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## Advances on somatic embryogenesis in forest trees

Marie-Anne Lelu-Walter, Daniel Cornu

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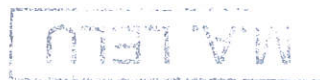
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COST "EUROSILVA" Working Group I : Growth and Development

Workshop :

"Advances on somatic embryogenesis in forest trees"

December 3rd-4th, 1998  
in Orléans, France

Organising committee

Marie-Anne Lelu, France

Daniel Cornu, France

INRA

Station de Génétique, Physiologie et Amélioration des Arbres forestiers,

Avenue de la Pomme de Pin BP 20619 ARDON

F-45 166 OLIVET Cedex FRANCE

tél. : 33 2 38 41 78 00 Fax : 33 2 38 41 48 09

Email : lelu@orleans.inra.fr ; cornu@orleans.inra.fr

Time Table

Wednesday, December 2.

6: 00 PM- 21: 00 PM

Registration Hôtel Ibis, Orléans

Thursday, December 3.

8: 00 AM

Registration

Hôtel Ibis, Orléans

8: 30 AM

Welcome : MA Lelu

Introduction : Björn Sundberg : Chairman of the Cost-Eurosilva Working Group I  
Jean-Charles Bastien : Director of INRA Station of Orléans

**I. Physiological aspects of somatic embryogenesis in forest trees**

**II. Induction of somatic embryogenesis :**

9: 00 AM

Helly Häggman : Factors affecting the induction of somatic embryogenesis in

pine

9: 20

Marc Pâques : Somatic embryogenesis from old conifers : a way to speed up the  
best clones availability for fibre-culture.

9: 40

Maria do Rosario Alvelos : induction of somatic embryogenesis in *Pinus pinaster*.

9: 45

Discussion Part II.

10: 05

COFFEE

**12. Proteins, markers of somatic embryogenesis :**

10: 35

Hélène David : Germin-like proteins are apoplastic markers of somatic and

zygotic embryos of pine.

10: 55

Ulrika Egertsdotter : Extracellular proteins in embryogenic suspension cultures  
of Norway spruce.

11: 15

Discussion Part 12.

11: 40

General Discussion (parts 11-12)

12: 40

LUNCH

### 13. Medium, hormonal analysis :

2: 00 PM	<b>Krystyna Klimaszewska</b> : Manipulation of embryogenic tissue water potential during somatic embryo maturation in conifers using <i>Pinus strobus</i> as an example.
2: 20	<b>Philippe Label</b> : Plant Hormones during hybrid larch and walnut somatic embryogenesis.
2: 40	<b>Helena Lipavska</b> : Carbohydrate status of Norway spruce somatic embryos during maturation- the effect of PEG.
2: 45	<b>Martin Vagner</b> : Endogenous phytohormones during Norway spruce somatic embryo development.
2: 50	<b>Karolina Niemi</b> : Ectomycorrhizal fungi as growth regulators in somatic embryogenesis of conifers.
2: 55	Discussion part 13.
3: 30	BREAK

### 14. Informations from zygotic embryogenesis:

4: 00	<b>Inger Hakman</b> : Aquaporins in Norway spruce - their putative functions during embryo formation and plant establishment.
4: 20	<b>Nicole Michaux-Ferriere</b> : Zygotic embryogenesis as a model for somatic embryogenesis ? Some examples from tropical trees.
4: 40	<b>Jana Albrechtova</b> : Anatomical and histochemical studies of Norway spruce somatic embryo maturation- the effect of PEG.
4: 45	Discussion part 14.
5: 15	General Discussions (parts 13-14)

### Friday, December 4.

## **II Towards an integration of somatic embryogenesis in a breeding program and other applications.**

### 11.1. Genetic transformation :

8: 30	<b>Josef Schmidt</b> : Aspect of genetic transformation in <i>Quercus</i> .
8: 50	<b>David Clapham</b> : Transformation of conifers by particle bombardment.
9: 10	<b>Gilles Pilate</b> : An overview of conifers transformation from embryogenic cultures with special emphasis on <i>Agrobacterium</i> transformation.
9: 30	<b>Jens Find</b> : <i>Abies nordmanniana</i> : Transformation and regeneration of transgenic plants



9: 35	Discussion part II.1	COFFEE
10: 00		
10: 30	<b>Bruno Florin</b> : Recent advances in long-term preservation of coffee and cocoa genetic resources.	
10: 50	<b>Marie-Anne Lelu</b> : desiccation and cryopreservation in conifers.	
11: 10	<b>Pawel Chmielarz</b> : somatic embryogenesis in <i>Quercus robur</i> L.	
11 : 15	Discussion part II.2	
11: 35	General discussions (parts II1 - II2)	
12: 35	LUNCH	
<b>II.3 Conformity:</b>		
2: 00 PM	<b>Jean-Luc Fourré</b> : Somatic variation and somatic embryogenesis in forest trees.	
2: 20	<b>Jean-Luc Verdel</b> : The mantled floral abnormality in oil palm clones : hormonal and molecular approach.	
2: 40	Discussion Part II.3	
3: 00	BREAK	
<b>II.4. Clonal propagation of forest trees via somatic embryogenesis ?</b>		
3: 30	<b>Krystyna Szczygiel</b> : Role of somatic embryogenesis in conservation of genetic diversity.	
3: 50	<b>Mike Becwar</b> : Clonal propagation of pines via somatic embryogenesis.	
4: 10	<b>D. Thompson</b> : Are we ready to integrate somatic embryogenesis into European forestry ?	
4: 30	<b>Mariano Toribio</b> : Bottlenecks in somatic embryogenesis in <i>Quercus suber</i> .	
4 : 35	Discussion part II.4.	
5: 00	General Discussions	
6:30	ADJOURN	

## Factors affecting induction of somatic embryogenesis in Scots pine.

Hely M. Häggman, Tuija S. Aronen and Jana Krájčáková

Finnish Forest Research Institute, Punkaharju Research Station, Finlandiantie 18, FIN-58450 Punkaharju, Finland.

Scots pine (*Pinus sylvestris* L.) is one of the most common conifers in Nordic countries. The traditional breeding programs have been focused on the species, and during the last few years also biotechnological studies (i.e. vegetative propagation, genetic transformation and molecular biology) have been emphasized. An efficient vegetative propagation technique is necessary both for biotechnological applications as well as for more effective transfer of genetic gain achieved through tree breeding to practical forestry. In Scots pine, regeneration has succeeded through somatic embryogenesis but the method is still under development. In this presentation we have focused on factors affecting induction and proliferation of embryogenic cultures.

One-year-old immature seed cones were collected from elite clones of Scots pine growing in Punkaharju clone collection (61°48'N; 29°17'E). Embryogenic cultures were established on immature female gametophytes including immature zygotic embryos with suspensor tissues. The effect of the following factors on induction was tested: time of collection, developmental phase of the explant tissues at the time of collection, the growth regulators and gelling agent concentration in the induction media, cold-treatment before inoculations, and the response of different seed families on induction of embryogenic cultures.

