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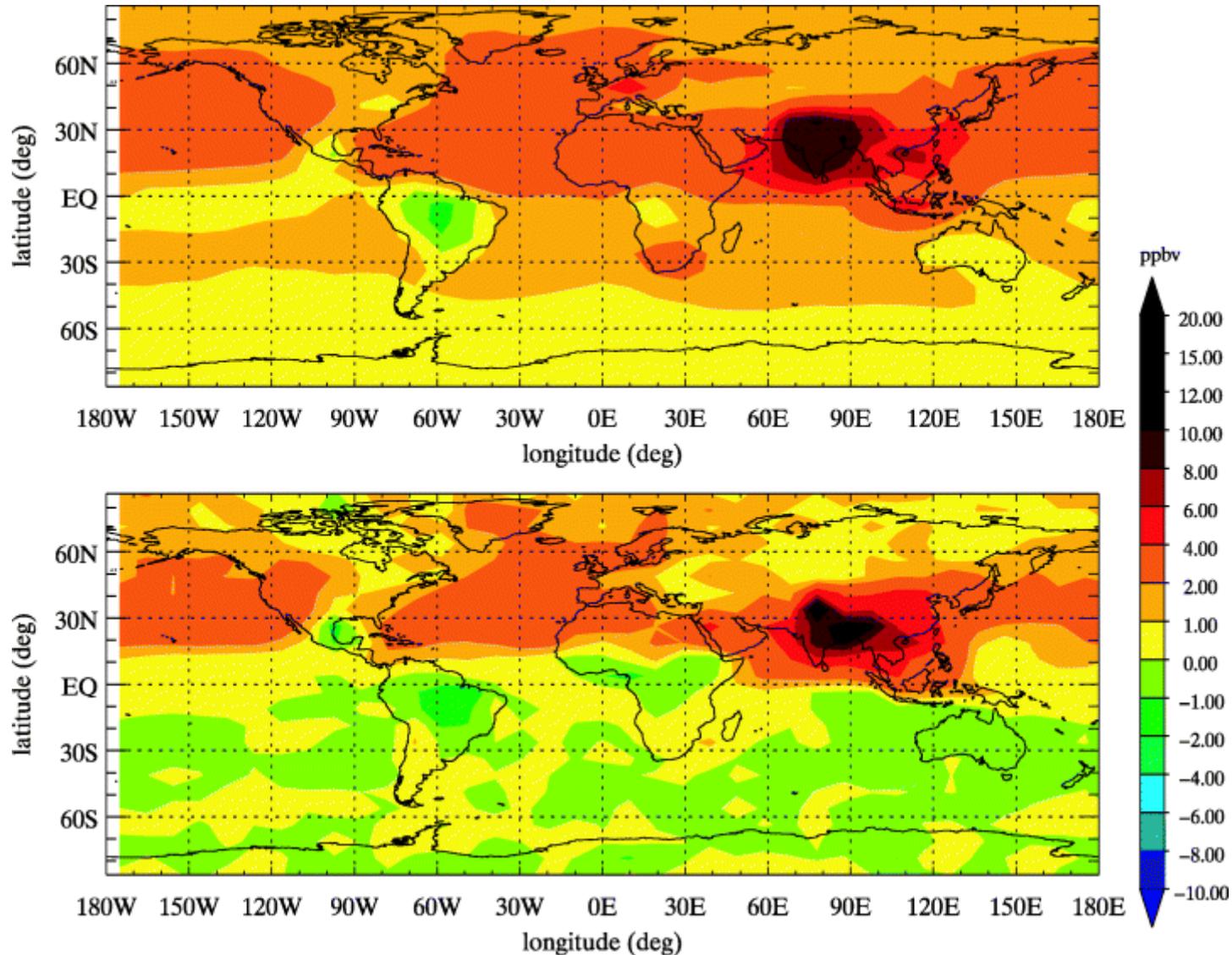


Improving the determination of the effective ozone flux in tree leaves for issuing a sub-model to be integrated in models predicting ozone risks to forest ecosystems

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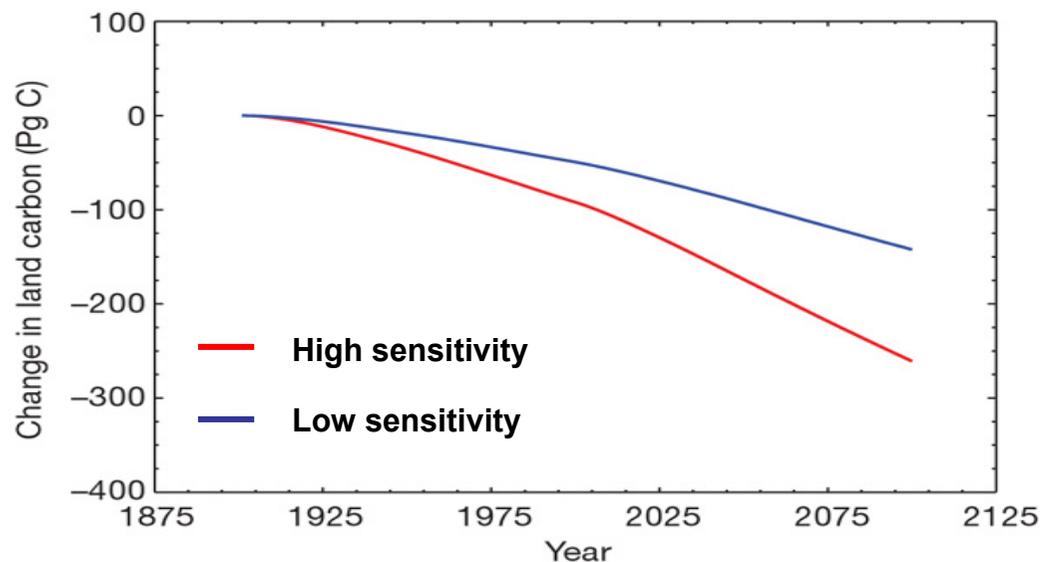


The concentration of OZONE, greenhouse gas, is expected to increase in the near future



Predicted differences in decadal annual mean surface ozone concentrations from the 1990s to the 2020s, for two global chemistry-transport models, under a 'Current Legislation' scenario. The upper diagram presents predictions for the TM3 model and the lower diagram presents predictions for the STOCHEM model. This figure is reproduced from fig. 11(a) of Dentener *et al.* (2005).

