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Maïa Meurillon, Magaly Angenieux, Frédéric Mercier, Patrick Blinet, Laurent Chaloin, et al.. Mitigation of heterocyclic aromatic amines in cooked meat. Part I: Informed selection of antioxidants based on molecular modeling. Food Chemistry, 2020, 331, pp.127264. 10.1016/j.foodchem.2020.127264. hal-02890485

## HAL Id: hal-02890485 https://hal.inrae.fr/hal-02890485v1

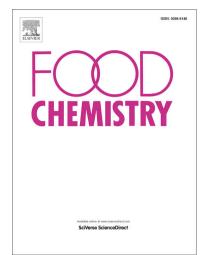
Submitted on 11 Nov 2020  $\,$ 

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| PII:           | S0308-8146(20)31126-2                          |
|----------------|--|
| DOI:           | https://doi.org/10.1016/j.foodchem.2020.127264 |
| Reference:     | FOCH 127264                                    |
| To appear in:  | Food Chemistry                                 |
| Received Date: | 5 February 2020                                |
| Revised Date:  | 4 June 2020                                    |
| Accepted Date: | 4 June 2020                                    |



Please cite this article as: Meurillon, M., Angenieux, M., Mercier, F., Blinet, P., Chaloin, L., Chevolleau, S., Debrauwer, L., Engel, E., Mitigation of heterocyclic aromatic amines in cooked meat Part I: Informed selection of antioxidants based on molecular modeling, *Food Chemistry* (2020), doi: https://doi.org/10.1016/j.foodchem. 2020.127264

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|          | Journal Pre-proofs  |
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| 1        | Mitigation of heterocyclic aromatic   |
| 2        | amines in cooked meat   |
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| 4        | Part I: Informed selection of antioxidants  |
| 5        | based on molecular modeling   |
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#### 26 Abstract:

This work aimed to develop a method permitting an informed choice of antioxidants to reduce 27 28 carcinogenic heterocyclic aromatic amine (HAA) formation during proteinaceous food 29 cooking. Therefore, a three-step approach was developed. First, the most promising 30 antioxidants were selected using molecular modeling approaches. For this, analog design was 31 used to highlight the most suitable antioxidants based on their diversification potential using bioisosteric replacement. Then, structure activity relationship studies allowed drawing the 32 33 relevant properties for inhibiting HAA formation and explained partly the inhibitory activity. 34 Secondly, the approved antioxidants were tested in ground beef patties to assess their inhibitory properties against HAA formation. Resveratrol was found to be the most efficient 35 36 as it totally inhibited MeIQ and reduced MeIQx and PhIP formation by 40 and 70%, 37 respectively. Finally, natural ingredients rich in these antioxidants were evaluated. Oregano 38 was found to totally inhibit MeIQ formation and to reduce by half MeIQx and PhIP formation. 39

40

41 Keywords: Heterocyclic aromatic amines, mitigation, antioxidants, molecular modeling,
42 analog design, Structure Activity Relationship (SAR), formulation.

43

#### 44 **1. Abbreviations**

Heterocyclic Aromatic Amine (HAA); 2-amino-3,4,8-trimethyl-imidazo[4,5-*f*]quinoxaline
(4,8-DiMeIQx); 2-amino-3-methyl-imidazo[4,5-*f*]quinoline (IQ); 2-amino-3,4-dimethylimidazo[4,5-*f*]quinoline (MeIQ); 2-amino-3,8-dimethyl-imidazo[4,5-*f*]quinoxaline (MeIQx);
2-amino-1-methyl-6-phenyl-imidazo-[4,5-*b*]pyridine (PhIP); 2-amino-3,4,7,8-tetramethyl-3*H*imidazo[4,5-*f*]quinoxaline (TriMeIQx).

50

#### 51 **2. Introduction**

52 The preparation and cooking of meat products impact their microbial load, palatability and 53 composition. However, some of these compositional changes such as the formation of 54 heterocyclic aromatic amines (HAAs) have been linked to the promotion of certain cancers (e. 55 g., colorectum, breast, prostate) in meat consumers (Zheng & Lee, 2009). Recently, the 56 International Agency for Research on Cancer classified red meat and processed meat as probably carcinogenic (group 2A) and carcinogenic (group 1), respectively (International 57 58 Agency for Research on Cancer, 2018). To reduce HAA impact on human health, mitigation 59 strategies were proposed. Today, one of the most promising approaches is to act on the formulation of red meat products by adding natural antioxidants that limit the formation of 60 61 HAAs (Meurillon & Engel, 2016). Although there is a large variety of antioxidants that can reduce HAA formation in meat (Lee et al., 2020), their choice remains mostly empirical. 62 63 Indeed, the antioxidants tested for HAA inhibition are often chosen arbitrarily because they 64 already display other medicinal properties (Tengilimoglu-Metin, et al., 2017) and are widely 65 used in cooking in a given region. Research works often focused on a particular type of meat 66 or on formulations based on cultural or geographical considerations such as green tea 67 formulation for Chinese research works (Khan, et al., 2017).

To our knowledge, no method for making a reasoned choice exists to date and therefore, there 68 69 is a real need to develop a well-reasoned mitigation strategy for choosing the best ingredients 70 or preferred diets. The present work hypothesized that medicinal chemistry approaches could help in choosing antioxidants and in explaining their reactivity against HAA formation. A 71 72 unique and original method based on medicinal chemistry knowledge was therefore 73 developed to choose antioxidants best suited to inhibit HAA formation. This method allowed 74 a rationalization of the choice of antioxidants, which is novel and could be subsequently 75 transposed to other process-induced toxicants such as polycyclic aromatic hydrocarbons. In

76 the first part of the paper, an approach adapted from medicinal chemistry and widely used to choose and optimize lead compounds with the help of molecular modeling methods 77 (Andricopulo, Salum, & Abraham, 2009; Bohacek, McMartin, & Guida, 1996) was 78 implemented in order to select potent antioxidants for the inhibition of HAA formation. The 79 80 rationale was to show which known antioxidants would be the most suitable for drug 81 development by the implementation of analog design strategies which rested upon the improvement of existing active molecules to get rid of some side-effects or unwanted 82 83 properties of the original compound. Analog design (Wermuth, 2010) permitted the discovery 84 of direct analogs (analogs possessing structural and functional similarities), structural analogs (analogs possessing only chemical similarities but presenting different pharmacological 85 86 properties) and functional analogs (chemically different compounds displaying similar 87 pharmacological properties). Following this selection step, other molecular modeling methods 88 were used to draw the relevant properties required for a good inhibition of HAA formation 89 and explain their reactivity. Then, in the second part, the different antioxidants put forward by 90 the analog design approaches were tested at different concentrations in meat samples in real 91 conditions, using a widely consumed meat product (ground beef patties) prepared by a 92 common cooking method (pan frying) according to WHO recommendations. In the final part, 93 natural ingredients known to be rich in the previous antioxidants were tested under similar 94 conditions in order to take into account complex systems more representative of common culinary practices. 95

96

#### 97 **3. Materials and Methods**

#### 98 **3.1.** Chemicals and standards

99 Dichloromethane, acetonitrile and methanol were organic trace analysis grade solvents100 (Sigma-Aldrich, France). Diatomaceous earth used for the preparation of accelerated solvent

101 extraction (ASE) cells was obtained from Sigma-Aldrich. Five HAA standards and TriMeIQx 102 used as internal standard were obtained from Toronto Research Chemicals (North York, 103 Canada). Resveratrol 99% and quercetin hydrate 95% were bought from Acros Organics 104 (Geel, Belgium). Carvacrol (>98%, FC, FG) and (-)-epicatechin (>90%, HPLC) were bought 105 from Sigma-Aldrich.

106

#### 107 3.2. Molecular modeling and structure--activity relationship (SAR) studies

108 A bibliographic research was performed to identify the active principles known as inhibitors 109 of HAA formation in order to achieve molecular modeling to select the best antioxidants 110 among them (Meurillon & Engel, 2016). A list of 17 compounds was obtained (figure 1).