The optimal stage for cone collections in southern part of Finland is usually in July, when the day degrees are between 400 and 650. Anatomical studies have shown that the immature zygotic embryos have the ability to initiate embryogenic cultures shortly after fertilization. Most of the Scots pine elite plus trees have been able to produce embryogenic cultures. Generally, the initiation frequencies have been relatively low and depending on the collection year as well as mother tree. Storing immature cones in cold room before inoculation is as good as immediate inoculation at the collection date or even increases, but not significantly, the number of embryogenic cell lines. For successful initiation and proliferation of embryogenic cultures auxin supplement has been necessary in the proliferation medium. The two auxin containing media DCR1 (13.6  $\mu$ M 2,4-D and 2.2  $\mu$ M BA) and DCR3 (9.05  $\mu$ M 2,4-D and 2.2  $\mu$ M BA) tested did not differ statistically from each other. Also the gelrite concentration affects the number of embryogenic cell lines but due to the low initiation frequency the difference is not significant.

Somatic embryogenesis from old conifers: a way to speed up the best clones

availability for fibre-culture.

**M. Paques and L. Harvengt**

AFOCCEL, Ressources du futur; Domaine de l'Étangon, F-77370 Nangis FRANCE

e-mail: paques@afocel.fr

AFOCCEL, a French company founded by the wood industries, is involved in Somatic embryogenesis since 1986 in order to be able to scale up the process at an industrial level. The demonstration of the possibility to produce currently a thousand quality plants of Norway spruce has been made by AFOCEL. The plantations now older than 5 years are indicating that the somatic seedlings quality is good and that the process could be used at an industrial level for the clonal propagation of elite clones. The estimated cost of a somatic seedling ready for acclimatization and for field plantation is respectively estimated around 2.5 and 5 Francs. A substantial reduction of those costs is still expected because the possibilities of reducing the hand labor both in the lab and in the nursery.

The interest of the technology by itself is relatively low if it is used to amplify untested very juvenile genotypes. The fact that the technology can be successfully combined with the cryopreservation is interesting : it allows to maintain juvenile the embryogenic tissues as long as requested for the field trials evaluation. However the valorization of somatic embryogenesis would be much more higher if this technology is applicable to old selected genotypes

As a consequence AFOCEL is focusing from years on the possibility to recover somatic embryos from " old " selected conifers.

In order to progress in that way, the research has been focused on the possibility to recover somatic embryos from more and more older plants starting on one hand from juvenile to mature trees and on the other hand from somatic to zygotic plants.

As results from those investigations :

we made the demonstration that it is possible to recover somatic plantlets from old conifers via somatic embryogenesis as recently described in a patent application.

we have learned a little more on the factors involved in the control of the initiation embryos from relatively old conifers.

Some of those information's will be presented in order to stimulate the discussion on the scientific investigations required to increase the level of embryogenesis initiation rate from juvenile and mature conifers.



Germ-in-like proteins are apoplastic markers of somatic and zygotic embryos of pine.  
Hélène David, Lionel Belingheri, Bertrand Dehorter and Godfrey Neuteilings  
Laboratoire de Physiologie des Parois Végétales, UPRES Différenciation Végétale, UFR de  
Biologie, USTL, 59655 Villeneuve d'Ascq Cedex-France.

In coniferous species, preglobular somatic embryos consist of embryonic heads composed of small cells rich in cytoplasm attached to long highly vacuolated suspensorial cells. These embryos are maintained as callus-like embryogenic tissues on a 2,4-D containing medium (Domon *et al.*, 1994). We previously characterized this early stage of pine development by the presence of germ-in-like proteins (GLPs) ionically bound to the cell surfaces (Domon *et al.*, 1995).  
Germ-ins and GLPs are proteins that belong to a multigenic family present in numerous species (Membre *et al.* 1997) and expressed in different physiological processes including development and stress tolerance. No precise function has been disclosed for them except that cereal germ-ins express an oxalate oxidase activity (Dumas *et al.*, 1993; Lane *et al.*, 1993).

In order to understand the involvement of GLPs in early embryo development, a cDNA library was constructed from pine preglobular somatic embryos. A 926bp full length cDNA named *PcGERI* (Genbank AF039201) encoding a predicted 220 amino acid protein precursor was isolated and further characterized. Expression of the recombinant *PcGERI* in *E. coli* confirmed the relationships between *PcGERI* and germ-ins (Neuteilings *et al.*, 1998).  
Very recently (Dunwell and Gane, 1998) it was emphasized on the absolute conservation of residues involved in structure determination of several procaroytic and eucaryotic proteins including germ-ins. The two germ-in conserved motifs described by the authors are found in *PcGERI*. We also PCR-amplified a pine genomic fragment of 852 bp named *gf-0.8*. The alignments with *PcGERI* cDNA confirmed the absence of introns in the corresponding genomic region, as it is the case for wheat germ-ins. Southern blot analysis was performed to estimate the number of *PcGERI* genes in the Carribean pine genome which appears to possess only a single copy of *PcGERI* while another related gene might be present.

*PcGERI* was highly expressed in the four embryogenic lines analysed as shown by the unique band corresponding to a 0.9 kb transcript. A strong signal was also obtained in quiescent zygotic embryos. Conversely, *PcGERI* was never expressed in nonembryogenic calli nor in haploid female megagametophytes and during germination.  
In conclusion, we have cloned a GLP cDNA, *PcGERI*, from pine which is expressed specifically in embryogenic lines and in quiescent embryos. Our objectives are now to study (1) the regulation of this GLP, (2) and the biochemical properties of the recombinant glycosylated polypeptide *PcGERI* produced in tobacco. Our aim is also to disclose whether the expression of GLPs is correlated with the bipolar structure of the conifer preglobular somatic embryos (small embryo head and large suspensor).

Domon *et al.*- Plant Physiol Biochem 32: 137-147 (1994).  
Domon *et al.* - Plant Physiol 108: 141-148 (1995).  
Dumas *et al.* - C R Acad Sci Paris 316: 793-798 (1993).  
Dunwell J, Gane P - J Mol Evol 46: 147-154 (1998).  
Lane *et al.* - J Biol Chem 268: 12239-12242 (1993).  
Membre *et al.* - Plant Mol Biol 35: 459-469 (1997).  
Neuteilings G *et al.*- Plant Mol Biol, sous presse (1998).

## Extracellular proteins in embryogenic suspension cultures of Norway spruce.

Ulrika Egertsdotter

NISK Høgskoleveien 12, N-1432 Ås Norway

Email : Ulrika.Egertsdotter@nisk.no

Different cell lines of Norway spruce contain somatic embryos that reach different developmental stages. The cell line-specific differences in embryo morphology can be linked to the embryos capacity to enter the maturation phase, and subsequently form normal-looking plantlets. We have divided our cell lines into two groups, A and B. Somatic embryos in the group A cell lines have a high maturation frequency and are characterised by embryos with a large embryonic region (> 400  $\mu\text{m}$ ). Group B embryos seldom forms mature embryos and characteristically have a small embryonic region (<200 $\mu\text{m}$ ). The size of the embryonic region, and the capacity to mature, is to some extent regulated by auxin and cytokinin. The morphology of the somatic embryos are also influenced by protein-related compounds of embryo origin, in the extracellular environment. We have been using well established suspension cultures of group A and B cell lines, to study how extracellular proteins regulate the development of somatic embryos. The effects of different compounds on embryo morphology have been studied in bioassays. The total secretion of proteins to the suspension culture media reflects the embryo morphology as shown by *in vivo* labelling techniques. Proteins and proteoglycans that differ in the extracellular environment of group A and B include zeamatin, chitinases, peroxidases and arabinogalactoproteins (AGPs).

