



Nitrate in 2020: Thirty Years from Transport to Signaling Networks

Elena A. Vidal, José M Alvarez, Viviana Araus, Eleodoro Riveras, Matthew Brooks, Gabriel Krouk, Sandrine Ruffel, Laurence Lejay, Nigel Crawford, Gloria Coruzzi, et al.

► To cite this version:

Elena A. Vidal, José M Alvarez, Viviana Araus, Eleodoro Riveras, Matthew Brooks, et al.. Nitrate in 2020: Thirty Years from Transport to Signaling Networks. *The Plant cell*, 2020, 32 (7), pp.2094-2119. 10.1105/tpc.19.00748 . hal-02894020

HAL Id: hal-02894020

<https://hal.inrae.fr/hal-02894020>

Submitted on 8 Dec 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

REVIEW

Nitrate in 2020: Thirty years from transport to signaling networks

Running title: Nitrate transport, signaling and responses

Elena A. Vidal^{1,2,3}, José M. Álvarez^{1,2,4}, Viviana Araus⁴, Eleodoro Riveras^{1,5,6}, Matthew D. Brooks⁴, Gabriel Krouk⁷, Sandrine Ruffel⁷, Laurence Lejay⁷, Nigel M. Crawford⁸, Gloria M. Coruzzi⁴ and Rodrigo A. Gutiérrez^{1,5,6}

¹ Millennium Institute for Integrative Biology (iBio), Santiago, Chile

² Centro de Genómica y Bioinformática, Facultad de Ciencias, Universidad Mayor, Santiago, Chile.

³ Escuela de Biotecnología, Facultad de Ciencias, Universidad Mayor, Santiago, Chile.

⁴ Center for Genomics and Systems Biology, Department of Biology, New York University, New York, NY 10003

⁵ Departamento de Genética Molecular y Microbiología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile

⁶ FONDAP Center for Genome Regulation, Santiago, Chile

⁷ BPMP, CNRS, INRA, Montpellier SupAgro, Univ Montpellier, Montpellier, France

⁸ Section of Cell and Developmental Biology, Division of Biological Sciences, University of California at San Diego, La Jolla, California

Corresponding author: Rodrigo A. Gutiérrez

Avenida Libertador Bernardo O'Higgins 340, Santiago, Chile

(+56) 2 686 2663

rgutierrez@bio.puc.cl

ABSTRACT

Nitrogen (N) is an essential macronutrient for plants, and a major limiting factor for plant growth and crop production. Nitrate is the main source of N available to plants in agricultural soils and in many natural environments. Sustaining agricultural productivity is of paramount importance in the current scenario of increasing world population, diversification of crop uses, and climate change. Plant productivity for major crops around the world is however still supported by excess application of N-rich fertilizers with detrimental economic and environmental impacts. Thus, understanding how plants regulate nitrate uptake and metabolism is key for developing new crops with enhanced N use efficiency (NUE) and to cope with future world food demands. The study of plant responses to nitrate has gained considerable interest over the last thirty years. This review provides an overview of key findings in nitrate research, spanning biochemistry, molecular genetics, genomics and systems biology. We discuss how we have reached our current view of nitrate transport, local and systemic nitrate sensing/signaling, and the regulatory networks underlying nitrate-controlled outputs in plants. We hope this summary will serve not only as a timeline and information repository, but also as a baseline to define outstanding questions for future research.

FOOTNOTES:

This review mentions over 150 genes by name. We will not spell out the acronyms in the main text for fear of interrupting the flow of the narrative. All gene acronyms, alternate names in the literature and their full names are listed in Supplemental Data Set 1.

INTRODUCTION

Nitrogen (N) is an essential macronutrient for plants and its availability is a key determinant for plant growth and productivity. N nutrients can be provided as inorganic (nitrate NO_3^- , ammonium NH_4^+) or organic forms (amino acids, urea). However, nitrate is the main source of N both in agricultural and in many natural systems (Crawford and Forde, 2002). N fertilizer consumption is currently approximately 118 million tons per year worldwide, representing 59% of total fertilizer nutrient used (FAO, 2017). The massive production and application of inorganic N fertilizers have contributed significantly to support and sustain the enormous rise in crop yield over the past century. However, fertilizer overuse has also led to the destruction of aquatic ecosystems and the formation of oceanic “dead zones” from eutrophication in coastal regions worldwide. N soil absorption is low, meaning that plants can only use about 50 to 75 % of the N input from fertilizers in a given season (Gutierrez, 2012). Nitrate that is not retained in the soil leaches into groundwater and is an important contributor to eutrophication. Furthermore, agriculture management practices are an important contributor to nitrous oxide emissions, a key greenhouse gas contributing to global warming. A key goal in driving N research is therefore to maintain high crop yields while reducing excess N-load to the environment by improving N use efficiency (NUE). A better understanding of the biology of N nutrition is key for achieving this goal.

The identification of genes encoding N uptake carriers, assimilation enzymes, and their regulators relied heavily on extensive biochemical and genetic studies in plants, algae and fungi. Figure 1 represents a timeline of some of the key publications in nitrate signaling over the last 70 years, and Supplemental Data Set 2 provides an extended (but non-exhaustive) list of the key publications involved in nitrate uptake, transport, and signaling, discussed in this section.

In 1953, soybean nitrate reductase (NR) was purified by Evans and Nason and its nitrate reductive activity confirmed *in vitro* (Evans and Nason, 1953). The 1980s saw the cloning of genes related to N-metabolism, including NR. Nitrate was also shown to control NR mRNA and protein levels in plants (Crawford et al., 1986; Cheng et al., 1986; Calza et al., 1987; Crawford et al., 1988). During the 1990s, several inorganic N transporters were identified. Tsay et al. isolated and characterized the first plant nitrate transporter, CHLORINA1 (CHL1), known today as NITRATE TRANSPORTER1.1 (NRT1.1) or NITRATE TRANSPORTER 1/PEPTIDE TRANSPORTER

FAMILY6.3 (NPF6.3), and showed that the mRNA levels of the encoding gene were induced by nitrate (Tsay et al., 1993). Although first described as a low-affinity transporter, NRT1.1/NPF6.3 (hereafter referred to as NRT1.1) was later shown to be a dual-affinity nitrate transporter, the switch from low to high affinity being mediated by phosphorylation of Thr 101 (Liu and Tsay, 2003). *NRT2.1* was isolated using degenerate oligonucleotides designed against conserved regions of nitrate transporters from fungi and algae (Zhuo et al., 1999), and its mRNA was also shown to be induced by nitrate treatments (Wang et al., 1998; Zhuo et al., 1999).

In the late 1990s, the first transcription factor (TF) involved in nitrate responses was identified in *Arabidopsis* (*Arabidopsis thaliana*): *ANR1*, a root-expressed and nitrate-induced gene encoding a MIKC-type MADS box TF. *ANR1* was shown to have a positive role in lateral root (LR) proliferation in nitrate-rich patches (Zhang and Forde, 1998). These results provided the first molecular basis for the effect of nitrate on controlling plant growth. During the first decade of the 2000s, genome-wide transcriptome studies began to unravel the full extent of nitrate impact on gene expression. Microarray and RNA-sequencing analysis showed that nitrate not only controlled the expression of genes involved in nitrate transport and assimilation, but also hundreds to thousands of genes involved in other processes such as carbon (C), sulfate, amino acid and nucleotide metabolism, hormone biosynthesis and response, as well as a myriad of TFs (Wang et al., 2000; Wang et al., 2001; Wang et al., 2003). At the same time, the increasing amount of data available on gene expression and molecular interactions allowed the rise of systems biology approaches to model nitrate-controlled regulatory networks (Gutiérrez et al., 2007; Gifford et al., 2008; Gutiérrez et al., 2008).

The identification of NRT1.1 as a nitrate sensor in 2009 was a major milestone in nitrate signaling (Ho et al., 2009). That same year, researchers identified NLP7, one of the best studied TFs in the N-response. NLP7 controls responses to nitrate by a nuclear retention mechanism that is triggered by nitrate availability (Castaings et al., 2009; Wang et al., 2009). At the beginning of the 2010s, high throughput small RNA (sRNA) sequencing was used to identify sRNAs involved in nitrate responses. A nitrate-induced regulatory module between microRNA 393 (miR393) and *AFB3* was identified: it integrated external nitrate availability, internal organic N signals and root auxin sensitivity to control root architecture (Vidal et al., 2010). Likewise, studies of nitrate responses in specific root cell types uncovered a role for miR167 and *ARF8* in LR emergence (Gifford et al., 2008). In addition, a function for NRT1.1 in transporting auxin provided a

molecular mechanism to explain LR elongation in response to external nitrate (Krouk et al., 2010b). Other TFs important to the nitrate response such as TGA1 and TGA4, NLP6, HRS1 and HHO1, NAC4, SPL9, TCP20, LBD37/38/39 and NRG2 were identified shortly after (Rubin et al., 2009; Krouk et al., 2010a; Vidal et al., 2013; Alvarez et al., 2014; Guan et al., 2014; Medici et al., 2015; Xu et al., 2016b). More recently, the development of ultrasensitive biosensors and considerable advances in live microscopy identified calcium as an important second messenger in nitrate signaling (Riveras et al., 2015; Liu et al., 2017). Coupled with the recent discovery of a calcium-dependent signaling pathway involving calcium-dependent kinases (CPKs) that phosphorylate NLP7 to control its subcellular localization, these results represent a major advance linking nitrate sensing to changes in the transcriptome, root and shoot growth (Liu et al., 2017).

In addition to primary nitrate responses, systemic N signaling has also received much attention. Development of the split-root system was instrumental, which allowed the identification of nitrate-cytokinin interactions in systemic N-signaling (Ruffel et al., 2011; Ruffel et al., 2016; Ruffel and Gojon, 2017). The relevant molecular players have also been identified, including the translocation of the peptide Cle and of the transcription factor HY5 from shoots to roots (Araya et al., 2014b ; Chen et al., 2016).

Today, advances in DNA sequencing and high-throughput genomic technologies have permitted substantial progress in deciphering gene regulatory networks controlling responses to nitrate. The analysis of time-series data (Krouk et al., 2010a; Varala et al., 2018; Brooks et al., 2019), yeast-1-hybrid technology (Gaudinier et al., 2018), and genome-wide chromatin accessibility and RNA polymerase II Chromatin Immunoprecipitation followed by deep-sequencing (ChIP-Seq) assays (Alvarez et al., 2019) all helped identify TF-DNA interactions in networks involved in nitrate responses. Perhaps not surprisingly, nitrate and hormone signaling networks are interconnected (Ristova et al., 2016).

Finally, the increasing availability of high-quality genomes and genetic resources for plants makes it possible to extend our vast knowledge to crops (e.g. the *OsNRT1.1b* gene in rice, (Hu et al., 2015)) and other plants of economic importance, with the aim to improve agronomic traits such as NUE, seed quality, and flowering time, among other potential uses.

MOLECULAR MECHANISMS OF NITRATE TRANSPORT

The molecular basis of nitrate uptake by the root has been studied intensively over the past 30 years. Plants employ two different uptake systems depending on how much external nitrate is available: the high (HATS) and low (LATS) affinity transport systems, operating at low (< 1mM) or high (> 1mM) nitrate concentrations, respectively (Crawford and Glass, 1998).

Nitrate transporters

The Arabidopsis genome has 53 *NRT1/NPF* genes and 7 *NRT2* genes (Supplemental Tables 1 and 2); the NPF family is responsible for LATS (Leran et al., 2014) while the NRT2/NNP (Nitrate-Nitrite Porter) functions in HATS (Orsel et al., 2002; Krapp et al., 2014). Although most NRT2 and NPF transporters have yet to be functionally characterized, four NRT2 transporters (NRT2.1, NRT2.2, NRT2.4 and NRT2.5) and two NPF transporters (NRT1.1 and NRT1.2) are components of root nitrate uptake (Tsay et al., 1993; Huang et al., 1999; Filleur et al., 2001; Kiba et al., 2012; Lezhneva et al., 2014). NRT1.1 is the only demonstrated dual-affinity nitrate transporter, and a putative nitrate transceptor (a portmanteau for a transporter-receptor) (Wang et al., 1998; Liu et al., 1999; Ho et al., 2009). Within the NRT2/NPF family, NRT2.1 predominantly localizes to the plasma membrane of root epidermal and cortical cells, where the bulk of nitrate uptake occurs (Chopin et al., 2007; Wirth et al., 2007).

Among NPF transporters, 13 have been found to be involved in root-to-shoot transport, seed development or N storage (Wang et al., 2018). NPF2.7, also called NAXT1, is the only known root nitrate efflux transporter identified to date (Segonzac et al., 2007). The gene is specifically expressed in seeds, where it controls nitrate content and dormancy (Chopin et al., 2007). In addition to NPF and NRT2 transporters, members of the CLC and SLAC/SLAH protein families facilitate nitrate transport inside the plant. SLAC/SLAH is a small protein family encoded by 5 genes that displays a common predicted structure of 10 transmembrane domains. Among them, SLAC1 and SLAH3 are characterized by NO₃⁻/Cl⁻ permeability and appear to be involved in the regulation of stomatal closure (Negi et al., 2008; Geiger et al., 2011). In contrast, *SLAH2* is expressed in the stele of the root and encodes an anion channel that transports nitrate exclusively (Maierhofer et al., 2014) and may facilitate nitrate transport between root and shoot. Seven *CLC* genes have been identified in Arabidopsis. CLCa and CLCb proteins are NO₃⁻/H⁺ antiporters involved in vacuolar nitrate storage (De Angeli et al., 2006; von der Fecht-Bartenbach et al., 2010), and are the only

two (out of over 20 nitrate transporters) that do not localize to the plasma membrane. Clearly, there is much to learn about the function of vacuolar nitrate transporters.

Transcriptional Control of Nitrate Transport

There are two major mechanisms that coordinate root nitrate uptake as a function of external nitrate supply and internal N demand (Crawford and Glass, 1998; Gojon et al., 2009). First, the expression of nitrate transporters *NRT2.1* and *NRT1.1* is rapidly induced shortly after nitrate treatment (Tsay et al., 1993; Filleur and Daniel-Vedele, 1999; Lejay et al., 1999; Zhuo et al., 1999; Okamoto et al., 2003). Second, a negative feedback exerted by a high N status (at the whole plant level or from external high nitrate), represses *NRT2.1*, *NRT2.2*, *NRT2.4* and *NRT2.5* expression once N needs are met (Lejay et al., 1999; Zhuo et al., 1999; Kiba et al., 2012; Lezhneva et al., 2014). This feedback repression is lifted when plants (re)experience N limitation, resulting in a strong increase in HATS capacity that improves N uptake. Feedback repression of *NRT2.1* is suppressed or strongly attenuated in *nrt1.1* mutants (Muños et al., 2004), although ammonium- or nitrate-fed *nrt1.1* mutants are not N-deficient, suggesting that *NRT2.1* expression is not only repressed by reduced N metabolites, but also by nitrate itself. In fact, further studies revealed that *NRT2.1* was highly induced under low nitrate/high ammonium availability even in wild-type plants. This shows that, at least for *NRT2.1*, feedback repression exerted by high N status is more complex and involves both 1) feedback repression by reduced N metabolites or mobile peptides and 2) *NRT1.1*-mediated repression by high external nitrate (Krouk et al., 2006; Ohkubo et al., 2017).

In addition, nitrate uptake is a highly integrated process, determined not only by nitrate availability and the N demand of the plant, but also by C produced by photosynthesis in order to balance levels of root nitrate uptake against the availability of C metabolites (Delhon et al., 1995; Lejay et al., 1999; Lejay et al., 2003). The coordination of photosynthesis (and thus C metabolism) with root N uptake involves the bZIP transcription factor HY5, which can translocate from shoots to roots upon light illumination to control *NRT2.1* expression, nitrate uptake and root growth (Chen et al., 2016). Light and sugars can also control root nitrate uptake by modulating *NRT2.1*, *NRT2.4* and *NRT1.1* expression, as can an Oxidative Pentose Phosphate Pathway (OPPP)-dependent signaling mechanism (Lejay et al., 2008; de Jong et al., 2014). It is interesting to note that the control of gene expression by an OPPP-derived signal extends to nitrate assimilatory genes in

response to sucrose in the roots (Bussell et al., 2013; de Jong et al., 2014). Understanding the signaling mechanism linked to OPPP could be key to determining how C and N signaling pathways are integrated to modulate root nitrate uptake.

Post-translational control of nitrate transport

Most studies of nitrate transporters have focused on their transcriptional regulation, but increasing evidence highlights the importance of post-translational control. Phosphorylation plays an important role in the regulation of NRT1.1 and AMT1.1. For NRT1.1, low nitrate concentrations trigger phosphorylation of Thr 101 (Liu and Tsay, 2003) and lead to a switch to its high affinity function. Conversely, NRT1.1 remains a low affinity transporter when Thr 101 is not phosphorylated at high nitrate concentrations.

Post-translational modifications also control the activity of NRT2 transporters. NRT2.1 shows no detectable nitrate transport activity in the absence of NRT3.1 (Okamoto et al., 2006a; Orsel et al., 2006). In agreement, frog (*Xenopus laevis*) oocytes co-injected with NRT3.1/NAR2.1 and NRT2.1 show pH-dependent and nitrate-elicited currents, while oocytes injected with either NRT2.1 or NRT3.1 alone do not (Zhou et al., 2000). Phosphorylation also appears to play a major role in post-translational regulation of the HATS activity of NRT2.1. NRT2.1 is phosphorylated at Ser 28 when plants are N-starved but rapidly dephosphorylated upon nitrate resupply (Engelsberger and Schulze, 2012). Furthermore, Ser 11 is dephosphorylated and Thr 521 phosphorylated upon nitrate starvation (Menz et al., 2016) and mimicking constitutive phosphorylation of Ser 501 prevents NRT2.1 nitrate uptake activity (Jacquot et al., 2019).

All NRT2 transporters in Arabidopsis, except NRT2.7, can interact with NRT3.1 in yeast. The presence of a Ser residue at or near the same position as in NRT2.1 is conserved in the six remaining NRT2 proteins in Arabidopsis (Kotur et al., 2012; Jacquot et al., 2019), and is well conserved across NRT2.1 homologs in green algae and land plants (Jacquot et al., 2019). This supports the idea that post-translational regulatory mechanisms are of strategic importance for root nitrate uptake not only in Arabidopsis but also in crops.

Research on nitrate transporters over the years has therefore greatly contributed to our current understanding of the nitrate transport machinery in plants and its regulation by external signals. Moreover, this body of work revealed unexpected functions of nitrate transporters in nitrate sensing and signaling. As we discuss on the next section, NRT1.1 has received particular

attention over the last decade due to its proposed role as a nitrate sensor controlling gene expression and plant development in response to nitrate availability, and also due to its ability to transport auxin, a key hormone controlling root growth.

MOLECULAR MECHANISMS OF LOCAL AND SYSTEMIC NITRATE SIGNALING

Nitrate, a Nutrient Signal Controlling Global Gene Expression

In the last 20 years, extensive use of transcriptomics analysis has shown that the expression of a considerable fraction of plant genes is modulated by nitrate treatments (approximately 10% of annotated genes in *Arabidopsis*) (Canales et al., 2014). Such genes are not limited to processes directly related to nitrate (e.g. nitrate transport, reduction or assimilation), but also include genes involved in metabolism of other nutrients (such as C, sulfate, phosphate (P), iron), hormone biosynthesis and signaling, and regulatory genes including kinases, phosphatases and TFs involved in growth and developmental processes (Wang et al., 2000; Wang et al., 2003; Wang et al., 2004; Gutiérrez et al., 2007; Krouk et al., 2010a). The transcriptome response to nitrate is highly dynamic, and changes in transcript expression can be detected as quickly as within 3 min of nitrate exposure (Krouk et al., 2010b; Varala et al., 2018). The nitrate response is also cell- and tissue-specific (Gifford et al., 2008; Walker et al., 2017). A meta-analysis of the transcriptomic response of nitrate-treated roots revealed that ~60% of nitrate-responsive genes respond to nitrate in only one experiment, highlighting the context-specific and complex nature of the nitrate response (Canales et al., 2014).

NR-null plants cannot reduce nitrate to nitrite, the first step in the nitrate assimilation pathway. Nitrate transport and assimilation genes, as well as genes coding for enzymes in C metabolism, are still controlled by changes in nitrate availability in NR-null plants, pointing to a role for nitrate as a signaling molecule (Scheible et al., 1997). Years later, microarray analysis showed that a significant fraction (~ 38%) of nitrate-responsive genes in *Arabidopsis* was able to respond to nitrate in both wild-type and NR-null plants, indicating that their response was independent of nitrate reduction and assimilation (Wang et al., 2004). These experiments were critical to demonstrating the signaling role of nitrate, in addition to its well-known nutritional role.

Nitrate can control long-term responses related to changes in growth and development, as discussed in further sections. Transcriptome reprogramming in response to nitrate is supported by

a temporal hierarchy of TFs that initiates dynamic gene regulatory networks (GRNs), which act at the cellular level to orchestrate organ responses (Varala et al., 2018). In Arabidopsis roots, nitrate triggers functionally distinct but coordinated responses across different cell types (Gifford et al., 2008; Walker et al., 2017). For example, nitrate regulation of auxin signaling in pericycle cells has been linked to LR formation in response to nitrate (Gifford et al., 2008; Vidal et al., 2013; Yu et al., 2016; Walker et al., 2017).

NRT1.1/NPF6.3 is a Nitrate Transceptor

The idea that nitrate could act as a signal led to a search for a nitrate sensor that would directly bind nitrate and activate downstream signaling pathways, similar to the NarX/NarL two-component system in bacteria (Nohn et al., 1989). Phenotypic and transcriptomic analyses of nitrate transporter mutants offered clues as to the identity of the nitrate-sensing apparatus. It turned out to be the transporter itself: *NRT1.1*, first identified as a gene responsible for chlorate sensitivity in Arabidopsis (Tsay et al., 1993). The resolution of the crystal structure of Arabidopsis NRT1.1 was key for understanding the molecular basis of its dual-affinity dependent on nitrate availability. Crystallized NRT1.1 adopts a dimer conformation, with each N-terminal half facing and interacting with each other (Parker and Newstead, 2014; Sun et al., 2014b), as suggested from in vitro crosslinking and Fluorescence Resonance Energy Transfer experiments in *Xenopus* oocytes (Sun et al., 2014b). Importantly, phosphorylation of Thr 101 interferes with NRT1.1 dimerization by introducing electrostatic and conformational changes into the monomers (Sun et al., 2014b). Unmodified NRT1.1 therefore adopts a dimer configuration suitable for low-affinity nitrate uptake, while low N-mediated phosphorylation at Thr 101 triggers conversion to a monomer with higher structure flexibility, which might explain the switch to high affinity (Parker and Newstead, 2014).

NRT1.1 expression in rapidly growing tissues such as the pericycle during LR formation, together with impaired LR primordia elongation in *nrt1.1* mutants, suggested that NRT1.1 function was not restricted to nitrate transport (Guo et al., 2001). Further work proposed a signaling role for NRT1.1 in the control of the high-affinity transporter NRT2.1 (Muños et al., 2004) and for the induction of LR elongation by localized high nitrate availability (Remans et al., 2006).

Demonstration that NRT1.1 acted as a nitrate transceptor came in 2009 (Ho et al., 2009). Transceptors are commonly involved in nutrient transport and sensing in many organisms

(reviewed by Kriel et al., 2011; Steyfkens et al., 2018). Evidence that NRT1.1 was a nitrate transceptor was obtained with an *nrt1.1* mutant that prevented nitrate uptake but retained induction of gene expression in response to nitrate, including *NRT2.1* expression (Ho et al., 2009). We now know that NRT1.1 can activate multiple mechanisms of nitrate sensing and signaling required for regulation of nitrate-responsive genes in Arabidopsis roots (Wang et al., 2009; Bouguyon et al., 2015), but also affects diverse biological processes including root development and architecture, auxin transport, seed dormancy, flowering time and stomatal movements (Fan et al., 2017; Zhang et al., 2018; Fredes et al., 2019).

