traits, accumulated through negative linkage disequilibrium, could prove to be an
45 important source of standing variation in self-fertilising species. Using a classical quantitative
46 genetics model, we demonstrate that selfing populations are better able to store cryptic genetic
47 variance than outcrossing populations, notably due to their lower recombination rate.
48 Following a shift in the environment, this hidden diversity can be partially released,
49 increasing the additive variance and adaptive potential of selfing populations. In such
50 conditions, even though the process of adaptation itself is mating system dependant, selfers
51 reach levels of fitness that are equal to or higher than outcrossing populations within a few
52 generations. Outcrossing populations respond better to selection for the new optimum, but
53 they maintain more genetic diversity resulting in a higher genetic load. In selfing populations,
54 genetic diversity is remobilised, and new close-to-optimum genotypes are generated and
55 quickly increase in frequency, leading to more homogenous populations. Our results bring
56 new insights into the role of standing genetic variation for adaptation in selfing populations.

57

58 ***INTRODUCTION***

59

60 Natural populations harbour a significant amount of genetic variation, especially at
61 loci governing polygenic traits (Mittell *et al.* 2015; Wood *et al.* 2016; Clo *et al.* 2019). This
62 variation, known as standing genetic variation, has been considered to be an important
63 predictor for the adaptive potential of populations (Orr and Betancourt 2001; Hermisson and

64 Pennings 2005; Barrett and Schluter 2008; Pritchard *et al.* 2010; Glémin and Ronfort 2013;
65 Matuszewski *et al.* 2015). Indeed, standing variation represents an easily accessible, hence
66 non-negligible, source of genetic variation, available for adaptation to changing conditions
67 (Hermisson and Pennings 2005; Barrett and Schluter 2008). Contrary to adaptation from *de*
68 *novo* mutations, the probability to adapt from standing variation is higher simply because
69 beneficial mutations already segregating in a population are expected to be present at higher
70 frequencies (Innan and Kim 2004; Barrett and Schluter 2008). It has also been suggested that
71 populations adapting from standing genetic variation can cope with more severe and more
72 rapid environmental change, as they are able to cross larger distances in phenotype space
73 (Matuszewski *et al.* 2015). The amount of heritable variance is thus expected to play a key
74 role in adaptation, and any forces affecting it may greatly influence whether or not
75 populations are able to survive environmental changes.

76 An important characteristic of populations, known to greatly affect the amount of
77 genetic variance, is the mating system. From both theoretical (Charlesworth and Charlesworth
78 1995; Lande and Porcher 2015; Abu Awad and Roze 2018) and empirical works
79 (Charlesworth and Charlesworth 1995; Geber and Griffen 2003; Clo *et al.* 2019), it is known
80 that, compared to outcrossing populations, self-fertilization reduces, on average, the amount
81 of additive genetic variance for polygenic or quantitative traits under stabilizing selection.
82 This diminution is due to more efficient purifying selection under selfing and linkage
83 disequilibria maintained between alleles at different loci: the so-called Bulmer effect (Lande
84 and Porcher 2015; Abu Awad and Roze 2018). Because of the low genetic variance
85 maintained in self-fertilizing populations, this mating system has been qualified as an
86 evolutionary dead-end (Stebbins 1957; Takebayashi and Morrell 2001; Igic and Busch 2013).
87 However, theoretical and some empirical works are now pointing towards the existence of
88 cryptic genetic variability (see Paaby and Rockman 2014 for a review), which should, in

89 addition to the “visible” genetic variation, contribute to the adaptive potential of natural
90 populations. Cryptic genetic variation has been defined as a part of a population’s standing
91 genetic variation that does not affect phenotypic variation in a stable environment, but can
92 increase heritable variation in environmental conditions rarely experienced (Gibson and
93 Dworkin 2004; Paaby and Rockman 2014). In other words, it defines a source of variability
94 that is not expressed in stable conditions (due to conditional neutrality, the genetic structure of
95 the population, etc), but which can contribute to adaptation in new conditions. Such “hidden”
96 variability has been detected in both outcrossing (in sticklebacks (McGuigan *et al.* 2011),
97 cavefish (Rohner *et al.* 2013), dung flies (Berger *et al.* 2011), gulls (Kim *et al.* 2013) or
98 spadefoot toads (Ledon-Rettig *et al.* 2010)) and selfing species (*Caenorhabditis elegans*,
99 Milloz *et al.* 2008; *Arabidopsis thaliana*, Queitsch *et al.* 2002). Two main mechanisms could
100 explain the accumulation and the release of such variance: interactions between loci (Badano
101 and Katsanis 2002; Carter *et al.* 2005; Shao *et al.* 2008), and phenotypic plasticity (Anderson
102 *et al.* 2013). In this paper, we focus on interactions between loci.

103 To maintain the population as close as possible to the phenotypic optimum, stabilizing
104 selection disfavors apparent genetic and phenotypic diversity (Lande and Porcher 2015; Abu
105 Awad and Roze 2018). However, the structuration of the additive variance also strongly
106 depends on the trait mutation rate and the prevalence of pleiotropy (Lande and Porcher 2015;
107 Abu Awad and Roze 2018). When the per-trait mutation rate is weak, associations between
108 loci are negligible, but when the rate increases, the creation and maintenance of co-adapted
109 gene complexes structure the additive variance into positive within-loci components and
110 negative among-loci components, reducing the observed additive variance (Abu Awad and
111 Roze 2018). The remobilization of alleles contributing to this last component of variance
112 could boost the evolvability of populations forced to undergo directional selection after a
113 change in the phenotypic optimum (Le Rouzic and Carlborg 2008). Indeed, if associations

114 between loci are broken, segregating alleles could express some or all of their additive effects
115 in new-genetic backgrounds. Classical models analyzing the effect of selfing on adaptation
116 from standing genetic variation have considered a single locus (Glémin and Ronfort 2013),
117 thus neglecting interactions among loci that could result in other forms of standing genetic
118 variation. As self-fertilization reduces the effective recombination rate (Nordborg 2000),
119 allowing the maintenance of co-adapted gene complexes, the storage of hidden genetic
120 diversity should be stronger in selfing populations (as suggested in Lande and Porcher 2015;
121 Abu Awad and Roze 2018), potentially increasing their probability to adapt to an
122 environmental change beyond that expected from single-locus models.

123 In this paper, we explore this hypothesis, using a quantitative genetics framework. We
124 describe and quantify how, to what degree, and under which conditions populations
125 accumulate hidden genetic variation in this theoretical framework of polygenic traits. Though
126 these polygenic trait models are based on simple hypotheses, they have so far proven to be
127 surprisingly accurate in predicting the distribution of epistatic coefficients (Martin *et al.* 2007)
128 and inferring the mean dominance coefficient of mutations (Manna *et al.* 2011). We show
129 that, in models allowing for hidden genetic diversity and when adaptation is only possible
130 from pre-existing standing genetic variation, selfing populations are able to perform just as
131 well as their mixed-mating and outcrossing counterparts.