Ozone is linked to temporal changes in land carbon storage

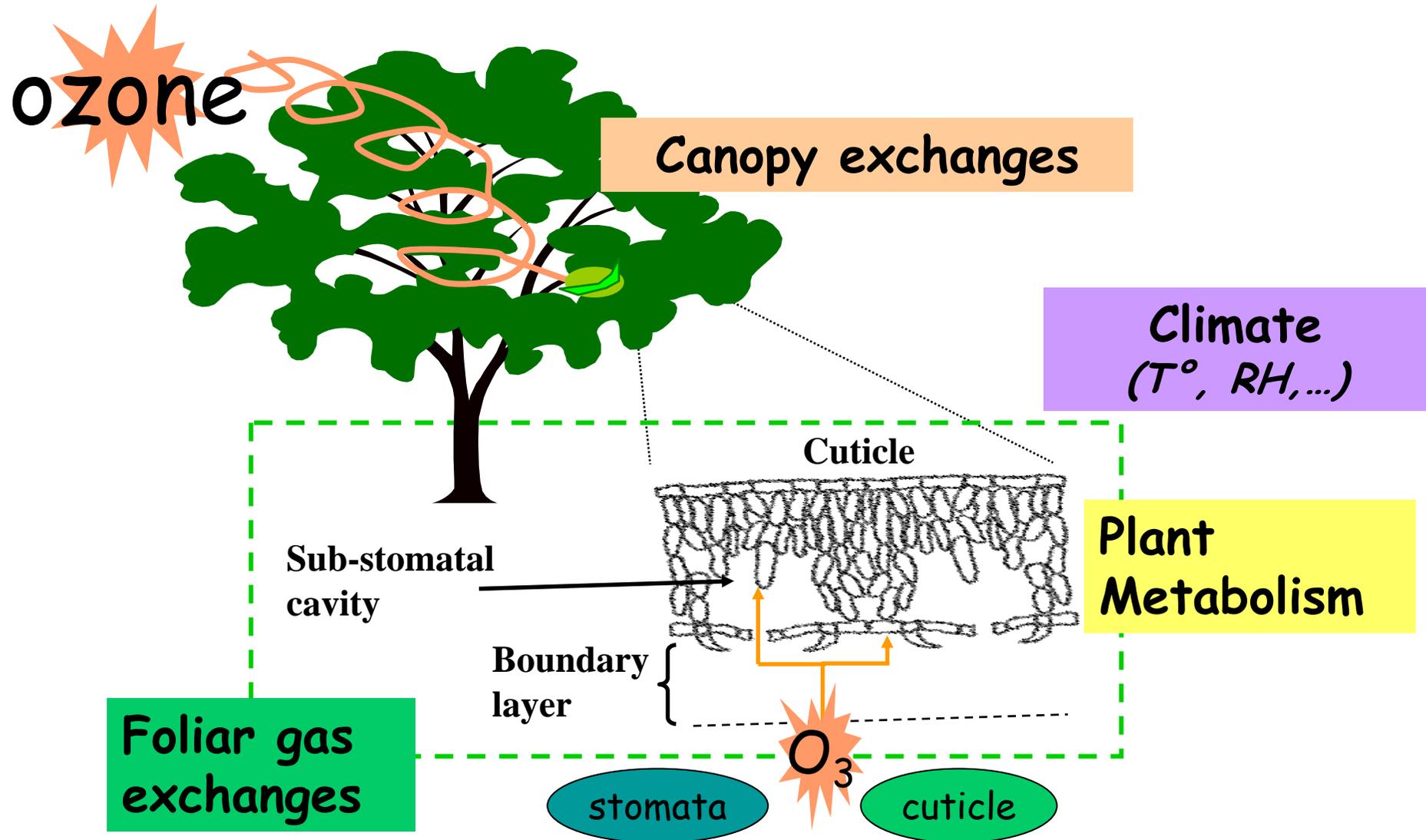


Simulated changes in land carbon storage due to O₃ increase
Sitch et al., 2007 *Nature*, 448, 791-95

Increasing ozone concentration in the atmosphere diminishes plant productivity

Ozone will lower the capacity of plants to sequester the increasing level of carbon (rise in atmospheric CO₂)

Exchanges of ozone with vegetation



Objectives :

Interlocking / integration of different models to assess risks for the coming 20 years

Needing more knowledge on :

- ozone deposit
- **cellular and foliar impact of ozone**
- impact on the yield in crop field and forest stand
- scenarii of evolution over 20 years
- socioeconomic recommendation

Indices of ozone exposure

AOT40 (accumulated exposure over 40 ppb):