The Backbone of N Signaling

While the identification of NRT1.1 as a nitrate transceptor was clearly critical, much remained unanswered as to how nitrate was sensed and transduced into changes in gene expression (Figure 2). For instance: how is NRT1.1 phosphorylation regulated? The kinase CIPK23 interacts with CBL9 or CBL1 and phosphorylates NRT1.1 at Thr 101 under low nitrate conditions, switching the transporter to its high-affinity function (Ho et al., 2009). Besides this switch in nitrate affinity, the phosphorylation state at Thr 101 also controls the expression of downstream genes such as the *NRT2.1* transporter and genes involved in N, sulfur and selenium metabolism (Ho et al., 2009; Bouguyon et al., 2015). The protein phosphatase 2C ABI2, normally involved in abscisic acid (ABA) signaling, can dephosphorylate CBL1 and CIPK23, thereby inactivating this calcium-sensor/kinase complex and preventing NRT1.1 phosphorylation (Léran et al., 2015). *abi2* and *nrt1.1* mutants have similar phenotypes, including the loss of low nitrate-dependent *NRT2.1* expression and failure to induce LR elongation in high nitrate versus low nitrate sides in a split-root experimental design (Léran et al., 2015). Since ABA binding to its receptors inactivates ABI2, this could represent a mechanism to decrease nitrate uptake and root remodeling under stress conditions to preserve cellular energy.

The involvement of calcium-related proteins (CBL1, CBL9 and CIPK23) in the control of NRT1.1 function prompted the idea that calcium might act as a second messenger in nitrate signaling. Early studies using maize and barley detached leaves showed that pretreating leaves with the calcium chelator EGTA or the calcium channel blocker LaCl₃ attenuated the expression of nitrate-responsive genes (Sakakibara et al., 1997; Sueyoshi et al., 1999). A more recent study showed that nitrate treatment caused a rapid increase of cytosolic calcium levels, which in turn

triggers changes in gene expression in Arabidopsis roots (Riveras et al., 2015). Downstream of calcium signaling, calcium-dependent kinases (CPKs) act as molecular links between NRT1.1 and NLP TFs, as detailed below. The transport activity of NRT1.1 induces calcium waves through the action of a phospholipase C and Inositol triphosphate IP₃ (Riveras et al., 2015) that are decoded by subgroup III CPKs, which then phosphorylate NLP7, thus promoting its nuclear localization and activation of target genes (Liu et al., 2017). Alignment of the nine Arabidopsis NLPs and four orthologous lotus (*Lotus japonicus*) NLPs identified a conserved Ser 205 residue in NLP7 as a candidate CPK phosphorylation site. Mass spectrometry results confirmed that Ser 205 in NLP7 was 1) phosphorylated in vivo in the presence of nitrate (Liu et al., 2017), and 2) abolished in a *cpk* triple mutant. Transcriptome analysis in the same *cpk* triple mutant revealed that CPK-dependent nitrate-induced genes largely overlapped with genes controlled by NLP7 (Figure 2). These results support a role for CPKs in NLP phosphorylation and in the control of nitrate responses.

Although there have been considerable advances in recent years in the identification of nitrate sensing and signaling components, many questions remain. For instance, the function of the NRT1.1 transceptor does not explain the full extent of nitrate responses, suggesting that additional sensing mechanisms exist in plants (Wang et al., 2009; Bouguyon et al., 2015). In this context, NRT2.1 was suggested as a putative nitrate sensor (Little et al., 2005). Some nitrate-responsive genes hint at both calcium-dependent and calcium-independent branches (Riveras et al., 2015; Liu et al., 2017). Other signaling molecules include phosphatidic acid, G-protein complexes, and reactive oxygen species (Shin et al., 2005; Hong et al., 2009; Liang et al., 2018; Wany et al., 2018; Chakraborty and Kanyuka, 2019) (Figure 2). However, their contribution or cross-talk with the canonical nitrate signaling pathway remains to be addressed.

Systemic Nitrate Signaling Mechanisms

Systemic signaling allows a plant to elicit a plant-wide response by communicating locally-perceived stimuli to distant organs. However, how different parts of the plant can perceive and communicate N status to the whole plant has been poorly studied. Potential systemic signals have been found such as malate, amino acids or the plant growth rate (reviewed in Gent and Forde, 2017). However, whether and how these signals report the plant nitrate status remains unclear (Gent and Forde, 2017). This gap in knowledge on systemic nitrate regulation may originate from the

difficulty to experimentally uncouple the intertwined local and systemic responses in nitrate signaling (Li et al., 2014).

Cytokinins (CKs) do play a central role in root-to-shoot communication in nitrate signaling. Nitrate nutrition, CK content, and shoot growth appear correlated in different plant species (Ruffel et al., 2011; Sasaki et al., 2014; Guan, 2017; Poitout et al., 2018). In *Arabidopsis*, nitrate induces CK synthesis, in particular through the up-regulation of *IPT3*, a gene encoding a key enzyme in CK biosynthesis (Sakakibara et al., 2006). Importantly, CKs move through the plant: the translocation of precursor (*trans*-zeatin) and active CK (*trans*-zeatin riboside) species requires the transporter ABCG14 to load the xylem with root-borne CKs (Ko et al., 2014; Zhang et al., 2014). Once above ground, active and precursor CKs control leaf expansion and shoot apical meristem activity, respectively (Osugi et al., 2017; Landrein et al., 2018). Root-to-shoot *trans*-zeatin translocation is also an important component of N-systemic signaling that controls root responses to nitrate provision. Indeed, plants subjected to homogeneous or heterogeneous nitrate treatment differ in their accumulation of *trans*-zeatin in shoots. Thus, disruption of CK biosynthesis or translocation leads to a lack of *trans*-zeatin accumulation and to repression of genes involved in nitrate transport and metabolism under nitrate-supplied conditions (Poitout et al., 2018). Moreover, CKs play a key role on the systemic N-signal that controls root architecture. When plants are grown in split-root conditions, in which half of the root system is grown in high nitrate concentrations and the other half without nitrate, *ipt* mutants show no proliferation of LR in the high nitrate side as compared with homogenous high nitrate, suggesting that CKs are a systemic signal involved in N-demand signaling. Furthermore, decapitated plants lose both N-demand and N-supply responses, indicating the existence of a root-to-shoot-to-root signal relay (Ruffel et al., 2011) that involves the accumulation of *trans*-zeatin (Ruffel et al., 2016). Accumulation of *trans*-zeatin modifies a N-assimilation pathway in shoots involving Gln (Poitout et al., 2018). Thus, Gln might serve as a shoot-to-root signal communicating nitrate availability in roots at a systemic level. Gln is a possible candidate since amino acids are commonly found in the phloem sap and their role as communicators of N status has been previously described (Muller and Touraine, 1992; Miller et al., 2008; Gent and Forde, 2017).

Besides hormones, small peptides can act as signaling molecules in nitrate signaling. N-deprivation induces the expression of CEP and CLE peptide-encoding genes in roots. CLE1, -3, -4 and -7 can interact with CLV1, repressing LR emergence and growth under N-deprivation. Since

CLE mRNAs are expressed on the pericycle, while *CLVI* is expressed in phloem companion cells, *CLE* peptides are proposed to serve as cell-cell mobile signals, integrating N signals into root responses (Araya et al., 2014b; Araya et al., 2014a).

CEPs are a family of short secreted peptides that serve as signaling molecules in plants. Overexpression of members of the CEP family causes repression of root growth, while their loss promotes root development. CEPs are recognized by the Leucine Rich Repeat-Receptor kinase LRR-RK receptors CEPR1 and CEPR2 in Arabidopsis (Tabata et al., 2014). Inactivation of *CEPRs* causes enhanced LR elongation, leaf chlorosis, dwarfism and anthocyanin accumulation, reminiscent of N-starved plants. In addition, expression of *NRT2.1* and *NRT3.1* as well as nitrate uptake are impaired in *cepr* mutants. Based on split-root experiments, CEPs are specifically induced during nitrate deficiency, while nitrate transporter genes *NRT2.1*, *NRT3.1* and *NRT1.1* are induced in the presence of high nitrate. Grafting experiments using *cepr1 cepr2* double mutant scions and wild-type rootstocks, together with analysis of CEP levels in xylem sap, demonstrated that CEPs act as mobile root-borne signals that are perceived by CEPRs in shoots and mediate systemic N-demand signaling. This systemic shoot-to-root signaling is due to the synthesis in shoots of two polypeptides, CEPD1 and CEPD2, which translocate to roots and activate expression of nitrate transporters specifically on the side exposed to high nitrate (Ohkubo et al., 2017). In contrast to CK signaling that depends directly on nitrate, CEP signaling responds to general N-deficiency, representing a different component of the N-demand systemic signaling.

Another long-distance communication system consists of translocation of TFs as reporters for tissue N-status, for example HY5, as detailed earlier (Chen et al., 2016).

The decoding of local and systemic signaling pathways and the identification of molecular components regulating nitrate responses may thus contribute to a better understanding of how plants sense and respond to changes in N availability, providing unique targets for improving NUE in crops.

REGULATORY NETWORKS OF THE NITRATE RESPONSE IN PLANTS

Genomics, high-throughput validation of protein–DNA interactions, and systems biology approaches have increased the speed at which TFs involved in nitrate responses are being identified. This includes the use of machine learning methods to predict networks (Krouk et al.,

2010a; Varala et al., 2018; Brooks et al., 2019), high throughput plant cell-based TF-perturbation assays such as TARGET (Transient Assay Reporting Genome-wide Effects of Transcription factors, illustrated in Supplemental Figure 1) (Bargmann et al., 2013; Varala et al., 2018; Brooks et al., 2019), yeast-1-hybrid (Y1H) (Gaudinier et al., 2018), and the measure of accessible chromatin regions in response to nitrate (Alvarez et al., 2019). To date, more than 40 TFs spanning several TF families have been identified that bind to the promoters or control the expression of genes involved in nitrate transport, nitrate reduction and nitrate assimilation (Gutiérrez et al., 2008; Rubin et al., 2009; Marchive et al., 2013; Alvarez et al., 2014; Guan et al., 2014; Para et al., 2014; Medici et al., 2015; Xu et al., 2016b; Maeda et al., 2018; Varala et al., 2018; Brooks et al., 2019). This data can be integrated into regulatory networks to further our understanding of how nitrate-dependent changes in gene expression impact plant physiology, growth and development (Figure 3, Table 1). Table 1 summarizes the data associated with the influential transcription factors that appear in Figure 3 and discussed below.

Identification of TFs Involved in Nitrate Signaling

The first TF identified in nitrate signaling pathways was ANR1, a MIKC-type MADS-box TF. *ANR1* was isolated using a classical molecular genetics approach in a screen for nitrate-inducible genes in roots (Zhang and Forde, 1998). *ANR1* loss of function mutants have impaired LR elongation in nitrate-rich localized patches, affecting plasticity of the root system (Zhang and Forde, 1998). It was later found that *ANR1* is controlled at the transcriptional level by NRT1.1 (Remans et al., 2006). Although the overexpression of *ANR1* promotes LR growth and initiation even in the absence of nitrate, the presence of nitrate further potentiates the promotion of LR growth, suggesting that nitrate exerts a post-translational control of ANR1 activity (Gan et al., 2012).

Although characterizing genes whose transcripts are induced has proven a powerful tool to identify important TFs involved in nitrate signaling, many TFs are controlled at the post-transcriptional level (e.g. post-translational modifications and/or control of subcellular localization). This is the case for NLP7, the best characterized TF in the nitrate response to date. NLP7 was initially identified as an orthologue to lotus NIN proteins, involved in early steps of nodulation in legumes (Schauser et al., 1999), and to the NIT2 transcription factor that controls *NR* expression in *Chlamydomonas reinhardtii* (Camargo et al., 2007). *nlp7* mutant plants have

longer primary roots (PRs) and more LR, a phenotype that is characteristic of N-starved plants, supporting the role of NLP7 as a key regulator of nitrate metabolism (Castaings et al., 2009). NLP7 activates the transcription of many nitrate-responsive genes, including nitrate reductases, *NRT2.1*, and *NRT2.2* (Castaings et al., 2009; Wang et al., 2009).

Members of the NLP family can be divided into four subgroups: NLP1/2, NLP4/5, NLP6/7, and NLP8/9 (Schauser et al., 2005; Castaings et al., 2009). Y1H screening using four copies of a 43-bp N response cis-element (NRE) revealed that all NLPs bind the NRE element (Konishi and Yanagisawa, 2013). The kinases CPK10, CPK30 and CPK32 phosphorylate NLP7 in the presence of nitrate, resulting in its nuclear localization and concomitant induction of nitrate-responsive gene expression (Liu et al., 2017) within minutes of nitrate treatment (Marchive et al., 2013). As discussed in the previous section, this mechanism depends on calcium signaling (Riveras et al., 2015; Liu et al., 2017) and coordinates nitrate availability to transcriptome reprogramming. Overexpression of NLP6 in the absence of nitrate is not sufficient to induce nitrate-inducible genes (Konishi and Yanagisawa, 2013), while its N-terminal GAF-like region is necessary for its nitrate-dependent transcriptional activity in protoplasts (Konishi and Yanagisawa, 2013). This suggests that post-translational modifications of NLP6 are similarly required for nitrate control of gene expression.

Previous studies have shown that a single TF does not explain the full induction of genes such as *NRT2.1*, *NRT2.2*, nitrate and nitrite reductases (Castaings et al., 2009; Alvarez et al., 2014; Guan et al., 2014) and that protein-protein interactions between TFs are important for gene expression of nitrate-responsive genes (Xu et al., 2016b; Guan et al., 2017). Nitrate transporters and nitrate metabolic genes are indeed convergent targets of many of the known nitrate-related TFs. The C-terminal PB1 domain of NLP proteins mediates protein-protein interactions in yeast two-hybrid assays. The PB1 domain is distinct from the DNA-binding domain but is necessary for full induction of nitrate-dependent expression of target genes (Konishi and Yanagisawa, 2019). These results are consistent with previous hypotheses on redundant and overlapping functions between different *Arabidopsis* NLP TFs.

NLP6 and NLP7 also physically interact with TCP20 (Guan et al., 2017). TCP20 binds to a 150 bp NRE and a 109 bp enhancer sequence found in the promoters of *NRT2.1* and *NIA1*, respectively, and acts in systemic nitrate signaling for root foraging, integrating cell cycle-related processes and root growth (Guan et al., 2014). NLP6, NLP7 and TCP20 bind to adjacent sites in

the upstream promoter region of the *NIA1* gene. The PB1 domains of NLP6 and NLP7 as well as the histidine- and glutamine-rich domain of TCP20 are important for their physical interaction (Guan et al., 2017).

Additional TFs for nitrate responses have been identified. Using an integrative bioinformatics approach, the genes encoding the TGA-type TFs *TGA1* and *TGA4* were consistently regulated by nitrate across many experiments and therefore were good candidates to mediate nitrate responses (Alvarez et al., 2014). 97% of the genes that have an altered expression in the *tga1 tga4* double mutant relative to wild-type plants are nitrate-responsive, indicating that TGA1 and TGA4 have a global role in nitrate response in roots. TGA1 and TGA4 can bind directly to the *NRT2.1* and *NRT2.2* promoters, and initiation of LR is affected in both *tga1 tga4* and *nrt2.1 nrt2.2* double mutants, suggesting that TGA1 and TGA4 regulate LR development at least partly via *NRT2.1* and *NRT2.2* (Alvarez et al., 2014). Induction of root hair development in response to nitrate treatments is also impaired in *tga1 tga4* double mutants (Canales et al., 2017). TGA1 and TGA4 directly modulate the expression of *CPC*, a key regulator of root hair cell fate, which in turn increases the production of root hairs and nitrate uptake (Canales et al., 2017). Besides the known interactions of TGA1 with other members of the TGA family in the plant stress response (Shearer et al., 2012), putative interactors of TGA1 have been revealed using a high-density Nucleic Acid Programmable Protein Array (NAPPA) to map protein-protein interaction networks (Yazaki et al., 2016). The TF-NAPPA dataset revealed that TGA1 can interact with proteins involved in nitrate signaling such as CIPK23, but also with proteins involved in hormone signaling such as HAI2 and the ABA receptor PYL6, positioning TGA1 as an integrator of hormone and nitrate signaling networks (Yazaki et al., 2016).

High-Throughput Assays to Identify TFs and TF Targets

After identification of key TFs involved in N-signaling, a second step to unravel the complex structure of regulatory networks is to systematically identify TF targets and the impact of these N-regulatory networks on organism function. Different approaches have been undertaken to fulfill this general goal, including integration of available high-throughput data on TF-DNA interactions obtained by technologies such as Y1H, protein-binding microarrays (Weirauch et al., 2014) or DNA Affinity Purification Sequencing (DAP-seq) (O'Malley et al., 2016).

To identify TFs and their targets involved in the regulation of root development in response to nitrate, Gaudinier et al. (2018) used an enhanced Y1H assay to screen for candidate TFs expressed in roots against 98 promoters of genes spanning different processes related to N. They generated a regulatory network comprised of 1,660 protein-DNA interactions between 345 TFs and all 98 promoters. TFs that bind genes participating in multiple N-related processes are more likely to be important for plant growth. This transcriptional network is highly enriched in genes regulated by ABA, auxin, methyl jasmonate, or ethylene. In agreement, mutants in transcription factors involved in ethylene (ERF4, ERF70 and ERF107) and auxin (ARF18) signaling show altered phenotypic responses to N availability (Gaudinier et al., 2018).

Functional assays of TFs in plants or plant cells offer a cellular context of regulatory networks, revealing when physical interactions lead to gene regulation. However, significant hurdles exist to identify the genes acting downstream of TFs within these networks. Mutants or overexpressors of the TF help define genes under the control of each TF, while a high-quality ChIP antibody or epitope-tagged transgenic line is needed to identify promoters bound by each TF; neither step is adaptable to high-throughput approaches. The development of TARGET, a plant cell-based temporal TF perturbation system, has been of tremendous help (Supplemental Figure 1) (Bargmann et al., 2013). The TARGET assay can validate direct TF-target interactions based on TF-induced changes in gene expression (Bargmann et al., 2013; Para et al., 2014; Medici et al., 2015; Doidy et al., 2016; Sparks et al., 2016) or by TF-binding (Para et al., 2014; Doidy et al., 2016). A scalable version of TARGET recently enabled the validation of genome-wide targets for seven N-responsive TFs in shoots (Varala et al., 2018) and 33 N-responsive TFs in roots (Brooks et al., 2019).

TARGET was successful in defining the genome-wide targets of HRS1 (Medici et al., 2015). Direct targets of HRS1 are enriched in genes related to auxin and ABA signaling pathways, as well as P metabolism (Medici et al., 2015). HRS1 and its close homolog HHO1 repress PR growth caused by P starvation, but only when nitrate is present in the medium, indicating that they participate in a pathway at the intersection of N and P signaling (Medici et al., 2015).

TARGET also captured the early and transient action of bZIP1 in the N-response cascade (Para et al., 2014). Time-series ChIP experiments conducted 1-3 min following TF nuclear import revealed that bZIP1 bound transiently to promoters of early N-response genes (Para et al., 2014). 4tU labeling of de novo transcripts to affinity-capture bZIP1-initiated mRNAs determined that

transiently bound bZIP1 targets were actively transcribed at times when the TF was no longer bound (Doidy et al., 2016). Importantly, these transiently-bound and regulated bZIP1 targets were the most relevant to early N-signaling in whole roots (Para et al., 2014). This provided the first genome-wide evidence for “Hit-and-Run” transcription in plants. This model posits that a TF trigger (the Hit) can organize a stable transcriptional complex, including recruitment of other TFs, so that transcription can continue even after the initiating TF is no longer bound (the Run). This opened a new perspective on dynamic aspects of transcriptional regulation in plants (Charoensawan et al., 2015; Varala et al., 2015).

Recently, DNase I hypersensitivity-seq assay (DNase-seq) uncovered TF-target interactions, taking into account the native chromatin structure during nitrate response of *Arabidopsis* roots (Alvarez et al., 2019). Local changes in DNase I cleavage patterns were observed in response to nitrate at specific loci bound by TFs. Rapid changes in TF occupancy correlated with RNA polymerase II (RNPII) recruitment and with changes in transcript accumulation in response to nitrate. By integrating genomic footprinting, transcriptional regulation of gene expression based on RNPII occupancy, and transcriptome data, a regulatory network was constructed that captures the relative contribution of known and new TFs in nitrate signaling. The network captured an interaction between NAP and *NRT2.1*, validated by ChIP and TARGET assays, illustrating NAP as a direct activator of *NRT2.1* expression and a novel positive regulator of nitrate uptake in roots (Alvarez et al., 2019).

Systems Approaches to Model Gene Regulatory Networks (GRNs) in Plants

Because of the complex underlying structure and relative recalcitrance to classical genetics, nitrate regulatory networks are an ideal framework for systems biology approaches to reveal features potentially considered as emergent properties (Gutierrez, 2012).

Interestingly, the N community has been among the first to pioneer modeling approaches and data integration in plant biology. MapMan was one of the first data integration tools (Thimm et al., 2004) and opened avenues towards the integration of transcriptomic datasets with metabolic insights (Gibon et al., 2006; Schwacke et al., 2019). MapMan was recently used to find enriched functional categories dependent on CPK10, CPK30 and CPK32 kinases that regulate early steps in nitrate signaling enriched in N metabolism, glycolysis, carbohydrate metabolism, and N transporters (Liu et al., 2017).

This was followed by the Arabidopsis MultiNetwork model, which integrates genes, metabolites, miRNA, and molecular interactions such as protein-protein, protein-DNA, and miRNA-target (Gutiérrez et al., 2007). This resource revealed relationships between N and the circadian clock via the CCA1 TF (Gutiérrez et al., 2008) and uncovered connections between miR167 and *ARF8* in the pericycle of roots (Gifford et al., 2008).

To broadly enable such approaches, the VirtualPlant website was created to provide a platform for systems biology where biologists can upload their genomic data and execute various data visualization and analysis tools (Katari et al., 2010). VirtualPlant's GeneCart supports the iterative nature of systems biology analysis and allows users to save results from one analysis and feed it into another. Applications range from Gene Ontology (GO) enrichment analysis to gene network visualization and analysis using the integrated Arabidopsis MultiNetwork (<http://virtualplant.bio.nyu.edu/cgi-bin/vpweb/>).

Myriad transcriptomics studies have evaluated different aspects of the plant N response, such as the effect of N source (Wang et al., 2004; Wang et al., 2007; Ristova et al., 2016), time after nitrate treatments (Krouk et al., 2010a; Varala et al., 2018), nitrate concentration (Wang et al., 2007), tissue type (Wang et al., 2003), and cell type (Gifford et al., 2008). An integrative meta-analysis of all data revealed the most consistently regulated genes by nitrate across different experimental conditions (Canales et al., 2014). The large number of TFs in this set of genes suggests that transcriptional reprogramming is at the core of plant adaptation to changes in nitrate availability. Weighted gene co-expression network analysis (WGCNA) (Langfelder and Horvath, 2008) identified gene modules associated with specific biological functions including C metabolism and trichoblast differentiation (Canales et al., 2014). It was later confirmed that nitrate regulates trichoblast differentiation through TGA1 (Canales et al., 2017).