111

112 To rationalize the selection of the antioxidants for further testing, analog design approaches 113 were used. This strategy relied on the "drug-like" concept (or thereafter the "lead-like" 114 concept) in medicinal chemistry (Lipinski, 2004). To choose between different active 115 compounds having the same biological target, medicinal chemists are looking for their 116 potential of structural diversification by bioisosteric replacement of functional groups, for 117 instance. This would permit in case of synthesis problems, unwanted side-effects or even 118 intellectual property matter to be able to develop quickly and efficiently a new drug with 119 similar activity from the pipeline of the initial drug analogs.

120

Search for structural analogs based on Tanimoto coefficient. To find new analogs of a drug or 121 as in the present study, antioxidants or natural active principles sharing equivalent inhibitory 122 123 properties against HAA formation, the most straightforward way is to find new entities with 124 highly similar chemical structures to that of the original compound. Binding and mode of action should be preserved between analogs and original compound. To this end, analogs 125

126 based on the high similarity of structural fingerprints measured by the Tanimoto coefficient 127 (describing the ratio of the shared structural elements in respect to the total number of 128 structural elements present in both molecules, Rogers & Tanimoto, 1960) were searched. This 129 strategy was applied here on the 17 already proven antioxidants (Figure 1) by screening the 130 ZINC (ZINC Is Not Commercial) database (Figure 2a) (Sterling & Irwin, 2015). The ZINC 131 database encompasses about 230 million purchasable compounds coming from different 132 databanks (natural products, synthetic compounds from chemical companies and Food & 133 Drug Administration (FDA) approved drugs). The rationale behind this step was that more 134 screened compounds (structurally similar to a studied antioxidant) would lead to more chance 135 of discovering a potent analog (i.e. potent HAA inhibitor). The studied antioxidant with the 136 most analogs would be therefore the most interesting from a biomolecular point of view, as it displayed a large panel of analogs with potentially the same reactivity against HAA 137 138 formation. This step only relied on atomic composition and the fact that structurally similar molecules would bind to similar targets and therefore display the same biological activity. It 139 140 permitted the assessment of a number of referenced compounds (in this case from databanks 141 of natural products and FDA approved products) that presented similar structural fingerprints 142 with the studied antioxidants and therefore determined the potential extent of such family to 143 inhibit HAA formation. The Tanimoto index used for this search was set to 0.8, which 144 indicated 80% of substructure homology between two molecules and then expanded to 0.7 to 145 broaden the scope of potentially relevant molecules.

146

*Functional analog search based on scaffold hopping.* To increase the diversity of active molecules with the aim to maintain the inhibitory activity while avoiding side effects, the solution was to look for functional analogues of the known antioxidants. For this purpose, scaffold hopping was used in the present work. This method consisted in the search for

151 compounds with different core structures but sharing a global 3D shape very similar to the 152 original compound. In medicinal chemistry it allows alternative starting points for lead 153 optimization or to generate backup candidates for advanced compounds. For this purpose, the 154 program LigCSRre (Ouintus, Sperandio, Grynberg, Petitjean, & Tuffery, 2009; Sperandio, Petitjean, & Tuffery, 2009) implemented on the RPBS (Ressource Parisienne en 155 156 Bioinformatique Structurale) platform (Alland, et al., 2005) (Figure 2b) combining 3D 157 maximal substructure search and molecular similarity was used. A databank of 2048 active 158 compounds was screened. It permitted determination of a range of new molecules from 159 different families potentially able to show an inhibitory effect on HAA formation. The 160 LigCSRre software returned a list of compounds ranked by similarity scores and a specific 161 file containing the best compounds superimposed onto the query (as to know the studied 162 antioxidants). The similarity score rested especially upon a calculated value, the Zscore, based 163 on the number of shared bonds and considered significant above 2. In this work, Zscore value was deliberately chosen above 2 in order to only restrict to the best results. However, for 164 165 antioxidants for which the query did not return results above 3.01, the Zscore was decreased 166 to 2 to evaluate the reply quality at this level of similarity (Supplementary Data 2).

167

168 Pharmacophore models by OSAR. Having identified the most relevant antioxidants, the 169 following step was to explain the reasons why they displayed such inhibitory activity. That 170 implied studying chemical scaffold and associated functional groups. This approach used the 171 pharmacophore concept. A pharmacophore is an abstract description of molecular features 172 that are necessary for molecular recognition of a ligand by a biological macromolecule. In 173 medicinal chemistry, the establishment of a pharmacophore model allows definition of all 174 parts of a molecule that are necessary for its biological activity. A pharmacophore model explains how structurally diverse ligands can bind to a common receptor site. Knowing that, it 175

becomes easier to develop new analogs possessing the same targeted activity. Typical 176 pharmacophore features include hydrophobic centroids, aromatic rings, hydrogen bond 177 178 acceptors or donors, cations and anions. The features need to match different chemical groups 179 with similar properties, in order to identify novel ligands. A well-defined pharmacophore 180 model includes both hydrophobic volumes and hydrogen bond vectors. In computational 181 chemistry, pharmacophores are used to define the essential features of one or more molecules 182 with the same biological activity and serve as the starting point for developing 3D-QSAR 183 models useful in drug discovery. In this research work, the pharmacophore model was built 184 after superposing validated antioxidants from the same structural family using the ZINC 185 PHARMER tool (Koes & Camacho, 2012). The model was created based on physicochemical 186 properties of the selected antioxidants as to know hydrophobicity, aromaticity, donor or 187 acceptor of hydrogen bonds, ionic or electrostatic bonds and used for screening the ZINC 188 database (Figure 2c).

189

190 Electrostatic potential surface computation. To increase the chance of discovering highly 191 similar compounds, another important component was added, the electrostatic potential 192 surface distribution. Indeed, to be able to compare different structurally similar compounds to 193 our selected antioxidants, the electrostatic surface was representative of both the electronic 194 distribution within aromatic cycles and partial charges of each atom. For structure-based drug 195 design, the visual representation of electrostatic potentials provides insights into protein-196 ligand interactions, and hence can play a major role in elucidating the relationship between 197 structure and reactivity of a biomolecular system. Nowadays, colorful plots of electrostatic 198 potential are used in drug design to rationalize trends in organic reactivity and binding in 199 host-guest complexes. In the present work, the different antioxidant molecules were modeled with Vega ZZ version 3.1 which allowed in particular charge assignment and energy 200

minimization necessary for the computation of surface electrostatic potentials. This was
computed using APBS ("Adaptive Poisson-Boltzmann Solver") plugin tool implemented in
the PyMol Molecular Graphics System, Version 1.3, Schrodinger, LLC. From this calculation,
the positive and negative isosurfaces were contoured at +5 and -5 kT/e, respectively (Figure
205 2d).

206

207 **3.3.** Meat sample preparation

Ground beef was obtained from Société Convivial (Gannat, France). The meat was certified
from Charolais cattle and contained 11% fat. Stored at -80 °C, it was thawed overnight in the
fridge at 4 °C one day before use.

211 The four natural products rich in phenolic antioxidants were found at the local supermarket: 212 'Extra Fine Capers' from Maille (France), Pinot Noir Burgundy wine bottles, organic green 213 tea in bags from Naturela and dried oregano from Ducros. They were added in two ways: 214 marinated or blended in. For red wine and green tea samples, the meat patties were fully 215 marinated in each solution for 2, 4 or 6 hours. For the caper and oregano samples, as well as 216 for the four pure chemical compounds, the necessary amount of antioxidant (0.1%, 0.25% and 217 0.5% mass concentration) was first manually ground in a stainless steel mortar and then added 218 to meat patties and mixed four times for 20 seconds. After mixing, patties of 26 g of the same 219 dimension were shaped with a round stainless steel ring. The patties were vacuum sealed and 220 stored at -18 °C.

