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### ► To cite this version:

Christelle Loncke, Pierre Noziere, J. Vernet, H. Lapierre, L. Bahloul, et al.. Net hepatic release of glucose from precursor supply in ruminants: a meta-analysis. *Animal*, 2020, 14 (7), pp.1422-1437. 10.1017/S1751731119003410 . hal-02900519

**HAL Id: hal-02900519**


**<https://hal.inrae.fr/hal-02900519>**

Submitted on 12 Nov 2021

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# Net hepatic release of glucose from precursor supply in ruminants: a meta-analysis

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(Received 16 July 2019; Accepted 28 November 2019; First published online 23 January 2020)

*For their glucose supply, ruminants are highly dependent on the endogenous synthesis in the liver, but despite the numerous studies that evaluated hepatic glucose production, very few simultaneously measured hepatic glucose production and uptake of all precursors. As a result, the variability of precursor conversion into glucose in the liver is not known. The present study aimed at investigating by meta-analysis the relationships between hepatic glucose net release and uptake of precursors. We used the FLuxes of nutrients across Organs and tissues in Ruminant Animals database, which gathers international results on net nutrient fluxes at splanchnic level measured in catheterized animals. Response equations were developed for intakes up to 41 g DM intake/kg BW per day of diets varying from 0 to 100 g of concentrate/100 g DM in the absence of additives. The net hepatic uptake of propionate,  $\alpha$ -amino-N and L-lactate was linearly and better related to their net portal appearance (NPA) than to their afferent hepatic flux. Blood flow data were corrected for lack of deacetylation of the para-aminohippuric acid, and this correction was shown to impact the response equations. To develop response equations between the availability of precursors (portal appearance and hepatic uptake) and net glucose hepatic release, missing data on precursor fluxes were predicted from dietary characteristics using previously developed response equations. Net hepatic release of glucose was curvilinearly related to hepatic supply and uptake of the sum of precursors, suggesting a lower conversion rate of precursors at high precursor supply. Factors of variation were explored for the linear portion of this relationship, which applied to NPA of precursors ranging from 0.99 to 9.60 mmol C/kg BW per h. Hepatic release of glucose was shown to be reduced by the portal absorption of glucose from diets containing bypass starch and to be increased by an increased uptake of  $\beta$ -hydroxybutyrate indicative of higher body tissue mobilization. These relationships were affected by the physiological status of the animals. In conclusion, we established equations that quantify the net release of glucose by the liver from the net availability of precursors. They provide a quantitative overview of factors regulating hepatic glucose synthesis in ruminants. These equations can be linked with the predictions of portal absorption of nutrients from intake and dietary characteristics, and provide indications of glucose synthesis from dietary characteristics.*

**Key words:** nutrient flux, gluconeogenesis, meta-analysis, glucose, liver

## Implications

Glucose, a key nutrient for ruminants, is synthesized in the liver from a range of precursors. The response equations developed from meta-analysis highlight the variable rate of conversion of precursors into glucose. This rate follows a law of diminishing return at high precursor availability and increases with peripheral glucose demand or negative energy balance. Also glucose

supplied from bypass starch partially substitutes for glucose synthesized in the liver. These results can be used in precision feeding strategies aiming at close coverage of glucose requirements. They will also contribute to the development of nutrient-based feeding systems for ruminants.

## Introduction

Glucose is a key energy yielding nutrient for all mammals. It is essential for brain, foetal growth, gut and muscle metabolism (Hammon *et al.*, 2016) and is the major precursor of lactose,

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the milk sugar. The ruminants have this peculiarity that they constantly need to synthesize glucose, because limited amounts, if any, are absorbed into the portal circulation unless high-starch diets are fed (Reynolds, 2006; Loncke *et al.*, 2009c). Glucose is synthesized in the liver and the kidneys, and the liver is the most important glucose synthesizing organ in ruminants (Brockman, 2005). The availability of glucose precursors is critical to support glucose synthesis.

Despite the numerous studies that evaluated net hepatic glucose release, a limited number of studies simultaneously measured the whole range of precursor uptake and conversion into glucose. Liver uptake of glucogenic precursors accounted for the majority of C released as glucose in maintenance fed sheep (e.g. El-Sabagh *et al.*, 2013) but failed to do so in *postpartum* dairy cows as reviewed by Larsen and Kristensen (2013), even after accounting for methodological effects on liver blood flow measurements. We postulated that a meta-analysis on published data would provide an overview of hepatic conversion of glucogenic precursors into glucose across a wide range of feeding and production conditions in ruminants, as in Loncke *et al.* (2015) for ketogenic nutrients across the liver.

The objectives of the present work were to investigate published knowledge on hepatic nutrient fluxes in both sheep and cattle in different physiological status and explore the quantitative relationships between the availability of major precursors, that is, propionate, amino acids and L-lactate, and the net hepatic release of glucose, as well as the regulatory factors which could impact these relationships. We first tested whether net hepatic fluxes of precursors responded to changes in total afferent flux (flux of nutrient supplied to the liver via the portal vein and the hepatic artery, see Supplementary Material S1) to the liver or to net portal appearance (NPA). Having demonstrated better relationships with NPA, the study focused on the incremental responses of hepatic fluxes to changes in NPA, thereby describing the responses to increments of 'net' supply of nutrients as in Loncke *et al.* (2015). The impact of analytical methods used to determine liver blood flow was tested. Factors driving the net hepatic uptake of insulin and interrelationships with glucose fluxes were also investigated.

## Material and methods

### *Selection of relevant publications from the FLuxes of nutrients across Organs and tissues in Ruminant Animals database*

We used the Flora (FLuxes of nutrients across Organs and tissues in Ruminant Animals; Vernet and Ortigues-Marty, 2006) database built from approximately 250 international publications on splanchnic fluxes in multi-catheterized ruminants. Publications anterior to 2008 were used for model development. To be eligible, publications had to report the net hepatic fluxes (see Supplementary Material S1 for definitions) of glucose or one of its major precursors, propionate, total amino acids (measured in the selected publication by its proxy

$\alpha$ -amino-N) and L-lactate (Hammon *et al.*, 2016). Publications or treatments reporting intra-venous infusion of nutrients, use of digestion or metabolism modifiers or non-steady state feeding were discarded. Five eligible datasets were identified to study propionate,  $\alpha$ -amino-N, L-lactate, glucose and infusion of propionate with respectively 29 (69), 26 (65), 25 (66), 46 (106) and 5 (16) experimental groups (treatments). No publication was available that reported total amino acids fluxes, only  $\alpha$ -amino-N fluxes were available. Insulin concentrations and fluxes were also considered in a separate dataset when available (9 experimental groups, 19 treatments). The list of references used for the meta-analysis is given in Supplementary Material S2.

Because the selected publications did not systematically report the chemical composition and nutritional value of feeds and diets, the feeds and diets from all relevant publications were described according to the INRA Feed Evaluation System (2007) as detailed by Loncke *et al.* (2009c and 2015). As the nutritional status of the animals differed among publications and treatments, the availability of endogenous precursors (alanine and glycerol) coming from tissue mobilization (Leng, 1970) was estimated, when relevant, from calculated energy balance (Loncke *et al.*, 2015) and tissue composition of mobilized energy (Supplementary Material Table S1). Because in all these experiments animals were fed with frequent meal distributions to keep nutrient supply and animal metabolism at steady state, it was considered that glycogen stores remained constant and did not contribute to net hepatic glucose output.

Attention was paid to the analytical methods used to determine net hepatic fluxes to ensure that all results could be combined in the meta-analysis, as described in Supplementary Material S3 and Supplementary Material Table S2. A special focus was made on blood or plasma flow measurements, usually based on the para-aminohippuric acid (pAH) down-stream dilution method. The pAH is acetylated across the liver and therefore incompletely recovered in the hepatic veins if not deacetylated, as demonstrated in sheep by Katz and Bergman (1969) and in cows by Kristensen *et al.* (2009) and Rodriguez-Lopez *et al.* (2014) but 80.4% of available publications did not correct for it.

To relate net glucose flux in the same unit as its precursors, the fluxes were expressed in C units when glucose and precursors were included in the same model (3 mol C/mol propionate, L-lactate or glycerol, 6 mol C/mol glucose). The C supplied from total amino acids was calculated from the average C content of total amino acids released in the portal blood and taken up by the liver (assumed to be 4 mol C/mol amino acid-N, based on Reynolds *et al.* (1988)) and the fluxes of total amino acids. For this, fluxes of  $\alpha$ -amino-N were converted to total amino acids (mmol/kg BW per h) as  $1.3958 \times$  moles of  $\alpha$ -amino-N (Martineau *et al.*, 2009). All absorbed amino acids were assumed to be glucogenic, no correction was applied for the non-glucogenic leucine and lysine which represented only *circa* 6% of net amino acid C uptake by the liver (Blouin *et al.*, 2002; Doepel *et al.*, 2009) that we considered as negligible. The alanine

mobilized from muscle was also considered as glucogenic based on the evidence reviewed by Larsen and Kristensen (2013) that alanine is the only amino acid involved in the inter-organ transfer of nitrogen in *postpartum* dairy cows. Subsequently and for the sake of accuracy, reference is made to  $\alpha$ -amino-N for all results derived from measured and reported  $\alpha$ -amino-N fluxes and expressed in moles of  $\alpha$ -amino-N, and reference is made to total amino acids for all results converted into moles of C from total amino acids from  $\alpha$ -amino-N.

#### Meta-analyses

The meta-analyses focused on the responses of the different net hepatic fluxes ( $Y$  variables) to variations in explanatory variables ( $X$ ) intra-studies, as detailed in Sauvant *et al.* (2008) and Loncke *et al.* (2015). Net hepatic fluxes are defined as the difference between hepatic efferent and afferent fluxes of nutrients measured in blood or plasma, as detailed in Loncke *et al.* (2015). Net fluxes can be either positive indicating a net hepatic release or negative indicating a net uptake. Net portal appearance is calculated from the difference between portal venous and arterial fluxes.

**Description of the meta-design.** The meta-design was described by statistics (mean, SD, range of values) generated for each parameter in the selected datasets. Species and physiological status effects were tested by one-way ANOVA. Normal data distribution and homogeneity of variances were tested by Shapiro–Wilk and Levene tests. All data (nutrient intake, NPA and net hepatic fluxes) were expressed as a function of BW to ensure normal distribution of the variables when combined across species (Loncke *et al.*, 2009c).

**Models of net hepatic flux of nutrients.** For each glucogenic precursor (propionate, total amino acids, L-lactate) it was initially tested whether their net hepatic uptake was best related to their total afferent flux or their NPA. Because net hepatic uptake was better related to net portal fluxes than to afferent fluxes, models developed included the net portal fluxes only. For L-lactate, the energy balance of the animals was also considered as a predictor because of the role of L-lactate in the C economy in the body. Second, the potential contribution of each precursor to net hepatic glucose release was evaluated by studying the relationships between the net hepatic glucose release and the availability of each precursor considered individually. Third, the net hepatic glucose release was considered in relation to the sum of all precursors. This last step required that all precursor and glucose fluxes had been measured in any given study. This was not always the case. An all-precursor dataset was constituted with studies that reported the net hepatic release of glucose and the NPA of at least one precursor ( $n_{\text{experimental groups}} = 45$ ;  $n_{\text{treatments}} = 104$ ). When missing, NPA of other precursors was estimated according to Loncke *et al.* (2009c), and their net hepatic uptakes were estimated according to the relationships developed in this work. Estimations represented 41%,

29% and 40% of NPA data and 43%, 28% and 46% of net hepatic flux data for propionate, L-lactate and  $\alpha$ -amino-N, respectively. Estimations were evaluated by comparing predicted and observed values when available. No biases were detected (results not shown).

**Data coding.** Experimental groups were coded to specifically explore intra-study variations in NPA or net hepatic uptake of nutrients ( $X$  predictors). The physiological status was also coded for each study because of its potential influence on nutrient net hepatic flux (Loncke *et al.*, 2015). Within experiments, data were considered only if the reported  $X$  variable varied by more than 2% within-study, as in Loncke *et al.* (2015). This conservative threshold eliminated studies with absolutely no variation in  $X$  variables but retained studies with small variations. No selection was imposed on the  $Y$  variables.