In contrast to correlation-based methods, time series-based machine learning approaches can predict causality, i.e. the regulatory influence of TFs on their targets in the dataset and in out-of-sample data. Many algorithms have been developed to address these tasks including Boolean networks, Bayesian networks, regularized linear regression or non-linear tree-based regression approaches (reviewed in Krouk et al., 2013).

To identify TFs in a temporal N-initiated cascade, TF-target edges were inferred using a time series-based machine learning method called Dynamic Factor Graph (DFG) (Mirowski and LeCun, 2009; Krouk et al., 2010a). This DFG approach learns network models that can predict

gene expression states at untested time points (Krouk et al., 2010a). An early application of this approach identified SPL9 as a network hub of early nitrate response genes in roots, which was confirmed to modify the nitrate response kinetics of *NIA2*, as well as of many nitrate-controlled genes (Krouk et al., 2010a). A more recent application of the DFG approach inferred a GRN containing 172 nitrate-responsive TFs and 2,174 dynamic nitrate-responsive genes in shoots (Varala et al., 2018). Using validated genome-wide targets for three TF hubs in this GRN identified by the functional TF-target assay (CRF4, SNZ and CDF1), a precision cutoff was experimentally determined to “prune” the TF-target predictions in the network to high-confidence 155 TFs and 608 targets (Varala et al., 2018). This network precision was reconfirmed using genome-wide TF-target regulation data for four additional TFs: TGA1, HHO5, HHO6, and PHL1 (Varala et al., 2018). This experimentally-refined GRN identified the temporal relationship of known and validated regulators of N signaling, including NLP7, NLP8, TGA1, TGA4, NAC4, HRS1, and LBD37, 38 and 39, as well as 146 novel TF regulators. CRF4, the earliest TF in the time-based N-response cascade, was validated to modulate ¹⁵N-uptake and NUE in planta (Varala et al., 2018).

Similarly, the DFG machine learning algorithm was used to infer TF-target edges for 145 N-responsive TFs in roots (Brooks et al., 2019). The inferred TF-target edges were pruned for high-confidence edges using 71,836 experimentally validated TF-target edges for 33 TFs identified using TARGET, which were then used in a precision/recall analysis to generate a GRN consisting of interactions between 145 TFs and 311 targets in the root N response (Brooks et al., 2019). To integrate data from TF-target interactions identified in root cells via TARGET with in planta TF perturbation data and high-confidence predicted edges in the GRN, the “Network walking” approach was developed. These combined datasets were used to connect TFs to their direct targets in root cells and to their indirect targets in planta via intermediate TFs. Network walking showed that TGA1 directly regulated 40% (508 of 1458) of the N-responsive genes in roots, including 63 of 145 N-responsive TFs. Moreover, 76% of the indirect targets of TGA1 in planta were linked to TGA1 through 49 intermediate TFs. This suggests that TGA1 is a high-level regulator whose effect on downstream secondary TFs amplifies a transcriptional cascade to control N-responses (Brooks et al., 2019). Moreover, TGA1 was among the most influential TFs in nitrate transport, nitrate reduction and nitrate assimilation when publicly available data of TF-target interactions for TFs involved in response to nitrate were interrogated (Figure 3, Table 1).

The flood of genomic data over the last decade in the plant nutrition community and others has been the impetus for increased collaboration between the fields of computer science and biology. Moving forward, there is an immediate need to make better use of existing data from Arabidopsis studies by developing new data integration tools and predictive modeling algorithms. Using Arabidopsis as a framework to learn how to integrate diverse data sets should facilitate similar analyses in species with fewer resources.

DEVELOPMENTAL OUTPUTS

Plants are highly plastic organisms that control growth and development of their organs in order to optimize nutrient acquisition and use. N availability and N-related GRNs exert a major control over endogenous developmental programs in plants, from germination to vegetative and reproductive development.

Nitrate Control of Seed Dormancy and Germination

Different environmental signals control seed dormancy, including temperature, light quality and quantity, oxygen, water potential, organic acids, pH and nitrate availability. The integration of these signals allows the seed to have temporal information about the time of the year and suitability for germination. Nitrate can act as a potent signal controlling seed dormancy and germination in different plant species. Arabidopsis plants exposed to high nitrate provision produce seeds that are less dormant than seeds from plants exposed to low nitrate. Similarly, seeds exposed to exogenous high nitrate are less dormant than low-nitrate exposed seeds. Both effects are independent of nitrate reduction, suggesting that nitrate itself functions as a signal for the induction of germination (Alboresi et al., 2005). Parts of this regulation are due to the signaling function of NRT1.1, since 1) a null *nrt1.1* mutant fails to induce germination in the presence of low nitrate (Alboresi et al., 2005) and 2) *NRT1.1* expression increases before relief of dormancy during seasonal cycling (Footitt et al., 2011).

Analysis of laboratory dormancy cycling in Cape Verde island (Cvi), an Arabidopsis accession with strong dormancy, documented low CIPK23 expression in low-dormant and non-dormant states, when seeds are more sensitive to nitrate, and high during primary and secondary dormancy, when sensitivity to nitrate is low (Cadman et al., 2006; Footitt et al., 2013). This

indicates that phosphorylation of NRT1.1 might play a role in seed nitrate sensitivity and control germination timing. However, transcriptome analysis of Cvi exposed to different treatments inducing germination, including drying after ripening, seed hydration, cold, light and exogenous nitrate showed that all these signals elicit similar changes to the transcriptome of the germinating seed (Finch-Savage et al., 2007). Exposure to nitrate or cold stratification translates into similar changes at the germinating seed proteome level (Arc et al., 2012) and in the metabolism of mother plants exposed to low nitrate and low temperature during seed maturation (He et al., 2016), suggesting that integrators of common downstream environmental signals may act to control seed dormancy and germination. The hormones ABA and gibberellic acid (GA) are attractive candidates, given their roles in promoting seed dormancy and seed germination, respectively. Nitrate reduces levels of ABA in dormant seeds (Ali-Rachedi et al., 2004), by inducing the cytochrome P450 *CYP707A2* (Matakiadis et al., 2009), encoding the main ABA catabolic enzyme present in the late-maturation stage of seed development and in hydrated seeds (Okamoto et al., 2006b). This regulatory mechanism is mediated by nitrate and involves NLP8, which is required for nitrate induction of *CYP707A2* and many nitrate-controlled genes in seeds including *NIA1*, *NIA2*, *NIR*, *RFNRI*, *G6PD2* and At1g25550, pointing to a key role for NLP8 in the seed nitrate response. NLP8 can bind to the *CYP707A2* promoter in three sites that are important for nitrate control. Nitrate induction of *CYP707A2* via NLP8 reduces ABA levels in seeds, relieving dormancy (Yan et al., 2016). As previously discussed, the nitrate sensor NRT1.1 is involved in nitrate-mediated seed dormancy relief. Although *nrt1.1* seeds are less sensitive to nitrate, normal germination levels can be achieved by increasing nitrate concentrations (Alboresi et al., 2005; Yan et al., 2016). *nlp8* mutants, on the other hand, are insensitive to nitrate over a wide range of concentrations (Yan et al., 2016), indicating that alternative nitrate sensors may act upstream of NLP8 in seeds in addition to NRT1.1.

Nitric oxide (NO) has been proposed as such an alternative signal. NO scavengers abolish the positive effect of nitrate on seed germination and NO induces *CYP707A2* (Bethke et al., 2006; Liu et al., 2009; Matakiadis et al., 2009). NO also controls the expression of *ABI5*, a key repressor of seed germination and post-germinative development (Finkelstein and Lynch, 2000; Lopez-Molina et al., 2001). NO triggers degradation of group VII ERF TFs, via the N-end rule pathway (Graciet and Wellmer, 2010). Degradation of group VII ERFs leads to a decrease in *ABI5*, releasing seed dormancy (Gibbs et al., 2014). Furthermore, NO can control *ABI5* protein levels. NO causes

S-nitrosylation of ABI5 in Cysteine 153 during seed germination. This S-nitrosylation controls ABI5 protein degradation via a proteasome-dependent pathway involving CUL4 and KEG E3 ubiquitin ligases, leading to the promotion of germination (Albertos et al., 2015). Besides ABI5, NO has been shown to S-nitrosylate SnRK2 proteins, key positive ABA signaling components, inactivating them and positively regulating germination and early seedling growth (Wang et al., 2015). In addition, NO induces Tyrosine nitration and inactivation of ABA receptors PYR1 and PYL8 in plants (Castillo et al., 2015), further reinforcing the repressive effect over ABA signaling.

Nitrate Influences the Heterotrophic-Autotrophic Switch and Seedling Establishment

After seed germination, the emerging seedling initially grows fueled by seed reserves. Seedling establishment is complete when the seedling acquires photosynthetic capability and transitions to autotrophy. This process involves a reprogramming for the efficient use of endogenous reserves as well as resources that can be incorporated from the environment. N availability is an important cue for early post-germinative growth, and positively influences cotyledon expansion and greening, hypocotyl growth, and the emergence of true leaves (Sato et al., 2009). Mobilization of triacylglycerols (TAGs), the main seed reserve in many plant species, is regulated by the C/N ratio, with high C/N ratios repressing eicosenoic acid breakdown. This represents a feedback mechanism in which accumulation of sucrose, the main product of TAG mobilization, represses further TAG breakdown. Genes encoding proteins involved in photosynthesis are also repressed by a high C/N ratio (Martin et al., 2002). C/N ratio signaling controlling post-germinative growth is mediated by the membrane-localized CNI1 ubiquitin ligase (Sato et al., 2009). Under high C/N ratios, the CIPK protein kinases CIPK7, CIPK12 and CIPK14 accumulate, physically interact with and phosphorylate CNI1, resulting in its stabilization. CIPK14 and CBL8 can also phosphorylate CNI1 in a calcium-dependent manner (Yasuda et al., 2017). Ser/Thr phosphorylation of the C-terminus of CNI1 is essential for its interaction with 14-3-3 proteins, leading to their ubiquitination and degradation (Sato et al., 2011; Yasuda et al., 2014). 14-3-3 proteins can bind phosphorylated motifs on enzymes involved in C and N metabolism such as nitrate reductase, glutamine synthetase, sucrose phosphate synthase and ADP-glucose pyrophosphorylase, regulating their activity in response to environmental signals (Bachmann et al., 1996; Comparot et al., 2003). Although a detailed mechanism is still elusive, the involvement of 14-3-3 proteins in downstream C/N control of post-germinative growth is an example of how

dynamic changes in Ca^{2+} and protein phosphorylation can convey nutritional signals into the developmental programs of plants.

Nitrate-Controlled Shoot Growth

Nitrate availability has profound effects on the growth of aerial tissues. The positive effect of high nitrate on leaf growth and branching have been attributed to CKs (Rahayu et al., 2005; Müller et al., 2015). CKs are synthesized in response to nitrate in roots and translocated via the xylem to the shoots, where they control shoot growth (Takei et al., 2001; Takei et al., 2004; Sakakibara et al., 2006; Osugi et al., 2017). In particular, translocated *trans*-zeatin controls leaf size while *trans*-zeatin riboside controls leaf size and shoot apical meristem activity (Osugi et al., 2017). Nitrate provision induces CK signaling in the shoot apical meristem, increasing expression of the homeodomain TF gene *WUS* and increasing the expression domain of *CLV3*. This leads to stem cell proliferation and increased meristem size resulting in bigger rosettes and more flowers (Landrein et al., 2018).

Compared to nitrate sufficiency, low nitrate concentrations repress secondary shoot formation by inhibiting the elongation of axillary buds, leading to the production of elongating branches only on the most apical cauline nodes. In contrast to other developmental responses to nitrate provision, this phenotype is not dependent on nitrate, but on overall N status of the plant. Strigolactones (SLs), hormones that are produced in the roots, are required for this N-limitation response, since mutants lacking SLs fail to repress branching. Auxin, a central negative regulator of root branching, is produced in leaves and transported rootward through the polar auxin transport system. In N-limited plants, the amount of exported auxin increases only in apical stem segments. This response is lost in mutants lacking SLs, indicating that an SL-auxin interaction is needed to control shoot branching in response to N availability (de Jong et al., 2014). On the other hand, CK mediates the positive effect of high nitrate on branching, since CK biosynthetic mutants *ipt3 ipt5 ipt7* and signaling mutants *arr3,4,5,6,7,15* have less active buds than wild-type plants (Müller et al., 2015). High nitrate can mimic the effect of CK supply on the accumulation of auxin transporters PIN3, PIN4 and PIN7 in the plasma membrane of xylem parenchyma cells of basal inflorescence internodes. This suggests that high nitrate might control branching by CK-mediated control of auxin transport by PIN3, PIN4 and PIN7 (Waldie and Leyser, 2018).

Root System Architecture Modulation by Nitrate

Roots acquire nutrients and provide an interface to interact with soil organisms and other plants. As such, roots system traits are attractive targets for breeding programs and biotechnological applications (Den Herder et al., 2010; Rogers and Benfey, 2015). The root system architecture (RSA), or the given spatial disposition of the root system, is determined by both internal, development-related cues and external biotic and abiotic cues. The RSA is highly plastic, which allows the plant to efficiently forage for nutrients and water in the soil, where these resources are commonly unevenly distributed. The effects of N on root growth depend on N source and availability and can impact development and growth of PRs, LRs and root hairs.

A breakthrough in our understanding of the molecular basis of the root response to nitrate came in the late 1990s (Zhang et al., 1999). In this now classic article, ANR1 was identified as a nitrate-responsive TF that controls LR elongation in response to localized high nitrate availability. This same phenotype was later reported for *nrt1.1* mutants, and it was shown that ANR1 acted downstream NRT1.1 in this pathway (Remans et al., 2006). Interestingly, the nitrate transporter NRT1.1 also acts as an auxin transporter (Krouk et al., 2010b). NRT1.1 represses the growth of pre-emerged LRs and LR primordia by preventing auxin accumulation in LR tips via basipetal transport of auxin when nitrate is absent or present in very low concentrations. The auxin transport function of NRT1.1 is inhibited by high nitrate, which allows for the outgrowth of LRs under sufficient nitrate availability (Krouk et al., 2010b). The repressive function of NRT1.1 on LR development depends on the phosphorylated form of NRT1.1, since the phosphomimetic mutant T101D presents a wild type phenotype, while a non-phosphorylatable mutant T101A exhibits a phenotype similar to an *nrt1.1* null mutant, which is characterized by a marked increase in LR density and length under low external nitrate concentration (Bouguyon et al., 2015). This increased LR development correlates with an increased auxin accumulation in root tips in the T101A mutant, confirming that the role of NRT1.1 on LR development is dependent on its auxin transporter function (Bouguyon et al., 2015).

Although the presence of nitrate strongly induces *NRT1.1*, the role of the encoded transporter is most evident in the absence of nitrate. To reconcile this fact, a detailed analysis of NRT1.1 protein dynamics throughout LR primordium development was conducted in the presence and absence of nitrate (Bouguyon et al., 2016). Nitrate repressed NRT1.1 protein accumulation

during pre-emergence of LR primordia, which promotes local auxin accumulation and induction of LR primordium outgrowth, supporting the fact that NRT1.1 prevents both auxin accumulation and LR growth in response to low nitrate availability (Bouguyon et al., 2016). This suggests a finely tuned mechanism for the control of NRT1.1 function in which post-transcriptional mechanisms such as phosphorylation and protein accumulation play an important role in nitrate-dependent LR development.

As previously discussed, a signaling cascade comprised of calcium and calcium-related kinases act downstream of NRT1.1 to control GRNs that shape RSA in response to nitrate. Although NLP6/7 have been proposed as key TFs within these GRNs, other TFs include TGA1/4 (Alvarez et al., 2014), NAC4 (Vidal et al., 2013), HRS1 and HHO1 (Medici et al., 2015; Kiba et al., 2018; Maeda et al., 2018), TCP20 (Guan et al., 2017), CRF4 (Varala et al., 2018), SPL9 (Krouk et al., 2010a) and NRG2 (Xu et al., 2016b).

In the last 15 years, various lines of evidence have shown that plant hormones play a key role as integrators of N-signals into root developmental programs. Transcriptome analysis has indicated that the interaction of N with hormones such as auxin, CKs and ABA generates a rapid reprogramming of the root transcriptome that impacts root development-related networks to enact quantifiable changes in root architecture (Ristova et al., 2016). Hormones in turn can control the expression of genes involved in N uptake, metabolism and signaling, providing feed-back control over N signals (Gaudinier et al., 2018).

Due to its key role as a master regulator of root development, auxin has been the preferred candidate to evaluate in nitrate-dependent root responses. Pioneering studies from the mid-1950s provided the first insights into the interaction between N and auxin signals. These studies described an inverse relationship between N supply and PR growth, attributed to changes in auxin levels (Bosemark, 1954). Later work with auxin-related mutants and treatment with auxin analogs or auxin transport inhibitors suggested that nitrate effects on PR and LR growth depend on auxin homeostasis and signaling (Zhang et al., 1999; Forde and Lorenzo, 2001; Guo et al., 2005; Tian et al., 2008). These findings were further supported by transcriptome analysis and genome-wide TF binding analysis demonstrating that nitrate availability can regulate the levels of auxin transporters, biosynthesis genes and signaling genes (Canales et al., 2014). For example, expression of *TAR2*, encoding the first step from L-tryptophan to indole-3-pyruvic acid, is induced by low nitrate, leading to an increase in IAA levels in LR primordia and emerging LR (Ma et al., 2014). Low

nitrate can also induce the local levels of IAA in LR primordia and LR roots by regulation of *AGL21*, encoding a TF closely related to *ANR1*. *AGL21* in turn regulates the levels of transcripts coding for enzymes involved in auxin biosynthesis *YUC1* and *YUC5*, *TAR3*, *NIT4* and *AAO1*, stimulating LR initiation and growth (Yu et al., 2014).

Auxin transport from shoots to roots has been proposed as a long-range signal controlling LR development in response to nitrate (Guo et al., 2005; Liu et al., 2010). Auxin transport is mainly facilitated by members of the PIN polar transporters. In rice, expression of the PIN transporters *PIN1c*, *PIN2*, *PIN9* and *PIN10a-b* is diminished in an *OsNAR2.1* knockdown line, and is accompanied by an inhibition in LR development under low nitrate condition (Huang et al., 2015). This effect might be due to a signaling function of the *OsNRT2.1* transporter, a partner of *OsNAR2.1*, since no differences in nitrate content were found in a *OsNAR2.1* knockdown line. As discussed above, *NRT1.1* is also an auxin transporter, repressing the growth of pre-emerged LR roots and LR primordia by preventing auxin accumulation in LR tips when nitrate is absent or very low (Krouk et al., 2010b).

Auxin perception and nitrate signaling are also closely interconnected. Nitrate induces transcription of the *AFB3* auxin receptor gene, while N metabolites produced by nitrate reduction and assimilation reset *AFB3* levels over time by post-transcriptional regulation via *miR393* (Vidal et al., 2010). Downstream of *AFB3*, a pericycle regulatory mechanism involving *IAA14* and the *NAC4* and *OBP4* transcription factors induce LR initiation and elongation in response to nitrate resupply of N-deficient roots (Vidal et al., 2013). By a still unknown mechanism, *AFB3* can also repress PR elongation in response to nitrate (Vidal et al., 2010; Vidal et al., 2013). Similarly, P deficiency represses the expression of the *TIR1* auxin receptor gene, leading to repression of LR development (Perez-Torres et al., 2008), indicating specific roles for auxin receptors in conveying signals from different nutrients. Still in the pericycle, a second auxin-dependent regulatory mechanism is mediated by *miR167*, where it is repressed by nitrate, which then leads to a rise in the mRNA levels of its target *ARF8*. *ARF8* in turn controls a module of nitrate-responsive genes, leading to LR elongation (Gifford et al., 2008).

Together with auxin, CK is also a key regulator of developmental and growth processes in roots. As with auxin, nitrate availability controls the levels of genes related to CK biosynthesis and signaling such as the *IPT* genes, which catalyze the rate-limiting step of CK biosynthesis. CKs are transported to the shoot, where they control vegetative growth (Takei et al., 2001; Takei et al.,

2004; Sakakibara, 2006), but can also control the expression of nitrate transporters and nitrate metabolism-related genes (Kiba et al., 2010). Moreover, as discussed previously, one of the main roles of CKs is as communicator of systemic N-signaling that controls root architecture (Ruffel et al., 2011; Ruffel et al., 2016; Poitout et al., 2018). Recently, a role for CKs was reported in the control of PR growth (Naulin et al., 2019). In plants grown in the presence of nitrate, PR growth is stimulated by promotion of cell proliferation and expansion, compared to plants grown without nitrate. Mutants in the key CK biosynthetic genes *IPT3* and *IPT5*, and CK receptors *AHK2*, *AHK3* and *AHK4* show reduced PR growth when grown on nitrate, indicating that cytokinin biosynthesis and perception are needed for nitrate-stimulated PR growth. This alteration in PR growth is due to a failure in cell division in the meristem at early stages of post-embryonic development, which is in part due to an altered expression of cell cycle checkpoint genes such as *KRP1*, *SKP2 α* and *AXR3* in *ahk* mutant plants in the presence of nitrate (Naulin et al., 2019).

ABA is a key stress hormone integrating external abiotic and biotic stimuli that controls stomatal closure and seed dormancy, but also PR and LR development. Work with ABA biosynthesis and signaling mutants showed that the repressive effect of high nitrate on LR elongation depends on ABA (Signora et al., 2001). Using an anti-ABA antibody, immunocytochemistry experiments showed that high nitrate availability triggers ABA accumulation in the endodermis and stele of the meristem and elongation zones of PRs and emerging LRs (Ondzighi-Assoume et al., 2016). Indeed, nitrate induces the accumulation of the ER-localized beta-1,3-glucanase 1 BG1, thus releasing bioactive ABA into the cytosol from inactive ABA D-glucopyranosyl ester conjugates. Nitrate can also induce the expression of ABA-responsive genes such as *RAB18*. ABA in turn induces the expression of nitrate-responsive genes *NIA1*, *NIA2* and *NIR* to control root growth (Ondzighi-Assoume et al., 2016). Interestingly, in a parallel to NRT1.1 and auxin, NRT1.2 transports ABA (Kanno et al., 2012). Similarly, the *Medicago truncatula* NPF member NPF6.8 transports nitrate and ABA, modulating root length (Pellizzaro et al., 2014). This highlights a complex mechanism of nitrate-ABA interaction in nitrate-dependent control of root growth.

Ethylene is a gaseous hormone that controls many developmental aspects in plants and has been associated with nutritional responses to P, iron and potassium (K) (Romera et al., 2016). Treating plants first grown on low nitrate with high doses of nitrate can induce a rapid burst of ethylene in LR primordia and initiating LRs, resulting from increased *ACC1* and *ACO1* expression

and corresponding enzymatic activities, key steps in ethylene biosynthesis (Tian et al., 2009). Ethylene regulates the expression of the nitrate transporters *NRT1.1* and *NRT2.1*, and mediates the repressive effect of high nitrate treatment on LR initiation and LR elongation (Tian et al., 2009). Similarly, nitrate deficiency, a condition that induces LR elongation (Khan et al., 2015), also triggers ethylene biosynthesis and stimulates ethylene signaling by inducing the expression of ethylene signaling components EIN3 and EIL1. Under low nitrate conditions, *NRT2.1* is upregulated and positively affects ethylene biosynthesis and signaling. In a negative feedback mechanism, ethylene in turn represses *NRT2.1* expression, reducing high-affinity nitrate uptake, and alleviating external nitrate deficiency stress (Zheng et al., 2013).