$$\text{AOT40} = \sum_d \cdot \sum_h ([\text{O}_3]_h - 40)$$

Critical level over a vegetation period: -forest trees: 10 ppm.h
-crops: 3 ppm h

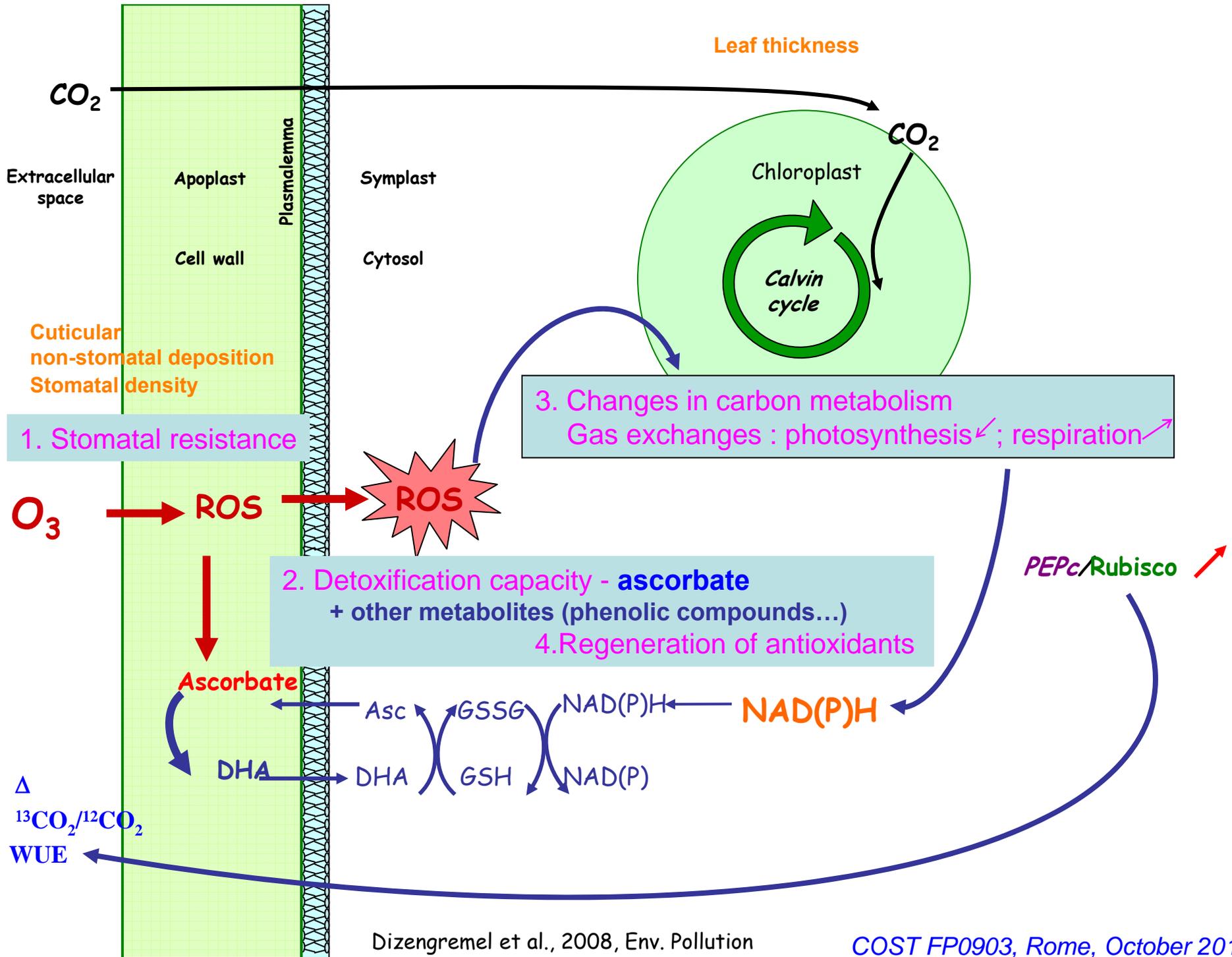
CUO (cumulative uptake of ozone) or POD

$$\text{CUO} = \sum_d \cdot \sum_h \left(F_{\text{O}_3} \times 3600 \right)$$

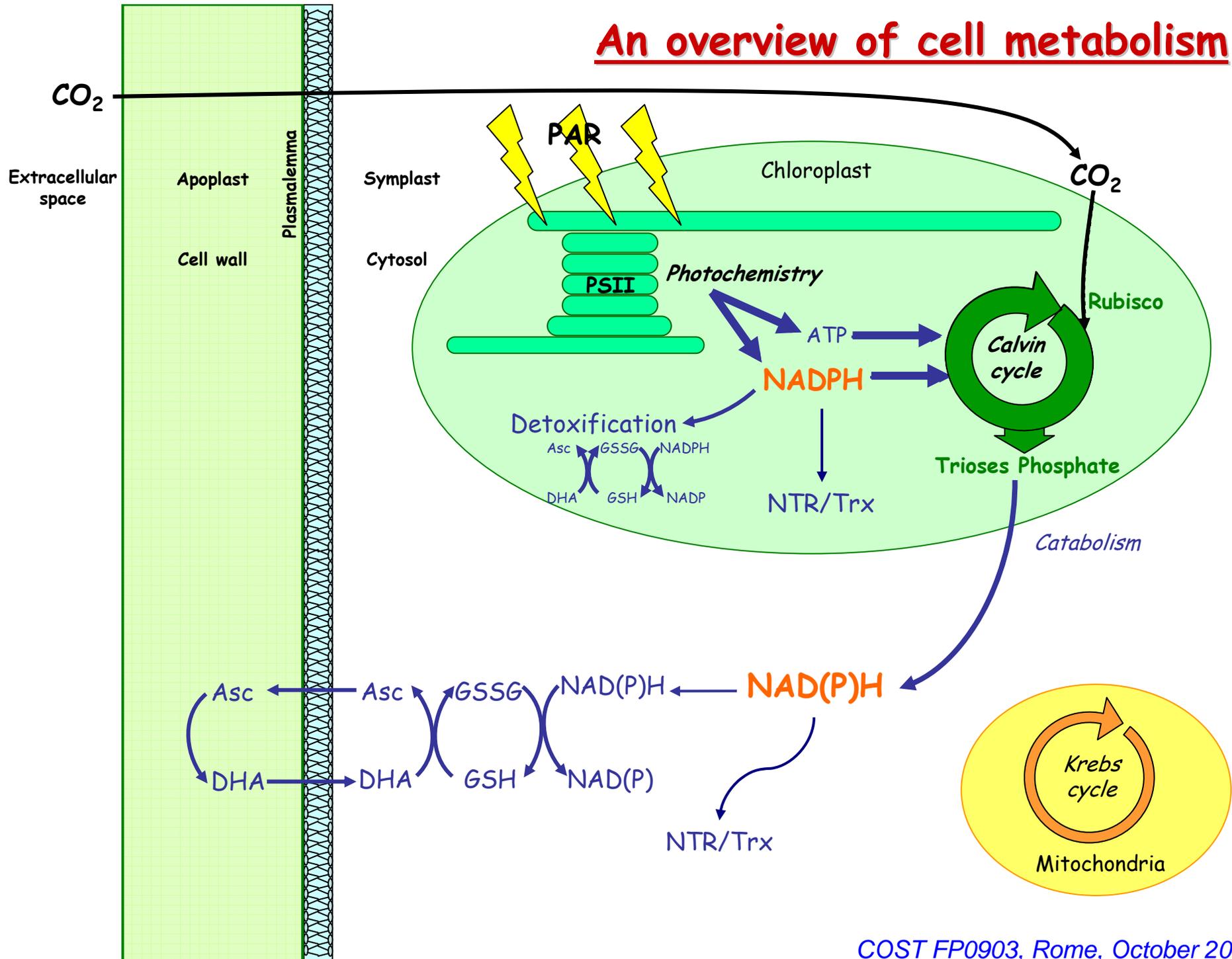
Karlsson *et al.* (2007)
Pleijel *et al.* (2007)

Instantaneous ozone flux: $F_{\text{O}_3} = [\text{O}_3] \times \frac{g_s}{16}$

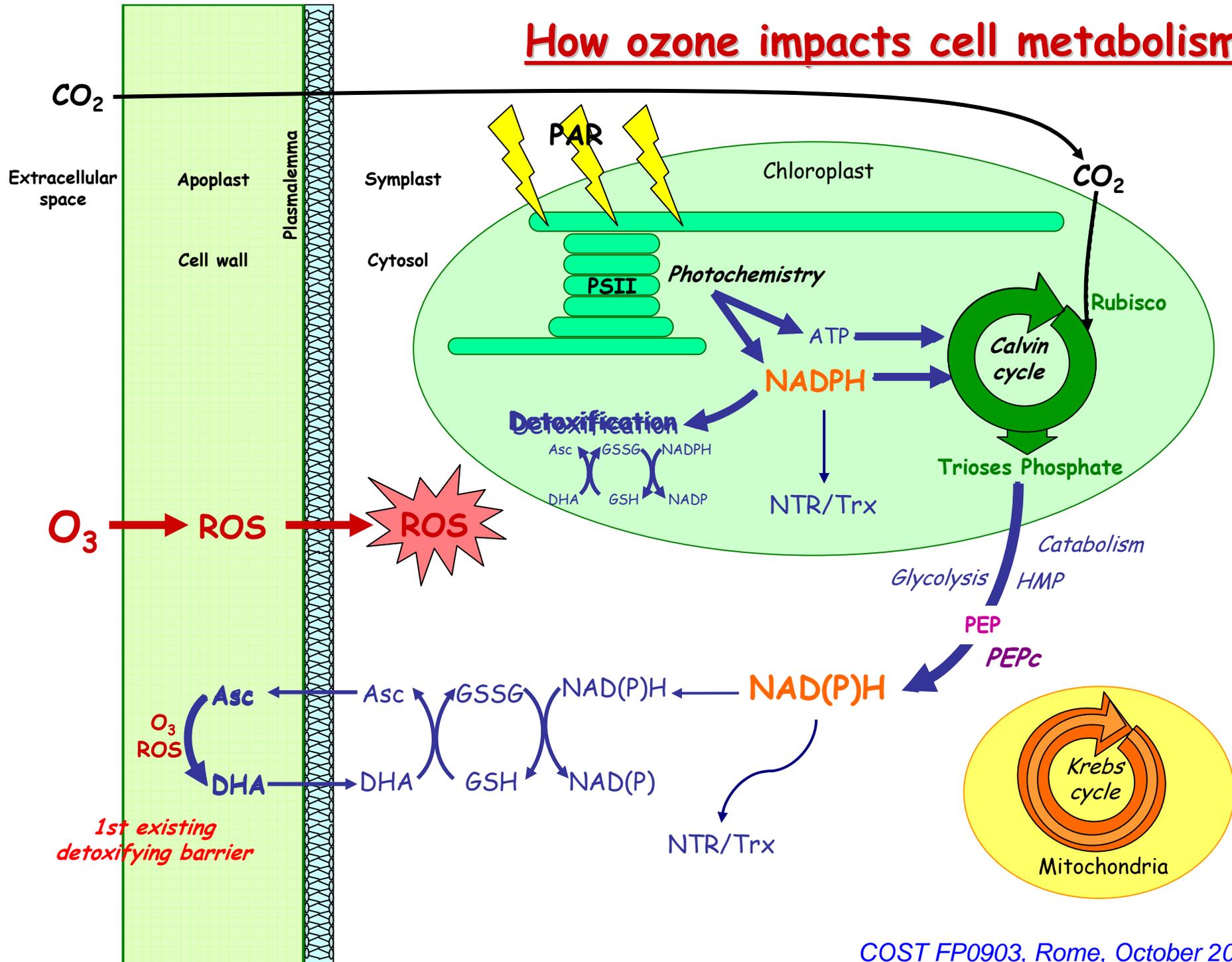
- Towards an effective ozone flux
- Detoxification/Metabolism must be taken into account