**Determination of response equations.** Within-study relationships between  $Y$  (net hepatic flux of nutrient) and the potential explanatory variable  $X$  were studied using a variance–covariance GLM model:

$$Y = \alpha + \beta X + \text{animal profile} + \alpha_i(\text{animal profile}) + \text{animal profile} \times X + \text{error}$$

where  $\alpha$  is the overall intercept and  $\alpha_i$  is the effect of study  $i$  on intercept  $\alpha$ . The study effect is nested within animal profile, that is, animal species or physiological state. Parameter  $\beta$  is the slope of the overall relationship. When relevant for hepatic glucose release, non-linear models were also considered:

$$Y = b \times (1 - e^{(-\alpha \times X)}) + \alpha_i + \text{error}$$

where  $\alpha$  is the overall intercept and  $\alpha_i$  is the effect of study  $i$  on intercept  $\alpha$ .

In all cases, the best fit model (based on the minimization of the RMSE and the SE associated with the parameters, the maximization of the adjusted  $R^2$  and the less number of interfering factors) was selected. Studies were considered as fixed factors in the models because experimental conditions and methods are specific to each study and could influence the study effect. This strategy has no impact on the statistical effect of the covariate (Sauvant *et al.*, 2008). The range of inference for this meta-analysis is, therefore, limited to the domain of the specific experiments in the dataset (Sauvant *et al.*, 2008). When the study effect was not considered, it was specified in the results. All statistical analyses were carried out using Minitab (Version 17). The normality of residuals was checked and outliers were identified on the basis of residues, HI leverage, Cook's distance and DFITS, as in Loncke *et al.* (2009c and 2015).

**Determination of factors influencing the response equations.** To explore the causes of heterogeneity between

**Table 1** Description of animals (non-productive, growing, gestating or lactating ruminants) and diets used for the meta-analysis<sup>a</sup>

|  | Non-productive adults <sup>b</sup>                                 |      |      |      |      | Growing animals  |      |      |      |      | Gestating animals   |       |      |      |      | Lactation <sup>c</sup>   |      |       |       |      | Physiological status effect<br><i>P</i> -value |
|--|--|------|------|------|------|--|------|------|------|------|---|-------|------|------|------|--|------|-------|-------|------|--|
|  | <i>n</i> <sub>t</sub> sheep = 81; <i>n</i> <sub>t</sub> cattle = 7 |      |      |      |      | <i>n</i> <sub>t</sub> sheep = 7; <i>n</i> <sub>t</sub> cattle = 26 |      |      |      |      | <i>n</i> <sub>t</sub> sheep = 5; <i>n</i> <sub>t</sub> cattle = 8 |       |      |      |      | <i>n</i> <sub>t</sub> sheep = 2; <i>n</i> <sub>t</sub> cattle = 21 |      |       |       |      |  |
|  | <i>n</i> <sub>t</sub>  | Mean | SD   | Min  | Max  | <i>n</i> <sub>t</sub>  | Mean | SD   | Min  | Max  | <i>n</i> <sub>t</sub>   | Mean  | SD   | Min  | Max  | <i>n</i> <sub>t</sub>  | Mean | SD    | Min   | Max  |  |
| Dietary composition <sup>d</sup> (g/kg DM) |  |      |      |      |      |  |      |      |      |      |   |       |      |      |      |  |      |       |       |      |  |
| Crude fibre                                | 88   | 258  | 92.5 | 39.5 | 404  | 33   | 194  | 94.6 | 87.9 | 371  | 13  | 259   | 62.2 | 177  | 374  | 23   | 175  | 57.9  | 107   | 280  | <0.001   |
| NDF  | 88   | 500  | 156  | 133  | 712  | 33   | 382  | 163  | 181  | 710  | 13  | 481   | 76.2 | 352  | 564  | 23   | 372  | 91.6  | 210   | 506  | <0.001   |
| ADF  | 88   | 279  | 96   | 43.8 | 393  | 33   | 211  | 104  | 93.8 | 403  | 13  | 288   | 57.7 | 204  | 367  | 23   | 196  | 63    | 108   | 287  | <0.001   |
| Starch                                     | 88   | 147  | 20.5 | 0    | 709  | 33   | 282  | 33.8 | 0    | 571  | 13  | 78.8  | 17.7 | 0    | 190  | 23   | 204  | 24.1  | 21    | 436  | <0.001   |
| CP   | 88   | 124  | 35   | 44.7 | 247  | 33   | 139  | 30.9 | 65.5 | 178  | 13  | 150   | 29.9 | 113  | 196  | 23   | 156  | 29.3  | 122   | 214  | <0.001   |
| Digestible OM                              | 88   | 622  | 90.9 | 458  | 852  | 33   | 674  | 109  | 461  | 789  | 13  | 629   | 76.9 | 512  | 736  | 23   | 706  | 42.5  | 605   | 776  | <0.001   |
| Digestible NDF                             | 88   | 292  | 102  | 94.9 | 474  | 33   | 215  | 82.7 | 97.3 | 400  | 13  | 268   | 34.7 | 212  | 323  | 23   | 235  | 84.1  | 121   | 403  | <0.001   |
| Digestible CP                              | 88   | 74.3 | 29.8 | 4.9  | 135  | 33   | 89.3 | 32.8 | 5    | 130  | 13  | 101   | 26.9 | 65.6 | 139  | 23   | 107  | 30.2  | 61.1  | 166  | <0.001   |
| Rumen fermentable OM                       | 88   | 503  | 40.4 | 413  | 586  | 33   | 481  | 31   | 421  | 586  | 13  | 488   | 38.5 | 450  | 544  | 23   | 541  | 44.1  | 436   | 631  | <0.001   |
| Rumen digestible NDF                       | 88   | 263  | 92   | 85.4 | 427  | 33   | 193  | 74.3 | 87.7 | 360  | 13  | 241   | 31.1 | 190  | 291  | 23   | 211  | 75.8  | 109   | 363  | <0.001   |
| Rumen fermentable CP                       | 88   | 72.9 | 23.9 | 27.6 | 128  | 33   | 75.2 | 22.9 | 28.5 | 119  | 13  | 105   | 31.1 | 58.7 | 149  | 23   | 102  | 25.5  | 66.1  | 167  | <0.001   |
| INRA (2007) PDI                            | 88   | 78.7 | 19.4 | 27.7 | 112  | 33   | 91.3 | 22   | 26.9 | 115  | 13  | 84.5  | 9.1  | 73.3 | 96.8 | 23   | 97.9 | 15.2  | 80.5  | 126  | <0.001   |
| ME (MJ/kg DM)                              | 88   | 9.51 | 1.51 | 6.75 | 13.1 | 33   | 10.4 | 1.79 | 6.76 | 12.2 | 13  | 9.55  | 1.19 | 7.75 | 11.4 | 23   | 11.2 | 0.955 | 9.17  | 12.5 | <0.001   |
| Proportion of concentrate (g/100 g DM)     | 88   | 29.5 | 35.8 | 0    | 100  | 33   | 47.3 | 38   | 0    | 98.8 | 13  | 35.3  | 22.7 | 0    | 75   | 23   | 48.5 | 29.4  | 10    | 100  | 0.025  |
| Intake (g/kg BW per day)                   |  |      |      |      |      |  |      |      |      |      |   |       |      |      |      |  |      |       |       |      |  |
| DM   | 88   | 21.2 | 7.96 | 0    | 41.5 | 33   | 20.6 | 7.79 | 8.21 | 38.8 | 13  | 19.5  | 7.41 | 0    | 29.6 | 23   | 26.4 | 6.4   | 19.1  | 36.9 | 0.014  |
| Digestible OM                              | 88   | 13.2 | 5.09 | 0    | 23   | 33   | 13.5 | 4.1  | 4.39 | 20.2 | 13  | 12.5  | 5.14 | 0    | 19.1 | 23   | 18.7 | 4.39  | 13.1  | 28.6 | <0.001   |
| Rumen fermentable OM                       | 88   | 10.7 | 4.17 | 0    | 20.8 | 33   | 9.93 | 3.74 | 3.45 | 18.5 | 13  | 9.63  | 3.83 | 0    | 13.8 | 23   | 14.2 | 3.05  | 11.2  | 17.4 | <0.001   |
| Rumen digestible NDF                       | 88   | 5.68 | 3.17 | 0    | 14.3 | 33   | 4.27 | 2.98 | 1.25 | 11.2 | 13  | 4.65  | 1.68 | 0    | 6.58 | 23   | 5.36 | 1.68  | 2.68  | 10.6 | 0.095  |
| Starch                                     | 88   | 3.13 | 4.18 | 0    | 15.2 | 33   | 5.27 | 3.78 | 0    | 12.4 | 13  | 1.76  | 1.6  | 0    | 5    | 23   | 5.75 | 3.77  | 0.4   | 16.1 | 0.001  |
| Energy balance (kJ/kg BW per day)          | 88   | 20.8 | 57.6 | -158 | 109  | 33   | 43.9 | 49.5 | -173 | 91.2 | 13  | -46.4 | 51.4 | -159 | 15.3 | 23   | -2.4 | 39    | -70.9 | 58.8 | <0.001   |

Min = minimum value; Max = maximum value; *n*<sub>t</sub> = number of treatments; OM = organic matter; ME = metabolizable energy.

<sup>a</sup>See the list of publications used in the meta-analysis in Supplementary Material S2.

<sup>b</sup>Fasting included.

<sup>c</sup>Dairy cattle were between 11 and 240 days in milk.

<sup>d</sup>Dietary composition and intake calculated by additivity according to INRA Feed Tables (INRA, 2007); PDI = protein digestible in the intestine, defined as the minimum between protein digestible in the intestine when energy is limiting and protein digestible in the intestine when nitrogen is limiting.

studies, all the factors which potentially altered the relationships between  $X$  and  $Y$  variables were considered and tested as interfering factors on the model parameters (residuals, individual within-study slopes and least-squares means (LSMeans)) as in Loncke *et al.* (2015). This recognizes the non-random character of some of the variability present in the data and evaluates a 'true' within-study response. The impact of qualitative factors (analytical methods, blood or plasma concentration, physiological state and animal species) was tested by one way ANOVA. The impact of quantitative factors (nutrient and hormonal concentrations and fluxes as available, intake level and diet composition) was established by regression between model parameters and potential quantitative interfering factors. Only the significant interfering factors are reported.

*Optimization of the response equations.* When an interfering factor had been detected as significant and if not correlated to the main explanatory variable  $X$ , the benefits of adding it as another  $X$  variable to the model were evaluated on the fit (Sauvant *et al.*, 2008; Loncke *et al.*, 2015). If improvement was not significant, the interfering factor was excluded from the final response equation. The model was considered stable when the optimization analyses showed no more significant interfering factors on slopes, residuals and LSMean or no significant improvement of fit. Parameters and correlations were considered significant at  $P \leq 0.05$ , whereas  $0.05 < P \leq 0.10$  indicated a trend. The RMSE was considered low when lower than 15% of the mean value of the predicted variable. This contributed to the good fit of the model.

## Results

### *Description of the meta-design*

Data were normally distributed and variances were homogeneous in the different datasets. Differences in intake, dietary composition and fluxes of nutrients were greater between physiological status (non-productive adults, growing, gestating or lactating animals; Table 1) than between species (bovine or ovine;  $P < 0.001$ ). As a result, the physiological status was included in the models, and interpretation accounted for the confounding effects of diet intake and composition, and animal species. Differences in intake and dietary composition and fluxes of nutrients according to species are however detailed in Supplementary Material S4 and Supplementary Material Tables S3a and b.