The role of other plant hormones in mediating changes in root development and growth in response to nitrate availability has been less investigated. However, regulatory network analysis supports a possible role for other hormones such as methyl-jasmonate, GA and brassinosteroids in root responses to nitrate (Gaudinier et al., 2018). In agreement, a recent genome-wide association study (GWAS) conducted to identify novel regulators of root foraging under N deficiency identified BSK3, a plasma membrane kinase associated with early brassinosteroid signaling (Ren et al., 2019). *bsk3* mutants have shorter PR and LR under mild N deficiency due to a failure to induce cell elongation (Jia et al., 2019). However, it is currently unknown whether this mechanism depends directly on nitrate or whether it interacts with nitrate signaling components such as NRT1.1.

Control of Flowering Time by Nitrate

The transition from vegetative to reproductive growth is a crucial developmental decision during the life cycle of a plant, and directly impacts plant yield, fertility and survival of the progeny. As such, the timing of this transition is finely regulated by both endogenous and exogenous cues, by a complex network of interacting signaling pathways whose outputs converge onto the flowering time integrators SOC1 and FT, which in turn activate the expression of floral meristem identity genes such as *API*, *LFY* and *CAL*. Distinct pathways that control flowering time integrate photoperiod, cold exposure (vernalization), plant age, ambient temperature, GA signaling and the autonomous pathway (Davis, 2009; Kim et al., 2009; Song et al., 2012; Wang et al., 2014; Capovilla et al., 2015). The plant nutritional status is another major environmental factor influencing flowering. The role of N in flowering was reported over a century ago (Klebs, 1913)

and fertilization strategies to modulate the timing of flowering to control yield are widely utilized in agriculture. Mutants in nitrate transport or metabolic genes have flowering time phenotypes (Tocquin et al., 2003; Seligman et al., 2008). The effect of nitrate over flowering can be described by a U-shaped curve, where both extremes of nitrate availability repress flowering, albeit by different mechanisms, while intermediate nitrate concentrations promote flowering (Lin and Tsay, 2017).

It is usually difficult to separate between the effects of nitrate availability on vegetative growth and those directly affecting developmental phase change, since plant nutritional status also impacts their overall growth rate. In order to do just that, Castro-Marin et al. (2011) designed an experimental system where 4 mM Gln was added to the medium to maintain constant internal levels of amino acids and proteins. Increasing nitrate concentrations (1, 10 or 35 mM) delayed flowering time in a dose-dependent manner. Plants at time of flowering had more leaves and produced more shoot biomass as nitrate levels increased, but retained the same relative growth rate. Interestingly, withholding Gln from the medium abolished the acceleration of flowering caused by low nitrate levels, possibly due to a general slow plant growth rate (Castro Marín et al., 2011). This is consistent with the U-shaped flowering response to nitrate, in which concentrations of nitrate severely limiting plant growth (as experienced with low nitrate and no Gln) delays flowering time (Lin and Tsay, 2017; Gras et al., 2018). However, flowering time mutants retained their response to nitrate (plants grown on low nitrate still flowered earlier), indicating that nitrate acts in an independent pathway to control flowering.

Further studies have shown that nitrate availability can control expression or activity of different floral regulator genes, thus supporting a mechanism by which nitrate signaling directly impacts flowering signaling pathways. Low N availability induces *FNR1* gene expression, as well as *CRY1*, encoding a blue light photoreceptor that acts in the photoperiod pathway and regulates the expression of *CO* and *FT*. Through FNR1, N availability regulates the NADPH/NADP⁺ ratio and ATP/AMP ratio, with low N increasing NADPH/NADP⁺ and ATP/AMP ratios. In contrast, high N decreases NADPH/NADP⁺ and ATP/AMP ratios, the latter positively regulating protein levels of AMPK α 1, the catalytic subunit of AMP-Activated Protein Kinase (AMPK), as well as its circadian oscillation amplitude. AMPK then phosphorylates CRY1 in the nucleus, leading to its degradation and late flowering. This control of CRY1 abundance may in turn control the amplitude

(but not the period) of the expression of central clock components *CCA1*, *LHY* and *TOC1*, controlling flowering time as a function of external N provision (Yuan et al., 2016).

Nitrate availability also regulates the expression of the flowering repressor *FLC*, as well as the *FT*, *LFY* and *API* genes. Indeed, high nitrate represses *FT*, *LFY* and *API* and induces *FLC*, leading to late flowering (Kant et al., 2011). In a different study, Arabidopsis plants first grown in high nitrate and then subjected to a short-term exposure to low nitrate experienced a rise in the levels of the floral integrator *SOC1*, of the GA biosynthesis gene *GAI* and the photoperiod gene *CO*, with no effect on *FRI* and *VIN3*, or *LD* and *FCA*, participating in the vernalization and autonomous pathways, respectively. Increased levels of *GAI* correlated with an increase in bioactive GA₃ when plants were grown on low nitrate. These results indicate that low nitrate might interact with the photoperiod pathway through CO and with the GA pathway through GA1, leading to an induction in *SOC1* transcript levels and promotion of flowering (Liu et al., 2013). Nitrate availability also controls the levels of DELLA proteins, known repressors of GA signaling: higher nitrate concentrations promote the accumulation of RGA, in turn inducing expression of the RGA targets *GNC* and *CGA1*, also known as *GNC-LIKE*. Expression of floral repressors *SMZ* and *SNZ* also depends on external nitrate provision: higher nitrate induces early accumulation of *SMZ* and *SNZ*, leading to a delayed peak of *FT* expression and therefore delays the timing of flowering under high nitrate. Interestingly, *SMZ*, *SNZ* and *FT* responses to nitrate are abolished in a *della* quintuple mutant. Furthermore, *GNC* and *CGA1* over-expressors exhibit higher expression of *SMZ* and *SNZ*, while *SMZ* and *SNZ* expression is diminished in the *gnc cga1* double mutant relative to wild-type. These results indicate that nitrate-dependent DELLA accumulation has a direct impact on the temporal expression of *SNZ*, *SMZ* and *FT*, resulting in differences in flowering time, and highlights the roles of components of the age-dependent pathway on nitrate-controlled flowering (Gras et al., 2018). Consistent with the involvement of the age pathway, mRNA levels in SPL TFs (miR156 targets, and therefore influencing flowering time), are temporally regulated by nitrate availability in the SAM. *SPL3* and *SPL5* levels showed a delayed peak when plants were grown in low nitrate conditions, which causes a late flowering phenotype (Olas et al., 2019). However, this regulation is independent of miR156 and may instead depend on direct control of *SPL* expression by nitrate in the SAM via the activity of NLP6 and NLP7 TFs (Olas et al., 2019).

NITRATE INTERACTION WITH OTHER NUTRIENTS

Crosstalk between nutrient signaling pathways is an important feature of plant adaptation and survival in natural environments. Under natural or agricultural field conditions, plants are exposed to variations in the levels of multiple nutrients at the same time. The macronutrients N, P and K are the most limiting nutrients for crop yield and the most widely required as fertilizers on agricultural soils. The response to combined nutrient deficiency or to combined changes in nutrient levels and their impact on plant physiology and growth is an emergent area of study in plant biology. Accordingly, the identification of molecular integrators involved in N, P and K interactions has increased its relevance in the last years.

Nitrate and P Interactions

The first evidence linking N and P nutrition was the discovery of the *NLA* gene, which plays a role on the crosstalk between nitrate and P in Arabidopsis. *NLA* was initially described as having a role in N limitation responses (Peng et al., 2007). Mutants in *NLA* exhibit N-limitation specific phenotypes (Peng et al., 2008) that are reverted by P limitation in an antagonistic crosstalk between N and P levels (Kant et al., 2010). *NLA* can also trigger the polyubiquitination and N-deficiency mediated degradation of the ORE1 TF (Park et al., 2018), leading to leaf senescence. Interestingly, *NLA* utilizes the E2 ubiquitin-conjugating enzyme PHO2 for ORE1 ubiquitination (Park et al., 2018). *PHO2* is regulated by miR399, which is induced by P and repressed by N deficiency. In agreement, *PHO2* expression is induced by N-limitation and repressed under high nitrate conditions (Medici et al., 2019). This suggests a miRNA-mediated post-transcriptional and ubiquitin-mediated post-translational regulation of leaf senescence under N limitation.

The HRS1 TF was identified as an integrator of N and P signaling for root growth. *HRS1* and its closest homolog *HHO1* are rapidly and strongly induced by nitrate resupply of N-limited Arabidopsis roots. These TFs act downstream of the NRT1.1 transporter, NLP6 and NLP7 TFs. However, HRS1 and HHO1 repress primary root growth in response to P deficiency conditions only when nitrate is present, suggesting a complex regulation of N/P sensing (Medici et al., 2015). A recent study in Arabidopsis further confirmed the role of the HRS/HHO family on nitrate-P crosstalk. The expression of the *HRS1* homologs *HHO1*, *HHO2*, *HHO3* and *HRS1* itself is directly induced by PHR1, a master regulator of P starvation responses. Under P deficiency, PHR1 is released from the repressors SPX1/2/3/4 and promotes expression of *HRS1*-clade genes. Therefore,

nitrate uptake is repressed by P-limitation via the PHR1-HRS1-NRT2.1 pathway. Hence, regulation of nitrate uptake by the PHR1-HRS1 pathway may be a strategy to adapt to P deficiency (Maeda et al., 2018).

The control of the P starvation response (PSR) by nitrate availability is likely conserved across a wide range of plant species. In rice, high nitrate concentrations increase P uptake as well as the transcript levels of P transporter genes and P starvation-induced genes. This correlates with an increase in rice biomass under high P and high nitrate conditions. Nitrate induction of P starvation-induced genes is abolished in mutants of the *OsNRT1.1b* transporter, the rice orthologue of *Arabidopsis* NRT1.1, indicating that nitrate-activated P responses are dependent on *OsNRT1.1B* function. *OsNRT1.1B* mediates the ubiquitination and degradation of the P signaling repressor *OsSPX4* under high nitrate conditions, which allows the translocation of OsPHR2, the key TF in P signaling, to the nucleus to initiate transcription of P utilization genes (Zhou et al., 2008). Thus, nitrate-triggered degradation of *OsSPX4* activates both P- and nitrate-responsive genes, generating a coordinated utilization of N and P (Hu et al., 2019).

In a recent work, Medici et al. proposed that PSR is actively turned off when nitrate is limiting, and this repression is due to a combination of local and systemic nitrate signaling pathways. *PHO2* transcript levels are regulated by nitrate supply, accumulating during nitrate depletion and under high nitrate (Medici et al., 2019). Most PSR genes are no longer controlled by nitrate in a *pho2* mutant, demonstrating that PHO2 integrates nitrate signals into the PSR (Medici et al., 2019). Moreover, *NRT1.1* is repressed by P starvation and PHO2 acts as positive regulator of *NRT1.1*. Importantly, this mechanism of PSR control by nitrate availability is conserved in plants. Rice P starvation-induced genes *OsIPS1*, *OsSPX1*, and P transporter *OsPT1* were induced by P starvation only in the presence of nitrate and nitrate deprivation prevents the PSR in rice. This phenomenon was also conserved in wheat (Medici et al., 2019). These studies highlight the intricacy of the nitrate and P responses, with NRT1.1 having a central conserved role in modulating the interaction.

Nitrate and K Interaction

Although K itself is not metabolized, it is fundamental for and plays a vital role in many aspects of plant metabolism. K is a known cofactor for the activity of no less than 46 enzymes, including critical enzymes like NR. In plants, K and nitrate absorption and transport are

coordinated (Blevins et al., 1978; Triplett et al., 1980; White, 2012). In particular, the nitrate transporter *NRT1.5* functions as a proton-coupled H⁺/K antiporter. *NRT1.5* is responsible for nitrate loading from pericycle cells into the xylem. *NRT1.5* directly mediates K release from the root parenchyma into the xylem for transport to the shoot, especially under low-K conditions (Li et al., 2017). The proton gradient across the plasma membrane promotes this *NRT1.5*-mediated K xylem loading. This study revealed that *NRT1.5* plays a crucial role in K translocation from root to shoot and that it is also involved in the coordination of K/nitrate distribution in plants (Li et al., 2017). A complementary study showed that the MYB59 TF modulates K and nitrate translocation. MYB59 directly binds the *NRT1.5* promoter and activates its expression to coordinate root-to-shoot nutrient distribution. When plants face K/nitrate deficient conditions, *MYB59* and subsequently *NRT1.5* are downregulated to achieve the proper K/nitrate distribution between roots and shoots (Du et al., 2019).

Nitrate, P and K Interaction

Contrary to nitrate-P or nitrate-K interactions, little is known about how the nitrate-P-K interaction is regulated in *Arabidopsis* or in crop species. A study evaluating the impact of modifying the availability of these nutrients along with Sulfate (S) on the root system is one of the few recent reports evaluating how plants integrate four nutritional stimuli into complex developmental programs (Kellermeier et al., 2014). The authors quantified 13 parameters from *Arabidopsis* roots under 32 binary combinations of nitrate, P, K, S, and daylength. P caused the most important single-nutrient effect, while nitrate effects were strongly daylength-dependent. As a proof of principle, they explored mutants in candidate genes known to be affected by nitrate-K interactions such as *HAK5*, *NRT2.1*, *AKT1*, *CIPK23* and *NRT1.1*, revealing combinations of transporters, receptors, and kinases acting in the modulation of those signals (Kellermeier et al., 2014).

NITRATE IN THE CONTEXT OF A CHANGING CLIMATE

A critical resource affecting plant growth is water availability. As with nitrate, water is globally limited in soils, creating marginal soils that are agriculturally unproductive. In the current projections of climate change, much of the impact, particularly on crops, will be felt through changes in patterns of water availability. To engineer or breed crop varieties that can thrive in

marginal environments, it is crucial to understand how plants sense and integrate responses to nitrate and water, but surprisingly little is known about how plants might do this (Humbert et al., 2013). Since water serves as a solvent for nitrate, a recent study used a unique approach to investigate whether plants respond to nitrate as the absolute amount of N (N-moles) or the concentration of nitrate in water (N-molarity) (Swift et al., 2019). The approach used a factorial matrix that systematically varied both N-dose and water-dose applied to rice plants. The response of plants to N-amount, water-volume, and their interactions was modeled by linear regression of the induced effects on phenotypic traits and gene expression. The model revealed that rice plants could distinguish changes in N-moles, water-volume, N-molarity (N/water), as well as a synergistic response $N \times \text{water}$, both at the level of gene expression and phenotype. These results were tested in the field using a set of 19 rice varieties. Interestingly, the same set of genes responding to combinations of N- and water-dose in the lab correlated with crop outcomes in the field over two seasons (Swift et al., 2019).

Another important effect of climate change is the constant increase in CO₂ concentration. Atmospheric CO₂ levels are projected to reach 530–970 $\mu\text{mol}\cdot\text{mol}^{-1}$ by 2100, well above the pre-industrial average of $\sim 280 \mu\text{mol}\cdot\text{mol}^{-1}$ (Bloom et al., 2012). While elevated CO₂ may benefit plant growth by enhancing photosynthesis, studies have shown that it can also lower nitrate assimilation (Bloom et al., 2010; J. Bloom et al., 2014). A recent study showed that CO₂ causes a bottleneck effect on root-to-shoot nutrient transport, which results in lower shoot nitrate and, consequently, lower photosynthesis and biomass (Cohen et al., 2018). In a study in tomato, a combination of elevated CO₂ and higher temperatures diminished the nitrate uptake rate by roots and total protein concentration in roots, also indicating an inhibition of nitrate assimilation. These studies highlight the need for additional research in order to understand how climate change may impact both plant productivity and food quality (Jayawardena et al., 2017).

ADVANCES IN NUE

Although the benefits of using N fertilizers for higher yield are beyond doubt, they involve substantial economic and environmental costs. Modern agriculture has remarkably affected the global N cycle. An impressive example of this detrimental effect is the cascade of impacts that N-reactive forms have on the environment and on humans: One N-atom can, in sequence, increase atmospheric ozone (human health impact), increase fine particulate matter (visibility impact), alter

forest productivity and acidify surface waters (biodiversity loss impact), increase coastal ecosystem productivity, promoting coastal eutrophication, and increase the greenhouse effect via nitrous oxide production (environmental impact) (Galloway et al., 2004). The importance of advances in research and technology in agriculture, and particularly in the area of nitrogen use efficiency (NUE), has prompted experts to call for a second Green Revolution, focused on increasing productivity using sustainable agricultural methods (Gutierrez, 2012; McAllister et al., 2012).

In the last decade, several genes regulating NUE have been studied (Table 2). Strategies targeting genes directly involved in nitrate transport and metabolism have been widely used to enhance NUE in crops. For example, genetic variation in the nitrate transporter *NRT1.1B* (Hu et al., 2015) or in the nitrate reductase gene *NR2* (Gao et al., 2019) determines NUE divergence of *indica* and *japonica* rice varieties, while concurrent introgression of *OsNR2-indica* and *OsNRT1.1B-indica* enhances increased effective tiller number, grain yield, and NUE in the *japonica* subspecies. Other approaches based on Quantitative Trait Loci (QTL) analyses have provided important regulators of NUE, including transcriptional regulators, signaling proteins and components of hormonal pathways. For example, a major rice NUE QTL (identified in rice by genetic analysis of a recurrent backcross population; Sun et al., 2014) mapped to *DEP1*, a heterotrimeric G protein gene that significantly increases yield and has been widely studied and used in rice breeding (Xu et al., 2016a). Plants carrying the dominant *dep1-1* allele exhibit N-insensitive vegetative growth and increased N uptake and assimilation, resulting in improved harvest index and yield at moderate levels of N fertilization (Sun et al., 2014a). Another QTL study for NUE was performed in semi-dwarf rice green revolution varieties for increased yield by preventing lodging. These semi-dwarf varieties have an increased accumulation of DELLA repressor proteins, however these varieties are also associated with reduced NUE. This study revealed a QTL corresponding to the TF gene *GRF4*. GRF4 controls N uptake and assimilation by binding to the promoters of genes such as *AMT1.1* and *GSI.2*. Interestingly, SLR1, a DELLA protein, can inhibit the activation of N-related genes by GRF4, by competitively inhibiting the interaction of GRF4 with the GIF1 co-activator. Increased abundance of GRF4 partially disconnects GA-DELLA regulation of growth from N metabolism, promoting biomass increase at low N supplies (Li et al., 2018). QTL analysis also led to the identification of NUE genes in other plant species such as maize. Plants normally remobilize N from senescing leaves to sustain growth

elsewhere. A novel QTL controlling a functional stay-green phenotype (whereby senescence and N remobilization are impaired) was found to map to the *NAC7* maize gene. Transgenic maize lines where *NAC7* was down-regulated by RNAi showed delayed senescence and both increased biomass and N accumulation in vegetative tissues and increases on grain yield (Zhang et al., 2019). Besides QTL analysis, GWAS analysis has also provided candidate genes related to NUE in major crops, for example with the recent identification of an elite haplotype of the nitrate transporter *NPF6.1* named *NPF6.1HapB*. This allele enhanced nitrate uptake and conferred high NUE by increasing yield under low N supply. *NPF6.1HapB* was found to be transactivated by the NAC42 TF conferring enhanced NUE through activation of N uptake, increasing effective panicle number and yield (Tang et al., 2019).

As discussed in this review, many efforts have been devoted to defining the molecular components and regulatory networks involved in nitrate transport, sensing, signaling and responses. Integrative and predictive systems biology approaches are providing new key molecular actors that have been successfully tested in crops in order to assess their impact in plant NUE and yield. As an example, a supervised machine learning algorithm predicted a NUE gene network in *Arabidopsis* (Araus et al., 2016). *BT2*, a member of the *BRIC-A-BRAC/TRAMTRACK/BROAD* gene family, was identified as the most central and connected gene in the NUE network. *BT2* and its close homolog, *BT1*, act as negative regulators of NUE in a conserved manner in *Arabidopsis* and rice. A *bt1 bt2* double mutant showed an increased expression of high-affinity transporters *NRT2.1* and *NRT2.4*, with a concomitant increase in nitrate uptake and NUE under low nitrate conditions. Consistently, NUE decreased in plants overexpressing *BT2* as compared to wild-type plants under limiting nitrate conditions. Inactivating the *BT1/BT2* orthologue in rice increased NUE under low N conditions (Araus et al., 2016).

NUE is a complex trait since it is the result of different metabolic, physiological, developmental and environmental interactions integrated over the entire life cycle of the plant. However, most advances in understanding the molecular mechanisms controlling nitrate responses have been done in controlled laboratory conditions and in unique genotypes of model plants. In the future, it will therefore be key to make a concerted effort to understand nitrate uptake, nitrate responses and N-resource allocation in crops under more realistic field conditions (Plett et al., 2018). In particular, an important part of this challenge is to study NUE under environmental

conditions that are predicted by climate change models. How will be NUE impacted by increasing CO₂ levels? How will NUE be affected by simultaneous variations in multiple nutrients in soils? How will NUE be affected by water availability, rise in temperatures, extreme drought or flooded conditions? We urgently need to address these questions to face this new millennium.

SUPPLEMENTAL DATA

Supplemental Figure 1. The scheme behind a typical TARGET (Transient Assay Reporting Genome-wide Effects of Transcription factors) experiment.

Supplemental Data Set 1. Complete list of gene acronyms, their full names and their organisms of origin, mentioned in this review.

Supplemental Data Set 2. Extended list of key publications on the topics of nitrate uptake, transport, and signaling.

Supplemental Table 1. Complete list of genes belonging to the *NRT1* gene family.

Supplemental Table 2. Complete list of genes belonging to the *NRT2* gene family.

Supplemental Table 3. Complete list of genes featured in Figure 3.

ACKNOWLEDGEMENTS

Research in R.A.G.'s laboratory is funded by FONDECYT 1180759, ANID/FONDAP/15090007, Instituto Milenio iBio - Iniciativa Científica Milenio MINECON and EvoNet project DE-SC0014377. Research in E.A.V.'s laboratory is funded by FONDECYT 1170926, CONICYT PCI-Redes Internacionales entre Centros de Investigación REDES180097 and Instituto Milenio iBio - Iniciativa Científica Milenio MINECON. Research in J.M.A.'s laboratory is funded by Instituto Milenio iBio – Iniciativa Científica Milenio MINECON. Research in G.M.C.'s laboratory is funded by NSF PGRP: IOS 1840761, NSF-IOS 1339362 to G.M.C. G.K. and S.R., and Zegar Family Foundation (A16-0051). Collaboration between G.K., S.R. and G.M.C is supported by NSF-IOS 1339362 and by Centre National de la Recherche Scientifique (CNRS LIA-CoopNet to G.M.C., G.K. and S.R.).

AUTHOR CONTRIBUTIONS

E.A.V., J.M.A., V.A., G.M.C. and R.A.G. conceived the article. Authors contributed to writing the following sections: Introduction (N.M.C.); Molecular Mechanisms of Nitrate Transport (L.L.); Nitrate Signaling (E.R., S.R. and R.A.G.); Nitrate Interaction with Other Nutrients, Nitrate in the Context of a Changing Climate, Advances in NUE (V.A.); Regulatory Networks of Nitrate Response in Plants (J.M.A., M.D.B., G.K. and G.M.C.); Developmental Outputs (E.A.V). E.A.V., J.M.A., V.A., E.R., G.M.C. and R.A.G. drafted the final version of the review. E.A.V., J.M.A., V.A. and E.R. constructed Figures and Tables.

All authors critically revised and approved the final manuscript.