132

133 ***MATERIAL AND METHODS***

134

135 **General assumptions**

136

137 We consider the evolution of a quantitative trait Z in a population of size N , made of
138 diploid individuals reproducing through partial self-fertilization, with a constant selfing rate σ .
139 The phenotypic value z of an individual is determined by the additive action of L loci each
140 with an infinite possible number of alleles and is given by

$$141 \quad z = g + e, \quad (1)$$

142 where g is the genetic component of the individual's phenotype, and is given by $g = \sum_j^L g_j^M +$
143 g_j^P , with g_j^M (respectively g_j^P) the additive allelic effect at locus j inherited from the maternal
144 (respectively paternal) gamete. The random environmental effect, e , is drawn from a Gaussian
145 distribution of mean 0 and variance V_E , and is considered to be independent from the genetic
146 components of fitness. The trait initially undergoes stabilizing selection around an optimal
147 phenotypic value (set arbitrarily at 0). The fitness value W_Z of an individual with phenotype z
148 is thus described by the Gaussian function:

$$149 \quad W_Z = e^{-d^2/2\omega^2}, \quad (2)$$

150 where d is the distance between the individual's phenotype z and the optimum trait value and
151 ω^2 is the width of the fitness function, and represents the strength of selection.

152

153 **Simulation model**

154

155 We implement the model described above into an individual based simulation model
156 written in C++, a modified version of the "continuum of alleles" program provided in Abu
157 Awad and Roze (2018). The simulation program is available in File S1 and online
158 (<https://github.com/dialaAbAw/SelfingAdaptation>). The life cycle can be summarized by five

159 successive events: (1) phenotype-dependent choice of the maternal parent (selection), (2)
160 mating-type choice (selfing versus outcrossing), (3) phenotype-dependent choice of the
161 paternal parent in the case of outcrossing, (4) production of gametes (recombination) and (5)
162 mutation. We simulate two phases. In the first one, the population evolves in a stable
163 environment (for a fixed trait optimum value $Z_{\text{OPT}} = 0$) until mutation-selection-drift
164 equilibrium (M-S-D) and we explore the effect of the mating system on the genetic
165 components and structure of a quantitative trait under stabilizing selection. In the second
166 phase, we consider the rate of adaptation following a brutal shift in the optimum (Z_{OPT} shift
167 from 0 to 2.5, the shift being of order $L \cdot a^2$).

168 Each generation before equilibrium, the number of new mutations per chromosome
169 per generation is sampled from a Poisson distribution with parameter U , the per-trait haploid
170 genomic mutation rate (ranging from 0.005 to 0.1, in accordance with the literature (Keightley
171 and Bataillon 2000; Shaw *et al.* 2002; Haag-Liautard *et al.* 2007)). The additive value of a
172 new mutant allele is drawn from a Normal distribution of mean 0 and variance a^2 . We use
173 parameter set values similar to those in Bürger *et al.* (1989) and Ronce *et al.* (2009), with the
174 number of freely recombining loci under selection $L = 50$, $a^2 = 0.05$, $V_E = 1$, $\omega^2 = 1$ (and hence
175 $V_S = \omega^2 + V_E$ is equal to 2). The mean deleterious effect of mutations \bar{s} ($\bar{s} = a^2 / 2V_S$, Martin
176 and Lenormand 2006) is equal to 0.0125. Although simulations were run over a large range of
177 selfing values, throughout the manuscript we show results run principally for three rates of
178 self-fertilisation, $\sigma = 0, 0.5$ and 0.95 , representing outcrossing, mixed-mating and
179 predominantly selfing respectively. They are representative of the three main patterns
180 observed over the whole range (σ from 0 to 1). We also considered two population sizes $N =$
181 250 and 1000.

182

183 **Simulation program:**

184

185 At the start of a simulation, all N individuals are homozygous for allele 0 at all L loci
186 and are thus at the fitness optimum. To form the next generation, N new zygotes are produced
187 through selfing with probability σ , and through random mating with probability $1-\sigma$. Selection
188 occurs during the sampling of parental individuals, occurring with probabilities proportional
189 to their fitness. During meiosis, the number of cross-overs is sampled from a Poisson
190 distribution with parameter R (which represents the genome map length), and the position of
191 each cross-over along the chromosome is sampled from an uniform distribution. According to
192 Haldane's mapping function, the recombination rate between two adjacent loci is $r =$
193 $\frac{1}{2}\left[1 - \exp\left(\frac{-2R}{L-1}\right)\right]$, with $R = 50$ in our simulations. Mutation occurs after recombination: the
194 number of new mutations per chromosome per generation is sampled from a Poisson
195 distribution with parameter U , the haploid genomic mutation rate ($U = L\mu$, with μ being the
196 per locus mutation rate). The additive value of a new mutant allele is drawn from a Normal
197 distribution of mean 0 and variance a^2 .

198 After reaching the M-S-D equilibrium, we introduce an environmental change by
199 shifting the phenotypic optimum. The only source of genetic variability to reach the new
200 optimum is the standing variation accumulated at M-S-D equilibrium (after the shift, U is set
201 to 0). We then let the population evolve for 200 generations.

202

203 *Effect of selfing on genetic variance structuration at Mutation-Selection-Drift equilibrium*

204

205 Following Turelli & Barton (Turelli and Barton 1990), we decompose the genetic variance of
206 a polygenic trait using the following equation:

$$207 \quad V_A = 2 \sum_j^L (C_{jj} + C_{j,j}) + 2 \sum_{j \neq k}^L (C_{jk} + C_{j,k}), \quad (3)$$

208 with

$$209 \quad C_{jj} = \frac{1}{2} E[(g_j^M - \bar{g}_j)^2 + (g_j^P - \bar{g}_j)^2] \quad (4)$$

210 and

$$211 \quad C_{jj} = E[(g_j^M - \bar{g}_j) \cdot (g_j^P - \bar{g}_j)], \quad (5)$$

212 where \bar{g}_j is the mean allelic effect on the phenotype at locus j and g_j^M (respectively g_j^P) is the
213 allelic effect at locus j inherited from the maternal (respectively paternal) gamete. The sum of
214 all values of C_{jj} represents the variance of allelic effects (the genic variance V_{genic} , the genetic
215 variance of a trait in a population harboring the same allelic frequencies as the population
216 under study, but without any genetic association between loci) and is computed from
217 simulation outputs following equation (4). The sum of all values of C_{jj} represents the
218 covariance in allelic effects on the maternally and paternally inherited chromosomes at locus
219 j , and represents the fraction of the genetic variance due to excess of homozygosity (named
220 V_{inbred}); we compute it following equation (5). This quantity represents $F \cdot V_{\text{genic}}$, where F is the
221 inbreeding coefficient of the population. These first term of equation (3) ($2 \sum_j^L (C_{jj} + C_{j,j})$)
222 represents the genetic variance due to within locus variation. The second term ($2 \sum_{j \neq k}^L (C_{jk} +$
223 $C_{j,k})$) represents the component of the variance due to associations between loci (noted V_{LD}),
224 and is obtained by subtracting V_{genic} and V_{inbred} from the total additive genetic variance. This
225 component is proportional to linkage disequilibrium (LD), and tends to be negative under
226 stabilizing selection due to associations between alleles from different loci with compensatory

227 effects (*i.e.* the allele on one locus is positive, the other negative, their effects on the
228 phenotype thus cancel out when both are present).

229

230 *Analyses after the environmental change*

231

232 Due to an increased probability of extinction during long periods of maladaptation, it
233 seems more relevant to focus only on the dynamics of adaptation during the first generations
234 after an environmental change. We follow the temporal dynamics of the additive variance and
235 its components, the dynamic of the mean population phenotype, and the dynamic of the mean
236 population fitness, as a function of the mating system and the mutation rate, over 20
237 generations.