The potential contribution of precursors to net hepatic release of glucose averaged  $51.4 \pm 19.9\%$ ,  $40.4 \pm 27.3\%$  and  $23.5 \pm 23.8\%$  for propionate,  $\alpha$ -amino-N and L-lactate, respectively, with large differences between physiological statuses (Table 2). Average values showed a general excess of C from total precursors, except in gestating and lactating animals. The amount of mobilized glycerol was estimated at  $1.59 \pm 1.26$  mmol/kg BW per h and the average amount of alanine mobilized from muscles was estimated at  $0.092 \pm 0.073$  mmol C/kg BW per h ( $n = 32$ , Table 2). Accuracy of these

calculated values could not be evaluated due to the lack of published data. Out of the analytical methods tested, only the pAH analysis significantly influenced the relationships (Supplementary Material S3 and Table S2).

*Net hepatic uptake of propionate, L-lactate and  $\alpha$ -amino-N*  
*Comparison of models using net portal appearance or afferent fluxes as predictor.* Quantitatively, NPA contributed to  $85.8 \pm 6.1\%$  (propionate),  $4.8 \pm 1.7\%$  ( $\alpha$ -amino-N) and  $15.4 \pm 6.3\%$  (L-lactate) of the hepatic afferent flux. For propionate, net hepatic uptake was similarly related to NPA and afferent flux. By contrast, for total amino acids and L-lactate net hepatic uptake was better related to NPA than to afferent flux (Figure 1 v. Figures 2 and 3; Supplementary Material Table S4). As a result, the subsequent meta-analysis considered NPA as the  $X$  variable.

*Models.* The net hepatic uptake of propionate,  $\alpha$ -amino-N and L-lactate increased linearly with their net absorption in the portal vein ( $P < 0.01$ , Table 3). For propionate and  $\alpha$ -amino-N, average intercepts were not significantly different from zero. The linear slopes suggested that for 1 mmol/kg BW per h appearing in portal vein, 0.910, 0.749 and 0.474 mmol/kg BW per h of propionate,  $\alpha$ -amino-N (corresponding to a slope of 0.536 for total amino acids) and L-lactate were taken up by the liver, respectively.

*Interfering factors.* Table 3 reports the significant interfering factors and Supplementary Material Table S5 reports the linear relationships between LSMean and interfering factors. For propionate (model 1, Figure 2a and b) and at similar NPA of propionate (LSMean effect), the net hepatic uptake increased with the energy balance of the animals ( $P < 0.01$ ) and starch intake ( $P \leq 0.05$ ). For  $\alpha$ -amino-N (model 2, Figure 2c and d) and at similar NPA of  $\alpha$ -amino-N, net hepatic uptake increased ( $P > 0.05$ ) with dietary N concentration (CP and protein digestible in the intestine) and net hepatic release of glucose ( $P \leq 0.05$ ), and decreased with the net hepatic uptake of propionate ( $P \leq 0.05$ ). When introduced in the model as additional covariates, none of those factors significantly improved the fit. For L-lactate (model 3, Figure 3a and b) and at similar NPA of L-lactate, the net hepatic uptake was significantly affected by propionate uptake. Inclusion of propionate uptake as a covariate significantly improved the model, removing all interfering factors (model 4). To account for the role of lactate to recycle C, energy balance was also tested as an  $X$  variable, but did not improve the fit and showed significant interfering factors (model 5, Figure 3c).

### *Net hepatic release of glucose*

*Influence of the net portal absorption of glucose.* The net hepatic release of glucose was quadratically related ( $P < 0.05$ ) to its NPA (model 6, Table 4; Figure 3d and e), indicating that the net release of glucose decreased when NPA of glucose increased from  $-0.334$  up to 0 mmol/kg BW per h, and increased thereafter.

**Table 2** Description of the reported nutrient arterial concentrations, net hepatic fluxes<sup>a</sup> and potential contribution to gluconeogenesis in ruminants used for the meta-analyses according to physiological status

|   | Non-productive adults <sup>b</sup> |        |        |        |        | Growing animals       |        |        |        |        | Gestating animals     |        |        |        |        | Lactation <sup>c</sup> |        |        |        |        | Effect <sup>d</sup> |
|---|------------------------------------|--------|--------|--------|--------|-----------------------|--------|--------|--------|--------|-----------------------|--------|--------|--------|--------|------------------------|--------|--------|--------|--------|---------------------|
|   | <i>n</i> <sub>t</sub>              | Mean   | SD     | Min    | Max    | <i>n</i> <sub>t</sub> | Mean   | SD     | Min    | Max    | <i>n</i> <sub>t</sub> | Mean   | SD     | Min    | Max    | <i>n</i> <sub>t</sub>  | Mean   | SD     | Min    | Max    | <i>P</i> -value     |
| Arterial concentration (mM)                                     |                                    |        |        |        |        |                       |        |        |        |        |                       |        |        |        |        |                        |        |        |        |        |                     |
| Propionate  | 25                                 | 0.0355 | 0.0347 | 0.012  | 0.16   | 14                    | 0.0586 | 0.0147 | 0.031  | 0.08   | 5                     | 0.046  | 0.013  | 0.032  | 0.061  | 6                      | 0.075  | 0.027  | 0.039  | 0.096  | 0.011               |
| α-amino-N   | 45                                 | 4.35   | 0.89   | 2.86   | 7.19   | 25                    | 2.95   | 0.419  | 2.23   | 3.7    | 3                     | 2.71   | 0.042  | 0      | 4.09   | 2                      | 3.02   | 0.0071 | 3.02   | 3.03   | <0.001              |
| L-lactate   | 28                                 | 0.803  | 0.372  | 0.34   | 1.42   | 25                    | 0.574  | 0.148  | 0.364  | 1.1    | 5                     | 0.492  | 0.133  | 0.361  | 0.65   | 11                     | 0.418  | 0.194  | 0.194  | 0.756  | <0.001              |
| Glucose   | 48                                 | 3.27   | 0.535  | 1.89   | 4.56   | 25                    | 4.42   | 0.535  | 3.3    | 5.49   | 5                     | 3.57   | 0.106  | 3.47   | 3.74   | 14                     | 3.52   | 0.439  | 2.73   | 4.2    | <0.001              |
| Insulin, μU/L   | 7                                  | 38.9   | 23     | 18.4   | 71.9   | 6                     | 31.9   | 11.8   | 21.3   | 51.5   | 2                     | 14.5   | 2.17   | 13     | 16     | 4                      | 9.43   | 3.32   | 6.31   | 13.8   | 0.045               |
| Net portal appearance (mmol/kg BW per h)                        |                                    |        |        |        |        |                       |        |        |        |        |                       |        |        |        |        |                        |        |        |        |        |                     |
| Propionate  | 37                                 | 0.656  | 0.417  | 0.085  | 2.11   | 16                    | 0.8    | 0.21   | 0.47   | 1.16   | 11                    | 0.64   | 0.18   | 0.382  | 0.447  | 11                     | 1.16   | 0.29   | 0.827  | 1.89   | <0.001              |
| α-amino-N   | 61                                 | 0.597  | 0.408  | -0.01  | 2.71   | 29                    | 0.354  | 0.198  | 0.029  | 0.95   | 3                     | 0.251  | 0.229  | 0      | 0.447  | 2                      | 0.636  | 0.106  | 0.561  | 0.711  | <0.001              |
| L-lactate   | 41                                 | 0.274  | 0.166  | 0.084  | 0.767  | 25                    | 0.203  | 0.083  | 0.117  | 0.399  | 5                     | 0.117  | 0.0217 | 0.091  | 0.141  | 15                     | 0.236  | 0.0816 | 0.074  | 0.363  | 0.031               |
| Glucose   | 61                                 | -0.091 | 0.102  | -0.373 | 0.168  | 25                    | -0.05  | 0.098  | -0.234 | 0.16   | 5                     | -0.026 | 0.0193 | -0.044 | 0      | 23                     | -0.026 | 0.136  | -0.166 | 0.352  | <0.001              |
| Net hepatic flux (mmol/kg BW per h)                             |                                    |        |        |        |        |                       |        |        |        |        |                       |        |        |        |        |                        |        |        |        |        |                     |
| Propionate  | 35                                 | -0.562 | 0.292  | -1.1   | -0.085 | 16                    | -0.705 | 0.183  | -1.034 | -0.409 | 11                    | -0.591 | 0.186  | -0.892 | -0.254 | 11                     | -1.08  | 0.279  | -1.69  | -0.757 | <0.001              |
| α-amino-N   | 63                                 | -0.422 | 0.203  | -0.923 | 0.436  | 23                    | -0.378 | 0.376  | -1.3   | -0.033 | 3                     | -0.23  | 0.237  | -0.474 | 0      | 2                      | -0.274 | 0.086  | -0.335 | -0.213 | <0.001              |
| L-lactate   | 41                                 | -0.356 | 0.346  | -1.94  | 0.029  | 27                    | -0.189 | 0.131  | -0.399 | -0.008 | 5                     | -0.079 | 0.126  | -0.297 | 0      | 17                     | -0.271 | 0.158  | -0.48  | -0.107 | 0.48                |
| Glucose   | 69                                 | 0.67   | 0.31   | 0.245  | 1.74   | 27                    | 0.551  | 0.161  | 0.317  | 0.975  | 5                     | 0.34   | 0.193  | 0      | 0.463  | 23                     | 0.995  | 0.313  | 0.467  | 1.405  | <0.001              |
| Potential contribution to gluconeogenesis (%)                   |                                    |        |        |        |        |                       |        |        |        |        |                       |        |        |        |        |                        |        |        |        |        |                     |
| Propionate  | 35                                 | 41.6   | 20.2   | 12.3   | 99.7   | 16                    | 66.5   | 13.3   | 46.3   | 94.7   | 4                     | 49.7   | 13.5   | 34     | 65.5   | 11                     | 60.9   | 10.1   | 49.4   | 81.1   | <0.001              |
| α-amino-N   | 45                                 | 46.3   | 29.3   | 0      | 180.2  | 17                    | 30.2   | 16.7   | 6.94   | 82.1   | 2                     | 19.2   | 27.2   | 0      | 38.4   | 2                      | 16.4   | 1.36   | 15.4   | 17.3   | 0.061               |
| L-lactate   | 41                                 | 31.9   | 30.8   | 0      | 149    | 27                    | 17.6   | 11.8   | 0.608  | 44.3   | 4                     | 12.6   | 18.4   | 0      | 39.8   | 17                     | 15.2   | 10     | 0      | 42.8   | 0.018               |
| Estimated <sup>e</sup> mobilized nutrients (mmol/kg BW per day) |                                    |        |        |        |        |                       |        |        |        |        |                       |        |        |        |        |                        |        |        |        |        |                     |
| Mobilized glycerol  | 18                                 | 1.68   | 1.26   | 0.058  | 4.31   | 0                     | ND     | ND     | ND     | ND     | 2                     | 2.91   | 2.55   | 1.11   | 4.72   | 8                      | 1.066  | 0.64   | 0.092  | 1.93   | 0.159               |
| Mobilized alanine   | 18                                 | 0.097  | 0.073  | 0.0034 | 0.249  | 0                     | ND     | ND     | ND     | ND     | 2                     | 0.169  | 0.148  | 0.064  | 0.273  | 8                      | 0.062  | 0.04   | 0.0053 | 0.112  | 0.159               |

Min = minimum value; Max = maximum value; *n*<sub>t</sub> = number of treatments; ND = not determined.

