

# Effectiveness of two intranasal vaccines for the control of bovine respiratory disease in newborn beef calves: A randomized non-inferiority multicentre field trial

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### 1 Original Article

2 3 4 Effectiveness of two intranasal vaccines for the control of bovine respiratory disease in 5 newborn beef calves: A randomized non-inferiority multicentre field trial 6 7 N. Masset <sup>a,b</sup> \*, F. Meurens <sup>a</sup>, M. Marie <sup>a,b</sup>, P. Lesage <sup>a,b</sup>, A. Lehébel <sup>a</sup>, N. Brisseau <sup>a</sup>, S. Assié <sup>a</sup> 8 9 <sup>a</sup> INRAE, Oniris, BIOEPAR, 44300 Nantes, France 10 <sup>b</sup> SELAS EVA, Réseau Cristal, 16 avenue du Général De Gaulle, 79150 Argentonnay, France 11 12 13 14 \* Corresponding author. Tel.: +33 (0)6 24 63 42 81. 15 E-mail address: nicolas.masset@oniris-nantes.fr (N. Masset).

#### 17 Abstract

18 Bovine respiratory syncytial virus (BRSV) and bovine parainfluenza-3 virus (bPI3V) 19 are major causes of bovine respiratory disease (BRD) in newborn calves worldwide. 20 Vaccination is widely used to prevent BRD, and intranasal vaccines for BRSV and bPI3V 21 were developed to overcome interference from BRSV and bPI3V-specific maternally derived 22 antibodies. Many experimental challenge trials have demonstrated that intranasal vaccines for 23 BRSV and bPI3V are efficacious, but effectiveness under field conditions has been 24 demonstrated less often, especially for newborn beef calves. The objective of this field trial 25 was to compare the effectiveness of a newly available commercial BRSV-bPI3V intranasal 26 vaccine with that of a benchmarked one in newborn beef calves reared in a cow-calf system. 27 A total of 935 calves from 39 farms were randomized into two vaccine groups (Bovalto Respi Intranasal [Vaccine A], n = 468; Rispoval RS + PI3 Intranasal [VaccineB], n = 467), and 28 29 monitored during the in-house risk period up to three months after vaccination. Non-30 inferiority analysis was performed by calculating the difference in BRD prevalence between 31 the two vaccine groups.

32

No significant differences were observed between vaccines regarding clinical outcomes of morbidity, mortality, duration between vaccination and BRD occurrence, or treatments required. Because the upper limit of the 2-sided 95% confidence interval of the difference in BRD prevalence between the two treatment groups (0.8%) was less than the margin of non-inferiority ( $\delta = 5\%$ ), a non-inferiority of Vaccine A was concluded. In conclusion, Vaccine A is at least as effective as Vaccine B for the prevention of BRD in newborn beef cattle in a cow-calf system under field conditions.

- *Keywords:* Bovine; Bovine respiratory syncytial virus; Bovine parainfluenza-3 virus; Calf;
- 42 Intranasal vaccine; Vaccination

#### 44 Introduction

45 Bovine respiratory disease (BRD) is one of the main health issues encountered in non-46 weaned beef calves, and can lead to high economic losses (Assié et al., 2004a; Wang et al., 47 2018). Viral infections generally initiate BRD and predispose animals to secondary bacterial 48 infections (Mosier, 2014). Bovine respiratory syncytial virus (BRSV), an Orthopneumovirus 49 of the *Pneumoviridae* family, is a major virus involved in the BRD complex and is highly 50 prevalent in both dairy and beef herds (Brodersen, 2010; Sacco et al., 2014; Valarcher and 51 Taylor, 2007). Likewise, bovine parainfluenza-3 virus (bPI3V), a Respirovirus of the 52 Paramyxoviridae family, is another virus involved in the BRD complex, widely prevalent in 53 herds (Ellis, 2010). Vaccines against BRSV and bPI3V are widely used to control BRD, 54 especially in beef calves. In a French study of 165 cow-calf herds in 2000, 116/186 (62%) 55 batches of beef calves were vaccinated against BRSV (Assié et al., 2009).

56

57 The neonatal period is a significant risk period for BRD. The immune system of 58 newborn calves differs from that of adults in several respects (Chase et al., 2008; Cortese, 59 2009). Although functional at birth, the immune system of a calf remains immature until six 60 months of age (Hauser et al., 1986; Tizard, 2018), and the immune response during this time 61 is weak, slow and more easily overcome by pathogenic microorganisms. In addition, 62 maternally derived antibodies (MDA), which are transmitted through colostrum and remain 63 present for up to six months, can interfere negatively with immunization of newborn calves 64 after vaccination (Ellis et al., 2014; Kimman et al., 1989). To overcome interference between 65 parenteral vaccines and MDA, intranasal vaccination strategies using modified live vaccines 66 for respiratory diseases have been developed and used widely for many years (Windeyer and 67 Gamsjäger, 2019). Intranasal vaccination is able to induce protective immunity in newborn 68 calves despite the presence of MDA by priming mucosal immunization of the upper

respiratory tract whereas protective immunity is inconsistent after parenteral vaccinations(Osman et al., 2018).

71

72 Veterinary vaccine efficacy is mainly evaluated in challenge trials under controlled 73 conditions (Knight-Jones et al., 2014). The efficacy of BRSV intranasal vaccines has been 74 proven in many challenge trials under controlled conditions even when vaccinations are performed in the presence of MDA (Ellis, 2017; Osman et al., 2018). However, these studies 75 76 generally do not consider variations that occur under field conditions, such as exposure to other pathogens, or host and environmental factors. Field trials are therefore needed to 77 78 reliably evaluate vaccine effectiveness (Knight-Jones et al., 2014). To our knowledge, only 79 one study dedicated to BRSV intranasal vaccination effectiveness has been carried out under 80 field conditions in newborn dairy calves. In that study, no decrease in BRD incidence or lung 81 lesions associated with pneumonia was demonstrated, but an increase in average daily gain 82 was observed (Ollivett et al., 2018). It should be noted, however, that the management of 83 dairy calves is quite different from that of beef suckler calves. Indeed, in cow-calf systems, 84 animals of different susceptibilities to respiratory diseases or with different immune statuses 85 are mixed in collective barns, whereas dairy calves are classically housed in individual pens 86 during the first eight weeks of life before being sorted and mixed into groups of similar age in 87 collective barns.

