HAL open science

A specific oligosaccharide-binding site in the alternansucrase catalytic domain mediates alternan elongation

Manon Molina, Claire Moulis, Nelly Monties, David Guieysse, Sandrine Morel, Gianluca Cioci, Magali Remaud-Siméon

To cite this version:

Manon Molina, Claire Moulis, Nelly Monties, David Guieysse, Sandrine Morel, et al.. A specific oligosaccharide-binding site in the alternansucrase catalytic domain mediates alternan elongation. Journal of Biological Chemistry, 2020, 295 (28), pp.9474-9489. 10.1074/jbc.RA120.013028 . hal-02918812

HAL Id: hal-02918812

https://hal.inrae.fr/hal-02918812

Submitted on 21 Aug 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Distributed under a Creative Commons Attribution 4.0 International License

JBC Papers in Press. Published on May 14, 2020 as Manuscript RA120.013028
The latest version is at https://www.jbc.org/cgi/doi/10.1074/jbc.RA120.013028

A specific oligosaccharide-binding site in the alternansucrase catalytic domain mediates alternan elongation

Manon Molina, Claire Moulis, Nelly Monties, David Guieysse, Sandrine Morel, Gianluca Cioci*, Magali Remaud-Siméon*

TBI, Université de Toulouse, CNRS, INRAE, INSA, Toulouse, France

Running title: *Oligosaccharide-binding sites in alternansucrase*

* Corresponding Authors
remaud@insa-toulouse.fr
cioci@insa-toulouse.fr

Keywords: glucansucrase, alternan, bio-sourced, polysaccharide, GH70, polymerase, sugar binding pockets, surface binding site, processivity, alternansucrase

ABSTRACT

Microbial α-glucans produced by glycoside hydrolase family 70 (GH70) glucansucrases are gaining importance because of the mild conditions for their synthesis from sucrose, their biodegradability, and their current and anticipated applications that largely depend on their molar mass. Focusing on the alternansucrase (ASR) from *Leuconostoc citreum* NRRL B-1355, a well-known glucansucrase catalyzing the synthesis of both high-molar mass (HMM) and low-molar mass alternans, we searched for structural traits in ASR that could be involved in the control of alternan elongation. The resolution of five crystal structures of a truncated ASR version (ASRΔ2) in complex with different gluco-oligosaccharides pinpointed key residues in binding sites located in the A and V domains of ASR. Biochemical characterization of three single mutants and three double mutants, targeting the sugar-binding pockets identified in domain V, revealed an involvement of this domain in alternan binding and elongation. More strikingly, we found an oligosaccharide binding site at the surface of domain A, distant from the catalytic site and not previously identified in other glucansucrases. We named this site surface binding site A1 (SBS-A1). Among the residues lining the SBS-A1 site, two (Gln-700 and Tyr-717) promoted alternan elongation. Their substitution to alanine decreased HMM alternan yield by a third, without significantly impacting enzyme stability or specificity. We propose that the SBS-A1 site is unique to alternansucrase and appears to be designed to bind alternating structures, acting as a mediator between the catalytic site and the sugar-binding pockets of domain V and contributing to a processive elongation of alternan chains.

INTRODUCTION

Since the last decades, microbial polysaccharides have gained attention as promising bio-sourced polymers that can be regularly supplied and are less sensitive to market and climate fluctuations than plant polymers (1, 2). In addition, progresses in structure-function studies and engineering of polymerases allow today to better control their structures and by extension their physicochemical properties. In this field, the glucansucrases belonging to the family 70 of glycoside hydrolases (GH70) (3) are very interesting enzymes. These α-transglucosylases catalyze the formation of high molar mass (HMM) and low molar mass (LMM) homopolysaccharides of D-glucose from sucrose, a low-cost and abundant substrate.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

The panel of polymers produced by glucansucrases varies dramatically in terms of size, type and arrangement of α-glycosidic linkages and degree of branching, all these features defining polymer structural properties and consequently, the range of ongoing or potential applications. Among them, dextran, an α-1,6 linked glucan, was the first microbial polymer to be commercialized and used as plasma substitute (4). Other applications of α-glucans are mainly in the biomedical and analytical fields but they are also used as thickening agents for food and cosmetics or as flocculants for ore mining (5, 6). Recently, the α-1,3 linked α-glucan named mutan was proposed to be used for biomaterial applications (7, 8).

To produce size-defined α-glucans directly from sucrose and avoid fractionation steps, it seems essential to identify the molecular determinants involved in the control of polymer size. Since 2010, the 3D structure resolution of five different glucansucrases established that they all adopt a *U-shaped* fold made up by four domains (A, B, C, IV), which are flanked by an additional domain V (9–13) formerly known as the Glucan Binding Domain (GBD). Structure-function studies that followed further showed that sugar binding pockets in the domain V of at least three GH70 family enzymes - the branching sucrase GBD-CD2 that catalyzes α-1,2 branches formation in dextran as well as the dextransucrase DSR-M from *Leuconostoc citreum* NRRL B-1299 and the dextransucrase DSR-OK from *Oenococcus kitaharae* - participate in polymer binding and were proposed to be involved in dextransucrase processivity leading to HMM polymer formation (12, 14, 15). Other residues located in the enzyme active site, particularly in subsites +2 or +3 (near subsites -1 and +1 where the glucosyl and fructosyl moieties of sucrose are accommodated, respectively (16)) were also found critical for HMM dextran formation in the dextransucrases DSR-S from *Ln. mesenteroides* NRRL B-512F, GTF180 from *Lactobacillus reuteri* 180 and DSR-M from *Ln. citreum* NRRL B-1299 (17–19).

Surprisingly, additional surface (or secondary) binding sites more distant from the active site were never described in the domains A, B or IV of GH70 dextransucrases contrary to what was observed for GH13 enzymes, which belong to the same clan as the GH70 enzymes and for which the distant surface binding sites were found to play a role in substrate targeting, allosteric regulation and also processivity (20–24).

Given the structural diversity of α-glucans, we wondered whether the role of sugar binding pockets in the synthesis of HMM dextran could be generalized to all glucansucrases, especially those producing polymers with a structure very different from that of dextran. With this in mind, we focused on the recombinant alternansucrase (ASR) from *Ln. mesenteroides* NRRL B-1355 strain, recently reclassified as *Ln. citreum* (25), which stands out as a distinct GH70 glucansucrase synthesizing a polysaccharide made of alternating α-1,3 and α-1,6 linkages, named « alternan » (26). From sucrose, ASR catalyzes the synthesis of a bi-modal population of alternan, one of high molar mass (around 1,700,000 $g.mol^{-1}$) and one of much lower molar mass (1,300 $g.mol^{-1}$) (27). These two populations are formed during the early stage of the reaction, suggesting that ASR follows a semi-processive mechanism of polymerization involving polymer anchoring regions in the enzyme to facilitate HMM alternan formation (28). HMM alternan is more soluble in water and less viscous than dextran, making it a good substitute of Arabic gum (29, 30). It was also recently shown to form nanoparticles of interest for biomedical applications (31) and to promote the proliferation, migration and differentiation of human mesenchymal stem cells (32). In addition, oligoalternans obtained by acceptor reaction with maltose (33–36) are hydrolyzed only by a limited number of microbial glycoside hydrolases such as isomaltodextranases (29) or alternanases (37)

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

and are resistant to mammalian enzymes, which is of interest for applications as prebiotics.

Recently, the resolution of ASR 3D-structure combined to mutagenesis and molecular docking unraveled the structural basis for α-1,3 and α-1,6 linkage specificity and for the alternance mechanism that is governed by the recognition of the terminal glucosidic linkage of the incoming acceptor, thanks to specific subsites (13). Several residues in the proximity of the active site were also found to be critical for HMM alternan formation. In particular, the replacement of Trp675 (subsite +2) and Trp543 (subsite +3') by Alanine resulted in 82% and 54% decrease of HMM polymer synthesis, respectively. In addition, four putative sugar binding pockets (V-A, V-B, V-C and V-D) homologous to those found in other GH70 family enzymes have been identified in ASR domain V. Surprisingly, removal of the entire domain V did not completely abolish polymer formation but reduced it by 86% (13).

The control of polymer elongation by ASR is thus complex, involving residues of both the catalytic site and domain V. To get deeper insight in this mechanism, we placed our effort in the resolution of several ASR complexes. To this end, ASR crystals were soaked with different sugars varying in terms of degree of polymerization (DP) and glycosidic linkages. We obtained several complexes with different types of oligosaccharides bound in domains A and V. Combined with mutant characterization, the study highlighted the contribution of the domain V in HMM alternan formation and shed light on the importance of a new surface/secondary binding site, found in domain A but remote from the catalytic core, which is defined by residues Tyr717 and Gln700 and was named SBS-A1 .

RESULTS

New complexes and identification of two sugar binding sites: Pocket V-B in domain V and SBS-A1 in domain A

To explore the structural determinants for alternan elongation, ASRΔ2 crystals were soaked with different acceptors varying in size and structures (Figure 1). The following ligands were tested: glucose, isomaltose (I2), nigerose (α-D-Glc*p*-(1→3)-D-Glc), isomaltotriose (I3), panose, isomaltononaose (I9), isomaltododecaose (I12, α-D-Glc*p*-(1→6)-[α-D-Glc*p*-(1→6)]$_{10}$-D-Glc) and a partially purified oligoalternan (OA) of DP around 8 that contains 39% of α-1,3 linkages and 61% of α-1,6 linkages (Figure S1 and S2).

For the complexes with isomaltose (I2), isomaltotriose (I3), panose and oligoalternan (OA), we unexpectedly observed a positive difference electron density in the domain A, in a site that we named Surface Binding Site A1 (SBS-A1) and that was never described before in GH70 family enzymes (Figure 1). For the same four ligands plus the isomaltononaose (I9), density was also observed in the domain V at a position corresponding to the sugar binding pocket V-B (Gly234-Thr304) previously proposed to be a putative sugar binding pocket (13). In contrast, no interpretable electron density was found in the pocket referred as V-A, also previously predicted to be a sugar-binding pocket. In all the complexes, the ASR crystallized in the space group $P2_12_12_1$ with the presence of two protein molecules in the asymmetric unit and very similar unit cell than the previously solved unliganded structure. Resolutions obtained are between 3.0Å to 3.5Å. Of note, in all the obtained complexes we will refer to the chain A where the electron density is better defined. The glucosyl units of each oligosaccharide will be numbered by ascending order from their reducing end.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

Domain V sugar binding pockets description and comparison

We used the I2 and I3 complexes for which we obtained the highest resolution (3.0Å) to model the isomaltose and isomaltotriose that are bound in pocket V-B. Comparison with the high resolution structures of the branching sucrase GBD-CD2 from *Ln. citreum* NRRL B-1299 in complex with I2 and I3 helped the assignment of protein-ligand interactions (Figure S3). The Glcp_2 is in CH-π stacking interaction with Tyr241 (ASR numbering) and interacts with Gln278, Gln270 and Lys280 through O2, O3 and O1 atoms respectively and also with the main chain oxygen of Thr297 through O3 (Figure 2A and S4). The Glcp_1 mainly interacts with the Thr249 through its O5 and with the Lys280 through O6 and O5 hydroxyls which suggests that Lys280 is a pivotal residue for the recognition of isomaltose moiety, as already observed in branching sucrase GBD-CD2 and dextransucrase DSR-M structures. In the I3 complex, the Glcp_3 occupies the remaining part of the pocket which is delimited by the Asp266 and weakly interacts with Asp266 and Asn268. Although obtained at lower resolution (Figure S4), the other complexes obtained in pocket V-B (panose, OA and I9) have also been modeled on the basis of the already established I2/I3 interaction. Panose interacts mainly with its isomaltose moiety as the presence of the short α-1,4 linkage probably destabilize the interaction between Lys280 and Glcp_1. For the OA complex only three monosaccharides could be modelled into the electron density which has a different shape than the I3 molecule (Figure S4). Assuming that an isomaltose moiety is still in interaction with the Gln278/Gln280, this ligand has been modeled as one isomaltose moiety further decorated by an α-1,3 linked glucose on its non-reducing end (Figure 2C). Unfortunately, the remaining part of the oligosaccharide is not visible and the lower resolution of this complex does not allow a complete modelling of this interaction. Lastly, it should be observed that in all complexes two protein monomers are arranged around a pseudo 2-fold axis in the asymmetric unit and the domain V of each unit is intertwined with its equivalent of the second chain. Only seven out of nine glucosyl units of isomaltononaose are visible into electron density and they appear to bind at the interface of the two pockets V-B of each chain (Figure S5).