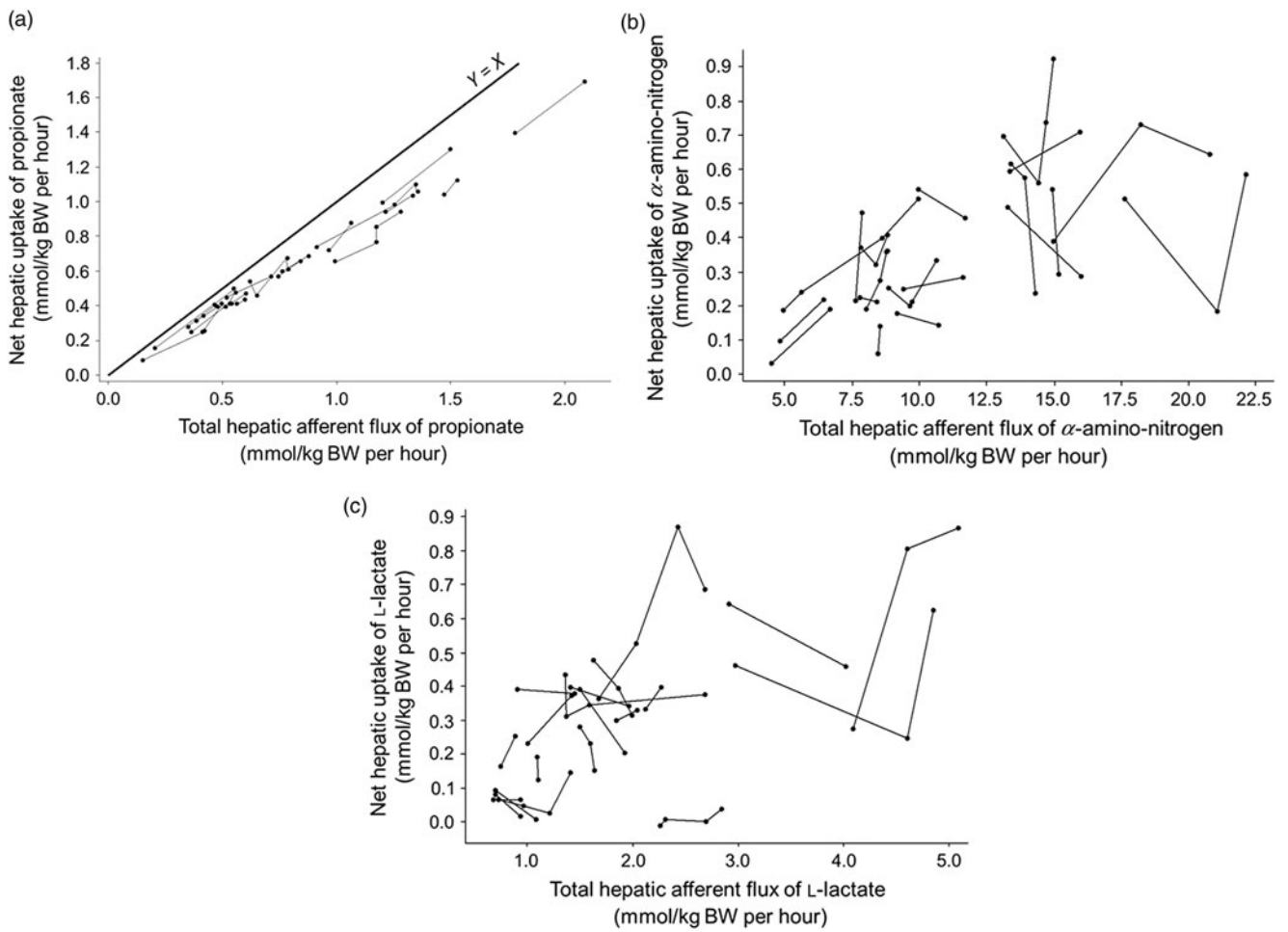
<sup>a</sup>A positive value indicates a net release; a negative value indicates a net uptake.

<sup>b</sup>Fasting included.

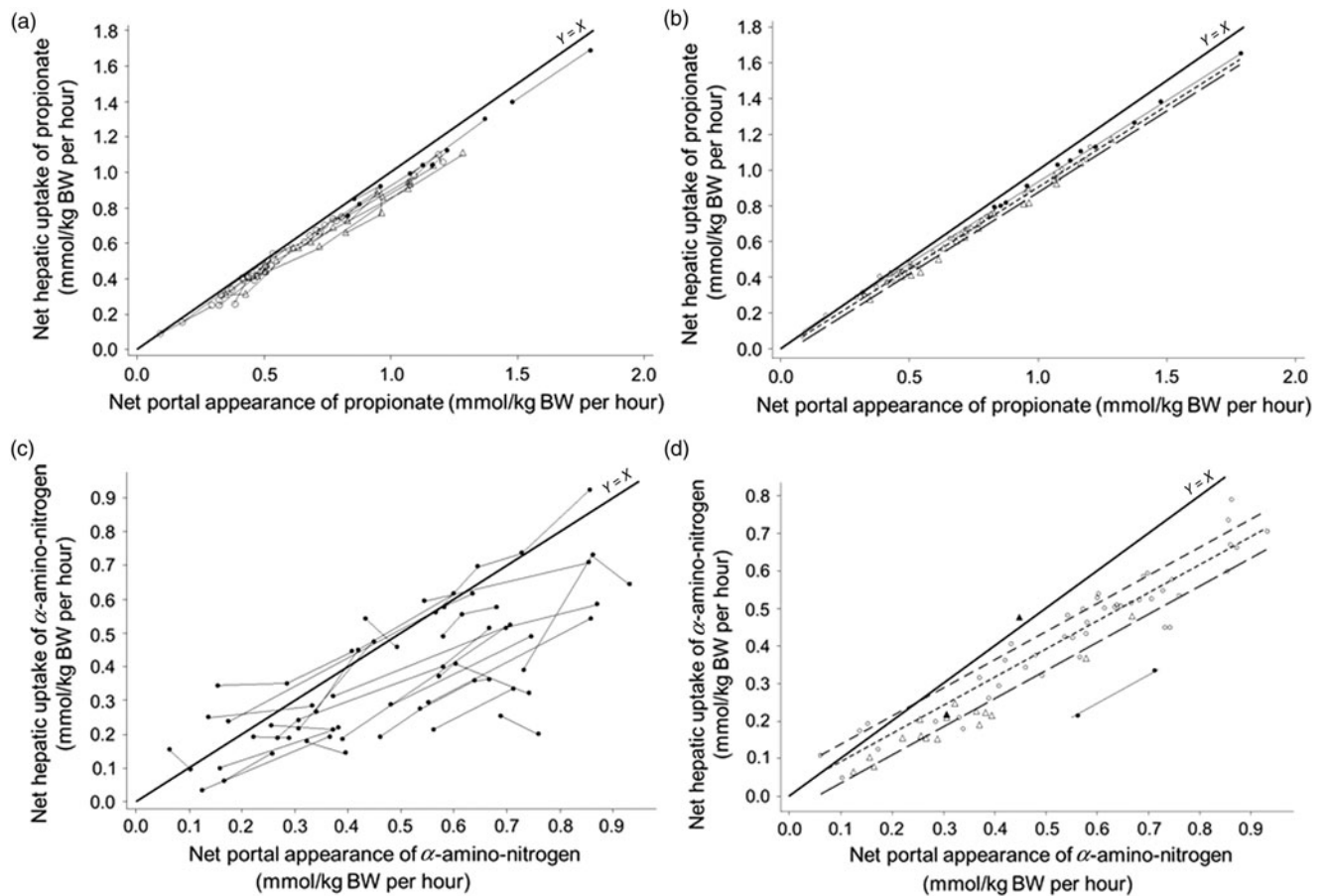
<sup>c</sup>The dairy cattle were between 11 and 240 days in milk.

<sup>d</sup>Physiological status effect.

<sup>e</sup>Values are reported only for animals in negative energy balance.

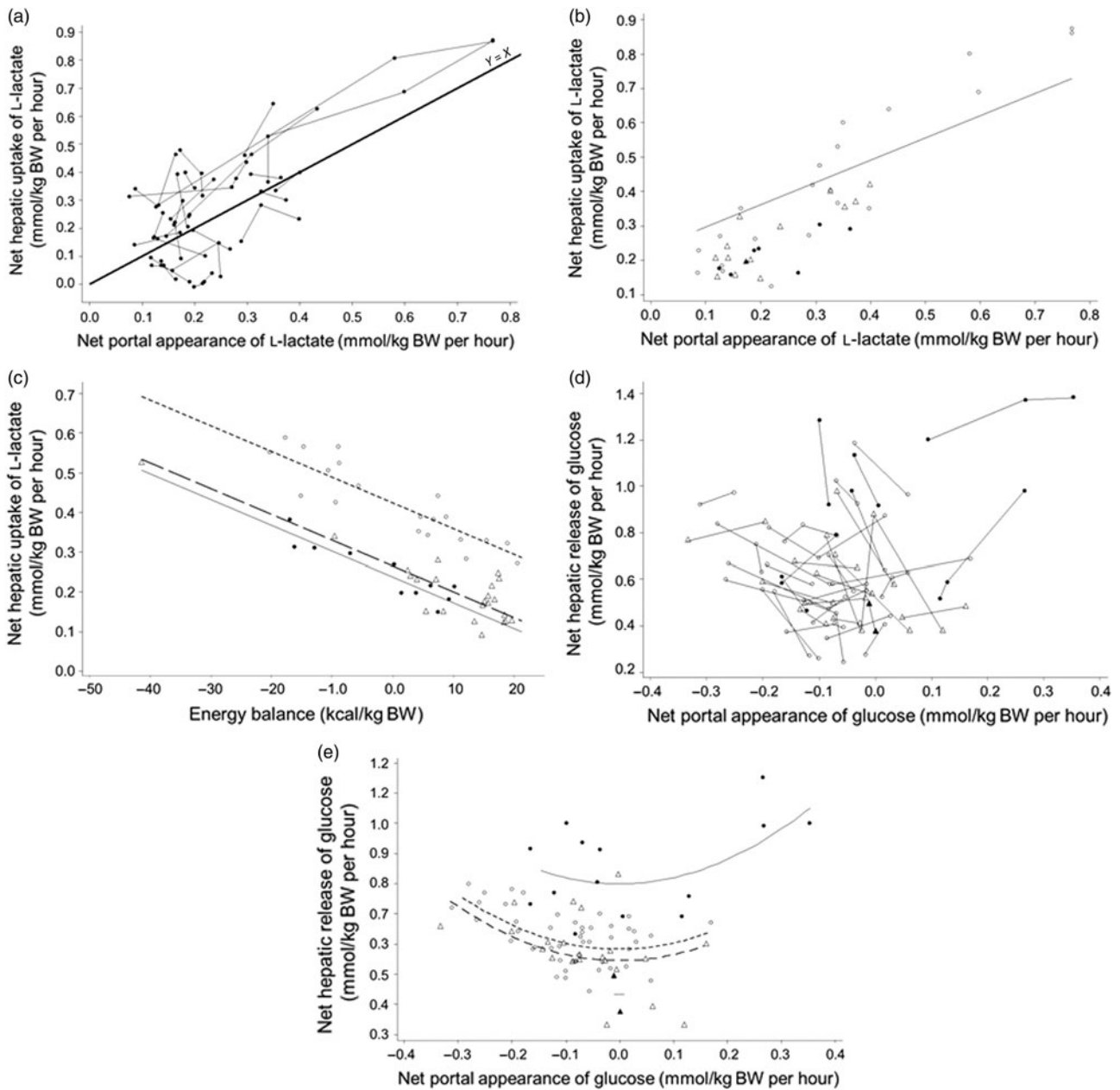


**Figure 1** Within-study relationships between the net hepatic uptake of propionate (a),  $\alpha$ -amino-N (b) and L-lactate (c) and their total hepatic afferent flux in ruminants. The propionate (a),  $\alpha$ -amino-N (b) and L-lactate (c) datasets were used.



**Figure 2** Within-study relationships between net hepatic uptake and net portal appearance of propionate (a for raw data and b for adjusted model) and  $\alpha$ -amino-N (c for raw data and d for adjusted model) in ruminants. The propionate (a, b) and  $\alpha$ -amino-N (c, d) datasets were used. Adjusted models are shown for non-productive adults ( $\circ$ ), growing animals ( $\triangle$ ), lactating cattle ( $\bullet$ ) and gestating animals ( $\blacktriangle$ ).