88

One BRSV-bPI3V intranasal vaccine authorized for use in newborn calves to prevent
BRD has been available for over 10 years in Europe (Vaccine B, Rispoval RS + PI3
Intranasal, Zoetis). The efficacy and the safety of Vaccine B have been demonstrated in
several experimental studies (Vangeel et al., 2009, 2007). With this vaccine, nasal shedding of
BRSV and bPI3V in vaccinated calves with or without MDA was reduced after challenges

94 with BRSV and bPI3V respectively. Additionally, the severity of clinical disease was also 95 reduced after BRSV in vaccinated calves with BRSV MDA. Moreover, this vaccine has been 96 widely used in Europe and is now a benchmark for BRSV-bPI3V intranasal vaccines. 97 Recently, several new BRSV intranasal vaccines have been launched in Europe. Our study 98 aimed to compare the effectiveness of a new intranasal vaccine against BRSV and bPI3V 99 (vaccine A, Bovalto Respi Intranasal, Boehringer Ingelheim) with that of the benchmarked 100 vaccine (vaccine B) in terms of decreasing BRD morbidity in newborn beef calves reared in a 101 cow-calf farming system. As these two vaccines were very similar in their composition (i.e. 102 bivalent modified live vaccines against BRSV and bPI3V) and their indication for use (i.e. 103 active immunization), a non-inferiority study was performed. 104 105 Materials and methods 106 The trial was carried out under the agreement of the Ethics Committee for Clinical and 107 Epidemiological Veterinary Research of Oniris (CERVO, Nantes-Atlantic National College 108 of Veterinary Medicine, Food Science and Engineering, France; Approval number, 109 CERVO-2018-8-V; Approval date, 8 October, 2018). 110 111 Vaccines Vaccines A and B contain BRSV and bPI3V strains administered as a single dose of 2 112 mL with an intranasal applicator. A dose of vaccine A (evaluated vaccine) contains between 113  $10^{4.0}$  and  $10^{6.0}$  TCID<sub>50</sub> of BRSV Bio 24/A strain and between  $10^{5.0}$  and  $10^{7.5}$  TCID<sub>50</sub> of bPI3V 114 115 strain Bio 23/A reconstituted with phosphate buffered saline. A dose of vaccine B (benchmarked vaccine) contains between  $10^{5.0}$  and  $10^{7.2}$  50% tissue culture infective doses 116 (TCID<sub>50</sub>) of BRSV 375 strain and between 10<sup>5.0</sup> and 10<sup>8.6</sup> TCID<sub>50</sub> of temperature-sensitive 117 118 mutant bPI3V strain RLB 103 reconstituted with saline.

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120 Study design

121 Type of trial

122 A randomized non-inferiority multicentre trial was carried out to assess whether 123 vaccine A was at least as effective as vaccine B, with a pre-stated margin of non-inferiority 124  $(\delta)$  for the prevention of BRD in newborn beef cattle. The null hypothesis (H<sub>0</sub>) was that 125 vaccine A was inferior to vaccine B in preventing BRD, whereas the alternative hypothesis 126 (H<sub>a</sub>) was that vaccine A was not inferior by more than the predefined non-inferiority margin. 127 The hypothesis statements may be summarized as follows: 128  $H_0$ :  $(P_{BRD}[vaccine A] - P_{BRD}[vaccine B]) \ge \delta$ 129  $(P_{BRD}[vaccine A] - P_{BRD}[vaccine B]) \le \delta$ H<sub>a</sub>: 130 where  $P_{BRD}$  was the prevalence rate of calves treated for BRD during the study period and  $\delta$ 131 the non-inferiority margin. 132 133 Determination of the non-inferiority margin 134 Due to the lack of published field trials of the effectiveness of vaccine B, it was not possible to determine the non-inferiority margin using a 2-step approach as described by 135 136 Freise et al. (2013). However, efficacy and the safety of vaccine B has been demonstrated in 137 controlled challenge trials, and this vaccine has been until now been considered as the 138 benchmarked BRSV-bPI3V IN vaccine. Thus, based on clinical judgment, because 5% was 139 the largest loss of effectiveness of vaccine A that would be considered clinically insignificant, 140 the non-inferiority margin  $\delta$  was defined as 5%. 141

142 Sample size determination

143	Sample size was determined using the package 'TrialSize' in R <sup>1</sup> based on a non-
144	inferiority trial with the prevalence of calves treated for BRD during the study period as the
145	primary outcome. Four hundred and forty-six (446) calves per group were needed to
146	demonstrate non-inferiority assuming $\alpha = 0.05$ , $\beta = 0.20$ , $\delta = 0.05$ and a prevalence of BRD in
147	the active control vaccine group B of 10%. As it was a stratified multicentre individually
148	randomized trial with two equal group sizes (A/B) and an equal number of calves in the two
149	groups on each farm, the inflating factor was defined as 1- $\rho$ , with $\rho$ the intraclass correlation
150	coefficient (Vierron and Giraudeau, 2009, 2007). After assigning this inflation factor with $\rho$ =
151	0.14 (Hendrick et al., 2013) and assuming 20% of loss to follow-up, it was decided to enrol at
152	least 920 calves in the study.
153	
154	Animals
154 155	Animals Herd selection
155	Herd selection
155 156	Herd selection Forty cow-calf farms with $110 \pm 55$ calvings (mean $\pm$ standard deviation, SD), located
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155 156 157 158	Herd selection Forty cow-calf farms with 110 ± 55 calvings (mean ± standard deviation, SD), located in four areas of France (Auvergne-Rhône-Alpes, Bretagne, Nouvelle-Aquitaine, Pays de La Loire), were selected for this study. Initial herd selection by veterinarians was based on the
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165

166 Calf selection

<sup>&</sup>lt;sup>1</sup> See: R Foundation for Statistical Computing, Vienna, Austria, <u>http://www.R-project.org</u> (Accessed 16 August 2020).