The zeamatin protein is specifically found in an A cell line, and an immunologically related protein only appear in the control B cell line when cultured with cytokinin alone. A 28 kDa chitinase is specifically secreted by group A cell lines, and in group B cell lines cultured with cytokinin alone. A related chitinase isolated from sugarbeet promote the formation of embryos in bioassays. However, an endogenous spruce chitinase has a detrimental effect on the growth of the embryos. The activity of peroxidases is higher in group A cultures than in group B cultures. Increasing the level of peroxidases in the culture medium is however not influencing the morphology of the group B embryos, indicating isoform specificity in a similar way to what appear to be the case for chitinases. Group A and B cell lines contain different amounts and species of extracellular AGPs. Isolated extracellular AGPs from group A cell lines, or from the crude extracts of seeds, can enhance the size of the embryonic regions of group B embryos. Additionally, the aggregation of embryos and the expansion of cells are influenced. Extracellular proteins in the suspension culture medium are also present in seeds. Extracts of whole seeds strongly stimulate growth rate and embryo morphology in both group A and B cell lines, and seed extract treated B embryos are forming mature somatic embryos in response to an ABA treatment. Proteins have been shown to be crucial components of the seed extract for its stimulatory effect, supporting the hypothesis that proteins play an important role in regulating embryo development.



Manipulation of embryogenic tissue water potential during somatic embryo maturation in conifers using *Pinus strobus* as an example.

Krzyszyna Klimaszewska, Natural Resources Canada, Canadian Forest Service, Laurentian Forestry Centre, 1055 du P.E.P.S., P.O.Box 3800, Sainte-Foy, Quebec, Canada G1V 4C7  
email: klimaszewska@cfl.forestry.ca

In conifers, the osmotic water stress to which the embryogenic tissue is exposed either initially (such as in the case of low molecular weight solutes) or continuously (such as in the case of high molecular weight solutes) triggers and/or maintains the maturation process of somatic embryos. Since in *P. strobus* high numbers of somatic embryos mature on medium with no significant changes in the solute composition, except for increased gellan gum concentration (Klimaszewska and Smith 1997), it represents a model to study the dynamics in the water status of the tissue during the maturation process.

Gellan gum is a bacterial (*Pseudomonas elodea*) polysaccharide composed of glucuronic acid, rhamnose and glucose. Agars are derived from seaweed (agarophytes) and represent a spectrum of closely related polysaccharides belonging to the family of galactans. These qualitative differences between the two gelling agent types are significant enough to pose a question concerning the possibility of stimulatory effects of certain gellan gum components on the maturation of somatic embryos of *P. strobus*. To determine if this was a viable hypothesis, *P. strobus* somatic embryo maturation was carried out on filter paper discs placed on media gelled with gellan gum or several types of agar. In order to make the comparison meaningful, gel strength of all the maturation media was measured and the embryonal masses' response compared at similar gel strength values. The results showed that the medium gel strength was a critical factor in exerting the maturation response regardless of the type of solidifying agent used and that both water availability and tissue water potential were negatively correlated with the gel strength of the maturation medium. There was a reduction in the water content of approximately 50 mg per 227 mg filter paper disc between medium gelled with 0.4 and 1.2% gellan gum and 70 mg between medium gelled with 0.8 and 2.8% agar Difco-Noble. It is concluded that the tissue was exposed to varying amounts of water during the culture period and this in turn affected the water status of the tissue and the maturation response. Initially, the water potential of the embryogenic tissue was in equilibrium with the water potential of the maturation medium (-0.43±0.01 MPa) but after two weeks of culture, a clear trend was established. The embryogenic tissue cultured on maturation medium with 0.4 and 0.6% gellan gum had a higher (less negative) water potential than that of the medium and the embryogenic tissue cultured on medium with 0.8 and 1.0% gellan gum had a lower water potential. This trend was maintained throughout week 12. The mature somatic embryos from medium with 1.0% gellan gum had water potential of -0.70±0.00 MPa and germinated at a high frequency. In contrast, somatic embryos matured on medium with 0.6% gellan gum had a water potential of -0.32±0.02 MPa and developed precociously after weeks 6 and 7.

In conclusion, embryogenic tissue water potential of *P. strobus* and other conifer species can be manipulated, without changing the solute composition of the medium, to procure production of somatic embryos that display high germination frequency. These somatic embryos germinate without any post-maturation treatment and are also amenable to desiccation.



Plant Hormones during hybrid walnut and hybrid larch somatic embryogenesis.

Philippe Label, Sandrine Drone, Markus Gutmann, Isabelle Lucas, Isabelle Jourdain, Sabine Saly, Claude Joseph, Daniel Cornu and Marie-Anne Lelu.

Station de Génétique, Physiologie et d'Amélioration des Arbres Forestiers  
INRA-CRO BP 20 619 Ardon F-45166 Olivet Cedex France

Vegetative propagation of hybrid walnut (*Juglans regia* X *J. nigra*) is difficult through traditional techniques like cuttings but more powerful techniques like somatic embryogenesis offer a good solution to produce vegetatively selected trees. Production of somatic embryos is performed through an adventive secondary SE and secondary somatic embryos (SSE) produced from the "primary" somatic embryo (PSE) are isolated, thus becoming PSE and subcultured iteratively. These steps occur without any exogenous plant growth regulator complementation. The effect of liquid endosperm on SSE production will be developed and correlations between endogenous hormonal levels and SSE formation will be outlined.

Among the conifers, the hybrid between European and Japanese larches has a great potential for reforestation programmes due to its fast growth and good wood quality. Somatic embryogenesis offers a solution to produce large quantities of propagules especially in hybrid larch (*Larix x leptoeuropaea*) species. The hormonal status at different stages of somatic embryo development will be presented, either during embryogenic induction, embryonal mass growth or somatic embryo maturation. Correlations between internal hormones and germination will also be discussed with a special emphasis on the role of desiccation.

The presentation will aim to draw general lines of hormonal regulation of somatic embryogenesis and will tentatively formulate fundamental questions on that subject that could be addressed and discussed during the present COST meeting.

Aquaporins in Norway spruce - their putative functions during embryo formation and plant establishment.