In our structures Gln270, Tyr241, Gln278 and Lys280 of pocket V-B are well aligned with Gln186, Tyr158, Gln194 and Lys196 of pocket V-A (Figure 2A, 2B), whereas different residues are found in the pocket V-C (Figure 2D, S6). On this basis we can propose that the sugar binding pockets V-A and V-B are functional whereas the pocket V-C is likely to be non-functional due to the absence of the QxK motif that, associated with a Tyr residue at the bottom of the pocket, was suggested to be a signature of sugar binding pocket functionality (Figure 2D) (12, 14). The structural comparison of pockets V-A and V-B revealed similarities but also subtle differences; in particular the pocket V-A seems to be more open with a longer distance between the aromatic platform (Tyr158 OH) and the top of the pocket (Thr213 O) that might explain the reduced affinity for oligosaccharides, at least in the crystalline form (Figure S6).

Mutation of the conserved tyrosine in the sugar binding pockets of domain V

To further investigate the role of V-A and V-B pockets, the central stacking residues, Tyr158 and Tyr241 respectively, were replaced by an alanine. The mutations resulted in a slight decrease of the HMM polymer yield, from 31.5% for the wild type to 27.2%, 28% and 27.6% for the Tyr158Ala mutant, the Tyr241Ala mutant and the Tyr158Ala+Tyr241Ala double mutant, respectively (Figure 3, Table 1). Enzyme specific activity, specificity and melting temperature were not significantly affected for these three mutants (Table 1).

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

To explore the affinity of the enzyme with glucan, we performed affinity gel electrophoresis of ASRΔ2, dextransucrase DSR-M (C-terminal truncated form DSR-MΔ1 that includes sugar binding pockets V-A, V-B and V-C (12), Figure 2D) and branching sucrase GBD-CD2 (N-terminal truncated form ΔN_{123}-GBD-CD2 (38) that includes sugar binding pockets V-J, V-K and V-L (14), Figure 2D) in native conditions. The two latter enzymes also possess functional sugar binding pockets in which oligosaccharides were experimentally shown to bind. Logically, the migration of the three enzymes is delayed in the presence of dextran as enzyme sugar binding pockets bind to a certain extend to the dextran containing gel matrix. In contrast, only ASR was slightly retained by the presence of alternan in the gel (Figure 4A). Focusing on ASR sugar binding pockets, both single and double mutations Tyr158Ala (V-A) and Tyr241Ala (V-B) affected the binding ability of the enzyme with dextran and alternan, confirming a participation of the sugar binding pocket stacking residue in glucan binding. A subtle difference is observed between the migration of Tyr158Ala and Tyr241Ala mutants in particular with the presence of alternan, the Tyr241Ala mutant being almost not delayed (Figure 4B). This is suggesting that the pocket V-B (Tyr241) has a slightly more affinity than the pocket V-A, thus being in agreement with our complexes where we clearly see the presence of oligosaccharides only in the pocket V-B.

Description of the surface binding site identified in the domain A: SBS-A1

The SBS-A1 site was never described before for any other GH70 enzymes. To model the various ligands found in SBS-A1, we first sought to determine the direction of the sugar chain in the binding site. To do so, I2 was fitted in the electron density considering two possible orientations with either Glcp_1 or Glcp_2 in stacking interaction with Tyr717 (Figure S7). The retained configuration corresponds to Glcp_1 in stacking interaction with Tyr717 as this positioning sets the 1-6 glucosidic linkage in a low energy conformation (Figure S8) and also allows the preferred interaction between alpha-D sugars and aromatic platforms that is, with the O1 pointing in the opposite direction of the stacking residue (39, 40). To further validate this binding mode, automatic docking of I2 has been performed, confirming that the lowest energy pose is with the Glcp_1 in stacking interaction with Tyr717 (Figure S7). A similar electron density was observed around the Tyr717 for both the isomaltotriose (I3) and the panose complexes suggesting that I2, I3 and panose all interact with their isomaltose moiety (Figure S7). Concerning the oligoalternan and assuming that this molecule is composed of alternative α-1,6 and α-1,3 linkages, we modeled its visible part as an hexasaccharide and succeeded to fit in the electron density with the Glcp_3 ring in stacking interaction with Tyr717 (Figure S7). Lastly and interestingly, no electron density could be observed in SBS-A1 for the longer isomaltooligosaccharides (I9, I12) that have been tested.

Isomaltotriose, isomaltose and panose adopt a similar positioning

Four residues are found in interaction with I3 (Figure 5A). Tyr717 is in CH-π stacking with Glcp_2 (parallel configuration) and Glcp_3 (T-shaped configuration). In addition, the O3 and O4 of Glcp_3 interact with the main chain O of Tyr717 and with the side chain of Gln700. The O4 of Glcp_2 interacts with Gln700 and Asn703, the O3 with Gln700, Ser713 and Asn703 and the O2 with Ser713 only. Glcp_1 is not stabilized by any interactions with the protein. Glcp_1 and Glcp_2 of I2 superimpose well with Glcp_2 and Glcp_3 of I3 and are bound through the same network of interactions (Figure S9). Panose binding involves the same four residues as those described for I2 or I3 complexes. Similarly, Glcp_1 of panose is not maintained by any interactions (Figure 5B). Noteworthy, we also attempted crystal soaking with glucose and nigerose but could not obtain any complex bound in SBS-A1. Hence, we

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

suggest that at least two α-1,6 linked glucosyl units are required for a correct positioning around Tyr717, one in parallel and the other in T-shaped stacking interaction.

Oligoalternan wraps around Tyr717

The glucosyl units Glcp_2, Glcp_3, Glcp_4 of the six visible glucosyl residues of the OA are in the same region as the three glucosyl residues of I3 and panose (Figure 5C). The units Glcp_1 and Glcp_2 are unbound. The O3 of Glcp_3 interacts with Ser713 and Gln700 and the O4 is coordinated with Gln700 and Asn703. There is a CH-π stacking interaction between Glcp_3 and Tyr717 and also a T-shaped stacking interaction with Glcp_4. The latter is also hydrogen bonded with Gln700 through O6 and Tyr717 carbonyl through O4. The unit Glcp_5 interacts with residues not identified previously with its O2 bound to Asp720 and the main chain of Gly722. Finally, the O2 of Glcp_6 interacts only with the hydroxyl group of Tyr717 side chain. For Glcp_5 and Glcp_6, there may also be a parallel-displaced stacking interaction with Trp716. Noteworthy, Tyr717 is in interaction with Glcp_3, Glcp_4 and Glcp_6. The OA literally wraps around this amino acid making it a pillar residue of SBS-A1 binding site.

The SBS-A1 is a mediator of HMM alternan formation

The five residues described in interaction with both I3 and OA (Gln700, Asn703, Ser713, Trp716 and Tyr717) were individually replaced by an alanine to evaluate their importance for enzyme specificity and polymer size distribution. The mutations did not affect enzyme melting temperature, linkage specificity, or hydrolysis rate (Table 1). The specific activity was also globally well-conserved, all mutants keeping between 76.4% and 95.8% of residual activity compared to wild type ASRΔ2. The product profile was unchanged for the Asn703Ala, Ser713Ala and Trp716Ala mutants (Figure S10). In contrast, the polymerization process was clearly affected by the mutations of Gln700 and Tyr717 confirming the importance of these residues for oligosaccharide binding. Indeed, the amount of HMM polymer decreased from 31.5% for the wild type enzyme to 20.4%, 18.8% and 18.3%, respectively, for the Gln700Ala, Tyr717Ala mutants and the Gln700Ala-Tyr717Ala double mutant (Table 1, Figure 6A). The peak apex of HMM polymer formed with the mutants is also slightly displaced towards higher masses. This may reflect a possible variation of the polymer supramolecular organization as suggested by Dynamic Light Scattering (DLS) assay used to compare the size distribution of wild type and mutant alternan in solution. Indeed, the wild type alternan has the ability to form nanoparticles with a diameter of ~90 nm and low polydispersity whereas the Tyr717Ala polymer is much more polydisperse and tend to aggregate (Figure S11). This could be due to variation of branching length (among other possible reasons) even if the global percentage of α-1,3 linkages was unchanged compared to the wild type alternan. Further analyses would be required to investigate the wild type and mutant alternan structures in more details and conclude. To note, alternan from ASRΔ2 and *L. citreum* ABK-1 alternansucrase (31) showed an almost identical nanoparticle size.

Chromatographic analysis by HPAEC-PAD of the sucrose reaction products obtained with ASRΔ2 and the Tyr717Ala mutant confirmed that the amount of oligosaccharides formed with the mutant is more abundant, hence corroborating the results of HPSEC (Figure 6B). However, when performing the reaction with maltose acceptor, the chromatograms of the reaction products from DP 2 to DP around 7/8 were perfectly stackable for all the seven mutants, as shown as example for Tyr717Ala mutant (Figure 6C). These products result from maltose glucosylation and correspond to previously characterized oligoalternans with a maltose unit at the reducing end (13, 41).

To assess binding interactions with alternan or glucans, we performed affinity gel electrophoresis of ASRΔ5 which is a truncated

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

form of ASR devoted of its domain V, or mutant ASRΔ5 Tyr717Ala in native conditions with dextran or alternan. The idea was to check whether SBS-A1 site could confer affinity for dextran and alternan in the absence of domain V. We did not observe any differences between the ASRΔ5 and ASRΔ5-Tyr717Ala migration in the presence of dextran or alternan indicating that the contribution of Tyr717 of SBS-A1 to glucan binding remains weaker than that conferred by Tyr158 or Tyr241 of sugar binding pockets from domain V (Figure 4A).

The Tyr717Ala mutation was combined with mutations of the stacking residues in the sugar binding pocket of the domain V, Tyr158Ala and Tyr241Ala for pocket V-A and V-B, respectively. The resulting HMM alternan yield was reduced from 18.8% for the Tyr717Ala single mutant to 13.3% and 8.5% for the Tyr717Ala+Tyr158Ala and Tyr717Ala+Tyr241Ala double mutants, respectively (Figure 7A). The mutations did not impact significantly the specificity and residual activity was higher than 60% for both mutants (Table 1). HMM polymer formation starts earlier with wild type ASRΔ2 than with the mutant Tyr717Ala (around 5 minutes and 10 minutes, respectively) and the production rate is almost twice faster (0.50 $g.L^{-1}.min^{-1}$ and 0.27 $g.L^{-1}.min^{-1}$, respectively) (Figure 7B, Figure S12). In parallel, oligoalternans formation is enhanced with the mutant Tyr717Ala, which is in accordance with a reduced formation of HMM polymer (Figure S13). Notably, HMM polymer formation rate is even lower for the double mutants Tyr717Ala+Tyr158Ala mutant (pocket V-A) and Tyr717Ala+Tyr241Ala mutant (pocket V-B). Mutation in pocket V-B affects more the kinetics and yield of HMM polymer formation than that in pocket V-A (Figure 7B).