On each farm, 1 to 3 blocks of 10 calves were enrolled (3, 6 and 32 farms had respectively 1, 2 or 3 blocks). As soon as there were more than 10 calves over 10 days of age, a clinical examination of each calf was performed by a veterinarian. Only calves in good health (no intercurrent illness detected during the clinical examination) at the time of enrolment were included. If a calf had been ill and treated for BRD or other diseases before the day of inclusion, enrolment was possible only after full clinical recovery of the animal.

#### 173

#### 174 Randomization and vaccination protocols

Randomization was performed with a 1:1 allocation ratio. In each block of 10 enrolled
calves from each farm, calves were sorted by decreasing age. A predefined randomized
spreadsheet was prepared for each block: a random drawing was performed using the Rand
function of MS Excel to assign the vaccine to the oldest calf, then the following calves were
vaccinated alternately with one of the two vaccines.

180

Vaccines were administered by one veterinary investigator with calves restrained by the farmer, while another veterinary investigator reconstituted vaccines and recorded data. The vaccines were administered according to the manufacturer's recommendations: for Vaccine A, 1 mL in each nostril with the specific intranasal applicator (cannula and disk) provided by the manufacturer; for Vaccine B, 2 mL in one nostril with the specific intranasal applicator (cannula) provided by the manufacturer.

187

Intranasal applicators were changed between each calf. Due to the different administration modalities for the two vaccines, it was not possible to carry out a blinded vaccination. However, data were collected and registered by another veterinary investigator who kept randomization sheets and was not involved in the monitoring of calves after vaccination. To avoid potential bias in later husbandry, farmers were asked to not read the tag
numbers of calves during vaccination. After vaccination, cow-calf couples were raised in the
same pen according to the usual rearing conditions on each farm.

195

196 *Outcomes and data collection* 

197 The protocol started with vaccination and ended three months later. The primary 198 outcome for analysis was BRD events, defined by at least one respiratory sign (such as cough, 199 dyspnea, and/or nasal discharge) associated with at least one general clinical sign 200 (hyperthermia, depression, and/or anorexia). Secondary outcomes included the time between 201 vaccine administration and the occurrence of BRD, calf mortality, and the number of calves 202 treated with antibiotics or anti-inflammatory drugs during the study.

203

For each calf, demographic data (tag number, date of birth, sex, and parity of the dam) were extracted from the official identification databases, and medical history was gathered from animal health records of the farm. For three months following vaccination, the farmers, under supervision of the veterinary investigators, had to monitor and record all observations and treatments carried out: date, clinical signs, diagnosis (respiratory disorders and others) and drug administration (antibiotic and/or anti-inflammatory preparations).

210

In the event of mortalities during the study, a veterinary investigator performed
required necropsy examinations to identify the cause of death. In the case of
bronchopneumonia, lung samples (approximately 5 cm<sup>3</sup>) were frozen at -20°C for PCR
analysis. Multiplex real-time PCR assays (Pack Respiratory 8 Bio-T kit, Biosellal) designed
for the detection of eight BRD agents (BRSV, bPI3V, bovine Coronavirus, Influenza D,

216 Mannheimia haemolytica, Pasteurella multocida, Mycoplasma bovis, Histophilus somni)
217 were performed in Agrivalys 71.

218

219 Post-admission exclusion

Calves with incomplete data, vaccinated before 10 days or after 60 days of age, or
treated for BRD without at least two clinical signs being recorded, or as part of a
metaphylaxis protocol, or vaccinated after the beginning of the grazing period were excluded.

224 Statistical analysis

225 The characteristics of calves assigned to the two vaccine groups were compared to 226 assess homogeneity: Student's t-test was used for continuous variables (i.e., age at 227 vaccination, risk period) after checking normality, and chi-squared or Fisher tests were used 228 for categorical variables (i.e., *parity of dam, sex*, and *disease occurrence before vaccination*). 229 The statistical analysis of the primary outcome was performed using a mixed logistic 230 regression model (with 'calf' as the statistical unit). The primary explanatory variable of 231 interest was vaccine. Other variables were also tested (sex, parity of dam, age at vaccination, 232 *risk period*) and kept in the model if P < 0.2 in the univariable analysis. A backward stepwise 233 elimination of variables was then performed until all explanatory variables with P < 0.05 were 234 included in the final model, taking into account potential confounders. Herd (categorical 235 variable) was included as a random effect. The variable risk period was kept in the model to 236 adjust BRD occurrence to the variation of the duration of exposition to pathogens between 237 calves. The variables *vaccine* and *risk period* were forced in the final model, written as:

238 
$$BRD_{ij} \sim Bernoulli(p_{ij})$$

239 
$$Logit(p_{ij}) = \ln\left(\frac{p_{ij}}{1-p_{ij}}\right) = \beta_0 + vaccine_{ij}\beta_1 + risk \ period_{ij}\beta_2 + X_{ij}\beta_k + v_j$$

240 
$$v_i \sim Normal(0, \sigma_v^2)$$

where BRD<sub>ij</sub> is the occurrence of a BRD case diagnosed during the study *Risk Period* for the calf i of the herd j with a probability of occurrence  $p_{ij}$ ,  $\beta_0$  is the intercept, *vaccine<sub>ij</sub>* is either vaccine A or B, *risk period<sub>ij</sub>* is the duration of the in-house risk period,  $X_{ij}$  is *sex*, *parity of dam* and *age at vaccination* variables and  $v_j$  is the random effect for herd j. Herd random effect followed a normal distribution with mean 0 and variance  $\sigma_v^2$ .

