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1 ***Title***

2 **Dynamics and diversity of mosquito vectors of Japanese encephalitis virus in Kandal**
3 **province, Cambodia**

4

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20 **Highlights**

- 21 • Vectors of Japanese Encephalitis Virus are abundant all the year in Cambodia
- 22 • One-year study and seasonality of JEV vectors
- 23 • Effect of host presence on JEV vector abundance in pig farms and cattle house
- 24 • The most abundant vectors are *Culex vishnui*, *Cx. gelidus* and *Cx. tritaeniorhynchus*

25

26 **Abstract**

27 The Japanese encephalitis virus (JEV) is one of the main causes of encephalitis in Asia,
28 including Cambodia. An understanding of the interactions between JEV hosts and vectors
29 (Diptera: Culicidae) remains rare in the context of expanding urbanization. The relative
30 abundance, species diversity and population dynamics of potential JEV vectors were studied
31 between August 2015 and July 2016 on a peri-urban and rural pig farm in Kandal province,
32 Cambodia, where JEV is circulating. Five similar environments in the two farms were
33 selected for mosquito trapping: pig farm, cattle house, river/canals, household/ponds and
34 paddy fields. The main objective was to describe the distribution and the dynamics of the
35 main JEV vector mosquito species. In total, 83,013 mosquitoes from 20 species were caught
36 in rural and peri-urban areas, and 82.3% of the mosquitoes were potential JEV vector species.
37 In peri-urban areas, *Culex (Cx.) gelidus* was the most abundant species, followed by *Cx.*
38 *vishnui* subgroup and *Cx. tritaeniorhynchus*. In rural areas, the same species were dominant:
39 *Cx. vishnui* subgroup, *Cx. gelidus* and *Cx. tritaeniorhynchus*. The vast majority of mosquitoes
40 (95.9%) were collected in close proximity to pigs and cattle. In conclusion, JEV vectors were
41 present at all study sites and throughout all months of the year, supporting a continuous
42 circulation of JEV in Cambodia.

43 **Keywords**

44 Japanese encephalitis; Vectors; Mosquitoes; *Culex vishnui*; *Culex gelidus*; *Culex*
45 *tritaeniorhynchus*; Cambodia

46 **Introduction**

47 Japanese encephalitis (JE), an arthropod-borne and zoonotic disease, is one of the main
48 causes of encephalitis, especially in the Asia Pacific region (Campbell et al. 2011). JE has an
49 estimated annual incidence rate of 67,900 cases among 24 JE endemic countries (van den
50 Hurk et al. 2009; Campbell et al. 2011). This vector-borne disease caused by Japanese
51 encephalitis virus (JEV) was first identified in Japan, in 1935 from an encephalitis patient's
52 brain (Lewis et al. 1947) and has since been found throughout Australasian and Asian
53 countries including Cambodia (van den Hurk et al. 2009; Rosen, 1986). JEV belongs to the
54 viral family *Flaviviridae* and the genus *Flavivirus*. It is maintained in the complex
55 transmission cycle between water birds, mosquitoes and domestic pigs. Humans and horses
56 are incidental hosts and considered dead-end hosts for JEV transmission because of low
57 viremia levels incapable of infecting mosquitoes (Scherer et al. 1959; Impoinvil et al., 2013).
58 Water bird species (heron [*Ardeola grayii*, *Nycticorax nycticorax*], and egrets [*Bulbucus ibis*,
59 *Egretta garzetta*, *Egretta intermedia*]) and wild ducks [*Anas platyrhynchos*] are reservoirs for
60 the maintenance and dissemination of the virus (Buescher et al. 1959; Soman et al. 1977;
61 Rodrigues et al. 1981). Pigs represent an important amplification host, and pig farming has
62 been identified as a risk factor for JEV infection in humans (Cao et al. 2010; Liu et al. 2010).
63 However, the role of pigs in JE epidemiology needs to be reassessed in order to identify and
64 implement efficient control strategies for both human and animal health (Ladreyt et al. 2019).
65 JEV transmission occurs primarily in rural areas but there are reports of occasional human
66 cases in urban areas (Gingrich et al. 1987). For example, in India, JEV occurs in peri-urban
67 and rural settings in nearly half of its 29 states (Pattan et al. 2009). In Bangkok, Thailand
68 (Thisyakorn and Nimmannitya, 1985; Gingrich et al. 1987), Vientiane, Lao PDR (Vallee et
69 al. 2009), and Can Tho, Vietnam (Lindhal et al. 2012), JEV or its vectors were identified in
70 sub-urban areas that are comparable to the outskirts of Phnom Penh, Cambodia.

71 To date, 34 mosquito species have been studied for their involvement in Japanese
72 encephalitis transmission: *Armigeres* species (n=1), *Ochlerotatus* (n=2), *Mansonia* (n=2),
73 *Aedes* (n=4), *Anopheles* (n=8) and mainly *Culex* (n=17). Of these 34 species, 14 species are
74 considered potential JEV vectors and seven species as confirmed vectors (following the
75 definitions of potential and confirmed vectors from Tantely et al. 2015). The seven confirmed
76 species are *Culex tritaeniorhynchus*, *Cx. bitaeniorhynchus*, *Cx. fuscocephala*, *Cx. gelidus*, *Cx.*
77 *quinquefasciatus*, *Cx. sitiens* group, and *Aedes vexans*. Notably, *Cx. tritaeniorhynchus* and
78 *Cx. vishnui* are considered the main vectors of JEV (Le Flohic et al. 2013; Oliveira et al.
79 2018).

80 Regarding Cambodia, JEV was first detected in 1965 from *Cx. tritaeniorhynchus* (Chastel
81 and Rageau, 1966) and again in 1967 from *Cx. tritaeniorhynchus* and *Cx. gelidus* (Chen et al.
82 1990). Several studies between 1996 and 2008, and in 2017, demonstrated serological
83 evidence of JEV infection in humans (Srey et al. 2002; Chhour et al. 2002; Touch et al. 2009;
84 Horwood et al. 2017). Furthermore, more than 60% of the 505 swine sera sampled from eight
85 provinces in Cambodia were positive to JEV antibodies (Duong et al. 2011). Recently,
86 evidence of infections in pigs, mosquitoes and humans demonstrated an intensive circulation
87 of JEV in peri-urban areas of Cambodia (Cappelle et al. 2016; Duong et al. 2017).

88 Importantly, Cappelle et al. (2016) showed that all female vector species have a low
89 minimum infection rate (MIR) of 0.091 / 1,000. These previous studies have established the
90 presence of JEV in Cambodia but there is limited data regarding the diversity and population
91 dynamics of JEV vectors driving the transmission. The main objectives of this study were to
92 (i) describe the diversity of potential JEV vectors in the vicinity of pig farms in a peri-urban
93 and a rural area of Cambodia and to (ii) compare their relative abundance and describe their
94 dynamics.

95 **Materials and methods**

96 Study sites

97 Between August 2015 and July 2016, mosquitoes were collected from two different
98 geographical areas classified as peri-urban and rural (Figure 1).