Marinade durations were chosen according to literature data on marinated meat (Busquets, Puignou, Galceran, & Skog, 2006; Melo, et al., 2008) to induce a decrease in HAAs while being realistic with common household practices. Concentrations in active principles and natural ingredients were chosen to be realistic from a consumer point of view: high enough to 225 display a potential inhibitory effect based on literature data (Gibis & Weiss, 2012; Friedman,

Zhu, Feinstein, & Ravishankar, 2009) but low enough to be accepted in meat recipes. 226

227 For each formulation and concentration, 3 samples were prepared and further cooked and 228 tested for HAA levels.

229

230

#### 3.4. Cooking method

231 To study the formation of HAAs during cooking, ground beef patties were cooked in a 232 stainless steel frying pan (17 cm diameter) on a controlled-temperature induction hob (Bosch 233 Electroménager, Saint-Ouen, France) according to the method described by Planche et al. 234 (2017). A sheet of 11 µm thick aluminum foil was laid on the bottom of the frying pan to recover juice released during meat cooking. Cooking conditions were used to simulate 235 236 medium meat (core 70 °C, according to WHO recommendations for ground meats). 237 According to Planche et al. (2017), these cooking conditions corresponded to 14 min heating (turned over three times) at 200 °C at the bottom of the pan. Temperatures at the core of the 238 239 meat and at the bottom of the pan were monitored by thermocouples (RS Components, 240 Beauvais, France).

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#### 242

#### 3.5. Quantification of HAAs in ground beef patties

Five different HAAs (IQ, MeIQ, MeIQx, 4,8-DiMeIQx, PhIP), likely to be found in meat, 243 244 were determined by ultra-high-performance liquid chromatography-atmospheric pressure 245 chemical ionization-tandem mass spectrometry (UHPLC-APCI-MS/MS) based on a liquid 246 chromatography-atmospheric pressure chemical ionization-tandem mass spectrometry (LC-247 APCI-MS/MS) method developed for chicken meat (Chevolleau, Touzet, Jamin, Tulliez, & 248 Debrauwer, 2007) and since adapted for analyzing beef samples (Kondjoyan, Chevolleau, Greve, et al., 2010). The choice of APCI was based on a previous work (Chevolleau, S., et al., 249

2007) which showed that positive APCI was better suited than ESI for the detection of 10
HAAs. These results were in agreement with other published works (Guy, Gremaud, Richoz,
& Turesky, 2000) and were further confirmed for all the studied HAAs. However, it should be
noted that several papers also describe ESI-based analytical methods for HAA analysis, since
ESI is much more widespread than APCI in laboratories.

255

256 Briefly, 1 g of lyophilized ground beef sampled from the whole cooked sample was first 257 treated with 1 M NaOH for protein denaturation. After accelerated solvent extraction (ASE) 258 (addition of 11 g of diatomaceous earth, extraction with dichloromethane) and purification of 259 the extract, the resulting residue was re-dissolved in 200  $\mu$ L of the starting LC mobile phase. 260 Separation was performed on an Accela 600 LC system (Thermo Fisher Scientific, France), 261 using a Hypersil GOLD C8 column (Thermo Fisher Scientific,  $50 \times 2.1$  mm,  $1.9 \mu$ m) with 262 prefilter. Five microliters of the final extract were injected, and separation was performed at 30 °C at a flow rate of 0.5 mL/min. Chromatographic separation was achieved using mobile 263 264 phases composed of A: AcONH<sub>4</sub> (30 mM, pH 6) / CH<sub>3</sub>CN/CH<sub>3</sub>OH (2/1) (90/10, v/v) and B: 265 AcONH<sub>4</sub> (30 mM, pH 6) / CH<sub>3</sub>CN/CH<sub>3</sub>OH (2/1) (10/90, v/v). The elution gradient was as 266 follows: 0 min 0% B, 0.24 min 6% B, 0.98 min 22% B, 1.86 min 22% B, 2.30 min 50% B, 2.74 min 50% B, 3.18 min 100% B, 4.21 min 100% B, 4.50 min 0% B. MS detection was 267 268 performed on a TSQ Vantage triple-stage quadrupole mass spectrometer (Thermo Fisher Scientific) 269 using positive APCI based on two specific transitions for each HAA. Typical working parameters 270 were as follows: discharge APCI current, 3 µA; nebulizer temperature, 420 °C; heated transfer 271 capillary temperature, 260 °C; sheath gas flow rate, 35 a.u.; auxiliary gas flow rate, 10 a.u. The 272 performance of the optimized method was characterized in terms of linearity ( $r^2 > 0.98$  between 1 and 273 500 pg/µL) with relative concentration residual deviation lower than 15% (20% LoQ-level) for all the 274 concentration levels of the calibration curve), measure repeatability (CV>15%) and relative bias 275 ( $\pm 15\%$ ), limit of detection (LoD) (0.1–0.3 pg/µL) and limit of quantification (LoQ) (1.0 pg/µL). The

276 LoD was defined as the lowest concentration of analyte that could be detected with acceptable 277 chromatographic peak shape, with quantification and confirmation ions present with a signal-278 to-noise ratio (S/N) greater than 3. The LoQ was the lowest concentration that met the LoD 279 criteria, but with a S/N of 10 and both bias% and relative standard deviation (RSD) below 280 20%. Both the LoD and LoQ have been precisely determined for each of the HAAs. However, 281 for convenience reasons, the LoQ was set at the value of 1  $pg/\mu L$ , which was common to all the 5 HAAs and corresponded to the lower point of the calibration range. Since APCI may be 282 283 subject to signal suppression or enhancement due to co-extracted matrix constituents, 284 evaluation of matrix effects (ME) was carried out by comparing the calibration curves 285 obtained by a calibration range in the solvent and in the matrix (n = 3). ME (%) = ((matrix 286 range slope/solvent range slope) -1 × 100. In this context, matrix effects were interpreted as 287 follows: negative and positive values for ME highlighted matrix-induced suppression and 288 enhancement, respectively. Between -20% and 20% the ME was considered small, between -289 50% and -20% or 20% and 50% the effect was medium, and for values of ME below -50% or 290 higher than 50% the effect was considered strong. The matrix effects precisely determined for 291 each of the HAAs were less than 20% and could be considered negligible. Each cooked meat sample 292 was extracted and analyzed in triplicate. HAA contents were expressed in nanograms of compound per 293 gram of freeze-dried product.

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295

#### **3.6.** Data analysis

Data were analyzed using Statistica software 10.0 (Statsoft, Maisons-Alfort, France). To determine whether the formulation (antioxidant or natural ingredient) had an effect on the level of HAAs in cooked meat, a one-way analysis of variance (ANOVA; p < 0.05) was performed on data from UHPLC–APCI-MS/MS analyses. A Dunnett mean comparison test was then performed on the resulting dataset, to determine which formulation concentration was distinct from the control sample (p < 0.05). When a significant difference was found, a 302 Duncan mean comparison test was performed on the dataset to differentiate the concentrations 303 of a given antioxidant (p < 0.05).

- 304
- 305 **4. Results and Discussion**

#### 306 4.1. Choice of relevant antioxidants by molecular modeling

The first part of this study was dedicated to adapting methods used for drug design in medicinal chemistry to select promising antioxidants and explain their inhibitory activity. The rationale behind this approach was, given known inhibitors of HAAs, **1**) which ones would be retained as good drug candidates in medicinal chemistry and **2**) how to explain their inhibitory activity.

Figure 1 shows a list of 17 antioxidants of different families chosen for this study based on a
previous review work of literature data on antioxidants known to be HAA inhibitors
(Meurillon & Engel, 2016).