246

The difference in BRD prevalence between vaccine groups and its 95% confidence interval (CI) was calculated from the model. Non-inferiority of vaccine A compared with vaccine B was concluded if the upper bound of the 2-sided 95% CI of the difference of BRD prevalence between the two vaccines was smaller than the non-inferiority margin  $\delta$  (Fig. 1). Secondary outcomes were compared between the two vaccine groups using the chi-squared test, Fisher test or Student's *t*-test. All analyses were carried out using R software <sup>1</sup> and a statistical significance at  $P \le 0.05$  was used.

254

#### 255 **Results**

256 Descriptive results

257 A total of 1,120 calves from 40 farms were enrolled in the study. Data from 185 calves 258 were excluded: 40 because of incomplete data, 85 for date of vaccination before 10 days or 259 after 60 days of age, 30 for being treated for BRD without at least two clinical signs or in a 260 metaphylaxis protocol, and 30 for being vaccinated after the beginning of the grazing period 261 (no housing period). Thus, a total of 935 calves from 39 herds were used in the study analysis: 262  $24 \pm 7$  calves (mean  $\pm$  SD) per herd. The two experimental groups were homogeneous in 263 regard to age at vaccination, duration of in-house risk period, parity of dams, sex ratio, and 264 occurrence of diseases before vaccination (Table 1).

#### 266 Primary outcome

267	The occurrence of BRD during the in-house risk period between the two vaccine
268	groups was similar (Table 2). Using least squares means of model outcome (BRD events), the
269	difference $P_{BRD}(vaccine A) - P_{BRD}(vaccine B)$ was estimated at -0.4% with a 95% CI between
270	-1.6% and 0.8%. Non-inferiority of Vaccine A compared to Vaccine B was concluded since
271	the upper limit of the 2-sided 95% CI ( $0.8\%$ ) of the difference in prevalence of calves
272	diagnosed with BRD between the two vaccines was smaller than $\delta$ (Fig. 1). BRD incidence in
273	our study was 0.74 cases per 1,000 calf-days at risk (Table 3).
274	
275	Secondary outcome
276	The two experimental groups were similar in regard to time between vaccination and
277	occurrence of BRD, treatments and mortality (Table 4). For the six calves which died during
278	the study period, BRSV was not detected from the samples collected during the necropsy
279	procedure (Table 5).
280	
281	
	Discussion
282	<b>Discussion</b> The objective of this study was to compare the effectiveness of the newly available

284

285 Vaccine A compared to Vaccine B was concluded. Due to the lack of studies dedicated to the

286 effectiveness of Vaccine B compared to a placebo, the non-inferiority margin  $\delta$  was defined

287 as 5% only, based on a clinical judgment. This margin is narrow compared to the one used in

- 288 most vaccine trials, with  $\delta$  usually fixed at 10% as reviewed by Donken et al. (2015).
- 289 Choosing a more conservative non-inferiority margin required an increased sample size and
- 290 improved the clinical significance of the trial. Indeed, based on the experiences of the authors,

an increase of more than 5% of BRD events (i.e., the primary outcome) in the vaccine A
group compared to the vaccine B group was considered unfavorable.

293

294 In this study, calves from the two vaccine groups were mixed together in order to 295 homogenize as much as possible environmental conditions and exposure to pathogens. This 296 design is often chosen in field studies dealing with vaccine effectiveness (Schunicht et al., 297 2003; Stilwell et al., 2008; Wildman et al., 2008). Moreover, in a cow-calf system, this design 298 enables the absence of separation of paired calves of the two vaccine groups after 299 randomization, and improves blind assessment of calf health in a single group. However, a 300 bias in vaccine effectiveness evaluation could be introduced with this method. The reduction 301 of virus shedding after vaccination of the calves of one vaccine group contributes to the 302 protection of the calves of the other vaccine group reared in the same environment (Smith, 303 2019, 2014; Stokka, 2010). Indeed, apparent effectiveness of the test vaccine could be 304 improved if the comparison involved a reference vaccine with a better shedding reduction 305 efficacy. Moreover, Vaccines A and B are both modified live vaccines. Cross-immunization 306 thus could occur between the two vaccine groups. In previous studies using commercial 307 vaccines including the reference vaccine, nasal shedding of vaccine-origin viruses was 308 detected by PCR in nasal swabs during 14 days in most of the vaccinated calves after 309 vaccination and was detected up to 28 days post vaccination in a few calves (Timsit et al., 310 2009; Walz et al., 2017).

311

The study was designed to determine the effectiveness of two commercial vaccines under conditions as close as possible to those encountered by calves reared in beef herds. Although the minimum age at vaccination recommended by the manufacturers is 9 and 10 days of age for Vaccines B and A, respectively, under our conditions the mean age (± SD) at vaccination was 26 ± 12 days. This delay in administrating vaccines was due to the
distribution of births on each farm and the packaging of the vaccines in 5-dose bottles. Since
both vaccines are available in a 5-dose bottle, 10 calves had to be over 10 days of age before
being vaccinated in order to randomize them into two equal groups of five calves. However,
this difference between the recommended and actual age at vaccination was the same in both
groups and is common in French cow-calf systems.