Inger Hakman

Dept. of Botany Stockholm University S-106 91 Stockholm

Sweden

Email : inger.hakman@botan.su.se

Aquaporins, water channel proteins that are members to the large MIP (major intrinsic protein) family of transmembrane channel proteins, have now been identified in several plant species. These proteins were named aquaporins (AQP) because of their high efficiency and selectivity to conduct water in the direction of an osmotic gradient. This capacity to transport water, is often demonstrated by increased water permeability of *Xenopus* oocytes expressing heterologous mRNA. The discovery of aquaporins in plants have suggested their involvement in a number of processes from long-distance transport of both water and assimilates to single cell expansion and osmoregulation, that requires the transport of water across cellular membranes, and, have directed new attention to the dynamics of water transport in plants particularly under water limiting conditions.

Plant MIPs are encoded by a relatively large gene family, and these genes display differences in their tissue and developmental expression patterns as well as in their response to various environmental conditions. In an earlier investigation we identified a protein isolated from mature seed of *Picea abies* (Norway spruce) as an a-TIP homologue by its biochemical characteristics such as molecular mass, partial amino acid sequence, and its location in protein storage vacuolar (PSV) membranes. We also identified similar proteins in *P. abies* female gametophytes of seeds prior to embryo development, in mature somatic embryos and in young seedlings where it was particularly abundant in the root.

In the biotechnology program somatic embryogenesis is generally regarded as the most important method to mass-produce selected clones. During this process an important step is the conversion of somatic embryos into embryos and their further establishment as soil-growing plants. Here root formation and growth is often a problem. For these reasons we are interested in processes involved in germination and seedling growth, and a better knowledge about the regulation of aquaporins may therefore be an advantage. Such knowledge may also be useful for improving plant production in the more conventional breeding programs such as through seedlings or cuttings.

In order to further our studies regarding aquaporin-related proteins in *P. abies* we have cloned a gene expressed in the female gametophyte during seed maturation and another isoform expressed in seedling roots. Probes from these clones have been used in Northern experiments and for *in situ* hybridization in order to identify the expression pattern of aquaporin homologues in various tissues of *P. abies*.



## Zygotic embryogenesis as a model for somatic embryogenesis ? Some examples from tropical trees.

N. Michaux-Ferrière

Histologie BIOTROP CIRAD BP 5035 34032 Montpellier

When induction conditions of somatic embryogenesis are well-known for a species, one of the keys for an efficient micropopagation method is to control development of the embryos obtained. A percentage of embryos as high as possible has to be well-formed, to be able to germinate and then converted to plantlets before transferring to the green-house and then in the field. For many species, and specially for woody species, blockages during ontogenesis or germination of the embryos were noticed (Oil Palm, Rubber tree, Cocoa, Coconut and Grapevine). To remove these blockages it is necessary first to identify them (structural, biochemical or physiological anomalies) by comparing somatic embryo development with zygotic embryos "in ovulo" considered as a model (Rangaswamy, 1986; Carman, 1989) Then, taking into account the differences registered between both the evolutions, new culture conditions can be defined to improve the quality of the embryos. Such an approach has been followed by our team to improve embryogenesis process in Rubber tree ( Etienne et al. 1993 a et b ), Oil Palm ( Morcillo et al. 1998) Cocoa ( Alemanno et al. 1997) among others. For the three considered species, the study of water parameters, of accumulation of reserves (protein, starch and lipids) of sugar content or ABA concentration throughout the zygotic embryo evolution have pointed out the importance of a "maturation" phase taking place after ontogenesis and growth were completed. It is during this phase (at the beginning or end) that embryos were dehydrating (more or less according to their recalcitrance level) and accumulating reserves. At the end of the somatic embryogenesis process, studies of some parameters demonstrated that embryos formed were more often non maturing even those morphologically well-formed. An additional phase of culture in conditions allowing a progressive dehydration and a synthesis of specific reserves (increasing sucrose or amino acid concentrations in the culture medium or adding ABA) has improved percentages of germinated embryos for the three considered species. The application of molecular biology technology to investigate the genes concerned with embryogenesis and their expression has provided interesting results in model plants (Goldberg et al. 1994; Vroemen et al. 1996; Komamine et al. 1998 ). "Differential Display" used to examine gene expression throughout the development of zygotic and somatic embryos could provide a better knowledge of both systems (Pullmann et al. 1998) and therefore improve the efficiency of somatic embryogenesis regeneration for the species we are studying.

Alemanno L. et al. 1997 - In vitro Cell Dev. Biol-Plant 33: 163-172

Carman J.G 1989 - In vitro Cell Dev. Biol-Plant 25 : 1155-1162

Etienne H. et al. 1993a- Plant Sci.92: 111-119

Etienne H. et al 1993b-J.Exp.Bot.44 : 1613-1619

Goldberg R.B et al 1994- Science 266: 605-614

Komamine A. et al.1998- IX Congress on Plant Tissue Culture. Jerusalem Israel June 14-19

Morcillo F. et al. 1998- Plant Physiol.Biochemistry

Pullman G. et al. 1998- IX Congress on Plant Tissue Culture. Jerusalem Israel June 14-19

Rangaswamy N.S 1986- Proc. Indian Acad.Sci 96 : 247-271

Vroemen C.W et al. 1996- Plant Cell 8:783-791

Aspects of genetic transformation in *Quercus*.  
E. Wilhelm, A. Burg, M. Berenyi, M. Endemann, R. Rodler and J. Schmidt

Austrian Research Centre Seibersdorf  
A-2444 Seibersdorf

Austria

Immature zygotic embryos were used as explant source to induce somatic embryogenesis throughout several years from several open pollinated trees at different sites. The suitable developmental window lasted from 3 to 18 weeks after anthesis with the highest initiation rate of nearly 60% occurring approximately 13 weeks after anthesis on 5 (M 2-4D plus 0.5 (M BAP. Embryogenic competence of the callus lines has been maintained by regular subculturing on 1 (M BAP or hormone free media for four years. Maturation media with increased agar concentration (1%) proved to enhance germination rate significantly. Conversion rate of matured somatic embryos into plantlets could be increased to 80% by a partial desiccation treatment.

Somatic embryogenic callus was shown to be infectable by *Agrobacterium tumefaciens*. Additional wounding of embryogenic tissue caused significant decreases in regeneration. A cocultivation period of two days seemed to enhance the regeneration capacity of oak somatic embryogenic callus more than four days cocultivation. An optimum of 70% regeneration could be observed after two passages on 50 mg/l kanamycin + 300 mg/l cefotaxime. More than 100 lines produced initially have been subcultured on 50 and 100 mg/l kanamycin for two consecutive years, respectively. Samples were repeatedly tested with selected PCR primers for the presence of the CAT (chloramphenicol acetyltransferase) gene. More than 50% of the highly embryogenic cultures exhibited the CAT fragment for up to 15 months, whereas *in vitro* plantlets germinated from four kanamycin tolerant cultures surviving up to two years did not contain the insert.