99 The peri-urban area was approximately 11 km south of Phnom Penh in a village of Ta Krapeu
100 Ha, Preaek Ruessei commune, Ta Khmau city (11.4739 °N, 104.9376 °E). This area is
101 located between a densely populated urban area and a rural landscape dominated by
102 cultivated areas (Figure 1). The peri-urban study site was located on an experimental pig farm
103 where 15 pigs were kept in a garden with no other domestic animals (Di Francesco et al.
104 2018).

105 The rural area was approximately 45 km south of Phnom Penh and adjacent to the Bassac
106 River (Figure 1), in a village of Kbal Chhroy, Porti Ban commune, Koh Thom district
107 (11.219846 °N, 105.039502 °E). Most inhabitants of the rural area are farmers and land usage
108 is predominantly agricultural and livestock, rearing mainly pigs, cattle, chickens and ducks
109 (Figure 1). Within this area, the selected dwelling had a backyard farm with a constant
110 presence of pigs, and 2 cows in the farm.

111 The number of pigs at each pig farm varied over the study period. In the peri-urban area, 15
112 pigs were present at the experimental pig farm between August and October 2015. No pigs
113 were present after the monitoring study of JEV circulation in pigs ended in October 2015. In
114 the rural area, more than 100 pigs were present between August 2015 and January 2016.
115 From February to July 2016, the number of pigs was reduced to 15 after the owner relocated
116 pigs to another pigsty.

117 Five different locations were defined in both peri-urban (PU) and rural (R) areas for mosquito
118 trapping: the pig farm (PU-P & R-P), a cattle house (PU-C & R-C) and the household to
119 estimate the mosquito abundance in proximity to humans (PU-H & R-H). Two aquatic sites

120 per study area were sampled: a canal and paddy field in the peri-urban area (PU-Ca and PU-
121 PF respectively); and a riverbank and paddy field in the rural area (R-R and R-PF
122 respectively (Figure 1).

123 Mosquito trapping and identification

124 Mosquitoes were captured over a 12-month period during the second or the third week of
125 each month. Each month a CDC light-trap was in operation at each location between 5-7 pm
126 until 6-7 am for three consecutive nights. In the pig farms, the suspended light-trap was
127 outside and close to the pig garden. In the human and cattle houses, the light-trap was
128 suspended outside from the ceiling under the stilt house. In the river/canals, ponds and paddy
129 fields, the traps were attached to a tree branch.

130 The morphological identification of adult mosquitoes was performed according to the
131 identification key to mosquitoes of Southeast Asia countries (Stojanovich and Scott 1966;
132 Reuben et al. 1994; Rattanarithikul et al. 2005). To avoid bias, any mosquitoes captured in
133 malfunctioning light-traps (i.e. light, fan or battery issues) were excluded from analysis.

134 Statistical analysis

135 Statistical analysis used RStudio software (Core Team (2017). R: A language and
136 environment for statistical computing. R Foundation for Statistical Computing, Vienna,
137 Austria. URL <https://www.R-project.org/>). Differences between species abundance within the
138 different areas and locations were assessed using ANOVA tests followed by a post-hoc
139 analysis, Tukey-HSD for multiple comparisons, to indicate differences between groups. The
140 ANOVA also tested the interactions between the location of the traps and the sampling area,
141 and between the trapping month and the sampling area.

142 **Results**

143 Diversity and relative abundance of mosquitoes in peri-urban and rural areas

144 In total, 360 collections were performed (10 traps were used) across both areas (PU and R)
145 during the 36 nights of capture. 340 (94.4%) traps operated correctly and 20 (5.6%) failed (12

146 times in rural and 8 in peri-urban). From correctly working traps, mosquitoes belonging to 10
147 genera and 20 species were collected from both areas, with 14 of these species known to be
148 JEV vectors: *Aedes* (n=2), *Anopheles* (n=2), *Armigeres* (n=1), *Culex* (n=7) and *Mansonia*
149 (n=2; Table 1).

150 Overall, 83,013 individual mosquitoes were captured from both areas: 59.4% (n=49,305)
151 from rural areas and 40.6% (n=33,708) from peri-urban areas. Of the total collection, 17.1%
152 (n=14,192) were not identified at the species level due to poor physical condition of the
153 specimen: 86.8% (n=12,317) of these mosquitoes belonged to the *Culex* genus. In total, *Culex*
154 mosquitoes represented 97.4% (n=32,843) and 96.2% (n=47,428) of the total mosquito
155 collection in peri-urban and rural areas respectively. (Table 1).

156 Furthermore, 99.3% (n=80,271) of these JEV vectors belong to the *Culex* genus, with 83.0%
157 (n=66,604) from three main species, namely *Cx. gelidus*, *Cx. tritaeniorhynchus*, and *Cx.*
158 *vishnui* subgroup. In the rural sites, *Cx. vishnui* subgroup accounted for 51.7% (n=25,488)
159 and were the most abundant, followed by *Cx. gelidus* (14.9%; n=7,322) and *Cx.*
160 *tritaeniorhynchus* (10.8%; n=5,327) (Table 1). In the peri-urban sites, the more abundant
161 species were *Cx. gelidus* (36.9%; n=12,421), *Cx. vishnui* subgroup (28.0%; n=9,453) and *Cx.*
162 *tritaeniorhynchus* (19.6%; n=6,593) (Table 1).

163 Main JEV vector species population dynamics

164 The three main mosquito species, *Cx. vishnui* subgroup, *Cx. gelidus* and *Cx.*
165 *tritaeniorhynchus*, were present throughout the year but with a variation in their abundance
166 depending on the month for the two areas (p=0.009 for rural area; p=0.049 for peri-urban
167 area). In the peri-urban area, the three species displayed a similar trend with an abundant
168 peak in October 2015 (rainy season), especially *Cx. gelidus* and *Cx. vishnui* with a maximum
169 relative density of approximately 300 mosquitoes per night. The population of those species
170 rapidly declined from November 2015 to April 2016, and then *Cx. vishnui* gradually started

171 to increase again with the beginning of the rainy season in May, and *Cx. gelidus* later in July
172 2016 (Figure 2). In the rural area, the three species were present throughout the year with two
173 peaks of abundance in December 2015 and July 2016 (Figure 2). Particularly, *Cx. vishnui* had
174 two peaks of abundance at the beginning and at the end of the rainy season, while *Cx. gelidus*
175 peaked at the beginning of the rainy season and *Cx. tritaeniorhynchus* to a lesser extent at the
176 end of the rainy season concomitantly *Cx. vishnui* (Figure 2).

177 Relative abundance of mosquitoes in different trap locations

178 More mosquitoes were trapped in pig farms and cattle houses compared to the household,
179 river/canals, ponds and paddy fields ($p < 10^{-4}$). In total, 95.9% of mosquitoes were caught in pig
180 farms (71.0%; $n=58,971$) and in cattle houses (24.8%; $n=20,606$), while only 4.1% of trapped
181 mosquitoes were from the other locations. In the peri-urban area, we captured a similar average
182 number of mosquitoes per night in the pig farm ($n=459$) compared to the cattle house ($n=463$)
183 ($p=1.00$). In the rural area, the average nightly capture rate was more than 8 times higher in the
184 pig farm ($n=1212$) compared to the cattle house ($n=146$) ($p < 10^{-4}$).