Table 1. Influential transcription factors in nitrate regulatory networks

| AGI Code | Name | TF family | # of edges in Network | # edges N Transport | # edges N Reduction | # edges N Assimilation | References |
|-----------|--------|----------------------|-----------------------|---------------------|---------------------|------------------------|---|
| At5g65210 | TGA1 | bZIP | 38 | 9 | 5 | 24 | Alvarez et al., (2014); Canales et al., (2017); Brooks et al., (2019) |
| At5g62430 | CDF1 | Dof-type zinc finger | 23 | 5 | 3 | 15 | Varala et al., (2018); Brooks et al., (2019) |
| At4g24020 | NLP7 | RWP-RK | 22 | 7 | 6 | 9 | Castaings et al., (2009); Marchive et al., (2013) |
| At2g22200 | ERF056 | ERF/AP2 | 21 | 1 | 3 | 17 | Brooks et al., (2019) |
| At3g49940 | LBD38 | ASL/LBD | 21 | 3 | 4 | 14 | Rubin et al., (2009); Brooks et al., (2019) |
| At1g66140 | ZFP4 | Zinc Finger | 20 | 5 | 1 | 14 | Brooks et al., (2019) |
| At3g16870 | GATA17 | GATA | 20 | 4 | 3 | 13 | Brooks et al., (2019) |
| At5g10970 | C2H2 | C2HC zinc fingers | 20 | 4 | 1 | 15 | Brooks et al., (2019) |
| At1g13260 | RAV1 | AP2/B3 | 19 | 5 | 2 | 12 | Brooks et al., (2019) |
| At5g15830 | bZIP3 | bZIP | 19 | 2 | 3 | 14 | Brooks et al., (2019) |
| At2g22430 | HB6 | Homeobox | 17 | 5 | 2 | 10 | Brooks et al., (2019) |
| At1g68670 | HHO2 | G2-like | 16 | 3 | 2 | 11 | Brooks et al., (2019) |
| At5g67420 | LBD37 | ASL/LBD | 16 | 6 | 3 | 7 | Rubin et al., (2009); Brooks et al., (2019) |
| At1g13300 | HRS1 | G2-like | 14 | 8 | 2 | 4 | Medici et al., (2015); Maeda et al., (2018) |
| At4g39780 | ERF060 | ERF/AP2 | 14 | 0 | 3 | 11 | Brooks et al., (2019) |

Table 2. Summary of genes regulating NUE in crop plants.

| Gene Name | Target plant | Physiological trait affected | Reference |
|----------------|-------------------|-------------------------------|----------------------------|
| <i>NAC2-5A</i> | wheat | root growth, yield | He et al., (2015) |
| <i>BZIP60</i> | wheat | LR branching, N uptake, yield | Yang et al., (2019) |
| <i>NAC42</i> | rice | plant height, NUE, yield | Tang et al., (2019) |
| <i>NAC7</i> | maize | senescence, biomass, yield | Zhang et al., (2019) |
| <i>AMT1.1</i> | rice | N uptake, yield | Ranathunge et al., (2014) |
| <i>NRT2.3b</i> | rice | growth, NUE, yield | Fan et al., (2016) |
| <i>NRT2.1</i> | rice | NUE, yield | Cheng et al., (2016) |
| <i>ARE1</i> | rice | senescence, NUE, yield | Wang et al., (2018) |
| <i>GRF4</i> | rice | biomass, yield | Li et al., (2018) |
| <i>NRT1.1B</i> | rice | NUE, yield | Hu et al., (2015) |
| <i>NFYA</i> | wheat | yield | Qu et al., (2015) |
| <i>PTR9</i> | rice | biomass, yield | Fang et al., (2013) |
| <i>BT1/BT2</i> | rice, Arabidopsis | yield, NUE | Araus et al., (2016) |
| <i>NAR2.1</i> | rice | NUE, yield | Cheng et al., (2017) |
| <i>GS1</i> | rice | NUE, yield | Baur et al., (2011) |
| <i>DOF1</i> | wheat, sorghum | biomass, yield | Pena et al., (2017) |
| <i>AAP1</i> | Pea | biomass, NUE, yield | Perchlik & Tegeder, (2017) |
| <i>DEP1</i> | Rice | biomass, yield | Sun et al., (2014) |
| <i>NR2</i> | Rice | yield, NUE | Gao et al., (2019) |

REFERENCES

- Albertos, P., Romero-Puertas, M.C., Tatematsu, K., Mateos, I., Sánchez-Vicente, I., Nambara, E., and Lorenzo, O.** (2015). S-nitrosylation triggers ABI5 degradation to promote seed germination and seedling growth. *Nature Communications* **6**, 8669.
- Alboresi, A., Gestin, C., Leydecker, M.T., Bedu, M., Meyer, C., and Truong, H.N.** (2005). Nitrate, a signal relieving seed dormancy in *Arabidopsis*. *Plant Cell Environ* **28**, 500-512.
- Ali-Rachedi, S., Bouinot, D., Wagner, M.H., Bonnet, M., Sotta, B., Grappin, P., and Jullien, M.** (2004). Changes in endogenous abscisic acid levels during dormancy release and maintenance of mature seeds: studies with the Cape Verde Islands ecotype, the dormant model of *Arabidopsis thaliana*. *Planta* **219**, 479-488.
- Alvarez, J.M., Riveras, E., Vidal, E.A., Gras, D.E., Contreras-López, O., Tamayo, K.P., Aceituno, F., Gómez, I., Ruffel, S., Lejay, L., Jordana, X., and Gutiérrez, R.A.** (2014). Systems approach identifies TGA1 and TGA4 transcription factors as important regulatory components of the nitrate response of *Arabidopsis thaliana* roots. *The Plant Journal* **80**, 1-13.
- Alvarez, J.M., Moyano, T.C., Zhang, T., Gras, D.E., Herrera, F.J., Araus, V., O'Brien, J.A., Carrillo, L., Medina, J., Vicente-Carbajosa, J., Jiang, J., and Gutiérrez, R.A.** (2019). Local changes in chromatin accessibility and transcriptional networks underlying the nitrate response in *Arabidopsis* roots. *Molecular Plant*.
- Araus, V., Vidal, E.A., Puelma, T., Alamos, S., Mieulet, D., Guiderdoni, E., and Gutiérrez, R.A.** (2016). Members of BTB Gene Family of Scaffold Proteins Suppress Nitrate Uptake and Nitrogen Use Efficiency. *Plant Physiology* **171**, 1523-1532.
- Araya, T., von Wiren, N., and Takahashi, H.** (2014a). CLE peptides regulate lateral root development in response to nitrogen nutritional status of plants. *Plant signaling & behavior* **9**, e29302.
- Araya, T., Miyamoto, M., Wibowo, J., Suzuki, A., Kojima, S., Tsuchiya, Y.N., Sawa, S., Fukuda, H., von Wiren, N., and Takahashi, H.** (2014b). CLE-CLAVATA1 peptide-receptor signaling module regulates the expansion of plant root systems in a nitrogen-dependent manner. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 2029-2034.
- Arc, E., Chibani, K., Grappin, P., Jullien, M., Godin, B., Cueff, G., Valot, B., Balliau, T., Job, D., and Rajjou, L.** (2012). Cold stratification and exogenous nitrates entail similar functional proteome adjustments during *Arabidopsis* seed dormancy release. *Journal of proteome research* **11**, 5418-5432.
- Bachmann, M., Huber, J.L., Athwal, G.S., Wu, K., Ferl, R.J., and Huber, S.C.** (1996). 14-3-3 proteins associate with the regulatory phosphorylation site of spinach leaf nitrate reductase in an isoform-specific manner and reduce dephosphorylation of Ser-543 by endogenous protein phosphatases. *FEBS letters* **398**, 26-30.
- Bargmann, B.O.R., Marshall-Colon, A., Efroni, I., Ruffel, S., Birnbaum, K.D., Coruzzi, G.M., and Krouk, G.** (2013). TARGET: a transient transformation system for genome-wide transcription factor target discovery. *Molecular plant* **6**, 978-980.
- Bethke, P.C., Libourel, I.G., Reinohl, V., and Jones, R.L.** (2006). Sodium nitroprusside, cyanide, nitrite, and nitrate break *Arabidopsis* seed dormancy in a nitric oxide-dependent manner. *Planta* **223**, 805-812.
- Blevins, D.G., Barnett, N.M., and Frost, W.B.** (1978). Role of Potassium and Malate in Nitrate Uptake and Translocation by Wheat Seedlings. *Plant Physiology* **62**, 784-788.

- Bloom, A.J., Burger, M., Asensio, J.S.R., and Cousins, A.B.** (2010). Carbon Dioxide Enrichment Inhibits Nitrate Assimilation in Wheat and Arabidopsis. *Science* **328**, 899-903.
- Bloom, A.J., Asensio, J.S., Randall, L., Rachmilevitch, S., Cousins, A.B., and Carlisle, E.A.** (2012). CO₂ enrichment inhibits shoot nitrate assimilation in C₃ but not C₄ plants and slows growth under nitrate in C₃ plants. *Ecology* **93**, 355-367.
- Bosemark, N.O.** (1954). The Influence of Nitrogen on Root Development. *Physiologia plantarum* **7**, 497-502.
- Bouguyon, E., Perrine-Walker, F., Pervent, M., Rochette, J., Cuesta, C., Benkova, E., Martinière, A., Bach, L., Krouk, G., Gojon, A., and Nacry, P.** (2016). Nitrate Controls Root Development through Posttranscriptional Regulation of the NRT1.1/NPF6.3 Transporter/Sensor. *Plant Physiology* **172**, 1237-1248.
- Bouguyon, E., Brun, F., Meynard, D., Kubeš, M., Pervent, M., Leran, S., Lacombe, B., Krouk, G., Guiderdoni, E., Zažímalová, E., Hoyerová, K., Nacry, P., and Gojon, A.** (2015). Multiple mechanisms of nitrate sensing by Arabidopsis nitrate transceptor NRT1.1. *Nature plants* **1**, 15015.
- Brooks, M.D., Cirrone, J., Pasquino, A.V., Alvarez, J.M., Swift, J., Mittal, S., Juang, C.-L., Varala, K., Gutiérrez, R.A., Krouk, G., Shasha, D., and Coruzzi, G.M.** (2019). Network Walking charts transcriptional dynamics of nitrogen signaling by integrating validated and predicted genome-wide interactions. *Nature Communications* **10**, 1569.
- Bussell, J.D., Keech, O., Fenske, R., and Smith, S.M.** (2013). Requirement for the plastidial oxidative pentose phosphate pathway for nitrate assimilation in Arabidopsis. *The Plant journal : for cell and molecular biology* **75**, 578-591.
- Cadman, C.S.C., Toorop, P.E., Hilhorst, H.W.M., and Finch-Savage, W.E.** (2006). Gene expression profiles of Arabidopsis Cvi seeds during dormancy cycling indicate a common underlying dormancy control mechanism. *The Plant Journal* **46**, 805-822.
- Calza, R., Huttner, E., Vincentz, M., Rouzé, P., Galangau, F., Vaucheret, H., Chérel, I., Meyer, C., Kronenberger, J., and Caboche, M.** (1987). Cloning of DNA fragments complementary to tobacco nitrate reductase mRNA and encoding epitopes common to the nitrate reductases from higher plants. *Molecular and General Genetics MGG* **209**, 552-562.
- Camargo, A., Llamas, A., Schnell, R.A., Higuera, J.J., González-Ballester, D., Lefebvre, P.A., Fernández, E., and Galván, A.** (2007). Nitrate signaling by the regulatory gene NIT2 in *Chlamydomonas*. *The Plant cell* **19**, 3491-3503.
- Canales, J., Moyano, T., Villarreal, E., and Gutiérrez, R.** (2014). Systems analysis of transcriptome data provides new hypotheses about Arabidopsis root response to nitrate treatments. *Frontiers in Plant Science* **5**.
- Canales, J., Contreras-López, O., Álvarez, J.M., and Gutiérrez, R.A.** (2017). Nitrate induction of root hair density is mediated by TGA1/TGA4 and CPC transcription factors in *Arabidopsis thaliana*. *The Plant Journal* **92**, 305-316.
- Capovilla, G., Schmid, M., and Pose, D.** (2015). Control of flowering by ambient temperature. *J Exp Bot* **66**, 59-69.
- Castaigns, L., Camargo, A., Pocholle, D., Gaudon, V., Texier, Y., Boutet-Mercey, S., Taconnat, L., Renou, J.P., Daniel-Vedele, F., and Fernandez, E.** (2009). The nodule inception-like protein 7 modulates nitrate sensing and metabolism in Arabidopsis. *The Plant Journal* **57**, 426-435.

- Castillo, M.C., Lozano-Juste, J., Gonzalez-Guzman, M., Rodriguez, L., Rodriguez, P.L., and Leon, J.** (2015). Inactivation of PYR/PYL/RCAR ABA receptors by tyrosine nitration may enable rapid inhibition of ABA signaling by nitric oxide in plants. *Science signaling* **8**, ra89.
- Castro Marín, I., Loef, I., Bartetzko, L., Searle, I., Coupland, G., Stitt, M., and Osuna, D.** (2011). Nitrate regulates floral induction in *Arabidopsis*, acting independently of light, gibberellin and autonomous pathways. *Planta* **233**, 539-552.
- Clarkson, D.T.** (1986). Regulation of the absorption and release of nitrate by plant cells: A review of current ideas and methodology. In *Fundamental, Ecological and Agricultural Aspects of Nitrogen Metabolism in Higher Plants: Proceedings of a symposium organized by the Department of Plant Physiology, University of Groningen and the Institute for Soil Fertility, Haren, 9–12 April 1985*, H. Lambers, J.J. Neeteson, and I. Stulen, eds (Dordrecht: Springer Netherlands), pp. 3-27.
- Cohen, I., Rapaport, T., Berger, R.T., and Rachmilevitch, S.** (2018). The effects of elevated CO₂ and nitrogen nutrition on root dynamics. *Plant Science* **272**, 294-300.
- Comparot, S., Lingiah, G., and Martin, T.** (2003). Function and specificity of 14-3-3 proteins in the regulation of carbohydrate and nitrogen metabolism. *J Exp Bot* **54**, 595-604.
- Crawford, N.M., and Glass, A.D.M.** (1998). Molecular and physiological aspects of nitrate uptake in plants. *Trends in Plant Science* **3**, 389-395.
- Crawford, N.M., and Forde, B.G.** (2002). Molecular and developmental biology of inorganic nitrogen nutrition. *The Arabidopsis Book/American Society of Plant Biologists* **1**.
- Crawford, N.M., Campbell, W.H., and Davis, R.W.** (1986). Nitrate reductase from squash: cDNA cloning and nitrate regulation. *Proceedings of the National Academy of Sciences* **83**, 8073-8076.
- Crawford, N.M., Smith, M., Bellissimo, D., and Davis, R.W.** (1988). Sequence and nitrate regulation of the *Arabidopsis thaliana* mRNA encoding nitrate reductase, a metalloflavoprotein with three functional domains. *Proceedings of the National Academy of Sciences* **85**, 5006-5010.
- Chakraborty, N., and Kanyuka, K.** (2019). GCR1 and GPA1 coupling regulates nitrate, cell wall, immunity and light responses in *Arabidopsis* **9**, 5838.
- Charoensawan, V., Martinho, C., and Wigge, P.A.** (2015). “Hit-and-run”: Transcription factors get caught in the act. *BioEssays : news and reviews in molecular, cellular and developmental biology* **37**, 748-754.
- Chen, X., Yao, Q., Gao, X., Jiang, C., Harberd, N.P., and Fu, X.** (2016). Shoot-to-Root Mobile Transcription Factor HY5 Coordinates Plant Carbon and Nitrogen Acquisition. *Current biology : CB* **26**, 640-646.
- Cheng, C.L., Dewdney, J., Kleinhofs, A., and Goodman, H.M.** (1986). Cloning and nitrate induction of nitrate reductase mRNA. *Proceedings of the National Academy of Sciences of the United States of America* **83**, 6825-6828.
- Chopin, F., Wirth, J., Dorbe, M.F., Lejay, L., Krapp, A., Gojon, A., and Daniel-Vedele, F.** (2007). The *Arabidopsis* nitrate transporter AtNRT2.1 is targeted to the root plasma membrane. *Plant physiology and biochemistry : PPB* **45**, 630-635.
- Davis, S.J.** (2009). Integrating hormones into the floral-transition pathway of *Arabidopsis thaliana*. *Plant, Cell & Environment* **32**, 1201-1210.

- De Angeli, A., Monachello, D., Ephritikhine, G., Frachisse, J.M., Thomine, S., Gambale, F., and Barbier-Brygoo, H.** (2006). The nitrate/proton antiporter AtCLCa mediates nitrate accumulation in plant vacuoles. *Nature* **442**, 939-942.
- de Jong, F., Thodey, K., Lejay, L.V., and Bevan, M.W.** (2014). Glucose elevates NITRATE TRANSPORTER2.1 protein levels and nitrate transport activity independently of its HEXOKINASE1-mediated stimulation of NITRATE TRANSPORTER2.1 expression. *Plant physiology* **164**, 308-320.
- Delhon, P., Gojon, A., Tillard, P., and Passama, L.** (1995). Diurnal regulation of NO₃⁻ uptake in soybean plants I. Changes in NO₃⁻ influx, efflux, and N utilization in the plant during the day/night cycle. *Journal of Experimental Botany* **46**, 1585-1594.
- Den Herder, G., Van Isterdael, G., Beeckman, T., and De Smet, I.** (2010). The roots of a new green revolution. *Trends in plant science* **15**, 600-607.
- Doidy, J., Li, Y., Neymotin, B., Edwards, M.B., Varala, K., Gresham, D., and Coruzzi, G.M.** (2016). Hit-and-Run" transcription: de novo transcription initiated by a transient bZIP1 "hit" persists after the "run. *BMC Genomics* **17**, 92.
- Du, X.-Q., Wang, F.-L., Li, H., Jing, S., Yu, M., Li, J., Wu, W.-H., Kudla, J., and Wang, Y.** (2019). The Transcription Factor MYB59 Regulates K⁺/NO₃⁻ Translocation in the Arabidopsis Response to Low K⁺ Stress. *The Plant cell* **31**, 699-714.
- Engelsberger, W.R., and Schulze, W.X.** (2012). Nitrate and ammonium lead to distinct global dynamic phosphorylation patterns when resupplied to nitrogen-starved Arabidopsis seedlings. *The Plant journal : for cell and molecular biology* **69**, 978-995.
- Evans, H.J., and Nason, A.** (1953). Pyridine Nucleotide-Nitrate Reductase from Extracts of Higher Plants. *Plant physiology* **28**, 233-254.
- Fan, X., Naz, M., Fan, X., Xuan, W., Miller, A.J., and Xu, G.** (2017). Plant nitrate transporters: from gene function to application. *Journal of Experimental Botany* **68**, 2463-2475.
- FAO.** (2017). World Fertiliser Trends and Outlook to 2020. Food and Agricultural Organisation of the United Nations.
- Filleur, S., and Daniel-Vedele, F.** (1999). Expression analysis of a high-affinity nitrate transporter isolated from *Arabidopsis thaliana* by differential display. *Planta* **207**, 461-469.
- Filleur, S., Dorbe, M.F., Cerezo, M., Orsel, M., Granier, F., Gojon, A., and Daniel-Vedele, F.** (2001). An arabidopsis T-DNA mutant affected in Nrt2 genes is impaired in nitrate uptake. *FEBS letters* **489**, 220-224.
- Finch-Savage, W.E., Cadman, C.S.C., Toorop, P.E., Lynn, J.R., and Hilhorst, H.W.M.** (2007). Seed dormancy release in Arabidopsis Cvi by dry after-ripening, low temperature, nitrate and light shows common quantitative patterns of gene expression directed by environmentally specific sensing. *The Plant Journal* **51**, 60-78.
- Finkelstein, R.R., and Lynch, T.J.** (2000). The Arabidopsis abscisic acid response gene ABI5 encodes a basic leucine zipper transcription factor. *The Plant cell* **12**, 599-609.
- Footitt, S., Douterelo-Soler, I., Clay, H., and Finch-Savage, W.E.** (2011). Dormancy cycling in Arabidopsis seeds is controlled by seasonally distinct hormone-signaling pathways. *Proceedings of the National Academy of Sciences of the United States of America* **108**, 20236-20241.
- Footitt, S., Huang, Z., Clay, H.A., Mead, A., and Finch-Savage, W.E.** (2013). Temperature, light and nitrate sensing coordinate Arabidopsis seed dormancy cycling, resulting in

- winter and summer annual phenotypes. *The Plant journal : for cell and molecular biology* **74**, 1003-1015.
- Forde, B., and Lorenzo, H.** (2001). The nutritional control of root development. *Plant and Soil* **232**, 51-68.
- Fredes, I., Moreno, S., Díaz, F.P., and Gutiérrez, R.A.** (2019). Nitrate signaling and the control of Arabidopsis growth and development. *Current opinion in plant biology* **47**, 112-118.
- Galloway, J.N., Dentener, F.J., Capone, D.G., Boyer, E.W., Howarth, R.W., Seitzinger, S.P., Asner, G.P., Cleveland, C.C., Green, P.A., Holland, E.A., Karl, D.M., Michaels, A.F., Porter, J.H., Townsend, A.R., and Vöosmarty, C.J.** (2004). Nitrogen Cycles: Past, Present, and Future. *Biogeochemistry* **70**, 153-226.
- Gan, Y., Bernreiter, A., Filleur, S., Abram, B., and Forde, B.G.** (2012). Overexpressing the ANR1 MADS-Box Gene in Transgenic Plants Provides New Insights into its Role in the Nitrate Regulation of Root Development. *Plant and Cell Physiology* **53**, 1003-1016.
- Gao, Z., Wang, Y., Chen, G., Zhang, A., Yang, S., Shang, L., Wang, D., Ruan, B., Liu, C., Jiang, H., Dong, G., Zhu, L., Hu, J., Zhang, G., Zeng, D., Guo, L., Xu, G., Teng, S., Harberd, N.P., and Qian, Q.** (2019). The indica nitrate reductase gene OsNR2 allele enhances rice yield potential and nitrogen use efficiency. *Nature Communications* **10**, 5207.
- Gaudinier, A., Rodriguez-Medina, J., Zhang, L., Olson, A., Liseron-Monfils, C., Bågman, A.-M., Foret, J., Abbitt, S., Tang, M., Li, B., Runcie, D.E., Kliebenstein, D.J., Shen, B., Frank, M.J., Ware, D., and Brady, S.M.** (2018). Transcriptional regulation of nitrogen-associated metabolism and growth. *Nature* **563**, 259-264.
- Geiger, D., Maierhofer, T., Al-Rasheid, K.A., Scherzer, S., Mumm, P., Liese, A., Ache, P., Wellmann, C., Marten, I., Grill, E., Romeis, T., and Hedrich, R.** (2011). Stomatal closure by fast abscisic acid signaling is mediated by the guard cell anion channel SLAH3 and the receptor RCAR1. *Science signaling* **4**, ra32.
- Gent, L., and Forde, B.G.** (2017). How do plants sense their nitrogen status? *J Exp Bot* **68**, 2531-2539.
- Gibbs, D.J., Md Isa, N., Movahedi, M., Lozano-Juste, J., Mendiando, G.M., Berckhan, S., Marín-de la Rosa, N., Vicente Conde, J., Sousa Correia, C., Pearce, S.P., Bassel, G.W., Hamali, B., Talloji, P., Tomé, D.F.A., Coego, A., Beynon, J., Alabadí, D., Bachmair, A., León, J., Gray, J.E., Theodoulou, F.L., and Holdsworth, M.J.** (2014). Nitric oxide sensing in plants is mediated by proteolytic control of group VII ERF transcription factors. *Mol Cell* **53**, 369-379.
- Gibon, Y., Usadel, B., Blaesing, O.E., Kamlage, B., Hoehne, M., Trethewey, R., and Stitt, M.** (2006). Integration of metabolite with transcript and enzyme activity profiling during diurnal cycles in Arabidopsis rosettes. *Genome biology* **7**, R76.
- Gifford, M.L., Dean, A., Gutierrez, R.A., Coruzzi, G.M., and Birnbaum, K.D.** (2008). Cell-specific nitrogen responses mediate developmental plasticity. *Proceedings of the National Academy of Sciences* **105**, 803-808.
- Gojon, A., Nacry, P., and Davidian, J.C.** (2009). Root uptake regulation: a central process for NPS homeostasis in plants. *Current opinion in plant biology* **12**, 328-338.
- Graciet, E., and Wellmer, F.** (2010). The plant N-end rule pathway: structure and functions. *Trends Plant Sci* **15**, 447-453.