The next step relied on the "drug-like"/"lead-like" concept of medicinal chemistry. The 315 316 rationale to choose antioxidants for HAA inhibition would therefore be linked to their 317 potential diversification. This suggested that many other natural or synthetic compounds 318 could play a similar inhibitory role than these known antioxidants. The selected strategy was 319 based on analog design approaches consisting of modifying an existing active molecule in 320 order to prepare a new molecule showing chemical and/or biological similarity to the original 321 model compound (with the aim for example in the present case to limit the impact on meat 322 organoleptic qualities and to deal with consumer acceptance issues). Thus, the more an 323 antioxidant would display analogs, the more it would be suitable for further inhibition 324 experiments. Indeed, an antioxidant inhibitor of HAA formation may present an aroma default 325 in mixture with meat and therefore may not be accepted by consumers; such strategy would

326 permit discovery of other compounds as inhibitors but less problematic from an organoleptic

327 point of view. The results from analog design approaches are presented in Table 1.

328

Concerning *closely-related analogs*, Figure 1 already displayed such compounds among known inhibitors of HAA formation. Epigallocatechin and epicatechin are direct analogs as they display structural and functional similarities, as do cafestol and kahweol. However, if the success likelihood (i.e. inhibitory effect against HAAs) is relatively high using one or the other direct analog, the chance to circumvent secondary effects would be low as the molecules are similar.

335

336 Structural analogs for the different antioxidants studied are displayed in Table 1. These 337 results already showed that rutin and allicin could be discarded from this study as they 338 displayed a poor diversification potential (i.e. no structural analogs) and therefore a poor "drug-like" potential. The detailed results obtained for all the targeted antioxidants were 339 340 reported in Supplementary Data 1. As an example, the similarity search done for quercetin 341 gave a list of 26 different referenced compounds. The variability was mostly displayed by the 342 presence of other functional groups such as chloride-like in ZINC299830137 or hydroxyl-like 343 in myricetin (ZINC3874317) or the replacement of one hydroxyl of quercetin by another 344 reactive group such as sulfate-like in quercetin 3'-sulfate (ZINC14644472) or methoxy like in 345 tamarixetin (ZINC6484604). This study evidenced that a lot of close compounds existed with 346 little modification as the main differences lied in functional groups and substituting positions. 347 This suggested that many other natural or synthetic compounds could play a similar inhibitory 348 role than these known antioxidants with however the inconvenience that the 3D structure was 349 not taken into account.

350

351 *Functional analogs* were then searched for and are also presented in Table 1. These analogs could display some equivalent functionalities (such as potentially similar inhibitory effects) 352 353 with wide structural diversity. The method used was scaffold hopping which considered 3D 354 structural similarity. Table 1 summarizes the data found for the different antioxidants studied 355 and confirms the conclusion of the previous step, that rutin and allicin could be discarded for 356 the rest of the study since they do not present any functional analogs. Carnosol, carnosic acid, 357 rosmanol, cafestol, kahweol, epigallocatechin gallate, vitamin E, pyridoxamine and diallyl 358 sulfide can also be dismissed from the study as they did not display results above 3.01 (and 359 few above 2). Supplementary Data 2 show the detailed results obtained for all the targeted 360 antioxidants. As an example, guercetin displayed 4 compounds with a Zscore above 3.01. 361 From the 3D representation, it appeared that the similar active site was the bicyclic part of the 362 molecule as the superimposition of the antioxidant and the found compounds was optimal for 363 this part of the molecule and of less good quality for the other parts.

364

Therefore, based on the similarity search and the scaffold hopping step, resveratrol, luteolin, quercetin, carvacrol, epigallocatechin and epicatechin displayed the most "drug-like" potential as they presented a large capacity for improvements given the number of structural and functional analogs they possessed.

369

To determine the relevant properties for a good inhibitory potential, pharmacophore studies were realized based on the antioxidant structures the most "drug-like" (i.e. resveratrol, luteolin, quercetin, carvacrol, epigallocatechin and epicatechin). The aim of this step was to define which functional groups were necessary for antioxidants to exhibit HAA inhibitory properties, which came back to define the functional groups necessary for molecular recognition of a ligand by a biomolecule. Using dedicated software, the pharmacophore study

376 was done for quercetin and luteolin (Supplementary Data 3) with the use of different 377 databases of bioactive products. The different properties of these two antioxidants focusing on 378 hydrophobicity, aromaticity, donor or acceptor of hydrogen bonds, ionic or electrostatic bonds 379 were studied by running the software with these data. This highlighted that these two 380 antioxidants were described by the same pharmacophore model and most importantly that 381 three hydrophobic regions and two of hydrogen bonds were the basis of this pharmacophore 382 model (Supplementary Data 3). This was most probably a prerequisite to promote significant 383 inhibitory activities against HAA formation.

384

385 To try to explain the reactivity of the selected antioxidants, their electrostatic potential 386 surfaces were computed (Supplementary Data 4). Bicyclic scaffolds of both quercetin and 387 luteolin displayed an important positive electrostatic potential in opposition to the *meta* 388 hydroxyl groups which were strongly electronegative. So finally, quercetin and luteolin 389 displayed a similar representation of electrostatic field thus explaining their similar reactivity. 390 Both quercetin and luteolin belong to the flavonoid family of polyphenolic antioxidants, are 391 among the most prevalent in fruits and vegetables and among the most potent flavonoids from 392 natural origin for their *in vitro* biological activity. They also displayed similar pharmacophore 393 results and electrostatic potential representation. Therefore, to select which one will be 394 studied in the rest of this research work, the price argument was used. Looking at Sigma-395 Aldrich quotation for these two active principles, it appeared that quercetin was much more 396 available (different kinds of packaging, of purity, etc.) and more than a thousand times less 397 expensive than luteolin for a similar purity (>95%). Quercetin was therefore selected for the 398 rest of this research work.

Concerning the choice of catechin antioxidant for the next step of the study, it appeared thatepicatechin was most promising as it displayed better results in term of similarity search and

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401 scaffold hopping results. From a pecuniary point of view, epicatechin also remained the less
402 expensive. Epicatechin was therefore selected for the following steps of the study.

403 The selection of carvacrol and resveratrol was less ambiguous as they were the only 404 representatives of their analog family.

405 As the result of these molecular modeling approaches, four antioxidants were selected to be 406 tested for the inhibition of HAA formation: carvacrol, resveratrol, (–)-epicatechin and 407 quercetin.

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#### 409 4.2. Assessment of the selected antioxidants on HAA formation inhibition

The inhibitory properties against HAA formation of the four antioxidants given by the analog design approaches were assessed. For each of them, three different concentrations were tested to evaluate the dose/answer effect. Based on litterature reviews (Alaejos & Afonso, 2011; Dong, Xian, Li, Bai, Zeng, 2020), this work focused on five thermic HAAs known to be found in pan fried ground beef patties: PhIP, IQ, MeIQ, MeIQx, and 4,8-DiMeIQx.

415 Table 2 shows the contribution of individual HAAs in each formulation. Control sample 416 (ground beef patty without any antioxidant) exhibited clearly four thermic HAAs: MeIQ, 417 MeIQx, 4,8-DiMeIQx, and PhIP, but no IQ. Their amounts were similar to those reported by 418 other groups for pan-fried ground beef patties as summarized by Alaejos & Afonso (2011). In 419 the present study, PhIP level in the control meat sample was found to be 1.3 ng/g of cooked 420 meat and in litterature reviews, it ranged between 0 and 32 ng/g for pan-fried ground beef 421 patties. IQ and MeIQx levels ranged from 0 to 7–8 ng/g of cooked meat in other research 422 studies and were found to be not detected and 4.0 ng/g, respectively, in this work. As for 4,8-423 DiMeIQx and MeIQ, their levels ranged from 0 to 3 ng/g based on litterature review and here 424 reached 0.4 ng/g and 0.2 ng/g of cooked meat respectively.