DISCUSSION

We disclose here five crystal structures of alternansucrase in complex with different oligosaccharides. They are the first complexes ever obtained with this enzyme, a α-transglucosylase showing a unique linkage specificity among the GH70 glucansucrases by alternating α-1,6 and α-1,3 linkages. These structures enabled us to locate several oligosaccharide binding sites in the enzyme. Two of them were located in the domain V and a new site was identified in the domain A. We have investigated their role on both the linkage specificity, stability and ability to synthesize high molar mass (HMM) alternans. It is important to emphasize on the fact that this enzyme naturally catalyzes the synthesis of both HMM and low molar mass (LMM) alternans. The idea behind this work was to identify structural parameters that could allow the design of ASR strictly specific for either HMM or LMM polymer synthesis in the future. What have we learned?

Involvement of ASR domain V and the sugar binding pockets in HMM alternan formation

For all the complexes we solved, we have found isomaltooligosaccharides and oligoalternans bound in domain V. This domain shares a high percentage of identity with its counterpart exhibited by dextransucrase DSR-M. As for DSR-M we have identified two sugar binding pockets, V-A and V-B, showing the structural traits close to those previously described. We have found clear electron density for isomaltooligosaccharides in the pocket V-B of alternansucrase. Affinity gel electrophoresis revealed that ASR binds dextran like dextransucrase DSR-M (12). More interestingly, we also found oligoalternans bound in pocket V-B and affinity gel confirmed the interaction between alternan and the domain V of the enzyme thus demonstrating the binding promiscuity of this domain. In contrast, dextransucrase DSR-M and branching sucrase GBD-CD2 were not retained by alternan, showing that despite structural similarities, the sugar binding pockets affinity could be subtly different from one enzyme to another. In particular, the positioning of isomaltooligosaccharide is

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

different in both DSR-M and GBD-CD2 with the presence of a second aromatic platform (Tyr187 and Trp1849 respectively) that could prevent the orientation observed in ASR pocket V-B (Figure S3). Lastly, in the complex with I9, the oligosaccharide seems to bind at the interface of the two domains V giving a striking example on how multiple glucansucrase chains can simultaneously bind to a single polymer chain and this is very much likely to take place in solution.

Analysis of the complexes revealed the same interaction network than dextransucrase DSR-M and branching sucrase GBD-CD2 involving a QxK motif and a conserved aromatic residue with the oligosaccharides. Replacing the aromatic residues to alanine in each pocket induces a slight but significant effect on the HMM polymer yield (Table 1), indicating that these pockets may interact with the alternan chain and promote its elongation by providing anchoring platforms for long chains. Interestingly, mutations in the pockets were less detrimental to HMM alternan formation than deletion of the entire domain V (truncated mutant named ASRΔ5). ASRΔ5 synthesized only 4.5% of HMM polymer (13), versus 31.5% for ASRΔ2 and around 27% for the single or double mutants targeting the aromatic residue of the sugar binding pockets V-A and V-B. The mutation of only the conserved aromatic residue of the pockets (Tyr158 and Tyr241) may not be sufficient to totally abolish the interactions, even in the double mutant, and to obtain similar effect to an entire deletion. However, it should be noted that deletion of the entire domain V (removal of 481 residues, ~1/3 of the protein (Figure S14)) may also modify the folding of the other domains as suggested by lower melting temperature of ASRΔ5 variant (-2°C compared to ASRΔ2) and this could have an impact on the production of HMM polymer. Altogether, our findings show that domain V plays a limited role in the formation of HMM polymer. Additional mutations targeting the conserved Gln and/or Lys of the QxK motif should help to conclude with more confidence on the role of the pockets in ASR domain V.

The Surface Binding Site A1, a signature of alternansucrase

SBS-A1 is the first functional surface binding site described for a GH70 family enzyme *e.g.* (i) located in the catalytic domain, (ii) where carbohydrates bound non-catalycally and (iii) at a fixed position relative to the catalytic site. This is in agreement with the presence of surface binding sites in GH13 and GH77 enzymes which belong to the same GH-H clan than GH70 enzymes (20, 22).

Of the various ligands tested in our soaking experiments, I2, I3 and panose did bind to SBS-A1 (Figure 1), nigerose did not as well as I9 nor I12 (data not shown). A close inspection of I3 complex revealed that, in the proposed configuration, a fourth α-1,6 linked glucose could not be added to the non-reducing end of I3 due to a steric clash with the protein surface. In contrast, the addition of an α-1,3 linked glucose enables the steric clash to be avoided and also supports the proposed binding mode of the OA in Figure 5C. The shape complementarity between protein surface and the OA indicates that the site is particularly well-designed to interact with oligosaccharides containing alternated α-1,6 and α-1,3 linkages above DP 4. Two residues appeared to be particularly important: Tyr717 and Gln700. Their replacement by Ala strongly reduced HMM polymer yield from 31.5% for ASRΔ2 to 18.8% and 20.4% for the mutants, showing that there is an important contribution of SBS-A1 to alternan elongation. In agreement with this assumption, mutagenesis of Tyr717 together with the deletion of domain V led to a further decrease of the percentage of HMM polymer produced from 4.5% to 2.3%. Thus, SBS-A1 would be involved in keeping the acceptor chain in close proximity to the catalytic site for further elongation, enhancing processivity, similarly to what was described for the GH13 amylosucrase of *Neisseria polysaccharea* (42), a transglucosidase that

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

belongs to the same GH-H clan than ASR and uses sucrose as substrate to catalyze the formation of an amylose-like polymer. Finally, the glucosylation of short oligosaccharides (DP 3 to DP 6) seems not at all impacted by the mutations operated in SBS-A1 confirming that SBS-A1, which is remote from the active center, comes on stage only when the formed oligosaccharides reach a sufficient length. Furthermore, we propose SBS-A1 binding site as a signature of alternansucrase specificity. Indeed, Gln700 and Tyr717 are conserved in all characterized (26, 31) and putative alternansucrase sequences identified by BLASTp search (from *Ln. citreum* NRRL B-1501, NRRL B-1498, LBAE-C11, KM20, and EFEL 2700 strains) (Figure 8A). Furthermore, of 64 characterized GH70 glucansucrases, residue Tyr717 is only found in alternansucrase sequences and is mainly replaced by an arginine (22/64) or a proline residue (20/64) in the other glucansucrases (Figure 8B). Finally, a question remains pending: is there an interplay between SBS-A1 site and the domain V of ASR? To address it, we constructed double mutants bearing the Tyr717Ala combined with mutation of the stacking tyrosine residue in each of the two pockets V-A and V-B. The HMM alternan yield changed from 31.5% (ASRΔ2) to 18.8% (Tyr717Ala) and further decreased to 13.3% and 8.5% for Tyr158Ala-Tyr717Ala and Tyr241Ala-Tyr717Ala double mutants respectively. This was also correlated with the slowdown of HMM alternan formation rate and provides evidence of an interplay between SBS-A1 and the sugar binding pockets, in particular with the pocket V-B. Therefore, SBS-A1 could act as a molecular bridge, favoring oligosaccharide capture and the semi-processive mode of elongation as previously suggested by Moulis *et al.* (28). This hypothesis adds a further level of complexity in the role of surface binding sites as not only the SBS-A1 and the domain V both seems to have a role in processivity, but they also probably cooperate. This is in agreement with the observation that SBSs and Carbohydrate Binding Modules (CBM) frequently co-occur in carbohydrate active enzymes (23).

CONCLUSION

To sum up, we described herein five new 3D structures of ASR in complex with different sugars (isomaltose, isomaltotriose, isomaltononaose, panose and oligoalternan) bound either in domain A (SBS-A1) or in domain V (sugar binding pocket V-B). The role of the domain V, as well as the new site SBS-A1 proposed as a signature of alternansucrase specificity, has been clarified. We have generated mutants quasi exclusively specific for the formation of oligoalternans (ASRΔ5-Y717A produces less than 3% of HMM alternan). Engineering alternansucrase for the exclusive formation of HMM alternan remains highly challenging. One could think to engineer the domain V and the sugar pockets in order to increase the affinity for alternan and shift the enzyme mechanism towards more processivity. Having a clear vision of the interaction with a longer alternan chain and possibly during the catalysis, would be a major achievement. As the resolution of crystal structures in complex with longer oligosaccharides (DP >9) is likely to be very difficult, the use of different techniques such as time resolved CryoEM and/or NMR could be of interest. Molecular dynamic simulations could also be performed to predict the positioning of long chains connecting the active site to the domain V *via* the site SBS-A1.

EXPERIMENTAL PROCEDURES

Production and purification of ASRΔ2 and ASRΔ5

ASRΔ2 and ASRΔ5 (Figure S14) were produced from cultures of *E. coli* BL21 DE3* transformed with plasmid pET53-*asrΔ2* or pET53-*asrΔ5* and purified using the conditions described in (13).

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

Crystallization and Data collection

Crystals of ASRΔ2 were obtained using the conditions identified previously (13). Crystals were soaked in the reservoir solution complemented with 15% (v/v) Ethylene glycol along with variable concentration of different oligosaccharide (Table 2). Isomaltose (α-D-Glc*p*-(1→6)-D-Glc), isomaltotriose (α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-D-Glc) and panose (α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc) were ordered from Carbosynth. Oligoalternan (α-(1→6)/α-(1→3) alternated) and Isomaltononaose (α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-D-Glc) were purified in-house. Crystals were then directly cryo-cooled in liquid nitrogen. To note, the soakings seemed to affect crystal integrity and data quality, especially those with longer ligands. This could explain the higher R-factors and also B-factors shown by some of our structures. Data collection was performed at the European Synchrotron Radiation Facility (Grenoble, France) on beamline ID23-1 for isomaltose and panose complexes and at ALBA synchrotron (Barcelona, Spain) on beamline XALOC for Isomaltotriose, Isomaltononaose and Oligoalternan complexes. Diffraction images were integrated using XDS (43) and converted to structure factors using CCP4 programs (44). The structure was solved by molecular replacement using PHASER and the unliganded ASRΔ2 structure (PDB ID: 6HVG) as search model. To complete the model and build the oligosaccharides in the density, cycles of manual rebuilding using COOT (45) were alternated to restrained refinement using REFMAC5 (46). The structures were validated using MolProbity (47) and deposited in the Protein Data Bank. Data collection and refinement statistics are shown in Table 2. The validation of oligosaccharide structures was done using the CARP server (http://www.glycosciences.de/tools/carp/) (48) and shown in Figure S8.

Automatic Docking in the SBS-A1

Starting conformations for isomaltose (I2) were generated using the Carbohydrate Builder at http://glycam.org. Receptor and ligand structures were prepared for docking with AutoDockTools (49, 50) and automatically docked with Vina-Carb (51) using standard settings.