An overview of cell metabolism



How ozone impacts cell metabolism



Effect of ozone on carboxylation

Rubisco & PEPc

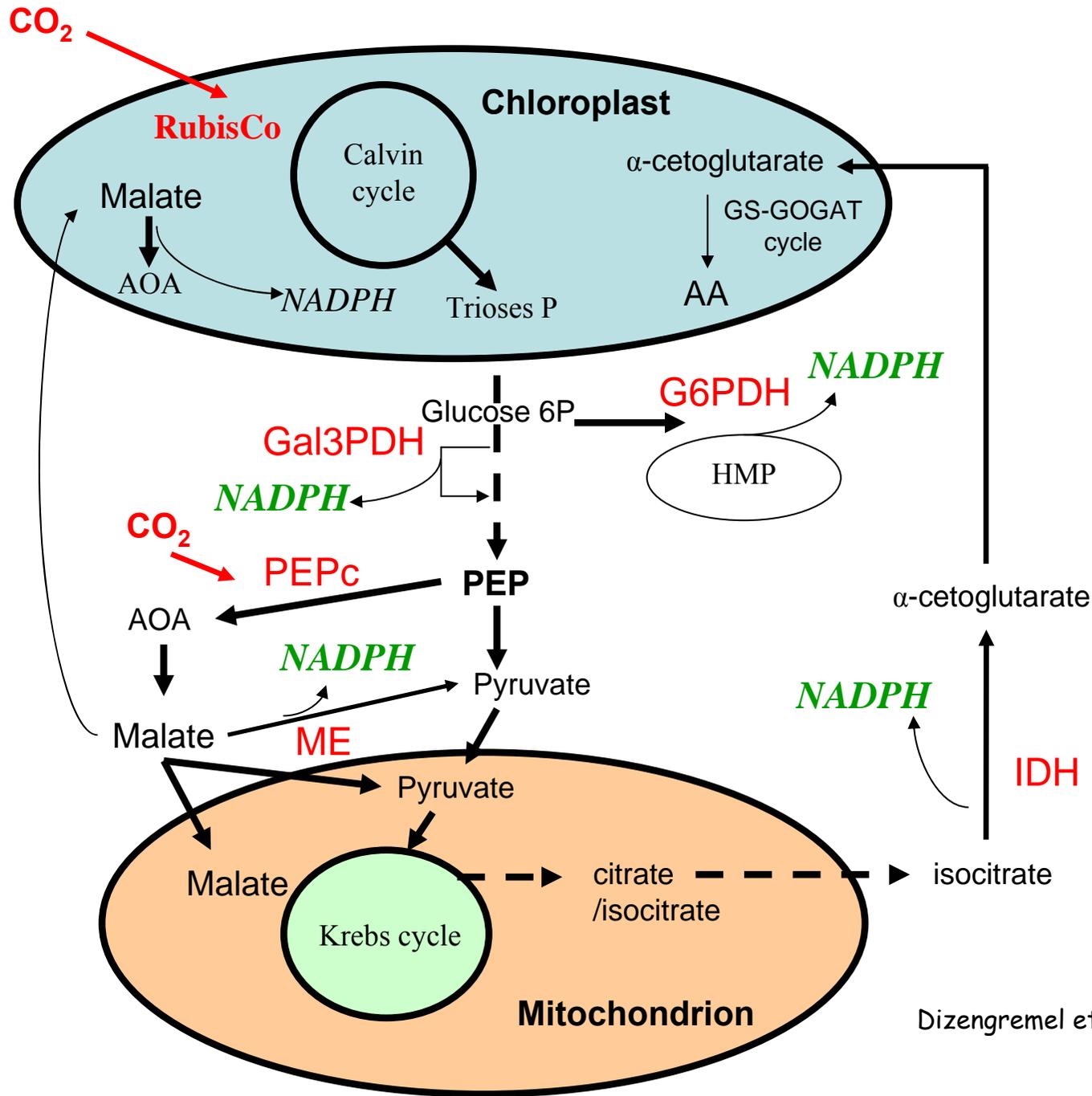
Specific activity (nkat.mg _{prot} ⁻¹)	Rubisco		PEPc		Rubisco / PEPc ratio	
	C	O ₃	C	O ₃	C	O ₃
Poplar 60 ppb O ₃ , 2 weeks	17.0	9.1	0.36	1.35	47.2	6.74
Norway spruce 200 ppb O ₃ , 12 weeks	13.2	6.2	0.45	2.55	29.3	2.43
Aleppo pine 200 ppb O ₃ , 5 weeks	13.6	7.5	0.55	1.96	24.7	3.82

