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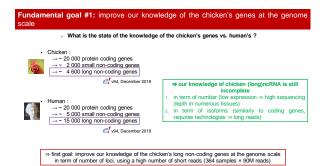


An atlas of chicken long non-coding RNAs gathering multiple sources: gene models and expression across more than twenty tissues

Frédéric Jehl*, Kévin Muret*, Maria Bernard*, Diane Esquerré, Hervé Acloque, Elisabetta Giuffra, Sarah Djebali, Sylvain Foissac, Thomas Derrien, Tatiana Zerjal, Christophe Klopp⁵ and Sandrine Lagarrigue⁵

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Context: IncRNAs, from cellular processes to complex traits variation

- Long non-coding RNAs (IncRNAs) transcripts longer than 200 nucleotides that are not translated into proteins
- IncRNAs have been showed to act on numerous cellular processes:
 chromatin compaction (Xist)

 - Chlomatin Comparison (Ass)
 gene expression :
 → transcription (*Fendir*, Grote et al., 2013)
 → translation (*lincRNA-p21*, Yoon et al., 2012)
- Influence on complex traits & diseases ?
 General agreement that complex traits and diseases are influenced by numerous loci = 90 % out of coding regions (Manilo *et al.*, 2009)
 These loci are located in regulatory regions (Maurano *et al.*, 2012)

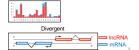
 - ⇒ effect on genes expression
 Some of them affect IncRNA expression (Kumar et al., 2013)
- Numerous, poor functional annotation: no function prediction rules (yet) lowly expressed (10-fold less than mRNAs): difficult to validate experimentally

Fundamental goal #2: annotate IncRNAs thanks to their expression profile in chicken focus on tissue-specificity

- Problematic: knowing the existence of a gene (coding or non-coding) doesn't inform on its function(s)
- with coding genes:
 prediction of the associated amino-acid sequence,
 prediction of the protein function by motif conservation across species.
- with long non-coding genes;
- poor sequence conservation, poor / no domains knowledge.
- ⇒ IncRNA are poorly annotated

 \Rightarrow second goal: annotate the lncRNAs applying the two following strategies

- Strategies to annotate IncRNAs function: expression profile: IncRNA expressed in tissue A and little or not in tissue B (= specific to tissue A) might have a role related to tissue A function
 - study their configuration with closest coding gene: e.g., close and divergent ⇒ common regulation ?



Context: the chicken (G. gallus), a specie of great economical and social important



1. Improvement of our knowledge of chicken's genes at the genome scale



- 90 millions reads per sample
 stranded reads
 2 × 150 pb paired-ends

1. Results of gene identification

Methods

- Methods Mapping on G. gallus reference genome (GalGal5) with STAR (Dobin *et al.*, 201 using Ensembl V94 annotation Transcriptome reconstruction using Cufflink2 (Trapnell *et al.*, 2013) Elimination of modelized genes overlapping on the same strand any Ensembl V94 ger IncRNA prediction using FEELICe (Wucher *et al.*, 2017)

Results : addition of 25 214 genes to the 24 881 Ensembl genes	isinna e	
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⇒ Total of 50 095 genes

	of our pipeline
13),	RNA-seq data (.fastq)
13),	
	Mapping (STAR)
ene	Mapped reads (.bam)
	Modelization (Cufflink2)
	Gene models (.gtf)

IncRNA predic (FEELnc) diction

1. Selection of the most robust modeled genes based on their expression pattern

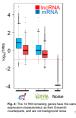
Reminder : lncRNA = lowly expressed (1/10th mRNA). Necessity to separate background noise from real expression

To improve reliability of the 25 214 newly modelized genes, we selected models according to their expression,

to ingore is inc____ ingo 2 expression criteria: • an normalized expression metric (TPM > 0.1) • number of reads supporting the model ⇒ selection of 58.5% of the 25 214 genes (14 760 models)

Comparison with background noise: • "No genes" = set of genomic regions out of the Ensembl and our genes models: our models have expression lavels supported to the background price

Comparison with mRNAs and Ensembl genes: than mRNAs



1. Selection of the most robust modeled genes based on their expression pattern

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INRA genes + Ensembl genes = **14 760** + 24 881 ⇒ **39 641 genes**