Oligoalternan preparation for soaking experiments

Acceptor reactions with a sucrose:glucose mass ratio of 2:1 were set up in the same conditions as above to produce oligoalternans. Oligoalternans were partially purified by size exclusion chromatography using two 1 m XK 26 columns (GE Healthcare) in series packed with Bio-Gel P6 and P2 resin (Biorad) and water as eluent at 1 mL/min flow rate. The fraction used for soaking experiments was analyzed using HPAEC-PAD (Figure S1), ^{1}H NMR in the same conditions as described below and Maldi-TOF-MS (Figure S2). The MW of the partially purified oligoalternan was determined in a Waters® MALDI-Micro MX-TOF Mass spectrometer. The measurements were performed with the mass spectrometer in positive reflectron mode using an accelerating voltage of 12 kV. Mass spectra were acquired from 400/m/z to 3000/m/z. Samples were dissolved in water (1 mg.ml^{-1}). A 0.75 ml of sample solution was mixed with 0.75 ml of the matrix solution (2,5-dihydroxybenzoic acid 10 mg.ml^{-1} in H2O:EtOH 0.5:0.5; v/v) and a total of 1.5 µl was applied to a stainless steel sample slide and dried at room temperature.

Site directed mutagenesis study

Mutants were constructed by inverse PCR using the pET53-*asr*-Δ2 or pET53-*asr*-Δ5 genes as template, Phusion® polymerase (NEB), and the primers described in Table S1. Following overnight *Dpn*I (NEB) digestion, the PCR product was transformed into competent *E. coli* DH5α and clones were selected on solid LB medium supplemented with ampicillin 100 µg.mL^{-1}. Plasmids were

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

extracted with the QIAGEN spin miniprep kit and mutated *asr* genes were checked by sequencing (GATC Biotech). All mutants were produced and purified as described above.

Activity measurement

Activity was determined in triplicate at 30°C in a Thermomixer (Eppendorf) using the 3,5-dinitrosalicylic acid method (52). 50 mM sodium acetate buffer pH 5.75, 292 mM sucrose and 0.05 $mg.mL^{-1}$ of pure enzyme were used. One unit of activity is defined as the amount of enzyme that hydrolyzes 1 µmol of sucrose per minute.

Enzymatic reaction and product characterization

Polymer productions were performed using 1 $U.mL^{-1}$ of pure enzyme with 292 mM sucrose in 50 mM sodium acetate buffer pH 5.75 at 30°C over a period of 24 hours. The products were analyzed using High Pressure Size Exclusion Chromatography (HPSEC) with Shodex OH-Pak 805 and 802.5 columns in series in a 70°C oven with a flow rate of 0.250 $mL.min^{-1}$ connected to RI detector. The eluent was 50 mM sodium acetate, 0.45 M sodium nitrate and 1% (v/v) ethylene glycol. The polymer yield was calculated using the area of the peak corresponding to HMM glucan divided by the sum of the areas of all the peaks arising on the chromatogram.

The products were also analyzed by High Pressure Anion Exchange Chromatography with Pulsed Amperometric Detection (HPAEC-PAD) using a CarboPac TM PA100 guard column upstream of a CarboPac TM PA100 analytical column (2 mm x 250 mm) at a flow rate of 0.250 $mL.min^{-1}$. The eluents were A: 150 mM NaOH and B: 500 mM sodium acetate with 150 mM NaOH. Sugars were eluted with an increasing 0 to 60% gradient of eluent B for 30 minutes. Quantification was performed using standards of glucose and sucrose at 5, 10, 15 and 20 $mg.L^{-1}$. The hydrolysis percentage was calculated by dividing the final molar concentration of glucose by the initial molar concentration of sucrose.

Acceptor reactions were set up in the presence of maltose (sucrose:maltose mass ratio 2:1) using 1 $U.mL^{-1}$ of pure enzyme with 292 mM sucrose in 50 mM NaAc buffer pH 5.75 at 30°C over a period of 24 hours. The products were analyzed by High Pressure Anion Exchange Chromatography with the same conditions than described above. The structures corresponding to the nomenclature used are: OD4: α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc; OD5: α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc; OA4: α-D-Glc*p*-(1→3)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc; OA5: α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→3)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc; OA6: α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→3)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc and α-D-Glc*p*-(1→3)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→3)-α-D-Glc*p*-(1→6)-α-D-Glc*p*-(1→4)-D-Glc.

NMR samples were prepared by dissolving 10 mg of the total products from sucrose in 0.5 mL D_2O. Deuterium oxide was used as the solvent, and sodium 2,2,3,3-tetradeuterio-3-trimethylsilylpropanoate ($TSPD_4$) was selected as the internal standard (δ^1H = 0 ppm, $\delta^{13}C$ = 0 ppm). ^1H and ^{13}C NMR spectra were recorded on a Bruker Avance 500-MHz spectrometer operating at 500.13 MHz for ^1H NMR and 125.75 MHz for ^{13}C using a 5-mm z-gradient TBI probe. The data were processed using TopSpin 3.0 software. 1D ^1H NMR spectra were acquired by using a zgpr pulse sequence (with water suppression). Spectra were performed at 298 K with no purification step, for all mutants.

Dynamic Light Scattering (DLS) was performed after the partial purification of the polymer using a 14 kDa cut-off cellulose dialysis tubing (Sigma) against water. The analysis was performed at 1% (w/v) in water with a DynaPro Nanostar instrument (Wyatt Corporation). Samples were pre-equilibrated

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

for 1 mn at 25°C and analyzed with a refractive index of 1.33, with 90° scattering optics at 658 nm.

Differential Scanning Fluorimetry (DSF) was performed with 7 µM of pure enzyme in 50 mM sodium acetate buffer pH 5.75 supplemented with 0.5 $g.L^{-1}$ of calcium chloride and 10 X of SYPRO orange (Life Technologies). A ramp from 20 to 80°C was applied with 0.3°C increments at the rate of 0.3°C per second on a CX100 Thermal Cycler (Biorad).

Affinity gel electrophoresis

4 µg of purified enzyme were loaded in 6.5% (w/v) acrylamide gels containing from 0 to 0.9% (w/v) of dextran 70 kDa, dextran 2,000 kDa (Sigma) or alternan produced by ASRΔ2 in the conditions described above. Alternan was purified by dialysis against water using a 14 kDa cut-off cellulose dialysis tubing (Sigma). Bovine Serum Albumine (BSA) in 1% (w/v) of NaCl and ladder All Blue Standard were used as negative control (BioRad). Migration was performed in mini PROTEAN system (BioRad) during 30 minutes at 65V followed by 2 hours at 95V in ice. Gels were stained with Colloidal Blue.

Multiple Sequence Alignment

Sequence alignment of putative sugar binding pockets was performed using Clustal Omega (https://www.ebi.ac.uk/Tools/msa/clustalo/), was inspected and corrected manually using the structural superimposition of pockets V-A, V-B of ASR (PDB ID: 6HVG) and DSR-M (5NGY) and pocket V-L of GBD-CD2 (4TVD) to align the first aromatic residue. Then, the alignment was submitted to WebLogo3 (http://weblogo.threeplusone.com/) (53).

Data Availability Statement

The crystal structures have been deposited in the Protein Data Bank (www.rcsb.org) and are freely accessible under the accession numbers 6SZI, 6SYQ, 6T16, 6T18 and 6T1P. All the other data are contained within the manuscript.

ACKNOWLEDGEMENTS

We thank Metasys, the Metabolomics & Fluxomics Center at the Toulouse Biotechnology Institute (France) for the NMR experiments. We are grateful to the European Synchrotron Radiation Facility (ESRF, Grenoble, France), the ALBA synchrotron (Barcelona, Spain) and the Structural Biophysics team of the Institute of Pharmacology and Structural Biology (IPBS, Toulouse, France) for access to their crystallization facility and help in synchrotron data collection. We also thank the ICEO facility, part of the Integrated Screening Platform of Toulouse (PICT), for access to HPLC and protein purification facilities. Technical assistance provided by Valérie Bourdon of the ICT-FR 2599 (Toulouse, France - ict.ups-tlse.fr) is gratefully acknowledged.

CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest with the contents of this article.

REFERENCES

1. Freitas, F., Alves, V. D., and Reis, M. A. M. (2011) Advances in bacterial exopolysaccharides: from production to biotechnological applications. *Trends Biotechnol.* **29**, 388–398
2. Moscovici, M. (2015) Present and future medical applications of microbial exopolysaccharides. *Front. Microbiol.* **6**, 1012
3. Lombard, V., Golaconda Ramulu, H., Drula, E., Coutinho, P. M., and Henrissat, B. (2014) The carbohydrate-active enzymes database (CAZy) in 2013. *Nucleic Acids Res.* **42**, D490-495
4. Moulis, C., André, I., and Remaud-Siméon, M. (2016) GH13 amylosucrases and GH70 branching sucrases, atypical enzymes in their respective families. *Cell. Mol. Life Sci.* **73**, 2661–2679
5. Leemhuis, H., Pijning, T., Dobruchowska, J. M., van Leeuwen, S. S., Kralj, S., Dijkstra, B. W., and Dijkhuizen, L. (2013) Glucansucrases: three-dimensional structures, reactions, mechanism, α-

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

glucan analysis and their implications in biotechnology and food applications. *J. Biotechnol.* **163**, 250–272