238 In addition, and in order to test if remobilization of V_{LD} plays a role in the adaptation
239 of selfing populations, we computed the slope of the trait mean dynamics just after the
240 environmental change (during the first five generations) as a function of the amount of
241 additive variance available at M-S-D equilibrium. If remobilization of V_{LD} is involved in the
242 adaptive process of selfing populations, the initial slope, for a similar amount of additive
243 variance, should be higher in selfing populations compared to mixed mating and outcrossing
244 ones.

245

246 **RESULTS**

247

248 Below we present results only for $N = 250$ and $L = 50$, as larger population sizes and
249 more loci did not qualitatively change the results. Results for $N = 1000$, as well as $L = 500$ are
250 given in the supplementary materials section (Figures S1-S6).

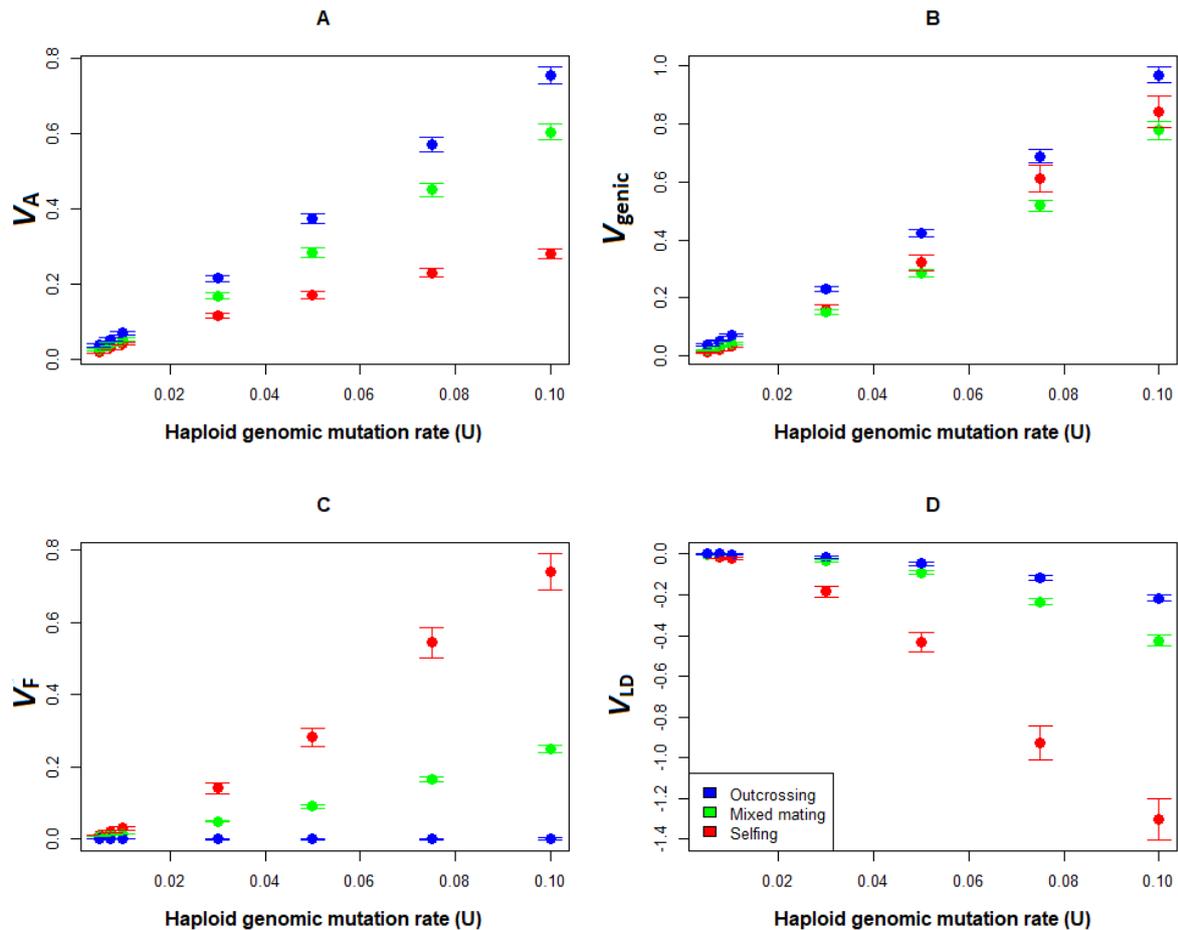
251

252 **DECOMPOSITION OF THE GENETIC VARIANCE AT EQUILIBRIUM**

253

254 As predicted, the additive genetic variance present at M-S-D equilibrium in our
255 simulations is negatively correlated with the selfing rate (figure 1A, see Abu Awad and Roze
256 2018 for the analytical model). By examining each component of the additive variance, we
257 can see that the variance due to within loci variation ($V_{\text{genic}} + V_F$) is higher in selfing
258 populations due to the higher rate of homozygosity (figure 1 B & C). These two components,
259 as well as the total additive variance, increase linearly with increasing mutation rates (figure 1
260 A, B & C). The among-loci component (V_{LD}) is negative (figure 1D), due to the build-up of
261 linkage disequilibrium between alleles with different signs, allowing phenotypes to be close to
262 the phenotypic optimum (0). Because recombination is less effective under selfing, this
263 negative component is responsible for the smaller additive variance observed under
264 predominant selfing. It also contributes significantly to the variance observed in mixed mating
265 and outcrossing populations with moderate to high mutation rates (figure 1D). Larger
266 population sizes or more loci do not change the results for outcrossing and mixed mating
267 populations (Figures S1-S2). For selfing populations, larger population sizes and higher per
268 locus mutation rates contribute to increasing negative linkage disequilibrium (V_{LD}) due to
269 negative linkage disequilibrium (Figure S3).

270



271

272 **Figure 1.** Additive genetic variance and its components as a function of the genomic mutation
273 rate and the mating system. **A.** Observed additive variance for the phenotypic trait. **B.** Genic
274 variance for the phenotypic trait (V_{genic}). **C.** Genetic variance due to inbreeding (V_F). **D.**
275 Genetic variance due to linkage disequilibrium (V_{LD}). Error bars stand for 95% confidence
276 interval ($n = 100$).

277

278 ADAPTATION THROUGH STANDING GENETIC VARIATION:

279

280 For simplicity, we focus on four mutation rates ($U = 0.005; 0.03; 0.05; 0.1$),
281 representing the different patterns of genetic variance observed at M-S-D equilibrium for the

282 outcrossing, mixed mating and selfing populations mentioned above. We first describe the
283 dynamics observed for outcrossing populations, which will serve as a reference for
284 comparison with the dynamics of selfing populations. We generally find that the dynamics for
285 mixed mating populations are similar to those of outcrossing populations (Figure S4).