Fontaine *et al.* (1999), *Physiologia Plantarum*

Pelloux *et al.* (2001), *Plant, Cell & Environment*

Dizengremel (2001), *Plant Physiol. Biochem.*

Fontaine *et al.* (2003), *Physiologia Plantarum*



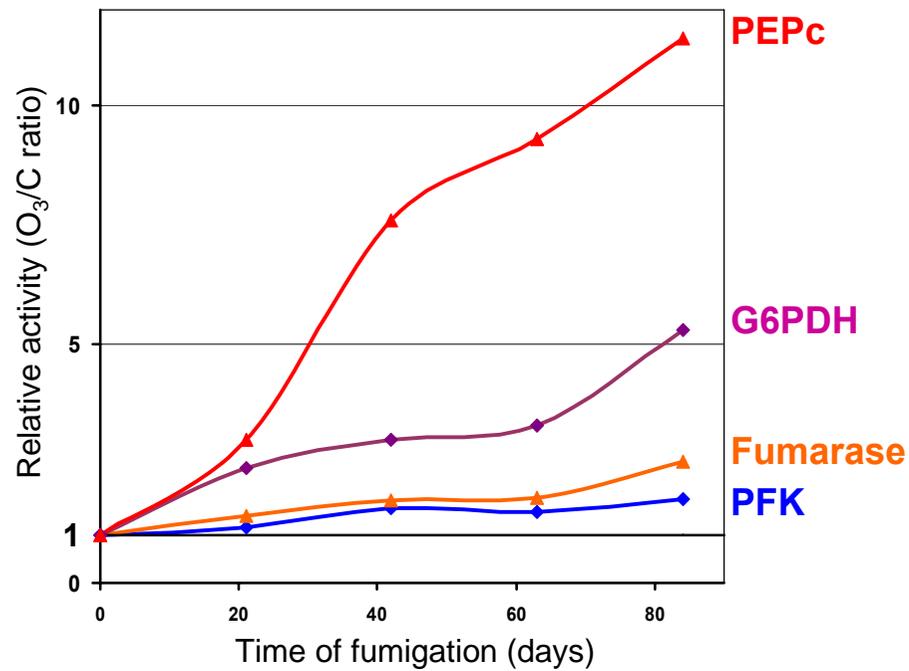
Dizengremel et al., 2009, Plant Biology

Effect of ozone on enzymes linked to catabolism

Glycolysis, Krebs cycle and anaplerotic pathway

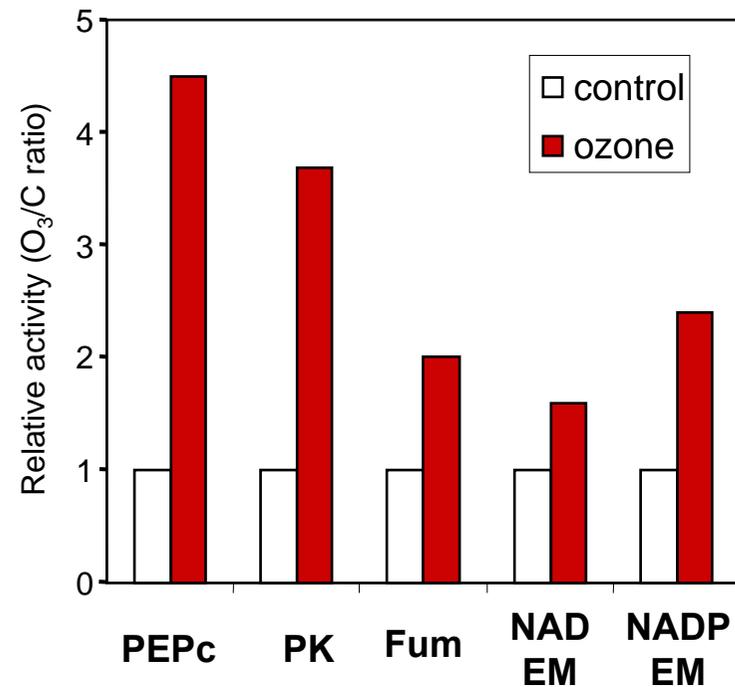
Norway spruce

200 ppb O₃, 12 weeks



Poplar

100 ppb O₃, 24 days



Sehmer *et al.* (1998), *Physiologia Plantarum*;

Dizengremel *et al.* (2009), *Plant Biology*

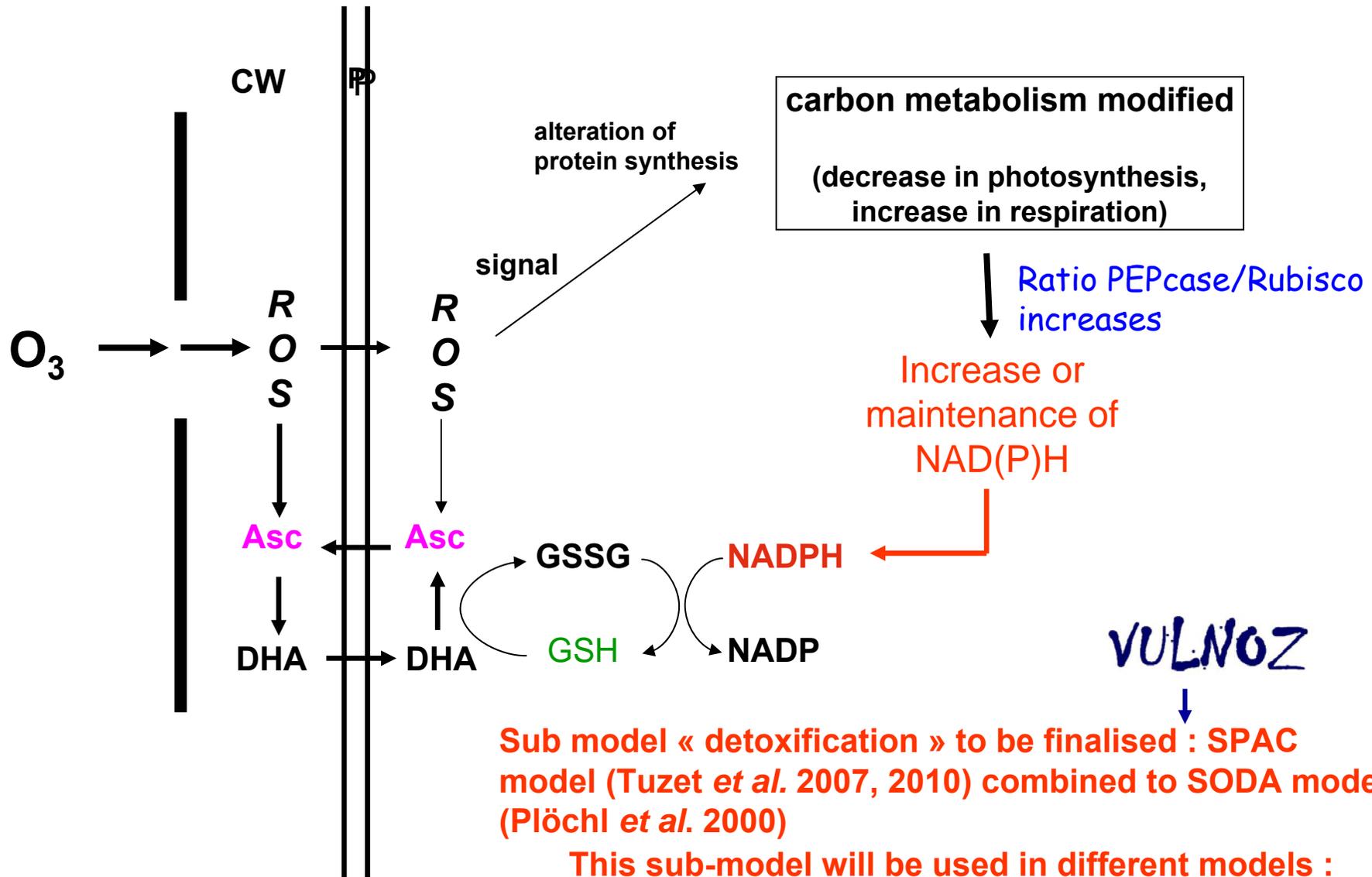
Which metabolites/enzymes are relevant to measure?

Ascorbate
Ascorbate peroxydases - total Asc (symplastic + apoplastic) could be a good indicator of ozone resistance.

Glutathione
Glutathione Reductase - this metabolite acts to reduce Asc through glutathione reductase activity.

Reducing power
(NADPH & NADH) - these nucleotides are the driving force of the detoxifying system (for regeneration of ascorbate pool).

It is necessary to confirm that ozone stimulates the activity of the cytosolic enzymes providing NADPH.