## Transformation of conifers by particle bombardment

David Clapham, Sara von Arnold

Department of Forest Genetics, Swedish Univ Agric Sci, Box 7027, S-750 07 Uppsala, Sweden

Transgenic plantlets of *Picea glauca* were produced by Ellis and coworkers in 1993 (Bio/Technology 11, 84-89) using particle bombardment and the method has subsequently been applied to other conifer species. In our department, a particle inflow gun enabled efficient production of transgenic plantlets of *Picea abies* from embryogenic suspension cultures. In studies of transient expression of the *gusA* reporter gene driven by various promoters, the *Zea* ubiquitin promoter was three times as active as the *Helianthus* ubiquitin promoter, four times as active as the *Eucalyptus* CAD promoter and 12-16 times as active as the 35S promoter. In four cell lines, placing the cells 1-3 h before bombardment on proliferation medium supplemented with 0.125-0.25 M myoinositol enhanced transient expression 15-40-fold. The procedure for stable transformation was developed using the plasmid pAHC25 which contains the *bar* gene and the *gusA* gene, both driven by the *Zea* ubiquitin promoter. Cells were maintained on proliferation medium supplemented with 0.25 M myoinositol and, from day 8 after bombardment, supplemented with Basta as selective agent. Callus resistant to Basta appeared from two months after bombardment. Of over 100 independent Basta-resistant sublines, 65% expressed the cotransformed reporter gene, even when it was not linked to the selectable marker. Over 80% of the sublines retained their embryogenic potential. Of eleven transformants analyzed, four contained transgenes in low copy number (1-3). Over 200 Basta-resistant sublines from four cell lines have been established, of which 138 are confirmed as transformed. Plantlets have been regenerated and potted.

Transformation of conifers via *Agrobacterium tumefaciens*  
Gilles Pilate, Jean-Charles Lepié, Daniel Cornu, Marie-Anne Lelu  
INRA, Unité Amélioration, Génétique et Physiologie Forestières,  
Ardon, F-45 160 Olivet, France.

For a long time, conifers were thought to be recalcitrant to *Agrobacterium* infection. However, the first conifer species - larch - was genetically transformed through cocultivation with *Agrobacterium rhizogenes* (Huang et al., 1991, In Vitro Cell Dev. Biol., 27, 201-207). We have developed an efficient procedure for the genetic transformation of hybrid larch (*Larix kaempferi* x *L. decidua*) using *Agrobacterium tumefaciens* as vector for DNA transfer (Levée et al. 1997, Plant Cell Reports, 16, 680-685). After cocultivation, embryogenic masses (EM) resistant to the selection agent were isolated. Using somatic embryogenesis numerous plantlets can easily be regenerated from each of antibiotic-resistant EM and only seven months are required from the cocultivation step to the acclimatisation of plantlets in the greenhouse. Since this first report, the procedure has been optimized in order to increase the yield of antibiotic-resistant EM. These improvements allow us to regularly obtain up to 50 different EM resistant to the selective agent per g FW. All of the lines analyzed by molecular techniques exhibited stable integration of the transgene. Copy number remains fairly low as 4 of 12 transgenic lines analyzed seems to have only one copy inserted. The high transformation yield obtained probably results, at least in part, from the high growth rate of the embryogenic lines used in transformation experiments. To be efficient, this method also requires a careful adjustment of selection pressure (timing of antibiotic application, antibiotic concentration). This procedure has been shown to be efficient on different hybrid larch genotypes. It still needs to demonstrate that it can be transferred to other larch species and further to other *Pinaceae* species.

This presentation will outline a description of the transformation procedure and detail the first results on the molecular characterisation of transgenic hybrid larch.



Recent advances in long-term preservation of coffee and cocoa genetic

resources

**Bruno Florin et Vincent Pétiard**

Centre de Recherche Nestlé - Tours, 101 Avenue Gustave Eiffel, Notre Dame d'Oé

B.P. 9716, 37097 Tours cedex 02, France.

Coffee and Cocoa are of great economical importance for many countries. In vitro techniques present a real interest in the development of new improved varieties but also for the preservation of genetic resources of these tropical species because they produce non-orthodox seeds, which are unsuitable for germplasm storage. Cryopreservation has proven to be reliable for long-term storage of biological materials. Cryopreservation of plant somatic embryos and/or embryogenic cells may therefore be used for germplasm storage and exchange, but also for a constant supply of embryogenic strains for propagation of elite clones or genetic modification experiments. Because of both the labour intensive and time consuming issues linked to the establishment of these cultures and the decline of their embryogenic potential with the subculture number, cryopreservation appears to be a key tool for clonal propagation of elite plants. Based on results obtained for somatic embryos and embryogenic strains of coffee and cocoa, cryopreservation approaches will be discussed.

Desiccation and cryopreservation in conifers.  
Marie-Anne Lelu, Geraldine Pflaum, Sandrine Dronne, Sophie Dannoux,  
Catherine Bastien and Philippe Label.  
Station de Génétique, Physiologie et d'Amélioration des Arbres Forestiers  
INRA-CRO BP 20 619 Ardon F-45166 Olivet Cedex France

Most seeds undergo desiccation during later stages of zygotic embryo development. This process has a role in the transition between embryo maturation and germination. In contrast, somatic embryo development is continuous without desiccation such as in zygotic embryos. To produce somatic embryos capable of synchronized germination, appropriate methodology has been developed. The rate of drying is a critical factor in acquisition of desiccation tolerance and generally slow drying is applied to somatic embryos (i.e. desiccation realized under high relative humidities 80-98%). Desiccation tolerance has also been enhanced by different treatments such as exogenous application of abscisic acid (ABA) and osmotic stress (PEG). In many conifers (*Picea glauca*, *P. glauca engelmannii*, *P. mariana*, *P. sitka*, *Larix leptocephala*), partial drying of somatic embryos promotes rapid and synchronous germination and plantlet development.

The use of dried somatic embryos might also constitute a method for storage of selected genotypes. For this purpose, desiccation treatments were tested under low relative humidities (43-59%) in order to reduce somatic embryo water content (close to stored seed embryos). In hybrid larch, survival of somatic embryos to these severe desiccations was depending on their maturation duration (Lelu *et al.* 1995).

In conifer species, production of somatic embryos that can be stored dry has not been so much studied due to the development of more efficient storage method such as cryopreservation. Indeed cryopreservation in which embryonal masses are frozen under controlled conditions and stored in liquid nitrogen (LN) is a reliable method and has been an important tool for long term preservation of germplasm. In general, embryonal masses are precultured with high concentrations of sucrose or sorbitol (0.4 to 0.6 M) and pre-treated with cryoprotectants such as DMSO (5%). Then successful cryopreservation has been achieved by slow prefreezing from -30°C to -40°C prior immersion in LN. It has been demonstrated that embryonal masses are amenable to cryopreservation and plantlets have been regenerated from cryostored embryonal masses of numerous conifer species (*Abies*, *Larix*, *Picea*, *Pinus* sp.).

M.A. Lelu, K. Klimaszewska, G. Pflaum and C. Bastien, 1995.  
Effect of maturation duration on desiccation tolerance in hybrid larch (*Larix x leptocephala*  
Dengler) somatic embryos. In Vitro Cell. Dev. Biol., 31, 15-20.