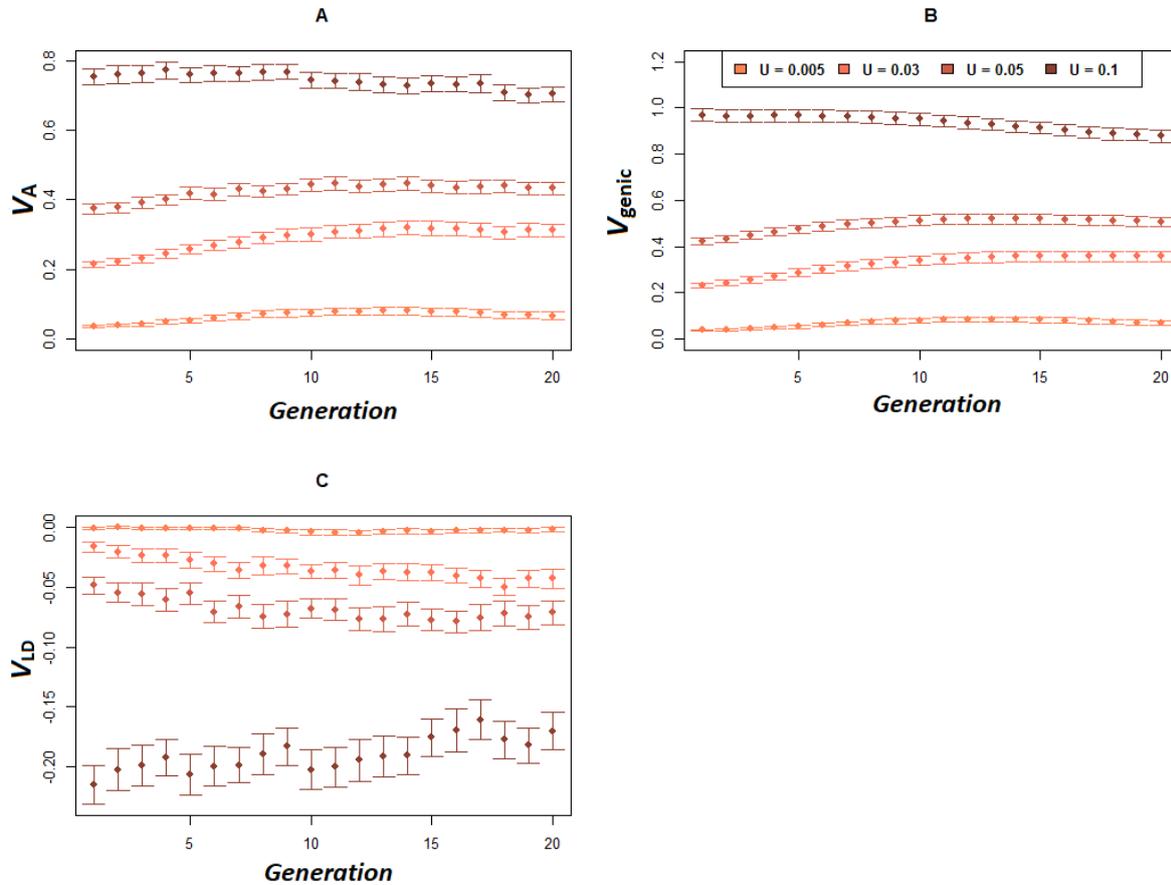
286

287 **DYNAMICS OF THE ADDITIVE VARIANCE AND OF ITS COMPONENTS:**

288

289 In outcrossing populations, the additive variance exhibits very tiny changes during the
290 adaptation process, so do its components (Figure 2). In all cases, the observed additive genetic
291 variance slightly increases during the first generations of adaptation (Figure 2A), then, it
292 either reaches an equilibrium (Figure 2A, $U < 0.1$) or slowly begins to decrease (Figure 2A, U
293 $= 0.1$). When V_{LD} is small at M-S-D equilibrium ($U < 0.1$), the observed increase of the
294 additive variance is mainly due to an increase of the genic variance (Figure 2B), probably due
295 to successive sweeps of rare alleles during the adaptation process. In these situations, V_{LD}
296 slightly decreases with time (Figure 2C), indicating that some associations between loci are
297 building up. When V_{LD} is significant at M-S-D equilibrium ($U = 0.1$), the genic variance
298 remains constant during the first generations (Figure 2B), but V_{LD} increases slightly (Figure
299 2C), contributing to the increase of the additive variance. The second phase during which the
300 observed additive variance decreases can be explained by the decrease of genic variance with
301 time, due to the purging of deleterious mutations (Figure 2B).

302



303

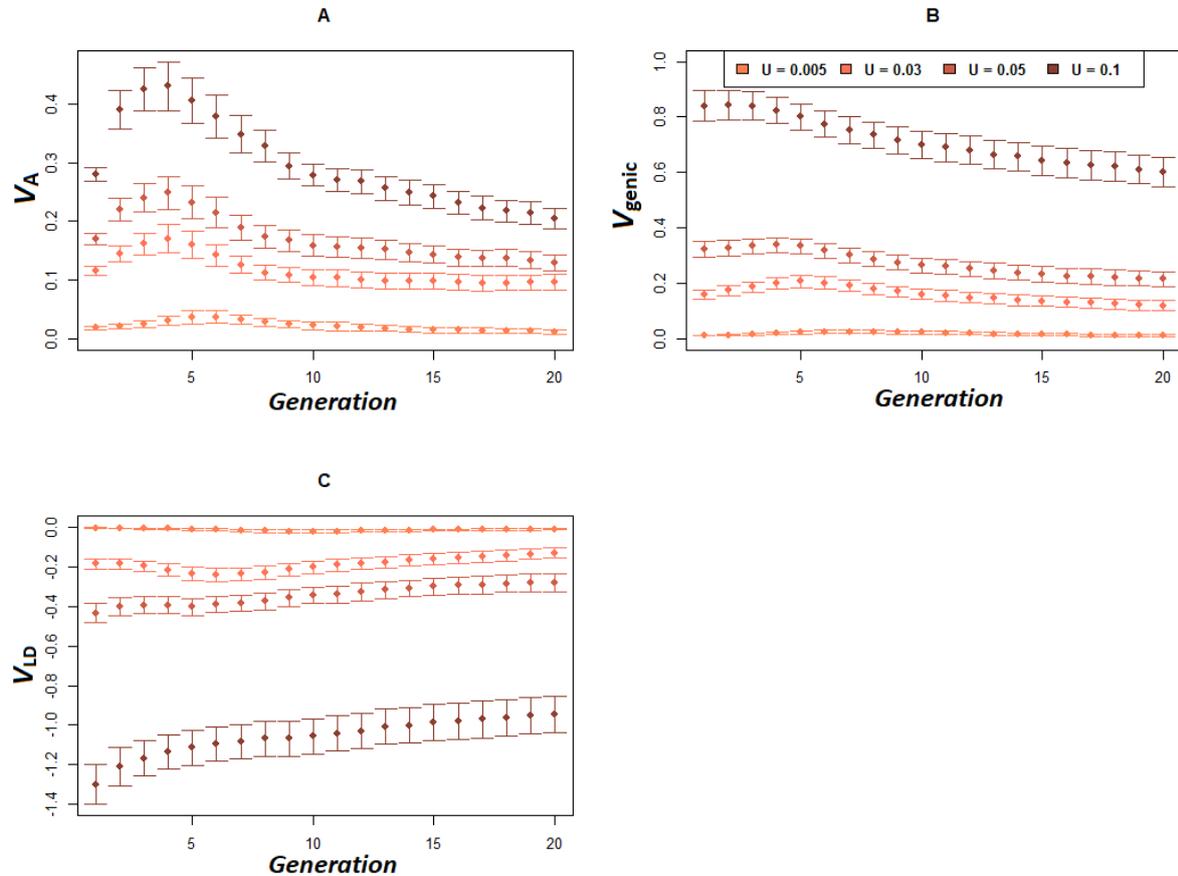
304 **Figure 2.** Dynamics of additive genetic variance and its components in function of the
305 haplotypic trait mutation rate, for outcrossing populations. **A.** Observed additive variance for
306 the phenotypic trait. **B.** Genic variance for the phenotypic trait (V_{genic}). **C.** Genetic variance of
307 the phenotype due to linkage disequilibrium (V_{LD}). Error bars stand for 95% confidence
308 interval (n=100).