**Figure 3** Within-study relationships between net hepatic uptake of L-lactate and net portal appearance of L-lactate (a for raw data, and b for adjusted model), and energy balance (c for adjusted model); and between net hepatic release of glucose and its net portal appearance (d for raw data, e for adjusted model) in ruminants. The L-lactate (a, b, c) and glucose (d, e) datasets were used. Adjusted models are shown for non-productive adults ( $\circ$ ), growing animals ( $\triangle$ ), lactating cattle ( $\bullet$ ) and gestating animals ( $\blacktriangle$ ).

*Influence of the hepatic uptake of each precursor.* Generally, the net hepatic release of glucose increased significantly and linearly with the availability of each precursor considered separately, in the portal vein (Table 4, models 7, 9, 11) and taken up by the liver (Table 4, models 8, 10, 12). Intercepts were generally significantly different from zero, indicating that glucose release also depends on other factors. At zero NPA of total amino acids, the uptake of total amino acids by the liver was higher with productive animals (growing and lactating) *v.* non-productive animals ( $P < 0.001$ , Table 4, model 9). At similar NPA of propionate (model 7, LSMeans effect,  $P = 0.001$ ) the net hepatic release of glucose

increased when the hepatic uptake of amino acids decreased, and when the hepatic release of  $\beta$ -hydroxybutyrate increased. No improvement of fit was gained when these factors were added in the model.

*Influence of the hepatic uptake of the sum of precursors.* When the all-precursor and the propionic infusion datasets were combined to evaluate the glucose response over a wide range of precursor supply, glucose release was curvilinearly related to precursor availability (Figure 4). In this combined dataset, the NPA of summed precursors ranged from 1.003 to 13.05 mmol C/kg BW per h (Figure 4a and b),

**Table 3** Response equations of the net hepatic fluxes (NHFs, mmol/kg BW per h) of propionate (C3),  $\alpha$ -amino-nitrogen ( $\alpha$ N) and L-lactate to variations in their net portal appearance (NPA, mmol/kg BW per h) and energy balance (EB, kcal/kg BW per day) in ruminants

| Model number       | Y           | Number <sup>a</sup> |                |       |       | Equation   | Adjustment |                | Effect <sup>b</sup> |        | Interfering factors <sup>c</sup>               |
|--------------------|-------------|---------------------|----------------|-------|-------|--|------------|----------------|---------------------|--------|--|
|                    |             | $n_{exp}$           | $n_{\alpha i}$ | $n_t$ | $n_r$ |  | RMSE       | Adjusted $R^2$ | PHY                 | X* PHY | LSMeans  |
| Net hepatic fluxes |             |                     |                |       |       |  |            |                |                     |        |  |
| 1                  | Propionate  | 27                  | 2              | 69    | 0     | $-0.0035 \pm 0.022^{NS} - 0.910 \pm 0.028^{***} \times \text{NPA-C3}$  | 0.028      | 0.992          | <0.001              | NS     | EB**; starch intake*                           |
| 2                  | $\alpha$ -N | 26                  | 11             | 65    | 3     | $0.0413 \pm 0.0353^{NS} - 0.749 \pm 0.068^{***} \times \text{NPA-}\alpha\text{N}$  | 0.062      | 0.898          | 0.001               | NS     | +CP <sup>†</sup> ; +PDI*; -NHF-C3*; +NHF-gluc* |
| 3                  | L-Lactate   | 25                  | 14             | 66    | 7     | $-0.0151 \pm 0.0430^{**} - 0.474 \pm 0.20^* \times \text{NPA-L-lactate}$   | 0.067      | 0.892          | NS                  | 0.058  | +NPA-gluc**; -NHF-C3*; +PDI*; +dOM**; +ME*     |
| 4                  |             | 18                  | 4              | 43    | 0     | $-0.2195 \pm 0.0621^{**} + 0.218 \pm 0.080^{**} \times \text{NPA-C3}$<br>$- 1.029 \pm 0.096 \times \text{NPA-L-lactate}$ | 0.060      | 0.916          | NS                  | NS     | None   |
| 5                  |             | 25                  | 17             | 54    | 4     | $-0.3090 \pm 0.0091^{***} + 0.00648 \pm 0.00142^{***} \times \text{EB}$  | 0.057      | 0.908          | 0.001               | NS     | +NPA-gluc**; -NPA- $\alpha$ N**                |

NS = not significant ( $P > 0.10$ ); RMSE = residual means square error; X = explanatory variable; LSMeans = least-squares means; EB = energy balance; PDI = protein digestible in the intestine; gluc = glucose; dOM = dietary concentrations of digestible organic matter; ME = metabolizable energy.

All models were based on reported measured fluxes.

<sup>†</sup> $P < 0.10$ ; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ .

<sup>a</sup> $n_{exp}$ : number of experimental groups in the model;  $n_{\alpha i}$ : number of experimental groups with  $\alpha i$  (intercept for the experimental group  $i$ ) significantly different from zero;  $n_t$ : number of treatments in the model;  $n_r$ : number of treatments rejected from the model (outliers).

<sup>b</sup>PHY: physiological status, effects of physiological status were detected on the intercept (Model 1,  $\Delta = -0.0005$  (NS), 0.032,  $-0.026$  ( $P = 0.06$ ),  $-0.0061$  (NS); Model 2,  $\Delta = -0.0584$ ,  $-0.0009$  (NS), 0.161,  $-0.104$ ; Model 5,  $\Delta = -0.1149$ , 0.0442, 0.0707, ND), on the slope (Model 3,  $\Delta = -0.507$ , 0.333 (NS),  $-0.174$  (NS), ND) for the non-productive adults, growing animals, lactation and gestation, respectively.

<sup>c</sup>No interfering factors were observed on individual slopes and residuals; these are interfering factors on the LSMeans: the minus sign indicates a negative effect on the net hepatic flux; the plus sign indicates a positive effect on the net hepatic flux.

**Table 4** Response equations of the net hepatic release of glucose (Y variable) to the following X variables: net portal appearance (NPA) and net hepatic flux (NHF) of propionate (C3), total amino acids (tAA), L-lactate, glucose and sum of precursors (mmol C/kg BW per h) in ruminants

| Model number | Predictors                  |                            | Numbers <sup>a</sup> |                 |                |                | Intercept            |       | Slope for X1       |       | Slope for X2 |       | RMSE  | R <sup>2</sup> <sub>adj</sub> | Effects <sup>b</sup> |        | Interfering factor <sup>c</sup> |
|--------------|-----------------------------|----------------------------|----------------------|-----------------|----------------|----------------|----------------------|-------|--------------------|-------|--------------|-------|-------|-------------------------------|----------------------|--------|---------------------------------|
|              | X1                          | X2                         | n <sub>exp</sub>     | n <sub>ai</sub> | n <sub>t</sub> | n <sub>r</sub> | α                    | SD    | β1                 | SD    | β2           | SD    |       |                               | PHY                  | X*PHY  | LSMeans                         |
| 6            | NPA-glucose                 | (NPA-glucose) <sup>2</sup> | 42                   | 16              | 93             | 0              | 0.591***             | 0.028 | NS                 |       | 2.0096*      | 0.826 | 0.13  | 0.725                         | <0.001               | NS     | None                            |
| 7            | NPA-C3                      |                            | 26                   | 8               | 57             | 0              | 2.259***             | 0.486 | 0.714***           | 0.185 | NS           | ND    | 0.532 | 0.848                         | <0.001               | NS     | +NHF-BHBA**; +NHF-αN**          |
| 8            | NHF-C3                      |                            | 27                   | 6               | 59             | 0              | 1.856***             | 0.66  | -0.923*            | 0.426 | NS           | ND    | 0.494 | 0.869                         | NS                   | 0.058  | None                            |
| 9            | NPA-tAA <sup>d</sup>        |                            | 26                   | 12              | 61             | 2              | 0.309 <sup>NS</sup>  | 0.733 | 1.288***           | 0.254 | NS           | ND    | 0.405 | 0.899                         | <0.001               | <0.001 | None                            |
| 10           | NHF-tAA <sup>d</sup>        |                            | 21                   | 8               | 52             | 0              | 2.134**              | 0.691 | -1.163**           | 0.404 | NS           | ND    | 0.694 | 0.75                          | 0.008                | 0.006  | None                            |
| 11           | NPA-L-lactate               |                            | 34                   | 10              | 72             | 0              | 2.403***             | 0.424 | 2.062**            | 0.66  | NS           | ND    | 0.794 | 0.698                         | <0.001               | NS     | +ME intake *                    |
| 12           | NHF-L-lactate               |                            | 35                   | 15              | 76             | 1              | 4.214***             | 0.207 | 0.537 <sup>†</sup> | 0.284 | NS           | ND    | 0.654 | 0.835                         | <0.001               | NS     | None                            |
| 13           | NPA-precursors <sup>e</sup> |                            | 39                   | 14              | 89             | 6              | -0.708 <sup>NS</sup> | 0.727 | 0.809***           | 0.112 | NS           | ND    | 0.424 | 0.896                         | <0.001               | <0.001 | NHF-BHBA*                       |
| 14           | NPA-precursors <sup>e</sup> | NPA-glucose                | 34                   | 12              | 82             | 0              | 1.364***             | 0.334 | 0.636**            | 0.094 | -0.613**     | 0.168 | 0.503 | 0.855                         | 0.001                | NS     | None                            |
| 15           | NHF-precursors <sup>e</sup> |                            | 30                   | 16              | 71             | 6              | -0.384 <sup>NS</sup> | 0.531 | -0.905***          | 0.096 | NS           | ND    | 0.375 | 0.925                         | <0.001               | <0.001 | None                            |

ND = not determined; NS = not significant ( $P > 0.10$ ); RMSE = residual means square error; LSMeans = least-squares means; BHBA = beta-hydroxybutyrate; ME = metabolizable energy; αN = α-amino-nitrogen.

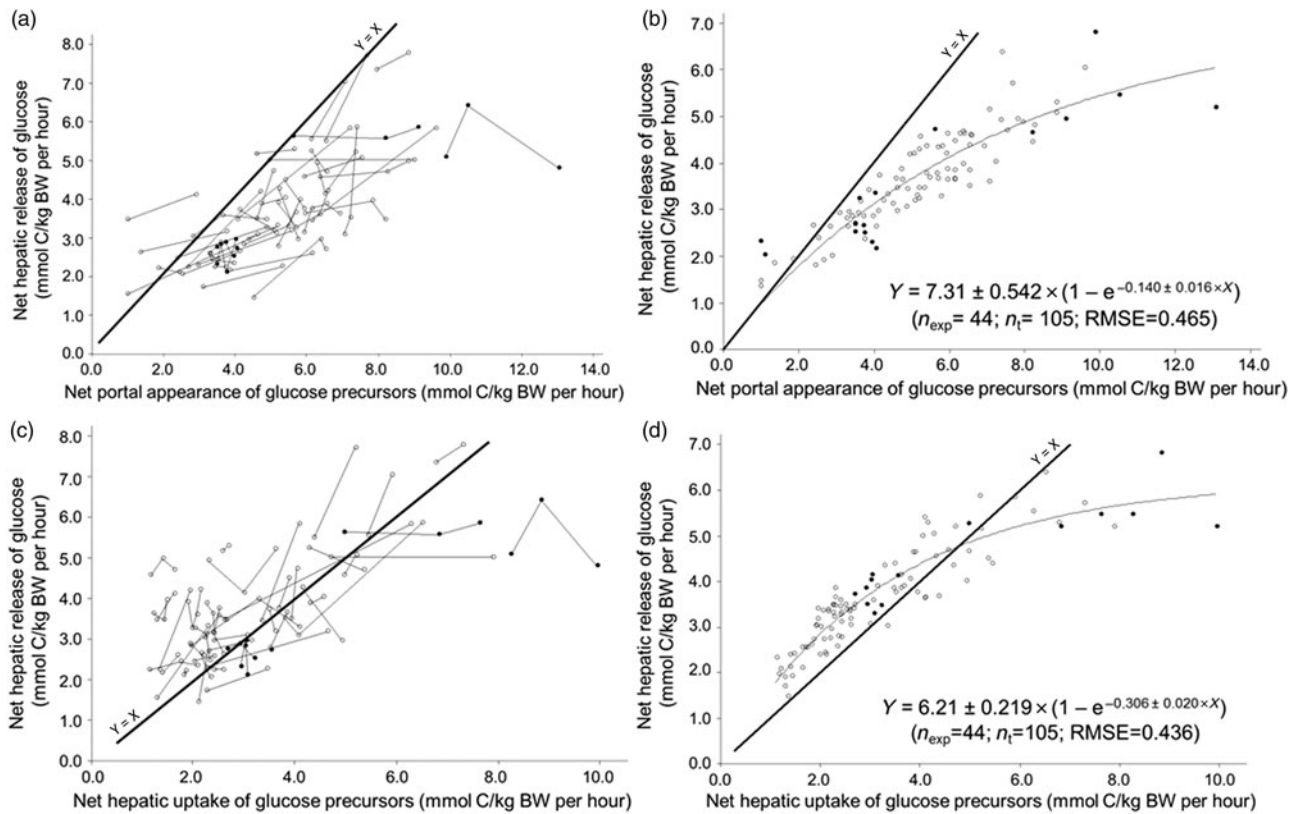
<sup>†</sup> $P < 0.10$ ; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ .

<sup>a</sup>n<sub>exp</sub>: number of experimental groups in the model; n<sub>ai</sub>: number of experimental groups with αi significantly different from zero; n<sub>t</sub>: number of treatments in the model; n<sub>r</sub>: number of treatments rejected from the model (outliers).  
<sup>b</sup>PHY = physiological state effect, effects of physiological status were detected on the intercept (Model 6, Δ = -0.007 (NS), -0.045 (NS), 0.210, -0.158; Model 7, Δ = 0.276, -0.773, 0.497; Model 9, Δ = 2.336, 0.317 (NS), -3.023 (NS), 0.369 (NS); Model 10, Δ = 1.533, -0.583 (NS), -0.457, -0.492 (NS); Model 11, Δ = -0.907, -0.092 (NS), 1.512, -0.512 (NS); Model 12, Δ = -0.286, -0.345, 2.047, -1.416; Model 13, Δ = 2.676, 0.984 (NS), -3.661, ND; Model 14, Δ = -0.199 ( $P = 0.1$ ), -0.341, 0.540, ND; Model 15, Δ = 2.478, -0.862, -1.615, ND), on the slope (Model 8, Δ = 0.237 (NS), 1.308, -0.934 (NS), -0.610 (NS); Model 9, Δ = -0.908, -0.059 (NS), 1.342, -0.374 (NS); Model 10, Δ = 1.157, 0.247 (NS), -2.070, 0.666 (NS); Model 13, Δ = -0.511, -0.129 (NS), 0.640, ND; Model 15, Δ = 0.573, -0.289, -0.285, ND) for the non-productive adults, growing animals, lactation and gestation, respectively.

<sup>c</sup>No interfering factors were observed on individual slopes and residuals: the minus sign indicates a negative effect on the net hepatic flux; the plus sign indicates a positive effect on the net hepatic flux.

<sup>d</sup>tAA fluxes calculating from α-amino-nitrogen fluxes according to Martineau *et al.* (2009).

<sup>e</sup>Percentage of estimated precursors : 30.6%, 29.6% and 31%, respectively, for models 13, 14 and 15.



**Figure 4** Within-study curvilinear relationships between net hepatic release of glucose and availability of summed precursors expressed as net portal appearance (a for raw data, b for adjusted model) and as net hepatic uptake (c for raw data, d for adjusted model) in ruminants. The combined all-precursor and propionic infusion datasets were used. Number of experimental groups ( $n_{exp}$ ) and number of treatments ( $n_t$ ) are given.

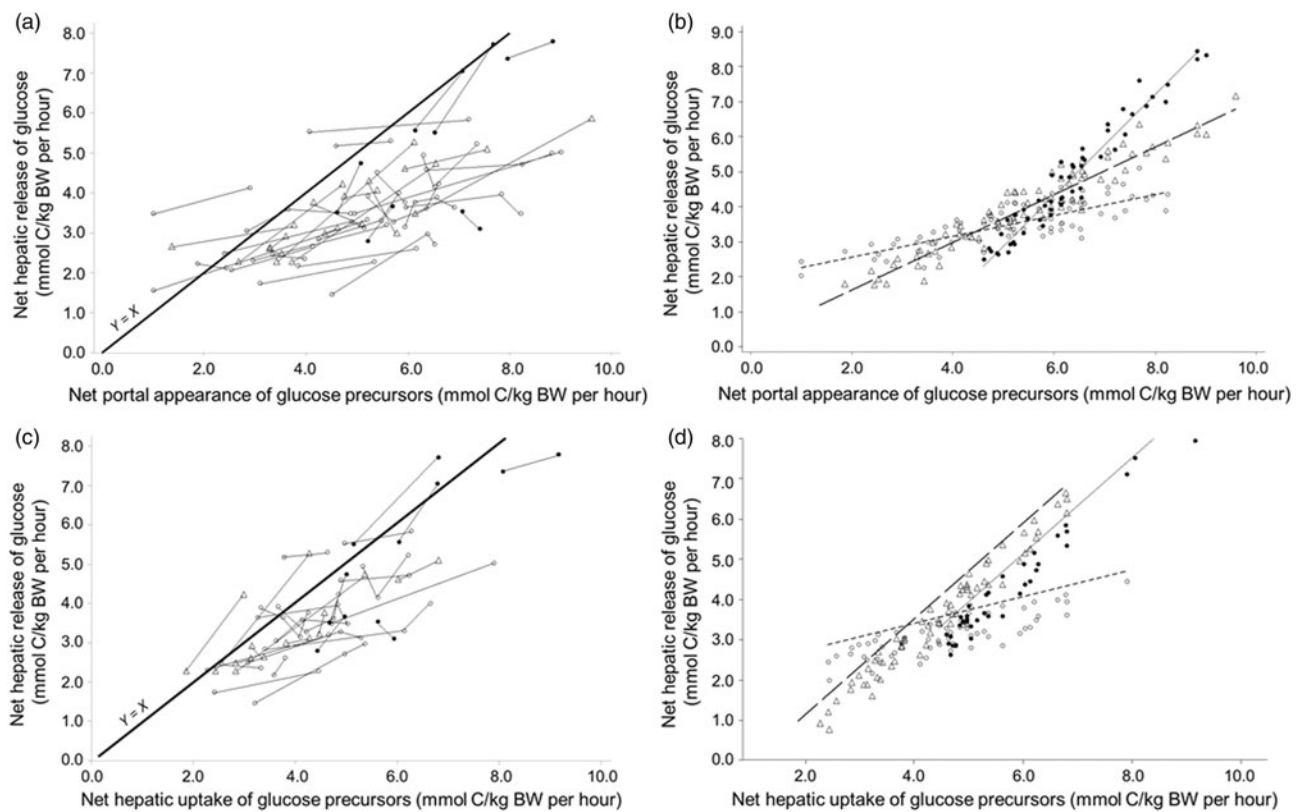
while net hepatic uptake ranged from 1.098 to 9.944 mmol C/kg BW per h (Figure 4c and d). At similar precursor availability (LSMeans effect on both models), glucose release was increased when the net hepatic release of  $\beta$ -hydroxybutyrate increased ( $P < 0.05$ ).

Another meta-analysis was conducted on the all-precursor dataset only, excluding the propionate infusion studies, to explore the more linear portion of the relationship. The NPA of summed precursors ranged from 0.994 to 9.69 mmol C/kg BW per h and was closer to the physiological range. A linear model (quadratic model not significant, results not shown) was applied and the net hepatic release of glucose increased by 0.809 mmol C/kg BW per h for each increment of the NPA of summed precursors (Table 4, model 13; Figure 5a and b) with a good adjustment (RMSE represented 13% of average release of glucose by the liver). The intercept was not different from zero ( $P < 0.001$ ) suggesting no net glucose release in the absence of precursor supply. Both the intercept and the slope were significantly affected by physiological status indicating a higher net release of glucose from the same net portal absorption of precursors for lactating animals. At similar NPA of precursors (LSMeans effect), glucose release was increased when hepatic emission of  $\beta$ -hydroxybutyrate increased. Interestingly when the NPA of glucose was added as covariate in the model (Table 4, model 14), it had a negative impact (slope effect) of the net hepatic release of glucose. Finally, the net hepatic release of glucose increased

linearly by 0.905 mmol C/kg BW per h for each increment in net hepatic uptake of precursors (in mmol C/kg BW per h; Table 4, model 15, Figure 5c and d), with a good adjustment (RMSE represented 10% of average release of glucose by the liver). Both the intercept and the slope were significantly affected by physiological status, indicating a higher conversion rate of precursors into glucose for productive animals (growing and lactating) v. non-productive adults.

#### Net hepatic insulin fluxes

The insulin dataset was limited to  $n = 19$  treatments. Arterial concentrations ranged from 6.3 to 71.9  $\mu$ UI/ml and increased (intra-study slope) by  $3.04 \pm 0.36 \mu$ UI/ml per g DMI per kg BW per day ( $P = 0.004$ ). The NPA of insulin also increased with intake (expressed in g/kg BW per day) by  $1.31 \pm 0.25 \mu$ UI/kg BW per h ( $P = 0.001$ ,  $n_t = 17$ ) per g DM intake, by  $9.81 \pm 1.61 \mu$ UI/kg BW per h per g of starch digested in the small intestine ( $P = 0.01$ ,  $n_t = 8$ ) and by  $8.02 \pm 1.43 \mu$ UI/kg BW per h per g NDF digested in the rumen ( $P = 0.001$ ,  $n_t = 17$ ). Moreover, the NPA of insulin tended to increase with the portal – arterial concentration difference (in mM) of propionate (slope =  $44.48 \pm 20.16$ ;  $P = 0.11$ ,  $n_t = 8$ ) and acetate (slope =  $10.82 \pm 4.43$ ;  $P = 0.09$ ,  $n_t = 8$ ), but not glucose ( $P = 0.23$ ). The net hepatic uptake of insulin increased with the NPA of insulin (slope =  $0.49 \pm 0.14$ ,  $P = 0.01$ ,  $n_t = 17$ ), the portal – arterial concentration difference of propionate



**Figure 5** Within-study linear relationships between net hepatic release of glucose and availability of summed precursors expressed as net portal appearance (a for raw data, b for adjusted model) and as net hepatic uptake (c for raw data, d for adjusted model) in ruminants. The all-precursor dataset was used. Adjusted models are shown for non-productive adults (○), growing animals (△), lactating cattle (●) and gestating animals (▲).

(slope =  $57.94 \pm 6.60$ ,  $P = 0.01$ ,  $n_t = 8$ ) and with the NPA of glucose (slope =  $27.65 \pm 9.96$ ,  $P = 0.05$ ,  $n_t = 16$ ).

## Discussion

The present work substantiates the use of the NPA, instead of total afferent fluxes, as the predictor for the net hepatic uptake of propionate,  $\alpha$ -amino-N and L-lactate in a database like the one used in the current study where within-study variations were mainly caused by dietary changes and where lactating cow data were obtained at mid-lactation. The higher the contribution of NPA to the afferent flux, the lower the difference in fit between the two predictors. Obviously, the relevance of using the NPA as predictor depends on the nutrient and on the range of nutritional situations under study. As a result, for propionate,  $\alpha$ -amino-N and to a lower extent for L-lactate their net hepatic uptakes follow mass action laws, with NPA as the main statistical driving force. The statistical driving force may differ from the biological one. This is especially the case for the amino acid fluxes, which are more accurate than the  $\alpha$ -amino-N ones, and for animals that are not representative of this dataset. Indeed, when comparing cows pre- and post-calving, it was clear that increased net portal absorption of amino acids post-calving accompanying increased DMI was not related to increased hepatic net removal, the latter declining in response to decreased arterial

amino acids concentrations related to the initiation of lactation (Doepel *et al.*, 2009). Also as a consequence of this statistical choice, endogenous L-lactate availability from mobilized tissues, as in early lactation, is not accounted for.