425 *PhIP inhibition.* Carvacrol and resveratrol were the most effective antioxidants tested here to inhibit PhIP formation. Carvacrol at 0.25% inhibited PhIP formation by 51% while resveratrol 426 427 at 0.25% inhibited PhIP formation by 71%. Quercetin and epicatechin needed higher concentrations to display an inhibitory effect. At a concentration of 0.5% of epicatechin 61% 428 429 inhibition was observed but none at lower concentrations (0.1 and 0.25%). Similarly, the 430 present study showed that quercetin at 0.5% tended towards reducing by half PhIP formation 431 (54% of inhibition). This result was in accordance with several other research works (Cheng, 432 Chen, & Wang, 2007; Oguri, Suda, Totsuka, Sugimura, & Wakabayashi, 1998; Zhu, et al., 433 2016) on model systems or on beef patties and using higher concentrations of this flavonoid 434 that induced inhibition ranging from 40 to 75% of PhIP formation. Zhu et al. (2016) found 435 that the presence of a hydroxyl group at different positions, especially the B-ring and C-ring 436 of the flavone skeleton, like in quercetin, significantly affected flavonoid inhibition effects on 437 PhIP formation. Thus, they suggested that the simultaneous occurrence in quercetin of a hydroxyl group at 3'-position of B-ring and 3-position of C-ring would greatly enhance the 438 439 resultant inhibition effects on PhIP formation. Salazar et al. analyzed the role of various 440 phenolic compounds on the PhIP produced in model systems to highlight the structural 441 characteristics that favored the inhibition of this HAA formation (Salazar, Arámbula-Villa, 442 Hidalgo, & Zamora, 2014). Among these compounds, they studied resveratrol, epicatechin 443 and guercetin. At a concentration of 10 µmol of phenolic compound (equimolar with 444 phenylalanine, the main precursor of PhIP), they found that resveratrol was a better inhibitor 445 of PhIP formation than guercetin and that epicatechin was the weakest inhibitor among the 446 three. The three of them have two hydroxyl groups in the *meta* position of the A-ring but the 447 difference of activity could be explained by the two hydroxyl groups in the ortho position of 448 the B-ring for quercetin and epicatechin.

449 The present results in pan-fried ground beef patties confirmed those obtained in model 450 systems by Salazar et al. (2014), as a better inhibition of PhIP was observed for resveratrol 451 than for quercetin and epicatechin. The similar results observed for epicatechin (61% 452 inhibition at 0.5%) and quercetin (54% inhibition at 0.5%) in this study could be explained by 453 their similar 3D structure (Supplementary Data 5). In the same way, as shown in 454 Supplementary Data 6, quercetin and epicatechin displayed similar representation of their 455 electrostatic potentials. Their bicyclic scaffold displayed an important positive electrostatic 456 potential in opposition to the *meta* hydroxyl groups which were strongly electronegative. The 457 hydroxyl group of resveratrol and carvacrol displayed the same electronegative potential but 458 contrary to the other two antioxidants, their scaffolds also displayed electronegative potential. 459 The electronegative potential around the hydroxyl group explained its reactivity with the 460 Strecker aldehyde intermediates of HAA formation.

461 To explain this reactivity, a mechanism was proposed in Figure 3 based on the work of Cheng 462 et al. on the inhibition of PhIP formation by reaction of epigallocatechin gallate on 463 phenylacetaldehyde, which is the key intermediate in this HAA formation (Cheng, et al., 464 2009). It rested upon an electrophilic substitution of phenylacetaldehyde by the A-ring (C-6 or 465 C-8) of quercetin, (-)-epicatechin and resveratrol. The electron-withdrawing property of the 466 conjugated carbonyl at the C-ring of quercetin could explain its lack of reactivity with the 467 Strecker aldehyde. The highest reactivity of resveratrol could be explained by a lower steric 468 hindrance as well as the repartition of its electrostatic field as displayed by our modeling 469 (Supplementary Data 4). Carvacrol, a monophenolic compound substituted by alkyl groups 470 was by far less reactive for this kind of reaction as alkyl groups, donors by inductive effect, 471 were less activating than hydroxyl (donor by conjugation) and therefore less ortho/para 472 director. The results of inhibition tests coupled to molecular modeling and structure-activity relationship studies showed the importance of a polyphenolic scaffold with hydroxyls in the 473

474 *meta* position to activate the electrophilic substitution sites. From the experimental results, steric hindrance and electron-withdrawing grouping near the A-ring had a negative effect on 475 476 the inhibitory properties of the antioxidants. Studies of the electrostatic field representation allowed to think that the reactive hydroxyl groups should be surrounded by negative 477 478 isosurface especially around the carbon involved in the electrophilic substitution and the 479 repartition of the electrostatic fields should be somewhat linear to promote the reaction. This 480 fact accounted for the difference of inhibitory properties between carvacrol and resveratrol on 481 one hand and quercetin and epicatechin on the other.

482 *MeIQx inhibition.* MeIQx was found in larger quantities (p < 0.01) in ground beef samples as 483 found by Ni et al. (2008). Of the four antioxidants studied, resveratrol and carvacrol were the 484 most promising to inhibit MeIQx formation at the concentrations tested. At 0.25%, resveratrol inhibited MeIQx by 40%. Carvacrol at 0.25% permitted an inhibition of 33% of MeIQx 485 486 formation. A previous study interested in antibacterial and HAA inhibitory properties of carvacrol in grilled ground beef patties described inhibition level of 72% for MeIQx 487 488 (Friedman, et al., 2009) but the concentration used (1% w/w) was more than in the present 489 study (0.1, 0.25 and 0.5% w/w).

490 4,8-DiMeIQx inhibition. 4,8-DiMeIQx, as MeIQx, belongs to the quinoxaline type of HAAs 491 and is most probably formed *via* a dialkyl pyrazine radical intermediate. Therefore, one would 492 have expected similar inhibition results for MeIOx and 4.8-DiMeIOx at least in term of 493 antioxidants having inhibitory properties. In the present work none of the antioxidants studied 494 displayed inhibitory activity against 4.8-DiMeIOx. These results are in accordance with other 495 research studies, such as the work of Zeng et al. (2016) that showed no inhibition of 4,8-496 DiMeIQx by quercetin in a model system even at a concentration higher than in the present 497 study. It could be thus postulated that either the levels of antioxidants employed were too low 498 to imply an inhibitory activity against 4,8-DiMeIQx or as the amount of 4,8-DiMeIQx was

lower than that of MeIQx, it was difficult to follow such a small variation in quantity. MeIQ

*inhibition.* The best results in HAA inhibition by antioxidants were obtained for MeIQ. Resveratrol even at the lowest concentration (0.1%) inhibited totally MeIQ formation. Epicatechin at a concentration of 0.25% or 0.5% achieved the same result. Concerning quercetin, a higher concentration (0.5%) was required to totally inhibit the formation of this HAA. Carvacrol was not found to inhibit MeIQ formation at the concentrations used in this study but Friedman et al. described an inhibitory effect of 58 % at a higher concentration (1% w/w) (Friedman, et al., 2009).

507 Based on the present results, resveratrol was found to be the most potent antioxidant tested as 508 it inhibited totally the formation of MeIQ, and by 40 and 71% the formation of MeIQx and 509 PhIP, respectively.