322

323 Most of the efficacy studies for BRSV and bPI3V intranasal vaccination include a 324 controlled challenge, but challenges may not reproduce natural exposure under variable host 325 and environmental factors (Knight-Jones et al., 2014). It has been observed that many BRSV 326 infection models failed to reproduce the severe clinical signs of the disease, complicating the 327 evaluation of vaccine efficacy (Belknap et al., 1995; Blodörn et al., 2015; Taylor, 2013). In 328 these studies, efficacy was demonstrated in newborn calves both in the absence of BRSV-329 specific MDAs (Ellis et al., 2007; Vangeel et al., 2007) and in their presence (Ellis et al., 330 2013; Hill et al., 2012). However, the absence of maternal antibody interference is not always 331 observed (Ellis et al., 2010). In our study, colostral immunity in calves was not systematically 332 controlled for practical reasons, and the BRSV and bPI3V serological status of calves at the 333 time of vaccination was unknown. Due to the high prevalence of BRSV in France, most of the 334 vaccinations were likely to have been performed in the presence of BRSV-specific MDAs.

335

As observed in a previous study, BRSV and bPI3V infections were found in 71% and 80% of cow-calf farms respectively (Assié et al., 2004b). Furthermore, challenge trials do not reproduce the variability of host and environmental conditions that may be encountered in the field, such as variable passive immune transfer (Raboisson et al., 2016), variable calf housing (Assié et al., 2009; Dubrovsky et al., 2019a; Maier et al., 2019) and variable seasonal or weather conditions (Buczinski et al., 2018; Dubrovsky et al., 2019b). For these reasons, a
multicentre study was chosen in order to reproduce this variability of environmental factors.
Both efficacy assessment in experimental challenges and effectiveness assessment in field
trials have limits but are complementary.

345

346 Contrary to challenge trials, the exposure of calves to pathogens, in particular to 347 BRSV and bPI3V, is rarely controlled in a field study (Ellis, 2017; Ollivett et al., 2018). The 348 authors acknowledge that monitoring BRSV and bPI3V exposure (by means such as PCR or 349 virus isolation on deep nasal swabs or fluid of transtracheal aspiration or bronchoalveolar 350 lavage, or serology on sentinels) would have allowed us to assess specifically the 351 effectiveness of vaccines against BRSV and bPI3V, and not only the prevention of BRD. As 352 previously reported, monitoring exposure to pathogens in a vaccinated population is very 353 difficult for both practical and economic reasons (Ellis, 2017). Indeed, the short viremia 354 would require repeated samplings of a large population. In a recent field trial evaluating 355 BRSV and bPI3V intranasal vaccination in dairy calves, Ollivett et al. (2018) similarly did not 356 assess the exposure of calves to BRSV and bPI3V, and monitored BRD morbidity alone.

357

358 However, the exposure of calves to respiratory pathogens in our study can be attested 359 by the measurement of BRD incidence, which was 0.74 cases per 1,000 calf-days at risk. 360 Although this BRD incidence was low, it remains consistent with the incidence observed in 361 another French study in a comparable breeding system in which respiratory vaccination was 362 inconsistent: 1.89 cases per 1,000 calf-days at risk in 137 farms (Assié et al., 2004a). 363 Circulation of respiratory pathogens in the study farms can also be attested by the viruses 364 (bPI3V, bovine Coronavirus) and bacteria (Mannheimia haemolytica, Pasteurella multocida, 365 *Histophilus somni*) identified at necropsy in dead animals. To overcome the variability in the 366 exposure to pathogens under field conditions, our study would need to be repeated.

367

#### 368 Conclusions

369 The effectiveness of a newly available commercial BRSV-bPI3V intranasal vaccine to 370 control BRD has been demonstrated under field conditions. To the authors' knowledge, this is 371 the first study under field conditions assessing BRSV and bPI3V intranasal vaccination 372 effectiveness in newborn beef calves in a cow-calf system. Data from challenge studies or 373 from dairy calf field studies cannot be extrapolated to beef calves. Beef cattle from different 374 age groups with different immune statuses against respiratory pathogens are mixed together in 375 a specific in-house environment, in contrast to dairy calves which are typically housed in 376 individual pens or in collective pens with animals of the same age.

377

#### **378 Conflict of interest statement**

This study was funded by Boehringer Ingelheim, which supports the European College of Bovine Health Management (ECBHM) residency program of the first author. Boehringer Ingelheim played no role in the study design, in data collection, analysis or interpretation, or in the writing of the report and the decision to submit the manuscript for publication. None of the authors has any other financial or personal relationships that could inappropriately influence or bias the content of the paper.

