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Hélène Martin-Yken, Cécile Formosa, Marion Schiavone, Jean Marie François,

Etienne Dague

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# **Stress, Drug resistance and Adhesion:** a closer look at the dark side of the wall.

# Hélène Martin-Yken, Cécile Formosa, Marion Schiavone, Jean Marie François and Etienne Dague.

1- Université de Toulouse; INSA, UPS, INP; LISBP, 135 Avenue de Rangueil, F-31077 Toulouse, France, 2- INRA, UMR792, Ingénierie des Systèmes Biologiques et des Procédés, F-31400 Toulouse, France, 3- CNRS, UMR5504, F-31400 Toulouse, France

Stress conditions and antifungal drugs induce significant changes in the cell wall composition of yeasts and fungi. They cause modifications of the cell wall molecular architecture, including nature, repartition and attachment of cell wall proteins to the cell surface. Atomic Force Microscopy (AFM) is a powerful tool for studying the morphology, nanomechanical and adhesive properties of live microorganisms under physiological conditions. We imaged and measured the biophysical consequences of various stresses on both S. cereviside and C. albicans cell morphology at the nanoscale, focusing on changes in cell surface aspect and characteristics: roughness, elasticity, and adhesive properties.

## **Heat Shock**

HS induces formation at the cell surface of a circular structure that takes its origin at a single punctuate source and propagates in a concentric manner to reach a diameter of 2–3  $\mu$ m<sup>1</sup>.



Our results suggest that this singular morphological event occurring at the cell surface is due to a dysfunction in the budding machinery and that this phenomenon is controlled by the CWI pathway.

Ultrastructure of the Circular Structure imaged by AFM

AFM deflection images of surface topology of living yeast cell at 30°C (unstressed, A, A') or exposed to heat shock for 1 h at 42°C (B, B'). BS: Bud scar, CS: circular structure.





## Nanoscale effects of Caspofungin on Yeast cell wall



(= 3D Image)

#### Adhesion Maps

A low dose of Caspofungin (0.5 x MIC) results in adhesins expression on the cell surface

Elasticity maps (z range = 0.5 MPa)

Caspofungin induces a deep cell wall remodeling, evidenced by nanomechanichal properties and dramatic chitin increase<sup>2</sup>.



## Measuring and mapping Cellular Adhesion

Molecular Biology tools and AFM established the contribution of previously uncharacterized PGA proteins to C. albicans adhesion mechanism<sup>3</sup>.





#### Adhesion force Maps of $1\mu m^2$ , corresponding histograms representing the adhesion force repartition, and a few representative force curves.



### **Conclusions and outlook**

AFM allowed us to unravel the morphologic effect on yeast cells of Heat Shock, Osmotic Shock, exposure to toxins and drugs. Moreover, using Single Molecule Force Spectroscopy (SMFS) we can now explore the organization of adhesins, map them on the cell surface and quantify their adhesion forces. Altogether, our studies establish the great interest of AFM to explore molecular mechanisms occurring on the cell surface of live fungal cells.

PGA22



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