Sub model « detoxification » to be finalised : SPAC model (Tuzet *et al.* 2007, 2010) combined to SODA model (Plöchl *et al.* 2000)

This sub-model will be used in different models :
CERES (yield/field) – **ORCHIDEE** (regional scale)
 and **AROPAj** (economic risks)

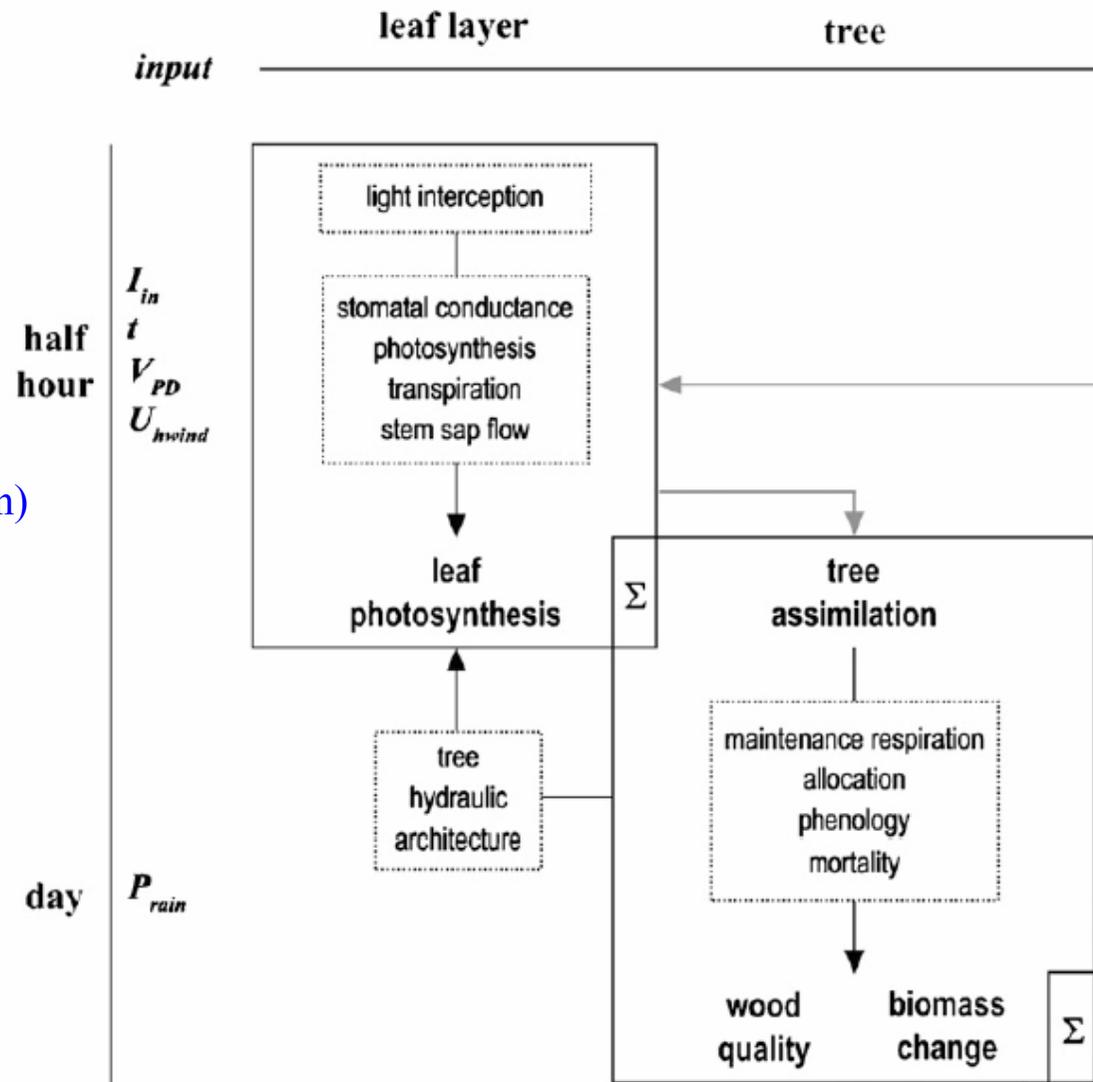
MODELING AND RISK UPSCALING

which models are available? example...

ANAFORE

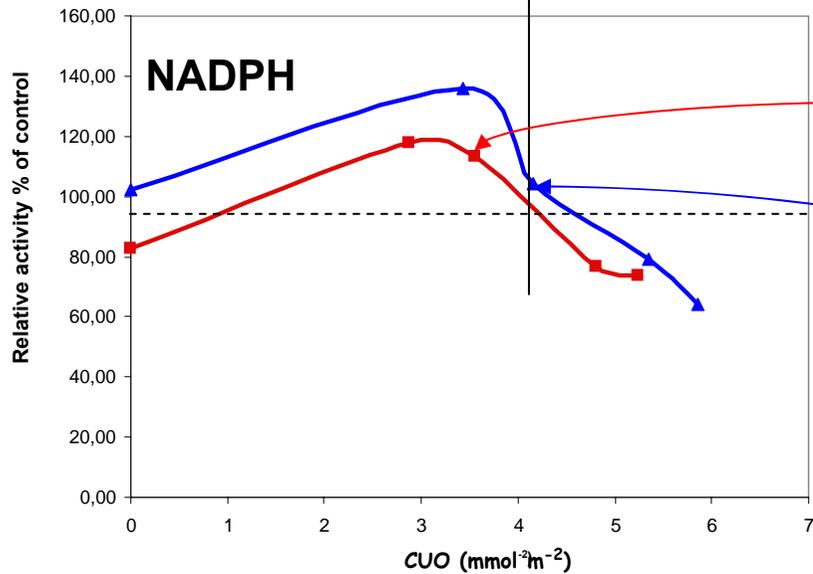
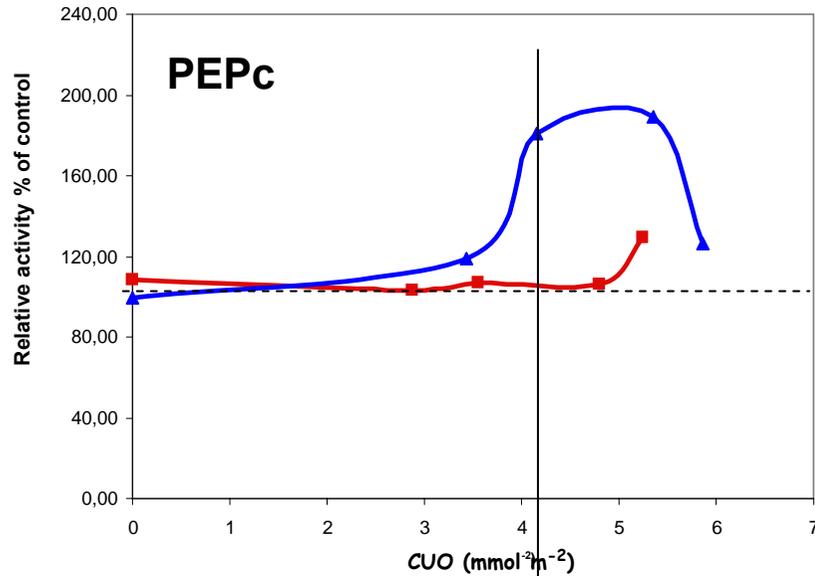
Deckmyn *et al.*, 2008

- New modules? (detoxification)
- New parameters :
 - PEPc/RubisCo
 - ascorbate , NADPH, ...
 - stomatal density...
- new parameterization



..... *Some preliminary results*

▲ Soissons ■ Caphorn



Winter wheat (*Triticum aestivum*) cultivars :