- Gras, D.E., Vidal, E.A., Undurraga, S.F., Riveras, E., Moreno, S., Dominguez-Figueroa, J., Alabadi, D., Blazquez, M.A., Medina, J., and Gutierrez, R.A.** (2018). SMZ/SNZ and gibberellin signaling are required for nitrate-elicited delay of flowering time in *Arabidopsis thaliana*. *J Exp Bot* **69**, 619-631.
- Guan, P.** (2017). Dancing with Hormones: A Current Perspective of Nitrate Signaling and Regulation in *Arabidopsis*. *Frontiers in Plant Science* **8**.
- Guan, P., Ripoll, J.-J., Wang, R., Vuong, L., Bailey-Steinitz, L.J., Ye, D., and Crawford, N.M.** (2017). Interacting TCP and NLP transcription factors control plant responses to nitrate availability. *Proceedings of the National Academy of Sciences* **114**, 2419-2424.
- Guan, P., Wang, R., Nacry, P., Breton, G., Kay, S.A., Pruneda-Paz, J.L., Davani, A., and Crawford, N.M.** (2014). Nitrate foraging by *Arabidopsis* roots is mediated by the transcription factor TCP20 through the systemic signaling pathway. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 15267-15272.
- Guo, F.Q., Wang, R., Chen, M., and Crawford, N.M.** (2001). The *Arabidopsis* dual-affinity nitrate transporter gene AtNRT1.1 (CHL1) is activated and functions in nascent organ development during vegetative and reproductive growth. *The Plant cell* **13**, 1761-1777.
- Guo, Y., Chen, F., Zhang, F., and Mi, G.** (2005). Auxin transport from shoot to root is involved in the response of lateral root growth to localized supply of nitrate in maize. *Plant Science* **169**, 894-900.
- Gutierrez, R.A.** (2012). Systems biology for enhanced plant nitrogen nutrition. *Science* **336**, 1673-1675.
- Gutiérrez, R.A., Lejay, L.V., Dean, A., Chiaromonte, F., Shasha, D.E., and Coruzzi, G.M.** (2007). Qualitative network models and genome-wide expression data define carbon/nitrogen-responsive molecular machines in *Arabidopsis*. *Genome biology* **8**, R7.
- Gutiérrez, R.A., Stokes, T.L., Thum, K., Xu, X., Obertello, M., Katari, M.S., Tanurdzic, M., Dean, A., Nero, D.C., McClung, C.R., and Coruzzi, G.M.** (2008). Systems approach identifies an organic nitrogen-responsive gene network that is regulated by the master clock control gene CCA1. *Proceedings of the National Academy of Sciences of the United States of America* **105**, 4939-4944.
- He, H., Willems, L.A., Batushansky, A., Falt, A., Hanson, J., Nijveen, H., Hilhorst, H.W., and Bentsink, L.** (2016). Effects of Parental Temperature and Nitrate on Seed Performance are Reflected by Partly Overlapping Genetic and Metabolic Pathways. *Plant & cell physiology* **57**, 473-487.
- Ho, C.-H., Lin, S.-H., Hu, H.-C., and Tsay, Y.-F.** (2009). CHL1 Functions as a Nitrate Sensor in Plants. *Cell* **138**, 1184-1194.
- Hong, Y., Devaiah, S.P., Bahn, S.C., Thamasandra, B.N., Li, M., Welti, R., and Wang, X.** (2009). Phospholipase D epsilon and phosphatidic acid enhance *Arabidopsis* nitrogen signaling and growth. *Plant J.* **58**, 376-387.
- Hu, B., Wang, W., Ou, S., Tang, J., Li, H., Che, R., Zhang, Z., Chai, X., Wang, H., Wang, Y., Liang, C., Liu, L., Piao, Z., Deng, Q., Deng, K., Xu, C., Liang, Y., Zhang, L., Li, L., and Chu, C.** (2015). Variation in NRT1.1B contributes to nitrate-use divergence between rice subspecies. *Nature genetics* **47**, 834.
- Hu, B., Jiang, Z., Wang, W., Qiu, Y., Zhang, Z., Liu, Y., Li, A., Gao, X., Liu, L., Qian, Y., Huang, X., Yu, F., Kang, S., Wang, Y., Xie, J., Cao, S., Zhang, L., Wang, Y., Xie, Q., Kopriva, S., and Chu, C.** (2019). Nitrate–NRT1.1B–SPX4 cascade integrates nitrogen and phosphorus signalling networks in plants. *Nature plants* **5**, 401-413.

- Huang, N.C., Liu, K.H., Lo, H.J., and Tsay, Y.F.** (1999). Cloning and functional characterization of an Arabidopsis nitrate transporter gene that encodes a constitutive component of low-affinity uptake. *The Plant cell* **11**, 1381-1392.
- Huang, S., Chen, S., Liang, Z., Zhang, C., Yan, M., Chen, J., Xu, G., Fan, X., and Zhang, Y.** (2015). Knockdown of the partner protein OsNAR2.1 for high-affinity nitrate transport represses lateral root formation in a nitrate-dependent manner. *Scientific reports* **5**, 18192-18192.
- Humbert, S., Subedi, S., Cohn, J., Zeng, B., Bi, Y.-M., Chen, X., Zhu, T., McNicholas, P.D., and Rothstein, S.J.** (2013). Genome-wide expression profiling of maize in response to individual and combined water and nitrogen stresses. *BMC Genomics* **14**, 3-3.
- J. Bloom, A., Burger, M., A. Kimball, B., and J. Pinter, J.P.** (2014). Nitrate assimilation is inhibited by elevated CO₂ in field-grown wheat. *Nature Climate Change* **4**, 477.
- Jacquot, A., Chaput, V., Mauries, A., Li, Z., Tillard, P., Fizames, C., Bonillo, P., Bellegarde, F., Laugier, E., Santoni, V., Hem, S., Martin, A., Gojon, A., Schulze, W., and Lejay, L.** (2019). NRT2.1 phosphorylation prevents root high affinity nitrate uptake activity in *Arabidopsis thaliana*. *bioRxiv*, 583542.
- Jayawardena, D.M., Heckathorn, S.A., Bista, D.R., Mishra, S., Boldt, J.K., and Krause, C.R.** (2017). Elevated CO₂ plus chronic warming reduce nitrogen uptake and levels or activities of nitrogen-uptake and -assimilatory proteins in tomato roots. *Physiol Plant* **159**, 354-365.
- Jia, Z., Giehl, R.F.H., Meyer, R.C., Altmann, T., and von Wirén, N.** (2019). Natural variation of BSK3 tunes brassinosteroid signaling to regulate root foraging under low nitrogen. *Nature Communications* **10**, 2378.
- Kanno, Y., Hanada, A., Chiba, Y., Ichikawa, T., Nakazawa, M., Matsui, M., Koshiba, T., Kamiya, Y., and Seo, M.** (2012). Identification of an abscisic acid transporter by functional screening using the receptor complex as a sensor. *Proceedings of the National Academy of Sciences* **109**, 9653.
- Kant, S., Bi, Y.-M., and Rothstein, S.J.** (2010). Understanding plant response to nitrogen limitation for the improvement of crop nitrogen use efficiency. *Journal of Experimental Botany* **62**, 1499-1509.
- Kant, S., Peng, M., and Rothstein, S.J.** (2011). Genetic Regulation by NLA and MicroRNA827 for Maintaining Nitrate-Dependent Phosphate Homeostasis in *Arabidopsis*. *PLoS genetics* **7**, e1002021.
- Katari, M.S., Nowicki, S.D., Aceituno, F.F., Nero, D., Kelfer, J., Thompson, L.P., Cabello, J.M., Davidson, R.S., Goldberg, A.P., Shasha, D.E., Coruzzi, G.M., and Gutierrez, R.A.** (2010). VirtualPlant: a software platform to support systems biology research. *Plant Physiol* **152**, 500-515.
- Kellermeier, F., Armengaud, P., Seditas, T.J., Danku, J., Salt, D.E., and Amtmann, A.** (2014). Analysis of the Root System Architecture of *Arabidopsis* Provides a Quantitative Readout of Crosstalk between Nutritional Signals. *The Plant cell* **26**, 1480-1496.
- Khan, M.I.R., Trivellini, A., Fatma, M., Masood, A., Francini, A., Iqbal, N., Ferrante, A., and Khan, N.A.** (2015). Role of ethylene in responses of plants to nitrogen availability. *Frontiers in plant science* **6**, 927-927.
- Kiba, T., Kudo, T., Kojima, M., and Sakakibara, H.** (2010). Hormonal control of nitrogen acquisition: roles of auxin, abscisic acid, and cytokinin. *Journal of Experimental Botany* **62**, 1399-1409.

- Kiba, T., Feria-Bourrellier, A.-B., Lafouge, F., Lezhneva, L., Boutet-Mercey, S., Orsel, M., Bréhaut, V., Miller, A., Daniel-Vedele, F., Sakakibara, H., and Krapp, A.** (2012). The Arabidopsis nitrate transporter NRT2.4 plays a double role in roots and shoots of nitrogen-starved plants. *The Plant cell* **24**, 245-258.
- Kiba, T., Inaba, J., Kudo, T., Ueda, N., Konishi, M., Mitsuda, N., Takiguchi, Y., Kondou, Y., Yoshizumi, T., Ohme-Takagi, M., Matsui, M., Yano, K., Yanagisawa, S., and Sakakibara, H.** (2018). Repression of Nitrogen Starvation Responses by Members of the Arabidopsis GARP-Type Transcription Factor NIGT1/HRS1 Subfamily. *The Plant cell* **30**, 925-945.
- Kim, D.H., Doyle, M.R., Sung, S., and Amasino, R.M.** (2009). Vernalization: winter and the timing of flowering in plants. *Annual review of cell and developmental biology* **25**, 277-299.
- Ko, D., Kang, J., Kiba, T., Park, J., Kojima, M., Do, J., Kim, K.Y., Kwon, M., Endler, A., Song, W.Y., Martinoia, E., Sakakibara, H., and Lee, Y.** (2014). Arabidopsis ABCG14 is essential for the root-to-shoot translocation of cytokinin. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 7150-7155.
- Konishi, M., and Yanagisawa, S.** (2013). Arabidopsis NIN-like transcription factors have a central role in nitrate signalling. *Nat Commun* **4**, 1617.
- Konishi, M., and Yanagisawa, S.** (2019). The role of protein-protein interactions mediated by the PB1 domain of NLP transcription factors in nitrate-inducible gene expression. *BMC Plant Biology* **19**, 90.
- Kotur, Z., Mackenzie, N., Ramesh, S., Tyerman, S.D., Kaiser, B.N., and Glass, A.D.** (2012). Nitrate transport capacity of the Arabidopsis thaliana NRT2 family members and their interactions with AtNAR2.1. *The New phytologist* **194**, 724-731.
- Krapp, A., David, L.C., Chardin, C., Girin, T., Marmagne, A., Leprince, A.-S., Chaillou, S., Ferrario-Méry, S., Meyer, C., and Daniel-Vedele, F.** (2014). Nitrate transport and signalling in Arabidopsis. *Journal of Experimental Botany* **65**, 789-798.
- Kriel, J., Haesendonckx, S., Rubio-Teixeira, M., Van Zeebroeck, G., and Thevelein, J.M.** (2011). From transporter to transceptor: signaling from transporters provokes re-evaluation of complex trafficking and regulatory controls: endocytic internalization and intracellular trafficking of nutrient transceptors may, at least in part, be governed by their signaling function. *BioEssays : news and reviews in molecular, cellular and developmental biology* **33**, 870-879.
- Krouk, G., Tillard, P., and Gojon, A.** (2006). Regulation of the high-affinity NO₃⁻ uptake system by NRT1.1-mediated NO₃⁻ demand signaling in Arabidopsis. *Plant Physiol* **142**, 1075-1086.
- Krouk, G., Mirowski, P., LeCun, Y., Shasha, D.E., and Coruzzi, G.M.** (2010a). Predictive network modeling of the high-resolution dynamic plant transcriptome in response to nitrate. *Genome biology* **11**, R123.
- Krouk, G., Lingeman, J., Colon, A.M., Coruzzi, G., and Shasha, D.** (2013). Gene regulatory networks in plants: learning causality from time and perturbation. *Genome Biology* **14**, 123.
- Krouk, G., Lacombe, B., Bielach, A., Perrine-Walker, F., Malinska, K., Mounier, E., Hoyerova, K., Tillard, P., Leon, S., Ljung, K., Zazimalova, E., Benkova, E., Nacry, P., and Gojon, A.** (2010b). Nitrate-Regulated Auxin Transport by NRT1.1 Defines a Mechanism for Nutrient Sensing in Plants. *Dev Cell* **18**, 927-937.

- Landrein, B., Formosa-Jordan, P., Malivert, A., Schuster, C., Melnyk, C.W., Yang, W., Turnbull, C., Meyerowitz, E.M., Locke, J.C.W., and Jönsson, H.** (2018). Nitrate modulates stem cell dynamics in Arabidopsis shoot meristems through cytokinins. *Proceedings of the National Academy of Sciences* **115**, 1382-1387.
- Langfelder, P., and Horvath, S.** (2008). WGCNA: an R package for weighted correlation network analysis. *BMC bioinformatics* **9**, 559.
- Lejay, L., Wirth, J., Pervent, M., Cross, J.M., Tillard, P., and Gojon, A.** (2008). Oxidative pentose phosphate pathway-dependent sugar sensing as a mechanism for regulation of root ion transporters by photosynthesis. *Plant Physiol* **146**, 2036-2053.
- Lejay, L., Tillard, P., Lepetit, M., Olive, F., Filleur, S., Daniel-Vedele, F., and Gojon, A.** (1999). Molecular and functional regulation of two NO₃- uptake systems by N- and C-status of Arabidopsis plants. *The Plant journal : for cell and molecular biology* **18**, 509-519.
- Lejay, L., Gansel, X., Cerezo, M., Tillard, P., Muller, C., Krapp, A., von Wiren, N., Daniel-Vedele, F., and Gojon, A.** (2003). Regulation of root ion transporters by photosynthesis: functional importance and relation with hexokinase. *The Plant cell* **15**, 2218-2232.
- Leran, S., Varala, K., Boyer, J.C., Chiurazzi, M., Crawford, N., Daniel-Vedele, F., David, L., Dickstein, R., Fernandez, E., Forde, B., Gassmann, W., Geiger, D., Gojon, A., Gong, J.M., Halkier, B.A., Harris, J.M., Hedrich, R., Limami, A.M., Rentsch, D., Seo, M., Tsay, Y.F., Zhang, M., Coruzzi, G., and Lacombe, B.** (2014). A unified nomenclature of NITRATE TRANSPORTER 1/PEPTIDE TRANSPORTER family members in plants. *Trends Plant Sci* **19**, 5-9.
- Léran, S., Edel, K.H., Pervent, M., Hashimoto, K., Corratgé-Faillie, C., Offenborn, J.N., Tillard, P., Gojon, A., Kudla, J., and Lacombe, B.** (2015). Nitrate sensing and uptake in Arabidopsis are enhanced by ABI2, a phosphatase inactivated by the stress hormone abscisic acid. *Science signaling* **8**, ra43-ra43.
- Lezhneva, L., Kiba, T., Feria-Bourrellier, A.B., Lafouge, F., Boutet-Mercey, S., Zoufan, P., Sakakibara, H., Daniel-Vedele, F., and Krapp, A.** (2014). The Arabidopsis nitrate transporter NRT2.5 plays a role in nitrate acquisition and remobilization in nitrogen-starved plants. *The Plant journal : for cell and molecular biology* **80**, 230-241.
- Li, H., Yu, M., Du, X.-Q., Wang, Z.-F., Wu, W.-H., Quintero, F.J., Jin, X.-H., Li, H.-D., and Wang, Y.** (2017). NRT1.5/NPF7.3 Functions as a Proton-Coupled H(+)/K(+) Antiporter for K(+) Loading into the Xylem in Arabidopsis. *The Plant cell* **29**, 2016-2026.
- Li, S., Tian, Y., Wu, K., Ye, Y., Yu, J., Zhang, J., Liu, Q., Hu, M., Li, H., Tong, Y., Harberd, N.P., and Fu, X.** (2018). Modulating plant growth-metabolism coordination for sustainable agriculture. *Nature* **560**, 595-600.
- Li, Y., Krouk, G., Coruzzi, G.M., and Ruffel, S.** (2014). Finding a nitrogen niche: a systems integration of local and systemic nitrogen signalling in plants. *Journal of experimental botany* **65**, 5601-5610.
- Liang, Y., Zhao, X., Jones, A.M., and Gao, Y.** (2018). G proteins sculp root architecture in response to nitrogen in rice and Arabidopsis. *Plant science : an international journal of experimental plant biology* **274**, 129-136.
- Lin, Y.-L., and Tsay, Y.-F.** (2017). Influence of differing nitrate and nitrogen availability on flowering control in Arabidopsis. *Journal of Experimental Botany* **68**, 2603-2609.
- Little, D.Y., Rao, H., Oliva, S., Daniel-Vedele, F., Krapp, A., and Malamy, J.E.** (2005). The putative high-affinity nitrate transporter NRT2.1 represses lateral root initiation in

- response to nutritional cues. *Proceedings of the National Academy of Sciences of the United States of America* **102**, 13693-13698.
- Liu, J., Mehdi, S., Topping, J., Tarkowski, P., and Lindsey, K.** (2010). Modelling and experimental analysis of hormonal crosstalk in Arabidopsis. *Mol Syst Biol* **6**, 373-373.
- Liu, K.-H., Huang, C.-Y., and Tsay, Y.-F.** (1999). CHL1 Is a Dual-Affinity Nitrate Transporter of Arabidopsis Involved in Multiple Phases of Nitrate Uptake. *The Plant cell* **11**, 865-874.
- Liu, K.-h., Niu, Y., Konishi, M., Wu, Y., Du, H., Sun Chung, H., Li, L., Boudsocq, M., McCormack, M., Maekawa, S., Ishida, T., Zhang, C., Shokat, K., Yanagisawa, S., and Sheen, J.** (2017). Discovery of nitrate–CPK–NLP signalling in central nutrient–growth networks. *Nature* **545**, 311.
- Liu, K.H., and Tsay, Y.F.** (2003). Switching between the two action modes of the dual-affinity nitrate transporter CHL1 by phosphorylation. *The EMBO journal* **22**, 1005-1013.
- Liu, T., Li, Y., Ren, J., Qian, Y., Yang, X., Duan, W., and Hou, X.** (2013). Nitrate or NaCl regulates floral induction in Arabidopsis thaliana. *Biologia* **68**, 215-222.
- Liu, Y., Shi, L., Ye, N., Liu, R., Jia, W., and Zhang, J.** (2009). Nitric oxide-induced rapid decrease of abscisic acid concentration is required in breaking seed dormancy in Arabidopsis. *The New phytologist* **183**, 1030-1042.
- Lopez-Molina, L., Mongrand, S., and Chua, N.H.** (2001). A postgermination developmental arrest checkpoint is mediated by abscisic acid and requires the ABI5 transcription factor in Arabidopsis. *Proceedings of the National Academy of Sciences of the United States of America* **98**, 4782-4787.
- Ma, W., Li, J., Qu, B., He, X., Zhao, X., Li, B., Fu, X., and Tong, Y.** (2014). Auxin biosynthetic gene TAR2 is involved in low nitrogen-mediated reprogramming of root architecture in Arabidopsis. *The Plant journal : for cell and molecular biology* **78**, 70-79.
- Maeda, Y., Konishi, M., Kiba, T., Sakuraba, Y., Sawaki, N., Kurai, T., Ueda, Y., Sakakibara, H., and Yanagisawa, S.** (2018). A NIGT1-centred transcriptional cascade regulates nitrate signalling and incorporates phosphorus starvation signals in Arabidopsis. *Nature Communications* **9**, 1376.
- Maierhofer, T., Lind, C., Hüttl, S., Scherzer, S., Papenfuß, M., Simon, J., Al-Rasheid, K.A.S., Ache, P., Rennenberg, H., Hedrich, R., Müller, T.D., and Geiger, D.** (2014). A Single-Pore Residue Renders the Arabidopsis Root Anion Channel SLAH2 Highly Nitrate Selective. *The Plant cell* **26**, 2554-2567.
- Marchive, C., Roudier, F., Castaings, L., Bréhaut, V., Blondet, E., Colot, V., Meyer, C., and Krapp, A.** (2013). Nuclear retention of the transcription factor NLP7 orchestrates the early response to nitrate in plants. *Nature Communications* **4**, 1713.
- Martin, T., Oswald, O., and Graham, I.A.** (2002). Arabidopsis seedling growth, storage lipid mobilization, and photosynthetic gene expression are regulated by carbon:nitrogen availability. *Plant Physiol* **128**, 472-481.
- Matakiadis, T., Alboresi, A., Jikumaru, Y., Tatematsu, K., Pichon, O., Renou, J.P., Kamiya, Y., Nambara, E., and Truong, H.N.** (2009). The Arabidopsis abscisic acid catabolic gene CYP707A2 plays a key role in nitrate control of seed dormancy. *Plant Physiol* **149**, 949-960.
- McAllister, C.H., Beatty, P.H., and Good, A.G.** (2012). Engineering nitrogen use efficient crop plants: the current status. *Plant biotechnology journal* **10**, 1011-1025.