385

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389

390 **References** 

391	Assié, S., Bareille, N., Beaudeau, F., Seegers, H., 2009. Management- and housing-related
392	risk factors of respiratory disorders in non-weaned French Charolais calves. Preventive
393	Veterinary Medicine 91, 218–225.
394	
395	Assié, S., Seegers, H., Beaudeau, F., 2004a. Incidence of respiratory disorders during housing
396	in non-weaned Charolais calves in cow-calf farms of Pays de la Loire (Western France).
397	Preventive Veterinary Medicine 63, 271–282.
398	
399	Assié, S., Seegers, H., Ogier de Baulny, M., Beaudeau, F., 2004b. Pathogens and incidence of
400	respiratory disorders on non-weaned calves in Charolais cow-calf farms of the Pays de
400	la Loire (France), in: 11ème Journées Rencontres Recherches Ruminants. Presented at
402	the 11ème Journées Rencontres Recherches Ruminants, Paris (France), pp. 329–332.
402	the Treme Journees Rencontres Remembers Rummants, Taris (France), pp. 529–552.
	Pollenon E.B. Cianovalvi D.K. Dolvon I.C. 1005 Experimental requirestory experticity into
404	Belknap, E.B., Ciszewski, D.K., Baker, J.C., 1995. Experimental respiratory syncytial virus
405	infection in calves and lambs. Journal of Veterinary Diagnostic Investigation 7, 285–
406	298.
407	
408	Blodörn, K., Hägglund, S., Gavier-Widen, D., Eléouët, JF., Riffault, S., Pringle, J., Taylor,
409	G., Valarcher, J.F., 2015. A bovine respiratory syncytial virus model with high clinical
410	expression in calves with specific passive immunity. BMC Veterinary Research 11, 76.
411	
412	Brodersen, B.W., 2010. Bovine Respiratory Syncytial Virus. Veterinary Clinics of North
413	America: Food Animal Practice 26, 323–333.
414	
415	Buczinski, S., Borris, M.E., Dubuc, J., 2018. Herd-level prevalence of the ultrasonographic
416	lung lesions associated with bovine respiratory disease and related environmental risk
417	factors. Journal of Dairy Science 101, 2423–2432.
418	
419	Chase, C.C.L., Hurley, D.J., Reber, A.J., 2008. Neonatal Immune Development in the Calf
420	and Its Impact on Vaccine Response. Veterinary Clinics of North America: Food
421	Animal Practice, Dairy Heifer Management 24, 87–104.
422	
423	Cortese, V.S., 2009. Neonatal Immunology. Veterinary Clinics of North America: Food
424	Animal Practice, Bovine Neonatology 25, 221–227.
425	
426	Donken, R., de Melker, H.E., Rots, N.Y., Berbers, G., Knol, M.J., 2015. Comparing vaccines:
427	A systematic review of the use of the non-inferiority margin in vaccine trials. Vaccine
428	33, 1426–1432.
429	
430	Dubrovsky, S.A., Van Eenennaam, A.L., Karle, B.M., Rossitto, P.V., Lehenbauer, T.W., Aly,
431	S.S., 2019a. Epidemiology of bovine respiratory disease (BRD) in preweaned calves on
432	California dairies: The BRD 10K study. Journal of Dairy Science 102, 7306–7319.
433	
434	Dubrovsky, S.A., Van Eenennaam, A.L., Karle, B.M., Rossitto, P.V., Lehenbauer, T.W., Aly,
435	S.S., 2019b. Bovine respiratory disease (BRD) cause-specific and overall mortality in
436	preweaned calves on California dairies: The BRD 10K study. Journal of Dairy Science
437	102, 7320–7328.
438	102, 1520 1520.
439	Ellis, J., Gow, S., Bolton, M., Burdett, W., Nordstrom, S., 2014. Inhibition of priming for
439	bovine respiratory syncytial virus-specific protective immune responses following

441 parenteral vaccination of passively immune calves. Canadian Veterinary Journal 55, 442 1180-1185. 443 444 Ellis, J., Gow, S., West, K., Waldner, C., Rhodes, C., Mutwiri, G., Rosenberg, H., 2007. 445 Response of calves to challenge exposure with virulent bovine respiratory syncytial 446 virus following intranasal administration of vaccines formulated for parenteral 447 administration. Journal of the American Veterinary Medical Association 230, 233-243. 448 449 Ellis, J.A., 2017. How efficacious are vaccines against bovine respiratory syncytial virus in 450 cattle? Veterinary Microbiology 206, 59-68. 451 452 Ellis, J.A., 2010. Bovine parainfluenza-3 virus. Veterinary Clinics of North America: Food 453 Animal Practice 26, 575–593. 454 455 Ellis, J.A., Gow, S.P., Goji, N., 2010. Response to experimentally induced infection with 456 bovine respiratory syncytial virus following intranasal vaccination of seropositive and 457 seronegative calves. Journal of the American Veterinary Medical Association 236, 991-458 999. 459 460 Ellis, J.A., Gow, S.P., Mahan, S., Leyh, R., 2013. Duration of immunity to experimental 461 infection with bovine respiratory syncytial virus following intranasal vaccination of 462 young passively immune calves. Journal of the American Veterinary Medical 463 Association 243, 1602–1608. 464 465 Freise, K.J., Lin, T.-L., Fan, T.M., Recta, V., Clark, T.P., 2013. Evidence-Based Medicine: 466 The Design and Interpretation of Noninferiority Clinical Trials in Veterinary Medicine. 467 Journal of Veterinary Internal Medicine 27, 1305–1317. 468 469 Hauser, M.A., Koob, M.D., Roth, J.A., 1986. Variation of neutrophil function with age in 470 calves. American Journal of Veterinary Research 47, 152-153. 471 472 Hendrick, S.H., Bateman, K.G., Rosengren, L.B., 2013. The effect of antimicrobial treatment 473 and preventive strategies on bovine respiratory disease and genetic relatedness and 474 antimicrobial resistance of Mycoplasma bovis isolates in a western Canadian feedlot. 475 Canadian Veterinary Journal 54, 1146–1156. 476 477 Hill, K.L., Hunsaker, B.D., Townsend, H.G., van Drunen Littel-van den Hurk, S., Griebel, 478 P.J., 2012. Mucosal immune response in newborn Holstein calves that had maternally 479 derived antibodies and were vaccinated with an intranasal multivalent modified-live 480 virus vaccine. Journal of the American Veterinary Medical Association 240, 1231-481 1240. 482 483 Kimman, T.G., Westenbrink, F., Straver, P.J., 1989. Priming for local and systemic antibody 484 memory responses to bovine respiratory syncytial virus: effect of amount of virus, virus 485 replication, route of administration and maternal antibodies. Veterinary Immunology 486 and Immunopathology 22, 145–160. 487 488 Knight-Jones, T.J.D., Edmond, K., Gubbins, S., Paton, D.J., 2014. Veterinary and human 489 vaccine evaluation methods. Proceedings of the Royal Society B: Biological Sciences 490 281.