6. Soetaert, W., Schwengers, D., Buchholz, K., and Vandamme, E. J. (1995) A wide range of carbohydrate modifications by a single micro-organism: *Leuconostoc mesenteroides*. in *Progress in Biotechnology* (Petersen, S. B., Svensson, B., and Pedersen, S. eds), pp. 351–358, Carbohydrate Bioengineering, Elsevier, **10**, 351–358
7. Dennes, T. J., Perticone, A. M., and Paullin, J. L. (2015) Cationic poly alpha-1,3-glucan ethers. *U.S. Patent No 9,957,334*. Washington, DC: U.S. Patent and Trademark Office.
8. Paullin, J. L., Perticone, A. M., Kasat, R. B., and Dennes, T. J. (2014) Preparation of poly alpha-1,3-glucan ethers. *U.S. Patent No 9,139,718*. Washington, DC: U.S. Patent and Trademark Office.
9. Vujičić-Žagar, A., Pijning, T., Kralj, S., López, C. A., Eeuwema, W., Dijkhuizen, L., and Dijkstra, B. W. (2010) Crystal structure of a 117 kDa glucansucrase fragment provides insight into evolution and product specificity of GH70 enzymes. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 21406–21411
10. Ito, K., Ito, S., Shimamura, T., Weyand, S., Kawarasaki, Y., Misaka, T., Abe, K., Kobayashi, T., Cameron, A. D., and Iwata, S. (2011) Crystal structure of glucansucrase from the dental caries pathogen *Streptococcus mutans*. *Journal of Molecular Biology*. **408**, 177–186
11. Pijning, T., Vujičić-Žagar, A., Kralj, S., Dijkhuizen, L., and Dijkstra, B. W. (2012) Structure of the α-1,6/α-1,4-specific glucansucrase GTFA from *Lactobacillus reuteri* 121. *Acta Crystallogr Sect F Struct Biol Cryst Commun*. **68**, 1448–1454
12. Claverie, M., Cioci, G., Vuillemin, M., Monties, N., Roblin, P., Lippens, G., Remaud-Siméon, M., and Moulis, C. (2017) Investigations on the Determinants Responsible for Low Molar Mass Dextran Formation by DSR-M Dextransucrase. *ACS Catal.* **7**, 7106–7119
13. Molina, M., Moulis, C., Monties, N., Pizzut-Serin, S., Guieysse, D., Morel, S., Cioci, G., and Remaud Siméon, M. (2019) Deciphering an undecided enzyme: investigations of the structural determinants involved in the linkage specificity of alternansucrase. *ACS Catal.* **9**, 2222–2237
14. Brison, Y., Malbert, Y., Czaplicki, G., Mourey, L., Remaud-Siméon, M., and Tranier, S. (2016) Structural Insights into the Carbohydrate Binding Ability of an α-(1→2) Branching Sucrase from Glycoside Hydrolase Family 70. *J. Biol. Chem.* **291**, 7527–7540
15. Claverie, M., Cioci, G., Vuillemin, M., Bondy, P., Remaud-Simeon, M., and Moulis, C. (2020) Processivity of dextransucrases synthesizing very high molar mass dextran is mediated by sugar-binding pockets in domain V. *J. Biol. Chem.* 10.1074/jbc.RA119.011995
16. Davies, G. J., Wilson, K. S., and Henrissat, B. (1997) Nomenclature for sugar-binding subsites in glycosyl hydrolases. *Biochem J*. **321**, 557–559
17. Claverie, M., Cioci, G., Guionnet, M., Schörghuber, J., Lichtenecker, R., Moulis, C., Remaud-Siméon, M., and Lippens, G. (2019) Futile Encounter Engineering of the DSR-M Dextransucrase Modifies the Resulting Polymer Length. *Biochemistry*. **58**, 2853–2859
18. Irague, R., Tarquis, L., André, I., Moulis, C., Morel, S., Monsan, P., Potocki-Véronèse, G., and Remaud-Siméon, M. (2013) Combinatorial Engineering of Dextransucrase Specificity. *PLoS One*. 10.1371/journal.pone.0077837
19. Meng, X., Pijning, T., Tietema, M., Dobruchowska, J. M., Yin, H., Gerwig, G. J., Kralj, S., and Dijkhuizen, L. (2017) Characterization of the glucansucrase GTF180 W1065 mutant enzymes producing polysaccharides and oligosaccharides with altered linkage composition. *Food Chem.* **217**, 81–90
20. Cuyvers, S., Dornez, E., Delcour, J. A., and Courtin, C. M. (2012) Occurrence and functional significance of secondary carbohydrate binding sites in glycoside hydrolases. *Crit. Rev. Biotechnol.* **32**, 93–107
21. Wilkens, C., Svensson, B., and Møller, M. S. (2018) Functional Roles of Starch Binding Domains and Surface Binding Sites in Enzymes Involved in Starch Biosynthesis. *Front. Plant Sci.* 10.3389/fpls.2018.01652
22. Cockburn, D., Wilkens, C., Ruzanski, C., Andersen, S., Willum Nielsen, J., Smith, A. M., Field, R. A., Willemoës, M., Abou Hachem, M., and Svensson, B. (2014) Analysis of surface binding sites

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

(SBSs) in carbohydrate active enzymes with focus on glycoside hydrolase families 13 and 77 — a mini-review. *Biologia*. **69**, 705–712
23. Cockburn, D., and Svensson, B. (2013) Surface binding sites in carbohydrate active enzymes: an emerging picture of structural and functional diversity. in *Carbohydrate Chemistry*, Lindhorst, T.K., and Rauter, A.P., pp. 204–221, Chemical and Biological Approaches, Cambridge: Royal Society of Chemistry, **39**, 204–221
24. Skov, L. K., Mirza, O., Sprogøe, D., Dar, I., Remaud-Simeon, M., Albenne, C., Monsan, P., and Gajhede, M. (2002) Oligosaccharide and Sucrose Complexes of Amylosucrase STRUCTURAL IMPLICATIONS FOR THE POLYMERASE ACTIVITY. *J. Biol. Chem.* **277**, 47741–47747
25. Bounaix, M.-S., Gabriel, V., Robert, H., Morel, S., Remaud-Siméon, M., Gabriel, B., and Fontagné-Faucher, C. (2010) Characterization of glucan-producing *Leuconostoc* strains isolated from sourdough. *Int. J. Food Microbiol.* **144**, 1–9
26. Côté, G. L., and Robyt, J. F. (1982) Isolation and partial characterization of an extracellular glucansucrase from *Leuconostoc mesenteroides* NRRL B-1355 that synthesizes an alternating (1→6),(1→3)-α-D-glucan. *Carbohydr. Res.* **101**, 57–74
27. Joucla, G., Pizzut-Serin, S., Monsan, P., and Remaud-Siméon, M. (2006) Construction of a fully active truncated alternansucrase partially deleted of its carboxy-terminal domain. *FEBS Lett.* **580**, 763–768
28. Moulis, C., Joucla, G., Harrison, D., Fabre, E., Potocki-Veronese, G., Monsan, P., and Remaud-Siméon, M. (2006) Understanding the polymerization mechanism of glycoside-hydrolase family 70 glucansucrases. *J. Biol. Chem.* **281**, 31254–31267
29. Côté, G. L. (1992) Low-viscosity α-d-glucan fractions derived from sucrose which are resistant to enzymatic digestion. *Carbohydr. Polym.* **19**, 249–252
30. Leathers, T. D., Nunnally, M. S., and Côté, G. L. (2009) Modification of alternan by dextranase. *Biotechnol. Lett.* **31**, 289–293
31. Wangpaiboon, K., Padungros, P., Nakapong, S., Charoenwongpaiboon, T., Rejzek, M., Field, R. A., and Pichyangkura, R. (2018) An α-1,6-and α-1,3-linked glucan produced by *Leuconostoc citreum* ABK-1 alternansucrase with nanoparticle and film-forming properties. *Sci. Rep.* **8**, 8340
32. Charoenwongpaiboon, T., Supraditaporn, K., Klaimon, P., Wangpaiboon, K., Pichyangkura, R., Issaragrisil, S., and Lorthongpanich, C. (2019) Effect of alternan versus chitosan on the biological properties of human mesenchymal stem cells. *RSC Adv.* **9**, 4370–4379
33. Côté, G. L., Holt, S. M., and Miller-Fosmore, C. (2003) Prebiotic Oligosaccharides via Alternansucrase Acceptor Reactions. in *Oligosaccharides in Food and Agriculture*, pp. 76–89, ACS Symposium Series, American Chemical Society, Washington, DC, **849**, 76–89
34. Hernandez-Hernandez, O., Côté, G. L., Kolida, S., Rastall, R. A., and Sanz, M. L. (2011) In vitro fermentation of alternansucrase raffinose-derived oligosaccharides by human gut bacteria. *J. Agric. Food Chem.* **59**, 10901–10906
35. Holt, S. M., Miller-Fosmore, C. M., and Côté, G. L. (2005) Growth of various intestinal bacteria on alternansucrase-derived oligosaccharides. *Lett. Appl. Microbiol.* **40**, 385–390
36. Sanz, M. L., Côté, G. L., Gibson, G. R., and Rastall, R. A. (2005) Prebiotic properties of alternansucrase maltose-acceptor oligosaccharides. *J. Agric. Food Chem.* **53**, 5911–5916
37. Biely, P., Côté, G. L., and Burgess-Cassler, A. (1994) Purification and Properties of Alternanase, a Novel Endo-α-1,3-α-1,6-d-Glucanase. *Eur. J. Biochem.* **226**, 633–639
38. Brison, Y., Pijning, T., Malbert, Y., Fabre, É., Mourey, L., Morel, S., Potocki-Véronèse, G., Monsan, P., Tranier, S., Remaud-Siméon, M., and Dijkstra, B. W. (2012) Functional and structural characterization of α-(1→2) branching sucrase derived from DSR-E glucansucrase. *J. Biol. Chem.* **287**, 7915–7924
39. Asensio, J. L., Ardá, A., Cañada, F. J., and Jiménez-Barbero, J. (2013) Carbohydrate–aromatic interactions. *Acc. Chem. Res.* **46**, 946–954
40. Hudson, K. L., Bartlett, G. J., Diehl, R. C., Agirre, J., Gallagher, T., Kiessling, L. L., and Woolfson, D. N. (2015) Carbohydrate–Aromatic Interactions in Proteins. *J. Am. Chem. Soc.* **137**, 15152–15160

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

41. Côté, G. L., and Sheng, S. (2006) Penta-, hexa-, and heptasaccharide acceptor products of alternansucrase. *Carbohydr. Res.* **341**, 2066–2072
42. Albenne, C., Skov, L. K., Tran, V., Gajhede, M., Monsan, P., Remaud-Siméon, M., and André-Leroux, G. (2007) Towards the molecular understanding of glycogen elongation by amylosucrase. *Proteins: Struct., Funct., Bioinf.* **66**, 118–126
43. Kabsch, W. (2010) XDS. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **66**, 125–132
44. Winn, M. D., Ballard, C. C., Cowtan, K. D., Dodson, E. J., Emsley, P., Evans, P. R., Keegan, R. M., Krissinel, E. B., Leslie, A. G. W., McCoy, A., McNicholas, S. J., Murshudov, G. N., Pannu, N. S., Potterton, E. A., Powell, H. R., Read, R. J., Vagin, A., and Wilson, K. S. (2011) Overview of the CCP4 suite and current developments. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **67**, 235–242
45. Emsley, P., and Cowtan, K. (2004) Coot: model-building tools for molecular graphics. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **60**, 2126–2132
46. Murshudov, G. N., Skubák, P., Lebedev, A. A., Pannu, N. S., Steiner, R. A., Nicholls, R. A., Winn, M. D., Long, F., and Vagin, A. A. (2011) REFMAC5 for the refinement of macromolecular crystal structures. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **67**, 355–367
47. Williams, C. J., Headd, J. J., Moriarty, N. W., Prisant, M. G., Videau, L. L., Deis, L. N., Verma, V., Keedy, D. A., Hintze, B. J., Chen, V. B., Jain, S., Lewis, S. M., Arendall, W. B., Snoeyink, J., Adams, P. D., Lovell, S. C., Richardson, J. S., and Richardson, D. C. (2018) MolProbity: More and better reference data for improved all-atom structure validation. *Protein Sci.* **27**, 293–315
48. Lütteke, T., Frank, M., and von der Lieth, C.-W. (2005) Carbohydrate Structure Suite (CSS): analysis of carbohydrate 3D structures derived from the PDB. *Nucleic Acids Res.* **33**, D242-246
49. Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., Goodsell, D. S., and Olson, A. J. (2009) AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* **30**, 2785–2791
50. Sanner, M. F. (1999) Python: a programming language for software integration and development. *J. Mol. Graph. Model.* **17**, 57–61
51. Nivedha, A. K., Thieker, D. F., Makeneni, S., Hu, H., and Woods, R. J. (2016) Vina-Carb: Improving Glycosidic Angles during Carbohydrate Docking. *J. Chem. Theory Comput.* **12**, 892–901
52. Miller, G. L. (1959) Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. *Anal. Chem.* **31**, 426–428
53. Crooks, G. E., Hon, G., Chandonia, J.-M., and Brenner, S. E. (2004) WebLogo: a sequence logo generator. *Genome Res.* **14**, 1188–1190
54. Robert, X., and Gouet, P. (2014) Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Res.* **42**, 320–324

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

Table 1: Biochemical data of the characterized mutants. Reaction from sucrose only at 30°C with 1 U.mL^{-1} of pure enzyme and sodium acetate buffer 50 mM pH 5.75. Specific activity of ASRΔ2: 30.2 ± 1.0 U.mg^{-1}. Specific activity was determined in triplicate. T_m was determined by DSF. *: NMR was performed on crude reaction medium.