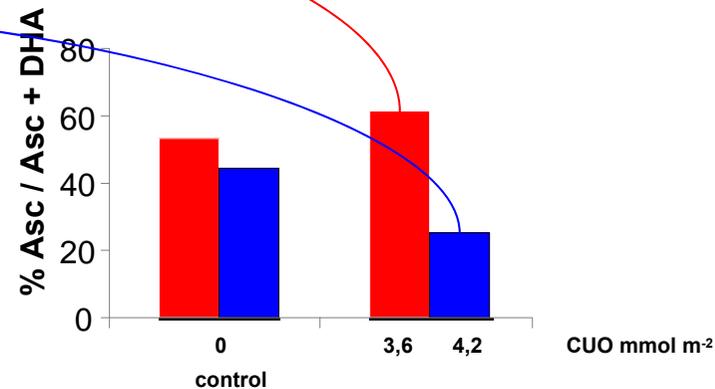
Soissons (sensitive)

Caphorn (tolerant)

Higher stimulation of PEPc activity in the sensitive cultivar

Higher content of NADPH as a response to ozone fumigation

Partial uncoupling between NADPH content and PEPc activity changes :
Role of NADPH generating enzymes ?





Poplar Experiment in phytotronic chambers

- 3 genotypes *Pop. deltoides* × *Pop. nigra* (Carpaccio, Cima, Robusta).
- Ozone 120ppb



Parameters :

Gas exchanges + Chlorophyll
Growth + Leaf surface
Ascorbate + glutathione
NADP(H) + NADPH generating enzymes
PEPc + RubisCo
Guard cells transcriptomic
Metabolomic



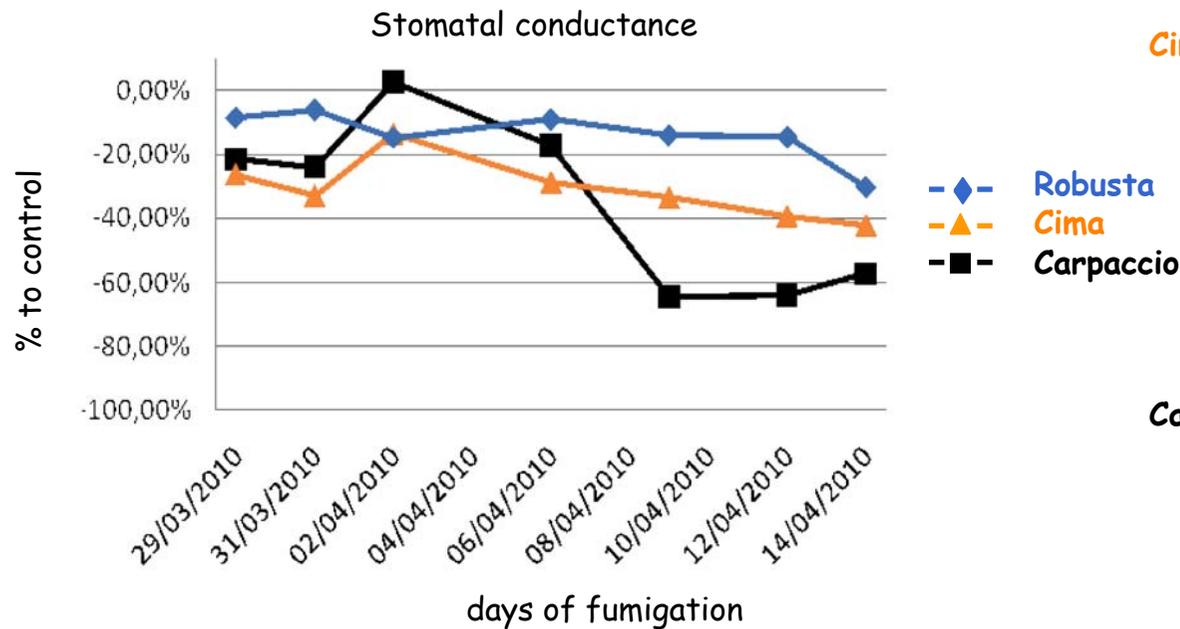
Robusta (sensitive +++)



Cima (tolerant)



Carpaccio (tolerant)



The ozone group at Nancy University



• Primary carbon metabolism

Photosynthesis

Didier Le Thiec, INRA Nancy

Assimilation / Stomatal conductance

Jennifer Dumont, PhD

Leaf anatomy



Photochemistry / Calvin cycle: Sacha Bohler PhD

Respiration

Respiratory chain Yves Jolivet



E. Oksanen

Photorespiration Marie-Paule Sauder

Marie-Noëlle Vaultier



Centre de Recherche Public
Gabriel Lippmann

J.-F. Hausman

J. Renaud

Detoxification Ata Dghim, PhD



Umeå Plant Science Centre

P. Gardeström

Comparison of C3/C4 plants

• Cell wall

Cellulose: Dany Afif

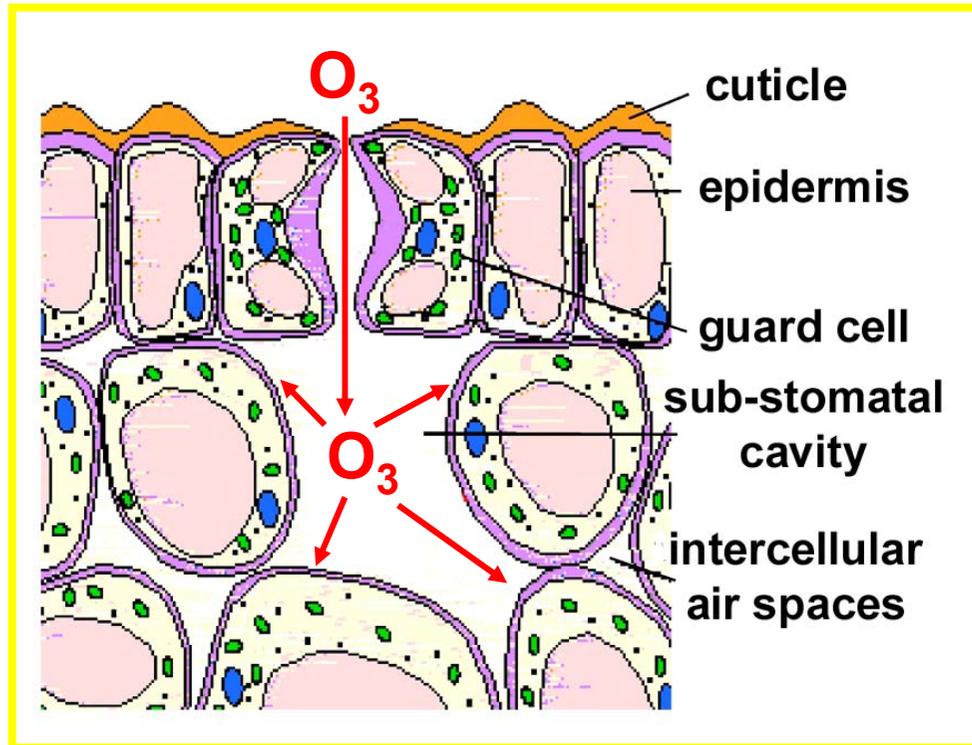
Lignins: Mireille Cabané

Nicolas Richet, PhD



Catherine Lapierre, INRA Grignon

Towards a model for Ozone Risk Assessment : from atmosphere to cell



The sensitivity of a leaf to ozone lies on the *stomatal conductance*, on the *existing scavenging system* and the *detoxifying capacity (regeneration)*.