185 There was no significant effect of the location of the traps and the sampling area on the number
186 of caught mosquitoes for the three main species ($p=0.33$) (Figure 3). However, in the rural area,
187 there were more *Cx. vishnui* subgroup (640 vs. 68, $p=0.0004$) and *Cx. tritaeniorhynchus* (127
188 vs. 20, $p=0.033$) mosquitoes in the pig farm ($n=640$) than in the cattle house ($n=68$) ($p=0.043$;
189 Figure 4). Even though the difference was not significant with *Cx. gelidus* ($p=0.053$), there
190 were more mosquitoes caught per night in pig farm ($n=191$) than in the cattle house ($n=15$).
191 The captured mosquito numbers were lower in the river/canal, household/pond and rice fields
192 compared to the pig farms and cattle house, and there was an overall difference between the
193 number of *Cx. vishnui* subgroup between rural and peri-urban areas ($p=0.008$; Figure 5). In the
194 rural areas, *Cx. vishnui* was more abundant in the river and canal than near households and
195 paddy fields.

196 **Discussion**

197 Japanese encephalitis is mainly considered a rural disease, but there is growing evidence of a
198 peri-urban and urban transmission in several countries, including Cambodia (Di Francesco et
199 al. 2018). Moreover, JE represents one of the most important cause of human viral
200 encephalitis in South East Asia and particularly in Cambodia (Horwood et al. 2017). Our
201 study focused on the dynamics of JEV mosquito vectors and was implemented to monitor
202 mosquito populations in peri-urban and rural areas in Cambodia. JE studies in Cambodia
203 mainly focused on JEV isolation from humans, pigs or mosquitoes, but few studies have
204 focused on JEV vectors. A previous study showed predominance of *Cx. quinquefasciatus* in
205 Phnom Penh during the rainy season (Kohn, 1990). Beyond the difference of habitats (urban
206 area vs peri-urban and rural areas in our study), this discrepancy could also be explained by
207 the different method of trappings and/or the locations of the study. Kohn (1990) used an
208 aspirator and netting in households that are likely to capture highly anthropophilic
209 mosquitoes (Gowda and Vijayan, 1992), while our study used light traps near pigs and cattle,
210 and likely captured mosquitoes that are more zoophilic. A more recent study demonstrated
211 similar findings in terms of species and vector diversity to our peri-urban area in Ta Khmau
212 city (Cappelle et al. 2016). They also found *Cx. tritaeniorhynchus* (69%), *Cx. gelidus* (17%)
213 and *Cx. vishnui* (12%) as the predominant species with a higher density between April and
214 July for the two first ones. The same abundance peaks are observed at the beginning of the
215 rainy season (June and July), and the beginning of the dry season (December).

216 Diversity and relative abundance of JEV vectors in peri-urban and rural areas

217 This entomological survey provides direct evidence for the abundance of JEV vectors in
218 close proximity to humans and in areas with known JEV circulation (Cappelle et al. 2016).
219 Among all mosquito species caught in either peri-urban or rural areas, *Cx. gelidus* and *Cx.*
220 *vishnui* subgroup were the predominant species in peri-urban areas and in rural areas,

221 respectively. In India, *Cx. gelidus* and *Cx. tritaeniorhynchus* were also the predominant
222 species in peri-urban and rural areas, respectively (Arunachalam et al. 2009; Murty et al.
223 2010). In both countries, the presence and importance of these two species could explain the
224 circulation of JEV.

225 In India, JE cases occurred with a seasonality shortly after the mosquito density reached its
226 peak (Mishra et al. 1984; Kanojia et al. 2003). A study in Thailand showed that *Cx.*
227 *tritaeniorhynchus* and *Cx. gelidus* populations increased during the rainy season (Gingrich et
228 al. 1992) with a peak in July when the farmers plough their rice fields (Somboon et al. 1989;
229 Changbunjong et al. 2013). Even if the JEV transmission is described as seasonal in Thailand
230 and Vietnam, the transmission was presumed to occur from May to October in Cambodia and
231 Laos (Pattan et al. 2009). However, the serological monitoring of pigs during the rainy and
232 dry seasons (Cappelle et al. 2016), and the high abundance of JEV vectors during the rainy
233 and dry season in the present study, strongly suggest that JEV transmission occurs all year
234 round. The abundance of *Culex* species can be explained by several factors such as the
235 irrigation of rice paddies two or three times per year, and the importance of several different
236 mosquito-breeding sites such as streams, rivers, seepages, marshy areas, water containers,
237 ponds and other water reservoirs during the dry season. Specifically, *Cx. gelidus*, *Cx.*
238 *tritaeniorhynchus* and *Cx. vishnui* subgroup breeding sites in Asia are rice fields and general
239 ground pools such as stagnant water with plants, ditches, ponds (Sirivanakarn, 1976; Abu
240 Hassan et al. 2010; Kumar et al. 2012) that are present in our areas. Further studies to define
241 the specific breeding sites for each species and to determine the impact of climate variability
242 on the dynamics of the species should be conducted.

243 JEV vector abundance and hosts

244 The vast majority of mosquitoes (95.8%) were captured near hosts, particularly pigs. It has to
245 be noted, notably in the rural area, that the number of hosts, especially the number of pigs,

246 certainly play a role as a better attractant than the 2 cows present in the cattle house. The
247 three main species *Cx. vishnui* subgroup, *Cx. tritaeniorhynchus* and *Cx. gelidus* are zoophilic
248 species mainly attracted to animals for blood feeding (Lord et al. 2016; Tuno et al. 2016). As
249 our results show, particularly in peri-urban areas, the mosquito number dramatically
250 decreased after the pigs were removed from the pigsty; pigs likely attract JEV vector species.
251 In Vietnam, the abundance of *Cx. tritaeniorhynchus* in an urban area was related to the
252 presence of pigs (Nitapattana et al. 2005). Studies in Indonesia and China also found that
253 JEV cases in human are associated to the proximity of the residence to rice fields and pig
254 ownership, respectively (Liu et al. 2010, Ren et al. 2017). Considering the role of pigs in JEV
255 amplification and transmission to humans, the attraction of JEV vectors to pigs is of
256 importance, especially pigs that are kept in close proximity to human residences.

257 **Conclusion: importance for Public Health**

258 In conclusion, this study has shown that JEV vectors are present throughout the year in both
259 peri-urban and rural areas, with a high abundance in both rainy and dry seasons in Cambodia.
260 More mosquitoes were collected near pigs and cattle. Based on these findings, we can
261 conclude that the JEV transmission can occur throughout the year in peri-urban and rural
262 areas where pigs are present. It is recommended for people to avoid the presence of pigs near
263 the house in order to limit the abundance of JEV vectors. In addition, it would be effective to
264 protect these animals with insecticide-treated nets even when breeding sites are unknown
265 (Dutta et al. 2011).