The within-study approach which was applied here focuses on incremental responses. It is testing the hypotheses that marginal changes in the X variables are inducing marginal changes in the Y variables of interest. This same strategy had also been applied to predict the net hepatic release of ketogenic nutrients by Loncke *et al.* (2015). As a result, the study of the incremental supply of nutrients, which is primarily driven by daily intake and the related gut metabolism, is dissociated from the baseline levels of nutrient fluxes, which also depend on animal characteristics. It is compatible with the static models of feed evaluation systems which calculate daily rations for animals, irrespective of the amount of nutrients already circulating in the animal from previous rations as developed in Ortigues-Marty *et al.* (2019).

### Domain of application of the models

Empirical prediction models apply to a strict range of validity. Because available data were balanced between cattle and sheep, models apply to both species. Species or physiological status significantly affected models of the present work, but as in Loncke *et al.* (2015) it was not always possible to evaluate whether it was a strict species or physiological status

effect, or if there was some confounding with nutritional conditions. These effects are discussed below for each nutrient. In all cases, using physiological status as a covariate was more discriminant than using species as in Loncke *et al.* (2015) for ketogenic nutrients. This covariate also encompasses differences in the level of production. As for other models developed from the same Flora database (Loncke *et al.*, 2009b and 2015), models apply to animals with intakes up to 41 g DMI/kg BW per day of diets varying from 0 to 100 g concentrate/100 g DM. They do not account for any feed additive effects (buffers, essential oils, ionophores, etc.) nor for lipid supplementation.

Out of the analytical methods considered, only pAH analysis and hence blood flow evaluation had a significant quantitative influence on the relationships (Supplementary Material S3). Acetylation of pAH was shown to result in overestimation of nutrient release and underestimation of nutrient uptake by Larsen and Kristensen (2013) and Rodriguez-Lopez *et al.* (2014), except for amino acids uptake which was reduced. When considering marginal rates instead of absolute values, both release and uptake rates were overestimated by liver acetylation of pAH. Despite these effects, we decided not to correct nutrient fluxes in order not to introduce further uncertainties to the data besides those inherent to the measurements (Rodriguez-Lopez *et al.*, 2016).

#### *Hepatic uptake of individual precursors*

**Propionate.** Results confirm the high hepatic uptake of propionate (Armentano, 1992). In the propionate dataset, no data exceeded the extraction capacity of the liver, propionate supply remained below levels obtained with infusions (e.g. Berthelot *et al.*, 2002) and portal propionate concentrations remained inferior to 1 mM. The intercept, not different from zero, was consistent with zero uptake in the absence of ruminal fermentations. The lowest hepatic uptake of propionate noted in growing animals was compatible with the known inhibitory effect of butyrate on propionate extraction (Berthelot *et al.*, 2002), as in the dataset hepatic supply and uptake of butyrate were elevated (Loncke *et al.*, 2015).

**L-Lactate.** Hepatic uptake of L-lactate was related to two different types of predictors. First, amounts of L-lactate and propionate appearing in the portal vein, which reflect dietary nutrient supply and synthesis of L-lactate from glucose in gut tissues (van der Walt *et al.*, 1983) that increases at high propionate levels (Harmon *et al.* 1991). Considering NPA of propionate as an additional predictor significantly improved the model (model 4 *v.* 3) and removed all interfering factors. We interpreted this improvement as the result of high propionate availability in the current dataset, and subsequently an increased hepatic conversion of propionate into L-lactate as shown *in vitro* by Demigné *et al.* (1991).

Second, the energy balance which reflects the nutritional status of the animal and the recycling of C through the Cori cycle when animals are in tissue mobilization (Reynolds, 2005). This predictor is relevant, even if there

was no gain in precision probably because of uncertainties of estimations. It did not fully eliminate the effect of physiological status suggesting an effect of physiological status per se, as reported, for example, in transition cows with an increase in L-lactate removal by the liver and in its potential contribution to liver glucose synthesis immediately after calving (Benson *et al.*, 2002; Reynolds *et al.*, 2003).

**$\alpha$ -amino-N.** The statistical mass action law which describes the impact of the portal appearance of  $\alpha$ -amino-N on its net hepatic uptake (also observed by Lescoat *et al.* (1996) and Reynolds (2006)) strictly applies to  $\alpha$ -amino-N as a proxy of the summed amino acids and does not apply to individual amino acids whose uptake is more related to their total afferent flux (Doepel *et al.*, 2009). Interestingly, the physiological and probably the underlying nutritional status of the animal (species, diet composition and intake, and level of production) influenced the baseline uptake (intercept) and not the marginal uptake rate (slope). Baseline net hepatic uptake (intercept) was higher in sheep than in cattle, probably reflecting lower production levels and lower tissue amino acid requirements (Blouin *et al.*, 2002; Reynolds, 2006). In a meta-analysis with a different dataset from the same Flora database, Martineau *et al.* (2011) attributed the higher recovery of ingested nitrogen in the portal vein of sheep *v.* cattle to a strict species effect. It was assumed to result from lower amino acid losses through portal-drained viscera metabolism in sheep or from a more efficient nitrogen recycling at lower nitrogen intake (Martineau *et al.*, 2011 and 2014). Our results also showed that the average level of  $\alpha$ -amino-N hepatic uptake (LSMeans) can be modified by the dietary N concentration, availability in propionate and hepatic glucose release. Direct experimental evidence was reported by Kraft *et al.* (2011) who showed that a N deficient diet significantly limited the net hepatic uptake of  $\alpha$ -amino-N, as a sparing mechanism, without modifying net glucose release (Loncke *et al.*, 2009a).

#### *Hepatic conversion of precursors into glucose*

An interesting outcome of this work is to show that the net hepatic release of glucose increases linearly with the availability of summed precursors (portal appearance or hepatic uptake) up to approximately 6 mmol C/kg BW per h, beyond which the conversion rate decreases. In the present dataset, the highest levels of precursor supply were met by intra-gastric infusion of individual nutrient (e.g. Majdoub *et al.*, 2003). Whether it is the high level of precursor supply or the imbalanced nutrient supply (as in Kraft *et al.*, 2011 with N deficient diets) which is responsible for this plateau would remain to be confirmed. This reduced efficiency of hepatic conversion of precursors into glucose suggests a supply of precursors in excess of glucose demand.

When considering a smaller range of summed precursor (i.e. excluding propionate infusion studies only) availability, the precursor C was converted to glucose at a rate (slope) ranging from 0.64 to 0.81 fold its net portal supply or

0.91 fold its hepatic uptake. Conversion rates remained below 1 and were subject to the influence of physiological status probably indicating some regulation in the conversion of precursors into glucose by the demand for glucose as discussed by Lapierre *et al.* (2010). At equal NPA of precursors (LSMean effect), the net release of glucose by the liver was higher for dairy cattle and lower for non-productive and growing animals, confirming the higher whole body glucose turnover rates observed for high *v.* low producing animals fed at similar metabolizable energy intake (Wilson *et al.*, 1983; Ortigues-Marty *et al.*, 2003). The impact of physiological status was also noted on the conversion rate of propionate into glucose (slope) which was highest for dairy cattle and lowest for growing animals (effect on intercept) as previously pointed out (Reynolds *et al.*, 2003).

Our results showed that for a same supply of precursors, the conversion of precursor C into glucose (LSMeans effect on model 13) and the conversion of portally absorbed propionate into glucose (model 7) increase when  $\beta$ -hydroxybutyrate uptake increases. They suggest increased glucose release when body reserves are mobilized as suggested by Loncke *et al.* (2009b), that is, animals in negative energy balance are more efficient in synthesizing glucose from precursors than when in positive energy balance. An increased conversion rate of precursors into glucose can be hypothesized because no increased contribution of endogenous precursors is expected as discussed above and reviewed by Larsen and Kristensen (2013).

Current results clearly show that bypass starch will limit the hepatic release of glucose reducing the conversion of precursors into glucose. The general relationship with precursor supply as well as the curvilinear relationship with NPA of glucose clearly demonstrated it. The supply of 117 g of bypass starch, equivalent to an NPA of 100 mmol glucose, will decrease the net hepatic release of glucose by 61 mmol of glucose, implying a substitution rate of 61% (model 14). Two mechanisms may be considered. First, starch intake also provides additional propionate, and the higher the propionate supply, the lower the propionate conversion rate into glucose as discussed above and the lower the net hepatic uptake of other precursors. Second, insulin/glucagon regulations might be involved. At negative NPA of glucose, that is, when bypass starch intake is null or low (lower than  $\approx 1.7$  g/kg BW per day, or lower than  $\approx 200$  g starch/kg DM, Loncke *et al.*, 2009c), precursor (mostly propionate; Majdoub *et al.*, 2003) absorption enhances hepatic insulin clearance and thereby the inhibitory effect of insulin on gluconeogenesis, possibly by reducing the entry of precursors in the gluconeogenesis pathway (Aschenbach *et al.*, 2010). At positive NPA of glucose, direct effects of insulin at hepatic level are unlikely because hepatic extraction of insulin was not modified by the NPA of glucose. Instead regulations may involve changes in the insulin/glucagon ratio (Brown and Allen, 2013). This ratio is curvilinearly affected by starch intake, and it decreases as a result of increased glucagonemia when the dietary starch concentration is higher than 160 g/kg DM (Garnsworthy *et al.*, 2008). In lactating dairy cows, glucose supply to

the animal will indeed increase with glucose absorption (Reynolds, 2005).

### Insulin

Insulin is an important regulator of hepatic metabolism of glucose and its precursors. Unfortunately, insulin net flux data were scarce, and it was a deliberate choice not to fully develop quantitative response models. It was also not possible to include it as additional covariate. However, the within-study response equations were sufficiently reproducible to draw useful conclusions. Individual nutrients have different insulin-secretory effects. Insulin secretion, reflected by its NPA, increased with the intake level and absorption (portal – arterial concentration difference) of propionate and acetate. The insulin-secretory effect of propionate shown experimentally by intra-abomasal (Casse *et al.*, 1994) or intra-ruminal (Majdoub *et al.*, 2003) infusions was almost seven times higher than that of acetate. The latter may not be a physiological regulator (Harmon, 1992). Furthermore, the sites of action of insulin may vary in relative terms with the nature of absorbed nutrients. On average,  $49 \pm 14\%$  of the NPA of insulin was taken up by the liver. But the hepatic clearance of insulin increased with the availability in volatile fatty acids (NPA), more specifically in propionate and butyrate (portal – arterial concentration difference), confirming experimental evidence for propionate (Majdoub *et al.*, 2003). This phenomenon is specific to the ruminants (Harmon *et al.*, 1991; Harmon, 1992). It suggests a proportionally greater action of insulin on liver functions, as compared to other tissues, when propionate and butyrate availability is high. By contrast, a positive NPA of glucose had no influence on the hepatic clearance of insulin, explaining why peripheral insulinaemia is more responsive to glucose than to volatile fatty acids (propionate) absorption and suggesting a proportionally greater action of insulin on tissues such as the adipose tissues and the muscles, as compared to the liver. This confirms that physiological levels of volatile fatty acids, especially propionate, and glucose differentially regulate insulin secretion, hepatic clearance and peripheral insulinaemia (Harmon, 1992; Gualdrón-Duarte and Allen, 2018).

In conclusion, glucose synthesis in the ruminant liver is tightly regulated. It does not strictly respond to precursor availability according to a single mass action law. Individual precursors are taken up by the liver in proportion of their availability but their uptake varies with the availability of other precursors. Also the rate of conversion of precursors into glucose is not fixed. Net glucose release varies curvilinearly with supply and uptake of precursors by the liver, suggesting a law of diminishing returns when precursor availability is in excess of glucose requirements. The linear portion of the response provides quantitative indications of the impacts of glucose demand, net portal absorption of glucose and negative energy balance on the net hepatic glucose release. These relationships can be linked with the predictions of portal absorption of nutrients from intake and dietary characteristics, and thereby provide indications of glucose synthesis from dietary and animal characteristics.