309

310 Contrary to the dynamics observed in outcrossing populations, additive variance
311 substantially varies during the adaptation process in selfing populations. For small mutation
312 rates ($U \leq 0.03$), the dynamics are very similar to those observed in outcrossing populations
313 (figure 3). The observed additive variance slightly increases during the first generations
314 (Figure 3A). This increase is due to a rise of the genic variance (Figure 3B), associated with a

315 decrease of V_{LD} (Figure 3C), probably due to an interaction between sweeps of beneficial
316 mutations that were rare before the environmental change and the creation of new beneficial
317 associations between loci. For higher mutation rates ($U \geq 0.05$), the dynamics are different.
318 The genic variance remains constant during the first generations and then decreases (figure
319 3B), whereas V_{LD} increases faster than the decrease of V_{genic} (figure 3C), explaining the
320 overall increase of additive variance. This behaviour can be explained as follows: there is a
321 complex interaction between the purging of deleterious mutations due to selfing, which tends
322 to quickly fix the best multi-locus genotypes, eroding genetic diversity and residual allogamy,
323 which allows the mobilization of a small fraction of the hidden genetic diversity. The outcome
324 of this interaction is an increase of the genetic diversity and of the populations' adaptive
325 potential. The fact that the rate of adaptation (*i.e* the slope of the change in the trait values
326 during the first generations) as a function of the initial level of additive variance is higher for
327 selfing populations when V_{LD} is large, confirms that the remobilisation of the hidden diversity
328 plays a role in the adaptation process (figure S5).

329



330

331 **Figure 3.** Dynamics of additive genetic variance and its components in function of the
332 haplotypic trait mutation rate, for selfing populations. **A.** Observed additive variance for the
333 phenotypic trait. **B.** Genic variance for the phenotypic trait (V_{genic}). **C.** Genetic variance of the
334 phenotype due to linkage disequilibrium (V_{LD}). Error bars stand for 95% confidence interval
335 ($n=100$).

336

337 **TRAIT AND FITNESS DYNAMICS DURING ADAPTATION PROCESS:**

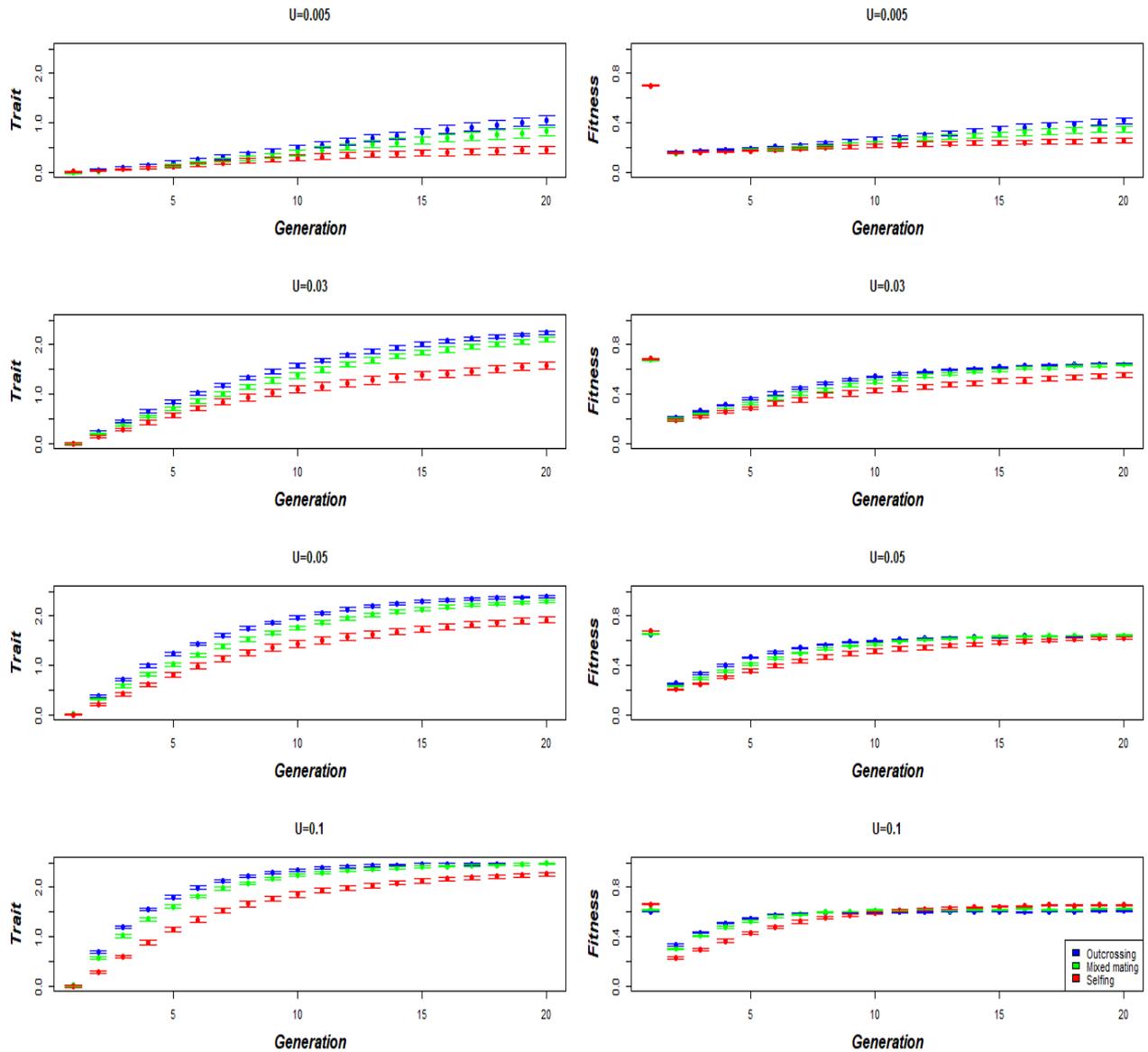
338

339 Here, we consider that there is adaptation if populations reach a similar level of fitness
340 as that observed before the environmental change. In all cases, compared to selfing

341 populations, outcrossing and mixed mating population respond better to selection, by being
342 closer to the new phenotypic optimum (figure 4). When the mutation rate is weak ($U =$
343 0.005), none of the populations are able to adapt, and thus the fitness remains low (figure 4).
344 For higher mutation rates ($U > 0.005$), outcrossing and mixed mating populations are always
345 close to the new optimum, and are able to return to fitness levels similar to those observed at
346 M-S-D equilibrium. Selfing populations require that the mutation rate is high enough ($U \geq$
347 0.05 , figure 4) in order to reach similar fitness levels as those observed in outcrossing
348 populations. Interestingly, in some cases, adaptation occurs more rapidly in selfing
349 populations, despite the higher genetic diversity of outcrossing populations. The larger the
350 population size and the higher the per-locus mutation rate, the higher the level of potentially
351 usable hidden genetic diversity, and the smaller the genomic mutation rate necessary to reach
352 similar levels of fitness in selfing populations (figures S6-S7).