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271 **References**

- 272 Abu Hassan, A., Dieng, H., Satho, T., Boots, M., Al Sariy, J. S., 2010. Breeding patterns of the
273 JE vector *Culex gelidus* and its insect predators in rice cultivation areas of northern peninsular
274 Malaysia. *Trop. Biomed.* 27, 404-416.
- 275 Arunachalam, N., Murty, U. S. N., Narahari, D., Balasubramanian, A., Samuel, P. P.,
276 Thenmozhi, V., Paramasivan, R., Rajendran, R., Tyagi, B. K., 2014. Longitudinal studies of
277 Japanese encephalitis virus infection in vector mosquitoes in Kurnool district, Andhra Pradesh,
278 South India. *J. Med. Entomol.* 46, 633-639.
- 279 Buescher, E. L., Scherer, W. F., McCluse, H., Moyer, J. T., Rosenberg, M. Z., Yoshii, Y.,
280 Okada, Y., 1959. Ecologic Studies of Japanese Encephalitis Virus in Japan. *Avian infection.*
281 *Am. J. Trop. Med. Hyg.* 8, 678-688.
- 282 Campbell, G. L., Hills, S. L., Fischer, M., Jacobson, J. A., Hoke, C. H., Hombach, J. M.,
283 Marfin, A. A., Solomon, T., Tsai, T. F., Tsu, V. D., Ginsburg, A. S., 2011. Estimated global
284 incidence of Japanese encephalitis: a systematic review. *Bull. W.H.O.* 89, 766-774.
- 285 Cappelle, J., Duong, V., Pring, L., Kong, L., Yakovleff, M., Prasetyo, D. B., Peng, B.,
286 Choeung, R., Duboz, R., Ong, S., Sorn, S., Dussart, P., Tarantola, A., Buchy, P., Chevalier, V.,
287 2016. Intensive circulation of Japanese encephalitis virus in peri-urban sentinel pigs near
288 Phnom Penh, Cambodia. *PLoS N.T.D.* 10, e0005149.
- 289 Cao, M., Feng, Z., Zhang, J., Ma, J., & Li, X., 2010. Contextual risk factors for regional
290 distribution of Japanese encephalitis in the People's Republic of China. *Trop. Med. Int. Health.*
291 15, 918-923.
- 292 Changbunjong, T., Weluwanarak, T., Taowan, N., Suksai, P., Chamsai, T., Sedwisai, P., 2013.
293 Seasonal abundance and potential of Japanese encephalitis virus infection in mosquitoes at the
294 nesting colony of ardeid birds, Thailand. *Asian Pac. J. Trop. Biomed.* 3, 207.

295 Chastel, C., Rageau, J., 1966. Isolement d'arbovirus au Cambodge a partir de moustiques
296 naturellement infectes. *Med. Trop.* 26, 391-400.

297 Chen, W. R., Tesh, R. B., Rico-Hesse, R., 1990. Genetic variation of Japanese encephalitis
298 virus in nature. *J. General Virol.* 71, 2915-2922.

299 Chhour, Y. M., Ruble, G., Hong, R., Minn, K., Kdan, Y., Sok, T., Nisalak, A., Aye Mint, K.
300 S., Vaughn, D. W., Endy, T. P., 2002. Hospital-based diagnosis of hemorrhagic fever,
301 encephalitis, and hepatitis in Cambodian children. *Emerg. Infect. Dis.* 8, 485.

302 Di Francesco, J., Choeung, R., Peng, B., Pring, L., Pang, S., Duboz, R., Ong, S., Sorn, S.,
303 Tarantola, A., Fontenille, D., Duong, V., Dussart, P., Chevalier, V., Cappelle, J., 2018.
304 Comparison of the dynamics of Japanese encephalitis virus circulation in sentinel pigs between
305 a rural and a peri-urban setting in Cambodia. *PLOS Negl. Trop. Dis.* 12, e0006644.

306 Duong, V., Sorn, S., Holl, D., Rani, M., Deubel, V., Buchy, P., 2011. Evidence of Japanese
307 encephalitis virus infections in swine populations in 8 provinces of Cambodia: implications for
308 national Japanese encephalitis vaccination policy. *Acta Trop.* 120, 146-150.

309 Duong, V., Choeung, R., Gorman, C., Laurent, D., Crabol, Y., Mey, C., Peng, B., Di Francesco,
310 J., Hul, V., Sothy, H., Santy, K., Richner, B., Pommier, J. D., Sorn, S., Chevalier, V., Buchy,
311 P., de Lamballerie, X., Cappelle, J., Horwood, P. F., Dussart, P., 2017. Isolation and full-
312 genome sequences of Japanese encephalitis virus genotype I strains from Cambodian human
313 patients, mosquitoes and pigs. *J. General Virol.* 98, 2287-2296.

314 Dutta, P., Khan, S. A., Khan, A. M., Borah, J., Sarmah, C. K., Mahanta, J., 2011. The effect of
315 insecticide-treated mosquito nets (ITMNs) on Japanese encephalitis virus seroconversion in
316 pigs and humans. *Am. J. Trop. Med. Hyg.* 84, 466-472.

317 Gingrich, J. B., Nisalak, A., Latendresse, J. R., Pomsdhit, J., Paisansilp, S., Hoke, C. H.,
318 Chantalakana, C., Satayaphantha, C., Uechiewcharnkit, K., 1987. A longitudinal study of
319 Japanese encephalitis in suburban Bangkok, Thailand. *S.E. Asian J. Trop. Med.* 18, 558-566.

320 Gingrich, J. B., Nisalak, A., Latendresse, J. R., Sattabongkot, J., Hoke, C. H., Pomsdhit, J.,
321 Chantalakana, C., Satayaphantha, C., Uechiewcharnkit, K., Innis, B. L., 1992. Japanese
322 encephalitis virus in Bangkok: factors influencing vector infections in three suburban
323 communities. *J. Med. Entomol.* 29, 436-444.

324 Gowda, N. N., Vijayan, V. A., 1992. Indoor resting density, survival rate and host preference
325 of *Culex quinquefasciatus* Say (Diptera: Culicidae) in Mysore City. *J. Communicable. Dis.* 24,
326 20-28.

327 Horwood, P. F., Duong, V., Laurent, D., Mey, C., Sothy, H., Santy, K., Richner, B., Heng, S.,
328 Cheval, J., Gorman, C., Dussart, P., de Jong, M. D., Kerleguer, A., Guillard, B., Murgue, B.,
329 Lecuit, M., de Lamballerie, X., Farrar, J. J., Tarantola, A., Eloit, M., Buchy, P., 2017. Aetiology
330 of acute meningoencephalitis in Cambodian children, 2010–2013. *Emerg. Microbes Infect.* 6,
331 1-8.

332 Impoinvil, D. E., Baylis, M., Solomon, T., 2012. Japanese encephalitis: on the one health
333 agenda. *Curr. Top. Microbiol. Immunol.* 365, 205-47.

334 Kanojia, P. C., Shetty, P. S., Geevarghese, G., 2003. A long-term study on vector abundance
335 & seasonal prevalence in relation to the occurrence of Japanese encephalitis in Gorakhpur
336 district, Uttar Pradesh. *Indian J. Med. Res.* 117, 104-110..

