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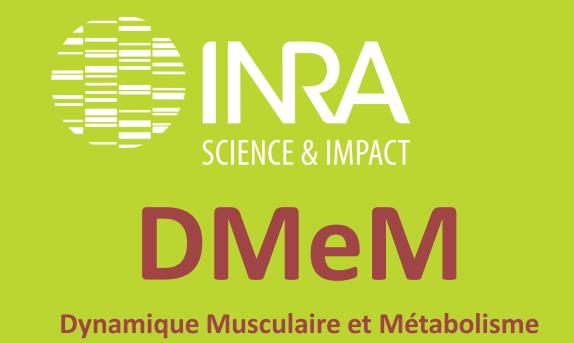
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Optimization of permeabilized fibers preparation for mitochondrial respiration measurements using Design of Experiments methodology

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Aim of the study

To optimize the permeabilized fibers (pf) preparation from mouse *Tibialis anterior* in our lab, we used the Design of Experiments (DoE) methodology that evaluates the impact of 6 experimental conditions or factors, on the pf respiration parameters (Pyruvate Malate Succinate respiration (PMS leak) and respiratory control ratio (RCR_{PMS})), to provide a maximum of information using a limited number of experiments and animals.

Materials and Methods

Test system Animals: C57BL/6 mice, 25 week-old, male and female (n=18)

Muscle: Tibialis anterior, n=2 per mice

<u>Device</u>: High-resolution Oxygraph-2k (OROBOROS Instruments) DoE software: NemrodW[®], version 2015, NewrodW SAS,

Marseille, France



Muscle fiber bundle (source : DMEM)

Fixed experimental conditions

Resting rate (PMS leak): 5 mM pyruvate, 5mM malate and 10 mM succinate

ADP-stimulated rate (PMS_p): addition of 5 mM ADP Respiratory Control Ratio (RCR_{PMS}) set as the ratio of oxygen consumption at

PMS leak (PMS₁) over oxygen consumption at PMS_P. Y_1 : PMS₁ level \Rightarrow to be maximized (at least 40 pmol $O_2/s*mg$ fibers)

Responses studied

Y₂: variability of RCR_{PMS}, estimated by coefficient of variation of 4 repeated experiments > to be minimized

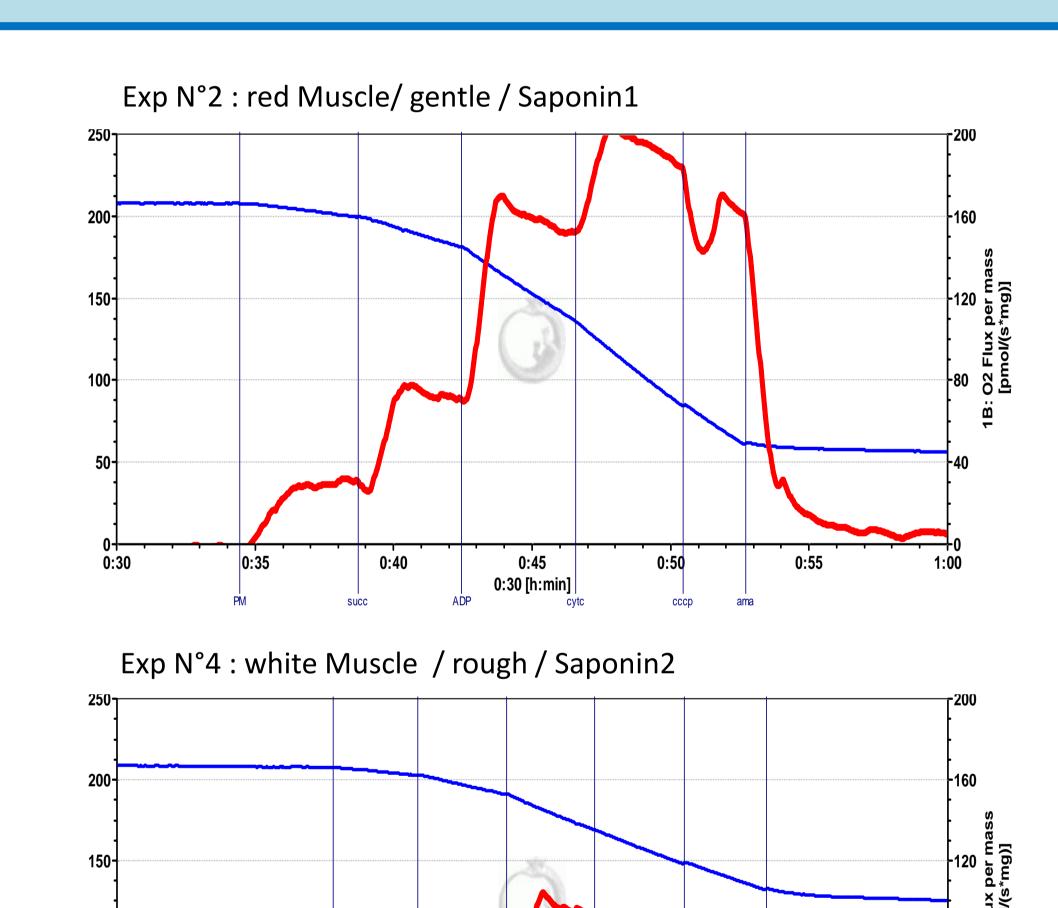
Design of Experiments

The influence of 6 factors on Y₁ and Y₂ responses has been evaluated using a Hadamard matrix with 8 experiments (instead of 64 experiments if all combinations had been tested with a « One-Factor-At-A-Time » (OFAT) method), see below. To evaluate experimental variance for Y₁ response, each experiment has been replicated 4 times. To evaluate experimental variance for Y₂ response, one experiment (n°6) has been replicated 4 additional times. In total, 36 experiments have been performed.