491 492	Maior C.U. Lovo W.L. Korlo D.M. Dukrovsky, S.A. Williams D.B. Champagna I.D.
	Maier, G.U., Love, W.J., Karle, B.M., Dubrovsky, S.A., Williams, D.R., Champagne, J.D.,
493	Anderson, R.J., Rowe, J.D., Lehenbauer, T.W., Van Eenennaam, A.L., Aly, S.S., 2019.
494	Management factors associated with bovine respiratory disease in preweaned calves on
495	California dairies: The BRD 100 study. Journal of Dairy Science 102, 7288–7305.
496	
497	Mosier, D., 2014. Review of BRD pathogenesis: the old and the new. Animal Health
498	Research Reviews 15, 166–168.
499	
500	Ollivett, T.L., Leslie, K.E., Duffield, T.F., Nydam, D.V., Hewson, J., Caswell, J., Dunn, P.,
501	Kelton, D.F., 2018. Field trial to evaluate the effect of an intranasal respiratory vaccine
502	protocol on calf health, ultrasonographic lung consolidation, and growth in Holstein
503	dairy calves. Journal of Dairy Science 101, 8159–8168.
504	
505	Osman, R., Malmuthuge, N., Gonzalez-Cano, P., Griebel, P., 2018. Development and
506	Function of the Mucosal Immune System in the Upper Respiratory Tract of Neonatal
507	Calves. Annual Review of Animal Biosciences 6, 141–155.
508	
509	Piaggio, G., Elbourne, D.R., Pocock, S.J., Evans, S.J.W., Altman, D.G., Group for the
510	CONSORT, 2012. Reporting of Noninferiority and Equivalence Randomized Trials:
511	Extension of the CONSORT 2010 Statement. Journal of the American Medical
512	Association 308, 2594–2604.
512	Association 508, 2574–2004.
515 514	Raboisson, D., Trillat, P., Cahuzac, C., 2016. Failure of Passive Immune Transfer in Calves:
514	
	A Meta-Analysis on the Consequences and Assessment of the Economic Impact. PloS
516	One 11, e0150452.
517	Sacco D.E. McCill II. Billetzki A.E. Delman M.V. Askermann M.D. 2014 Despiratory
518	Sacco, R.E., McGill, J.L., Pillatzki, A.E., Palmer, M.V., Ackermann, M.R., 2014. Respiratory
519	syncytial virus infection in cattle. Veterinary Pathology 51, 427–436.
520	Schweicht O.C. Declare C.W. Free C.K. Creicher D.T. Wildman D.K. Hill D.W. 2002
521	Schunicht, O.C., Booker, C.W., Jim, G.K., Guichon, P.T., Wildman, B.K., Hill, B.W., 2003.
522	Comparison of a multivalent viral vaccine program versus a univalent viral vaccine
523	program on animal health, feedlot performance, and carcass characteristics of feedlot
524	calves. Canadian Veterinary Journal 44, 43–50.
525	
526	Smith, D.R., 2019. Herd immunity. Veterinary Clinics of North America: Food Animal
527	Practice 35, 593–604.
528	
529	Smith, D.R., 2014. Field epidemiology to manage BRD risk in beef cattle production systems.
530	Animal Health Research Reviews 15, 180–183.
531	
532	Stilwell, G., Matos, M., Carolino, N., Lima, M.S., 2008. Effect of a quadrivalent vaccine
533	against respiratory virus on the incidence of respiratory disease in weaned beef calves.
534	Preventive Veterinary Medicine 85, 151–157.
535	
536	Stokka, G.L., 2010. Prevention of Respiratory Disease in Cow/Calf Operations. The
537	Veterinary Clinics of North America: Food Animal Practice, Bovine Respiratory
538	Disease 26, 229–241.
539	
540	Taylor, G., 2013. Bovine Model of Respiratory Syncytial Virus Infection, in: Challenges and

541 542	Opportunities for Respiratory Syncytial Virus Vaccines, Current Topics in Microbiology and Immunology. Springer, Berlin, Heidelberg, pp. 327–345.
543 544 545 546 547	Timsit, E., Le Dréan, E., Maingourd, C., Belloc, C., Guattéo, R., Bareille, N., Seegers, H., Douart, A., Sellal, E., Assié, S., 2009. Detection by real-time RT-PCR of a bovine respiratory syncytial virus vaccine in calves vaccinated intranasally. The Veterinary Record 165, 230–233.
548 549 550	Tizard, I., 2018. Immunity in the Fetus and Newborn. In: Veterinary Immunology. Elsevier, Saint Louis, Missouri, pp. 247–260.
551 552 553 554	Valarcher, JF., Taylor, G., 2007. Bovine respiratory syncytial virus infection. Veterinary Research 38, 153–180.
555 556 557 558 559	Vangeel, I., Antonis, A.F.G., Fluess, M., Riegler, L., Peters, A.R., Harmeyer, S.S., 2007. Efficacy of a modified live intranasal bovine respiratory syncytial virus vaccine in 3- week-old calves experimentally challenged with BRSV. The Veterinary Journal 174, 627–635.
560 561 562 563 564	Vangeel, I., Ioannou, F., Riegler, L., Salt, J.S., Harmeyer, S.S., 2009. Efficacy of an intranasal modified live bovine respiratory syncytial virus and temperature-sensitive parainfluenza type 3 virus vaccine in 3-week-old calves experimentally challenged with PI3V. The Veterinary Journal 179, 101–108.
565 566 567	Vierron, E., Giraudeau, B., 2009. Design effect in multicenter studies: gain or loss of power? BMC Medical Research Methodology 9, 39.
568 569 570	Vierron, E., Giraudeau, B., 2007. Sample size calculation for multicenter randomized trial: Taking the center effect into account. Contemporary Clinical Trials 28, 451–458.
570 571 572 573 574 575	Walz, P.H., Newcomer, B.W., Riddell, K.P., Scruggs, D.W., Cortese, V.S., 2017. Virus detection by PCR following vaccination of naive calves with intranasal or injectable multivalent modified-live viral vaccines. Journal of Veterinary Diagnostic Investigation 29, 628–635.
576 577 578 579	Wang, M., Schneider, L.G., Hubbard, K.J., Smith, D.R., 2018. Cost of bovine respiratory disease in preweaned calves on US beef cow-calf operations (2011-2015). Journal of the American Veterinary Medical Association 253, 624–631.
579 580 581 582 583 584	Wildman, B.K., Perrett, T., Abutarbush, S.M., Guichon, P.T., Pittman, T.J., Booker, C.W., Schunicht, O.C., Fenton, R.K., Jim, G.K., 2008. A comparison of 2 vaccination programs in feedlot calves at ultra-high risk of developing undifferentiated fever/bovine respiratory disease. Canadian Veterinary Journal 49, 463–472.
585 586 587 588	Windeyer, M.C., Gamsjäger, L., 2019. Vaccinating calves in the face of maternal antibodies: Challenges and opportunities. Veterinary Clinics of North America: Food Animal Practice 35, 557–573.