337 Kohn, M., 1990. A survey on indoor resting mosquito species in Phnom Penh, Kampuchea.
338 *Folia Parasitol.* 37, 165-174.

339 Kumar, G., Karthik, L., Rao, K. B., Kirthi, A. V., Rahuman, A. A., 2012. Larvicidal, repellent
340 and ovicidal activity of *Calotropis gigantea* against *Culex gelidus*, *Culex tritaeniorhynchus*
341 (Diptera: Culicidae). *Int. J. Agr. Tech.* 8, 869-80.

342 Ladreyt, H., Durand, B., Dussart, P., Chevalier, V., 2019. How Central Is the Domestic Pig in
343 the Epidemiological Cycle of Japanese Encephalitis Virus? A Review of Scientific Evidence
344 and Implications for Disease Control. *Viruses.* 11, 949.

345 Le Flohic, G., Porphyre, V., Barbazan, P., Gonzalez, J. P., 2013. Review of climate, landscape,
346 and viral genetics as drivers of the Japanese encephalitis virus ecology. *PLoS Neg. Trop. Dis.*
347 7, e2208.

348 Lewis, L., Taylor, H. G., Sorem, M. B., Norcross, J. W., Kindsvatter, V. H., 1947. Japanese B
349 encephalitis: clinical observations in an outbreak on Okinawa Shima. *Arch. Neurol. Psychiatry.*
350 57, 430-63.

351 Lindahl, J., Chirico, J., Boqvist, S., Thu, H. T. V., Magnusson, U., 2012. Occurrence of
352 Japanese encephalitis virus mosquito vectors in relation to urban pig holdings. *Am. J. Trop.*
353 *Med. Hyg.* 87, 1076-1082.

354 Liu, W., Gibbons, R. V., Kari, K., Clemens, J. D., Nisalak, A., Marks, F., Xu, Z. Y., 2010. Risk
355 factors for Japanese encephalitis: a case-control study. *Epidemiol. Infect.* 138, 1292-97.

356 Lord, J. S., Al-Amin, H. M., Chakma, S., Alam, M. S., Gurley, E. S., Pulliam, J. R., 2016.
357 Sampling design influences the observed dominance of *Culex tritaeniorhynchus*:
358 Considerations for future studies of Japanese encephalitis virus transmission. *PLoS Neg. Trop.*
359 *Dis.* 10, e0004249.

360 Mishra, A. C., Jacob, P. G., Ramanujam, S., Bhat, H. R., Pavri, K. M., 1984. Mosquito vectors
361 of Japanese encephalitis epidemic (1983) in Mandya district (India). *Indian J. Med. Res.* 80,
362 377.

363 Murty, U. S., Rao, M. S., Arunachalam, N., 2010. The effects of climatic factors on the
364 distribution and abundance of Japanese encephalitis vectors in Kurnool district of Andhra
365 Pradesh, India. *J. Vector Borne Dis.* 47, 26.

366 Nitatpattana, N., Apiwathnasorn, C., Barbazan, P., Leemingsawat, S., Yoksan, S., Gonzalez,
367 J., 2005. First isolation of Japanese encephalitis from *Culex quinquefasciatus* in Thailand. *S.E.*
368 *Asian J. Trop. Med. Public Health.* 36, 875.

369 Oliveira, A. R. S., Cohnstaedt, L. W., Strathe, E., Etcheverry, L., McVey, D. S., Piaggio, J.,
370 Cernicchiaro, N., 2018. Meta-Analyses of Japanese Encephalitis Virus Infection,
371 Dissemination, and Transmission Rates in Vectors. *Am. J. Trop. Med. Hyg.* 98, 883-890.

372 Pattan, S. R., Dighe, N. S., Nirmal, S. A., Bhawar, S. B., Dighe, S. B., Gaware, V. M., Hole,
373 M. H., 2009. Japanese Encephalitis Disease: A review. *Pharmacologyonline.* 2, 550-60.

374 Rattanarithikul, R., Harbach, R. E., Harrison, B. A., Panthusiri, P., Jones, J. W., Coleman, R.
375 E., 2005. Illustrated keys to the mosquitoes of Thailand. II. Genera *Culex* and *Lutzia*. *The*
376 *Southeast Asian J. Trop. Med. Public Health.* 36, 1-97.

377 Ren, X., Fu, S., Dai, P., Wang, H., Li, Y., Li, X., Lei, W., Gao, X., He, Y., Lv, Z., Cheng, J.,
378 Wang, G., Liang, G., 2017. Pigsties near dwellings as a potential risk factor for the prevalence
379 of Japanese encephalitis virus in adult in Shanxi, China. *Infect. Dis. Poverty.* 6, 100.

380 Reuben, R., Tewari, S. C., Hiriyan, J., Akiyama, J., 1994. Illustrated keys to species of *Culex*
381 (*Culex*) associated with Japanese encephalitis in Southeast Asia (Diptera: Culicidae). *Mosquito*
382 *Systematics.* 26, 75-96.

383 Rodrigues, F. M., Guttikar, S. N., Pinto, B. D., 1981. Prevalence of antibodies to Japanese
384 encephalitis and West Nile viruses among wild birds in the Krishna-Godavari Delta, Andhra
385 Pradesh, India. *Trans. R. Soc. Trop. Med. Hyg.* 75, 258-262.

386 Rosen, L., 1986. The natural history of Japanese encephalitis virus. *Ann. Rev Microbiol.* 40,
387 395-414.

388 Scherer, W. F., Kitaoka, M., Okuno, T., Ogata, T., 1959. Ecologic studies of Japanese
389 encephalitis virus in Japan. VII. Human infection. *Am. J. Trop. Med. Hyg.* 8, 707-15.

390 Sirivanakarn, S., 1976. Medical entomology studies III. A revision of the subgenus *Culex* in
391 the Oriental region (Diptera: Culicidae). *Cont. Am. Entomol. Inst.* 12, 1-271.

392 Soman, R. S., Rodrigues, F. M., Guttikar, S. N., Guru, P. Y., 1977. Experimental viraemia and
393 transmission of Japanese encephalitis virus by mosquitoes. *Indian J. Med. Res.* 66, 709-18.

394 Somboon, P., Choochote, W., Khamboonruang, C., Keha, P., Suwanphanit, P., Sukontasan, K.,
395 Chaivong, P., 1989. Studies on the Japanese encephalitis vectors in Amphoe Muang, Chiang
396 Mai, Northern Thailand. *SE Asian J. Trop. Med. Public Health.* 20, 9-17.

397 Srey, V. H., Sadones, H., Ong, S., Mam, M., Yim, C., Sor, S., Grosjean, P., Reynes, J. M.,
398 2002. Etiology of encephalitis syndrome among hospitalized children and adults in Takeo,
399 Cambodia, 1999-2000. *Am. J. Trop. Med. Hyg.* 66, 200-207.

400 Stojanovich, C. J., Scott, H. G., 1966. *Illustrated Key to Mosquitoes of Vietnam*, U. S.
401 Department of Health, Education and Welfare: Public Health Service, Communicable Disease
402 Center.