353 Nevertheless, different mating systems exhibit different dynamics of adaptation.
354 Outcrossing populations adapt by reaching a new phenotypic optimum, but they also maintain
355 a high level of genetic diversity (figure 3), resulting in populations having both mal-adapted
356 and well-adapted individuals. Selfing populations are always further away from the new
357 phenotypic optimum (figure 4), but both the remobilization of the V_{LD} , and the fast fixation of
358 the “best” phenotypes allow for the production of close-to-optimum individuals. These
359 phenotypes will then quickly increase in frequency, leading to more homogenous populations
360 than observed for an outcrossing mating system (figure 4).

361



362

363 **Figure 4.** Dynamics of the trait and fitness, in function of the trait haplotypic mutation rate

364 and the mating system. Error bars stand for 95% confidence interval (n=100).

365

366 **DISCUSSION**

367

368 In accordance with Stebbins' definition of the dead-end hypothesis (Stebbins 1957),

369 single-locus models predict that adaptation from standing genetic variation is less likely in

370 selfing populations compared to outcrossing ones, notably due to the reduced genetic
371 variation resulting from purging (Glémin and Ronfort 2013). Considering a polygenic trait,
372 and associated interactions among loci, we find that this is not always the case. Indeed, we
373 find that for low mutation rates, our results support the expectations of single locus models.
374 However, for higher mutation rates, and as predicted by previous work (Abu Awad and Roze
375 2018), associations between loci are no longer negligible. In this case, stabilizing selection
376 shelters an important amount of hidden additive variance, especially in self-fertilising
377 populations. We show that some of this variance can be released during phases of directional
378 selection to new environmental conditions. Our results support that it is possible to observe
379 similar levels of adaptation in selfing and outcrossing populations, despite unconditionally
380 lower levels of observed additive variance under predominant selfing.

381

382 **Hidden genetic variation, its remobilization and genotypic selection**

383

384 Stabilizing selection is expected to favour the build-up and maintenance of co-adapted
385 gene complexes, and this will structure the additive variance into positive within-loci
386 components and negative among-loci components, reducing the observed additive variance
387 (Abu Awad and Roze 2018). In a changing environment, alleles involved in the negative
388 among loci component of variance may be unveiled and mobilized to respond to directional
389 selection (Le Rouzic and Carlborg 2008). Our analysis shows that such associations between
390 loci, coupled with hidden genetic variation, are more likely to emerge in selfing than in
391 outcrossing populations, because of less efficient recombination under selfing (Lande and
392 Porcher 2015; Abu Awad and Roze 2018). This prediction is in accordance with empirical
393 observations of more frequent transgressive segregation (progeny of a cross being outside the

394 phenotypic parental range) in inbred compared to outbred species (Rieseberg *et al.* 1999;
395 Johansen-Morris and Latta 2006). Our analysis also shows that the less effective
396 recombination rate associated with selfing avoids the complete release of the negative linkage
397 disequilibria and thus limits the response to selection. As a result, selection mostly operates at
398 the genotypic level while allelic selection is more prevalent in more recombining populations
399 (Neher and Shraiman 2009). Interestingly, if the rate of self-fertilization is changed during the
400 adaptation process (σ changing from 0.95 to zero after the environmental change), the
401 dynamics of the fitness remains similar (figures S8-S9). Indeed, under complete outcrossing
402 all the hidden genetic variance can be mobilized and this allows initially selfing populations
403 to reach the new phenotypic optimum but recombination also generates less adapted
404 genotypes which reduces the mean population fitness.

405

406 ***De novo* mutations vs. standing genetic variation: rethinking adaptation in selfing**
407 **species?**

408

409 It has been a long accepted paradigm that the advantage procured by selfing, was the
410 rapid fixation of *de novo* mutations more efficiently than in outcrossing populations,
411 independently of the dominance of new mutations, a process known as “Haldane sieve”
412 (Haldane 1927). Indeed, from one locus theory, adaptation through new mutations is more
413 likely in selfing species, and is more likely than adaptation from standing genetic variation
414 (Glémin and Ronfort 2013). However, recent works have suggested that the reduced effective
415 recombination rate of selfing populations adds a disadvantage even when it comes to the
416 fixation of new mutations. Unlike what is expected in outcrossing populations, the fixation of
417 beneficial mutations in selfing populations can be hindered if they appear during the selective

418 sweep triggered by a beneficial allele at another locus (Hartfield and Glémin 2016). This
419 observation as well as the results presented here show that predictions from the point of view
420 of polygenic models are less dichotomist: the ability of selfing populations to adapt from
421 either *de novo* mutations or standing variation strongly depends on the parameters considered
422 (the mutation rate in particular), with selfers being often as able to adapt as outcrossers, even
423 if the underlying mechanisms strongly differ.

424 We have only considered a simple architecture of quantitative traits in which epistatic
425 interactions emerge naturally. Epistasis, and notably its directionality, is known to play a key
426 role in adaptation (Hansen 2013). Positive epistasis, with genes that reinforce each other's
427 effects in the direction of selection, inflate the additive variance and thus the ability of
428 populations to adapt to an environmental change (Carter *et al.* 2005; Monnahan and Kelly
429 2015), contrary to the non-directional epistatic scenario (Carter *et al.* 2005). Negative
430 epistasis, where genes tend to mute each other's effects, reduces the additive variance of the
431 character, thus limiting adaptive potential (Carter *et al.* 2005). Few empirical estimations of
432 the directionality of epistasis are available in literature (Le Rouzic 2014; Monnahan and Kelly
433 2015; Oakley *et al.* 2015, all detecting positive epistatic interactions), despite numerous
434 methods and the diversity of data used to infer it (Le Rouzic 2014). Developing methods to
435 detect and measure the directionality of epistatic interactions in relation to the mating system
436 may bring us closer to understanding the differences in patterns of adaptation observed in
437 selfing and outcrossing populations.

438

439 **New insights into the role of standing genetic variation in the adaptation dynamics of**
440 **selfing populations**

441

442 The overwhelming success of selfing species in the domestication process and as
443 invasive species is not considered as resulting from a higher adaptive ability of selfing species
444 compared to outcrossing ones. For instance, the invasive success of selfing populations is
445 attributed to reproductive assurance, since a single individual is able to colonize a new
446 environment (Rambuda and Johnson 2004; van Kleunen *et al.* 2008), and to reduce gene flow
447 which are expected to limit maladapted gene exchanges between populations (Levin 2010).
448 Regarding domestication, it has been argued that selection in selfing populations is most
449 probably due to new mutations, because the standing genetic variation is lower in such
450 populations due to more efficient purging of deleterious mutations that could be involved in
451 the domestication process (Glémin and Bataillon 2009). This idea is reinforced by the fact that
452 selfing species are expected to quickly fix a rare beneficial mutation, independently from its
453 dominance level (Ross-Ibarra 2005). In their review on mating system variation in
454 domesticated plant species, Glémin and Bataillon (2009) have suggested that the high
455 frequency of self-fertilising crop species could be related to an increase in additive variance
456 during domestication; this idea has however never been tested theoretically or empirically.
457 Here we show that this increase in additive variance could indeed be an advantage when
458 selfing species are faced with new environments. However, our results hold true only if
459 bottlenecks during the domestication and invasion processes are not too strong or if mutation
460 rates are high.