591 Calf characteristics in vaccine groups A and B

Variable	Vaccine g	Р		
	A ( $n = 468$ )	B ( <i>n</i> = 467)	-	
Age at vaccination in days (mean ± SD)	25.97 ± 11.62	25.85 ± 11.54	0.88	
Duration of in-house risk period in				
days (mean $\pm$ SD)	$56.52 \pm 28.25$	$57.00 \pm 27.93$	0.80	
Parity				
1 (n = 279)	143	136		
2(n = 157)	72	85	0.51	
3 and more $(n = 499)$	253	246		
Sex				
Male $(n = 447)$	235	212		
Female $(n = 488)$	233	255	0.14	
Occurrence of diseases before vaccination				
	127	126	1	
No disease $(n = 873)$	437	436	1	
Respiratory $(n = 6)$ Other than respiratory <sup>a</sup> $(n = 56)$	3 28	3 28		

592 SD, Standard deviation

593 <sup>a</sup> All neonatal diseases were diagnosed and treated (i.e., septicaemia, diarrhoea, umbilical

594 infection, others)

597 Multivariable results of mixed logistic regression model of bovine respiratory disease (BRD)

Variable	Category	Number of calves	Odds ratio of BRD	95% Confidence interval	P <sup>a</sup>
		evaluated	occurrence		
Vaccine	Vaccine B	467	Reference		
	Vaccine A	468	0.61	0.30-1.25	0.17
Duration of in-house	(0-45)	313	Reference		
risk period in days	(45-67.5)	243	8.88	1.07-73.66	0.04
- •	(67.5-90)	379	6.61	0.86-50.99	0.07

598 prevalence after intranasal vaccination of non-weaned beef calves.

- <sup>a</sup> For each variable, refers to level of significance between the category under consideration
- 600 and the reference category.

Vaccine group	Number of	Number of		
	Calf-days at risk	Cases		
Vaccine A	25,538	15	0.59	
Vaccine B	26,031	23	0.88	
Total	51,569	38	0.74	

# 603 Incidence of bovine respiratory disease cases in the two vaccine groups

04 <sup>a</sup> per 1,000 calf-days at risk

Outcome		Vaccine group $(n = 935)$		
	A ( <i>n</i> = 468)	B ( <i>n</i> = 467)		
Calves treated for BRD with				
Antibiotics (%)	15 (3.2)	23 (4.9)	0.23	
Non-steroidal anti-inflammatories (%)	12 (2.6)	17 (3.6)	0.34	
Steroidal anti-inflammatories (%)	1 (0.2)	2 (0.4)	0.50	
Mortalities (%)	1 (0.2)	5 (1.1)	0.11	
Time between vaccination and occurrence of BRD i days (mean ± SD)	n $33 \pm 20$	28 ± 22	0.45	
BRD, Bovine respiratory disease; SD, Standard dev	iation			

607 Comparisons of secondary outcomes between the two vaccine groups

611 Results of multiplex real-time PCR on lung samples for detection of eight respiratory

Calf number	Vaccine group	Herd number	Pathogen detected in multiplex real-time PCR <sup>a</sup>						'R <sup>a</sup>	
			BRSV	bPI3V	Mh	Pm	Mb	Hs	bCo	ID
14	В	1			No	samp	le			
244	В	13			No	samp	le			
913	В	37	-	+	+	+	-	+	-	-
941	А	38	-	-	-	+	-	+	+	-
955	В	38	-	-	-	+	-	-	+	-
967	В	38	-	+	-	+	-	-	-	-

612 pathogens from necropsies of dead calves

613 BRSV, Bovine respiratory syncytial virus; bPI3V, bovine Parainfluenza Virus type 3; Mh,

614 *Mannheimia haemolytica*; Pm, *Pasteurella multocida*; Mb, *Mycoplasma bovis*; Hs, *Histophilus somni*;
615 bCo, bovine Coronavirus; ID, Influenza D virus

616 <sup>a</sup> Pack Respiratory 8 Bio-T kit, Biosellal

#### 618 Figure legends

619

620 Fig.1. Four possible scenarios of a non-inferiority trial comparing vaccine A to vaccine B for 621 preventing BRD. The margin of non-inferiority ( $\delta$ ) is drawn by a vertical dashed line. P<sub>BRD</sub> is 622 the prevalence of BRD cases diagnosed during the study risk period of housing after 623 vaccination. Error bars indicate 2-sided 95% confidence interval (CI) of the difference in 624 BRD incidence (Piaggio et al., 2012). S: if the CI lies wholly to the left of zero, vaccine A is 625 superior. NI: if the CI lies to the left of  $\delta$  and includes zero, vaccine A is non-inferior. IC: if 626 the CI includes  $\delta$  and zero, the difference is non-significant but the result regarding noninferiority is inconclusive. I: if the CI is wholly above  $\delta$ , vaccine A is inferior. VT is the 627 628 representation of the main outcome of this non-inferiority trial. The black block indicates the 629 difference in BRD incidence between vaccine A group and vaccine B group. Non-inferiority 630 of vaccine A compared to vaccine B at a margin of 5% is demonstrated because the 95% CI 631 lies to the left of  $\delta$  (=5%) and includes zero.