- Medici, A., Marshall-Colon, A., Ronzier, E., Szponarski, W., Wang, R., Gojon, A., Crawford, N.M., Ruffel, S., Coruzzi, G.M., and Krouk, G.** (2015). AtNIGT1/HRS1 integrates nitrate and phosphate signals at the Arabidopsis root tip. *Nature Communications* **6**, 6274.
- Medici, A., Szponarski, w., Dangeville, P., Safi, A., Dissanayake, I.M., Saenchai, C., Emanuel, A., Rubio, V., Lacombe, B., Ruffel, S., Tanurdzic, M., Rouached, H., and Krouk, G.** (2019). Identification of molecular integrators shows that nitrogen actively controls the phosphate starvation response in plants. *The Plant cell*, tpc.00656.02018.
- Menz, J., Li, Z., Schulze, W.X., and Ludewig, U.** (2016). Early nitrogen-deprivation responses in Arabidopsis roots reveal distinct differences on transcriptome and (phospho-) proteome levels between nitrate and ammonium nutrition. *The Plant Journal* **88**, 717-734.
- Miller, A.J., Fan, X., Shen, Q., and Smith, S.J.** (2008). Amino acids and nitrate as signals for the regulation of nitrogen acquisition. *Journal of Experimental Botany* **59**, 111-119.
- Mirowski, P., and LeCun, Y.** (2009). *Dynamic Factor Graphs for Time Series Modeling* (Berlin, Heidelberg: Springer Berlin Heidelberg), pp. 128-143.
- Muller, B., and Touraine, B.** (1992). Inhibition of NO₃⁻ Uptake by Various Phloem-Translocated Amino Acids in Soybean Seedlings. *Journal of Experimental Botany* **43**, 617-623.
- Müller, D., Waldie, T., Miyawaki, K., To, J.P.C., Melnyk, C.W., Kieber, J.J., Kakimoto, T., and Leyser, O.** (2015). Cytokinin is required for escape but not release from auxin mediated apical dominance. *The Plant Journal* **82**, 874-886.
- Muños, S., Cazettes, C., Fizames, C., Gaymard, F., Tillard, P., Lepetit, M., Lejay, L., and Gojon, A.** (2004). Transcript profiling in the chl1-5 mutant of Arabidopsis reveals a role of the nitrate transporter NRT1.1 in the regulation of another nitrate transporter, NRT2.1. *The Plant cell* **16**, 2433-2447.
- Naulin, P.A., Armijo, G.I., Vega, A.S., Tamayo, K.P., Gras, D.E., de la Cruz, J., and Gutiérrez, R.A.** (2019). Nitrate Induction of Primary Root Growth Requires Cytokinin Signaling in Arabidopsis thaliana. *Plant and Cell Physiology*.
- Negi, J., Matsuda, O., Nagasawa, T., Oba, Y., Takahashi, H., Kawai-Yamada, M., Uchimiya, H., Hashimoto, M., and Iba, K.** (2008). CO₂ regulator SLAC1 and its homologues are essential for anion homeostasis in plant cells. *Nature* **452**, 483-486.
- Nohn, T., Noji, S., Taniguchi, S., and Saito, T.** (1989). The narX and narL genes encoding the nitrate-sensing regulators of Escherichia coli are homologous to a family of prokaryotic two-component regulatory genes. *Nucleic Acids Res* **17**, 2947-2957.
- O'Malley, R.C., Huang, S.-S.C., Song, L., Lewsey, M.G., Bartlett, A., Nery, J.R., Galli, M., Gallavotti, A., and Ecker, J.R.** (2016). Cistrome and Epicistrome Features Shape the Regulatory DNA Landscape. *Cell* **165**, 1280-1292.
- Ohkubo, Y., Tanaka, M., Tabata, R., Ogawa-Ohnishi, M., and Matsubayashi, Y.** (2017). Shoot-to-root mobile polypeptides involved in systemic regulation of nitrogen acquisition **3**, 17029.
- Okamoto, M., Vidmar, J.J., and Glass, A.D.M.** (2003). Regulation of NRT1 and NRT2 Gene Families of *Arabidopsis thaliana*: Responses to Nitrate Provision. *Plant and Cell Physiology* **44**, 304-317.
- Okamoto, M., Kumar, A., Li, W., Wang, Y., Siddiqi, M.Y., Crawford, N.M., and Glass, A.D.** (2006a). High-affinity nitrate transport in roots of Arabidopsis depends on expression of the NAR2-like gene AtNRT3.1. *Plant Physiol* **140**, 1036-1046.

- Okamoto, M., Kuwahara, A., Seo, M., Kushiro, T., Asami, T., Hirai, N., Kamiya, Y., Koshiba, T., and Nambara, E.** (2006b). CYP707A1 and CYP707A2, which encode abscisic acid 8'-hydroxylases, are indispensable for proper control of seed dormancy and germination in *Arabidopsis*. *Plant Physiol* **141**, 97-107.
- Olas, J.J., Van Dingenen, J., Abel, C., Dzialo, M.A., Feil, R., Krapp, A., Schlereth, A., and Wahl, V.** (2019). Nitrate acts at the *Arabidopsis thaliana* shoot apical meristem to regulate flowering time. *The New phytologist* **223**, 814-827.
- Ondzighi-Assoume, C.A., Chakraborty, S., and Harris, J.M.** (2016). Environmental Nitrate Stimulates Absciscic Acid Accumulation in *Arabidopsis* Root Tips by Releasing It from Inactive Stores. *The Plant cell* **28**, 729-745.
- Orsel, M., Krapp, A., and Daniel-Vedele, F.** (2002). Analysis of the NRT2 nitrate transporter family in *Arabidopsis*. Structure and gene expression. *Plant physiology* **129**, 886-896.
- Orsel, M., Chopin, F., Leleu, O., Smith, S.J., Krapp, A., Daniel-Vedele, F., and Miller, A.J.** (2006). Characterization of a two-component high-affinity nitrate uptake system in *Arabidopsis*. Physiology and protein-protein interaction. *Plant Physiol* **142**, 1304-1317.
- Osugi, A., Kojima, M., Takebayashi, Y., Ueda, N., Kiba, T., and Sakakibara, H.** (2017). Systemic transport of trans-zeatin and its precursor have differing roles in *Arabidopsis* shoots **3**, 17112.
- Para, A., Li, Y., Marshall-Colón, A., Varala, K., Francoeur, N.J., Moran, T.M., Edwards, M.B., Hackley, C., Bargmann, B.O., and Birnbaum, K.D.** (2014). Hit-and-run transcriptional control by bZIP1 mediates rapid nutrient signaling in *Arabidopsis*. *Proceedings of the National Academy of Sciences*, 201404657.
- Park, B.S., Yao, T., Seo, J.S., Wong, E.C.C., Mitsuda, N., Huang, C.H., and Chua, N.H.** (2018). *Arabidopsis* NITROGEN LIMITATION ADAPTATION regulates ORE1 homeostasis during senescence induced by nitrogen deficiency. *Nature plants* **4**, 898-903.
- Parker, J.L., and Newstead, S.** (2014). Molecular basis of nitrate uptake by the plant nitrate transporter NRT1.1. *Nature* **507**, 68-72.
- Pellizzaro, A., Clochard, T., Cukier, C., Bourdin, C., Juchaux, M., Montrichard, F., Thany, S., Raymond, V., Planchet, E., Limami, A.M., and Morere-Le Paven, M.C.** (2014). The nitrate transporter MtNPF6.8 (MtNRT1.3) transports abscisic acid and mediates nitrate regulation of primary root growth in *Medicago truncatula*. *Plant Physiol* **166**, 2152-2165.
- Peng, M., Bi, Y.M., Zhu, T., and Rothstein, S.J.** (2007). Genome-wide analysis of *Arabidopsis* responsive transcriptome to nitrogen limitation and its regulation by the ubiquitin ligase gene NLA. *Plant molecular biology* **65**, 775-797.
- Peng, M., Hudson, D., Schofield, A., Tsao, R., Yang, R., Gu, H., Bi, Y.-M., and Rothstein, S.J.** (2008). Adaptation of *Arabidopsis* to nitrogen limitation involves induction of anthocyanin synthesis which is controlled by the NLA gene. *Journal of experimental botany* **59**, 2933-2944.
- Perez-Torres, C.A., Lopez-Bucio, J., Cruz-Ramirez, A., Ibarra-Laclette, E., Dharmasiri, S., Estelle, M., and Herrera-Estrella, L.** (2008). Phosphate availability alters lateral root development in *Arabidopsis* by modulating auxin sensitivity via a mechanism involving the TIR1 auxin receptor. *The Plant cell* **20**, 3258-3272.
- Plett, D.C., Holtham, L.R., Okamoto, M., and Garnett, T.P.** (2018). Nitrate uptake and its regulation in relation to improving nitrogen use efficiency in cereals. *Seminars in Cell & Developmental Biology* **74**, 97-104.

- Poitout, A., Crabos, A., Petřík, I., Novák, O., Krouk, G., Lacombe, B., and Ruffel, S.** (2018). Responses to Systemic Nitrogen Signaling in Arabidopsis Roots Involve trans-Zeatin in Shoots. *The Plant cell* **30**, 1243-1257.
- Rahayu, Y.S., Walch-Liu, P., Neumann, G., Römheld, V., von Wirén, N., and Bangerth, F.** (2005). Root-derived cytokinins as long-distance signals for NO₃--induced stimulation of leaf growth. *Journal of Experimental Botany* **56**, 1143-1152.
- Remans, T., Nacry, P., Pervent, M., Filleur, S., Diatloff, E., Mounier, E., Tillard, P., Forde, B.G., and Gojon, A.** (2006). The Arabidopsis NRT1.1 transporter participates in the signaling pathway triggering root colonization of nitrate-rich patches. *Proceedings of the National Academy of Sciences* **103**, 19206-19211.
- Ren, H., Willige, B.C., Jaillais, Y., Geng, S., Park, M.Y., Gray, W.M., and Chory, J.** (2019). BRASSINOSTEROID-SIGNALING KINASE 3, a plasma membrane-associated scaffold protein involved in early brassinosteroid signaling. *PLoS genetics* **15**, e1007904.
- Ristova, D., Carré, C., Pervent, M., Medici, A., Kim, G.J., Scalia, D., Ruffel, S., Birnbaum, K.D., Lacombe, B., and Busch, W.** (2016). Combinatorial interaction network of transcriptomic and phenotypic responses to nitrogen and hormones in the *Arabidopsis thaliana* root. *Sci. Signal.* **9**, rs13-rs13.
- Riveras, E., Alvarez, J.M., Vidal, E.A., Osés, C., Vega, A., and Gutiérrez, R.A.** (2015). The Calcium Ion Is a Second Messenger in the Nitrate Signaling Pathway of Arabidopsis. *Plant Physiology* **169**, 1397-1404.
- Rodgers, C.O., and Barneix, A.J.** (1988). Cultivar differences in the rate of nitrate uptake by intact wheat plants as related to growth rate. *Physiologia plantarum* **72**, 121-126.
- Rogers, E.D., and Benfey, P.N.** (2015). Regulation of plant root system architecture: implications for crop advancement. *Current Opinion in Biotechnology* **32**, 93-98.
- Romera, F.J., Smith, A.P., and Pérez-Vicente, R.** (2016). Editorial: Ethylene's Role in Plant Mineral Nutrition. *Frontiers in plant science* **7**, 911-911.
- Rubin, G., Tohge, T., Matsuda, F., Saito, K., and Scheible, W.R.** (2009). Members of the LBD family of transcription factors repress anthocyanin synthesis and affect additional nitrogen responses in Arabidopsis. *The Plant cell* **21**, 3567-3584.
- Ruffel, S., and Gojon, A.** (2017). Systemic nutrient signalling: On the road for nitrate. *Nature plants* **3**, 17040.
- Ruffel, S., Poitout, A., Krouk, G., Coruzzi, G.M., and Lacombe, B.** (2016). Long-distance nitrate signaling displays cytokinin dependent and independent branches. *Journal of integrative plant biology* **58**, 226-229.
- Ruffel, S., Krouk, G., Ristova, D., Shasha, D., Birnbaum, K.D., and Coruzzi, G.M.** (2011). Nitrogen economics of root foraging: transitive closure of the nitrate-cytokinin relay and distinct systemic signaling for N supply vs. demand. *Proceedings of the National Academy of Sciences of the United States of America* **108**, 18524-18529.
- Safi, A., Medici, A., Szponarski, W., Marshall-Colon, A., Ruffel, S., Gaymard, F., Coruzzi, G., Lacombe, B., and Krouk, G.** (2018). HRS1/HHOs GARP transcription factors and reactive oxygen species are regulators of Arabidopsis nitrogen starvation response. *bioRxiv*, 164277.
- Sakakibara, H.** (2006). CYTOKININS: Activity, Biosynthesis, and Translocation. *Annual review of plant biology* **57**, 431-449.
- Sakakibara, H., Takei, K., and Hirose, N.** (2006). Interactions between nitrogen and cytokinin in the regulation of metabolism and development. *Trends in Plant Science* **11**, 440-448.

- Sakakibara, H., Kobayashi, K., Deji, A., and Sugiyama, T.** (1997). Partial Characterization of the Signaling Pathway for the Nitrate-Dependent Expression of Genes for Nitrogen-Assimilatory Enzymes Using Detached Maize Leaves. *Plant cell physiology* **38**, 837-843.
- Sasaki, T., Suzaki, T., Soyano, T., Kojima, M., Sakakibara, H., and Kawaguchi, M.** (2014). Shoot-derived cytokinins systemically regulate root nodulation. *Nat Commun* **5**, 4983.
- Sato, T., Maekawa, S., Yasuda, S., Domeki, Y., Sueyoshi, K., Fujiwara, M., Fukao, Y., Goto, D.B., and Yamaguchi, J.** (2011). Identification of 14-3-3 proteins as a target of ATL31 ubiquitin ligase, a regulator of the C/N response in Arabidopsis. *The Plant Journal* **68**, 137-146.
- Sato, T., Maekawa, S., Yasuda, S., Sonoda, Y., Katoh, E., Ichikawa, T., Nakazawa, M., Seki, M., Shinozaki, K., Matsui, M., Goto, D.B., Ikeda, A., and Yamaguchi, J.** (2009). CNI1/ATL31, a RING-type ubiquitin ligase that functions in the carbon/nitrogen response for growth phase transition in Arabidopsis seedlings. *The Plant journal : for cell and molecular biology* **60**, 852-864.
- Schauser, L., Wieloch, W., and Stougaard, J.** (2005). Evolution of NIN-like proteins in Arabidopsis, rice, and Lotus japonicus. *Journal of molecular evolution* **60**, 229-237.
- Schauser, L., Roussis, A., Stiller, J., and Stougaard, J.** (1999). A plant regulator controlling development of symbiotic root nodules. *Nature* **402**, 191-195.
- Scheible, W.R., Gonzalez-Fontes, A., Lauerer, M., Muller-Rober, B., Caboche, M., and Stitt, M.** (1997). Nitrate Acts as a Signal to Induce Organic Acid Metabolism and Repress Starch Metabolism in Tobacco. *The Plant cell* **9**, 783-798.
- Schwacke, R., Ponce-Soto, G.Y., Krause, K., Bolger, A.M., Arsova, B., Hallab, A., Gruden, K., Stitt, M., Bolger, M.E., and Usadel, B.** (2019). MapMan4: A Refined Protein Classification and Annotation Framework Applicable to Multi-Omics Data Analysis. *Molecular Plant* **12**, 879-892.
- Segonzac, C., Boyer, J.-C., Ipotesi, E., Szponarski, W., Tillard, P., Touraine, B., Sommerer, N., Rossignol, M., and Gibrat, R.** (2007). Nitrate Efflux at the Root Plasma Membrane: Identification of an Arabidopsis Excretion Transporter. *The Plant cell* **19**, 3760-3777.
- Seligman, K., Saviani, E.E., Oliveira, H.C., Pinto-Maglio, C.A.F., and Salgado, I.** (2008). Floral Transition and Nitric Oxide Emission During Flower Development in *Arabidopsis thaliana* is Affected in Nitrate Reductase-Deficient Plants. *Plant and Cell Physiology* **49**, 1112-1121.
- Shearer, H.L., Cheng, Y.T., Wang, L., Liu, J., Boyle, P., Despres, C., Zhang, Y., Li, X., and Fobert, P.R.** (2012). Arabidopsis clade I TGA transcription factors regulate plant defenses in an NPR1-independent fashion. *Molecular plant-microbe interactions : MPMI* **25**, 1459-1468.
- Shin, R., Berg, R.H., and Schachtman, D.P.** (2005). Reactive Oxygen Species and Root Hairs in Arabidopsis Root Response to Nitrogen, Phosphorus and Potassium Deficiency. *Plant and Cell Physiology* **46**, 1350-1357.
- Signora, L., De Smet, I., Foyer, C.H., and Zhang, H.** (2001). ABA plays a central role in mediating the regulatory effects of nitrate on root branching in Arabidopsis. *The Plant Journal* **28**, 655-662.
- Song, Y.H., Smith, R.W., To, B.J., Millar, A.J., and Imaizumi, T.** (2012). FKF1 conveys timing information for CONSTANS stabilization in photoperiodic flowering. *Science (New York, N.Y.)* **336**, 1045-1049.

- Sparks, E.E., Drapek, C., Gaudinier, A., Li, S., Ansariola, M., Shen, N., Hennacy, J.H., Zhang, J., Turco, G., Petricka, J.J., Foret, J., Hartemink, A.J., Gordân, R., Megraw, M., Brady, S.M., and Benfey, P.N.** (2016). Establishment of Expression in the SHORTROOT-SCARECROW Transcriptional Cascade through Opposing Activities of Both Activators and Repressors. *Dev Cell* **39**, 585-596.
- Steyfkens, F., Zhang, Z., Van Zeebroeck, G., and Thevelein, J.M.** (2018). Multiple Transceptors for Macro- and Micro-Nutrients Control Diverse Cellular Properties Through the PKA Pathway in Yeast: A Paradigm for the Rapidly Expanding World of Eukaryotic Nutrient Transceptors Up to Those in Human Cells. *Front Pharmacol* **9**, 191-191.
- Sueyoshi, K., Mitsuyama, T., Sugimoto, T., Kleinhofs, A., Warner, R.L., and Oji, Y.** (1999). Effects of inhibitors for signaling components on the expression of the genes for nitrate reductase and nitrite reductase in excised barley leaves. *Soil Science and Plant Nutrition* **45**, 1015-1019.
- Sun, H., Qian, Q., Wu, K., Luo, J., Wang, S., Zhang, C., Ma, Y., Liu, Q., Huang, X., Yuan, Q., Han, R., Zhao, M., Dong, G., Guo, L., Zhu, X., Gou, Z., Wang, W., Wu, Y., Lin, H., and Fu, X.** (2014a). Heterotrimeric G proteins regulate nitrogen-use efficiency in rice. *Nature genetics* **46**, 652-656.
- Sun, J., Bankston, J.R., Payandeh, J., Hinds, T.R., Zagotta, W.N., and Zheng, N.** (2014b). Crystal structure of the plant dual-affinity nitrate transporter NRT1.1. *Nature* **507**, 73-77.
- Swift, J., Adame, M., Tranchina, D., Henry, A., and Coruzzi, G.M.** (2019). Water impacts nutrient dose responses genome-wide to affect crop production. *Nature Communications* **10**, 1374.
- Tabata, R., Sumida, K., Yoshii, T., Ohyama, K., Shinohara, H., and Matsubayashi, Y.** (2014). Perception of root-derived peptides by shoot LRR-RKs mediates systemic N-demand signaling. *Science* **346**, 343-346.
- Takei, K., Sakakibara, H., Taniguchi, M., and Sugiyama, T.** (2001). Nitrogen-dependent accumulation of cytokinins in root and the translocation to leaf: implication of cytokinin species that induces gene expression of maize response regulator. *Plant & cell physiology* **42**, 85-93.
- Takei, K., Ueda, N., Aoki, K., Kuromori, T., Hirayama, T., Shinozaki, K., Yamaya, T., and Sakakibara, H.** (2004). AtIPT3 is a key determinant of nitrate-dependent cytokinin biosynthesis in Arabidopsis. *Plant & cell physiology* **45**, 1053-1062.
- Tang, W., Ye, J., Yao, X., Zhao, P., Xuan, W., Tian, Y., Zhang, Y., Xu, S., An, H., Chen, G., Yu, J., Wu, W., Ge, Y., Liu, X., Li, J., Zhang, H., Zhao, Y., Yang, B., Jiang, X., Peng, C., Zhou, C., Terzaghi, W., Wang, C., and Wan, J.** (2019). Genome-wide associated study identifies NAC42-activated nitrate transporter conferring high nitrogen use efficiency in rice. *Nature Communications* **10**, 5279.
- Thimm, O., Blasing, O., Gibon, Y., Nagel, A., Meyer, S., Kruger, P., Selbig, J., Muller, L.A., Rhee, S.Y., and Stitt, M.** (2004). MAPMAN: a user-driven tool to display genomics data sets onto diagrams of metabolic pathways and other biological processes. *The Plant journal : for cell and molecular biology* **37**, 914-939.
- Tian, Q., Chen, F., Liu, J., Zhang, F., and Mi, G.** (2008). Inhibition of maize root growth by high nitrate supply is correlated with reduced IAA levels in roots. *Journal of Plant Physiology* **165**, 942-951.

- Tian, Q.Y., Sun, P., and Zhang, W.H.** (2009). Ethylene is involved in nitrate-dependent root growth and branching in *Arabidopsis thaliana*. *New Phytologist* **184**, 918-931.
- Tillard, P., Passama, L., and Gojon, A.** (1998). Are phloem amino acids involved in the shoot to root control of NO₃ uptake in *Ricinus communis* plants? *Journal of Experimental Botany* **49**, 1371-1379.
- Tocquin, P., Corbesier, L., Havelange, A., Pieltain, A., Kurtem, E., Bernier, G., and Périlleux, C.** (2003). A novel high efficiency, low maintenance, hydroponic system for synchronous growth and flowering of *Arabidopsis thaliana*. *BMC Plant Biology* **3**, 2.
- Touraine, B., Muller, B., and Grignon, C.** (1992). Effect of Phloem-Translocated Malate on NO₃ Uptake by Roots of Intact Soybean Plants. *Plant Physiol* **99**, 1118-1123.
- Triplett, E.W., Barnett, N.M., and Blevins, D.G.** (1980). Organic Acids and Ionic Balance in Xylem Exudate of Wheat during Nitrate or Sulfate Absorption. *Plant Physiology* **65**, 610-613.
- Tsay, Y.-F., Schroeder, J.I., Feldmann, K.A., and Crawford, N.M.** (1993). The herbicide sensitivity gene CHL1 of *Arabidopsis* encodes a nitrate-inducible nitrate transporter. *Cell* **72**, 705-713.
- Varala, K., Li, Y., Marshall-Colón, A., Para, A., and Coruzzi, G.M.** (2015). “Hit-and-Run” leaves its mark: Catalyst transcription factors and chromatin modification. *BioEssays : news and reviews in molecular, cellular and developmental biology* **37**, 851-856.
- Varala, K., Marshall-Colón, A., Cirrone, J., Brooks, M.D., Pasquino, A.V., Léran, S., Mittal, S., Rock, T.M., Edwards, M.B., Kim, G.J., Ruffel, S., McCombie, W.R., Shasha, D., and Coruzzi, G.M.** (2018). Temporal transcriptional logic of dynamic regulatory networks underlying nitrogen signaling and use in plants. *Proceedings of the National Academy of Sciences* **115**, 6494-6499.
- Vidal, E.A., Moyano, T.C., Riveras, E., Contreras-López, O., and Gutiérrez, R.A.** (2013). Systems approaches map regulatory networks downstream of the auxin receptor AFB3 in the nitrate response of *Arabidopsis thaliana* roots. *Proceedings of the National Academy of Sciences* **110**, 12840-12845.
- Vidal, E.A., Araus, V., Lu, C., Parry, G., Green, P.J., Coruzzi, G.M., and Gutierrez, R.A.** (2010). Nitrate-responsive miR393/AFB3 regulatory module controls root system architecture in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 4477-4482.
- von der Fecht-Bartenbach, J., Bogner, M., Dynowski, M., and Ludewig, U.** (2010). CLC-b-mediated NO₃/H⁺ exchange across the tonoplast of *Arabidopsis* vacuoles. *Plant & cell physiology* **51**, 960-968.
- Waldie, T., and Leyser, O.** (2018). Cytokinin targets auxin transport to promote shoot branching. *Plant physiology*, pp. 01691.02017.
- Walker, L., Boddington, C., Jenkins, D., Wang, Y., Grønlund, J.T., Hulsmans, J., Kumar, S., Patel, D., Moore, J.D., and Carter, A.** (2017). Changes in gene expression in space and time orchestrate environmentally mediated shaping of root architecture. *The Plant cell* **29**, 2393-2412.
- Wang, C.Q., Guthrie, C., Sarmast, M.K., and Dehesh, K.** (2014). BBX19 interacts with CONSTANS to repress FLOWERING LOCUS T transcription, defining a flowering time checkpoint in *Arabidopsis*. *The Plant cell* **26**, 3589-3602.
- Wang, P., Du, Y., Hou, Y.J., Zhao, Y., Hsu, C.C., Yuan, F., Zhu, X., Tao, W.A., Song, C.P., and Zhu, J.K.** (2015). Nitric oxide negatively regulates abscisic acid signaling in guard