461

462 **CONCLUSION AND PERSPECTIVES**

463

464 In this work, we argue that selfing populations under stabilizing selection are able to
465 accumulate hidden genetic variation through negative linkage disequilibrium. Thanks to this

466 hidden variation, adaptation under high self-fertilisation rates is not always limited by the
467 expected reduction in additive genetic variance due to purging. We therefore suggest that a
468 simple estimation of additive variance of quantitative traits is not adequate when speculating
469 on the long-term adaptive capacity of a population. Complementary analyses should also be
470 carried out when quantifying the long-term evolvability of a population. Such analyses
471 include looking for transgressive segregation, or carrying out experimental evolutionary
472 experiments in which directional selection is induced. More empirical evidence is required to
473 determine how frequent is cryptic diversity in natural populations of selfing species, and
474 whether or not this property is sufficient to allow for selfing species to adapt to a changing
475 environment.

476

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485 **Bibliography:**

486

487 Abu Awad D., and D. Roze, 2018 Effects of partial selfing on the equilibrium genetic variance,
488 mutation load, and inbreeding depression under stabilizing selection. *Evolution* 72: 751–769.

- 489 Anderson J. T., C.-R. LEE, C. A. Rushworth, R. I. Colautti, and T. MITCHELL-OLDS, 2013 Genetic
490 trade-offs and conditional neutrality contribute to local adaptation. *Mol. Ecol.* 22: 699–708.
- 491 Badano J. L., and N. Katsanis, 2002 Human genetics and disease: Beyond Mendel: an evolving view
492 of human genetic disease transmission. *Nat. Rev. Genet.* 3: 779.
- 493 Barrett R. D., and D. Schluter, 2008 Adaptation from standing genetic variation. *Trends Ecol. Evol.*
494 23: 38–44.
- 495 Berger D., S. S. Bauerfeind, W. U. Blanckenhorn, and M. A. Schäfer, 2011 High temperatures reveal
496 cryptic genetic variation in a polymorphic female sperm storage organ. *Evol. Int. J. Org. Evol.*
497 65: 2830–2842.
- 498 Bürger R., G. P. Wagner, and F. Stettinger, 1989 HOW MUCH HERITABLE VARIATION CAN BE
499 MAINTAINED IN FINITE POPULATIONS BY MUTATION–SELECTION BALANCE?
500 *Evolution* 43: 1748–1766. <https://doi.org/10.1111/j.1558-5646.1989.tb02624.x>
- 501 Carter A. J., J. Hermisson, and T. F. Hansen, 2005 The role of epistatic gene interactions in the
502 response to selection and the evolution of evolvability. *Theor. Popul. Biol.* 68: 179–196.
- 503 Charlesworth D., and B. Charlesworth, 1995 Quantitative genetics in plants: the effect of the breeding
504 system on genetic variability. *Evolution* 49: 911–920.
- 505 Clo J., L. Gay, and J. Ronfort, 2019 How does selfing affect the genetic variance of quantitative traits?
506 An updated meta-analysis on empirical results in angiosperm species. *Evolution*.
- 507 Geber M. A., and L. R. Griffen, 2003 Inheritance and natural selection on functional traits. *Int. J. Plant*
508 *Sci.* 164: S21–S42.
- 509 Gibson G., and I. Dworkin, 2004 Uncovering cryptic genetic variation. *Nat. Rev. Genet.* 5: 681.
- 510 Glémin S., and T. Bataillon, 2009 A comparative view of the evolution of grasses under
511 domestication. *New Phytol.* 183: 273–290. <https://doi.org/10.1111/j.1469-8137.2009.02884.x>

- 512 Glémin S., and J. Ronfort, 2013 Adaptation and maladaptation in selfing and outcrossing species: new
513 mutations versus standing variation. *Evol. Int. J. Org. Evol.* 67: 225–240.
- 514 Haag-Liautard C., M. Dorris, X. Maside, S. Macaskill, D. L. Halligan, *et al.*, 2007 Direct estimation of
515 per nucleotide and genomic deleterious mutation rates in *Drosophila*. *Nature* 445: 82.
- 516 Haldane J. B. S., 1927 A mathematical theory of Natural and Artificial Selection. *Math. Proc. Camb.*
517 *Philos. Soc.* 23: 607–615. <https://doi.org/10.1017/S0305004100011750>
- 518 Hansen T. F., 2013 WHY EPISTASIS IS IMPORTANT FOR SELECTION AND ADAPTATION.
519 *Evolution* 67: 3501–3511. <https://doi.org/10.1111/evo.12214>
- 520 Hartfield M., and S. Glémin, 2016 Limits to Adaptation in Partially Selfing Species. *Genetics* 203:
521 959. <https://doi.org/10.1534/genetics.116.188821>
- 522 Hermisson J., and P. S. Pennings, 2005 Soft sweeps: molecular population genetics of adaptation from
523 standing genetic variation. *Genetics* 169: 2335–2352.
- 524 Igic B., and J. W. Busch, 2013 Is self-fertilization an evolutionary dead end? *New Phytol.* 198: 386–
525 397.
- 526 Innan H., and Y. Kim, 2004 Pattern of polymorphism after strong artificial selection in a
527 domestication event. *Proc. Natl. Acad. Sci.* 101: 10667–10672.
- 528 Johansen-Morris A. D., and R. G. Latta, 2006 Fitness consequences of hybridization between ecotypes
529 of *Avena barbata*: hybrid breakdown, hybrid vigor, and transgressive segregation. *Evolution*
530 60: 1585–1595.
- 531 Keightley P. D., and T. M. Bataillon, 2000 Multigeneration Maximum-Likelihood Analysis Applied to
532 Mutation-Accumulation Experiments in *Caenorhabditis elegans*. *Genetics* 154:
533 1193.