The aim is to determine a sub-cellular model taking into account these aspects.

This model will have to be further integrated into a plant model and in a stand model.

The ultimate aim is to improve ozone risk assessment by the determination of more accurate critical levels of exposure than those currently used by EU.

Increasing tolerance to ozone by elevating foliar ascorbic acid confers greater protection against ozone than increasing avoidance (Chen and Gallie, 2005, *Plant Physiology*, 138, 1673-1689)

Increasing the level of ascorbate through enhanced ascorbate recycling provided greater protection against oxidative damage than reducing stomatal area aperture.

Our data lead to hypothesize that O₃-tolerance in bean depends more on a superior potential cultivar-specific ability more than stress-induced physiological and biochemical adjustments to avoiding and countering stress-induced oxidative damage. (Guidi et al., *Environmental pollution*, 2010, 158, 3164-3171)

Good correlation between tolerance to ozone and high endogenous levels of antioxidant metabolites such as AA and GSH in tobacco (Pasqualini et al., *Plant cell Environment*, 2001, 24, 245-252)

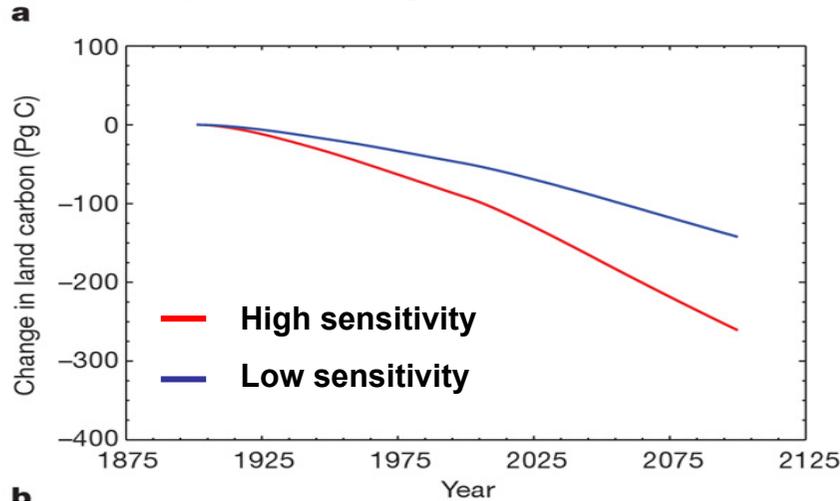
Ozone flux was best explained by stomatal conductance and symplastic (rather than apoplastic) ascorbate explaining 66% of the total variation in leaf flux. It is important to measure processes other than stomatal conductance to explain steady-state leaf-level fluxes of pollutant gases. (Eller and Parks, 2006, *Plant Cell Environment*, 29, 1742-1750).

Other factors than constitutive total apoplastic ascorbate contribute to the differential ozone tolerance of two clones of *trifolium repens*. The symplastic redox status could play a role (D'Haese et al., 2005, *Plant Cell Environment*, 28, 623-632).

The increase in foliar ascorbate under ozone stress in the sensitive poplar clone Eridano, with a greater stomatal conductance, was insufficient to counteract ROS accumulation and the consequent oxidative stress. The higher influx of ozone into Eridano leaves compared with the tolerant clone I-214 resulted in a lower *potential detoxification capacity per unit of ozone influx*. (Di Baccio et al., 2008, *Tree Physiol.*, 28, 1761-1772).

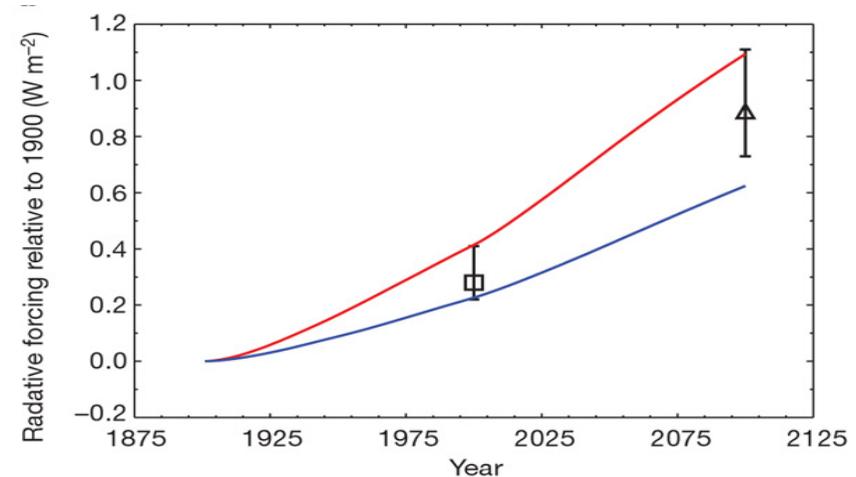
Recent developments highlight the complexity of redox-dependent defence reactions and the importance of interactions between the reduction state of soluble redox couples and their concentration in mediating dynamic signalling in response to stress - i.e. reactive oxygen species (Noctor, 2006, *Plant cell Environment*, 29, 409-425).

Temporal changes in land carbon storage and radiative forcing due to ozone



Simulated changes in land carbon storage due to O₃ increase

Sitch et al., 2007 *Nature*, 448, 791-95



Simulated changes in indirect radiative forcing due to O₃ increase (part of non-assimilated CO₂)

For comparison, estimates of the *direct* radiative forcing due to O₃ increase only are shown by the square and the triangle

Increasing ozone concentration in the atmosphere decreases plant productivity

Ozone will lower the capacity of plants to sequester the increasing level of carbon (rise in atmospheric CO₂)

Consequence: indirect radiative forcing of ozone, due to additional level of CO₂ in the atmosphere, which is added to the direct radiative forcing

Which links could exist between metabolism and ozone flux related to stomatal conductance?

$$WUE = A / g_s \quad (1)$$

From (1), (2) and (3):

$$WUE = (C_a - C_i) / 1.6 \quad (4)$$

According to Farquhar *et al.* (1982) :

$$\Delta = a + (b - a) (C_i / C_a) \quad (5)$$

From (4) and (5) :

$$WUE = C_a / 1.6 \times (b - \Delta / b - a) \quad (6)$$

According to Farquhar & Richards (1984) :

$$b = \beta \cdot b_1 + (1 - \beta) b_2$$

$$\beta = \text{PEPc activity} / (\text{PEPc activity} + \text{Rubisco activity})$$

b_1 = isotopic discrimination due to PEPc

b_2 = isotopic discrimination due to Rubisco

WUE = Water Use Efficiency

A = CO_2 assimilation = $g_{CO_2} (C_a - C_i)$ (2)

g_s = stomatal conductance to water = $1.6 \cdot g_{CO_2}$ (3)

C_a = atmospheric [CO_2]

C_i = internal [CO_2]

Δ = discrimination between ^{12}C and ^{13}C isotopes

a = stomatal diffusion

b = isotopic discrimination due to carboxylation

- As ozone induces a decrease in the Rubisco/PEPc ratio, β , b and consequently WUE should be modified under ozone treatment.