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## 4.3. Assessment of the inhibitory properties of the natural products rich in the chosen

#### antioxidants on HAA formation in cooked meat

513 To each active principle studied was associated a natural product known to be rich in that 514 antioxidant in order to determine the inhibitory potential against HAA formation of the 515 antioxidant pure or in a mixture found in nature and used in household preparations. Thus in 516 parallel to resveratrol, epicatechin, guercetin and carvacrol studies for HAA inhibition, red 517 wine, green tea, caper and oregano formulations, respectively, were investigated. Different 518 concentrations and marinade times were chosen to be representative of consumer's practices 519 and high enough to display a potential inhibitory effect based on litterature data (Gibis & 520 Weiss, 2012; Friedman, Zhu, Feinstein, & Ravishankar, 2009) but low enough to be accepted in meat recipes. 521

522 Table 3 shows the contribution of individual HAAs in each formulation.

523 *PhIP inhibition.* Concerning natural products rich in antioxidants, oregano (rich in carvacrol) 524 at higher concentrations (0.25 & 0.5%) had a tendancy to decrease PhIP formation by more 525 than two fold (60 and 54% inhibition, respectively). This result agreed with those described by Damašius et al. (2011) which showed a similar range of inhibitory effect of oregano on 526 527 PhIP formation. The range of PhIP inhibition with oregano was also in accordance with the 528 results found for carvacrol in the previous part of this work (51% of PhIP inhibition at a 529 concentration of 0.25%). From the results of Quelhas et al. (2010), a better inhibition of PhIP 530 formation was expected for green-tea-based formulations but in their work PhIP control level 531 was higher. The difference in reactivity could be explained by the variable content in active 532 ingredient in food matrices. Indeed, as shown by Henning et al. (2003) the content of 533 epicatechin in green tea varied from 6.5 to 15.4 mg/100 mL among similar kinds of green tea 534 and from 0 mg/100 mL in decaffeinated green tea to 19.7 mg/100 mL in green tea 535 supplement. This variation could explain the differences observed from one study to another, 536 especially since in the previous part of this study, the active principle (epicatechin) at a 537 concentration of 0.5% displayed an inhibition of PhIP formation of 61%. From the results of 538 Busquets et al. (Busquets, et al. 2006; Melo, et al., 2008), better inhibition results for red wine 539 were expected but the lower inhibitory effect observed could be potentially explained by 540 confounding factors including the meat matrix used, the red wine used (different antioxidant 541 content depending on the wine), the duration of marinade. Indeed, as was the case for green 542 tea, the level of resveratrol in similar wines (pinot noir) could vary from 2.3 to 10.5 mg/L 543 according to its geographical origin (Stervbo, Vang, & Bonnesen, 2007). This explained the 544 difference of reactivity found between resveratrol (inhibition of 71% at a concentration of 545 0.25%) and red wine (no inhibition).

546 *MeIQx inhibition.* Poor MeIQx inhibition was observed with the natural ingredient 547 formulations. Oregano could be the most inhibiting among the products tested as at a

548 concentration of 0.25%, it inhibited by 36% MeIQx formation. This result was similar to the 549 inhibition found with carvacrol at 0.25% (33% inhibition). Contrary to resveratrol, red wine 550 did nott exhibit inhibitory properties against MeIQx formation, which again could be due to 551 variation of resveratrol content among pinot noir red wines or the existence of antagonist 552 effects with other red wine constituents. Green tea seemed to display no inhibitory effect, 553 which was in accordance with the results found by Quelhas et al. (2010), who observed no 554 reduction of MeIQx for marinated meat samples during 1 to 6 h in green tea. The absence of 555 inhibition of MeIQx observed for caper and green tea was in agreement with the results found 556 for quercetin and epicatechin, respectively, in the previous part of this study.

557 4,8-DiMeIQx inhibition. The different formulations studied did not inhibit significantly 4,8-558 DiMeIQx formation. These results were in accordance with previous studies revealing that 559 wine (Melo, et al., 2008) and green tea (Quelhas, et al., 2010) marinades were poor inhibitors 560 of 4,8-DiMeIQx formation in pan-fried beef. They were also in agreement with the lack of 561 inhibition found for the different active principles in the previous part of this work.

562 MeIQ inhibition. Green tea and oregano were the best natural products tested in this work to 563 inhibit MeIQ formation. The addition of 0.25% or more of oregano inhibited totally MeIQ 564 formation. As for green tea, 2 h or 4 h marination totally inhibited MeIQ formation, while 6 h 565 marination induced an increase of its level. The fact that there was no inhibition with red wine 566 as opposed to resveratrol (which totally inhibited MeIO formation at a concentration of 0.1% 567 and more) suggested that either the concentration of resveratrol in wine was too low to 568 achieve a good inhibition or an antagonist effect existed with other wine constituents. This antagonistic effect could be due to the large amount of phenolic acids in red wine and in 569 particular gallic acid and caftaric acid (Van Leeuw, Kevers, Pincemail, Defraigne, & 570 571 Dommes, 2014) that were both polyphenolic compounds with hydroxyl groups in ortho position and electron withdrawing group, strongly deactivating, in *meta* position from the 572

573 hydroxyl group. These phenolic acids, given their chemical structure, could also act as 574 inhibitors of MeIQ but would be less active than resveratrol due to the presence of ortho 575 deactivating group. As they were in large amounts in red wine, they could limit the inhibitory effect of resveratrol by competitive agonist reaction. In the same way, the better MeIO 576 577 inhibition of oregano than carvacrol (total inhibition vs 33% of inhibition respectively at 578 concentration of 0.25%) could suggest that the concentrations of pure carvacrol used for the 579 experiments were too low compare to that of oregano. Indeed as for the other natural 580 ingredients tested, the content of active principle in oregano was highly variable. Dambolena 581 et al. (2010) found that the relative percentage of carvacrol in oregano species varied from 582 trace amounts to 3.57%. Another probable explanation of the difference of reactivity between 583 oregano and carvacrol against MeIQ formation could be that other oregano constituents 584 displayed an additive effect with carvacrol increasing the inhibition potential of MeIQ 585 formation. This agonist effect could be explained by the high amount of thymol, an isomer of carvacrol, in common oregano species (such as *Oreganum majorana* and *Oreganum vulgare*) 586 587 (Dambolena, et al., 2010). The low inhibition displayed by caper formulation could be 588 explained by the varying amount of quercetin (from 0.03 to 1.45 mg/g depending on caper 589 origin) and by the fact that an important part of the quercetin load in caper sample was in the 590 form of flavonol glycosides (Inocencio, Rivera, Alcaraz, & Tomás-Barberán, 2000). The 591 presence of sugar moieties added to the steric hindrance of the compound and could therefore 592 explain its lack of reactivity against HAA inhibition under this form.

593

#### 594 **5.** Conclusion

595 The implementation of a method based on molecular modeling allowed the reasoned choice of 596 antioxidants inhibiting HAA formation (using analog design methods) and the explanation of 597 their reactivity (using structure–activity relationships). The first part of this study, based on

medicinal chemistry approaches, highlighted the mandatory presence of a polyphenolic 598 599 moiety for HAA inhibition with two hydroxyl groups in *meta* position to enhance the reaction 600 of electrophilic aromatic substitution of the antioxidant on the Strecker aldehydes, 601 intermediates of the Maillard reaction forming thermic HAAs. The selected antioxidants, as to 602 know four phenolic active principles and four corresponding natural ingredients rich in these 603 active principles, were tested in ground beef patties cooked under common household 604 conditions. The inhibition study (Sections 4.2 & 4.3) highlighted that the reactivity of 605 antioxidants, as active principles or in natural ingredients, depended on the HAA targeted. It 606 was therefore difficult to conclude in a global manner about antioxidant capacity to inhibit 607 HAA formation in the meat matrix. Resveratrol was shown to be the most efficient in 608 inhibiting the HAAs studied but similar results were not found for red wine rich in this active 609 principle. On the contrary, oregano displayed the best inhibitory properties, suggesting 610 probable additive effect of other antioxidants present in this herb. This study pointed out the 611 importance to consider ingredients in their entirety. Indeed, molecular modeling permitted the 612 selection of inhibitors of HAA formation among a wide range of active principles but when 613 looking at natural ingredients known to be rich in these antioxidants, the results were not 614 linear. This highlighted the necessity to consider: 1) ingredient variety, as antioxidant 615 concentration varied from one species to another, as well as 2) the antioxidant environment in 616 the natural ingredient, in order to take into account potential agonist or antagonist effects on 617 the desired inhibitory activity. In the future it would be interesting to test different varieties of 618 a natural ingredient to take into account varietal diversity. The idea would be to see how to 619 make a reasoned choice of the ingredient and not only of the active principle. The use of 620 medicinal chemistry approaches developed in this study could be expanded to natural 621 ingredient composition. It would permit consideration of the impact of other components of 622 the natural ingredients, either agonist or antagonist, on HAA inhibition.