## Somacalonal variations and somatic embryogenesis in forest trees.

Jean-Luc Fourré

UCL-Unité des eaux et forêts Place Croix du Sud 2 bte 9

B-1348 Louvain-la-Neuve (BELGIUM)

Tel: +32-10-473707 Fax: +32-10-473697 Email: fourre@efor.ucl.ac.be

Aside from traditional propagation techniques, somatic embryogenesis is often considered to be a very promising technique for vegetative multiplication of forest trees. Moreover, this method is already used as a regeneration model for genetic transformation experiments. However, plantlets derived from *in vitro* culture may exhibit phenotypical anomalies called somacalonal variations (Larkin and Scowcroft 1981). For the "Tissue Culture Association Terminology Committee", a somacalonal variation is a phenotypic variation, either genetic or epigenetic (i.e. non genetic) in origin, displayed among somacalones (Schaeffer 1990). Somacalonal variation is caused by the *in vitro* culture system itself or may be the result of pre-existing physiological or genetic variation in somatic. Virus elimination can also explain some cases of somacalonal variation. A variety of studies on somacalonal variation have been performed only on *in vitro* growing cells or tissues (calli, cellular suspensions, protoplasts, embryogenic tissues), rather than on acclimated regenerants. The diminution or the loss of regeneration capacities of embryogenic cell lines is often ascribed to somacalonal variations. For forest trees with a long life cycle there is a lack of information concerning the transmissibility of somacalonal variation to the offspring, but for herbaceous species with a short life cycle, it has been demonstrated that phenotypical anomalies produced by *in vitro* propagation systems can be heritable, partially heritable or not transmitted through meiosis. These last two cases of variation often called epigenetic or non-genetic variation, appear in regenerated plants as a result of physiological responses. For instance, shoot regeneration from dedifferentiated cultures can result in an immature form of the parental clone that may or may not be stable over time. Beside these epigenetic phenomena, somacalonal variation that is fixed as mutations may be essentially identical to sports and other somatic mutations. Especially in tissue culture, the combination of a high rate of mitotic division, and the occurrence of unusual combinations of nutrients and phytohormones active in both the production and repair of mutational events, may explain at least in part, the high rate of mutations. Many factors influence the level of somacalonal variation like the regeneration system used, the culture conditions and the number of subcultures or other specific procedures like protoplast isolation, gene transfer or cryopreservation. Some species or clones are also more susceptible than others. Morphogenetic, cytotogenic, cytomeric, and molecular approaches have been used to study somacalonal variation. This talk will review advantages and drawbacks of these different approaches.

The mantled floral variation in oil palm clones : hormonal and molecular approach  
J.L. Verdeil, A. Rival, I. Besse, R. Maldiney\*, J. Tregear, T. Beulé, Y. Duval and  
E. Miginiac\*

CIRAD-CP/ORSTOM - Laboratoire GenéTrop - B.P. n° 5045 - 34032 Montpellier Cedex 01 -

France.

\* Lab physiologie du développement des plantes, Université Paris VI, 4, place Jussieu, 75 252 Paris

cedex 05 - France.

Email : verdeil@melusine.mpl.orstom.fr

Clonal propagation of oil palm (*Elaeis guineensis* Jacq.) through somatic embryogenesis has led to the production of several million clonal plantlets to date. Evaluation in the field has revealed the occurrence of variant palms (below 5%) showing an abnormal flower development, called "mantled" (development of stamens in carpell-like organs). Furthermore, clonal oil palm plantlets originating from Nodular Compact Calli (NCC) have been shown to exhibit the "mantled" variant phenotype at an average level below 5%, whereas this rate has been found to attain 100% in plantlets derived from Fast Growing Calli (FGC).

Endogenous levels of cytokinins were measured in the two type of callus of oil palm by an HPLC-ELISA methodology. No significant amounts of cytokinins were detected in FGC whereas they accumulated in NCC. Furthermore, it has been demonstrated that an increase in the 2,4-D level produces a decrease in the NC endogenous cytokinins content and promote the evolution of NC into FGC. Analysis of inflorescences issued from abnormal and normal individuals confirm the disorder characterized by lower levels of cytokinins.

Studies of ploidy level and RAPD polymorphism have not revealed any genomic changes that could be linked with the variant phenotype. These results are consistent with the hypothesis of an epigenetic origin for the abnormality.

Levels of global DNA methylation were estimated after enzymatic hydrolysis of genomic DNA to nucleosides and HPLC quantification of 5-Methyl deoxyCytidine. Methylation rates of genomic DNA [(5mdC) / (5mdC+dC)] have been investigated in regenerated oil palms, with the aim of comparing normal and variant regenerants within the same clonal line. Global levels of genomic DNA methylation were found to range 20 to 25%. DNA methylation has been found to discriminate the variants at the adult age. In some clonal lines, analyses of leaf genomic DNA have revealed a substantial demethylation in severely "mantled" palms (20.6% versus 22.2%).

A significant hypomethylation in FGC (23.2% vs 18.7%) has been found when compared to NCC from the same clonal line.

Evidence for a direct relationship in oil palm between hypomethylation of genomic DNA and the determinism of the "mantled" somaclonal variant phenotype is still yet to be obtained, but our studies on DNA methylation have already opened promising lines of research.



## Role of somatic embryogenesis in conservation of genetic diversity.

Krzyszyna Szczygieł

Department of Genetics and Forest Tree Physiology, Forest Research Institute, Warsaw, Poland.

Mass decline of forest trees populations in Poland was first observed in the eighties at the higher elevation (800 m.) of Western Sudety mountains. This forest population decline resulted in increased necessity of its geneticall diversity protection. This disaster caused by air pollution resulted at first in Norway spruce population decline. Total deforestation area was estimated at the level of 15,000 hectares. Near half of Norway spruce stands in Poland are under influence of industrial emissions. Additionally, silver fir and oak populations decline is observed. Other species as larch, or beech produce seeds irregularly. On the other hand, oak, chestnut and sycamore recalcitrant seeds storage is difficult for a long time. All these reasons resulted in Program of Forest Genetic Resources Conservation. This program includes establishment of gene banks as protection plots *in-situ* and *ex-situ* for long storage of seeds, pollens and plant tissues.

For genetic resources and geneticall diversity conservation, storage of tissues or organs at the temperatures of  $0 - 10^{\circ}\text{C}$  or in liquid nitrogen can be used. More efficient method of trees tissue and organs long term storage is use of cryopreservation. This method is useful also in respect to recalcitrant seeds. Embryogenic callus can be stored for a long time in liquid nitrogen and it can be use in future for production of planting material for reforestation and conservation of forest trees genetic resources in gene bank. For this reason methods of somatic embryogenesis and cryopreservation need more work to be improved. Wide range of genotypes have to be used for induction of somatic embryogenesis which should protect genetic diversity. Additionally use of this method should avoid mutations and adaptation of embliings to natural conditions have to be improved. Very important will be also possibility of embryogenic tissue induction on organs collected from old trees. At present in Forest Research Institute in Warsaw, we study of somatic embryogenesis for long term storage in liquid nitrogen and propagation of Norway spruce, European larch and silver fir Polish provenances.

