

The GEMO project: Hitchhiking DNA in Magnaporthe oryzae

Ludovic Mallet, Cyprien Guerin Guérin, Joelle J. Amselem, Elisabeth E. Fournier, Helene H. Chiapello

► To cite this version:

Ludovic Mallet, Cyprien Guerin Guérin, Joelle J. Amselem, Elisabeth E. Fournier, Helene H. Chiapello. The GEMO project: Hitchhiking DNA in Magnaporthe oryzae. JOBIM 2015. Meeting of working Group Medicago sativa, Jul 2015, Clermont-Ferrand, France. , 1123 p., 2015. hal-02801084

HAL Id: hal-02801084 https://hal.inrae.fr/hal-02801084

Submitted on 5 Jun2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



The GEMO project: Hitchhiking DNA in Magnaporthe oryzae

MALLET L.^{1,2}, GUÉRIN C.², AMSELEM J.^{3,4}, FOURNIER E.⁵ and CHIAPELLO H.⁶

¹ Evolutionary Bioinformatics University of Münster, DE ² INRA, MalAGE Jouy-en-Josas, FR ³ INRA, URGI Versailles, FR ⁴ INRA, UMR BIOGER Thiverval-Grignon, FR ⁵ INRA, UMR BGPI Montpellier, FR

⁶INRA, MIAT Castanet-Tolosan, FR

Abstract

ludovic.mallet@wwu.de

The Evolutionary Genomics of *Magnaporthe oryzae* (GEMO) project is an attempt to identify the genomic determinants and evolutionary events involved in pathogenesis, host specificity and adaptation. Ten closely related genomes of *M.oryzae* strains and one of the sister species *M. grisea* with different main host and host range were sequenced and analysed [1].

We focused here on the detection and analysis of potentially **horizontally acquired DNA regions** that we characterized using compositional methods.

We examined the general content, the **functional profiles** and the **potential donors** of the putative acquired genes. We reviewed the acquired regions and investigated a few target candidates for pathogenicity.

Methods

Horizontal Transfer detection [2]

- Tetranucleotide composition in sliding windows, 5kb long, 100bp step
- Kullback-Leibler divergence of composition: windows vs whole genome
- Parametric determination of threshold

——— Horizontal Transfer filtering

- To avoid false positive detection from known repeats, we filtered out:
- Transposable elements, annotated with REPET [3].
- rRNA, annotated with REPET base and blast.
- Homopolymers >100bp, Nannotator, PC
- Simple repeats, annotated with RepeatMasker [4]

Potential origin identification

- Inference with GOHTAM [1], a database of species composition
- Neighbor species with a distance over 160 arbitrary units were discarded

Functional profile

- Functional annotation was predicted with InterProScan [5]
- Analysis and presentation with DAVID [6], REVIGO [7] and Genes2WordCloud [8]

Potential origin

 Class
 # of regions
 Species

 Virus
 3
 Corilla gorilla cytomegalovirus 2.1 (x2) ; Simian adenovirus 18

 Fungi
 6
 Humicola grisea ; Colletotrichum higginsanum (x4) ; Chrysosporium Lucknowense

 Arthropods
 3
 Drosophila parabipectinata ; Lychas mucronatus; Richardia teevani peniculus; Cruoriaceae sp.

 Alguae
 3
 Pyropia yezoensis ; Acetabularia peniculus; Cruoriaceae sp.

 Choanof lagellate
 2
 Monosiga ovata

Table 2: Most credible donors

Figure 4: Potential donors

Candidate(s)

The putative transfered region is in red.

The composition of this region is very similar to the whole genome composition of another fungi: *Colletotrichum higginsianum*, *a* plant pathogen.

The gene is a repeat assemlby of 3 protein domains.

The condensation domain is found in many enzymes which synthesise peptide antibiotics. It catalyses a reaction to form peptide bonds in **non-ribosomal peptide biosynthesis**. We hypotesise an NRPS activity











Table 1 : Genomes and statistics

Genomes	70-15	† BR29	BR32	CD156	FR13	GY11	TH12	PH14	TH16	US71
Main host	rice	rice	wheat	Eleusine	rice	rice	rice	rice	rice	Setaria
Size (Mb)	40.9	40.9	41.9	42.7	43.1	46.3	48.5	49.8	39.1	41.2
Predicted genes	12,827	12,616	14,781	14,415	15,035	20,621	19,811	20,067	13,725	14,013
HT DNA (Mb)	1.22	1.23	2.59	1.55	0.14	0.37	0.67	0.52	0.47	1.26
HT DNA %	2.97	3.0	6.18	3.62	0.32	0.79	1.38	1.05	1.19	3.06
HT Genes	502	374	948	438	41	229	252	175	176	511
\pm . The reference strain M or 70-15 is sequenced with Sanger technology \pm RP29 is a strain from the M origon species										

** The reference strain M.o. 70-15 is sequenced with Sanger technology. * BR29 is a strain from the M. grised species



Figure 3: GO enrichment



References

Chiapello *et al.*, 2015, GBE, in revision
 Ménigaud *et al.*, 2012, BMC Bioinfo.
 Flutre *et al.*, 2011, Plos One
 Flutre *et al.*, 2011, Plos One
 Stobnov *et al.*, 2001, Bioinformatics
 Huang *et al.*, 2009, Nature Protoc.
 Supek *et al.*, 2011, PloS One
 Baroukh *et al.*, 2011, Source Code Biol Med.

Conclusion & perspectives

The genomes of *Magnaporthe oryzae* contain between 0.14 (0.3%) to 3.59 (6.2%) of putative horizontal transfers, likely coming from **various kingdoms**. The functional profiling suggests their implication in various processes, including **pathogenicity**, **regulation and cell motion**. Early analyses yielded interesting candidates but further work is required for other functional classes. The strain-specific transfers will be analysed in the future to assess their implication in the variable host range of the strains. These investigations are expected to enhance the understanding of the evolutionary origins of the features driving pathogenicity phenotype and host specificity.

Acknowledgments

Funding: **ANR** 2009-GENM-029-01. Travel grant from the **SFBI**, many many thanks! Special thanks to Sophie Schbath and Philippe Leroy.