	Residual activity (%)	ΔT_m with the wild type enzyme (°C)	% of α-1,3 linkages (NMR)*	% of α-1,6 linkages (NMR)*	% of polymer formed (area HPSEC)	t_R polymer (min, HPSEC)	Hydrolysis (%)
	Wild type						
ASRΔ2	100 ± 3.3	0	35	65	31.5 ± 1.6	37 ± 0.2	4.4 ± 0.4
	Mutations in Domain V						
ASRΔ2 Y158A (pocket V-A)	80.2 ± 3.0	+0.1	34	66	27.2	36.8	4.7
ASRΔ2 Y241A (pocket V-B)	78.4 ± 3.2	+0.2	35	65	28.0	36.6	4.7
ASRΔ2 Y158A+Y241A (pockets V-A+V-B)	77.5 ± 2.4	+0.2	33	67	27.6	36.5	4.7
ASRΔ5[14]	79.1 ± 2.7	-2	30	70	4.5	38.2	5.8
	Mutations in SBS-A1						
ASRΔ2 N703A	95.3 ± 6.0	-0.1	35	65	33.5	36.9	4.3
ASRΔ2 S713A	95.8 ± 3.3	-0.3	35	65	31.7	36.6	4.3
ASRΔ2 W716A	76.4 ± 7.0	-1.0	35	65	30.0	36.4	5.0
ASRΔ2 Q700A	80.5 ± 2.8	-0.2	35	65	20.4	35.7	5.6
ASRΔ2 Y717A	80.2 ± 2.4	-0.1	33	67	18.8	35.6	5.2
ASRΔ2 Q700+Y717A	26.1 ± 7.7	-0.9	35	65	18.3	35.5	5.8
	Mutations in Domain V + SBS-A1						
ASRΔ2 Y158A+Y717A	62.9 ± 3.9	-0.1	32	68	13.3	35.3	5.5
ASRΔ2 Y241A+Y717A	78.3 ± 5.8	-0.2	31	69	8.5	35.4	5.9
ASRΔ5 Y717A	63.1 ± 3.6	-2.4	30	70	2.3	36.4	6.2

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

Table 2: Crystallographic statistics. Values in parentheses refer to the high resolution shell. *Soakings, in particular those with longer ligands, affected diffraction quality and refinement statistics.

Enzyme	**pET53-His-ASRΔ2-Strep**				
Ligand	Isomaltose	Isomaltotriose	Panose	Oligoalternan	Isomaltononaose
PDB ID	6SZI	6SYQ	6T16	6T18	6T1P
Soaking concentration and duration	100 mM 1 minute	100 mM 5 minutes	100 mM 5 minutes	100 g/L 5 minutes	100 mM 5 minutes
Data collection					
Wavelength (Å)	0.9734	0.9793	0.9979	0.9793	0.9979
Space group	$P2_12_12_1$	$P2_12_12_1$	$P2_12_12_1$	$P2_12_12_1$	$P2_12_12_1$
Molecules per asymmetric unit	2	2	2	2	2
Cell constants a, b, c (Å)	100.74 134.69 236.04	100.72 135.65 238.94	101.00 134.30 235.37	100.87 135.65 239.42	100.57 135.21 237.20
Resolution (Å)	50.00-3.00	50.00-3.00	50.00-3.10	50.00-3.15	50.00-3.50
Measured reflections	440819	352514	275779	255328	187213
Unique reflections	65077	61391	58714	57606	39240
Data completeness %	99.9	92.4	99.7	99.9	94.9
Rmerge	0.11 (0.77)	0.07 (0.77)	0.10 (0.78)	0.09 (0.72)	0.14 (0.76)*
< I/σ (I) >	6.1 (1.0)	8.5 (1.0)	6.7 (1.0)	7.5 (1.1)	4.9 (1.0)
$CC_{1/2}$	0.99 (0.85)	0.99 (0.82)	0.99 (0.80)	0.99 (0.71)	0.99 (0.71)
Wilson B-factor (Å²)	74.2	73.7	83.0	78.5	99.4*
Refinement					
R_{work}/R_{free}	0.207/0.237	0.201/0.237	0.223/0.253	0.200/0.226	0.206/0.266*
RMSD bonds (Å)	0.010	0.010	0.010	0.008	0.009
RMSD angles (°)	1.332	1.308	1.282	1.222	1.250
Ramachandran's:					
Favored/Allowed/Outliers(%)	96/4/0	96/4/0	96/4/0	96/4/0	96/4/0
Number of atoms					
Protein	19318	19242	19251	19221	19271
Calcium	2	2	2	2	2
Carbohydrates	69	103	136	146	78
Average B-factor (Å²)	82.0	73.7	105.6	90.0	108.2*
CC_{all} / CC_{free}	0.94/0.92	0.95/0.93	0.94/0.92	0.94/0.92	0.94/0.89
Clashscore (percentile)	3 (100th)	3 (100th)	4 (100th)	3 (100th)	3 (100th)

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

FIGURES

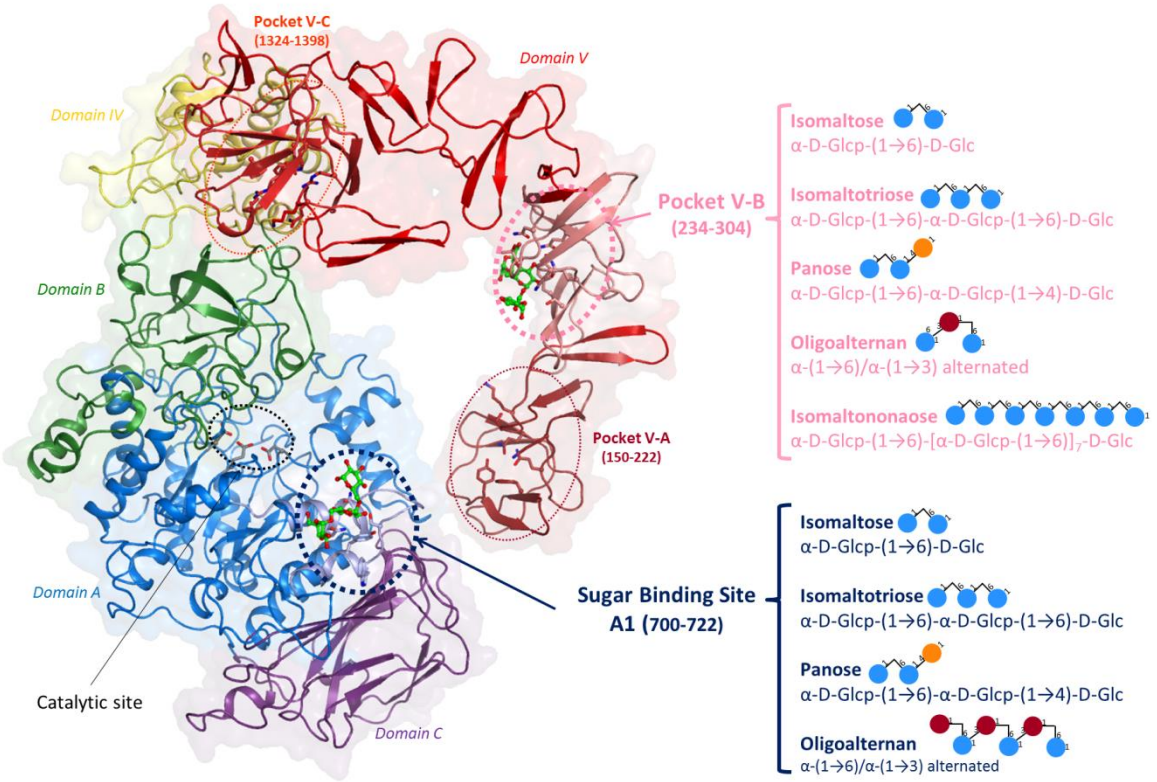

Figure 1: Identification of the new sugar binding sites: Surface Binding Site A1 (SBS-A1) and Sugar binding pocket V-B in ASR. The list of ligands found at these binding sites is indicated on the right panel with an illustration of the structure visible in the complexes. Grey: catalytic residues Asp635, Glu673 and Asp767. See Figures S4 and S7 for electron density maps sugar binding pocket V-B and in surface binding site A1 respectively.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

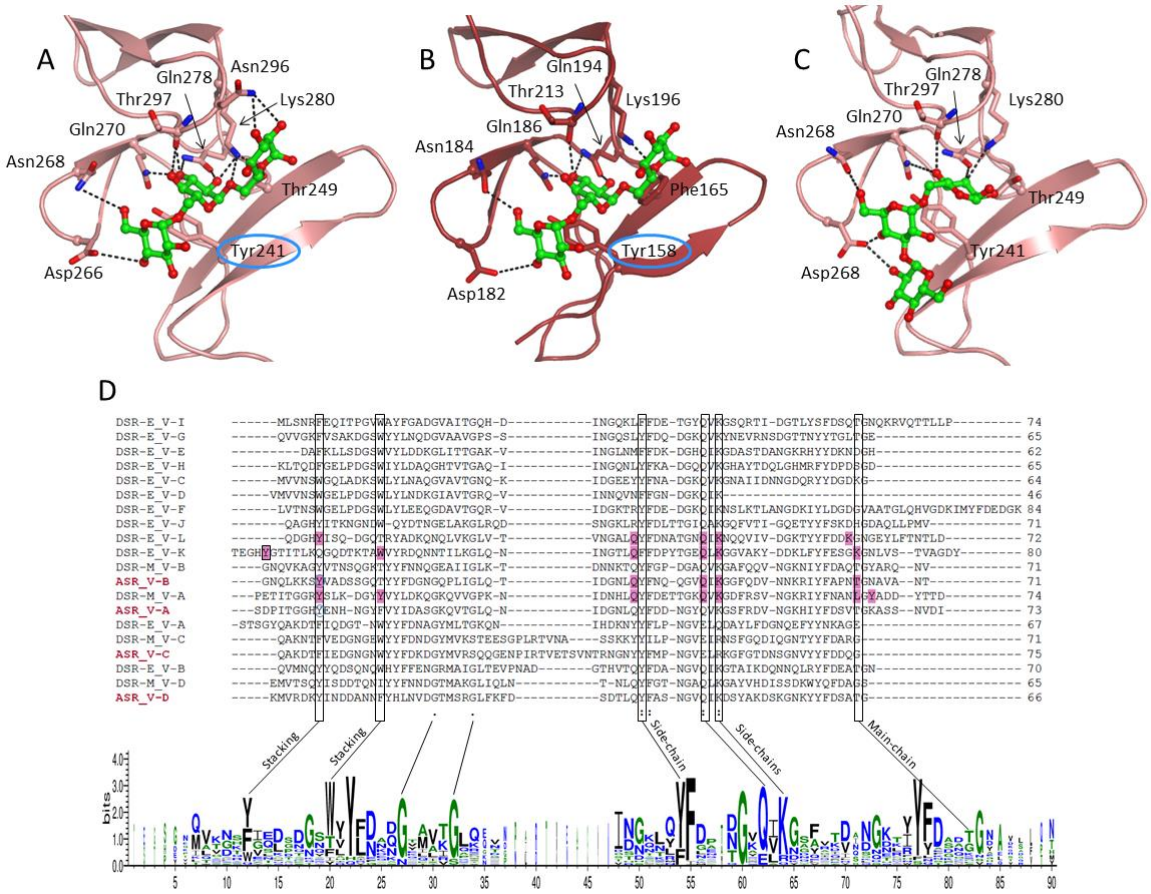

Figure 2: Complexes in sugar binding pockets. (A) Crystal structure of isomaltotriose binding in pocket V-B (PDB ID: 6SYQ). **(B)** Possible location of isomaltotriose in pocket V-A as superimposed from pocket V-B. **(C)** Crystal structure of the OA bound in pocket V-B (PDB ID: 6T18). The probable interaction network is displayed as dashed lines. **(D)** Sequence alignment of the sugar binding pockets identified in ASR, DSR-M and DSR-E glucansucrases. Pink highlighted residues were shown to directly interact with sugar ligands in 3D structures. Blue circled residues have been mutated to Ala in this study.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