403 Tantely, L. M., Boyer, S., Fontenille, D., 2015. A review of mosquitoes associated with Rift
404 Valley fever virus in Madagascar. *Am. J. Trop. Med. Hyg.* 92, 722-729.

405 Thisyakorn, U., Nimmannitya, S., 1985. Japanese encephalitis in Thai children, Bangkok,
406 Thailand. *SE Asian J. Trop. Med. Public Health.* 16, 93-97.

407 Touch, S., Hills, S., Sokhal, B., Samnang, C., Sovann, L., Khieu, V., Soeung, S. C., Toda, K.,
408 Robinson, J., Grundy, J., 2009. Epidemiology and burden of disease from Japanese encephalitis
409 in Cambodia: results from two years of sentinel surveillance. *Trop. Med. Int. Health.* 14, 1365-
410 1373.

411 Tuno, N., Tsuda, Y., Takagi, M., 2016. How zoophilic Japanese encephalitis vector mosquitoes
412 feed on humans. *J. Med. Entomol.* 54, 8-13.

413 Vallée, J., Dubot-Pérès, A., Ounaphom, P., Sayavong, C., Bryant, J. E., Gonzalez, J. P., 2009.
414 Spatial distribution and risk factors of dengue and Japanese encephalitis virus infection in urban
415 settings: the case of Vientiane, Lao PDR. *Trop. Med. Int. Health.* 14, 1134-1142.

416 Van den Hurk, A. F., Ritchie, S. A., Mackenzie, J. S., 2009. Ecology and geographical
417 expansion of Japanese encephalitis virus. *Ann. Rev. Entomol.* 54, 17-35.

418 **Figure Legends**

419 **Table 1.** Number and percentage of each species captured in peri-urban and rural areas.
420 Number of mosquito species captured between August 2015 and July 2016 in peri-urban and
421 rural areas. Highlighted species with * are the potential know JEV vectors.

422 **Figure 1.** Geographic locations of traps in the peri-urban and rural areas. Peri-urban pig farm
423 (PU-P), Peri-urban household (PU-H), Peri-urban canal (PU-Ca), Peri-urban cattle house
424 (PU-C), Peri-urban paddy fields (PU-PF). Rural pig farm (R-P), Rural cattle house (R-C),
425 Rural river (R-R), Rural ponds (R-Po), Rural paddy fields (R-PF).

426 **Figure 2.** Population dynamic of the three main JEV vectors in rural and peri-urban areas.

427 **Figure 3.** Average number of the three main species collected per night in the pig farm and
428 cattle house and compared between peri-urban and rural areas. Average comparison of the
429 number of mosquitoes / night was tested with a Tukey HSD test (NS meaning non-significant
430 with $p > 0.05$; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

431 **Figure 4.** Average number of the three main species collected per night in peri-urban and
432 rural areas and compared between the pig farm and cattle house. Average comparison of the
433 number of mosquitoes / night was tested with a Tukey HSD test (NS meaning non-significant
434 with $p > 0.05$; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$).

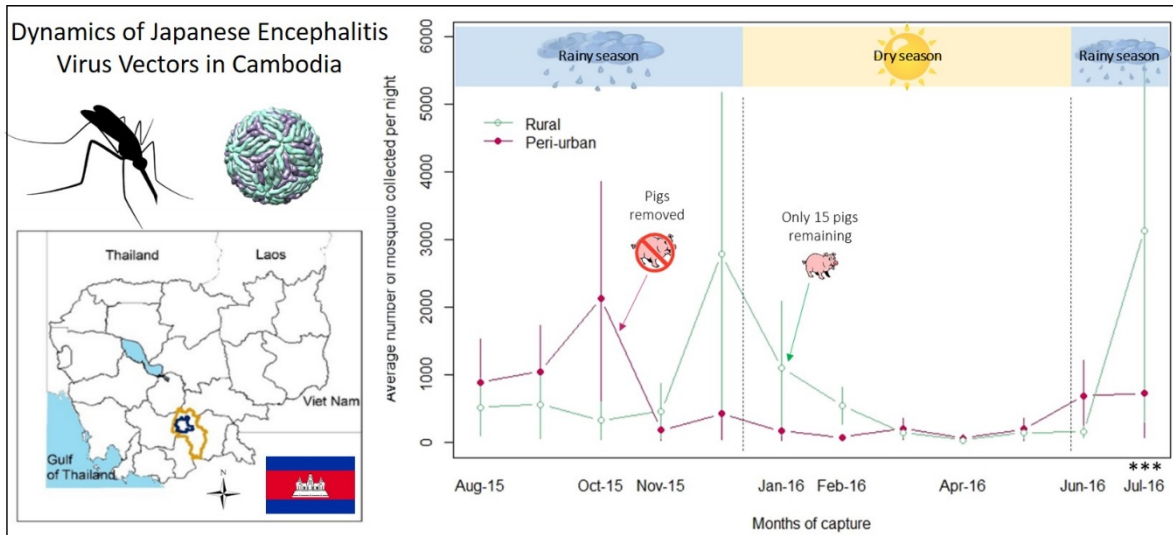
435 **Figure 5.** Average number of the three main species collected per night in paddy fields,
436 river/canals and household/ponds trap locations in peri-urban and rural areas. Average
437 comparison of the number of mosquitoes / night was tested with a Tukey HSD test (NS
438 meaning non-significant with $p > 0.05$; for *Cx vishnui*, a and b means there is a statistical
439 difference with $p < 0.05$).

440 **Highlights.**

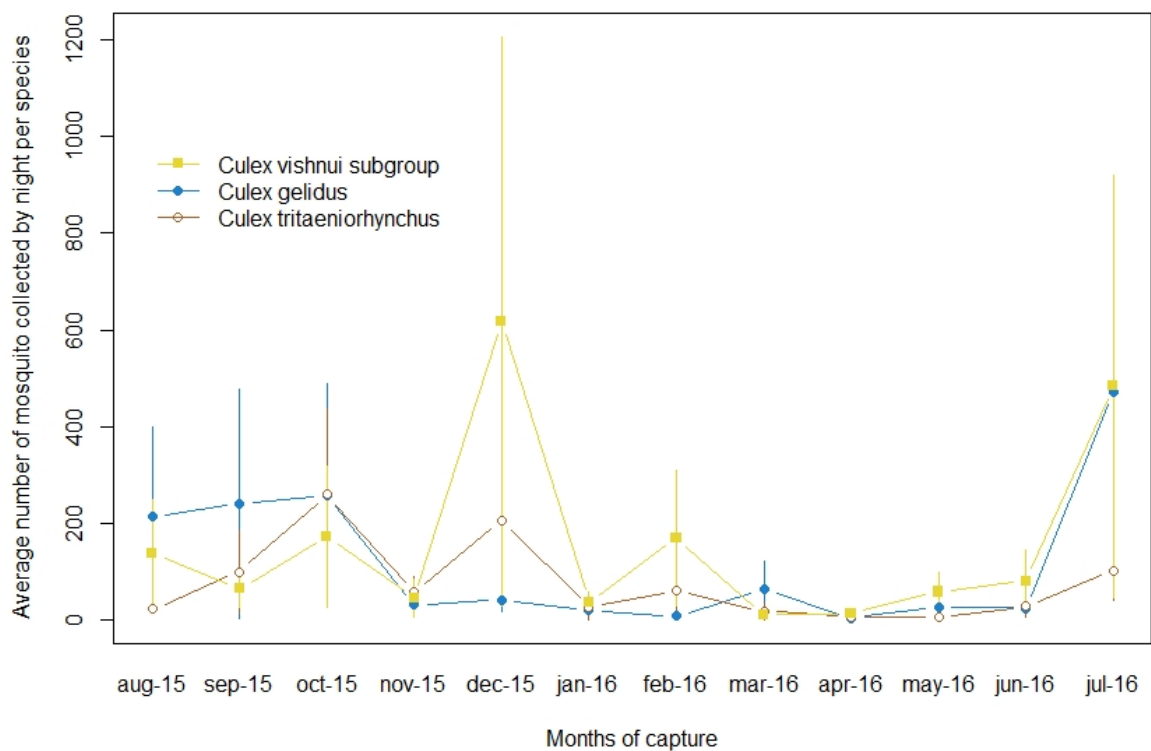
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- 442 • Vectors of Japanese Encephalitis Virus are abundant all the year in Cambodia