## Acknowledgements

The authors thank Neovia and Limagrain as well as the *Association Nationale de la Recherche et de la Technologie* for partly funding this project. This work was part of the Ph.D. thesis of the first author, C. Loncke.

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## Declaration of interest

None.

## Ethics statement

This meta-analysis study on published data did not require ethical approval.

## Software and data repository resources

The Flora database has been deposited at the Agence pour la Protection des Programmes, 249 rue de Crimée, 75019 Paris, France, IDDN.FR.001.380017.000.R.P.2010.000.10300. Authors are open to collaborations.

## Supplementary material

To view supplementary material for this article, please visit <https://doi.org/10.1017/S1751731119003410>

## References

- Armentano LE 1992. Ruminant hepatic metabolism of volatile fatty acids, lactate and pyruvate. *Journal of Nutrition* 122, 838–842.
- Aschenbach JR, Kristensen NB, Donkin SS, Hammon HM and Penner GB 2010. Gluconeogenesis in dairy cows: the secret of making sweet milk from sour dough. *IUBMB Life* 62, 869–877.
- Benson JA, Reynolds CK, Aikman PC, Lupoli B and Beaver DE 2002. Effects of abomasal vegetable oil infusion on splanchnic nutrient metabolism in lactating dairy cows. *Journal of Dairy Science* 85, 1804–1814.
- Berthelot V, Pierzynowski SG, Sauvant D and Kristensen NB 2002. Hepatic metabolism of propionate and methylmalonate in growing lambs. *Livestock Production Science* 74, 33–43.
- Blouin JP, Bernier JF, Reynolds CK, Lobley GE, Dubreuil P and Lapierre H 2002. Effect of supply of metabolizable protein on splanchnic fluxes of nutrients and hormones in lactating dairy cows. *Journal of Dairy Science* 85, 2618–2630.
- Brockman RP 2005. Glucose and short-chain fatty acid metabolism. In *Quantitative aspects of ruminant digestion and metabolism* (ed. J Dijkstra, JM Forbes and J France), pp. 291–310. CABI Publishing, Wallingford, UK.
- Brown WE and Allen MS 2013. Effects of intrajugular glucose infusion on feed intake, milk yield, and metabolic responses of early postpartum cows fed diets varying in protein and starch concentration. *Journal of Dairy Science* 96, 7132–7142.
- Casse EA, Rulquin H and Huntington GB 1994. Effect of mesenteric vein infusion of propionate on splanchnic metabolism in primiparous Holstein cows. *Journal of Dairy Science* 77, 3296–3303.
- Demigné C, Yacoub C, Morand C and Rémy C 1991. Interaction between propionate and amino acid metabolism in isolated sheep hepatocytes. *The British Journal of Nutrition* 65, 301–317.
- Doepel L, Lobley GE, Bernier JF, Dubreuil P and Lapierre H 2009. Differences in splanchnic metabolism between late gestation and early lactation dairy cows. *Journal of Dairy Science* 92, 3233–3243.
- EL-Sabagh M, Goto M, Sugino T, Obitsu T and Taniguchi K 2013. Energy metabolism by splanchnic tissues of mature sheep fed varying levels of Lucerne hay cubes. *Animal* 7, 1622–1630.
- Garnsworthy PC, Lock A, Mann GE, Sinclair KD and Webb R 2008. Nutrition, metabolism, and fertility in dairy cows: 1. Dietary energy source and ovarian function. *Journal of Dairy Science* 91, 3814–3823.
- Gualdrón-Duarte LB and Allen MS 2018. Fuels derived from starch digestion have different effects on energy intake and metabolic responses of cows in the postpartum period. *Journal of Dairy Science* 101, 5082–5091.
- Hammon HM, Schäff CT, Gruse J and Weber C 2016. Hepatic metabolism of glucose in the adaptation of the transition period in the dairy cow. In *Energy and protein metabolism and nutrition* (ed. J Skomial and H Lapierre), pp. 41–52. Wageningen Academic Publishers, Wageningen, Netherlands.
- Harmon DL 1992. Impact of nutrition on pancreatic exocrine and endocrine secretion in ruminants: a review. *Journal of Animal Science* 70, 1290–1301.
- Harmon DL, Gross KL, Krehbiel CR, Kreikemeier KK, Bauer ML and Britton RA 1991. Influence of dietary forage and energy intake on metabolism and acyl-coa synthetase activity in bovine ruminal epithelial tissue. *Journal of Animal Science* 69, 4117–4127.
- INRA 2007. Alimentation des bovins, ovins et caprins – besoins des animaux – valeurs des aliments – Tables INRA 2007. Editions Quae, Versailles, France.
- Katz ML and Bergman EN 1969. Hepatic and portal metabolism of glucose, free fatty acids, and ketone bodies in the sheep. *American Journal of Physiology* 16, 953–960.
- Kraft G, Ortigues-Marty I, Durand D, Rémond D, Jardé T, Bequette B and Savary-Auzeloux I 2011. Adaptations of hepatic amino acid uptake and net utilization contributes to nitrogen economy or waste in lambs fed nitrogen- or energy-deficient diets. *Animal* 5, 678–690.
- Kristensen NB, Røjen BA, Raun BML, Storm AC, Puggaard L and Larsen M 2009. Hepatic acetylation of the blood flow marker p-aminohippuric acid affect measurement of hepatic blood flow in cattle. In *Ruminant physiology* (ed. Y Chilliard, F Glasser, Y Faulconnier, F Bocquier, I Veissier and M Doreau), pp. 558–559. Wageningen Academic Publishers, Wageningen, Netherlands.
- Lapierre H, Galindo CE, Lemosquet S, Ortigues-Marty I, Doepel I and Ouellet D 2010. Protein supply, glucose kinetics and milk yield in dairy cows. In *Energy and protein metabolism and nutrition* (ed. GM Croveto), pp. 275–286. Wageningen Academic Publishers, Wageningen, Netherlands.
- Larsen M and Kristensen NB 2013. Precursors for liver gluconeogenesis in periparturient dairy cows. *Animal* 7, 1640–1650.
- Leng RA 1970. Glucose synthesis in ruminants. *Advances Veterinary Science and Comparative Medicine* 14, 209–260.
- Lescoat P, Danfaer A and Sauvant D 1996. Quantitative aspects of blood and amino acid flows in cattle. *Reproduction Nutrition Development* 36, 137–174.
- Loncke C, Kraft G, Savary-Auzeloux I and Ortigues-Marty I 2009a. Adaptation of hepatic glucose uptake and metabolism in growing lambs fed energy and nitrogen imbalanced diets. In *Ruminant physiology* (ed. Y Chilliard, F Glasser, Y Faulconnier, F Bocquier, I Veissier and M Doreau), pp. 586–587. Wageningen Academic Publishers, Wageningen, Netherlands.
- Loncke C, Nozière P, Bahloul L, Vernet J, Lapierre H, Sauvant D and Ortigues-Marty I 2015. Empirical prediction of net splanchnic release of ketogenic nutrients, acetate, butyrate and  $\beta$ -hydroxybutyrate in ruminants: a meta-analysis. *Animal* 9, 449–463.
- Loncke C, Nozière P, Vernet J, Lapierre H, Sauvant D and Ortigues-Marty I 2009b. Empirical prediction of net splanchnic release of  $\beta$ -hydroxybutyrate in ruminants. In *Ruminant physiology* (ed. Y Chilliard, F Glasser, Y Faulconnier, F Bocquier, I Veissier and M Doreau), pp. 444–445. Wageningen Academic Publishers, Wageningen, Netherlands.
- Loncke C, Ortigues-Marty I, Vernet J, Lapierre H, Sauvant D and Nozière P 2009c. Empirical prediction of net portal appearance of volatile fatty acids, glucose, and their secondary metabolites ( $\beta$ -hydroxybutyrate, lactate) from dietary characteristics in ruminants: a meta-analysis approach. *Journal of Animal Science* 87, 253–268.
- Majdoub L, Vermorel M and Ortigues-Marty I 2003. Intraruminal propionate supplementation modifies hindlimb energy metabolism without changing the splanchnic release of glucose in growing lambs. *The British Journal of Nutrition* 89, 39–50.
- Martineau R, Côrtes C, Ortigues-Marty I, Ouellet DR and Lapierre H 2014. Evaluation of equations predicting the net portal appearance of amino acid nitrogen in ruminants. *Journal of Dairy Science* 97, 1670–1681.
- Martineau R, Ortigues-Marty I, Vernet J and Lapierre H 2009. Technical note: Corrections of net portal flux of nitrogen compounds for differences in methods: first-step of a meta-analysis. *Journal of Animal Science* 87, 3300–3303.



- Martineau R, Sauvant D, Ouellet DR, Côrtes C, Vernet J, Ortigues-Marty I and Lapiere H 2011. Relation of net portal flux of nitrogen compounds with dietary characteristics in ruminants: a meta-analysis approach. *Journal of Dairy Science* 94, 2986–3001.
- Ortigues-Marty I, Agabriel J, Vernet J, Sepchat B, Al-Jammas M and Nozière P 2019. Contribution of nutrient fluxes to the evolution of the net energy systems, example of the INRA feeding system for beef cattle. *Translational Animal Science* 3, 1048–1063.
- Ortigues-Marty I, Vernet J and Majdoub L 2003. Whole body glucose turnover in growing and non-productive adult ruminants: meta-analysis and review. *Reproduction Nutrition Development* 43, 371–383.
- Reynolds CK 2005. Glucose balance in cattle. In *Proceedings of the 16th Florida Ruminant Nutrition Conference*, 1–2 February 2005, Gainesville, FL, USA, pp. 143–154.
- Reynolds CK 2006. Splanchnic amino acid metabolism in ruminants. In *Ruminant physiology: Digestion, metabolism and impact on gene expression, immunology and stress* (ed. K Sejrsen, TH Velplund and MO Nielsen), pp. 225–248. Wageningen Academic Publisher, Wageningen, Netherlands.
- Reynolds CK, Aikman PC, Lupoli B, Humphries DJ and Beaver DE 2003. Splanchnic metabolism of dairy cows during the transition from late gestation through early lactation. *Journal of Dairy Science* 86, 1201–1217.
- Reynolds CK, Huntington GB, Tyrrell HF and Reynolds PJ 1988. Net portal drained visceral and hepatic metabolism of glucose, l-lactate and nitrogenous compounds in lactating Holstein cows. *Journal of Dairy Science* 71, 1803–1812.
- Rodriguez-Lopez JM, Cantalapiedra-Hijar G, Durand D, Isserty-Thomas A and Ortigues-Marty I 2014. Influence of the PAH analysis method on the net hepatic flux of nutrients in lactating cows. *Journal of Animal Science* 92, 1074–1082.
- Rodriguez-Lopez JM, Feinberg M, Durand D, Cantalapiedra-Hijar G, Thomas A and Ortigues-Marty I 2016. Evaluation of the expanded measurement uncertainty in the determination of the net flux of nutrients and carbon atoms through the liver in ruminants. *Accreditation and Quality Assurance* 21, 131–142.
- Sauvant D, Schmidely P, Daudin JJ and St-Pierre NR 2008. Meta-analyses of experimental data in animal nutrition. *Animal* 2, 1203–1214.
- van der Walt JG, Baird GD and Bergman EN 1983. Tissue glucose and lactate metabolism and interconversions in pregnant and lactating sheep. *The British journal of nutrition* 50, 267–280.
- Vernet J and Ortigues-Marty I 2006. Conception and development of a bibliographic database of blood nutrient fluxes across organs and tissues in ruminants: data gathering and management prior to meta-analysis. *Reproduction Nutrition Development* 46, 527–546.
- Wilson RT, De Leuw PN and De Haan C 1983. Recherche sur les systèmes des zones arides du Mali : résultats préliminaires. *ILCA Research Report* 5, 89.