- 534 Kim S.-Y., J. C. Noguera, A. Tato, and A. Velando, 2013 Vitamins, stress and growth: the availability
535 of antioxidants in early life influences the expression of cryptic genetic variation. *J. Evol.*
536 *Biol.* 26: 1341–1352.
- 537 Kleunen M. van, J. C. Manning, V. Pasqualetto, and S. D. Johnson, 2008 Phylogenetically
538 Independent Associations between Autonomous Self-Fertilization and Plant Invasiveness.
539 *Am. Nat.* 171: 195–201. <https://doi.org/10.1086/525057>
- 540 Lande R., and E. Porcher, 2015 Maintenance of quantitative genetic variance under partial self-
541 fertilization, with implications for evolution of selfing. *Genetics* 200: 891–906.
- 542 Le Rouzic A., and Ö. Carlborg, 2008 Evolutionary potential of hidden genetic variation. *Trends Ecol.*
543 *Evol.* 23: 33–37.
- 544 Le Rouzic A., 2014 Estimating directional epistasis. *Front. Genet.* 5: 198.
545 <https://doi.org/10.3389/fgene.2014.00198>
- 546 Ledon-Rettig C. C., D. W. Pfennig, and E. J. Crespi, 2010 Diet and hormonal manipulation reveal
547 cryptic genetic variation: implications for the evolution of novel feeding strategies. *Proc. R.*
548 *Soc. B Biol. Sci.* 277: 3569–3578.
- 549 Levin D. A., 2010 Environment-enhanced self-fertilization: implications for niche shifts in adjacent
550 populations. *J. Ecol.* 98: 1276–1283. <https://doi.org/10.1111/j.1365-2745.2010.01715.x>
- 551 Manna F., G. Martin, and T. Lenormand, 2011 Fitness Landscapes: An Alternative Theory for the
552 Dominance of Mutation. *Genetics* 189: 923. <https://doi.org/10.1534/genetics.111.132944>
- 553 Martin G., and T. Lenormand, 2006 A GENERAL MULTIVARIATE EXTENSION OF FISHER'S
554 GEOMETRICAL MODEL AND THE DISTRIBUTION OF MUTATION FITNESS
555 EFFECTS ACROSS SPECIES. *Evolution* 60: 893–907. [https://doi.org/10.1111/j.0014-](https://doi.org/10.1111/j.0014-3820.2006.tb01169.x)
556 [3820.2006.tb01169.x](https://doi.org/10.1111/j.0014-3820.2006.tb01169.x)

- 557 Martin G., S. F. Elena, and T. Lenormand, 2007 Distributions of epistasis in microbes fit predictions
558 from a fitness landscape model. *Nat. Genet.* 39: 555–560. <https://doi.org/10.1038/ng1998>
- 559 Matuszewski S., J. Hermisson, and M. Kopp, 2015 Catch me if you can: adaptation from standing
560 genetic variation to a moving phenotypic optimum. *Genetics* 200: 1255–1274.
- 561 McGuigan K., N. Nishimura, M. Currey, D. Hurwit, and W. A. Cresko, 2011 Cryptic genetic variation
562 and body size evolution in threespine stickleback. *Evol. Int. J. Org. Evol.* 65: 1203–1211.
- 563 Milloz J., F. Duveau, I. Nuez, and M.-A. Félix, 2008 Intraspecific evolution of the intercellular
564 signaling network underlying a robust developmental system. *Genes Dev.* 22: 3064–3075.
- 565 Mittell E. A., S. Nakagawa, and J. D. Hadfield, 2015 Are molecular markers useful predictors of
566 adaptive potential? *Ecol. Lett.* 18: 772–778.
- 567 Monnahan P. J., and J. K. Kelly, 2015 Epistasis is a major determinant of the additive genetic variance
568 in *Mimulus guttatus*. *PLoS Genet.* 11: e1005201.
- 569 Neher R. A., and B. I. Shraiman, 2009 Competition between recombination and epistasis can cause a
570 transition from allele to genotype selection. *Proc. Natl. Acad. Sci.* 106: 6866.
571 <https://doi.org/10.1073/pnas.0812560106>
- 572 Nordborg M., 2000 Linkage disequilibrium, gene trees and selfing: an ancestral recombination graph
573 with partial self-fertilization. *Genetics* 154: 923–929.
- 574 Oakley C. G., J. Ågren, and D. W. Schemske, 2015 Heterosis and outbreeding depression in crosses
575 between natural populations of *Arabidopsis thaliana*. *Heredity* 115: 73.
- 576 Orr H. A., and A. J. Betancourt, 2001 Haldane’s sieve and adaptation from the standing genetic
577 variation. *Genetics* 157: 875–884.
- 578 Paaby A. B., and M. V. Rockman, 2014 Cryptic genetic variation: evolution’s hidden substrate. *Nat.*
579 *Rev. Genet.* 15: 247.

- 580 Pritchard J. K., J. K. Pickrell, and G. Coop, 2010 The genetics of human adaptation: hard sweeps, soft
581 sweeps, and polygenic adaptation. *Curr. Biol.* 20: R208–R215.
- 582 Queitsch C., T. A. Sangster, and S. Lindquist, 2002 Hsp90 as a capacitor of phenotypic variation.
583 *Nature* 417: 618.
- 584 Rambuda T. D., and S. D. Johnson, 2004 Breeding systems of invasive alien plants in South Africa:
585 does Baker’s rule apply? *Divers. Distrib.* 10: 409–416. [https://doi.org/10.1111/j.1366-](https://doi.org/10.1111/j.1366-9516.2004.00100.x)
586 [9516.2004.00100.x](https://doi.org/10.1111/j.1366-9516.2004.00100.x)
- 587 Rieseberg L. H., M. A. Archer, and R. K. Wayne, 1999 Transgressive segregation, adaptation and
588 speciation. *Heredity* 83: 363–372.
- 589 Rohner N., D. F. Jarosz, J. E. Kowalko, M. Yoshizawa, W. R. Jeffery, *et al.*, 2013 Cryptic variation in
590 morphological evolution: HSP90 as a capacitor for loss of eyes in cavefish. *Science* 342:
591 1372–1375.
- 592 Ronce O., F. H. Shaw, F. Rousset, and R. G. Shaw, 2009 IS INBREEDING DEPRESSION LOWER
593 IN MALADAPTED POPULATIONS? A QUANTITATIVE GENETICS MODEL. *Evolution*
594 63: 1807–1819. <https://doi.org/10.1111/j.1558-5646.2009.00678.x>
- 595 Ross-Ibarra J., 2005 Quantitative trait loci and the study of plant domestication, pp. 197–204 in
596 *Genetics of Adaptation*, edited by Mauricio R. Springer Netherlands, Dordrecht.
- 597 Shao H., L. C. Burrage, D. S. Sinasac, A. E. Hill, S. R. Ernest, *et al.*, 2008 Genetic architecture of
598 complex traits: large phenotypic effects and pervasive epistasis. *Proc. Natl. Acad. Sci.* 105:
599 19910–19914.
- 600 Shaw F. H., C. J. Geyer, and R. G. Shaw, 2002 A COMPREHENSIVE MODEL OF MUTATIONS
601 AFFECTING FITNESS AND INFERENCES FOR ARABIDOPSIS THALIANA. *Evolution*
602 56: 453–463. <https://doi.org/10.1111/j.0014-3820.2002.tb01358.x>

- 603 Stebbins G. L., 1957 Self fertilization and population variability in the higher plants. *Am. Nat.* 91:
604 337–354.
- 605 Takebayashi N., and P. L. Morrell, 2001 Is self-fertilization an evolutionary dead end? Revisiting an
606 old hypothesis with genetic theories and a macroevolutionary approach. *Am. J. Bot.* 88: 1143–
607 1150.
- 608 Turelli M., and N. H. Barton, 1990 Dynamics of polygenic characters under selection. *Theor. Popul.*
609 *Biol.* 38: 1–57. [https://doi.org/10.1016/0040-5809\(90\)90002-D](https://doi.org/10.1016/0040-5809(90)90002-D)
- 610 Waddington C. H., 1953 Genetic assimilation of an acquired character. *Evolution* 7: 118–126.
- 611 Wood J. L., M. C. Yates, and D. J. Fraser, 2016 Are heritability and selection related to population
612 size in nature? Meta-analysis and conservation implications. *Evol. Appl.* 9: 640–657.
- 613