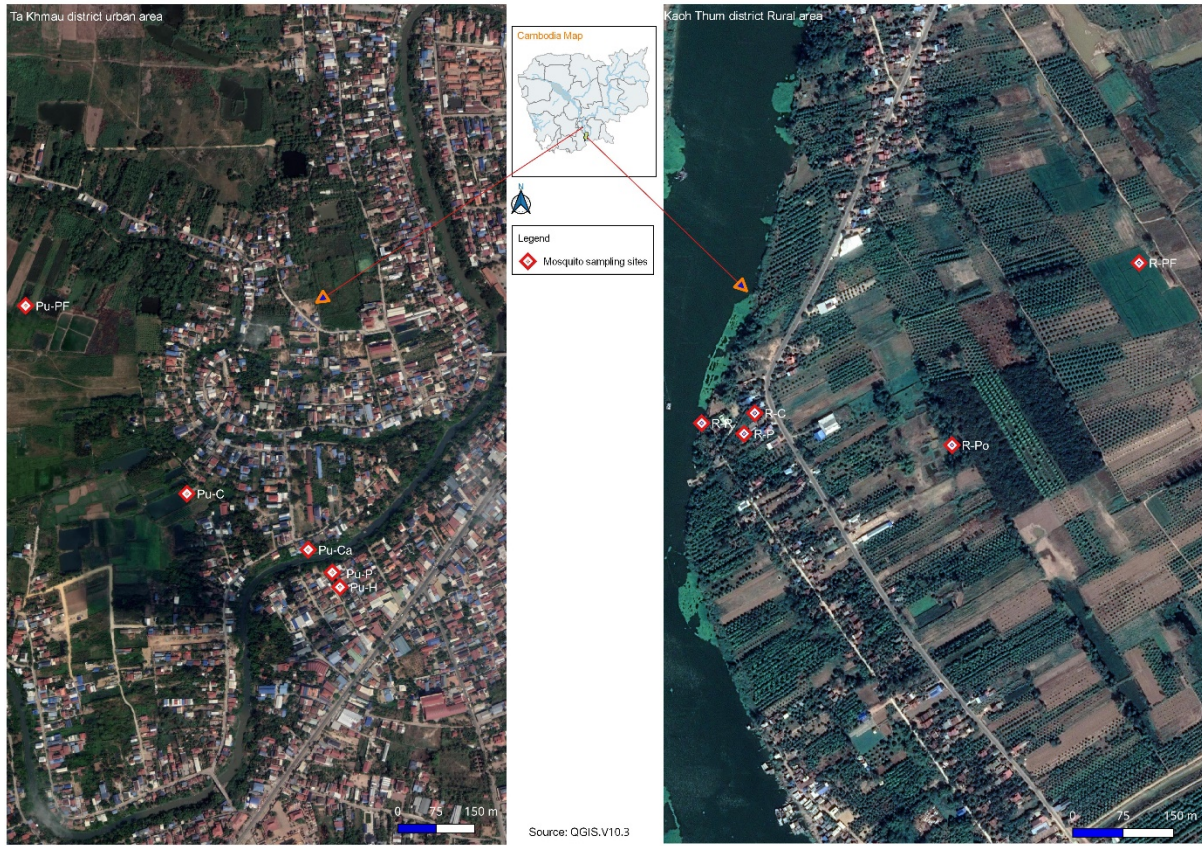
- 443 • One-year study and seasonality of JEV vectors
- 444 • Effect of host presence on JEV vector abundance in pig farms and cattle house
- 445 • The most abundant vectors are *Culex vishnui*, *Cx. gelidus* and *Cx. tritaeniorhynchus*
- 446
- 447