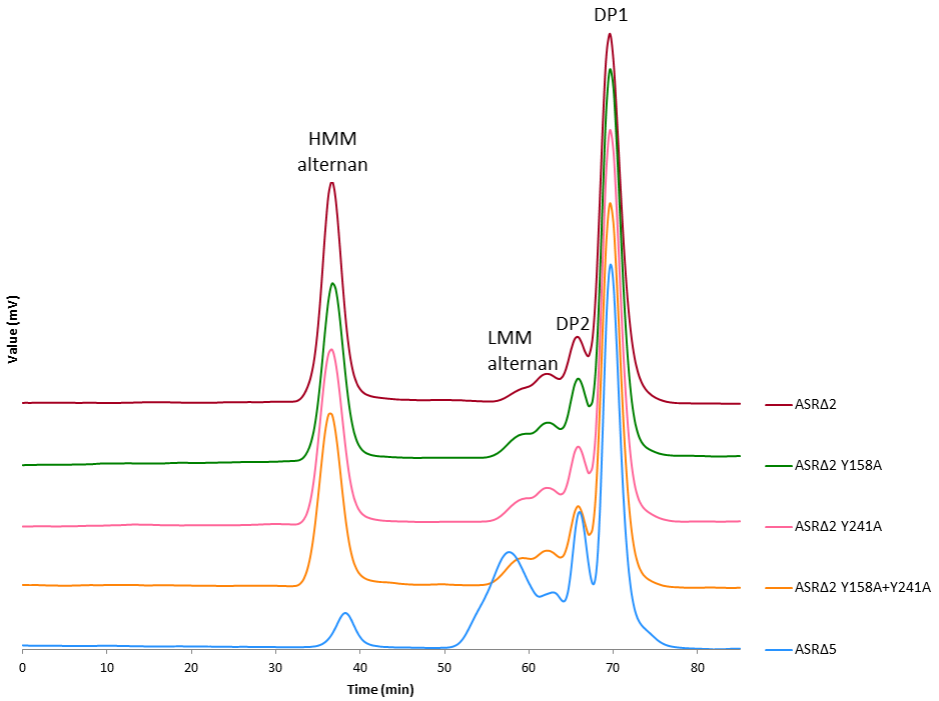

Figure 3: HPSEC analysis of the mutants in the sugar binding pockets of domain V. Reaction from sucrose at 30°C with 1 U.mL^{-1} of pure enzyme and sodium acetate buffer 50 mM pH 5.75.

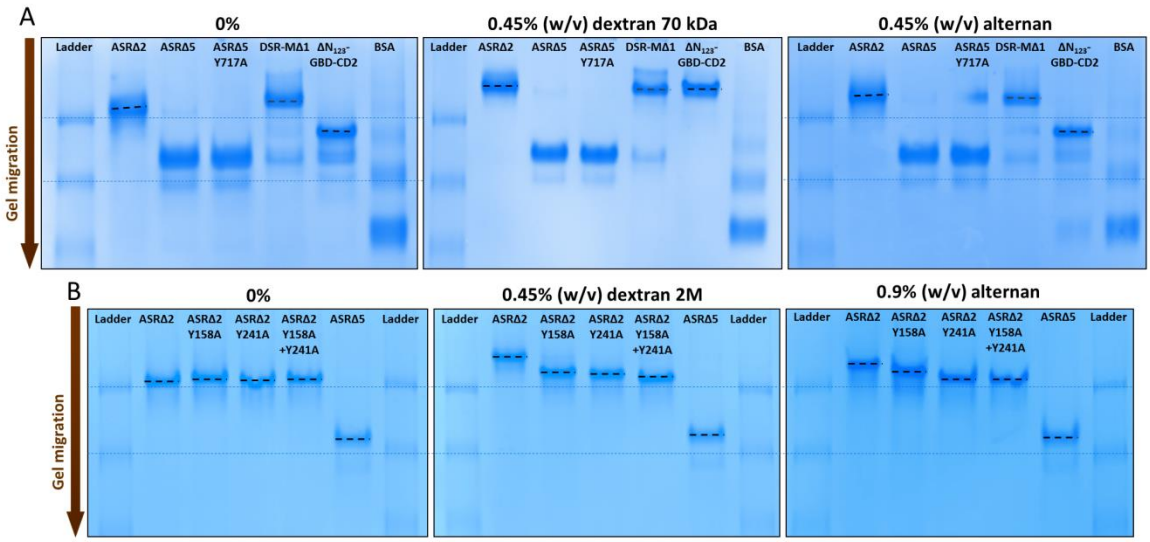

Figure 4: Affinity gel electrophoresis. **(A)** Affinity gel electrophoresis of ASRΔ2, ASRΔ5, ASRΔ5-Tyr717Ala. DSR-MΔ1 and ΔN_{123}-GBD-CD2 (branching sucrase) are used as a positive control (12, 14). BSA 1% and Protein standard (ladder) are used as negative controls. Gels were made in the presence or absence of 0.45% (w/v) dextran 70,000 g.mol^{-1} or alternan. **(B)** Affinity gel electrophoresis of ASRΔ2, ASRΔ2-Tyr158Ala, ASRΔ2-Tyr241Ala, ASRΔ2-Tyr158Ala+Tyr241Ala and ASRΔ5. Protein standard (ladder) is used as negative control. Gels were made in the presence or absence of 0.45% (w/v) dextran 2,000,000 g.mol^{-1} or 0.9% (w/v) alternan.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

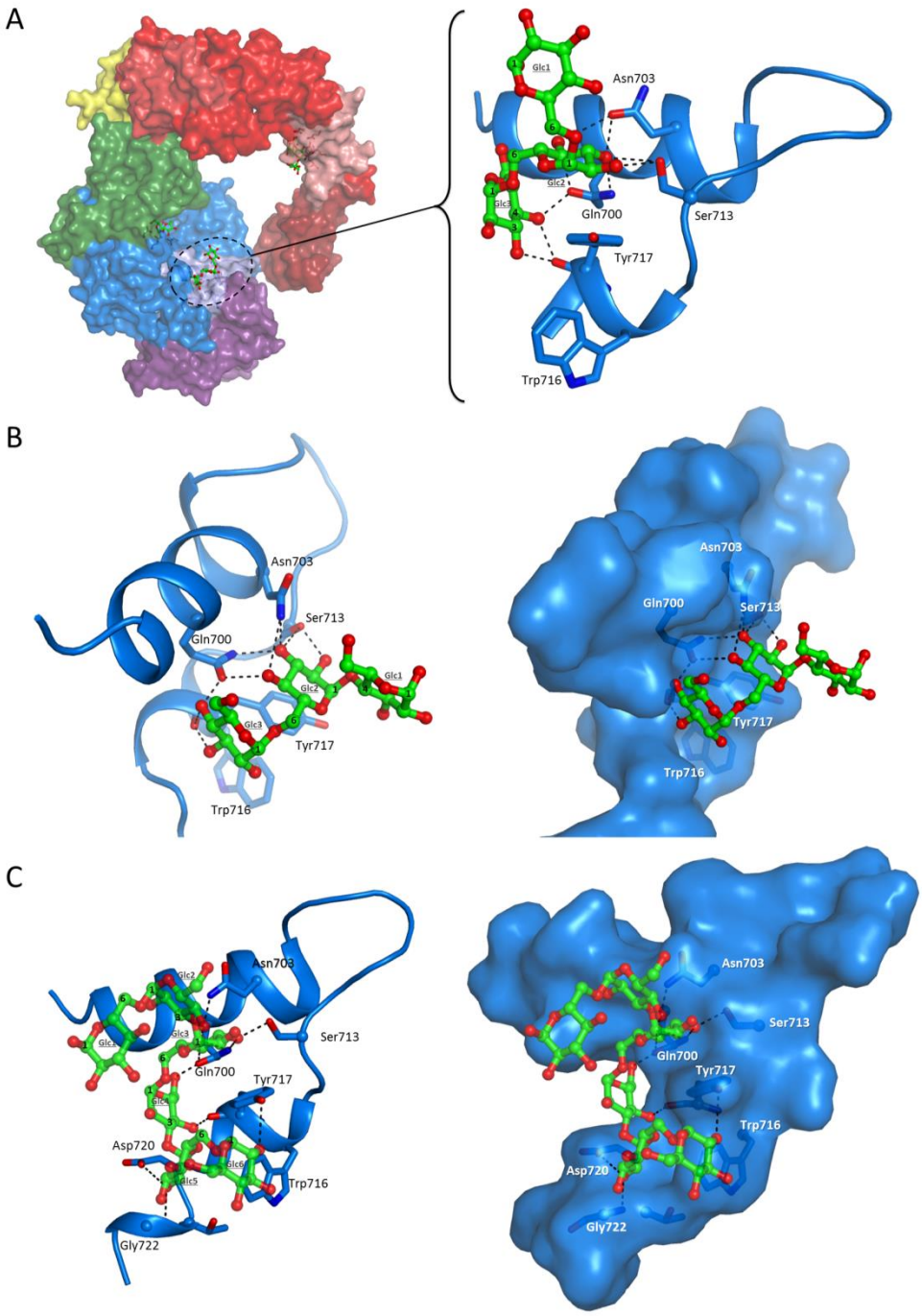

Figure 5: Complexes in Surface Binding Site A1. (A) Crystal structure of isomaltotriose binding in Surface Binding Site A1 (PDB ID: 6SYQ). Sucrose molecule in the active site was positioned from the superimposition of the ASR catalytic domain with GTF-180:sucrose complex (PDB ID: 3HZ3). **(B)** Crystal structure of panose binding in Surface Binding Site A1 (PDB ID: 6T16). **(C)** Crystal structure of OA binding in Surface Binding Site A1 (PDB ID: 6T18).