623 While this study gave a method adapted from medicinal chemistry to select antioxidants 624 efficient in inhibiting HAA formation in proteinaceous food and explain their inhibitory 625 activity, the natural ingredients rich in these antioxidants should also be tested for their 626 organoleptic quality in meat preparations to assess consumer acceptance of these formulated 627 products. Indeed, finding inhibitory solutions is essential but useless if the corresponding formulation is not accepted by consumers. Therefore the effects of the identified natural 628 629 antioxidant addition on the meat aroma profile were assessed in a second part of this research 630 work and compounds responsible for the differences between formulations were identified by an original approach using hedonic rating, non-verbal analysis, quantitative descriptive 631 analysis, and gas chromatography-olfactometry. It would permit a comprehensive validation 632 considering both safety and sensory aspects. 633

634

#### 635 6. Acknowledgements

This research was conducted as part of the MARMEAT project supported by the
TRANSFORM division of the French National Research Institute for Agriculture, Food and
the Environment (INRAE).

639

#### 640 **7. Conflicts of interest**

641 The authors declare that there are no conflicts of interest.

642

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- 770 CRediT author statement

## 772 Mitigation of heterocyclic aromatic amines in cooked meat

# Part I: Informed selection of antioxidants based on molecular modeling

775

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- 783 Sylvie CHEVOLLEAU- Investigation, Resources, Writing Review & Editing
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## 787788 Declaration of interests

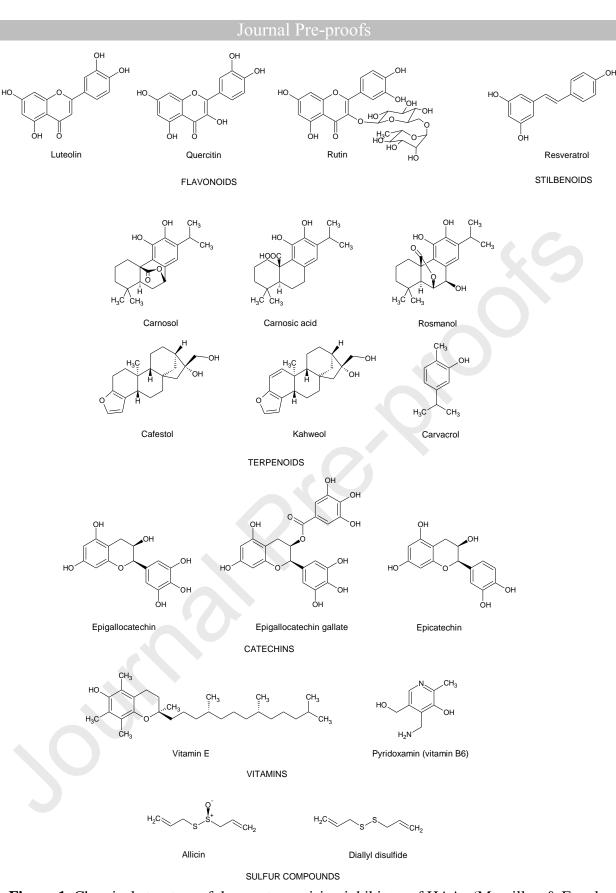
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- 790  $\square$  The authors declare that they have no known competing financial interests or personal
- relationships that could have appeared to influence the work reported in this paper.
- 792

793 □The authors declare the following financial interests/personal relationships which may be794 considered as potential competing interests:

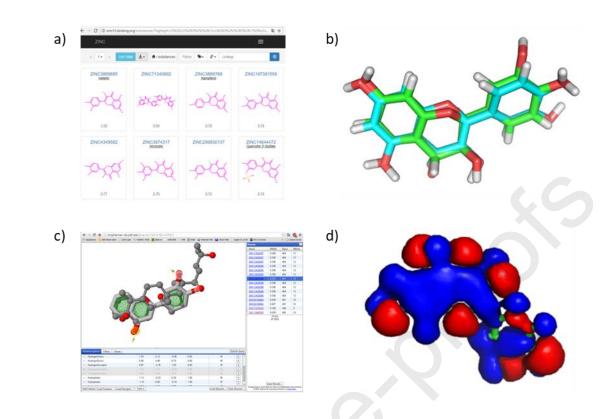
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| 796<br>797<br>798<br>799 |   |
|--------------------------|---|
| 800                      |   |
| 801                      | Highlights:   |
| 802                      | • Analog design strategy to make a reasoned choice of antioxidants for HAA inhibition |
| 803                      | • Determination of the structural properties required for inhibiting HAA formation    |
| 804                      | • Tests of HAA inhibition by 4 active principles and 4 natural ingredients            |
| 805                      | Best formulations to reduce HAA formation   |
| 806<br>807               |   |



809 Figure 1. Chemical structure of the most promising inhibitors of HAAs (Meurillon & Engel

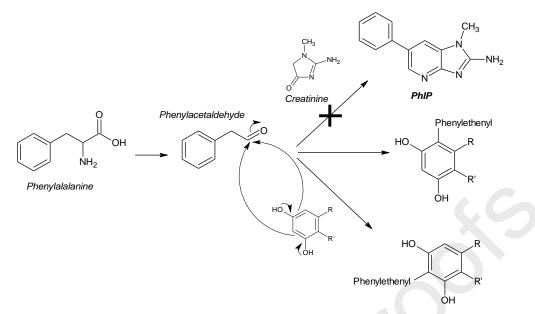
2016)





816 Figure 2. The different steps of molecular modelling and analog search based on quercetin scaffold. a) 817 Similarity search based on Tanimoto index (based on fingerprints defined only by the chemical formula) 818 allowing collection of all compounds having 70% similarity with quercetin, b) Scaffold hopping based on 3D 819 shape similarity and physicochemical properties permitting enrichment of compound libraries with similar core 820 structures, c) Pharmacophore model defined by the main molecular features of quercetin (aromatic cycle, 821 hydrogen bond donors and acceptors) allowing the screening of chemical databases for compounds fulfilling 822 these criteria, d) *Electrostatic potential surface determination* to help in defining organic reactivity by 823 examining electron distribution within the chemical structure. Electrostatic potential surface was computed using 824 Adaptive Poisson-Boltzmann Solver with a probe radius of 1.4 Å. Positive (blue) and negative (red) isosurfaces 825 at  $\pm$  5 kT/e and visualized by VMD.

826 827



**Figure 3.** Postulated pathways for inhibitory activity of polyphenolic compounds on PhIP

- formation based on the work of Cheng et al. (2009).

Table 1. Number of structural analogs displaying a Tanimoto coefficient > 0.7 and number of
functional analogs given by scaffold hopping based on 3D and physicochemical similarity for

| 835 | each antioxidant | described i | n the literati | ure as a HA | A inhibitor. |
|-----|------------------|-------------|----------------|-------------|--------------|
|-----|------------------|-------------|----------------|-------------|--------------|

| Compounds                   | Structural analogs <sup>a</sup>  | Functional analogs <sup>b</sup> |
|-----------------------------|--|---------------------------------|
| Resveratrol                 | <b>17 different compounds</b> (47 compounds including different diastereoisomers)  | 17 compounds                    |
| Luteolin                    | 36 different compounds   | 11 compounds                    |
| Quercetin                   | 26 different compounds   | 4 compounds                     |
| Rutin                       | 1 compound with different diastereoisomers   | 0 (& 0 with a Zscore >2)        |
| Carnosol                    | 8 different compounds (51 compounds including different diastereoisomers)          | 0 (& 5 with a Zscore >2)        |
| Carnosic acid               | <b>9 different compounds</b> (37 compounds including different diastereoisomers)   | 0 (& 7 with a Zscore >2)        |
| Rosmanol                    | <b>11 different compounds</b> (84 compounds including different diastereoisomers)  | 0 (& 1 with a Zscore >2)        |
| Cafestol                    | 6 different compounds (100 compounds including different diastereoisomers)         | 0 (& 3 with a Zscore >2)        |
| Kahweol                     | <b>3 different compounds</b> (57 compounds including different diastereoisomers)   | 0 (& 5 with a Zscore >2)        |
| Carvacrol                   | 5 different compounds  | 41 compounds                    |
| Epigallocatechin            | <b>11 different compounds</b> (46 compounds including different diastereoisomers)  | 1 compound                      |
| Epigallocatechin<br>gallate | <b>28 different compounds</b> (100 compounds including different diastereoisomers) | 0 (& 4 with a Zscore >2)        |