Gene type	Number of genes	% of total	Ensembl
IncRNAs	13 009	88.14%	4 641
mRNAs	1 199	8.12%	18 346
Others	552	3.74%	1 894
TOTAL	14 760	100%	24 881

1. Sequential building of an extended annotation using 6 external data source

- From this set of 14 760 INRA genes (88% IncRNA) + 24 881 Ensembl genes, extension of the chicken IncRNA annotation using external sources: 1. NONCOE (Fang et al., 2016) : 9 322 IncRNAs 2. ADB (I of al., 2015) : 5728 IncRNAs

- 3. ALDB (Li et al., 2015) : 5 752 INCKINAS 4. FR-AGENCODE (Foissac et al., submitted) : 6 089 IncRNAs

For each source, we kept only the loci with no overlap with the extended annotation:

Existing catalogue	A
External source (e.g. NONCODE, NCBI,)	B
(e.g. NUNCODE, NOBI,)	↓ overlap
Result	Ab

As a result, we increased gene numbers from 14 760 + 24 881 to →→→→ 52 075 genes including 30 084 incRNAs : ×6.5 compared to Ensembla alone 19 345 mRNAs : ×1.1 compared to Ensembla alone

This work was done at the locus level since isoforms are poorly known and expression analysis are done at the gene level

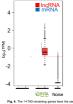
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Fig. 4: express

2. Annotation of the IncRNAs thanks to their expression profile in chicken, focus on tissue-specificity

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Thanks to the previous step : extended IncRNA catalogue (from 4 640 to 30 084) role or function of the IncRNAs ?

objective of this second part: provide an annotation of the potential IncRNA functions using their expression profiles

2. Annotation of the IncRNAs thanks to their expression profile in chicken, focus or tissue-specificity

Hypothesis: if a gene is expressed in one or a few tissues, its function is likely to be related to the function of the tissue(s)

To study tissue-specificity, necessity to have numerous tissues ⇒ public data from other teams.

3 datasets were analyzed:

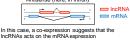
- INRA : 5 tissues (Jehl *et al*, in preparation) Sishuan University : 6 tissues (Tang *et al.*, 2017) Roslin Institute : 21 tissues (used by Ensembl for the chicken genome annotation)

▲ IncRNAs are lowly expressed ⇒ sensitivity to genotype, physiological status, experimental conditions, ...

Preliminary question : can these 3 datasets be studied together after expression normalization, or is there a batch

2. IncRNA classification using FEELnc

Long non-coding genes are classified by position relatively to the nearest coding genes using FEELnc (Wucher et al., 2017) • some configurations suggest potential (co)-regulations Examples Divergent Antisense (here, in intron) ;_____ **;___**^ In this case, if the genes are close, a co-expression suggests a common regulation, and therefore a common function

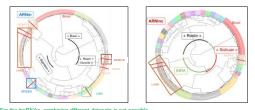


In both case, we infer the function of the IncRNA using the function of the mRNA

	Divergent	Antisense of intron
In the extended catalogue	4 895	2 105
with τ ≥ 0.95	977	418

2. Datasets comparisons

The mRNAs expression clusters the sample based on tissues ar The IncRNAs expression clusters the sample mainly by projects ession clusters the sample based on tissues and functions



⇒ For the IncRNAs, combining different datasets is not pr ⇒ focus on the 21 tissues from the Roslin Institute data

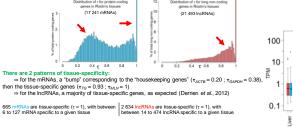
- We provide a large catalogue of chicken lncRNAs at the gene level from 4 640 (Ensembl v94) to 30 084 lncRNAs
- We also provided a rough annotation of all these genes, based on : their expression pattern across 21 chicken tissues their position relative to the nearest coding gene

⇒ this will allow the scientific community to work on different scientific problematics related to IncRNAs in chicken

Perspective: • we will use these data to study the genetic component of feed efficiency in layer chicken ⇒ feed ≈ 60% of production cost in monogastrics ⇒ reduction of feed vs. food competition ⇒ lower environmental impact

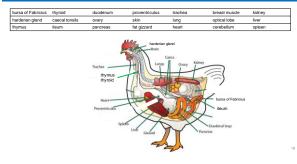


2. IncRNAs and mRNAs have different tissue-specificity patterns Use of the tissue specificity index τ , which ranges from 0 (same expression in all tissues) to 1 (expression in one tissue only) (Yanai et al., 2005)





Tissues present in Roslin Institute data



IncRNA	IncRNA
normalization using all genes	normalization using IncRNA only
ANNIC Regins C Statuurs	Roslin

1. Selection of the most robust modeled genes based on their expression pattern

Reminder : lncRNA = lowly expressed (1/10th mRNA). Necessity to separate background noise from real expression.