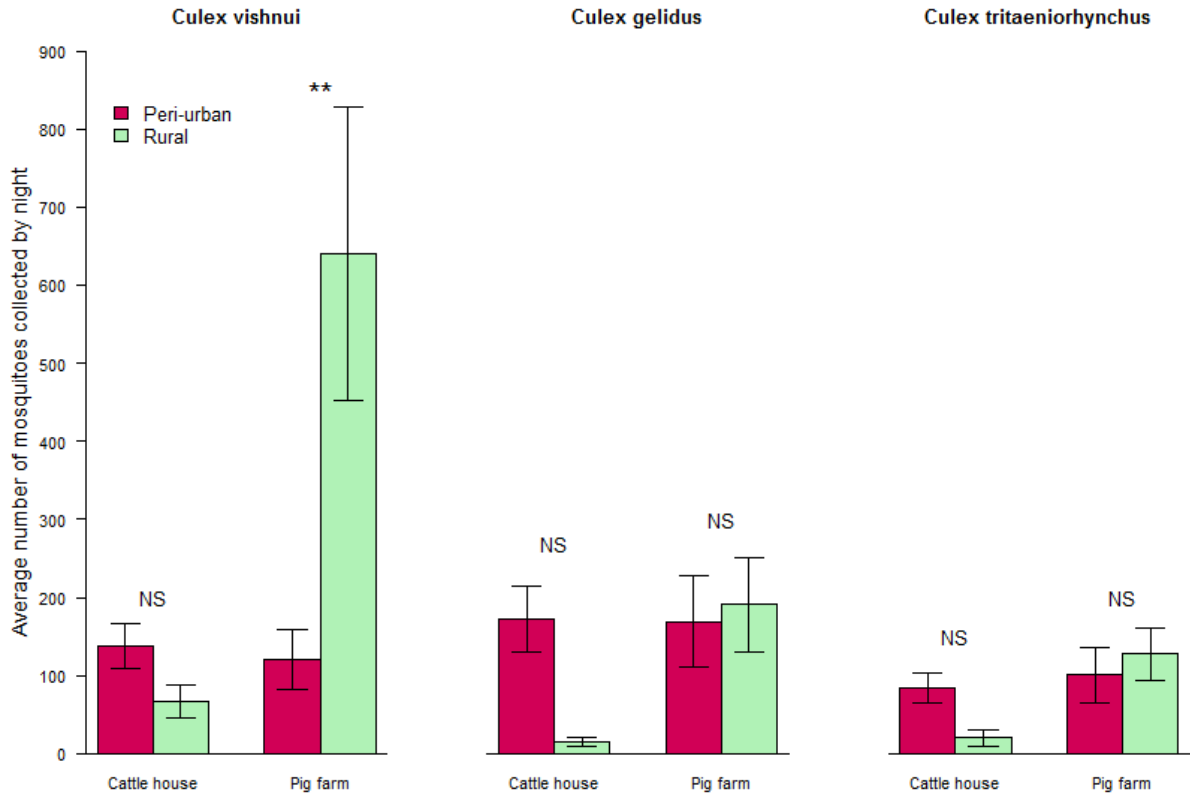
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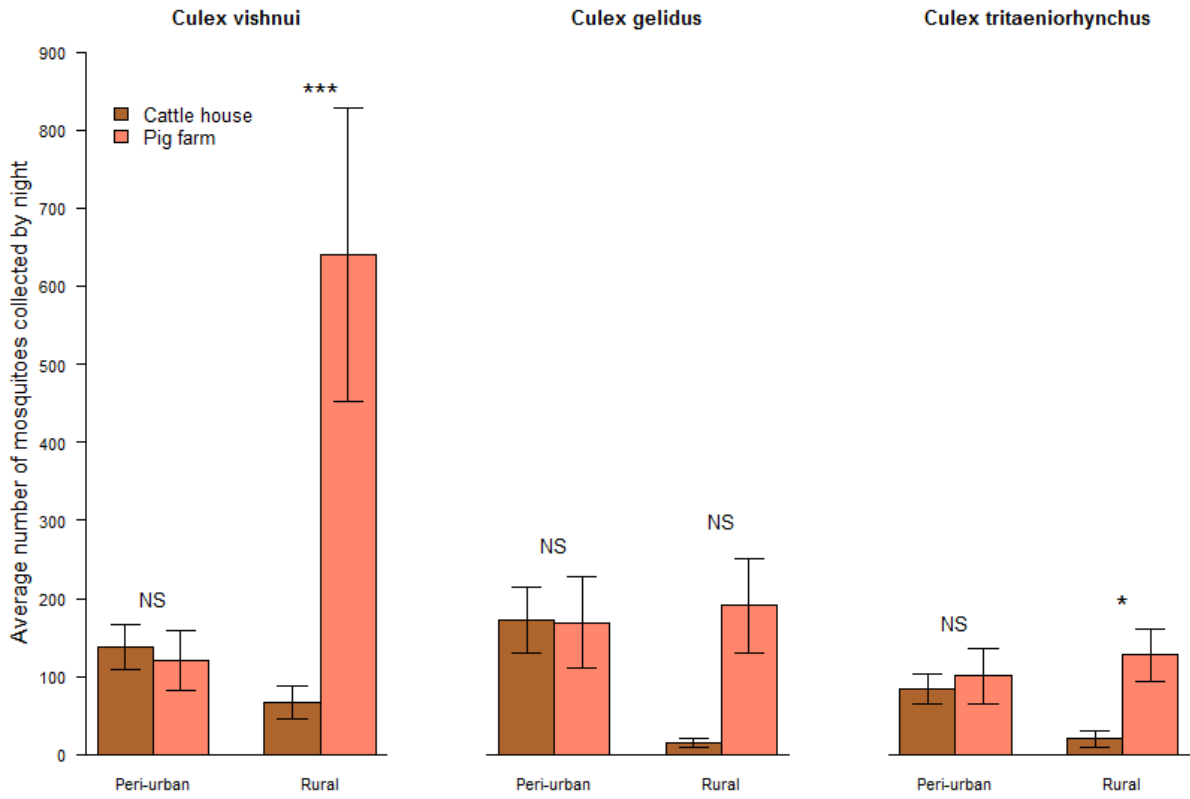
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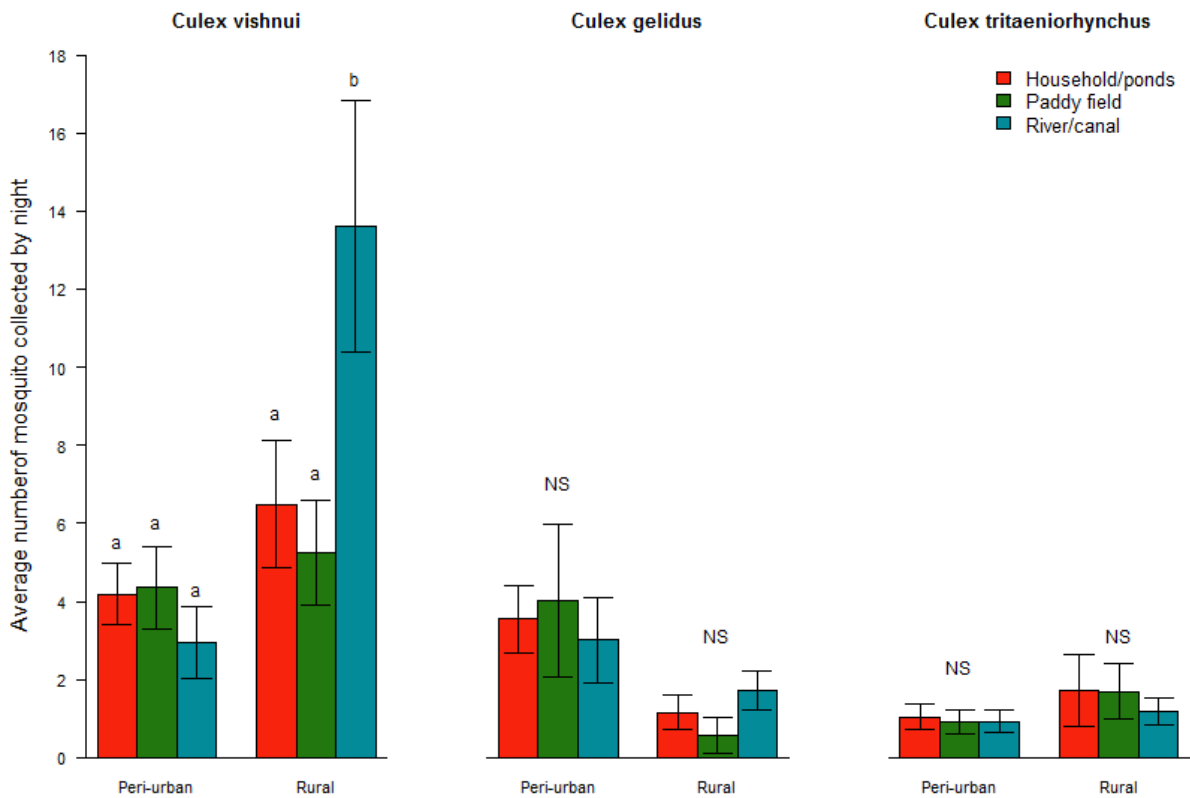
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