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|--------------------|--|---------------------------|--|--|--|
| Epicatechin        | <b>20 different compounds</b> (82 compounds including different diastereoisomers)  | 7 compounds               |  |  |  |
| Vitamin E          | <b>14 different compounds</b> (100 compounds including different diastereoisomers) | 0 (& 0 with a Zscore >2)  |  |  |  |
| Pyridoxamine       | 17 different compounds   | 0 (& 26 with a Zscore >2) |  |  |  |
| Allicin            | 1 compound with different diastereoisomers   | 0 (& 0 with a Zscore >2)  |  |  |  |
| Diallyl sulfide    | 7 different compounds  | 0 (& 1 with a Zscore >2)  |  |  |  |

<sup>a</sup> Structural analogs based on chemical structure displaying a Tanimoto coefficient > 0.7; <sup>b</sup> Functional analogs based on scaffold hopping results and displaying a Zscore > 3.01

#### 840 Table 2. Amount of the different HAAs found in cooked ground beef patties formulated with

841 or without antioxidants.

| Samples           | IQ   | MelQx             | MelQ          | 4,8-DiMelQx   | PhIP              |
|-------------------|------|-------------------|---------------|---------------|-------------------|
| Control           | N.D. | 7.2 ± 2.2         | $0.4 \pm 0.1$ | 0.7 ± 0.3     | 2.3 ± 0.9         |
| Epicatechin 0.1%  | N.D. | $10.8 \pm 1.7$    | 0.3 ± 0.1     | 1.1 ± 0.2     | $3.1 \pm 0.9$     |
| Epicatechin 0.25% | N.D. | $10.6 \pm 4.1$    | N.D.***       | $1.1 \pm 0.4$ | 2.3 ± 1.5         |
| Epicatechin 0.5%  | N.D. | 8.9 ± 3.3         | N.D.***       | 0.8 ± 0.3     | $1.0 \pm 0.4$     |
| Resveratrol 0.1%  | N.D. | $4.9 \pm 1.1^{*}$ | N.D.***       | 0.5 ± 0.2     | 0.8 ± 0.2**       |
| Resveratrol 0.25% | N.D. | 4.5 ± 0.6*        | N.D.***       | $0.5 \pm 0.1$ | 0.7 ± 0.1**       |
| Resveratrol 0.5%  | N.D. | 8.2 ± 1.6         | N.D.***       | $0.8 \pm 0.2$ | $1.9 \pm 0.9$     |
| Quercetin 0.1%    | N.D. | 8.9 ± 2.6         | 0.3 ± 0.0     | $0.8 \pm 0.3$ | $2.4 \pm 0.9$     |
| Quercetin 0.25%   | N.D. | 11.0 ± 6.3        | 0.3 ± 0.2     | $1.0 \pm 0.5$ | $3.1 \pm 2.0$     |
| Quercetin 0.5%    | N.D. | 5.6 ± 0.8         | N.D.          | $0.5 \pm 0.0$ | $1.0 \pm 0.2$     |
| Carvacrol 0.1%    | N.D. | 6.1 ± 0.5         | $0.3 \pm 0.1$ | $0.6 \pm 0.0$ | 1.5 ± 0.2         |
| Carvacrol 0.25%   | N.D. | 5.1 ± 0.4**       | $0.3 \pm 0.1$ | $0.5 \pm 0.1$ | $1.2 \pm 0.1^{*}$ |
| Carvacrol 0.5%    | N.D. | 6.1 ± 1.2         | $0.3 \pm 0.1$ | $0.6 \pm 0.1$ | $1.4 \pm 0.9$     |

Values expressed in ng/g of freeze-dried product; each experiment was repeated 3 times; The detection and quantification limits (expressed in ng/g of freeze-dried product) were, respectively, 0.02 and 0.2 for PhIP, IQ and 4.8-DiMeIQx and 0.05 and 0.2 for MeIQx and MeIQ; N.D.:

845 not detected; \*: significant difference to control value, p < 0.05; \*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.001.

- **Table 3.** Amount of the different HAAs found in cooked ground beef patties formulated with
- 851 or without natural ingredients.

| Samples       | IQ   | MelQx     | MelQ          | 4,8-DiMelQx   | PhIP          |
|---------------|------|-----------|---------------|---------------|---------------|
| Control       | N.D. | 7.2 ± 2.2 | $0.4 \pm 0.1$ | 0.7 ± 0.3     | 2.3 ± 0.9     |
| Green Tea 2 h | N.D. | 6.2 ± 1.7 | N.D.***       | $0.6 \pm 0.0$ | $2.1 \pm 0.5$ |
| Green Tea 4 h | N.D. | 4.9 ± 0.7 | N.D.***       | $0.6 \pm 0.1$ | 1.7 ± 0.9     |
| Green Tea 6 h | N.D. | 5.9 ± 2.2 | $0.3 \pm 0.0$ | 0.6 ± 0.2     | 2.1 ± 1.5     |
| Red Wine 2 h  | N.D. | 7.1 ± 2.1 | $0.3 \pm 0.0$ | 0.8 ± 0.2     | $1.6 \pm 0.7$ |
| Red Wine 4 h  | N.D. | 8.4 ± 2.8 | $0.4 \pm 0.0$ | $1.0 \pm 0.3$ | $1.6 \pm 0.7$ |

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|--------------------|------|-----------|---------------|---------------|---------------|--|
| Red Wine 6 h       | N.D. | 8.2 ± 1.4 | 0.3 ± 0.1     | $1.0 \pm 0.1$ | 1.7 ± 0.6     |  |
| Caper 0.1%         | N.D. | 5.6 ± 1.2 | 0.3 ± 0.0     | $0.6 \pm 0.1$ | $1.2 \pm 0.5$ |  |
| Caper 0.25%        | N.D. | 7.0 ± 3.7 | $0.2 \pm 0.1$ | 0.7 ± 0.4     | 2.2 ± 1.5     |  |
| Caper 0.5%         | N.D. | 7.1 ± 2.4 | $0.2 \pm 0.1$ | 0.7 ± 0.2     | $1.8 \pm 0.6$ |  |
| Oregano 0.1%       | N.D. | 9.2 ± 3.2 | $0.5 \pm 0.1$ | $0.8 \pm 0.2$ | $3.0 \pm 1.4$ |  |
| Oregano 0.25%      | N.D. | 4.6 ± 0.3 | N.D.***       | $0.5 \pm 0.1$ | $1.0 \pm 0.2$ |  |
| Oregano 0.5%       | N.D. | 5.7 ± 0.9 | N.D.***       | $0.6 \pm 0.1$ | $1.1 \pm 0.3$ |  |

853 Values expressed in ng/g of freeze-dried product; each experiment was repeated 3 times; Red wine is rich in resveratrol, green tea in

854 epicatechin, caper in quercetin and oregano in carvacrol; The detection and quantification limits (expressed in ng/g of freeze-dried product)

were, respectively, 0.02 and 0.2 for PhIP, IQ and 4,8-DiMeIQx and 0.05 and 0.2 for MeIQx and MeIQ; N.D.: not detected; \*: significant difference to control value, p < 0.05; \*\*: significant difference to control value, p < 0.05; \*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.05; \*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant difference to control value, p < 0.01; \*\*\*: significant

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