- cells by S-nitrosylation of OST1. Proceedings of the National Academy of Sciences of the United States of America **112**, 613-618.
- Wang, R., Liu, D., and Crawford, N.M.** (1998). The Arabidopsis CHL1 protein plays a major role in high-affinity nitrate uptake. Proceedings of the National Academy of Sciences **95**, 15134-15139.
- Wang, R., Xing, X., and Crawford, N.** (2007). Nitrite acts as a transcriptome signal at micromolar concentrations in Arabidopsis roots. Plant physiology **145**, 1735-1745.
- Wang, R., Guegler, K., LaBrie, S.T., and Crawford, N.M.** (2000). Genomic analysis of a nutrient response in Arabidopsis reveals diverse expression patterns and novel metabolic and potential regulatory genes induced by nitrate. The Plant cell **12**, 1491-1509.
- Wang, R., Okamoto, M., Xing, X., and Crawford, N.M.** (2003). Microarray analysis of the nitrate response in Arabidopsis roots and shoots reveals over 1,000 rapidly responding genes and new linkages to glucose, trehalose-6-phosphate, iron, and sulfate metabolism. Plant Physiol **132**, 556-567.
- Wang, R., Xing, X., Wang, Y., Tran, A., and Crawford, N.M.** (2009). A Genetic Screen for Nitrate Regulatory Mutants Captures the Nitrate Transporter Gene NRT1.1. Plant Physiology **151**, 472-478.
- Wang, R., Tischner, R., Gutierrez, R.A., Hoffman, M., Xing, X., Chen, M., Coruzzi, G., and Crawford, N.M.** (2004). Genomic analysis of the nitrate response using a nitrate reductase-null mutant of Arabidopsis. Plant Physiol **136**, 2512-2522.
- Wang, Y.-Y., Cheng, Y.-H., Chen, K.-E., and Tsay, Y.-F.** (2018). Nitrate Transport, Signaling, and Use Efficiency. Annual review of plant biology **69**, 85-122.
- Wang, Y.H., Garvin, D.F., and Kochian, L.V.** (2001). Nitrate-induced genes in tomato roots. Array analysis reveals novel genes that may play a role in nitrogen nutrition. Plant Physiol **127**, 345-359.
- Wany, A., Foyer, C.H., and Gupta, K.J.** (2018). Nitrate, NO and ROS Signaling in Stem Cell Homeostasis. Trends in Plant Science **23**, 1041-1044.
- Weirauch, M.T., Yang, A., Albu, M., Cote, A.G., Montenegro-Montero, A., Drewe, P., Najafabadi, H.S., Lambert, S.A., Mann, I., Cook, K., Zheng, H., Goity, A., van Bakel, H., Lozano, J.-C., Galli, M., Lewsey, M.G., Huang, E., Mukherjee, T., Chen, X., Reece-Hoyes, J.S., Govindarajan, S., Shaulsky, G., Walhout, A.J.M., Bouget, F.-Y., Ratsch, G., Larrondo, L.F., Ecker, J.R., and Hughes, T.R.** (2014). Determination and inference of eukaryotic transcription factor sequence specificity. Cell **158**, 1431-1443.
- White, P.J.** (2012). Chapter 2 - Ion Uptake Mechanisms of Individual Cells and Roots: Short-distance Transport. In Marschner's Mineral Nutrition of Higher Plants (Third Edition), P. Marschner, ed (San Diego: Academic Press), pp. 7-47.
- Wirth, J., Chopin, F., Santoni, V., Viennois, G., Tillard, P., Krapp, A., Lejay, L., Daniel-Vedele, F., and Gojon, A.** (2007). Regulation of root nitrate uptake at the NRT2.1 protein level in *Arabidopsis thaliana*. The Journal of biological chemistry **282**, 23541-23552.
- Xu, H., Zhao, M., Zhang, Q., Xu, Z., and Xu, Q.** (2016a). The DENSE AND ERECT PANICLE 1 (DEP1) gene offering the potential in the breeding of high-yielding rice. Breed Sci **66**, 659-667.
- Xu, N., Wang, R., Zhao, L., Zhang, C., Li, Z., Lei, Z., Liu, F., Guan, P., Chu, Z., Crawford, N.M., and Wang, Y.** (2016b). The Arabidopsis NRG2 Protein Mediates Nitrate

- Signaling and Interacts with and Regulates Key Nitrate Regulators. *The Plant cell* **28**, 485-504.
- Yan, D., Easwaran, V., Chau, V., Okamoto, M., Ierullo, M., Kimura, M., Endo, A., Yano, R., Pasha, A., Gong, Y., Bi, Y.-M., Provart, N., Guttman, D., Krapp, A., Rothstein, S.J., and Nambara, E.** (2016). NIN-like protein 8 is a master regulator of nitrate-promoted seed germination in Arabidopsis. *Nature Communications* **7**, 13179.
- Yasuda, S., Aoyama, S., Hasegawa, Y., Sato, T., and Yamaguchi, J.** (2017). Arabidopsis CBL-Interacting Protein Kinases Regulate Carbon/Nitrogen-Nutrient Response by Phosphorylating Ubiquitin Ligase ATL31. *Mol Plant* **10**, 605-618.
- Yasuda, S., Sato, T., Maekawa, S., Aoyama, S., Fukao, Y., and Yamaguchi, J.** (2014). Phosphorylation of Arabidopsis ubiquitin ligase ATL31 is critical for plant carbon/nitrogen nutrient balance response and controls the stability of 14-3-3 proteins. *The Journal of biological chemistry* **289**, 15179-15193.
- Yazaki, J., Galli, M., Kim, A.Y., Nito, K., Aleman, F., Chang, K.N., Carvunis, A.-R., Quan, R., Nguyen, H., Song, L., Alvarez, J.M., Huang, S.-s.C., Chen, H., Ramachandran, N., Altmann, S., Gutiérrez, R.A., Hill, D.E., Schroeder, J.I., Chory, J., LaBaer, J., Vidal, M., Braun, P., and Ecker, J.R.** (2016). Mapping transcription factor interactome networks using HaloTag protein arrays. *Proceedings of the National Academy of Sciences* **113**, E4238-E4247.
- Yu, L.-H., Miao, Z.-Q., Qi, G.-F., Wu, J., Cai, X.-T., Mao, J.-L., and Xiang, C.-B.** (2014). MADS-box transcription factor AGL21 regulates lateral root development and responds to multiple external and physiological signals. *Molecular plant* **7**, 1653-1669.
- Yu, P., Baldauf, J.A., Lithio, A., Marcon, C., Nettleton, D., Li, C., and Hochholdinger, F.** (2016). Root Type-Specific Reprogramming of Maize Pericycle Transcriptomes by Local High Nitrate Results in Disparate Lateral Root Branching Patterns **170**, 1783-1798.
- Yuan, S., Zhang, Z.-W., Zheng, C., Zhao, Z.-Y., Wang, Y., Feng, L.-Y., Niu, G., Wang, C.-Q., Wang, J.-H., Feng, H., Xu, F., Bao, F., Hu, Y., Cao, Y., Ma, L., Wang, H., Kong, D.-D., Xiao, W., Lin, H.-H., and He, Y.** (2016). Arabidopsis cryptochrome 1 functions in nitrogen regulation of flowering. *Proceedings of the National Academy of Sciences* **113**, 7661-7666.
- Zhang, G.-B., Meng, S., and Gong, J.-M.** (2018). The Expected and Unexpected Roles of Nitrate Transporters in Plant Abiotic Stress Resistance and Their Regulation. *Int J Mol Sci* **19**, 3535.
- Zhang, H., and Forde, B.G.** (1998). An Arabidopsis MADS Box Gene That Controls Nutrient-Induced Changes in Root Architecture. *Science* **279**, 407-409.
- Zhang, H., Jennings, A., Barlow, P.W., and Forde, B.G.** (1999). Dual pathways for regulation of root branching by nitrate. *Proceedings of the National Academy of Sciences* **96**, 6529-6534.
- Zhang, J., Fengler, K.A., Van Hemert, J.L., Gupta, R., Mongar, N., Sun, J., Allen, W.B., Wang, Y., Weers, B., Mo, H., Lafitte, R., Hou, Z., Bryant, A., Ibraheem, F., Arp, J., Swaminathan, K., Moose, S.P., Li, B., and Shen, B.** (2019). Identification and characterization of a novel stay-green QTL that increases yield in maize. *Plant biotechnology journal* **17**, 2272-2285.
- Zhang, K., Novak, O., Wei, Z., Gou, M., Zhang, X., Yu, Y., Yang, H., Cai, Y., Strnad, M., and Liu, C.J.** (2014). Arabidopsis ABCG14 protein controls the acropetal translocation of root-synthesized cytokinins. *Nat Commun* **5**, 3274.

- Zheng, D., Han, X., An, Y., Guo, H., Xia, X., and Yin, W.** (2013). The nitrate transporter NRT2. 1 functions in the ethylene response to nitrate deficiency in *Arabidopsis*. *Plant, cell & environment* **36**, 1328-1337.
- Zhou, J., Jiao, F., Wu, Z., Li, Y., Wang, X., He, X., Zhong, W., and Wu, P.** (2008). OsPHR2 is involved in phosphate-starvation signaling and excessive phosphate accumulation in shoots of plants. *Plant Physiol* **146**, 1673-1686.
- Zhou, J.J., Fernandez, E., Galvan, A., and Miller, A.J.** (2000). A high affinity nitrate transport system from *Chlamydomonas* requires two gene products. *FEBS letters* **466**, 225-227.
- Zhuo, D., Okamoto, M., Vidmar, J.J., and Glass, A.D.** (1999). Regulation of a putative high-affinity nitrate transporter (Nrt2;1At) in roots of *Arabidopsis thaliana*. *The Plant journal : for cell and molecular biology* **17**, 563-568.
- Zioni, A.B., Vaadia, Y., and Lips, S.H.** (1971). Nitrate Uptake by Roots as Regulated by Nitrate Reduction Products of the Shoot. *Physiologia plantarum* **24**, 288-290.

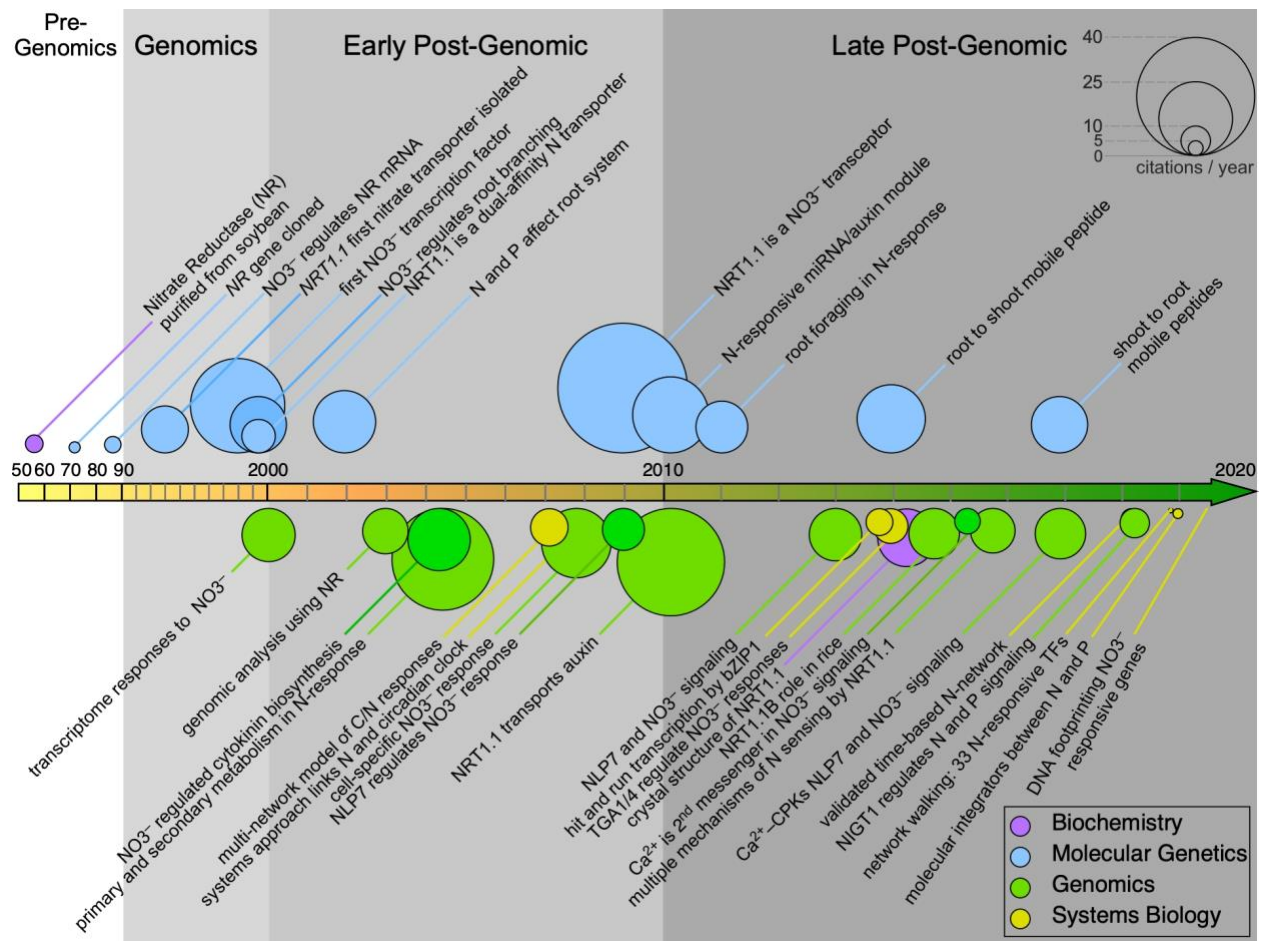


Figure 1. A timeline with milestone publications in nitrate signaling.

The “nitrate” and “Arabidopsis” terms were used to retrieve articles and their citation numbers from web of science (www.webofknowledge.com). Please note that the size of circles is proportional to the number of citations normalized by years from publication. Top articles related to nitrate signaling were included for this timeline.

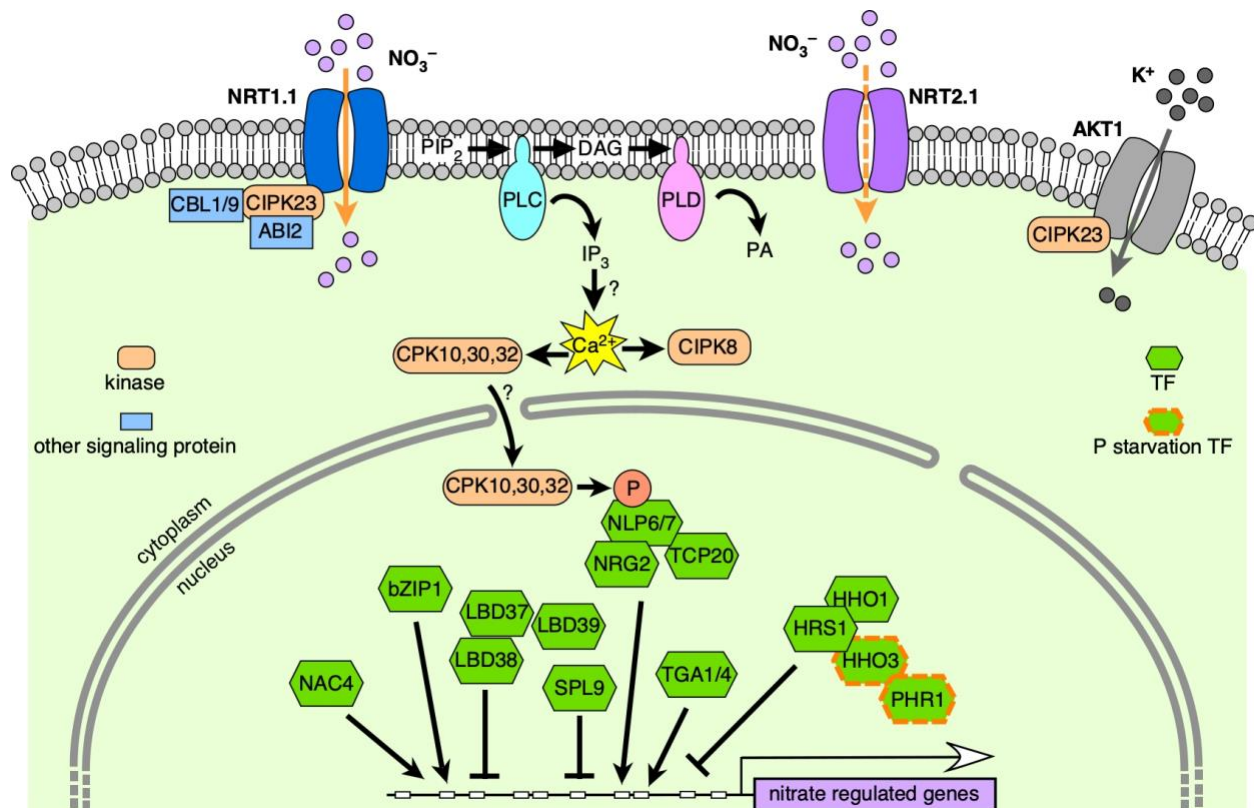


Figure 2. A summary of nitrate signaling pathways.

NRT1.1 is at the first layer of the nitrate signaling pathway. CIPK23, CBL1/9, and ABI2 control the NRT1.1 phosphorylation status switching its nitrate affinity. The transduced nitrate signal through NRT1.1 activates PLC activity, increasing calcium accumulation. Then, the calcium signals are decoded by subgroup III CPKs kinases, which in turn phosphorylate the NLP7 transcription factor, promoting its nuclear retention and the activation of nitrate-responsive target genes. NLP7 physically interacts with NLP6, TCP20 and NRG2 to activate the expression of genes involved in nitrate metabolism. NRT2.1 has also been proposed as a nitrate sensor. A myriad of TFs have been discovered as important regulators of gene expression in response to nitrate, or as integrators of N and P signaling (HRS1, HHO1 and PHR1). See text and Supplemental Data Set 2 for relevant references.

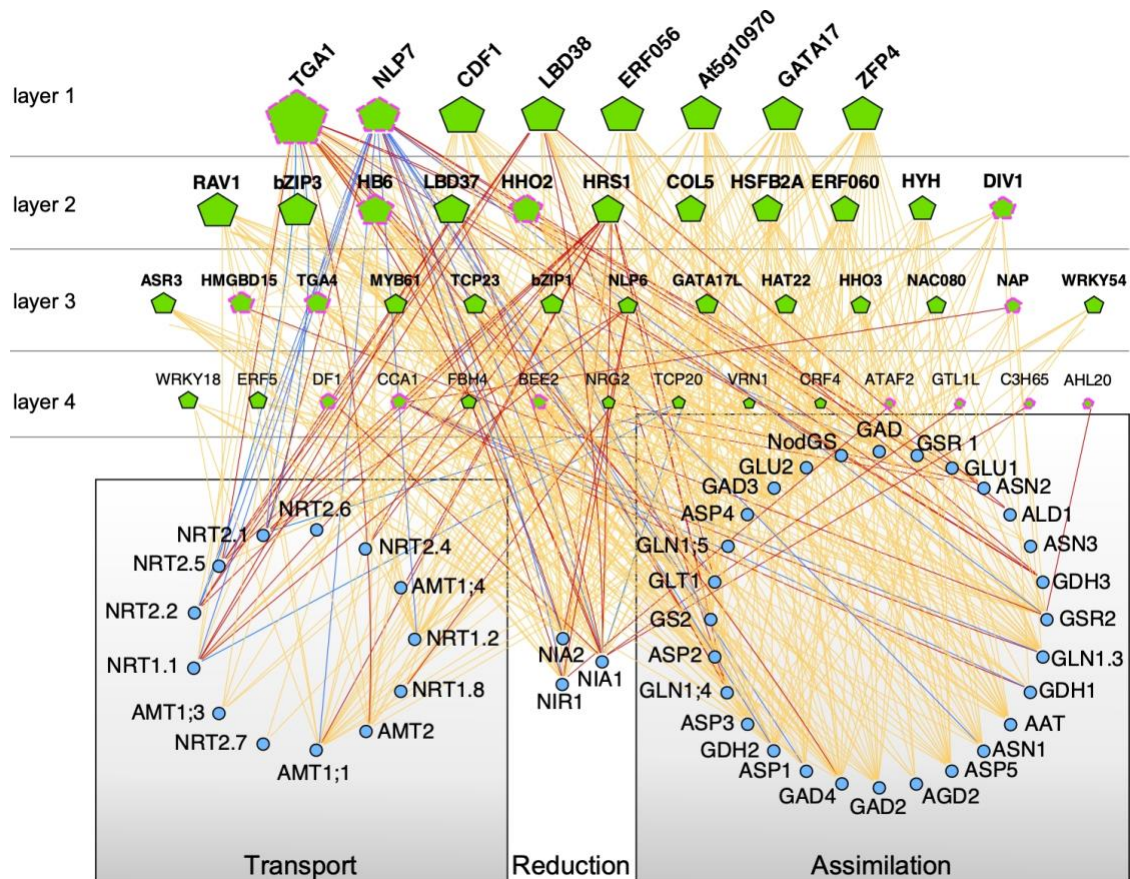


Figure 3. Integrative gene regulatory network analysis with the most influential TFs in N-regulatory networks.

Genes are drawn as pentagons (transcription factors) or circles (target genes). The size of the triangle is proportional to the number of targets bound or regulated by each TF (outdegree). The four TF layers were organized according to their outdegree. TFs were selected based on published evidence of TF-regulation or TF-binding to genes involved in N-transport, N-reduction and N-assimilation in Arabidopsis. Blue edges: Chromatin immunoprecipitation (ChIP) binding data (Gutiérrez et al., 2008; Marchive et al., 2013; Alvarez et al., 2014; Guan et al., 2014; Para et al., 2014). Orange edges: TARGET and Yeast one hybrid data (Para et al., 2014; Medici et al., 2015; Varala et al., 2018; Brooks et al., 2019; Gaudinier et al., 2018). Red edges: in planta TF-regulation and chromatin accessibility (Gutiérrez et al., 2008; Rubin et al., 2009; Marchive et al., 2013; Xu et al., 2016b; Maeda et al., 2018; Brooks et al., 2019; Alvarez et al., 2019). The pink border of the pentagon denotes TF regulated in response to nitrate (Alvarez et al., 2019).

Nitrate 2020: Thirty years from transport to signaling networks

Elena A Vidal, José M Alvarez, Viviana Araus, Eleodoro Riveras, Matthew Brooks, Gabriel Krouk, Sandrine Ruffel, Laurence Lejay, Nigel Crawford, Gloria M. Coruzzi and Rodrigo A Gutiérrez
Plant Cell; originally published online March 13, 2020;
DOI 10.1105/tpc.19.00748

This information is current as of July 8, 2020

| | |
|---------------------------------|---|
| Supplemental Data | /content/suppl/2020/03/13/tpc.19.00748.DC1.html |
| Permissions | https://www.copyright.com/ccc/openurl.do?sid=pd_hw1532298X&issn=1532298X&WT.mc_id=pd_hw1532298X |
| eTOCs | Sign up for eTOCs at: http://www.plantcell.org/cgi/alerts/ctmain |
| CiteTrack Alerts | Sign up for CiteTrack Alerts at: http://www.plantcell.org/cgi/alerts/ctmain |
| Subscription Information | Subscription Information for <i>The Plant Cell</i> and <i>Plant Physiology</i> is available at: http://www.aspb.org/publications/subscriptions.cfm |