Results / Interpretations

Experimental matrix and results

Experiments	Fiber types	Manual teasing	Saponin content	Saponin concentration (µg/ml)	Permeabilization time (min)	Resting period (h)	Y ₁ : PMS _L level (pmol O ₂ /s*mg)				Y ₂ : RCR _{PMS} variability
N°1	White	Gentle	S1	25	30	0	30.7	24.6	12	33.5	0.196
N°2	Red	Gentle	S1	50	10	6	80.1	56.1	81	61.2	0.194
N°3	Red	Rough	S1	50	30	0	56.4	63.8	68.6	90.2	0.084
N°4	White	Rough	S2	50	30	6	31.3	42.9	27.6	39.5	0.204
N°5	Red	Gentle	S2	25	30	6	51.2	53.4	53.3	63.5	0.096
N°6	White	Rough	S1	25	10	6	32.8	30.3	44.8	43.1	0.094
							67.9	81.9	35.2	49.5	0.099
N°7	White	Gentle	S2	50	10	0	31.8	24.3	25.1	22.2	0.120
N°8	Red	Rough	S2	25	10	0	62.4	61.4	69.6	45.9	0.202



Experimental domain

Level -1

Red

Gentle

 $25 \mu g/ml$

10 mn

0 hour

Level +1

White

Rough

 $50 \, \mu g/ml$

30 mn

6 hours

8-25% (S1) 20-35% (S2)

Factors evaluated

Fiber Types

Manual Teasing

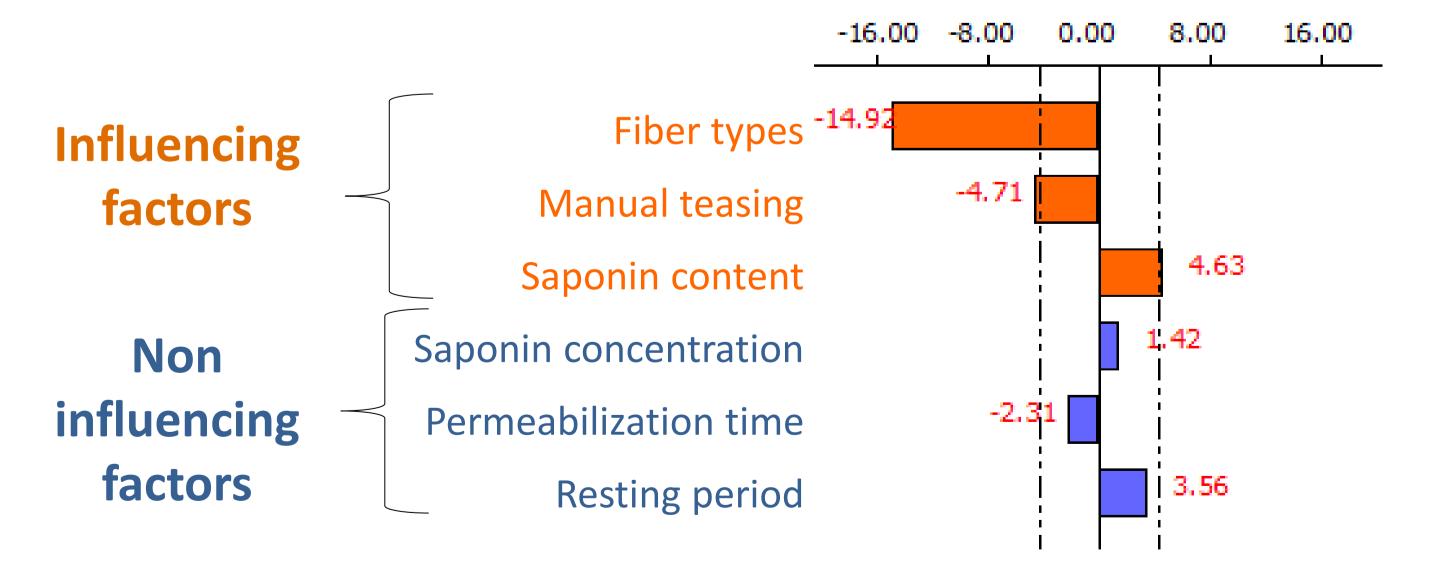
Saponin content

Saponin concentration for permeabilization

Permeabilization time

Resting period before permeabilization

Y₁: PMS, level



Best experimental conditions to maximize PMS, level are red fibers, rough manual teasing and 20-35% saponin content.

0:35

To be noticed:

- ✓ Six hours resting period had no deleterious impact on PMS₁ level, allowing a more convenient organization of the protocole schedule.
- ✓ Objective level of 40 pmol $O_2/s*mg$ fibers is reached with both saponin content allowing to choose between the more pratical/less toxic mode of preparation.

Y₂: RCR_{PMS} variability

Evaluation of experimental variance with only one replicate of one experiment over 8 was not accurate enough to discriminate with confidence which of the 6 tested factors are really influencing RCR_{PMS} variability. Nevertheless, it seems that levels of influencing factors that maximize PMS_I level were not deleterious in minimizing RCR_{PMS} variability.

Conclusion

Using a DoE analysis, we were able to optimize pf assay conditions with a reduced number of experiments and animals, and rapidly obtain valuable data in accordance with ethical recommendations (3Rs). The optimization of pf preparation by DoE will be pursued with two objectives (i) studying the possible interactions existing between the 3 factors related to saponin (saponin concentration and incubation time), (ii) calculating the optimal sample size (n) needed to observe statistically significant differences between two animal groups.