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

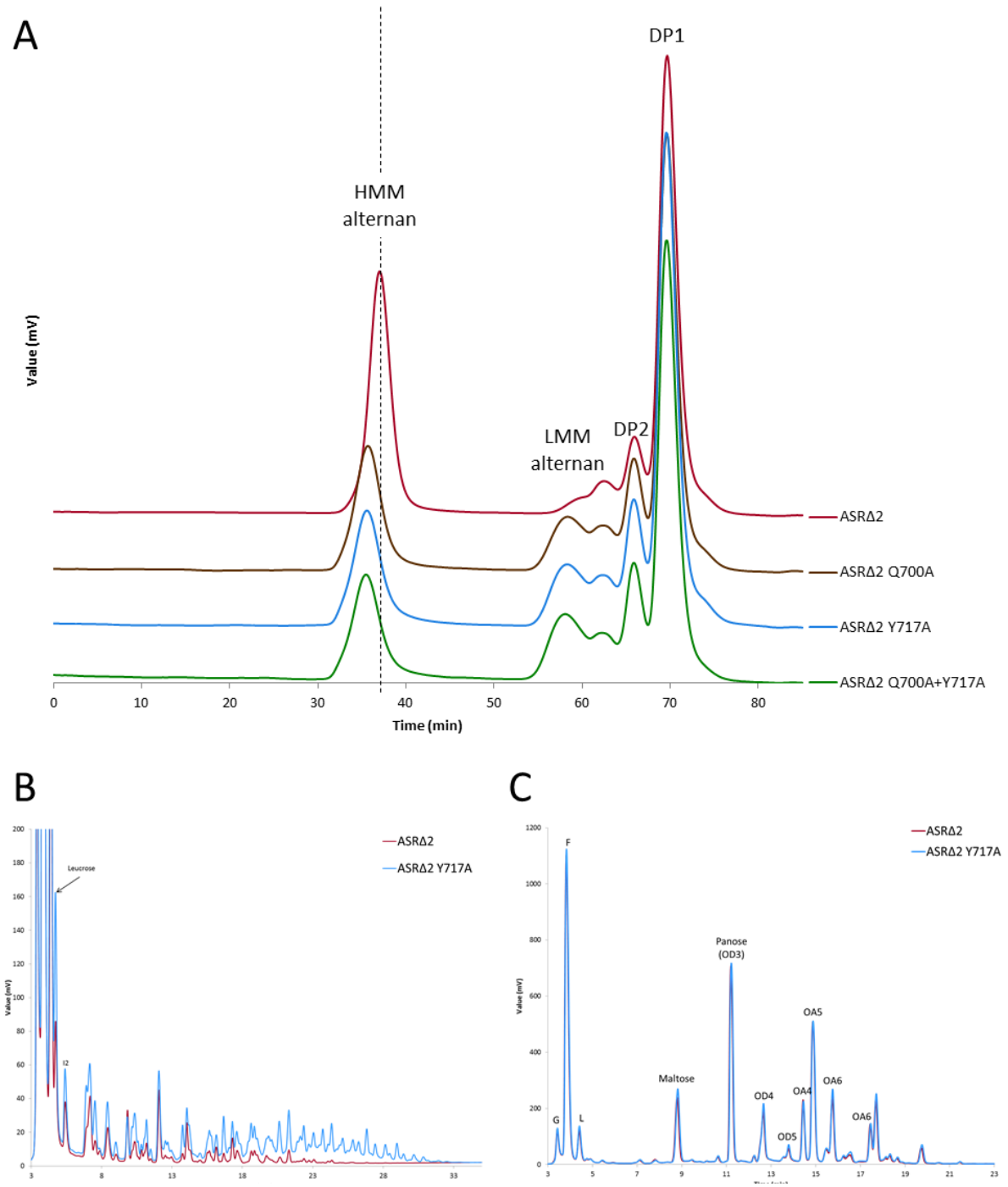

Figure 6: Reaction product analysis of SBS-A1 mutants. (A) HPSEC chromatograms of the products synthesized by SBS-A1 mutants. **(B)** HPAEC-PAD chromatogram of the oligoalternans produced from sucrose with ASRΔ2 and mutant Y717A. Reaction from sucrose at 30°C with 1 U.mL-1 of pure enzyme and sodium acetate buffer 50 mM pH 5.75. **(C)** HPAEC-PAD chromatogram of the acceptor reaction products from maltose, with ASRΔ2 and mutant Y717A. Reaction from sucrose and maltose with sucrose:maltose mass ratio 2:1 at 30°C with 1 U.mL-1 of pure enzyme and sodium acetate buffer 50 mM pH 5.75. G: glucose, F: fructose, L: leucrose. For detailed structures, see Experimental procedures.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

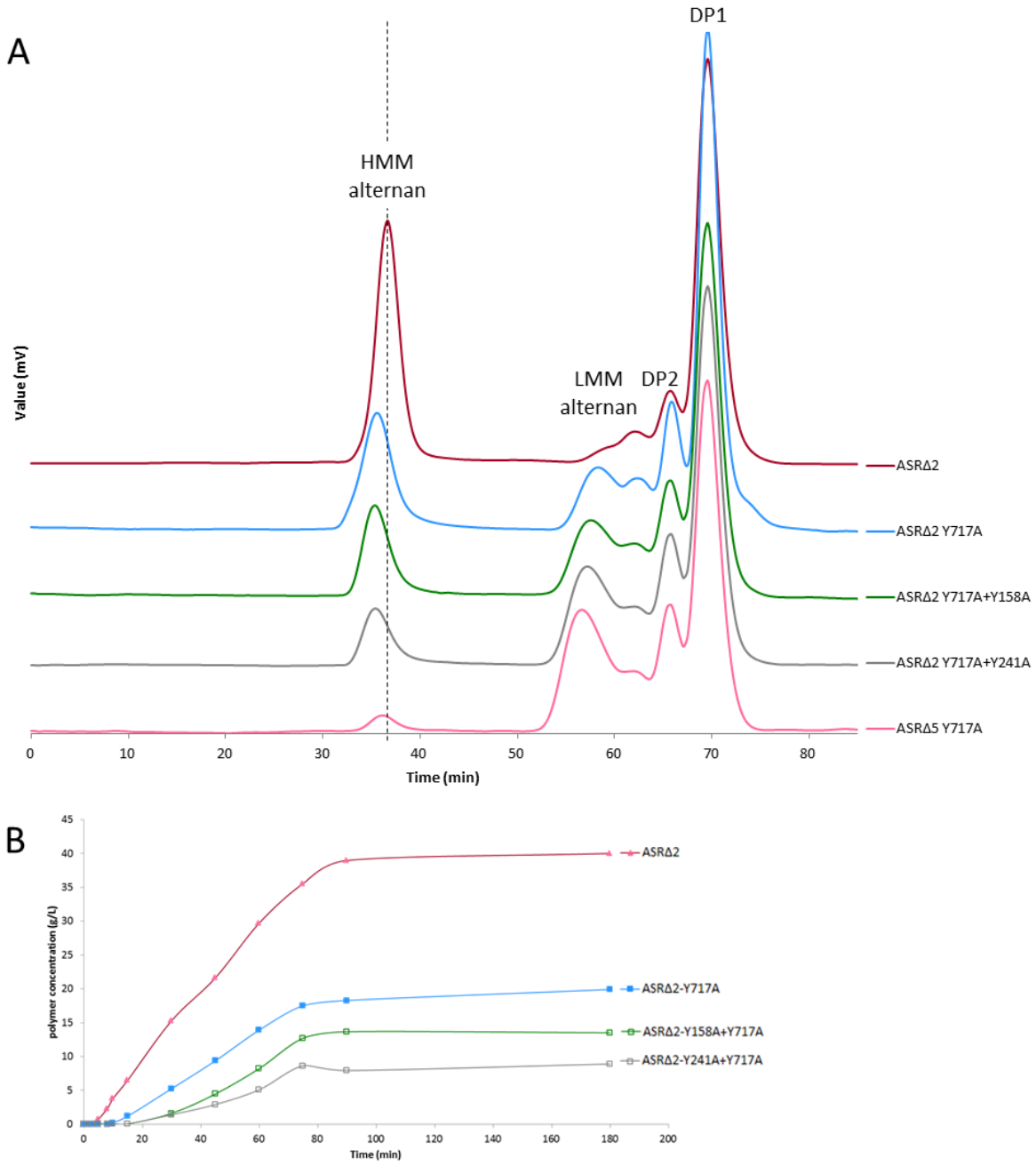

Figure 7: Analysis of mutants combined to the Tyr717Ala mutation. (A) HPSEC chromatograms of alternan populations produced with Tyr717Ala and the double mutants Tyr717Ala+Tyr158Ala mutant (pocket V-A) and Tyr717Ala+Tyr241Ala mutant (pocket V-B). Reaction from sucrose at 30°C with 1 U.mL-1 of pure enzyme and sodium acetate buffer 50 mM pH 5.75. **(B)** Monitoring of polymer formation with time. Reaction from sucrose at 30°C with 1 U.mL-1 of pure enzyme and sodium acetate buffer 50 mM pH 5.75. Production rate was calculated from 5 minutes to 75 minutes (R^2 of 0.997) and from 10 minutes to 75 minutes (R^2 of 0.999) for ASRΔ2 and ASRΔ2 Y717A, respectively.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

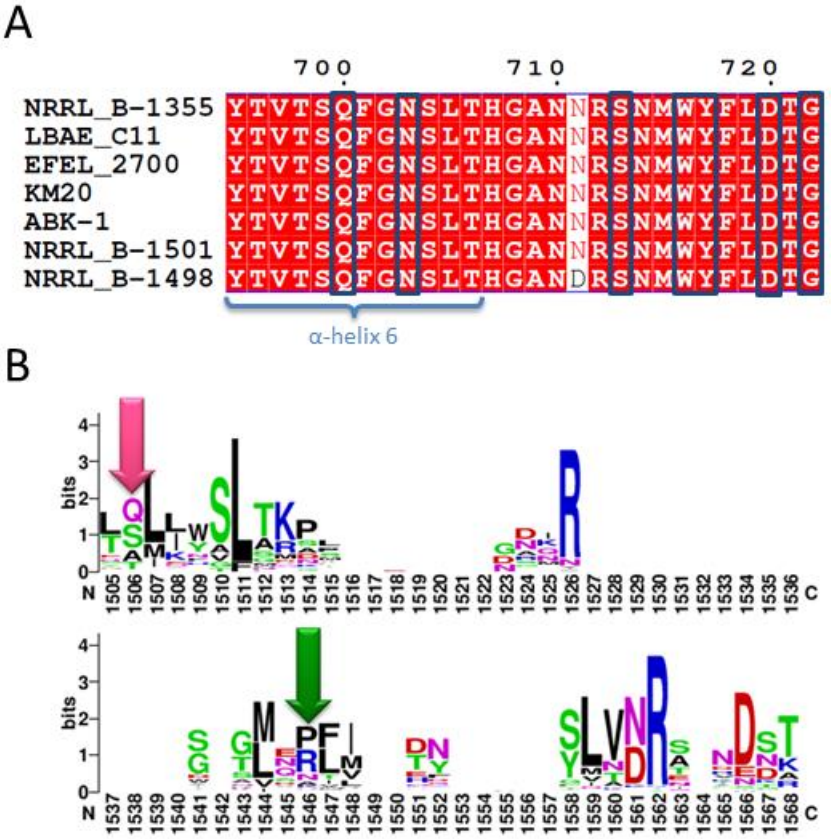

Figure 8: SBS-A1 sequence comparison. (A) Alignment of the residues corresponding to SBS-A1 (in black boxes) in all characterized and putative alternansucrases. Only the strain name is indicated. Species: Leuconostoc citreum or mensenteroides. Alignment created with ENDscript 2 (54) **(B)** Web logo of all GH70 characterized enzymes. Pink arrow corresponds to Gln700 position and green arrow, to Tyr717 position.

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020

A specific oligosaccharide-binding site in alternansucrase catalytic domain mediates alternan elongation

Manon Molina, Claire Moulis, Nelly Monties, David Guieysse, Sandrine Morel, Gianluca Cioci and Magali Remaud-Simeon

J. Biol. Chem. published online May 14, 2020

Access the most updated version of this article at doi: 10.1074/jbc.RA120.013028

Alerts:
- When this article is cited
- When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

Downloaded from http://www.jbc.org/ at INRAE Inst Natl Recherche pour l'Agriculture, l'Alimentation et l'Environnement on August 21, 2020