## Clonal propagation of pines via somatic embryogenesis.

Michael R. Becwar, Lewis W. Handley and Mark R. Rutter

Westaco Forest Science and Technology, PO Box 1950, Summerville, SC 29484 USA

mrbecwa@westaco.com

The development and use of somatic embryogenesis (SE) for the clonal propagation of pines will be discussed. Primary advantages of SE over other conventional propagation systems include (1) the ability to cryo-preserve cell lines to overcome problems associated with maturation of stock plants, (2) the potential to automate embryo production and synthetic seed manufacture, and (3) the flexibility associated with having an indefinite long-term cryogenic storage bank to use in clonal testing. We have developed an efficient SE system for use in clonal testing of loblolly (*Pinus taeda*) and other pine species. Components of the system will be described, including (1) improved culture initiation, enabling a higher genotype capture frequency, (2) increased somatic embryo production, and (3) methods to enhance embryo quality that translates into better germination and conversion to vigorous somatic embryo-derived planting stock. Establishment of clonal field tests of pine somatic embryo-plants will enable the selection of desirable genotypes for plantation deployment and use in conjunction with genetic engineering. Even with the above improvements, pine species remain recalcitrant relative to some conifers. SE in pines appears to be under strong genetic control. This high degree of genotype specificity makes the application of SE to clonal propagation challenging. Fundamental genetic and biochemical research is needed to elucidate mechanisms that control induction of SE and completion of the regeneration process. Results will be presented on factors that may contribute to genotype specificity during the culture induction step in pine species.



## Are We Ready to Integrate Somatic Embryogenesis in European Forestry ?

D.G. Thompson

Coillte Teoranta- The Irish Forestry Board

Research Laboratory Newtownmounkenedy

Country Wicklow Ireland

Among forest tree species, conifer somatic embryogenesis is the most well developed and nearest to integration in tree improvement programmes. Although this process was originally reported from three European research laboratories, presently it is at the "pilot-scale" in parts of the world outside of Europe. Therefore, the most pressing question for people at this Workshop is not "Should we integrate somatic embryogenesis with conventional tree improvement?" (the answer is clearly yes) or "How can we integrate this technology?" (we will work this out when we begin to integrate) but rather the more important "What do we need to do to advance the application of this technology in Europe?" or "How can we catch up with the others ?

Research over the past 15 years on conifer somatic embryogenesis first concentrated on developing an understanding and then attempting to overcome the technical bottlenecks in the process. This is where we are at present. We have in some species such as the spruces, a process, while not completely understood, controllable or predictable, can, nevertheless, produce enough plants to demonstrate the potential of the technology. What we need to do now (like others elsewhere), is to create interest among potential customers and to develop a market for this technology.

Conifer somatic embryogenesis is not yet "market driven" because very few foresters and tree breeders are demanding emblys. This technology is actually "technology driven" where we need to find an application, a market and a customer for emblys. This is mainly due to the fact that most customers (foresters and tree breeders) do not fully understand and appreciate both the applications and the implications of this technology. The challenge now is to create and develop this market.

We need to continue to work on improving our basic understanding of the process as well as overcoming the bottlenecks such as low initiation rates, maturation/germination problems and the production of high quality emblys. At the same time we need to stimulate interest on the part of tree breeders and foresters in the process. To accomplish this we need to start work on scale-up of the technology, establishment of both field demonstrations and scientific trials and to illustrate and document the biological and economic aspects of emblys. Development of the application of this technology is essential if interest and support for further basic research in this technology is to continue.

**Jana Albrechtova**  
Dept. Plant Physiology, Fac. Sci. Charles University  
Vinicna 128 44 Praha 2  
Czech Republic  
Fax : 420 2 21953306  
Email : albrecht@mail.natur.cuni.cz

**Maria do Rosario Alvelos Alegre Marques**  
Universidade de Coimbra Departamento de Botanica  
3049 Coimbra Codex  
Portugal  
Fax : 351 039820780  
Email : mra@condor.ci.uc.pt

**Maiгуill Appelgen**  
Department of Horticulture and Crop Sciences  
P.O. Box 5022  
N-1432 Aas  
Norway  
Fax : 47 64 94 78 02  
Email : maiгуill.appelgen@ipf.nih.no

**Jean-Charles Bastien**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex France  
Fax : 33 2 38 41 78 79  
Fax : 33 2 38 41 78 79  
Email : bastien@orleans.inra.fr

**Michael R. Bewar**  
Westvaco Forest Science and Technology  
PO Box 1950, 180 Westvaco road  
Summerville, SC 29484  
USA  
Fax : 1 803 875 7185  
Email : mrbecwa@westvaco.com

**Krystyna Bojarczuk**  
Institute of Dendrology  
62-035 Kornik  
Poland  
Fax : 48 61 8170166  
Email : idkornik@rose.man.poznan.pl

**Marc Bonnet-Masimbert**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex France  
Fax : 33 2 38 41 78 79  
Email : masimbert@orleans.inra.fr

**Christian Breton**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex France  
Fax : 33 2 38 41 78 79  
Email : breton@orleans.inra.fr

**Maurizio Capuana**  
Istituto Miglioramento Genetico Pianta Forestali  
V. San Bonaventura 13  
50145 Firenze  
Italy  
Fax : 39 055 486604  
Email : ist\_selv@CESITI.UNIFI.IT

**Marc-Philippe Carron**  
CIRAD  
Av. agropolis BP 5035  
34 032 Montpellier Cedex 1  
France  
Fax : 33 4 67 61 55 96  
Email : carron@cirad.fr

**Pawel Chmielez**  
Polish Academy of Sciences Institut of Dendrology  
62-035 Kornik Parkowa 5  
Poland  
Fax : 48 61 8170166  
Email : idkornik@rose.man.poznan.pl

**David Clapham**  
Swedish Univ. Agric. Sci., Dept of Forest Genetics  
Box 7027, S-75007 Uppsala  
Sweden  
Fax : 46 18 67 32 79  
Email : David.Clapham@sgen.slu.se

**Daniel Cornu**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20619  
Ardon F-45166 Olivet Cedex France  
Fax : 33 2 38 41 78 79  
Email : cornu@orleans.inra.fr

**Hélène David**  
Laboratoire de Physiologie des Parois Végétales  
Université de Lille I, Bât. SN2 2ème étage  
59 655 Villeneuve d'Ascq  
France  
Fax : 33 3 20 33 63 02  
Email : helene.david@univ-lille1.fr

**David Dunstan**  
Crop Science Department  
Horticulture Research International  
East Malling, West Malling Kent ME19 6BJ  
England  
Fax : 44 1732 849 067  
Email : David.Dunstan@hri.ac.uk

**Ulrika Egertsdotter**  
NISK  
Høgskoleveien 12, N-1432 As  
Norway  
Fax : 47 64 94 29 80  
Email : Ulrika.Egertsdotter@nisk.no



**Julie Fauve**  
Michelin, lab. CPN Place des Carmes  
63 040 Clermont-Ferrand  
France  
Fax : 04 73 32 75 31

**Helena Fernández González**  
Lab. F. Vegetal Dpto BOS  
Faculta de Biología Universidad de Oviedo  
c/Catedrático Rodrigo Uria s/n.  
Oviedo E.33071  
Spain  
Fax : 34 985104867  
Email : helenaf@sci.cpd.uniovi.es