To improve reliability of the 25 214 newly modelized genes, we selected models using two criteria: an expression metric (TPM) number of supporting reads

Additional criterion

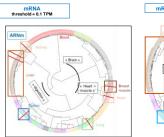
5 paired-reads or more in 25% of the samples of a tissue: at least 75 supporting reads.
 ⇒ leaves 14 760 modelized genes from the 25 214 (= 58.5%).

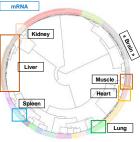
Number of IncRNAs and mRNAs in the remaining catalogue :

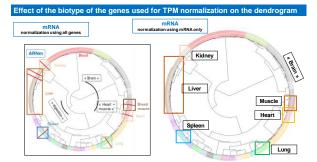


INRA genes + Ensembl genes = 14 760 + 24 881 = **39 641 genes**

Effects of the expression threshold chosen on the dendrogram : 1 TPM

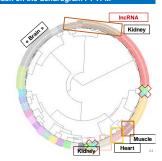






Effects of the expression threshold chosen on the dendrogram : 1 TPM



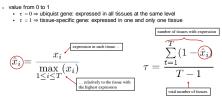


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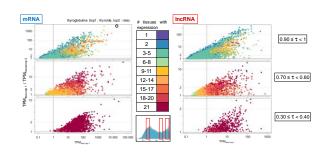
A tissue-specificity metrics: the tau (τ)

To study tissue-specificity, we used the tau metrics from Yanai et al., 2005:

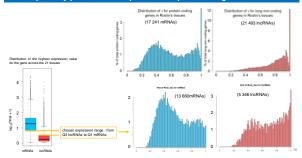
- associates a value (r) to each gene
- associates a value (r) to each gene of reach gene, it accounts for :
 the expression in each tissue, relatively to the tissue with the highest expression
 the number of tissues with expression the number of tissues with expression
 the total number of tissues



Gene expression at different τ values



Tissue-specificity patterns are independant of expression ranges

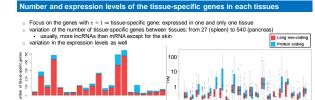


IncRNA classification using FEELnc

Examples

Long non-coding genes are classified relatively to the nearest coding genes using FEELnc (Wucher *et al.*, 2017) ⇒ some configurations suggest potential (co)-regulations

	Examples:						
		Divergent		Antisense (here	, in intron)		
	7		<u>,</u>			mrna mrna	
Туре					Total	Total classified	Number of genes
Position same strand divergent	distar ≤ 1kb	> 1kb	Total				
g	same strand	1 403	8 663	10 066	18 204		
te	divergent	1 476	3 419	4 895			
-	convergent	584	2 659	3 243		23 129	30 08
Genic	Position	direct sense	ion antisense	Total	4 925		30 00
Ger	exonic	2	2 494	2 496	4 925		
5	intronic	324	2 105	2 429			
Unclassified					6 955		



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Bursa of Fal Caecal tonsi Cerebellur Duodanum Fat gozzand Hand, gand Broast 1 048 mRNAs are tissue-specific (0.95 ≤ t ≤ 0.99), with between 2 788 IncRNAs are tissue-specific (0.95 ≤ t ≤ 0.99), with between 5 to 248 mRNA specific to a given tissue 17 to 589 IncRNA specific to a given tissue 665 mRNAs are tissue-specific (r = 1), with between 6 to 127 2 634 IncRNAs are tissue-specific (r = 1), with between 14 to 474 mRNA specific to a given tissue

tand teart ideurn idrey Liver Lung nuscle orary Ovary Skin Spleen Thymus Thyroid