**Jens Find**  
Tissue Culture Laboratory, Botanical Garden,  
DK-1353 Copenhagen K, Denmark  
E-mail : JensF@Bot.Ku.DK  
Fax : 45 3532 2221

**Bruno Florin**  
Centre de Recherche Nestlé  
101 av. Gustave Eiffel  
37 390 Notre Dame d'Oé  
France  
Fax : 33 2 47 49 14 14  
Email : bruno.florin@rdto.nestle.com

**Jean-Luc Fourré**  
UCL-Unité des Eaux et Forêts  
Place Croix du Sud 2 bte 9  
B-1348 Louvain-la-Neuve  
Belgium  
Fax : 32 10 47 36 97  
Email : fourre@efor.ucl.ac.be

**Elisabeth Garin**  
AFOCCEL-Station de Biotechnologies  
Domaine de l'Etangon  
77370 Nangis  
France  
Fax : 33 1 60 67 54 04

**Inger Hakman**  
Dept. of Botany Stockholm University  
S-106 91 Stockholm  
Sweden  
Fax : 46 8-16 55 25  
Email : inger.hakman@botan.su.se

**Luc Harvent**  
AFOCCEL-Station de Biotechnologies  
Domaine de l'Etangon  
77370 Nangis France  
Fax : atobio@afocel.fr  
Email : 33 1 60 67 54 04

**Hely Häggman**  
Finnish Forest Research Institute  
Punkaharju Research Station  
Finlandiantie 18, FIN-58450 Punkaharju,  
Finland  
Fax : 358 15 644333  
Email : hely.haggman@metla.fi

**Christian Jay-Allemand**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex  
France  
Fax : 33 2 38 41 78 79  
Email : jay@orleans.inra.fr

**Josiane Julien**  
Michelin, lab. CPN Place des Carmes  
63 040 Clermont-Ferrand  
Fax : 04 73 32 75 31  
Email : jjulien@francemultimedia.fr

**Krystyna Klimaszewska**  
Natural Resources Canada, Canadian Forest Service  
Laurentian Forestry Center, 1055 rue du P.E.P.S.  
CP 3800 Ste Foy Quebec, G1V 4C7  
Canada  
Fax : 1 418 648 5849  
Email : KKKlimaszewska@exchange.cfl.forestry.ca

**Philippe Label**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex  
France  
Fax : 33 2 38 41 78 79  
Email : label@orleans.inra.fr

**Marie-Anne Lelu**  
Research Unit on Genetic, Physiology and  
Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex France  
Fax : 33 2 38 41 78 79  
Email : lelu@orleans.inra.fr

**Céline Le Pichon**  
CEMAGREF  
Division Ressources Génétiques et Plants Forestiers  
Domaine des Barres  
45 290 Nogent sur Vernisson France  
Fax : 33 2 38950346  
Email : lepichon@rainbow.cemagref.fr

**Helena Lipavska**  
Dept. Plant Physiology, Fac. Sci. Charles University  
Vinicna 128 44 Praha 2  
Czech Republic  
Fax : 420 2 2195 3306  
Email : lipavska@mail.natur.cuni.cz

**Nicole Michaud-Ferrière**

Histologie BIOTROP  
CIRAD BP 5035

F-34032 Montpellier France  
Fax : 33 4 67 61 55 96

Email : ferriere@cirad.fr

**Simon Moorhouse**

Crop Science Department

Horticulture Research International  
East Malling, West Malling Kent ME19 6BJ  
England

Fax : 44 1732 849 067

Email : simon.moorhouse@hri.ac.uk

**Karoliina Niemi**

Dept. of Ecology and Environmental Science

University of Kuopio B.O.X 1627  
FIN-70211 Kuopio Finland

Fax : 358 17 163 230

Email : karoliina.niemi@uku.fi

**Luc Pâques**

Research Unit on Genetic, Physiology and

Improvement of Forest Trees INRA-CRO BP 20619  
Ardon F-45166 Olivet Cedex France

Fax : 33 2 38 41 78 79

Email : paques@orleans.inra.fr

**Marc Pâques**

AFOCEL-Station de Biotechnologies

Domaine de l'Étangon  
77370 Nangis France

Fax : paques@afocel.fr  
Email : 33 1 60 67 54 04

**Vincent Pétiard**

Centre de Recherche Nestlé

101 av. Gustave Eiffel  
37 390 Notre Dame d'Oé France

Fax : 33 2 47 49 14 14

**Gilles Pilate**

Research Unit on Genetic, Physiology and

Improvement of Forest Trees INRA-CRO BP 20 619  
Ardon F-45166 Olivet Cedex France

Fax : 33 2 38 41 78 79

Email : pilate@orleans.inra.fr

**Amely Potes**

Dto. de Biologia

Nucleo da Mitra Ap. 94  
7001 Évora Codex Portugal

Fax : 351 667409821

Email : apotes@uevora.pt

**Aro Ramarosandratana**

AFOCEL-Station de Biotechnologies

Domaine de l'Étangon  
77370 Nangis France

Fax : 33 1 60 67 54 04

**Josef Schmidt**

Biotechnology Unit

Austrian Research Centers  
A-2444 Seibersdorf

Austria

Fax : 43 2254 780 3653

Email : schmidt@zdfs.arcs.ac.at

**Bjorn Sundberg**

Dept. of Forest Genetics and Plant Physiology

Swedish University of Agricultural Sciences  
901 83 Umea  
Sweden

Fax : 46-90-786 5901

E-mail : Bjorn.Sundberg@genfys.slu.se

**Krystyna Szczygiel**

Forest Research Institute

Section of Seed Science and Selection  
05-550 Raszyn Sekocin, k/Warszawy  
Poland

Fax : 48 22 8 22 49 35

Email : rakowskk@ibles.waw.pl

**David Thompson**

Coillte Teoranta - The Irish Forestry Board

Research Laboratory Newtownmounkennedy  
County Wicklow  
Ireland

Fax : 353 1281 0465

Email : btdetal@gpo.iol.ie

**Mariano Toribio**

Instituto Madrileño de Investigación

Agraria y Alimentaria (IMIA)  
Apdo. 127, 28800 Alcalá de Henares, Madrid.

Fax : 34 91 8880726

Email : encmti@encin.alcala.es

**Martin Wagner**

Institute of Experimental Botany ASCR

Rozvojova 135, CZ-16502 Prague 6,  
Czech Republic

Fax : 42 02 20390419

Email : wagner@ueb.cas.cz

**Jean-Luc Verdeil**

Orstom genetrop

BP 5045 911 av. Agropolis  
34032 Montpellier Cedex 1 France

Fax : 33 4 67 54 78 00

Email : verdeil@mpl.orstom.fr

**Kurt Zoglauner**

Humboldt University, Institut of Biology

Invalidenstr. 43  
D-10115 Berlin Germany

Fax : 49 30 20 93 87 25

Email : h1299egj@rz.hu-berlin.de